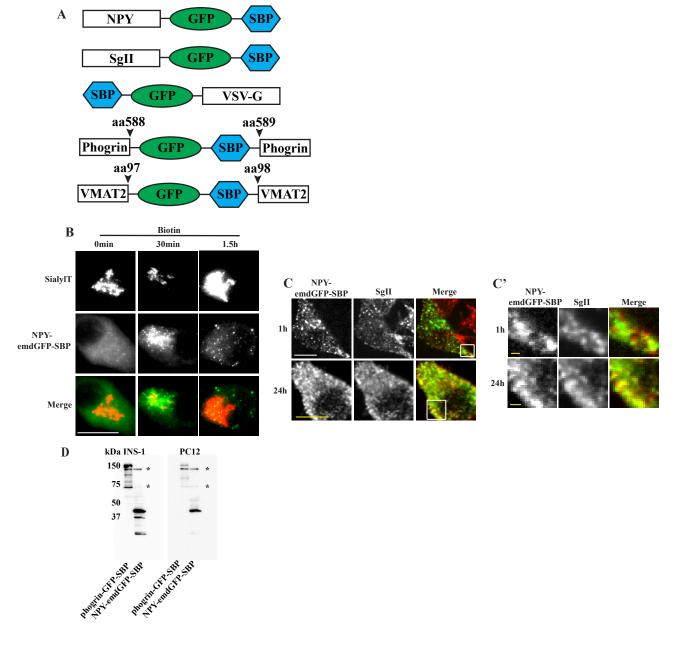
Supplemental Materials Molecular Biology of the Cell

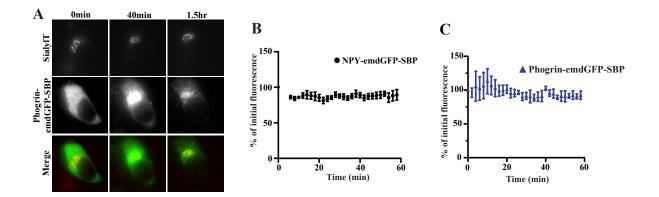
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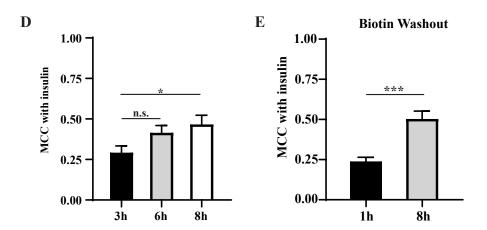


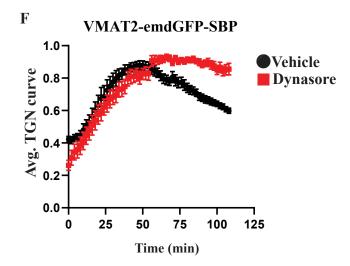
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Figure S1

(A) Diagrams depicting the insertion sites of GFP-SBP for the RUSH constructs used in this study. (B) PC12 cells transfected with NPY-emdGFP-SBP as in Fig. 1, incubated with biotin for the indicated times, fixed and imaged using a widefield epifluorescence microscope. (C) PC12 cells transfected with NPY-emdGFP-SBP and incubated with biotin for 1h (B) or 24h (C) and co-stained for SgII. Insets are shown in C'. (D) Western blot using lysates from INS-1 or PC12 cells transfected with phogrin-emdGFP-SBP or NPY-emdGFP-SBP as indicated. Asterisks indicate non-specific bands. Scale bar indicates 10 μm and 1 μm for insets.







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Figure S2

(A) INS-1 cells were transfected with phogrin-emdGFP-SBP together with the TGN marker sialyltransferase-TagRFP657 and ER-hook, incubated with biotin for the indicated times, fixed and imaged using a widefield epifluorescence microscope. (B,C) INS-1 cells were co-transfected with indicated RUSH cargoes together with an ER-hook and imaged using an inverted widefield microscope. Cells not treated with biotin were imaged as in Fig. 2 to measure photobleaching. n = 3 cells for NPY-emdGFP-SBP and n = 2 for phogrin-emdGFP-SBP. Data shown indicate mean ± SEM. (D) INS-1 cells were transfected with phogrin-emdGFP-SBP and co-stained for insulin at the indicated times. The extent of colocalization with insulin was determined by Manders Correlation Coefficient (MCC) as in Fig. 3. ****p<0.0001 by one-way ANOVA followed by posthoc Tukey t-test (3hr Biotin: n = 13 cells, 6h biotin biotin: n = 12 cells, 8h biotin: n = 8cells). (E) INS-1 cells were transfected as in D, incubated with biotin for 30min, and rinsed extensively. Cells were fixed and co-stained for insulin at 1h or 8h after the washout. The extent of colocalization was determined as in **D**. ***p<0.0001 (1h biotin: n = 10 cells, 8h biotin: n = 10cells). (F) INS-1 cells transfected with VMAT2-emdGFP-SBP were incubated with biotin in presence or absence of 80µM dynasore. Fluorescence within the TGN region was monitored as described in Fig. 3. Average curves of TGN fluorescence are shown. (n=12 and 4 cells from 2 independent transfections for VMAT2-emdGFP-SBP vehicle and dynasore, respectively). Data shown indicate mean \pm SEM.

Movie S1

INS-1 cells transfected with sialyltransferase-TagRFP657 (red), NPY-emdGFP-SBP (green), or SgII-emdGFP-SBP (green), or phogrin-emdGFP-SBP (green), or VMAT2-emdGFP-SBP (green), or SBP-emdGFP-VSV-G (green) and ER hook. Movie shown are played at 15 frames per second. Scale bar indicates 10 µm.

Movie S2

INS-1 cells transfected with NPY-mCherry-SBP (red), sialyltransferase-TagRFP657 (blue), SgII-emdGFP-SBP (green), or phogrin-emdGFP-SBP (green), or SBP-emdGFP-VSV-G (green), and ER hook in presence or absence of dynasore as indicated. Movie shown is 2 min and played at 10 frames per second. Movie begins 40 min after biotin addition. Scale bar indicates 5 µm.