

Supplementary Information

**High-throughput identification of synthetic riboswitches  
by barcode-free amplicon-sequencing in human cells**

Strobel *et al.*

## Supplementary Figure 1

a

b

aagcttgttaccatcgacgttatcccagtccaaacaaaacaaaaggccggcatggtccagccctcgcgtggccggctggcaannntataatcgctggatatggcacgcaagtttacccggcaccgtaaatgtccgactannngcgaatggacgcacaatctcttagcaaacaaaacaagacctggtaaaattgttatccgcgttagatct

C

aagcttgttaccatgcggcgtttcccaactccaaacaaaggcgccgttcgttatccnnnnntataatcgctggatatggcacgcaatgttcta  
ccgggcacccgttaatgtccgactanntacatccagctgtatggatcccataatggacgaaacgcgtcaaacaacaaaggcttgttgcgttatcc  
gttagatct

d

aagcttgtaccatgcggcgttcccgactccaaacaaacaaaccccttaaaggcggttacaaggccgcaaaaatagcagagtaacnnngagaggtaagaatacgcaccacctaggctcgaaagagcctaaacatacnngttaatgcagctttaaaggcaaacaacaaacgcctggtaatgttatccgcttagatct

e

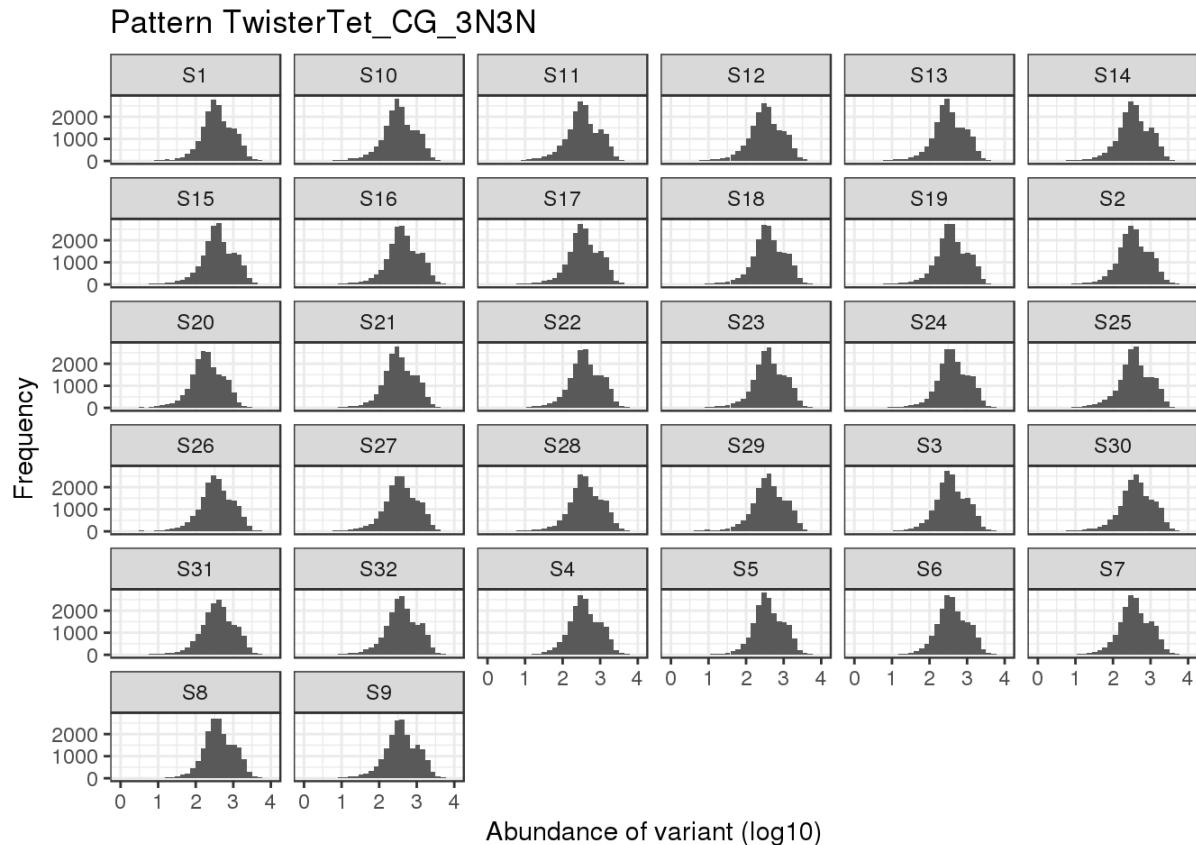
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AAV2 ITRs	CMV promoter	eGFP	riboswitch	randomized	amplification primer
<i>Hind</i> III and <i>Bgl</i> III RE-sites	inserted in "CG"-design	deleted in "noCG"-design			

**Fig. 1: Sequences of riboswitch plasmids.** (a) Tet-hammerhead library plasmid. The same construct with defined instead of randomized nucleotides was also used for the functional characterization of individual aptazyme candidates. (b) Gua-HDV library construct. (c) Gua-HHR library construct. (d) Tet-Twister library construct. The sequence shown here represents the “CG\_3N3N” sub-library. Optionally inserted CG pairs in the so-called “CG”- and “noCG”-designs are indicated (see legend). (e) Gua-U1 library construct. ITR, inverted terminal repeat, CMV, cytomegalovirus, eGFP, enhanced green fluorescent protein, RE= restriction enzyme.

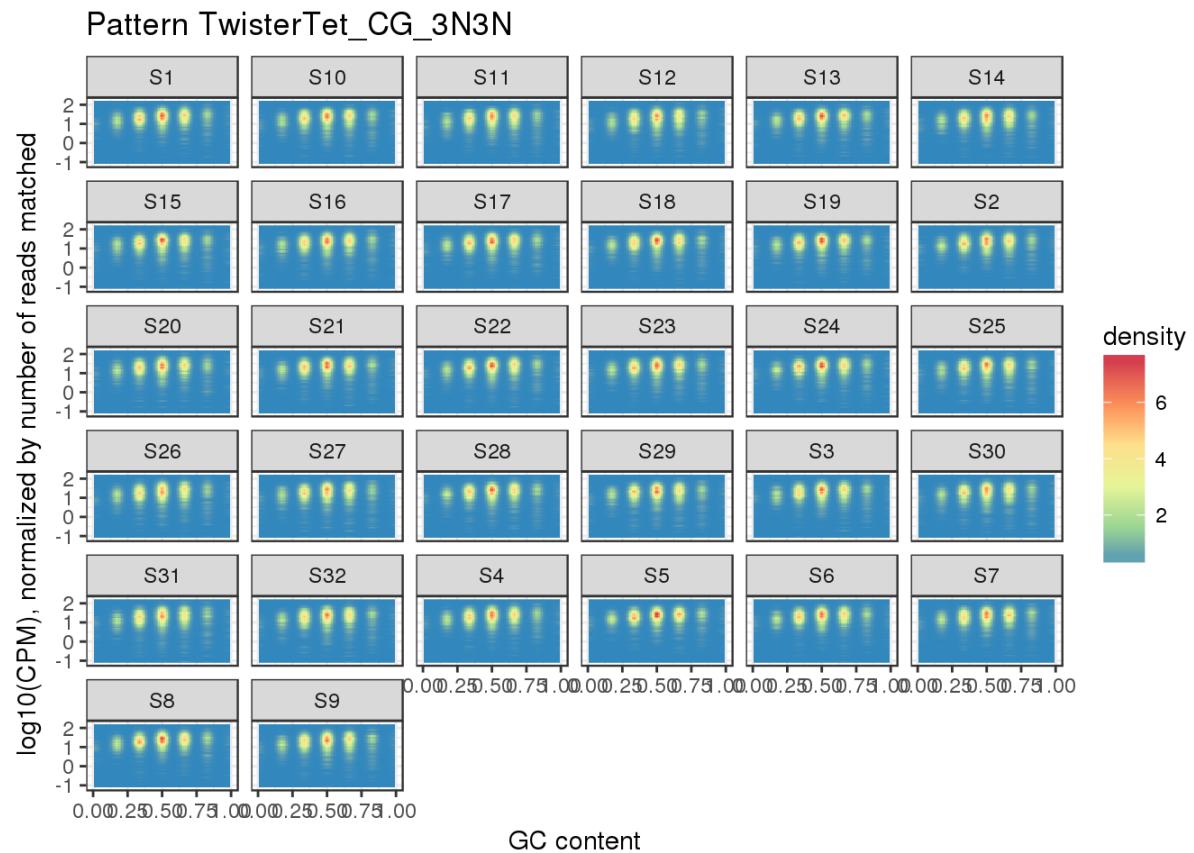
The data analysed in the context of this study were processed using a computational analysis pipeline implemented in bpipe. The plots and tables in Suppl. Figs. 2-6 and 8 are taken out of the HTML report generated by the pipeline.

## Supplementary Figure 2



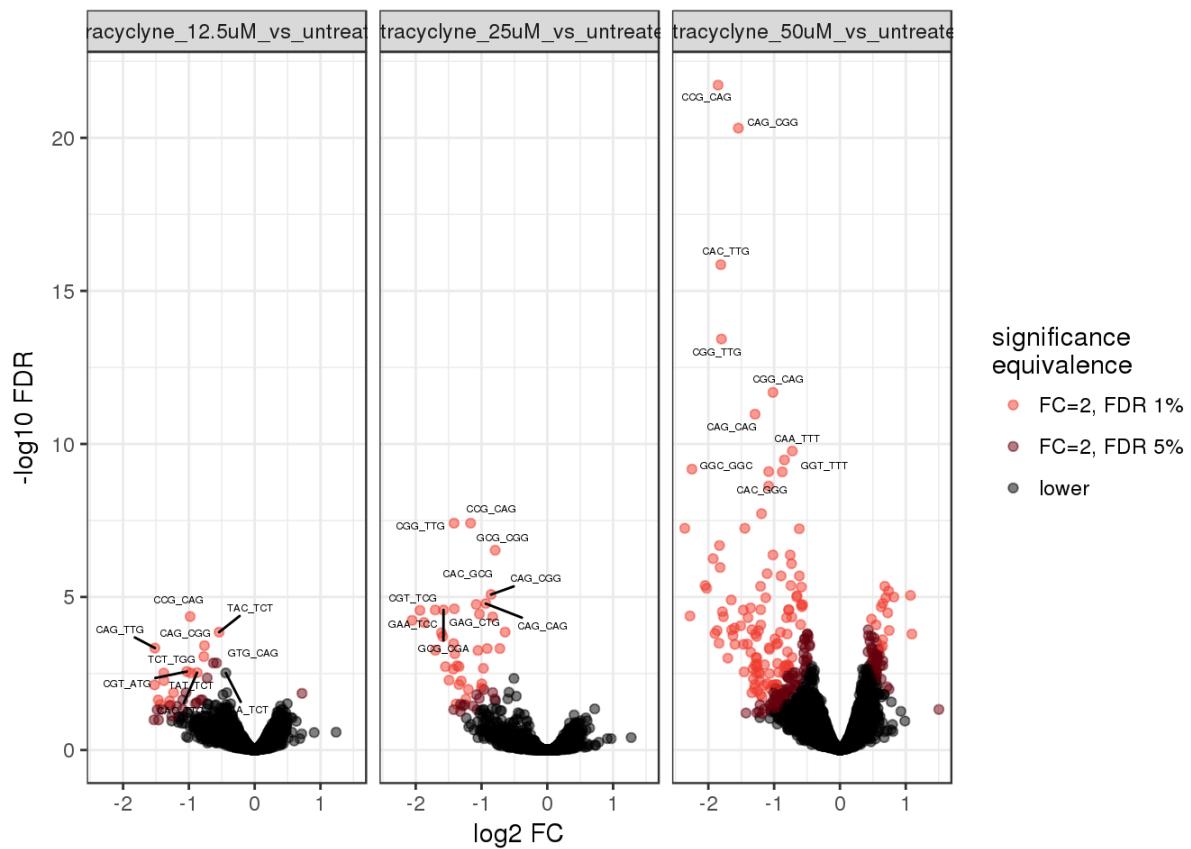
**Fig. 2: Per-sample variant abundance of library constructs (quality measure).** Exemplarily shown is the analysis for the CG\_3N3N Tet-Twister sub-library, screened using 32 replicate samples stimulated with increasing doses of Tet (n=8 per group. S1-S8, untreated; S9-16, 12.5 µM Tet; S17-S24, 25 µM Tet; S25-S32, 50 µM Tet). For each sample, the distribution of raw counts per variant is plotted on a log10 scale. The abundance distribution is consistent across all samples.

### Supplementary Figure 3



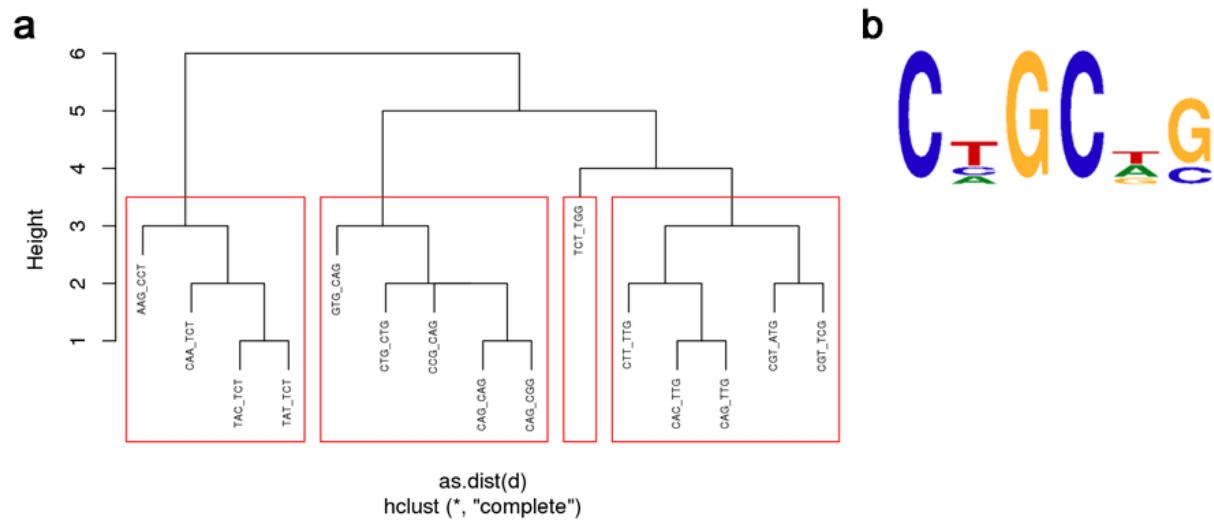
**Fig. 3: GC-dependence of variant abundance (quality measure).** Per-sample plots of the GC-content per variant vs. the log<sub>10</sub>(counts per million, CPM) per variant. The discrete values for GC-content result from the limited possibilities for 6 different nucleotides in the CG\_3N3N example. The dependence of overall variant abundance on the GC-content is generally low, and the samples do not show differences.

## Supplementary Figure 4



**Fig. 4: Volcano plot for differentially abundant variants.** Fold-changes and false discovery rate (FDR), as calculated using edgeR for different contrasts (Tet concentrations 12.5  $\mu\text{M}$ , 25  $\mu\text{M}$  and 50  $\mu\text{M}$  vs. untreated). Different significance levels (FDR 1% and 5%) are marked in red. The variable sequences of the variants with the lowest FDRs are annotated in the plot. In the present example of the Tet-Twister sub-library CG\_3N3N, the number of variants with differential abundance relative to the untreated control grows with increasing Tet doses.

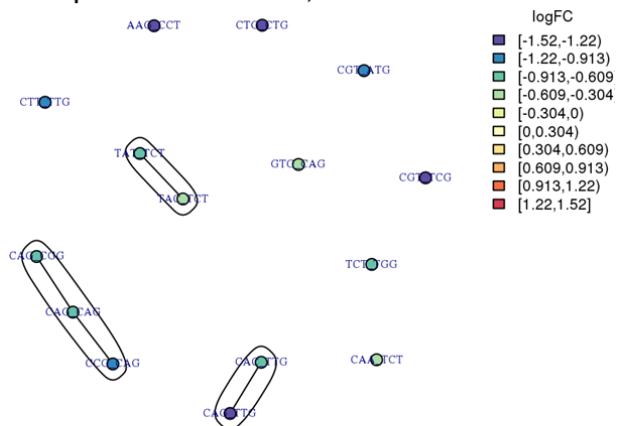
## Supplementary Figure 5



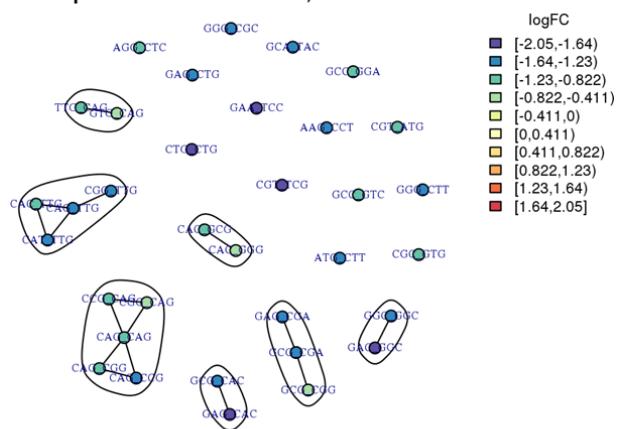
**Fig. 5: Sequence similarity analysis.** (a) Hierarchical clustering of variants with significant differential abundance at an FDR of 1%, based on the Hamming distance. The example shows differentially abundant variants for the Tet-Twister sub-library CG\_3N3N for 12.5  $\mu$ M Tet vs. untreated. (b) Position weight matrix of the variable regions of the second cluster.

## Supplementary Figure 6

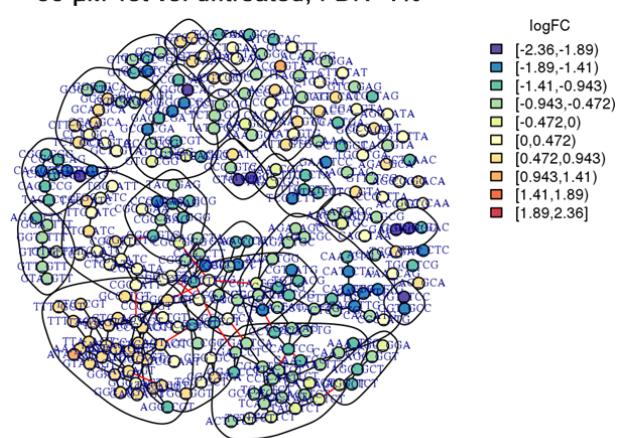
12.5  $\mu$ M Tet vs. untreated, FDR<1%



25  $\mu$ M Tet vs. untreated, FDR<1%

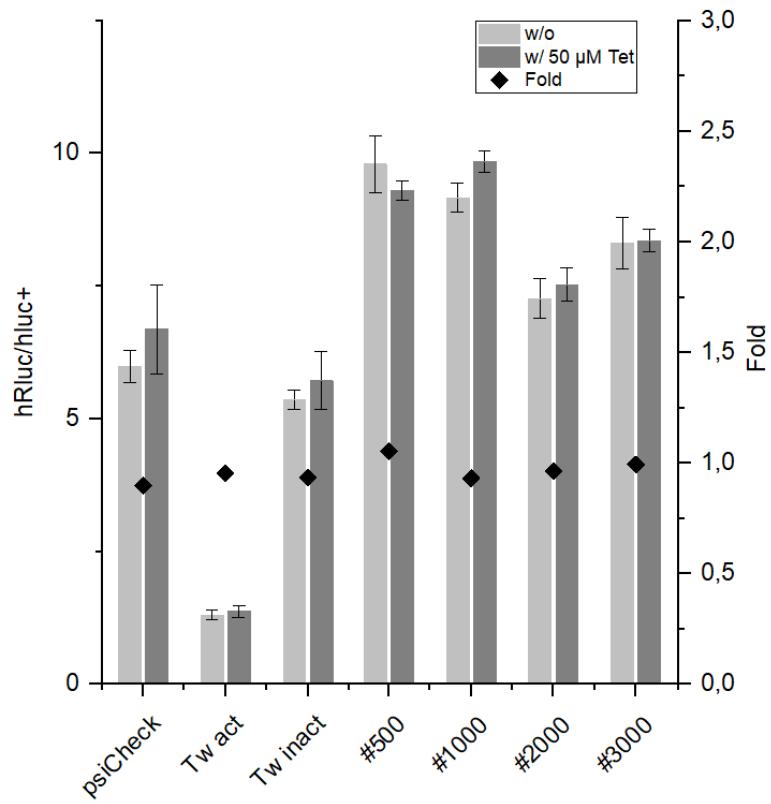


50  $\mu$ M Tet vs. untreated, FDR<1%



**Fig. 6: Network modules of similar variants.** Significant differentially abundant motifs are joined if they have a Hamming distance of 1 (single nucleotide difference only). The resulting network is clustered by community inference based on “edge betweenness”. Additionally, the individual variants are colored by logFC. Blue color indicates significantly lower abundance (negative fold change) in the respective contrast, while red color indicates significantly higher abundance (positive fold change). In this example, differentially abundant variants for the Tet-Twister sub-library CG\_3N3N at 12.5  $\mu$ M Tet vs. untreated, 25  $\mu$ M Tet vs. untreated, and 50  $\mu$ M Tet vs. untreated at an FDR of 1% are shown.

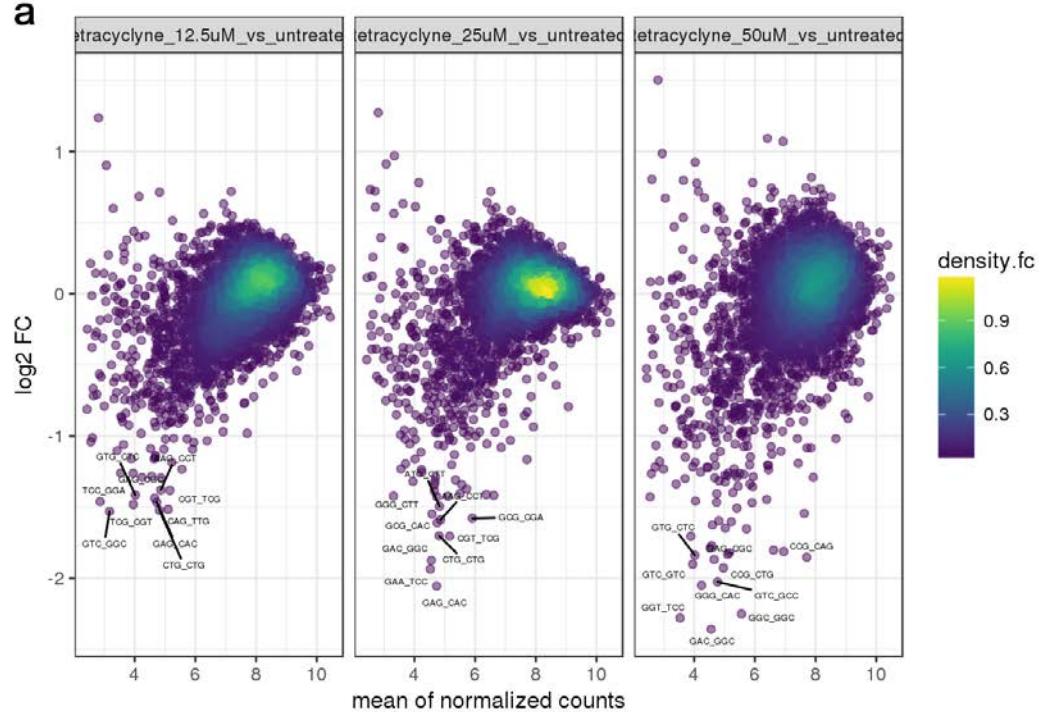
### Supplementary Figure 7



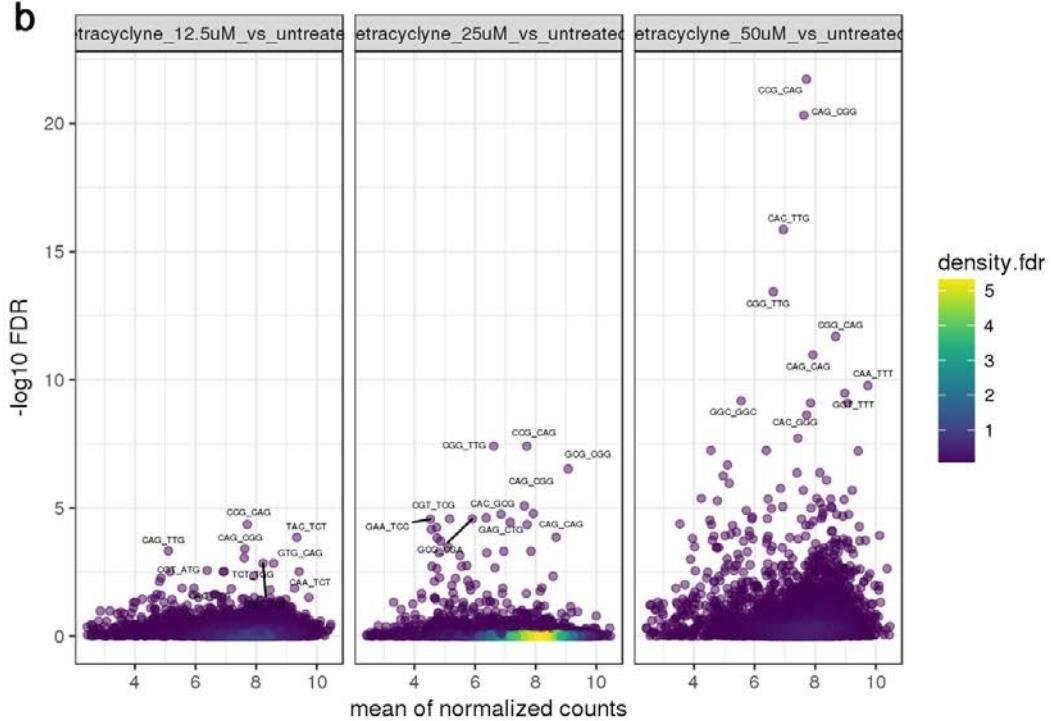
**Fig. 7: Switching performance of Twister constructs not fulfilling positive hit selection criteria.**  
 Shown are luciferase reporter gene expression data in HeLa cells for the constructs that ranked #500, #1000, #2000 and #3000 in the list of all CG\_3N3N constructs that showed a  $\log_2$ -fold change of  $>-0.6$  and FDR $>0.01$ , sorted by  $\log_2$ -fold change at 50  $\mu\text{M}$  Tet stimulation (n=3 replicates, mean  $\pm$  SD).  
 psiCheck: ribozyme-free control; Tw act/inact: constitutively active/inactive ribozyme controls.

## Supplementary Figure 8

**a**

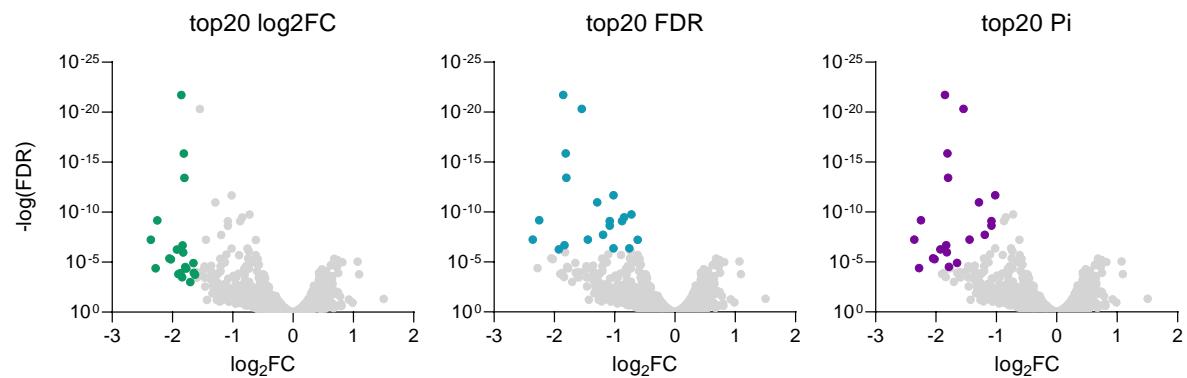


**b**



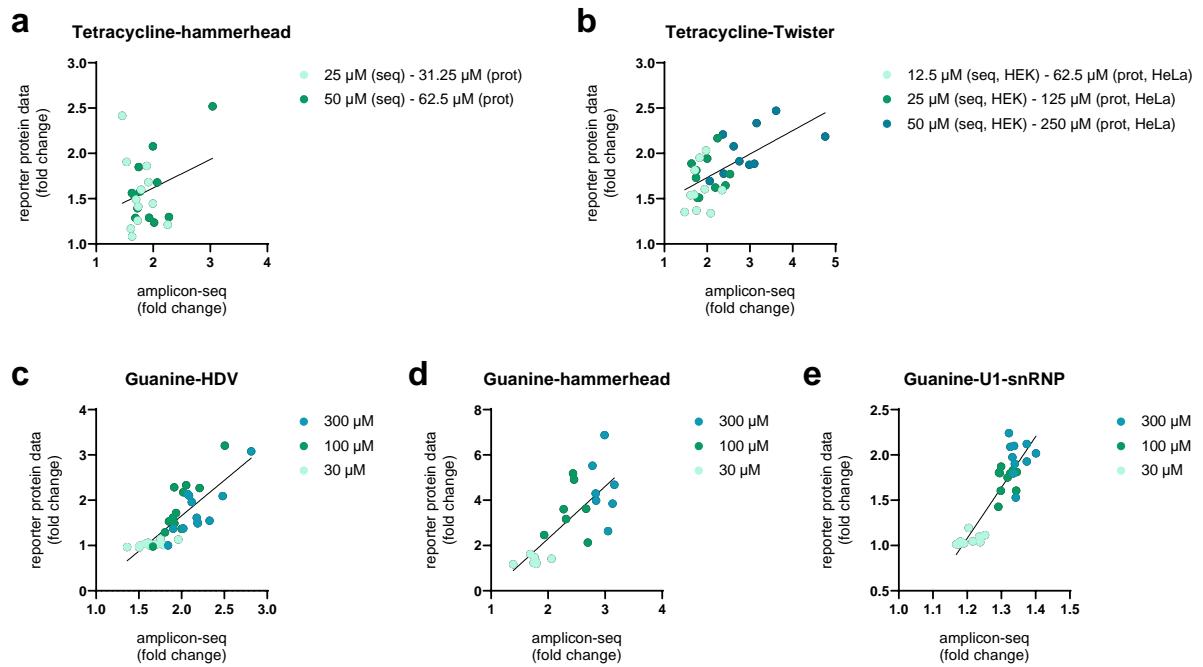
**Fig. 8:  $\log_2$ FC and FDR as a function of construct abundance.** Correlation of mean normalized counts with (a)  $\log_2$ FC and (b)  $-\log_{10}$  FDR, exemplarily shown for the Tet-Twister sub-library CG\_3N3N.

## Supplementary Figure 9



**Fig. 9: Dependence of hit selection on selection parameter used.** Shown are identical, exemplary volcano plots for the CG\_3N3N sub-library contained in the Tet-Twister screen. The top-20 constructs selected based on either ranking of  $\log_2 FC$ , FDR or Pi-values are marked in each graph.

## Supplementary Figure 10



**Fig. 10: Correlation of amplicon-seq-derived and experimentally determined (cellular reporter protein assay) mean fold changes for all functional riboswitch constructs.** (a) Tetracycline hammerhead hit constructs (compare main Fig. 2f). (b) Guanine-HDV constructs (compare main Fig. 3d). (c) Guanine-hammerhead constructs (compare main Fig. 4d). (d) Tetracycline-Twister constructs (compare main Fig. 5c). (e) Guanine-U1-snRNP constructs (compare main Fig. 6e). seq= amplicon-seq-derived data, prot= reporter protein assay-derived data. Regression curves were generated using the “linear regression” feature in GraphPad Prism.

**Supplementary Table 1**

#	Motif	K	logFC		Pvalue		FDR		FDR	
			25 μM	50 μM	logCPM	25 μM	50 μM	25 μM	50 μM	25 μM
1	TAACAG	1,453	2,428	0,944	9,3E-04	3,8E-08	0,330	0,000		
2	GTAATC	0,544	1,605	1,961	1,3E-01	9,0E-06	1,000	0,004		
3	AATAGA	1,031	1,455	2,254	3,5E-03	4,2E-05	0,537	0,012		
4	CGTCTG	0,756	1,193	2,713	1,8E-02	1,9E-04	0,921	0,040		
5	GTATGC	1,173	1,185	3,929	4,6E-08	3,4E-08	0,000	0,000		
6	GATAAC	1,009	1,142	1,800	1,1E-02	3,9E-03	0,777	0,274		
7	GTAAAC	0,211	1,126	1,288	5,9E-01	3,4E-03	1,000	0,256		
8	TAAATC	0,527	1,122	1,919	1,3E-01	1,4E-03	1,000	0,151		
9	CGAACG	0,247	1,113	1,950	5,0E-01	2,2E-03	1,000	0,200		
10	AAAAAA	0,545	1,108	0,998	1,6E-01	4,0E-03	1,000	0,274		
11	CATAAG	0,841	1,059	1,143	2,4E-02	4,5E-03	0,932	0,288		
12	GTGGTG	0,937	1,050	5,166	1,8E-11	5,7E-14	0,000	0,000		
13	GTAGGA	0,938	1,020	2,310	3,8E-03	1,7E-03	0,552	0,170		
14	GTAGGC	0,703	1,011	5,809	1,5E-08	4,7E-16	0,000	0,000		
15	CACCACT	0,627	1,002	1,823	6,3E-02	2,9E-03	1,000	0,235		
16	CAACCG	0,356	1,000	1,411	3,1E-01	4,1E-03	1,000	0,279		
17	GTGGTAC	K19	0,617	0,995	3,405	1,8E-02	1,4E-04	0,921	0,031	
18	GTGTTA	0,394	0,973	4,546	2,5E-02	3,6E-08	0,952	0,000		
19	AATATG	0,762	0,953	1,694	3,7E-02	9,1E-03	1,000	0,379		
20	GTATTAA	0,789	0,948	4,680	1,2E-06	6,0E-09	0,003	0,000		
21	GTGGC GT	0,658	0,941	7,750	3,2E-13	2,6E-25	0,000	0,000		
22	GTAAGCT	0,535	0,938	4,849	6,9E-04	3,0E-09	0,277	0,000		
23	CAAATG	0,070	0,934	0,375	8,7E-01	3,1E-02	1,000	0,535		
24	TAACAG	1,310	0,933	1,287	1,1E-03	2,0E-02	0,352	0,467		
25	ATAATAG	0,411	0,929	1,711	2,5E-01	9,1E-03	1,000	0,379		
26	GTGAGCT	0,660	0,926	5,493	1,5E-07	2,0E-13	0,000	0,000		
27	AAAGATG	0,561	0,925	2,781	5,6E-02	1,6E-03	1,000	0,168		
28	GTAGAAC	0,452	0,910	2,749	1,3E-01	2,6E-03	1,000	0,223		
29	GATAACG	0,620	0,909	2,172	7,7E-02	9,6E-03	1,000	0,383		
30	TTGAACC	0,634	0,905	2,367	5,1E-02	5,5E-03	1,000	0,308		
31	CACATGG	-0,408	0,884	1,391	3,2E-01	2,8E-02	1,000	0,516		
32	CACGAG	0,388	0,864	2,372	2,3E-01	7,3E-03	1,000	0,350		
33	CAAGGCC	0,279	0,842	4,139	1,6E-01	1,9E-05	1,000	0,006		
34	TAAGACG	0,694	0,839	2,790	1,9E-02	4,5E-03	0,921	0,289		
35	TAATAAG	0,883	0,835	0,502	5,8E-02	7,3E-02	1,000	0,649		
36	GTACCGCG	0,995	0,827	3,354	5,1E-05	7,6E-04	0,038	0,103		
37	TAATAAT	0,541	0,818	2,323	9,5E-02	1,2E-02	1,000	0,403		
38	CTTAAAGG	-0,152	0,814	1,547	6,7E-01	2,0E-02	1,000	0,462		
39	GTAGGTC	0,916	0,804	4,367	6,7E-07	1,3E-05	0,002	0,005		
40	GTGATAT	0,600	0,803	5,235	1,1E-05	4,3E-09	0,011	0,000		
41	CAGTCGG	-0,290	0,799	1,353	4,5E-01	3,7E-02	1,000	0,555		
42	AAACATA	-0,453	0,797	1,470	2,1E-01	2,5E-02	1,000	0,506		
43	CGATAG	0,580	0,797	1,552	1,3E-01	3,7E-02	1,000	0,555		
44	TAACAG	0,400	0,789	2,048	2,5E-01	2,3E-02	1,000	0,483		
45	TCATAAG	0,618	0,788	1,929	9,0E-02	3,1E-02	1,000	0,535		
46	CTAACGT	0,797	0,786	3,954	1,0E-04	1,3E-04	0,063	0,029		
47	CAGGC GG	0,203	0,786	0,336	6,0E-01	4,1E-02	1,000	0,574		
48	CGAGTAC	-0,118	0,783	3,480	6,3E-01	1,5E-03	1,000	0,160		
49	CGGTA GG	0,398	0,781	2,675	1,9E-01	9,6E-03	1,000	0,383		
50	GTAACGA	0,503	0,776	5,690	2,6E-05	9,4E-11	0,021	0,000		
51	GTAATGT	0,484	0,774	5,000	1,8E-03	6,6E-07	0,442	0,000		
52	CACTTCA	0,692	0,772	2,841	1,9E-02	9,1E-03	0,921	0,379		
53	CTTTACG	0,672	0,771	2,421	4,3E-02	2,0E-02	1,000	0,467		
54	TAGTAC	0,495	0,769	1,632	1,7E-01	3,2E-02	1,000	0,536		
55	ATGAAAC	0,524	0,767	2,834	9,7E-02	1,5E-02	1,000	0,431		
56	AGTTAAG	0,277	0,760	3,271	3,0E-01	4,8E-03	1,000	0,297		
57	CATCCGG	0,443	0,758	1,230	2,8E-01	6,5E-02	1,000	0,635		
58	CTAGTGC	0,685	0,756	4,858	5,6E-06	5,6E-07	0,008	0,000		
59	GTATCGT	0,336	0,755	5,564	5,8E-03	5,8E-10	0,641	0,000		
60	AGACTAG	0,352	0,752	3,132	1,8E-01	4,5E-03	1,000	0,288		
61	GAAGGAC	0,191	0,752	2,230	5,5E-01	1,8E-02	1,000	0,448		
62	CACAACT	0,997	0,750	2,293	4,6E-03	3,3E-02	0,599	0,541		
63	AGAACAA	0,419	0,749	2,080	2,3E-01	3,2E-02	1,000	0,536		
64	GTGAAAT	0,607	0,747	3,806	4,7E-03	5,2E-04	0,611	0,083		
65	TGTTAA	-0,131	0,747	2,498	6,6E-01	1,2E-02	1,000	0,408		
66	GTATCGG	K5	0,527	0,744	5,897	2,7E-05	3,2E-09	0,021	0,000	
67	CGCGCAC	0,599	0,743	1,772	6,3E-02	2,1E-02	1,000	0,475		
68	GGAGTAC	0,767	0,738	4,363	1,5E-05	3,2E-05	0,014	0,010		
69	CACTCGG	0,985	0,734	1,572	1,0E-02	5,6E-02	0,777	0,615		
70	TATCAAG	0,411	0,734	1,914	2,3E-01	3,1E-02	1,000	0,535		
71	ATGATAA	0,159	0,728	2,181	6,2E-01	2,2E-02	1,000	0,479		
72	CGTTAAG	0,220	0,728	2,515	4,8E-01	1,9E-02	1,000	0,459		
73	CAATAAT	0,344	0,725	2,119	3,1E-01	3,4E-02	1,000	0,543		
74	TAGAAAC	0,337	0,723	0,564	4,0E-01	7,3E-02	1,000	0,649		
75	CTATACC	-0,398	0,723	1,947	2,3E-01	2,9E-02	1,000	0,526		
76	GTACCGCT	0,088	0,720	3,787	6,9E-01	1,3E-03	1,000	0,146		
77	CTAGGCC	0,536	0,715	4,925	6,1E-04	4,9E-06	0,255	0,002		
78	CGCAAGG	0,078	0,713	2,830	7,8E-01	1,1E-02	1,000	0,390		
79	CCAACAA	-0,088	0,713	1,151	8,2E-01	6,7E-02	1,000	0,636		
80	GTGTTTC	0,605	0,711	4,664	2,2E-04	1,5E-05	0,126	0,005		
81	GTGTTTA	0,403	0,710	3,561	9,0E-02	2,9E-03	1,000	0,233		
82	GTAGGTT	0,422	0,708	5,974	4,5E-04	3,8E-09	0,215	0,000		
83	GAGTAC	0,366	0,704	2,506	2,5E-01	2,6E-02	1,000	0,509		
84	AAAATCC	0,532	0,704	2,213	9,4E-02	2,7E-02	1,000	0,514		
85	GTGGAATC	0,837	0,701	5,019	6,3E-08	5,9E-06	0,000	0,002		
86	GAAACCG	0,590	0,700	0,776	1,5E-01	8,5E-02	1,000	0,677		
87	GTAGTAT	K11	0,539	0,699	4,116	9,5E-03	7,8E-04	0,766	0,104	
88	GCACCG	0,499	0,697	4,703	1,7E-03	1,2E-05	0,441	0,004		
89	AAGAGCC	0,352	0,697	2,025	3,7E-01	7,6E-02	1,000	0,656		
90	TAATCAG	0,987	0,696	2,221	3,8E-03	4,1E-02	0,552	0,574		
91	AAAATAG	-0,265	0,695	0,833	5,6E-01	1,3E-01	1,000	0,724		
92	GATATCT	0,511	0,694	3,358	3,5E-02	4,3E-03	0,998	0,287		
93	CTTAAAG	0,178	0,690	3,261	5,2E-01	1,3E-02	1,000	0,409		
94	GGTAAAC	0,077	0,690	2,316	8,1E-01	3,4E-02	1,000	0,543		
95	CACTGG	-0,498	0,689	1,771	1,6E-01	4,8E-02	1,000	0,606		
96	CAGCCGG	0,276	0,688	3,694	2,2E-01	2,2E-03	1,000	0,200		
97	TAGTCAG	1,218	0,685	1,578	6,2E-04	5,4E-02	0,255	0,614		
98	GACGC	0,396	0,685	1,870	2,2E-01	3,3E-02	1,000	0,541		
99	GTACGAC	0,731	0,685	2,510	1,8E-02	2,7E-02	0,921	0,514		
100	CGATTCT	0,528	0,684	3,019	5,1E-02	1,2E-02	1,000	0,403		

**Tab. 1: Top 100 constructs from the Tet-hammerhead screen, ranked by their fold change at 50 μM Tet (here, no further cutoffs were applied). K\* constructs were contained in the original publication by Beilstein and colleagues.**

**Supplementary Table 2**

#	Motif	GuaM*	logCPM	logFC 30 µM	logFC 100 µM	logFC 300 µM	P-value 30 µM	P-value 100 µM	P-value 300 µM	FDR 30 µM	FDR 100 µM	FDR 300 µM
1	TGC_GTA	GuaM8HDV	6.68	-0.971	-1.325	-1.492	2.20E-16	2.20E-16	2.20E-16	2.05E-14	1.50E-14	1.07E-14
2	TGG_CTA		6.68	-0.837	-1.144	-1.311	2.20E-16	2.20E-16	2.20E-16	2.05E-14	1.50E-14	1.07E-14
3	TGA_TTA		8.16	-0.588	-1.000	-1.218	2.20E-16	2.20E-16	2.20E-16	2.05E-14	1.50E-14	1.07E-14
4	CGG_CTG		5.19	-0.768	-0.926	-1.129	2.20E-16	2.20E-16	2.20E-16	2.05E-14	1.50E-14	1.07E-14
5	CGG_TCG		5.39	-0.703	-0.951	-1.129	2.20E-16	2.20E-16	2.20E-16	2.05E-14	1.50E-14	1.07E-14
6	TGT_ATA	GuaM12HDV	7.58	-0.447	-0.852	-1.121	2.20E-16	2.20E-16	2.20E-16	2.05E-14	1.50E-14	1.07E-14
7	CGT_ACG		5.25	-0.687	-0.937	-1.083	2.20E-16	2.20E-16	2.20E-16	2.05E-14	1.50E-14	1.07E-14
8	CGC_GCG		3.99	-0.809	-1.038	-1.061	2.20E-16	2.20E-16	2.20E-16	2.05E-14	1.50E-14	1.07E-14
9	CGT_GCG		3.83	-0.728	-1.014	-1.050	2.20E-16	2.20E-16	2.20E-16	2.05E-14	1.50E-14	1.07E-14
10	CAA_TTG		7.01	-0.656	-0.888	-1.014	2.20E-16	2.20E-16	2.20E-16	2.05E-14	1.50E-14	1.07E-14
11	CGC_GTG		4.18	-0.596	-0.938	-0.927	2.20E-16	2.20E-16	2.20E-16	2.05E-14	1.50E-14	1.07E-14
12	CAC_GTG		5.50	-0.594	-0.736	-0.881	2.20E-16	2.20E-16	2.20E-16	2.05E-14	1.50E-14	1.07E-14
13	CGA_TCG		5.27	-0.663	-0.777	-0.880	2.20E-16	2.20E-16	2.20E-16	2.05E-14	1.50E-14	1.07E-14
14	CGA_TTG		6.00	-0.423	-0.673	-0.878	2.20E-16	2.20E-16	2.20E-16	2.05E-14	1.50E-14	1.07E-14
15	CGA_CCG		6.76	-0.303	-0.678	-0.877	2.20E-16	2.20E-16	2.20E-16	2.05E-14	1.50E-14	1.07E-14
16	CGT_ATG		6.56	-0.405	-0.642	-0.828	2.20E-16	2.20E-16	2.20E-16	2.05E-14	1.50E-14	1.07E-14
17	CGG_CCG		4.35	-0.591	-0.845	-0.800	2.20E-16	2.20E-16	2.20E-16	2.05E-14	1.50E-14	1.07E-14
18	CGC_ACG		6.06	-0.221	-0.540	-0.798	1.22E-08	2.20E-16	2.20E-16	9.27E-07	1.50E-14	1.07E-14
19	CCA_TGG		4.14	-0.527	-0.648	-0.780	2.20E-16	2.20E-16	2.20E-16	2.05E-14	1.50E-14	1.07E-14
20	CAT_ATG		5.97	-0.495	-0.614	-0.756	2.20E-16	2.20E-16	2.20E-16	2.05E-14	1.50E-14	1.07E-14
21	CAG_CTG		5.60	-0.494	-0.612	-0.739	2.20E-16	2.20E-16	2.20E-16	2.05E-14	1.50E-14	1.07E-14
22	TGG_TTA		8.13	-0.243	-0.480	-0.733	2.20E-16	2.20E-16	2.20E-16	2.05E-14	1.50E-14	1.07E-14
23	CCG_TGG		3.92	-0.440	-0.534	-0.732	1.08E-10	7.56E-15	2.20E-16	8.81E-09	4.92E-13	1.07E-14
24	CGA_GCG		6.22	-0.095	-0.436	-0.718	1.01E-02	2.20E-16	2.20E-16	2.36E-01	1.50E-14	1.07E-14
25	CAG_TTG		6.39	-0.283	-0.519	-0.696	2.20E-16	2.20E-16	2.20E-16	2.05E-14	1.50E-14	1.07E-14
26	TGT ACA		8.08	-0.430	-0.555	-0.682	2.20E-16	2.20E-16	2.20E-16	2.05E-14	1.50E-14	1.07E-14
27	CAT_GTG		5.76	-0.317	-0.494	-0.680	2.20E-16	2.20E-16	2.20E-16	2.05E-14	1.50E-14	1.07E-14
28	CGG_GCG		4.83	-0.144	-0.387	-0.675	4.08E-03	3.47E-14	2.20E-16	1.38E-01	2.18E-12	1.07E-14
29	CGT_CCG		5.63	-0.218	-0.429	-0.664	1.05E-07	2.20E-16	2.20E-16	7.65E-06	1.50E-14	1.07E-14
30	CTA_TAG		7.42	-0.431	-0.563	-0.636	2.20E-16	2.20E-16	2.20E-16	2.05E-14	1.50E-14	1.07E-14
31	TGT_GCA		7.39	-0.319	-0.501	-0.636	2.20E-16	2.20E-16	2.20E-16	2.05E-14	1.50E-14	1.07E-14
32	CGG_TTG		6.57	-0.163	-0.378	-0.601	1.38E-09	2.20E-16	2.20E-16	1.09E-07	1.50E-14	1.07E-14
33	CGT_TCG		6.15	-0.163	-0.449	-0.599	5.22E-08	2.20E-16	2.20E-16	3.89E-06	1.50E-14	1.07E-14
34	TGA_TCA		8.43	-0.425	-0.561	-0.597	2.20E-16	2.20E-16	2.20E-16	2.05E-14	1.50E-14	1.07E-14
35	CAG_CCG		6.57	-0.242	-0.509	-0.595	2.20E-16	2.20E-16	2.20E-16	2.05E-14	1.50E-14	1.07E-14
36	TGC_GCA		6.83	-0.461	-0.591	-0.579	2.20E-16	2.20E-16	2.20E-16	2.05E-14	1.50E-14	1.07E-14
37	CGG_ACG		6.36	-0.117	-0.357	-0.560	2.92E-05	2.20E-16	2.20E-16	1.96E-03	1.50E-14	1.07E-14
38	TCA_TGA		7.11	-0.391	-0.523	-0.549	2.20E-16	2.20E-16	2.20E-16	2.05E-14	1.50E-14	1.07E-14
39	TCG_TGA		6.22	-0.286	-0.418	-0.536	2.20E-16	2.20E-16	2.20E-16	2.05E-14	1.50E-14	1.07E-14
40	CTC_GAG		5.91	-0.317	-0.449	-0.502	2.20E-16	2.20E-16	2.20E-16	2.05E-14	1.50E-14	1.07E-14
41	CCT_AGG		5.93	-0.382	-0.448	-0.492	2.20E-16	2.20E-16	2.20E-16	2.05E-14	1.50E-14	1.07E-14
42	CGA_ACG		6.24	-0.091	-0.190	-0.470	1.21E-02	1.82E-07	2.20E-16	2.51E-01	8.98E-06	1.07E-14
43	CAC_GCG		4.53	-0.320	-0.397	-0.466	8.29E-09	1.33E-12	2.20E-16	6.41E-07	8.00E-11	1.07E-14
44	CCG_CGG		3.35	-0.182	-0.310	-0.460	1.99E-02	8.69E-05	7.02E-09	3.36E-01	2.95E-03	2.55E-07
45	CCT_GGG		5.14	-0.167	-0.412	-0.456	1.41E-03	7.93E-15	2.20E-16	6.08E-02	5.08E-13	1.07E-14
46	TCC_GGA		6.52	-0.299	-0.392	-0.447	2.20E-16	2.20E-16	2.20E-16	2.05E-14	1.50E-14	1.07E-14
47	TGG_TCA		7.66	-0.259	-0.362	-0.444	2.20E-16	2.20E-16	2.20E-16	2.05E-14	1.50E-14	1.07E-14
48	CTT_AAG		7.76	-0.258	-0.363	-0.443	2.20E-16	2.20E-16	2.20E-16	2.05E-14	1.50E-14	1.07E-14
49	CGC_TCG		4.83	-0.143	-0.263	-0.437	7.64E-03	1.09E-06	9.71E-16	2.09E-01	4.85E-05	4.57E-14
50	TCT_AGA		7.55	-0.273	-0.374	-0.422	2.20E-16	2.20E-16	2.20E-16	2.05E-14	1.50E-14	1.07E-14
51	TTC_GTA		7.83	-0.174	-0.319	-0.418	2.20E-16	2.20E-16	2.20E-16	2.05E-14	1.50E-14	1.07E-14
52	TAA_TTA	GuaM7HDV	8.82	-0.186	-0.323	-0.394	2.20E-16	2.20E-16	2.20E-16	2.05E-14	1.50E-14	1.07E-14
53	CCC_GGG		5.65	-0.305	-0.435	-0.394	2.20E-16	2.20E-16	2.20E-16	2.05E-14	1.50E-14	1.07E-14
54	CCT_TGG		6.37	-0.074	-0.279	-0.385	2.11E-02	2.20E-16	2.20E-16	3.40E-01	1.50E-14	1.07E-14
55	TCG_CGA		6.31	-0.275	-0.394	-0.371	4.27E-12	2.20E-16	2.20E-16	3.64E-10	1.50E-14	1.07E-14
56	CGC_CCG		4.58	-0.157	-0.260	-0.364	1.99E-02	1.21E-04	8.24E-08	3.36E-01	3.87E-03	2.84E-06
57	CCA_CGG		5.34	-0.067	-0.161	-0.356	1.65E-01	9.35E-04	4.04E-13	7.32E-01	2.29E-02	1.78E-11
58	TCT_GGA		7.51	-0.087	-0.208	-0.342	1.29E-04	2.20E-16	2.20E-16	8.01E-03	1.50E-14	1.07E-14
59	TGT_GTA	GuaM11HDV	8.05	-0.059	-0.185	-0.341	9.72E-04	2.20E-16	2.20E-16	4.37E-02	1.50E-14	1.07E-14
60	TAC_GTA	GuaM1HDV	7.24	-0.240	-0.286	-0.336	2.20E-16	2.20E-16	2.20E-16	2.05E-14	1.50E-14	1.07E-14

**Tab. 2: Top 60 constructs from the Gua-HDV screen, ranked by their fold change at 300 µM Gua (here, no further cutoffs were applied). GuaM\* constructs were contained in the original publication by Nomura and colleagues.**

**Supplementary Table 3**

Sub-Library	#	Motif	logCPM	log2FC 12.5 µM	log2FC 25 µM	log2FC 50 µM	p value 12.5 µM	p value 25 µM	p value 50 µM	FDR 12.5 µM	FDR 25 µM	FDR 50 µM
2NBN	1	CG_CGG	7,718	-0.808	-0.856	-1,043	1,15E-10	7,88E-12	7,20E-17	5,86E-08	8,07E-09	3,69E-14
	2	GG_CCA	7,933	-0.429	-0.532	-0.916	7,02E-04	2,65E-05	5,81E-13	1,24E-02	5,42E-03	7,44E-11
	3	GG_CCG	7,800	-0.553	-0.580	-0.908	5,05E-07	1,27E-07	1,56E-16	3,77E-05	4,34E-05	3,99E-14
	4	CG_CGC	7,666	-0.650	-0.451	-0.804	6,46E-08	1,50E-04	1,60E-11	8,06E-06	2,20E-02	1,64E-09
	5	CG_CGA	7,299	-0.832	-0.771	-0.755	1,99E-10	2,98E-09	4,19E-09	6,81E-08	1,53E-06	2,68E-07
	6	CC_GGT	7,966	-0.528	-0.243	-0.697	1,57E-06	2,55E-02	1,83E-10	8,92E-05	4,95E-01	1,25E-08
	7	CG_CGT	6,538	-0.614	-0.526	-0.656	3,57E-04	2,05E-03	1,11E-04	7,46E-03	1,34E-01	1,23E-03
	8	GC_GCG	9,099	-0.361	-0.342	-0.624	3,19E-06	9,65E-06	6,33E-16	1,63E-04	2,47E-03	1,30E-13
	9	GA_TCG	8,641	-0.480	-0.276	-0.620	6,61E-07	4,02E-03	1,09E-10	3,98E-05	1,96E-01	8,61E-09
	10	TC_CTG	9,973	-0.103	-0.017	-0.580	1,83E-01	8,28E-01	9,21E-14	4,34E-01	9,68E-01	1,35E-11
3N2N	1	ACG_CG	6,553	-1,380	-1,178	-1,545	6,15E-24	1,54E-18	2,51E-30	6,30E-21	5,27E-16	2,57E-27
	2	CGG_CC	7,354	-0,763	-0,804	-1,389	6,43E-08	1,13E-08	2,52E-22	9,40E-06	1,65E-06	6,45E-20
	3	AAG_CT	7,061	-1,043	-1,139	-1,297	9,00E-16	1,49E-18	9,78E-24	4,61E-13	5,27E-16	3,34E-21
	4	GCG_CTT	7,879	-0,698	-0,747	-1,178	6,25E-10	3,32E-11	3,23E-25	1,72E-07	6,81E-09	1,65E-22
	5	ACC_GG	7,172	-0,790	-0,648	-0,979	1,61E-09	5,91E-07	5,10E-14	3,31E-07	4,32E-05	5,23E-12
	6	GTG_CC	7,381	-0,534	-0,700	-0,973	5,17E-05	1,16E-07	1,85E-13	1,41E-03	1,19E-05	1,72E-11
	7	GCC_GG	7,462	-0,779	-0,822	-0,939	1,44E-08	1,96E-09	6,38E-12	2,46E-06	3,35E-07	5,03E-10
	8	AGC_GC	7,215	-0,621	-0,860	-0,921	2,58E-07	1,19E-12	1,74E-14	2,20E-05	3,05E-10	1,98E-12
	9	CCG_CG	9,043	-0,299	-0,295	-0,902	2,63E-03	2,89E-03	1,85E-19	3,02E-02	7,97E-02	3,80E-17
	10	GCG_GC	7,465	-0,637	-0,538	-0,879	1,54E-05	2,37E-04	2,18E-09	5,43E-04	1,01E-02	9,69E-08
3N3N	1	GCG_CGT	4,243	-0,999	-1,208	-2,150	1,35E-04	4,17E-06	5,19E-15	1,88E-02	5,18E-03	2,13E-11
	2	GTG_TAC	4,744	-0,665	-0,658	-2,017	1,26E-02	1,28E-02	6,51E-13	1,96E-01	4,91E-01	6,67E-10
	3	GAG_CTT	4,885	-0,960	-1,132	-1,579	3,06E-05	9,64E-07	1,35E-11	8,97E-03	2,88E-03	9,21E-09
	4	ACG_CGG	5,579	-1,037	-1,023	-1,557	3,18E-05	3,71E-05	6,32E-10	8,97E-03	1,90E-02	2,27E-07
	5	GGG_CTC	3,384	-0,784	-0,322	-1,544	1,48E-02	2,95E-01	2,36E-06	2,04E-01	9,26E-01	1,90E-04
	6	CCG_TGG	5,987	-0,723	-0,602	-1,369	1,35E-04	1,32E-03	1,03E-12	1,88E-02	1,29E-01	8,47E-10
	7	TGG_CTA	4,911	-1,002	-0,767	-1,343	4,09E-05	1,38E-03	3,61E-08	8,97E-03	1,29E-01	5,48E-06
	8	CGA_TTG	5,561	-0,593	-0,469	-1,292	4,31E-03	2,26E-02	8,95E-10	1,14E-01	5,54E-01	2,63E-07
	9	TGC_GTA	5,699	-0,341	-0,421	-1,275	1,21E-01	5,50E-02	1,49E-08	5,51E-01	7,26E-01	2,77E-06
	10	CTG_CAG	4,401	-0,858	-0,788	-1,271	5,22E-04	1,25E-03	2,91E-07	3,69E-02	1,25E-01	3,41E-05
CG_2N3N	1	CG_CGA	7,013	-0,868	-1,000	-1,658	2,13E-06	4,41E-08	6,76E-19	2,96E-04	9,04E-06	2,31E-16
	2	CG_CGG	8,119	-1,059	-1,279	-1,634	4,22E-14	9,29E-20	1,07E-30	4,32E-11	9,51E-17	1,09E-27
	3	CG_CTG	6,517	-0,416	0,017	-0,993	4,46E-02	9,34E-01	2,06E-06	2,14E-01	9,96E-01	6,20E-05
	4	CG_CGC	7,709	-0,641	-0,568	-0,965	7,93E-06	6,52E-05	1,65E-11	5,07E-04	5,13E-03	1,87E-09
	5	CA_TGG	8,928	-0,693	-0,618	-0,887	3,56E-10	1,83E-08	7,83E-16	1,22E-07	4,68E-06	1,60E-13
	6	GA_TCA	7,259	-0,381	-0,362	-0,811	2,21E-02	2,87E-02	1,16E-06	1,37E-01	4,41E-01	3,83E-05
	7	CG_CGT	7,647	-0,761	-0,526	-0,772	4,25E-06	1,30E-03	2,38E-06	3,35E-04	5,12E-02	6,95E-05
	8	GG_CTG	7,987	-0,522	-0,562	-0,758	1,02E-03	3,92E-04	1,70E-06	1,96E-02	2,36E-02	5,43E-05
	9	TG_CAG	10,113	-0,352	-0,543	-0,749	2,03E-06	2,48E-13	5,86E-24	2,96E-04	1,27E-10	3,00E-21
	10	AG_TGC	10,827	0,006	0,042	-0,708	9,45E-01	6,19E-01	1,76E-16	9,81E-01	9,62E-01	4,50E-14
CG_3N2N	1	GAG_CG	5,244	-0,653	-0,804	-1,593	1,51E-03	8,61E-05	6,56E-14	1,60E-02	3,27E-03	5,17E-12
	2	CGG_CG	7,089	-0,688	-0,799	-1,462	1,95E-06	2,95E-08	3,03E-23	8,70E-05	4,32E-06	7,75E-21
	3	CGG_CC	6,660	-0,812	-0,741	-1,358	6,40E-07	4,23E-06	1,35E-16	3,45E-05	2,40E-04	1,97E-14
	4	GCG_GC	6,370	-1,157	-1,468	-2,245	4,45E-10	3,87E-15	8,83E-12	5,70E-08	2,24E-12	5,65E-10
	5	CAG_CT	8,381	-0,774	-0,709	-1,244	5,79E-12	2,07E-10	2,99E-28	1,97E-09	5,30E-08	1,53E-25
	6	CGT_CG	8,188	-0,500	-0,652	-1,226	1,80E-05	2,17E-08	3,34E-25	6,83E-04	3,71E-06	1,14E-22
	7	CGA_TC	8,501	-0,572	-0,555	-0,963	1,71E-07	3,31E-07	1,33E-18	1,35E-05	2,83E-05	2,72E-16
	8	ACG_CG	7,152	-0,779	-0,580	-0,954	7,54E-08	4,67E-05	2,87E-11	7,02E-06	1,91E-03	1,55E-09
	9	GAA_TC	5,711	-0,590	-0,619	-0,945	1,99E-03	1,03E-03	6,29E-07	1,88E-02	2,93E-02	1,43E-05
	10	CTG_CTC	8,284	-0,460	-0,516	-0,940	6,96E-05	7,43E-06	5,51E-16	1,84E-03	3,62E-04	5,64E-14
CG_3N3N	1	GAC_GGC	4,558	-1,282	-1,874	-2,358	2,98E-04	2,34E-07	2,02E-10	3,49E-02	6,85E-05	5,69E-08
	2	GGT_TCC	3,535	-1,260	-0,577	-2,279	3,65E-03	1,61E-01	5,30E-07	1,13E-01	7,66E-01	4,17E-05
	3	GCG_GGC	5,555	-1,233	-1,342	-2,250	5,87E-05	1,16E-05	1,47E-12	1,34E-02	1,89E-03	6,67E-10
	4	GGG_CAC	4,242	-1,291	-1,254	-2,049	3,21E-04	3,90E-04	2,80E-08	3,55E-02	3,33E-02	4,24E-06
	5	GTC_GCC	4,769	-0,673	-0,657	-2,025	5,48E-02	5,84E-02	3,94E-08	3,49E-01	5,64E-01	5,21E-06
	6	CCG_CTG	4,961	-1,093	-1,067	-1,927	5,66E-04	6,67E-04	2,74E-09	4,73E-02	4,80E-02	5,61E-07
	7	GTC_GTC	3,949	-1,261	-1,318	-1,900	1,54E-03	8,32E-04	2,67E-06	7,83E-02	5,59E-02	1,56E-04
	8	GAG_CGC	4,656	-1,436	-1,386	-1,868	2,48E-04	3,51E-04	2,05E-06	3,28E-02	3,13E-02	1,25E-04
	9	CCG_CAG	7,707	-0,980	-1,165	-1,852	1,06E-08	1,61E-11	4,61E-26	4,36E-05	3,88E-08	1,89E-22
	10	GTG_CTC	4,020	-1,415	-1,162	-1,838	5,63E-04	3,55E-03	7,14E-06	4,73E-02	1,45E-01	3,25E-04
noCG_2N3N	1	CT_GCG	9,823	-0,216	-0,046	-0,730	3,18E-03	5,27E-01	2,85E-23	5,93E-02	9,88E-01	1,46E-20
	2	CT_AGC	10,795	-0,127	-0,084	-0,686	3,88E-02	1,71E-01	8,11E-29	2,78E-01	9,72E-01	8,31E-26
	3	CT_GGT	10,832	-0,200	-0,109	-0,634	1,77E-02	1,97E-01	6,11E-14	1,93E-01	9,77E-01	6,25E-12
	4	TC_GGT	9,970	-0,455	-0,087	-0,614	3,22E-13	1,57E-01	4,85E-23	1,65E-10	9,59E-01	1,65E-20
	5	CC_GGT	9,476	-0,700	-0,363	-0,614	2,31E-15	3,29E-05	2,28E-12	2,37E-12	2,06E-02	1,55E-10
	6	CA_TGC	10,526	-0,034	0,027	-0,596	6,67E-01	7,39E-01	1,06E-13	8,60E-01	9,88E-01	9,89E-12
	7	CT_TGT	9,719	-0,105	-0,158	-0,564	2,24E-01	6,70E-02	6,69E-11	5,71E-01	9,06E-01	3,26E-09
	8	CA_TGT	10,082	-0,137	-0,101	-0,562	1,24E-01	2,57E-01	3,63E-10	4,57E-01	9,88E-01	1,49E-08
	9	AC_AGC	10,895	-0,246	-0,089	-0,556	7,21E-05	1,51E-01	2,83E-19	4,10E-03	9,59E-01	7,25E-17
	10	AC_GGC	11,689	-0,523	-0,130	-0,547	6,77E-13	7,35E-02	5,05E-14	2,31E-10	9,16E-01	5,75E-12
noCG_3N2N	1	AGG_CC	7,795	-0,647	-0,729	-0,968	1,97E-08	2,29E-10	3,31E-17	1,91E-05	2,35E-07	3,38E-14
	2	AGC_GC	8,031	-0,543	-0,467	-0,649	6,14E-07	1,59E-05	1,70E-09	9,09E-05	8,14E-03	1,25E-07
	3	CGC_GC	9,553	-0,268	-0,158	-0,621	1,46E-03	6,05E-02	1,64E-13</			

## Supplementary Table 4

## Gua-HDV ribozyme library and hit validation

Name	Sequence	Comment
Gua-HDV ribozyme library fwd	ttttacccggccaccgttaatqtcgcactaNNNcgaaatgggacgcacaatct	Insertion of guanine xpt aptamer into HDV ribozyme with randomized communication module
Gua-HDV ribozyme library rev	cttgctgcgcataccacggcattataNNNtgcggccaggccgg	Insertion of HDV ribozyme
HDV act fwd	gaggggaccgcctccctcgtaatggcgcacaaatctctcgaaacaaaaaaaggctgttggaaatttgttat	
HDV act rev	ggaaatgttgcggccaggccggccaggaggctgggacatqccggccctttgtttggggactggaaatcag	
HDV inact fwd	tgaatggacgcacaaaatctctcg	Inactivation of HDV ribozyme
HDV inact rev	ccataccgtggggacgtt	
GuaM8HDV fwd	ttttacccggccaccgttaatqtcgcactaGTAGcgaatggggacgcacaatct	Insertion of guanine xpt aptamer into HDV ribozyme with GuaM8HDV communication module
GuaM8HDV rev	cttgctgcgcataccacggcattataCATtgcggccaggccgg	Insertion of guanine xpt aptamer into HDV ribozyme with GuaM8HDV communication module
GuaB1HDV fwd	ttttacccggccaccgttaatqtcgcactaTAgcgaatggggacgcacaatct	Insertion of guanine xpt aptamer into HDV ribozyme with GuaB1HDV communication module
GuaB1HDV rev	cttgctgcgcataccacggcattataCATtgcggccaggccgg	Insertion of guanine xpt aptamer into HDV ribozyme with GuaB1HDV communication module
GuaB2HDV fwd	ttttacccggccaccgttaatqtcgcactaTAgcgaatggggacgcacaatct	Insertion of guanine xpt aptamer into HDV ribozyme with GuaB2HDV communication module
GuaB2HDV rev	cttgctgcgcataccacggcattataCATtgcggccaggccgg	Insertion of guanine xpt aptamer into HDV ribozyme with GuaB2HDV communication module
GuaB3HDV fwd	ttttacccggccaccgttaatqtcgcactaTAgcgaatggggacgcacaatct	Insertion of guanine xpt aptamer into HDV ribozyme with GuaB3HDV communication module
GuaB3HDV rev	cttgctgcgcataccacggcattataCGGtgcggccaggccgg	Insertion of guanine xpt aptamer into HDV ribozyme with GuaB3HDV communication module
GuaB4HDV fwd	ttttacccggccaccgttaatqtcgcactaTAgcgaatggggacgcacaatct	Insertion of guanine xpt aptamer into HDV ribozyme with GuaB4HDV communication module
GuaB4HDV rev	cttgctgcgcataccacggcattataCGGtgcggccaggccgg	Insertion of guanine xpt aptamer into HDV ribozyme with GuaB4HDV communication module
GuaM12HDV fwd	ttttacccggccaccgttaatqtcgcactaATAgcgaatggggacgcacaatct	Insertion of guanine xpt aptamer into HDV ribozyme with GuaM12HDV communication module
GuaM12HDV rev	cttgctgcgcataccacggcattataATtgcggccaggccgg	Insertion of guanine xpt aptamer into HDV ribozyme with GuaM12HDV communication module
GuaB5HDV fwd	ttttacccggccaccgttaatqtcgcactaCGGcgaatggggacgcacaatct	Insertion of guanine xpt aptamer into HDV ribozyme with GuaB5HDV communication module
GuaB5HDV rev	cttgctgcgcataccacggcattataCGGtgcggccaggccgg	Insertion of guanine xpt aptamer into HDV ribozyme with GuaB5HDV communication module
GuaB6HDV fwd	ttttacccggccaccgttaatqtcgcactaCGGcgaatggggacgcacaatct	Insertion of guanine xpt aptamer into HDV ribozyme with GuaB6HDV communication module
GuaB6HDV rev	cttgctgcgcataccacggcattataCGGtgcggccaggccgg	Insertion of guanine xpt aptamer into HDV ribozyme with GuaB6HDV communication module
GuaB7HDV fwd	ttttacccggccaccgttaatqtcgcactaCGGcgaatggggacgcacaatct	Insertion of guanine xpt aptamer into HDV ribozyme with GuaB7HDV communication module
GuaB7HDV rev	cttgctgcgcataccacggcattataCGGtgcggccaggccgg	Insertion of guanine xpt aptamer into HDV ribozyme with GuaB7HDV communication module
GuaB8HDV fwd	ttttacccggccaccgttaatqtcgcactaTAgcgaatggggacgcacaatct	Insertion of guanine xpt aptamer into HDV ribozyme with GuaB8HDV communication module
GuaB8HDV rev	cttgctgcgcataccacggcattataTAgcgaatggggacgcacaatct	Insertion of guanine xpt aptamer into HDV ribozyme with GuaB8HDV communication module
GuaB9HDV fwd	ttttacccggccaccgttaatqtcgcactaTAgcgaatggggacgcacaatct	Insertion of guanine xpt aptamer into HDV ribozyme with GuaB9HDV communication module
GuaB9HDV rev	cttgctgcgcataccacggcattataTAgcgaatggggacgcacaatct	Insertion of guanine xpt aptamer into HDV ribozyme with GuaB9HDV communication module
GuaB10HDV fwd	ttttacccggccaccgttaatqtcgcactaTAgcgaatggggacgcacaatct	Insertion of guanine xpt aptamer into HDV ribozyme with GuaB10HDV communication module
Ampl fwd	agataaaatggtttaccatcgacgttat	Insertion of the constructs into CMV-EGFP-pA expression plasmid backbone using HindIII and BglII restriction sites
Ampl rev	agataaaatgttaccatcgacgttat	

## Gua-HHR ribozyme library and hit validation

Name	Sequence	Comment
Gu-aHHR ribozyme library fwd	ttttacccggccaccgttaatgtccgactaNNTatcatccgcgtatgtqagtcccaaataagg	Insertion of guanine xpt aptamer into HHR ribozyme with randomized communication module
Gu-aHHR ribozyme library rev	cttgcggtccatataccacggcattataNNNNGaaatccaggacggccgttttgc	
Hh act fwd	catgtatgtggatccaaataatggacgaaaacccgcgtcaaaacaacaaagccctgtgttataatgttt	Insertion of HHR ribozyme
Hh act rev	gtatgtacccgtggaaatccaggacgcgcctttgtttgttggactggaaatcg	
Hh inact fwd	gaccgcgtcaaaaaacaaaa	Inactivation of HHR ribozyme
Hh inact rev	tgcgttatattggacte	
Guab1HHR fwd	tttttacccggccaccgttaatgtccgactaTGTatcatccgcgtatgtqagtcccaaataagg	Insertion of guanine xpt aptamer into HHR ribozyme with Guab1HHR communication module
Guab1HHR rev	cttgcggtccatataccacggcattataATGGaaatccaggacggccgttttgc	
Guab2HHR fwd	tttttacccggccaccgttaatgtccgactaGtacatccgcgtatgtqagtcccaaataagg	Insertion of guanine xpt aptamer into HHR ribozyme with Guab2HHR communication module
Guab2HHR rev	cttgcggtccatataccacggcattataATGGaaatccaggacggccgttttgc	
Guab3HHR fwd	tttttacccggccaccgttaatgtccgactaTtacatccgcgtatgtqagtcccaaataagg	Insertion of guanine xpt aptamer into HHR ribozyme with Guab3HHR communication module
Guab3HHR rev	cttgcggtccatataccacggcattataATGCaaatccaggacggccgttttgc	
Guab4HHR fwd	tttttacccggccaccgttaatgtccgactaGtacatccgcgtatgtqagtcccaaataagg	Insertion of guanine xpt aptamer into HHR ribozyme with Guab4HHR communication module
Guab4HHR rev	cttgcggtccatataccacggcattataACTCAaaatccaggacggccgttttgc	
Guab5HHR fwd	tttttacccggccaccgttaatgtccgactaTtacatccgcgtatgtqagtcccaaataagg	Insertion of guanine xpt aptamer into HHR ribozyme with Guab5HHR communication module
Guab5HHR rev	cttgcggtccatataccacggcattataGCTGaaatccaggacggccgttttgc	
Guab6HHR fwd	tttttacccggccaccgttaatgtccgactaGtacatccgcgtatgtqagtcccaaataagg	Insertion of guanine xpt aptamer into HHR ribozyme with Guab6HHR communication module
Guab6HHR rev	cttgcggtccatataccacggcattataATGGaaatccaggacggccgttttgc	
Guab7HHR fwd	tttttacccggccaccgttaatgtccgactaGtacatccgcgtatgtqagtcccaaataagg	Insertion of guanine xpt aptamer into HHR ribozyme with Guab7HHR communication module
Guab7HHR rev	cttgcggtccatataccacggcattataATGCaaatccaggacggccgttttgc	
Ampl fwd	atgtaaaatgttgtaccatcgactgttt	Insertion of the constructs into CMV-pGK-pAV expression plasmid backbone using HindIII and BglII restriction sites
Ampl rev	atgtaaaatgttgtaccatcgactgttt	

TetTwister Hit validation

Name	Sequence	Comment
Tw act fwd	agatactcgaccaaaacactcccttaaagcggttacaad	Amplification of Twister ribozyme and insertion into psICHCK-2 plasmid using Xhol and NotI restriction sites
Tw act rev	agatagccgcgttttttttcgtttaaaagcgccattaaatg	
Tw inact fwd	agatactcgaccaaaaacactcccttaaagcggttacaad	Amplification of inactive Twister ribozyme and insertion into psICHCK-2 plasmid using Xhol and NotI restriction sites
Tw inact rev	agatagccgcgttttttcgtttaaaagcgccattaaatg	
T1 fwd	aggctcgaaaaaggcttacaaaacatacCGGtaatgcgcgtttaaagaaaaaaaagc	Insertion of tetracycline aptamer into Twister ribozyme with T1 communication module
T1 rev	agtgttgtatcttcacccctccGttactcgcttattttcgcggcttgc	
T2 fwd	aggctcqaaaaaggcttacaaaacatacCttaaatgcgcgtttaaagaaaaaaaagc	Insertion of tetracycline aptamer into Twister ribozyme with T2 communication module
T2 rev	agtgttgtatcttcacccctccGttactcgcttattttcgcggcttgc	
T3 fwd	aggctcqaaaaaggcttacaaaacatacCGttaaatgcgcgtttaaagaaaaaaaagc	Insertion of tetracycline aptamer into Twister ribozyme with T3 communication module
T3 rev	agtgttgtatcttcacccctccGttactcgcttattttcgcggcttgc	
T4 fwd	aggctcqaaaaaggcttacaaaacatacCttaaatgcgcgtttaaagaaaaaaaagc	Insertion of tetracycline aptamer into Twister ribozyme with T4 communication module
T4 rev	agtgttgtatcttcacccctccGttactcgcttattttcgcggcttgc	
T5 fwd	aggctcqaaaaaggcttacaaaacatacCGGttaaatgcgcgtttaaagaaaaaaaagc	Insertion of tetracycline aptamer into Twister ribozyme with T5 communication module
T5 rev	agtgttgtatcttcacccctccGttactcgcttattttcgcggcttgc	
T6 fwd	aggctcqaaaaaggcttacaaaacatacCGGttaaatgcgcgtttaaagaaaaaaaagc	Insertion of tetracycline aptamer into Twister ribozyme with T6 communication module
T6 rev	agtgttgtatcttcacccctccGttactcgcttattttcgcggcttgc	
T7 fwd	aggctcqaaaaaggcttacaaaacatacCTttaaatgcgcgtttaaagaaaaaaaagc	Insertion of tetracycline aptamer into Twister ribozyme with T7 communication module
T7 rev	agtgttgtatcttcacccctccGttactcgcttattttcgcggcttgc	
T8 fwd	aggctcqaaaaaggcttacaaaacatacCGttaaatgcgcgtttaaagaaaaaaaagc	Insertion of tetracycline aptamer into Twister ribozyme with T8 communication module
T8 rev	agtgttgtatcttcacccctccGttactcgcttattttcgcggcttgc	
T9 fwd	aggctcqaaaaaggcttacaaaacatacCGttaaatgcgcgtttaaagaaaaaaaagc	Insertion of tetracycline aptamer into Twister ribozyme with T9 communication module
T9 rev	agtgttgtatcttcacccctccGttactcgcttattttcgcggcttgc	
T10 fwd	aggctcqaaaaaggcttacaaaacatacCGGttaaatgcgcgtttaaagaaaaaaaagc	Insertion of tetracycline aptamer into Twister ribozyme with T10 communication module
T10 rev	agtgttgtatcttcacccctccGttactcgcttattttcgcggcttgc	
Neg. control #500 fwd	aggctcqaaaaaggcttacaaaacatacCGttaaatgcgcgtttaaagaaaaaaaagc	Insertion of tetracycline aptamer into Twister ribozyme with communication module of neg. control #500
Neg. control #500 rev	agtgttgtatcttcacccctccGttactcgcttattttcgcggcttgc	
Neg. control #1000 fwd	aggctcqaaaaaggcttacaaaacatacCGGttaaatgcgcgtttaaagaaaaaaaagc	Insertion of tetracycline aptamer into Twister ribozyme with communication module of neg. control #1000
Neg. control #1000 rev	agtgttgtatcttcacccctccGttactcgcttattttcgcggcttgc	
Neg. control #2000 fwd	aggctcqaaaaaggcttacaaaacatacCGttaaatgcgcgtttaaagaaaaaaaagc	Insertion of tetracycline aptamer into Twister ribozyme with communication module of neg. control #2000
Neg. control #2000 rev	agtgttgtatcttcacccctccGttactcgcttattttcgcggcttgc	
Neg. control #3000 fwd	aggctcqaaaaaggcttacaaaacatacCGGttaaatgcgcgtttaaagaaaaaaaagc	Insertion of tetracycline aptamer into Twister ribozyme with communication module of neg. control #3000
Neg. control #3000 rev	agtgttgtatcttcacccctccGttactcgcttattttcgcggcttgc	

**Tab. 4: Cloning primers used in this study.**

## Supplementary Table 4 - continued

### Gu-U1 library and hit validation

Name	Sequence	Comment
Gu-U1 library fwd	tttctaccggcaccgttaaatgtccgactaNNNacttacctgcaaaacaaaacaagcctggtaaatttttat	Insertion of guanine xpt aptamer onto U1 snRNP binding site with randomized communication module
Gu-U1 library rev	cttgcgtggccatataccacccgattataANNNacttacctgttttgttggacttggaaatcacg	
U1 bs fwd	caaaaaaaaatccacccgattata	Insertion of U1 snRNP binding site
U1 bs rev	acttaccgttttgttggacttggaaatcacg	
Guau1B1 fwd	tttctaccggcaccgttaaatgtccgactaGGCacttacctgtttttttggacttggaaatttttat	Insertion of guanine xpt aptamer onto U1 snRNP binding site with Guau1B1 communication module
Guau1B1 rev	cttgcgtggccatataccacccgattataGCacttacctgtttttttggacttggaaatcacg	
Guau1B2 fwd	tttctaccggcaccgttaaatgtccgactaGGCacttacctgtttttttggacttggaaatttttat	Insertion of guanine xpt aptamer onto U1 snRNP binding site with Guau1B2 communication module
Guau1B2 rev	cttgcgtggccatataccacccgattataGTGacttacctgtttttttggacttggaaatcacg	
Guau1B3 fwd	tttctaccggcaccgttaaatgtccgactaAGTacttacctgtttttttggacttggaaatttttat	Insertion of guanine xpt aptamer onto U1 snRNP binding site with Guau1B3 communication module
Guau1B3 rev	cttgcgtggccatataccacccgattataAGCacttacctgtttttttggacttggaaatcacg	
Guau1B4 fwd	tttctaccggcaccgttaaatgtccgactaAGGacttacctgtttttttggacttggaaatttttat	Insertion of guanine xpt aptamer onto U1 snRNP binding site with Guau1B4 communication module
Guau1B4 rev	cttgcgtggccatataccacccgattataAGAacttacctgtttttttggacttggaaatcacg	
Guau1B5 fwd	tttctaccggcaccgttaaatgtccgactaGTTacttacctgtttttttggacttggaaatttttat	Insertion of guanine xpt aptamer onto U1 snRNP binding site with Guau1B5 communication module
Guau1B5 rev	cttgcgtggccatataccacccgattataGTGacttacctgtttttttggacttggaaatcacg	
Guau1B6 fwd	tttctaccggcaccgttaaatgtccgactaTGGacttacctgtttttttggacttggaaatttttat	Insertion of guanine xpt aptamer onto U1 snRNP binding site with Guau1B6 communication module
Guau1B6 rev	cttgcgtggccatataccacccgattataTCGacttacctgtttttttggacttggaaatcacg	
Guau1B7 fwd	tttctaccggcaccgttaaatgtccgactaAGCacttacctgtttttttggacttggaaatttttat	Insertion of guanine xpt aptamer onto U1 snRNP binding site with Guau1B7 communication module
Guau1B7 rev	cttgcgtggccatataccacccgattataACGacttacctgtttttttggacttggaaatcacg	
Guau1B8 fwd	tttctaccggcaccgttaaatgtccgactaACTacttacctgtttttttggacttggaaatttttat	Insertion of guanine xpt aptamer onto U1 snRNP binding site with Guau1B8 communication module
Guau1B8 rev	cttgcgtggccatataccacccgattataACGacttacctgtttttttggacttggaaatcacg	
Guau1B9 fwd	tttctaccggcaccgttaaatgtccgactaATTacttacctgtttttttggacttggaaatttttat	Insertion of guanine xpt aptamer onto U1 snRNP binding site with Guau1B9 communication module
Guau1B9 rev	cttgcgtggccatataccacccgattataATGacttacctgtttttttggacttggaaatcacg	
Guau1B10 fwd	tttctaccggcaccgttaaatgtccgactaACacttacctgtttttttggacttggaaatttttat	Insertion of guanine xpt aptamer onto U1 snRNP binding site with Guau1B10 communication module
Guau1B10 rev	cttgcgtggccatataccacccgattataTGCActtacctgtttttttggacttggaaatcacg	
AmpI fwd	adataaaagtgttggatccccatgtcgcttatt	Insertion of the constructs into CMV-eGFP pAAV expression plasmid backbone using HindIII and BglII restriction sites
AmpI rev	adataaaadatctggatcacatccaccaggc	

**Supplementary Table 5**

TwisterTet_3N2N	AGTAA([ACGT]{3})GAGAGG.{37}ACATAC([ACGT]{2})TTAAT
TwisterTet_2N3N	AGTAA([ACGT]{2})GAGAGG.{37}ACATAC([ACGT]{3})TTAAT
TwisterTet_3N3N	AGTAA([ACGT]{3})GAGAGG.{37}ACATAC([ACGT]{3})TTAAT
TwisterTet(CG)_3N2N	AGTAAC([ACGT]{3})GAGAGG.{37}ACATAC([ACGT]{2})GTTAAT
TwisterTet(CG)_2N3N	AGTAAC([ACGT]{2})GAGAGG.{37}ACATAC([ACGT]{3})GTTAAT
TwisterTet(CG)_3N3N	AGTAAC([ACGT]{3})GAGAGG.{37}ACATAC([ACGT]{3})GTTAAT
TwisterTet_noCG_3N2N	AGTAA([ACGT]{3})AGAGG.{37}ACATA([ACGT]{2})TTAAT
TwisterTet_noCG_2N3N	AGTAA([ACGT]{2})AGAGG.{37}ACATA([ACGT]{3})TTAAT
TwisterTet_noCG_3N3N	AGTAA([ACGT]{3})AGAGG.{37}ACATA([ACGT]{3})TTAAT
Spikein	TTAGAACAG

**Tab. 5: Regular expressions used in the variant extraction.**