## **Supplemental Methods**

*Subjects.* Bangladeshi infants were recruited from the urban slum areas of Kamrangir Char and Hazaribagh in Dhaka; study procedures were approved by the Institutional Review Board of the International Centre for Diarrhoeal Disease Research, Bangladesh (icddr,b). In Guatemala, subjects were recruited from several urban centers (Santa Catarina Pinula and La Comunidad, Mixco) and from 2 semi-urban communities (San Miguel Dueñas and Santa María de Jesus); procedures were approved by the Institutional Ethics Committee of INCAP. Filipino children were recruited from low-income neighborhoods in Mandaluyong City within Metro-Manila; the study protocol was approved by the Institutional Review Board at the University of California, Davis and by the Research Ethics Board of the University of the Philippines, Manila. All studies were also approved by the Research Ethics Board of Newcastle University.

Assessment of vitamin A intake. Vitamin A intake was estimated for the Bangladeshi children based on measured breastmilk vitamin A concentrations and published estimates of breastmilk intake (Supplemental References 1.2) and by doing 24 h dietary recalls on 2 separate days and a 30 d food frequency questionnaire. Estimates for dietary intake were determined using a food composition table from the Institute of Nutrition and Food Science at the University of Dhaka and included both preformed vitamin A and provitamin A carotenoids [as RAE based on the IOM conversion factors (Supplemental Reference 3)]. For the Guatemalan children, dietary vitamin A intake was assessed by using a 30 d food frequency questionnaire as well as 24 h recalls done on 3 separate days during the study; the latter included weighing mother-reported portions of consumed foods. Estimated intake, based on the INCAP Food Composition Table, included both preformed vitamin A and provitamin A carotenoids [as RAE using the IOM conversion factors (Supplemental Reference 3)]. For the Filipino children, dietary vitamin A intake was assessed by conducting 24 h recalls on 4 – 5 separate days during the study, a 12 h in-home observation to weigh and record the child's food intake (with 12 h recall to complete the 24 h period), and a 30 d questionnaire to assess use of vitamin A-containing supplements. For breastfeeding children, breast milk intake was measured using the <sup>2</sup>H<sub>2</sub>O dose-to-mother method (Supplemental References 4,5). Results were combined to estimate usual vitamin A intake using the US National Cancer Institute method (Supplemental Reference 6); the estimate included preformed vitamin A and provitamin A carotenoids [as RAE based on the IOM conversion factors (Supplemental Reference 3)].

*Calculation of vitamin A intake for modeling.* Because assessed vitamin A intake included both preformed and provitamin A, we adjusted the value for provitamin A carotenoids (as RAE) to the equivalent amount of preformed vitamin A. That is, because the conversion of provitamin A carotenoids to RAE [using the IOM conversion factors (Supplemental Reference 3)] includes absorption and conversion efficiency, and we assumed an absorption efficiency of 80% for preformed vitamin A for modeling, the value for vitamin A intake used as weighted input in the model was calculated as: preformed vitamin A + (provitamin A / 0.8). More specifically, for the Bangladeshi and Guatemalan children, the individual assessed preformed and provitamin A intakes were used in the above calculation; the geometric mean of the individual adjusted estimates was used in the model. For the Filipino children, the above calculation. Preformed vitamin A contributed, on average, >95% of total vitamin A intake for the Bangladeshi subjects, >90% for the Filipinos, and ~76% for the Guatemalan children.

Retinol analyses. Retinol was extracted from plasma using a modification of the method of Aebischer et al. (16). Briefly, 400  $\mu$ L of plasma was diluted with 400  $\mu$ L diH<sub>2</sub>0 before addition of 800 µL ethanol containing 50 pmol [<sup>12</sup>C]retinyl acetate as internal standard. The denatured sample was extracted twice with 2 mL of hexane and hexane in the combined extracts was evaporated under N<sub>2</sub>. The residue was resuspended in 100  $\mu$ L of ethanol and 10  $\mu$ L was used for LC-MS/MS analysis. Liquid chromatographic conditions were adapted from Kane and Napoli (17) to a Shimadzu Prominence HPLC equipped with a 100 mm x 2.1 mm 3 µm Supelcosil ABZ PLUS column (Supelco) and a 4 x 2 mm SecurityGuard (Phenomenex) C<sub>18</sub> cartridge maintained at 25°C. The binary mobile phase system consisted of 0.1% formic acid aqueous (A) and 0.1% formic acid in acetonitrile (B); separation of analytes was done at a flow rate of 400 µL/min and a linear gradient of 60% B to 95% B from 0.0 – 6.0 min; 100% B from 6.1 – 13.0 min; 60% B from 13.0 – 14.0 min; and then 60% B from 14.0 – 17.0 min to re-equilibrate the column. The HPLC was coupled to an API 4000 MS/MS system (AB SCIEX) with atmospheric pressure chemical ionization in positive ion mode. Optimized atmospheric pressure chemical ionization source parameters were: CUR 20 psig: GS1 40 psig: GS2 40 psig: CAD 6 psig: TEM 300°C: NC 5 µA. Selected reaction monitoring and quantitation of [<sup>12</sup>C]retinol, [<sup>12</sup>C]retinyl acetate, and  $[^{13}C_{10}]$  retinol was done as previously described (18).

*Plasma volume estimation.* Calculation of fraction of dose in plasma required an estimate of plasma volume for each child. For this work, we estimated plasma volume using the regression equation of Linderkamp et al. (19). Specifically, blood volume (L) was estimated based on body weight (kg) and height (cm); then, hemoglobin concentration was used to estimate hematocrit as hemoglobin × 3 (Supplemental Reference 7), and plasma volume was estimated as blood volume × (1 – hematocrit).

Rationale for updated hypotheses in the compartmental model. The model shown in Figure 1 was adapted from a previously-published model for vitamin A kinetics in children (8). Specifically, in addition to irreversible loss from the larger storage compartment 6, the current model includes an additional site of loss from extrahepatic tissues that do not recycle retinol to plasma (component 8). The hypothesis that 2 tissue sites of irreversible loss are required was supported by modeling tracer data for plasma and loss in rats [unpublished observations, MH Green, based on data presented in (Supplemental Reference 8)] in which, to adequately fit the loss data, the model required output from both an exchangeable and a nonexchangeable extravascular compartment (analogous to compartment 6 and component 8, respectively). In that analysis, a delay of 75 min was required to fit the loss data. Thus, we fixed the delay time in component 8 [DT(8)] at 75 min for all 3 groups in the current analysis. Note that the value assigned to DT(8) does not influence model results and was chosen to provide a more physiologically-sound model. Since further testing using theoretical data (unpublished observations, JL Ford and MH Green) indicated that, without loss data, one cannot accurately distinguish the proportion of output from these 2 sites, for the current work, we partitioned the rate of loss from tissues so that 50% of the output (i.e., disposal rate) was from tissue component 8 and 50% was from storage compartment 6.

## **Supplemental Discussion**

*Future recommendations for the super-child approach.* Based on the current analysis, the following revised design is recommended for future super-child studies (**Supplemental Figure 1**). We recommend that there be a sample size of at least 60 children per group and that 2 blood samples be collected from each child at times (n=13) from 5 h to 42 d. Based on sensitivity analysis performed using WinSAAM (Supplemental Reference 9), the recommended sampling times are 5, 8, and 12 h, and 1, 2, 4, 7, 10, 14, 21, 28, 35, and 42 d. All children should be sampled at 7 d (see below) and at one additional time, randomly assigned at recruitment, such that 5 children will be sampled at each of the 12, non-7 d times.

Accuracy related to administration of the labeled oral dose is crucial. That is, the time and volume or, preferably, the weight of the dose administered needs to be precisely recorded at the time of dosing (d 0). The amount of label administered to each child must be adequate to ensure accurate detection 42 d later, based on detection capability of the equipment that will be used for analysis.

For the current modeling analysis, absorption efficiency for preformed vitamin A was assumed to be 80% (24,27). To improve the accuracy of predictions by the model, investigators should consider actually determining absorption of vitamin A. For example, the dual isotope absorption method (Supplemental Reference 10) can be used to measure vitamin A absorption by administering an oral dose of labeled vitamin A followed ~3 h later by an intravenous dose of a differently-labeled vitamin A in a physiological lipid emulsion; absorption can be calculated as the ratio of fraction of dose of orally ingested vitamin A / intravenously injected vitamin A at a time after the ratio in plasma has stabilized (e.g., 14 d in rats).

In addition to predicting a group mean value for TBS from modeling the composite dataset, data from the plasma samples collected on d 7 can be used in an RID equation to estimate TBS in individual children. Since in more recent work (15), we found that 7 d was a better time than 4 d for doing RID in children, we recommend the former time (7 d) for future studies. Specifically, the super-child model can be used to calculate time-variant values for the equation coefficients (*Fa* and *S*) to be applied in the equation along with each child's retinol specific activity in plasma (SA<sub>p</sub>) at the corresponding time (d 7). Note that values for the coefficients can be calculated at all times and values from 4 to 42 d can be used in the equation to predict TBS for individuals sampled at those times. It is important to mention that, while we hypothesize that 7 d will be more accurate than 4 d for estimating the absolute value for TBS in individual children by RID when a group model-predicted value for *FaS* is used, TBS predictions for the group should be approximately the same at both times. In addition, use of 4 d data can still be valuable for ranking subjects within a group from low to high TBS and estimating the distribution and range of TBS for the group.

	Bangladesh	Philippines	Guatemala
Parameter			
DT(3), d	0.240	0.207	0.191
L(4,3), d <sup>-1</sup>	0.8	0.8	0.8
L(0,3), d <sup>-1</sup>	0.2	0.2	0.2
L(5,4), d <sup>-1</sup>	6.37	1.17	1.17
L(7,5), d <sup>-1</sup>	42.8	2.74	2.47
L(5,7), d <sup>-1</sup>	3.31	0.270	0.316
L(6,5), d <sup>-1</sup>	12.8	11.5	12.0
L(5,6), d <sup>-1</sup>	0.0227	0.00806	0.00856
L(10,6), d <sup>-1</sup>	0.00248	0.00194	0.00106
L(8,5), d <sup>-1</sup>	1.30	2.21	1.34
DT(8), d	0.052	0.052	0.052
L(10,8), d <sup>-1</sup>	1	1	1
Compartment masses			
M(4), µmol	0.154	1.74	1.94
M(5), µmol	0.382	0.462	0.848
M(6), µmol	194	530	1061
M(7), µmol	4.94	4.71	6.63
TBS, µmol	199	535	1068

**Supplemental Table 1** Group-independent model-derived retinol kinetic parameters and steady state compartment masses for 3 groups of young children<sup>1</sup>

<sup>1</sup>Values are model-predicted parameters and steady state compartment masses calculated by the model when applied independently to geometric mean datasets for Bangladeshi, Filipino, and Guatemalan children. Parameters include fractional transfer coefficients [L(I,J)s, or the fraction of retinol in compartment J transferred to compartment I each day] and delay times [DT(I)s, or delay time spent in compartment I]. Compartment masses [M(I), or µmol of vitamin A in compartment I] were calculated using the steady state model solution in WinSAAM. See Supplemental Table 2 for statistical uncertainties for these parameters. The model is shown in Figure 1. TBS, total body stores.

	Banglad	esh	Philippir	nes	Guatemala			
Parameter	Independent	Parallel	Independent	Parallel	Independent	Parallel		
DT(3)	0.033	0.013	0.019	0.013	0.031	0.013		
L(5,4)	0.36	0.12	0.048	0.039	0.057	0.039		
L(7,5)	0.30	0.27	0.13	0.10	0.18	0.10		
L(5,7)	0.26	0.20	0.14	0.094	0.17	0.094		
L(6,5)	0.027	0.015	0.028	0.015	0.027	0.015		
L(5,6)	0.035	0.030	0.057	0.032	0.051	0.032		
L(10,6)	0.12	0.12	0.13	0.13	0.12	0.12		

**Supplemental Table 2** Fractional standard deviations for group-independent versus partially-parallel model-derived kinetic parameters for 3 groups of young children<sup>1</sup>

<sup>1</sup>Shown are statistical uncertainties (fractional standard deviations) for adjustable kinetic parameters when super-child datasets for the 3 groups of young children were fit independently versus using a partially-parallel model. Parameters are delay times [DT(I)s, or delay time spent in compartment I] and fractional transfer coefficients [L(I,J)s, or the fraction of retinol in compartment J transferred to compartment I each day]. The model is shown in Figure 1.

Time (d)	Bangladesh	Philippines	Guatemala			
4	1.58	2.44	2.93			
5	1.04	1.67	2.03			
6	0.788	1.29	1.58			
7	0.673	1.06	1.31			
8	0.618	0.909	1.12			
9	0.592	0.796	0.984			
10	0.578	0.712	0.882			
11	0.571	0.649	0.806			
12	0.566	0.601	0.748			
13	0.562	0.564	0.704			
14	0.559	0.536	0.670			
15	0.556	0.515	0.644			
16	0.554	0.498	0.624			
17	0.551	0.485	0.609			
18	0.549	0.475	0.597			
19	0.546	0.467	0.588			
20	0.544	0.461	0.581			
21	0.542	0.456	0.575			
22	0.539	0.452	0.570			
23	0.537	0.448	0.567			
24	0.534	0.445	0.564			
25	0.532	0.442	0.561			
26	0.530	0.440	0.559			
27	0.527	0.438	0.557			
28	0.525	0.436	0.555			

**Supplemental Table 3** Population-specific values for the composite coefficient (*FaS*) at various times for use in a retinol isotope dilution equation<sup>1</sup>

<sup>1</sup>Shown are model-calculated values for the time-variant composite coefficient *FaS* that can be applied in a retinol isotope diluatio equation such as Equation 1 to estimate TBS for individual children in each group. Values are calculated as  $[F(6)_t + F(7)_t] \times \{F(5)_t/M(5) / [F(6)_t + F(7)_t / M(6)+M(7)]\}$ , where F(I) is the fraction of tracer in compartment I as a function of time and M(I) is the mass of vitamin A in compartment I. The model is shown in Figure 1.

[ <sup>13</sup> C]Retin	iyl ace 子	etate	Retinol isotope dilution										
Child ID	5 h	8 h	12 h	1 d	2 d	4 d	7 d	10 d	14 d	21 d	28 d	35 d	42 d
1-5	✓						✓						
6-10		✓					✓						
11-15			~				~						
16-20				~			~						
21-25					✓		✓						
26-30						✓	✓						
31-35							✓	✓					
36-40							✓		✓				
41-45							✓			✓			
46-50							✓				✓		
51-55							1					✓	
56-60							1						✓
# sampled	5	5	5	5	5	5	60	5	5	5	5	5	5

**Supplemental Figure 1.** Recommended design for future super-child studies of vitamin A kinetics and status. The labeled dose is administered on d 0 and data from d 7 are used to calculated vitamin A total body stores using a retinol isotope dilution equation. See Supplemental Discussion for further details about the recommended design.

### Supplemental WinSAAM Deck

P-B-G GM ALL U(3) PARTIALLY-PARALLEL [29-JUN-19] A SAAM31 CC GEOMETRIC MEAN PLASMA FRACTION OF DOSE VERSUS TIME FOR 3 GROUPS CC INCLUDING BANGLADESHI, FILIPINO AND GUATEMALAN CHILDREN CC COMPOSITE DATASETS COMBINED AND FIT USING PARTIALLY-PARALLEL MODEL CC 6-COMPONENT MODEL FOR VITAMIN A KINETICS IN CHILDREN CC DIETARY INTAKE [U(3)] USED AS WEIGHTED INPUT CC FOR FILIPINO AND GUATEMALAN GROUPS: VALUES FOR ALL ADJUSTABLE CC PARAMETERS SET EQUAL EXCEPT THAT L(10,6) INDEPENDENT AND ADJUSTABLE CC FOR BANGLADESHI GROUP: VALUES FOR L(5,4), L(7,5), L(5,7), L(5,6), L(10,6) CC WERE INDEPENDENT AND ADJUSTABLE CC L(8,5) CALCULATED FOR EACH GROUP AS [(DIETARY INTAKE\*0.8)/2]/M(5) AND FIXED CC PHILIPPINES (N=120) CC PARAMETER H PAR CC VALUE LOWER LIMIT UPPER LIMIT CC IC(I)=INITIAL CONDITION (FRACTION OF DOSE) IN COMPARTMENT I AT TIME 0 IC(3) = 1CC DN(I)=NUMBER OF ELEMENTS IN DELAY COMPONENT I CC DT(I)=DELAY TIME IN COMPONENT I (DAY) DT(3) 2.057179E-01 0.000000E+00 1.000000E+02 DN(3) 8 CC L(I,J)=FRACTION OF J TRANSFERRED TO I PER DAY (DAY^-1) CC ASSUMING 80% ABSORPTION EFFICIENCY L(4,3) 0.8 CC L(0,3)=FRACTIONAL LOSS OF UNABSORBED TRACER L(0,3) 0.2 L(5,4) 1.223622E+00 0.000000E+00 1.000000E+02 3.071890E+00 0.000000E+00 1.000000E+02 L(7,5) 3.405374E-01 0.000000E+00 1.000000E+02 L(5,7)L(6,5) 1.231655E+01 0.000000E+00 1.000000E+02 L(5,6) 8.835580E-03 0.000000E+00 1.000000E+02 CC L(10,6) = FRACTIONAL CATABOLIC RATE OF COMPARTMENT 6; CC COMPARTMENT 10 IS LOSS FROM THE SYSTEM L(10,6) 1.939394E-03 0.000000E+00 1.000000E+02 CC L(8,5) = [(GM DIETARY INTAKE\*0.8)/2]/M(5)CC L(8,5) WAS FIXED AT THE CALCULATED VALUE L(8,5) 2.214 CC DT(8) FIXED AT 75 MIN 0.052 DT(8) DN(8) 8 L(10, 8) = 1CC STEADY STATE SOLUTION CC U(I)=VITAMIN A INTAKE RATE (UMOL/D) CC M(I) = MASS OF VITAMIN A IN COMPARTMENT I (UMOL) H STE 2.561592E+00 0 U(3) 100 CC M(5)=PLASMA RETINOL POOL CC GM M(5) CALCULATED FOR ALL CHILDREN AT ALL TIMES M(5) 0.4625 CC MODEL-PREDICTED MASSES IN COMPARTMENTS 6 & 7 CC M(6) = 529CC M(7) = 4.17H DAT CC DIETARY VITAMIN A INTAKE [U(3)] AS WEIGHTED INPUT

#### Supplementary Data

CC TIME (D) U(3) FRACTIONAL STANDARD DEVIATION 100 FSD=0.05 CC GM ASSESSED INTAKE=704 UG PREFORMED VITAMIN A/D + 23 UG PROVITAMIN A RAE/D CC WEIGHTED U(3)=704+(23/0.8)=733 UG RAE/D=2.56 UMOL RAE/D U(3) 0 2.56 CC GEOMETRIC MEAN PLASMA FRACTION OF ORAL [13C10]RETINYL ACETATE DOSE CC INCLUDED 9-13 CHILDREN/TIME AND 112 CHILDREN AT 4 D 105 FSD=0.05 CC TIME (D) PLASMA FRACTION OF ORAL DOSE 0 0 0.24819662 0.026361583 0.369220693 0.04586997 0.493377025 0.043454498 0.9580861 0.027169027 0.009556731 1.963451096 CC INCREASE WEIGHT TO ACCOUNT FOR INCREASE NUMBER OF CHILDREN AT 4 D 105 FSD=0.025 3.995176137 0.002181794 105 FSD=0.05 6.993033117 0.001045432 10.92628846 0.000553514 15.93270898 0.000522024 21.95402784 0.000388812 28.03872142 0.000352375 CC SIMULATIONS IN COMPARTMENTS 5, 6 AND 7 105 0 2 0.05 20 2 1 27 2 10 10 106 0 2 0.05 20 2 27 1 10 2 10 107 0 2 0.05 20 2 27 1 2 10 10 CC BANGLADESH CC PARAMETER H PAR IC(13)=1 DT(13) = DT(3)DN(13) 8 L(14,13) 0.8 L(0,13) 0.2 L(15,14) 2.152658E+00 0.000000E+00 1.00000E+02 L(17,15) 1.392411E+01 0.000000E+00 1.00000E+02 L(15,17) 1.593562E+00 0.000000E+00 1.00000E+02 L(16, 15) = L(6, 5)L(15,16) 2.168768E-02 0.000000E+00 1.00000E+02 L(20,16) 2.462724E-03 0.000000E+00 1.00000E+02 CC L(18, 15) = [(DIETARY INTAKE\*0.8)/2]/M(15)L(18,15) 1.298

DT(18) 0.052 DN(18) 8 L(20, 18) = 1CC STEADY STATE H STE U(13) 1.219523E+00 0 100 CC M(15) = PLASMA RETINOL POOL CC GM M(15) CALCULATED FOR ALL CHILDREN AT ALL TIMES M(15) 0.382 CC MODEL-PREDICTED MASSES IN COMPARTMENTS 16 & 17 CC M(16)=195 CC M(17) = 3.34H DAT CC DIETARY VITAMIN A INTAKE [U(13)] AS WEIGHTED INPUT TIME (D) U(13) FRACTIONAL STANDARD DEVIATION CC 100 FSD=0.05 CC GM ASSESSED INTAKE=352 UG RAE/D CC U(13) = GM OF INDIVIDUAL CHILD ESTIMATES FOR PREFORMED VA+(PROVITAMIN A/0.8) CC WEIGHTED U(13)=356 UG RAE/D=1.24 UMOL RAE/D U(13) 0 1.24 CC GEOMETRIC MEAN PLASMA FRACTION OF ORAL [13C10]RETINYL ACETATE DOSE CC INCLUDED 5-6 CHILDREN/TIME AND 40 CHILDREN AT 4 D 115 FSD=0.05 TIME (D) PLASMA FRACTION OF ORAL DOSE CC 0 0 0.255070515 0.039304484 0.379283989 0.054793611 0.498899587 0.046096319 0.993991584 0.019386832 2.001794497 0.015871874 CC INCREASE WEIGHT TO ACCOUNT FOR INCREASE NUMBER OF CHILDREN AT 4 D 115 FSD=0.025 3.927820427 0.003016679 115 FSD=0.05 6.993672175 0.001439822 10.99022327 0.00094098 16.00226079 0.001032172 22.00956505 0.001148056 0.001200537 28.04110436 CC SIMULATIONS IN COMPARTMENTS 15, 16 AND 17 115 0 2 0.05 20 2 1 27 2 10 10 116 0 2 0.05 20 2 1 27 2 10 10 117 0 2 20 0.05 2 27 1 2 10 10 CC GUATEMALA CC PARAMETER

H PAR IC(23)=1 DT(23) = DT(3)DN(23) 8 L(24,23) 0.8 L(0,23) 0.2 L(25, 24) = L(5, 4)L(27, 25) = L(7, 5)L(25, 27) = L(5, 7)L(26, 25) = L(6, 5)L(25, 26) = L(5, 6)L(30,26) 1.077104E-03 0.000000E+00 1.000000E+02 CC L(28, 25) = [(DIETARY INTAKE\*0.8)/2]/M(25)L(28,25) 1.335 DT(28) 0.052 DN(28) 8 L(30,28) 1 CC STEADY STATE H STE U(23) 2.833704E+00 0 100 CC M(25)=PLASMA RETINOL POOL CC GM M(25) CALCULATED FOR ALL CHILDREN AT ALL TIMES M(25) 0.848 CC MODEL-PREDICTED MASSES IN COMPARTMENTS 26 & 27 CC M(26) = 1054CC M(27) = 7.65H DAT 100 FSD=0.05 CC GM ASSESSED INTAKE=764 UG RAE/D CC U(23)=GM OF INDIVIDUAL CHILD ESTIMATES FOR PREFORMED VA+(PROVITAMIN A/0.8) CC WEIGHTED U(23)=810 UG RAE/D=2.83 UMOL RAE/D U(23) 0 2.83 CC GEOMETRIC MEAN PLASMA FRACTION OF ORAL [13C10]RETINYL ACETATE DOSE CC INCLUDED 8-16 CHILDREN/TIME AND 112 CHILDREN AT 4 D 125 FSD=0.05 0 0 0.253637423 0.035501178 0.368441789 0.044641924 0.491466246 0.045459892 1.022452771 0.026679538 2.050807582 0.00975629 CC INCREASE WEIGHT TO ACCOUNT FOR INCREASE NUMBER OF CHILDREN AT 4 D 125 FSD=0.025 3.992746907 0.002251418 125 FSD=0.05 7.116623055 0.001131295 11.05266214 0.000565315 16.06009991 0.000534231 22.04308722 0.000400275 28.13302869 0.000506924 CC SIMULATIONS IN COMPARTMENTS 25, 26 AND 27 125 0 2 0.05 20 2 27 1 2 10 10 126 0

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2
             0.05
                                               20
2
             1
                                               27
2
             10
                                               10
127
              0
2
              0.05
                                               20
2
             1
                                               27
2
                                               10
             10
CC FaS PHILIPPINES
135G(35)
XG(35) = (F(6) + F(7)) * (F(5) / M(5)) /
          ((F(6)+F(7))/(M(6)+M(7)))
             1
2
             1
                                               28
CC FaS BANGLADESH
145G(45)
XG(45) = (F(16) + F(17)) * (F(15) / M(15)) /
           ((F(16) + F(17)) / (M(16) + M(17)))
             1
2
             1
                                               28
CC FaS GUATEMALA
155G(55)
XG(55) = (F(26) + F(27)) * (F(25) / M(25)) /
           ((F(26)+F(27))/(M(26)+M(27)))
             1
2
             1
                                               28
100
   M(4)
   M(5)
   M(6)
   M(7)
   U(3)
   R(10,6)
   R(8,5)
   M(14)
   M(15)
   M(16)
   M(17)
   U(13)
   R(20,16)
   R(18,15)
   M(24)
   M(25)
   M(26)
   M(27)
   U(23)
   R(30,26)
   R(28,25)
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