

Title: Metabolite therapy guided by liquid biopsy proteomics delays retinal neurodegeneration

Authors: Katherine J. Wert^{1*}, Gabriel Velez^{1,2,*}, Vijaya L. Kanchustambham³, Vishnu Shankar³, Lucy P. Evans⁴, Jesse D. Sengillo⁵, Richard N. Zare³, Alexander G. Bassuk⁴, Stephen H. Tsang^{6,7,8}, Vinit B. Mahajan^{1,9}

Affiliations:

¹Omics Laboratory, Byers Eye Institute, Department of Ophthalmology, Stanford University, Palo Alto, CA 94304

²Medical Scientist Training Program, Carver College of Medicine, University of Iowa, Iowa City, IA 52242

³Department of Chemistry, Stanford University, Stanford, CA 94305

⁴Departments of Pediatrics and Neurology, Carver College of Medicine, University of Iowa, Iowa City, IA 52242

⁵Bascom Palmer Eye Institute, University of Miami Miller School of Medicine, Miami, FL 33136

⁶Edward S. Harkness Eye Institute, New York-Presbyterian Hospital, New York, NY 10032

⁷Jonas Children's Vision Care and Bernard & Shirlee Brown Glaucoma Laboratory, Department of Ophthalmology, Vagelos College of Physicians and Surgeons, Columbia University, New York, NY 10032

⁸Department of Pathology & Cell Biology, Stem Cell Initiative (CSCI), Institute of Human Nutrition, Vagelos College of Physicians and Surgeons, Columbia University, New York, NY 10032

⁹Veterans Affairs Palo Alto Health Care System, Palo Alto, CA 94304

*Co-first authors

Corresponding Author: Vinit B. Mahajan, Byers Eye Institute, Department of Ophthalmology, Stanford University, Palo Alto, CA, 94304 USA. Phone: 650-723-6995. Fax: 650-498-1528. E-mail: vinit.mahajan@stanford.edu

List of Supplementary Materials:

Supplementary Figures

- Fig. S1.** Gene ontology of the *Pde6a*^{D670G} retina at different stages of degeneration.
- Fig. S2.** Gene ontology of the *Pde6a*^{D670G} vitreous at different stages of degeneration.
- Fig. S3.** Gene ontology distributions of candidate vitreous biomarkers.
- Fig. S4.** DESI-MSI chemical maps of retina from wild-type, arRP, and α -KG treated mice.
- Fig. S5.** DESI-MSI chemical maps of whole eye from wild-type, arRP, and α -KG treated mice.
- Fig. S6.** Tandem-MS analysis of extracted metabolites from the retinal tissue.
- Fig. S7.** DESI-MS analysis of extracted metabolites from the retinal tissue.
- Fig. S8.** Distribution of α -KG across wild-type, arRP, and α -KG treated mice.

Supplementary Tables

- Table S1.** Vitreous biopsies from arRP patients and unaffected controls.
- Table S2.** Proteomic content of human vitreous samples by LC-MS/MS analysis.
- Table S3.** Relative quantification of proteins identified in human vitreous samples.
- Table S4.** Proteomic content of *Pde6a*^{D670G} retina and vitreous by LC-MS/MS analysis.
- Table S5.** Distinct peptides identified in *Pde6a*^{D670G} retina and vitreous samples.
- Table S6.** Protein groups identified in *Pde6a*^{D670G} retina and vitreous samples.
- Table S7.** Relative quantification and comparison of proteins identified in *Pde6a*^{D670G} retina samples.
- Table S8.** Relative quantification and comparison of proteins identified in *Pde6a*^{D670G} vitreous samples.
- Table S9.** Differentially-expressed *Pde6a*^{D670G} retina and vitreous proteins in all disease stages.
- Table S10.** Differentially-expressed *Pde6a*^{D670G} retina and vitreous proteins in early disease stages.
- Table S11.** Candidate biomarkers from *Pde6a*^{D670G} vitreous samples.
- Table S12.** Pathway representation of down-regulated proteins in the *Pde6a*^{D670G} retina.
- Table S13.** Down-regulated proteins in the *Pde6a*^{D670G} retina involved in oxidative phosphorylation.
- Table S14.** Down-regulated proteins in the *Pde6a*^{D670G} retina involved in the tricarboxylic acid (TCA) cycle.
- Table S15.** Down-regulated proteins in the *Pde6a*^{D670G} retina involved in rod outer segment phototransduction.
- Table S16.** Tandem-MS analysis of extracted metabolites from the retinal tissue.

Supplementary Figures

Fig. S1

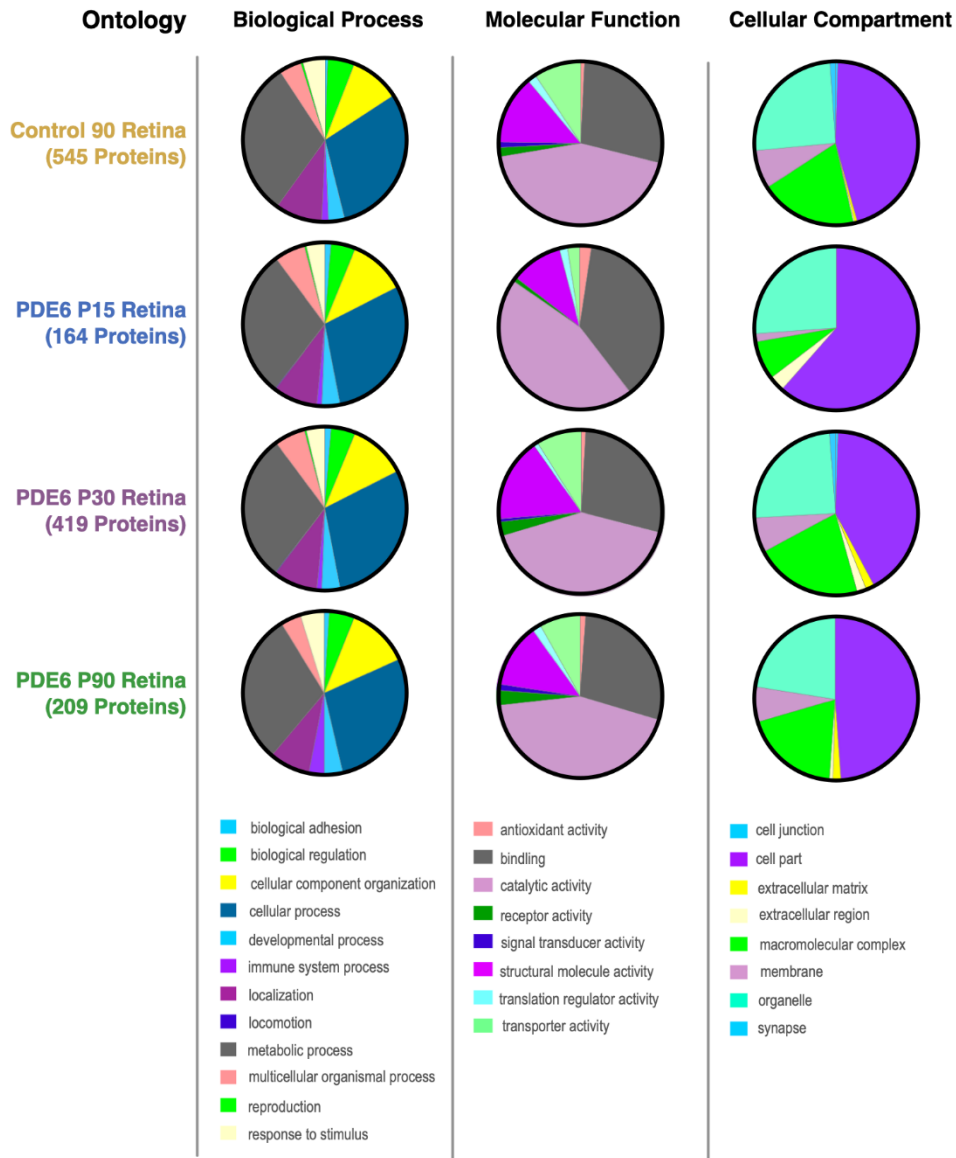


Fig. S2

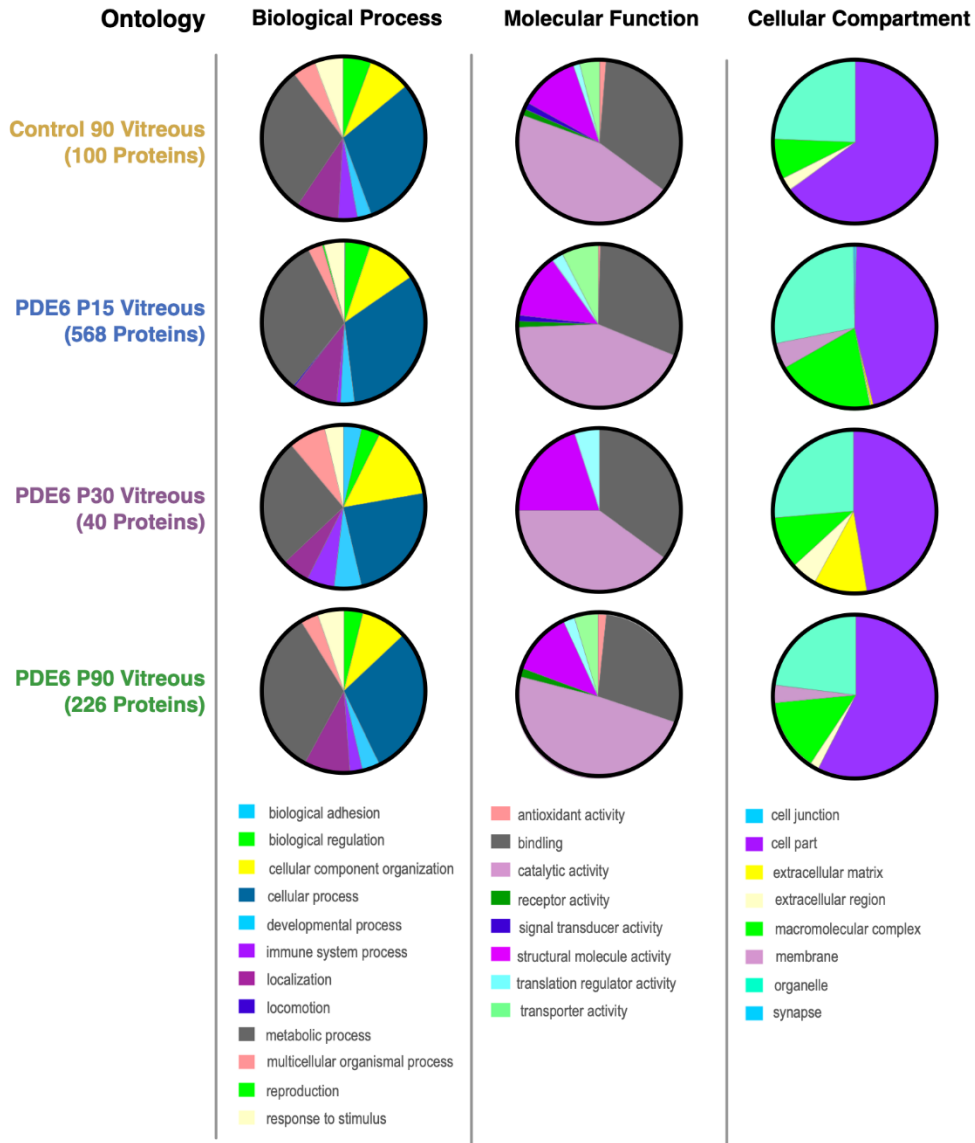


Fig. S3

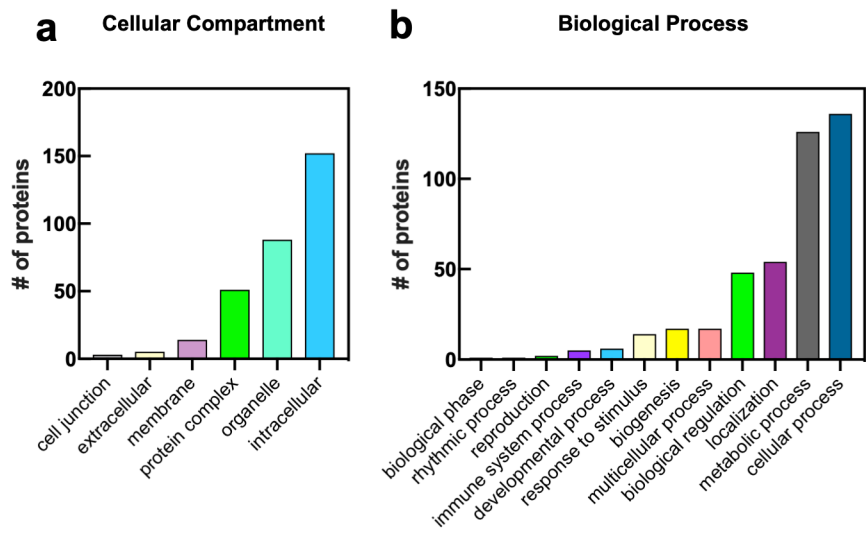


Fig. S4

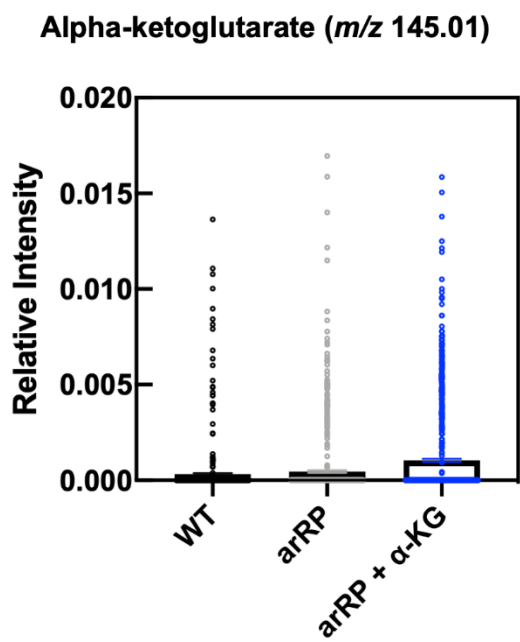


Fig. S5

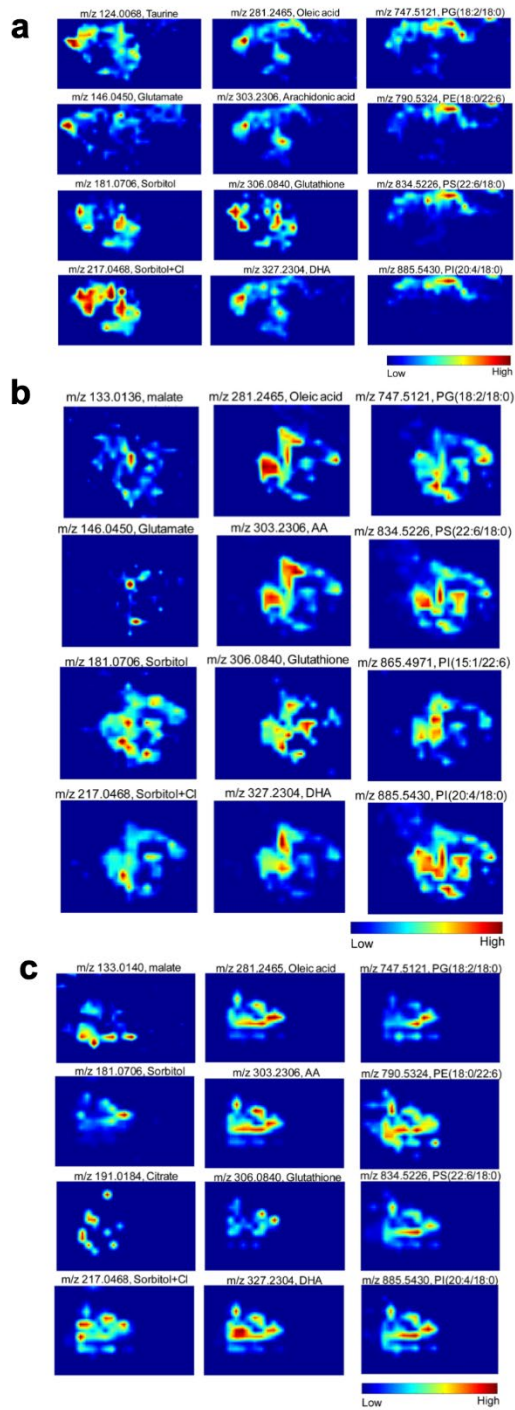


Fig. S6

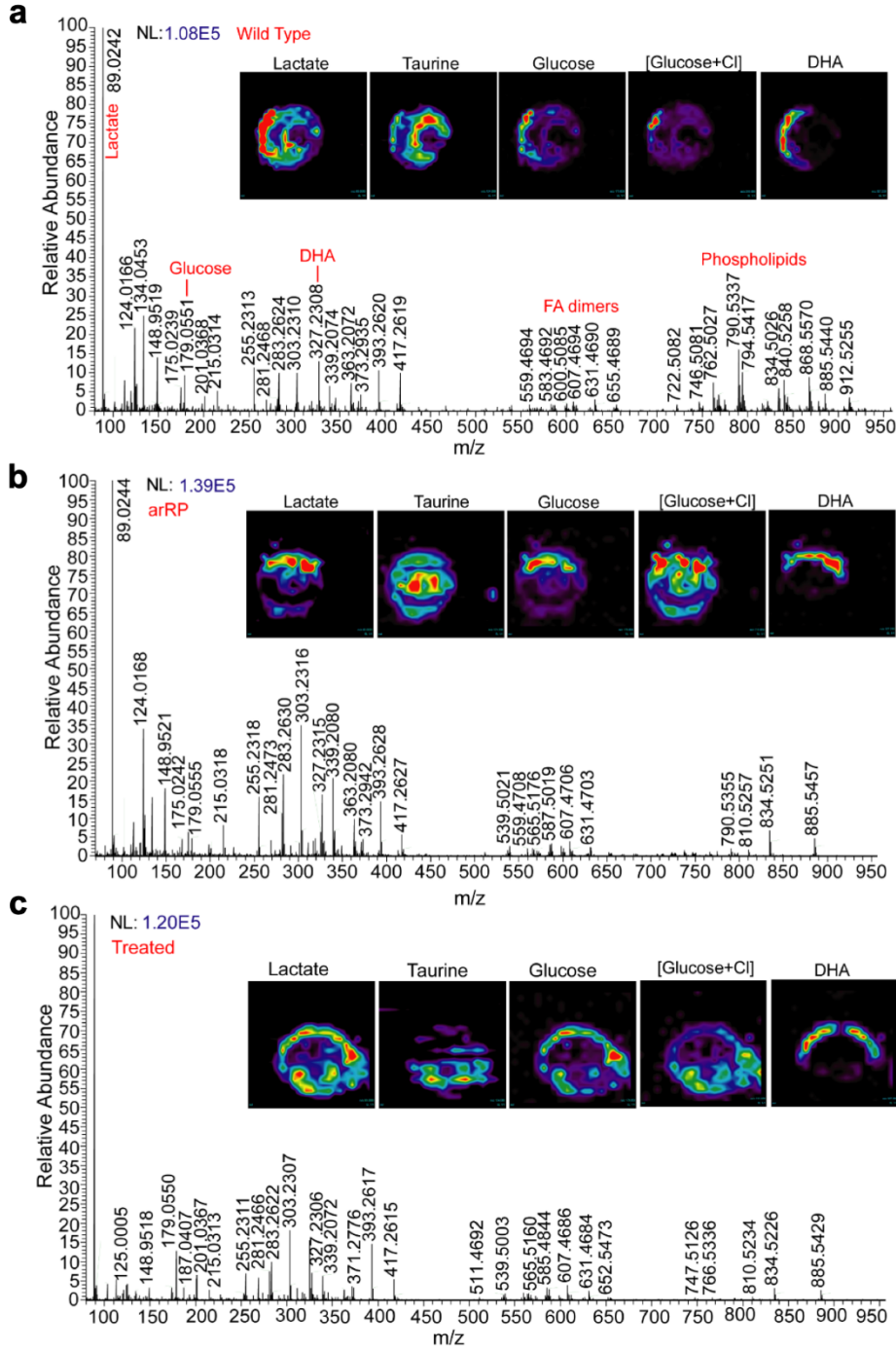
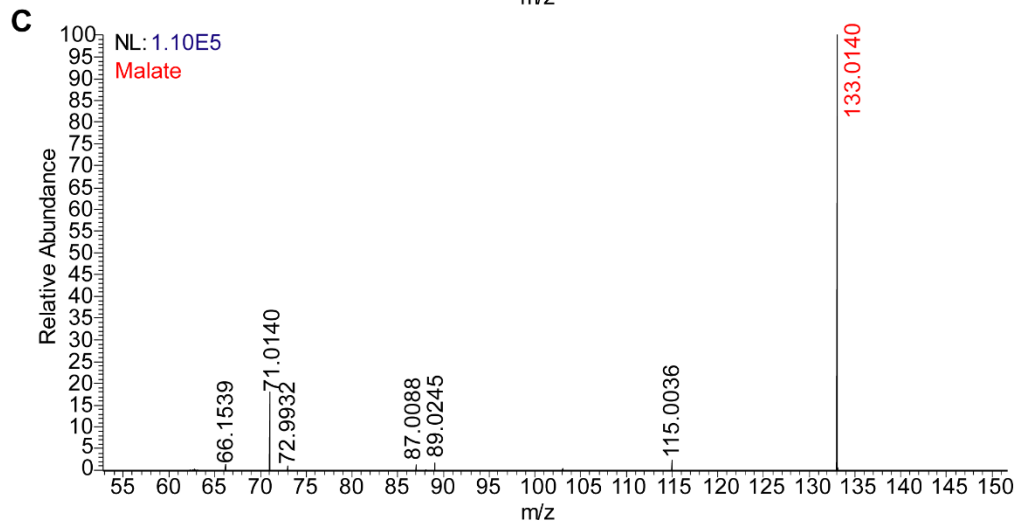
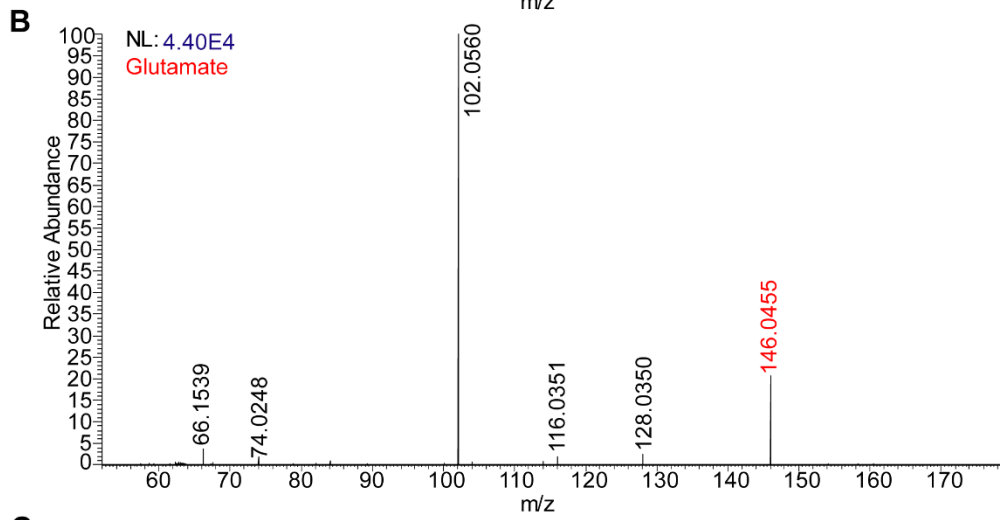
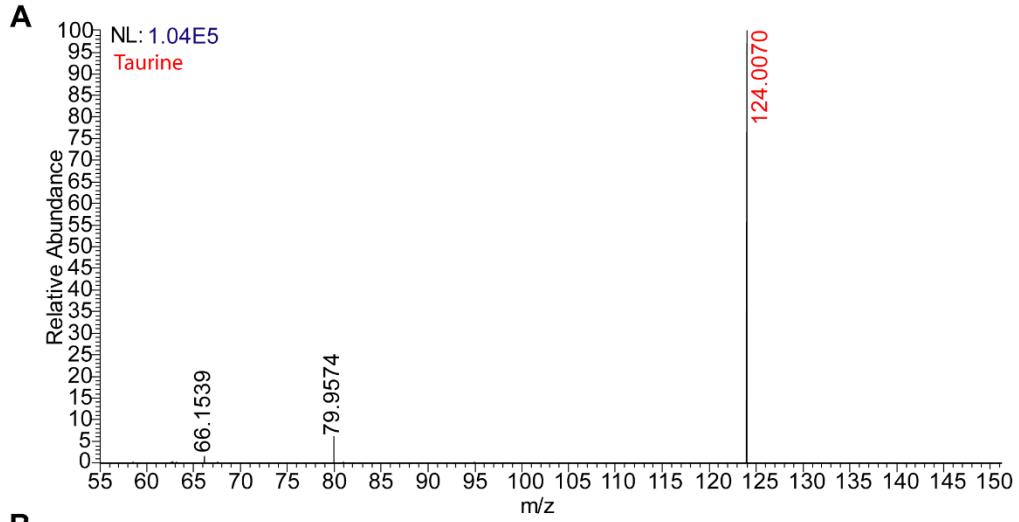
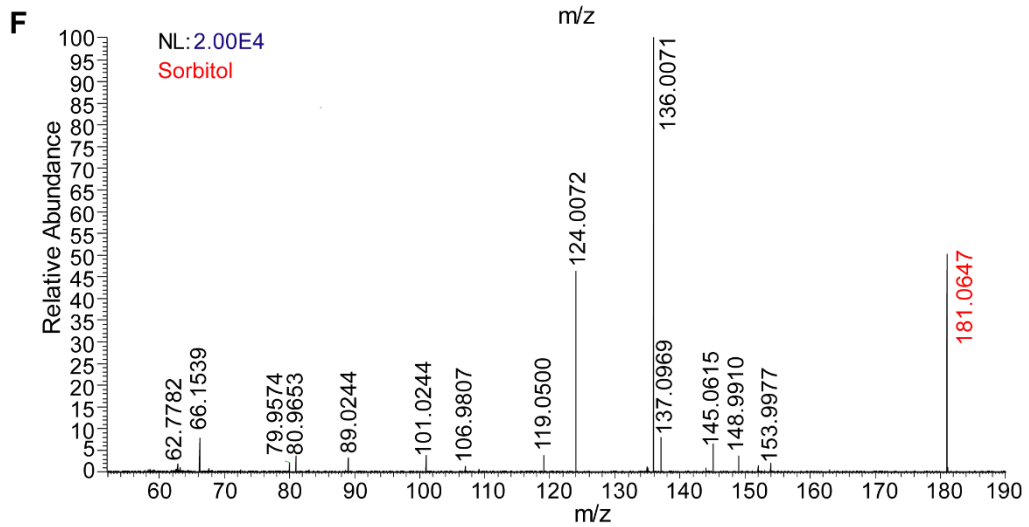
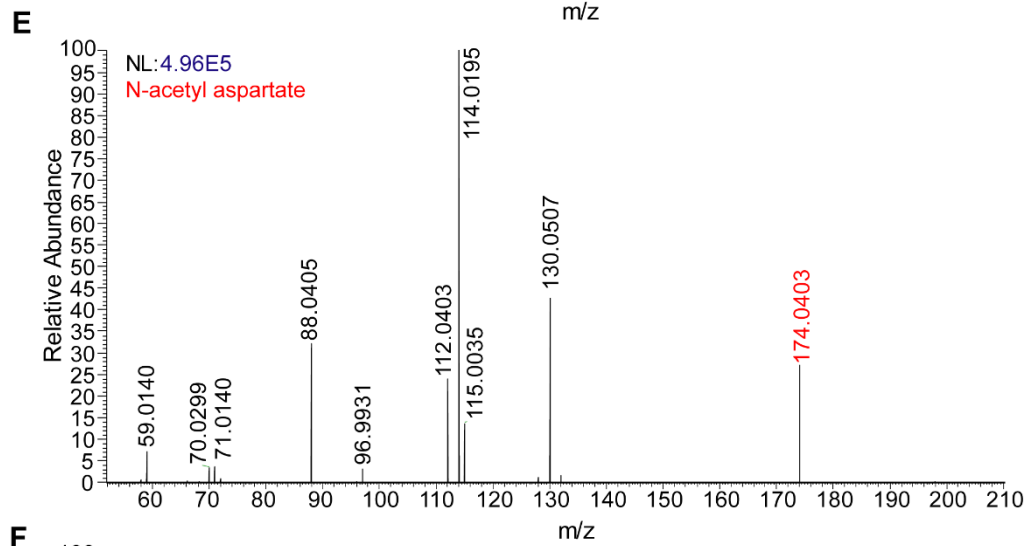
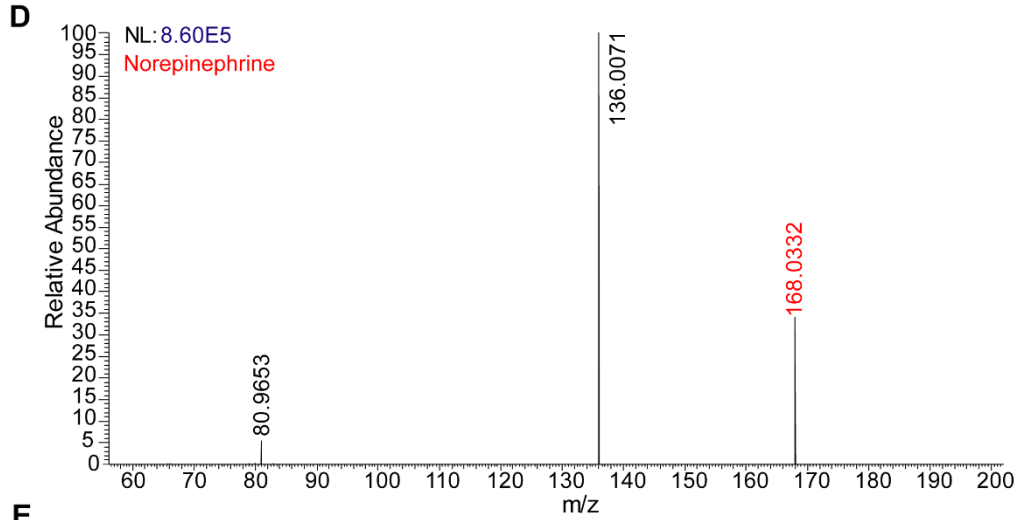
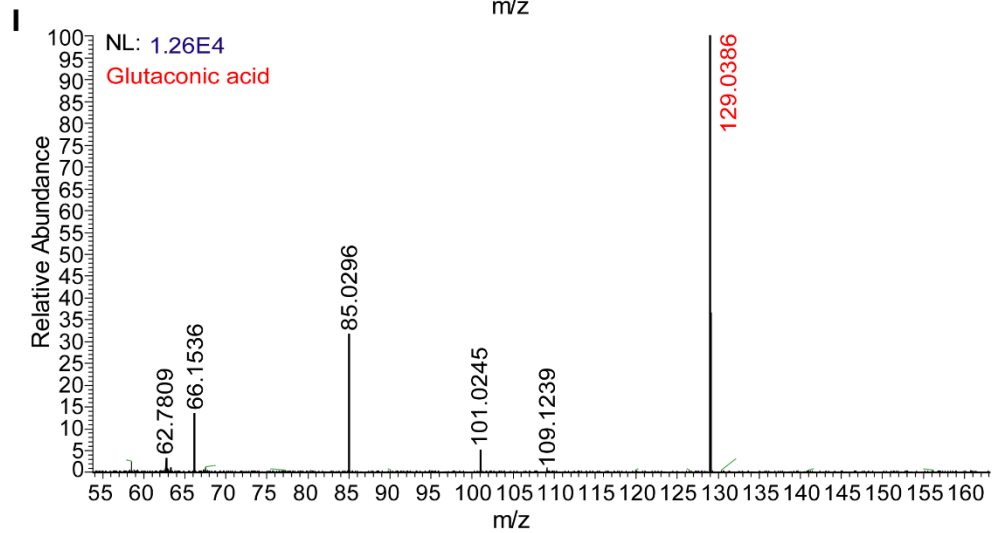
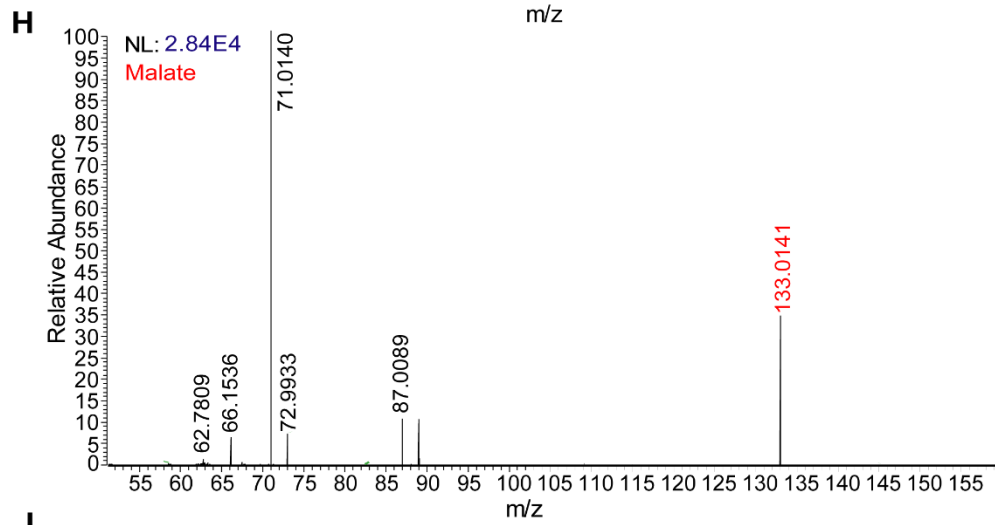
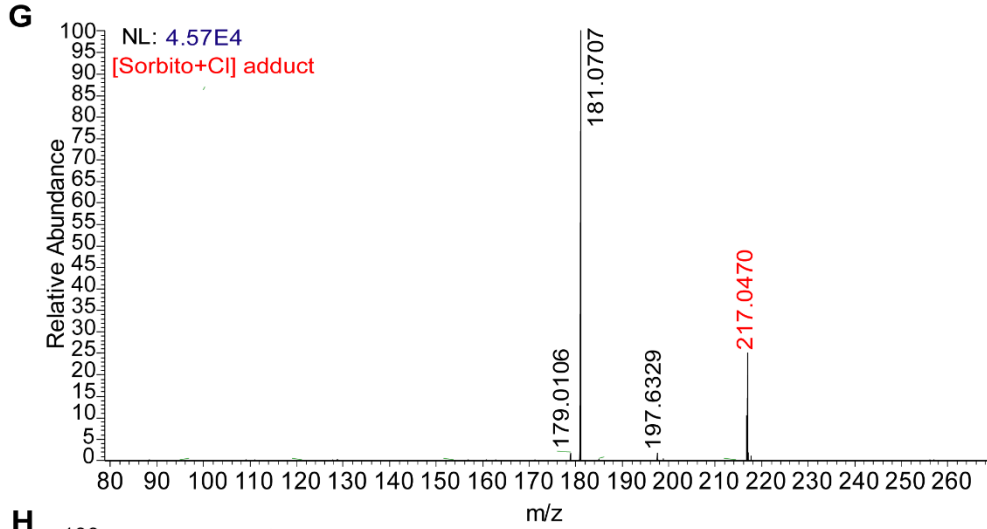
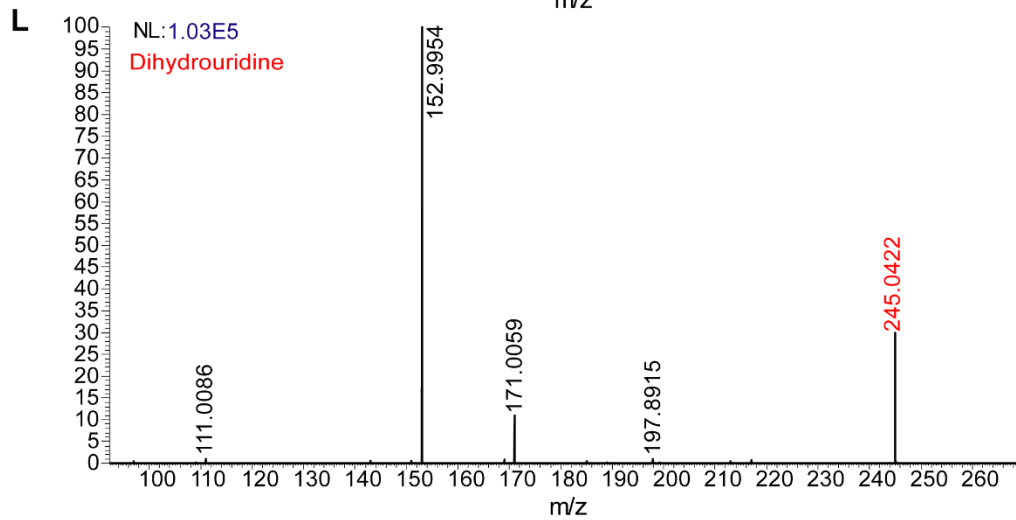
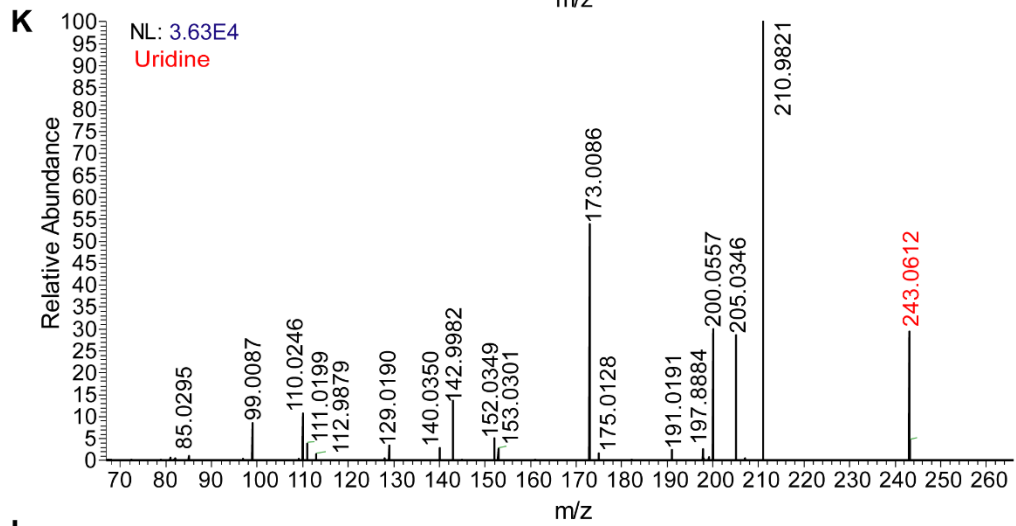
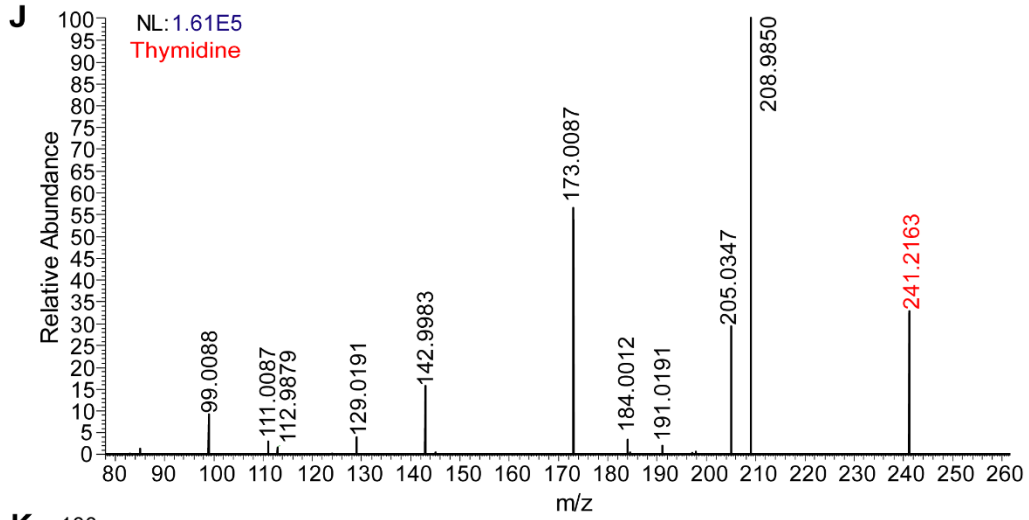


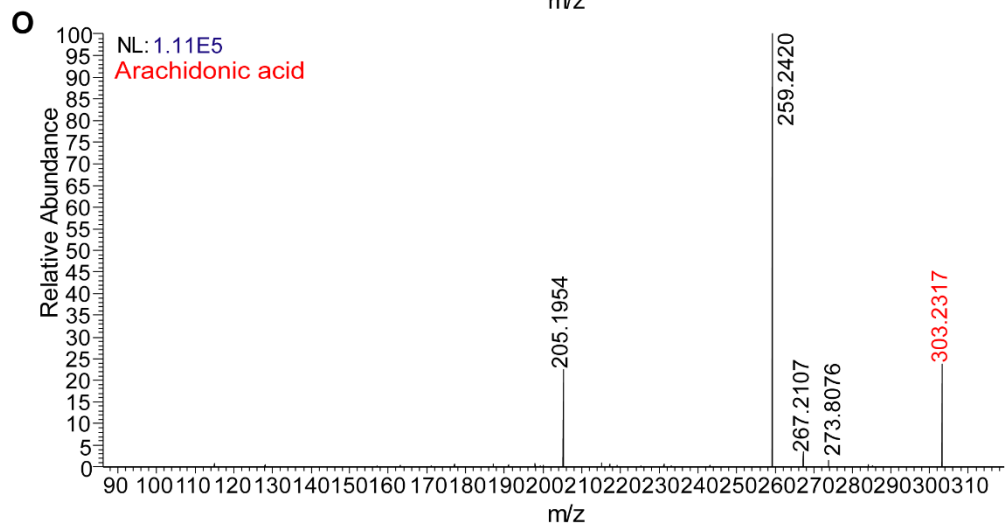
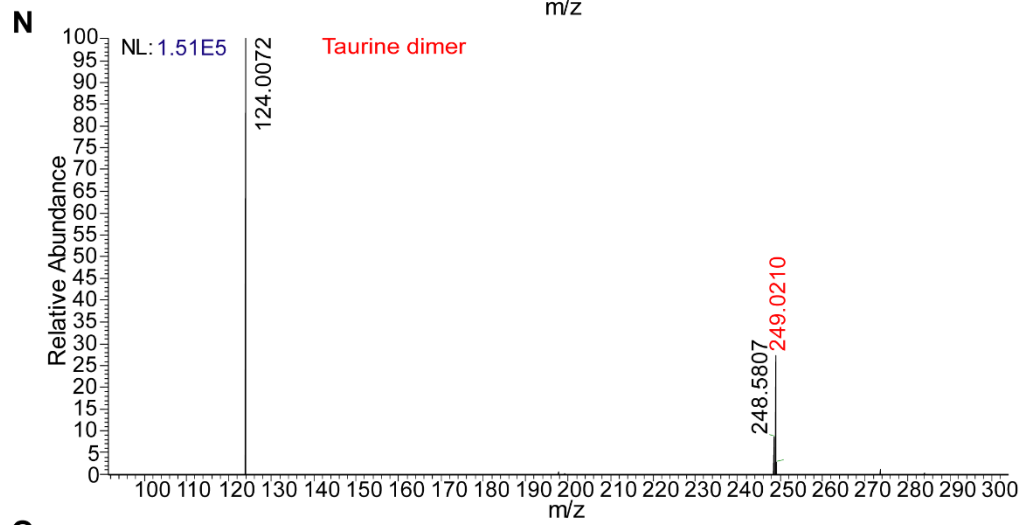
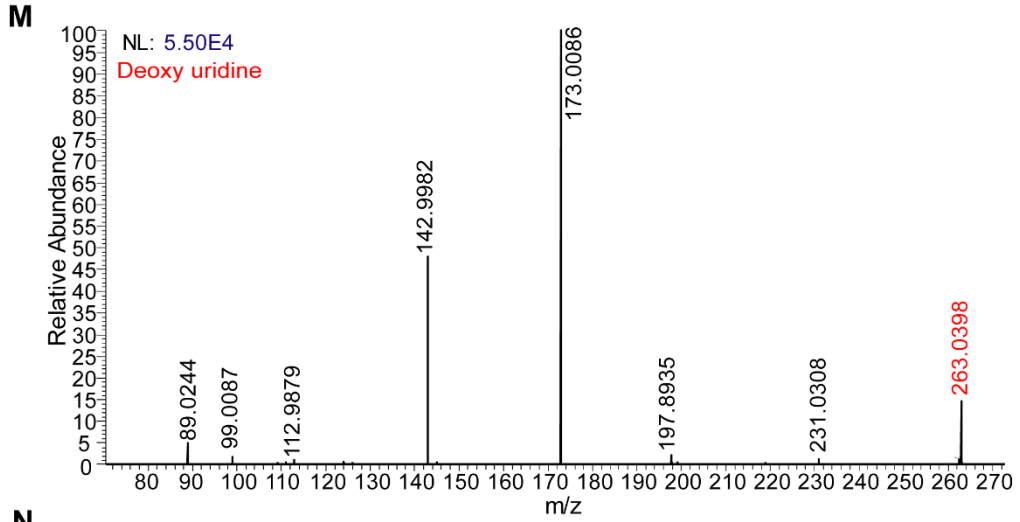
Fig. S7

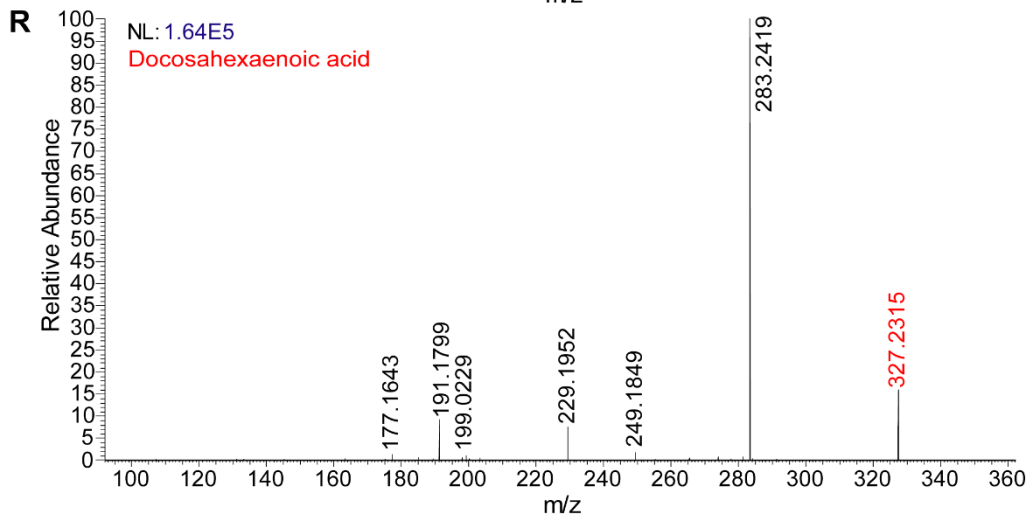
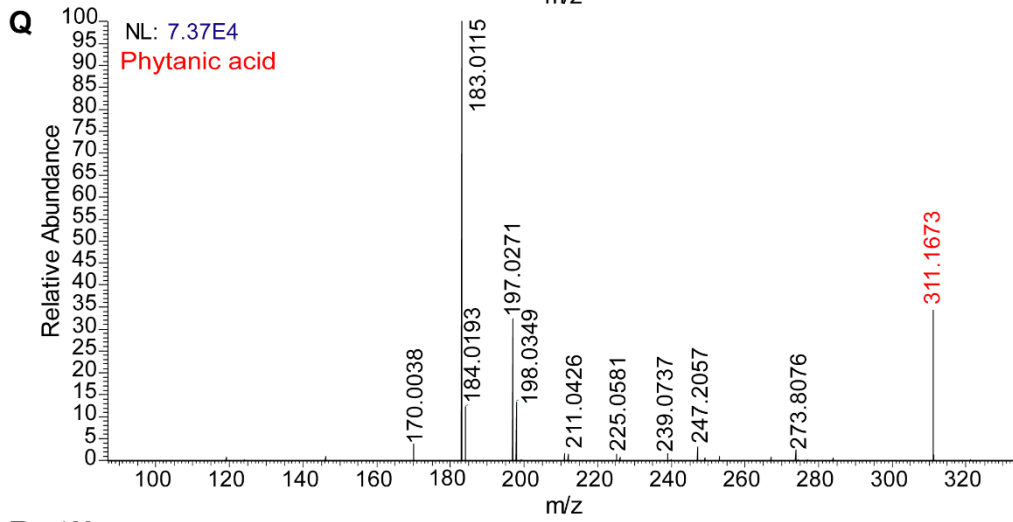
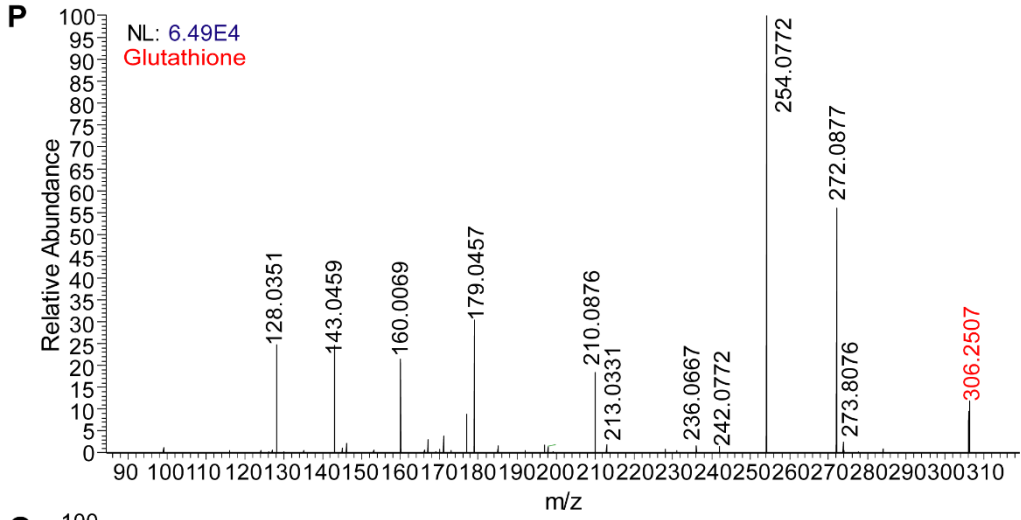


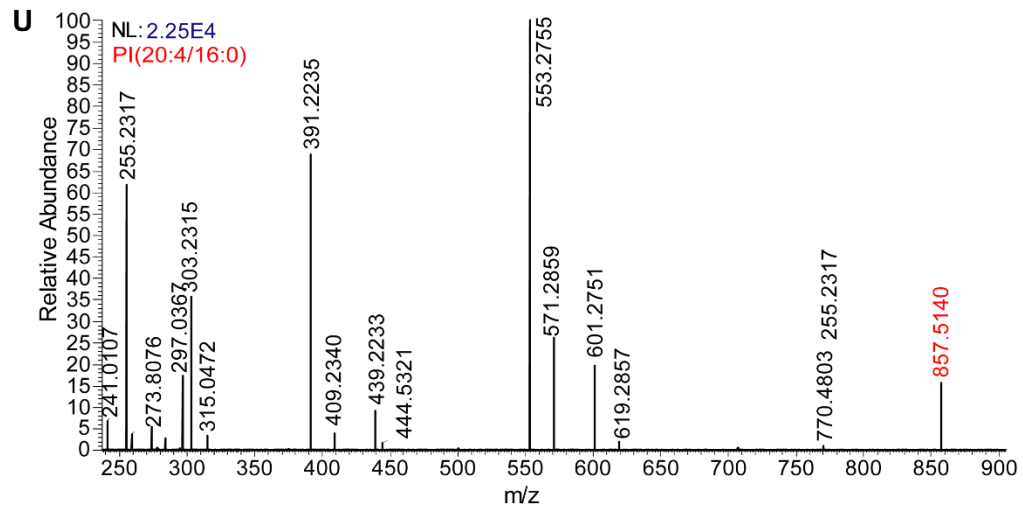
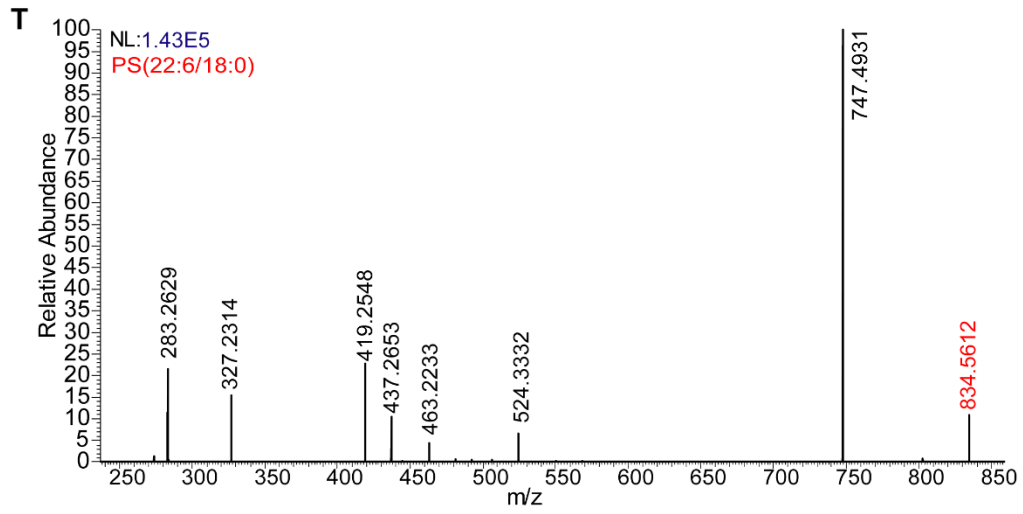
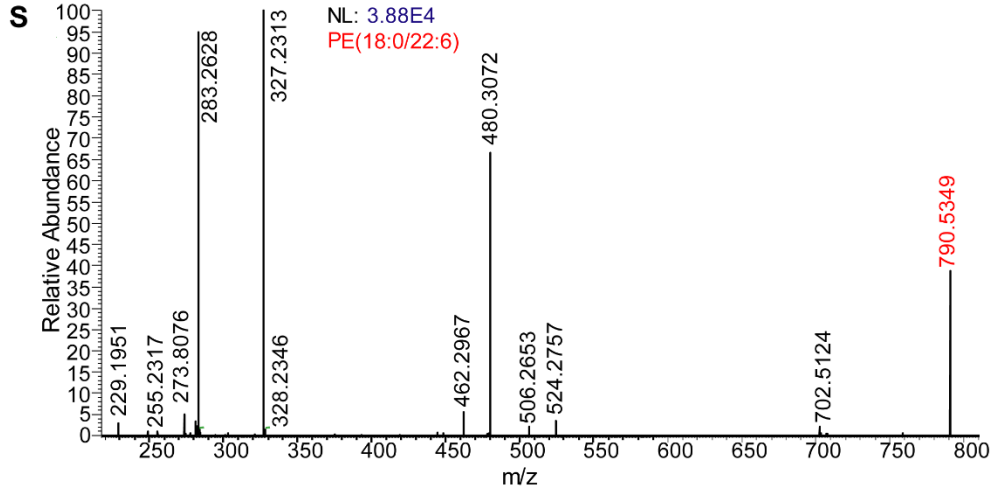












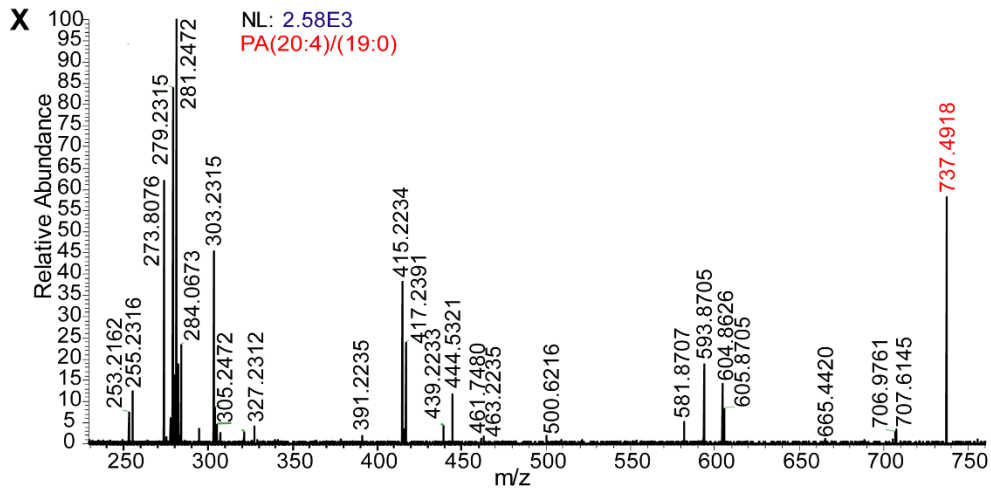
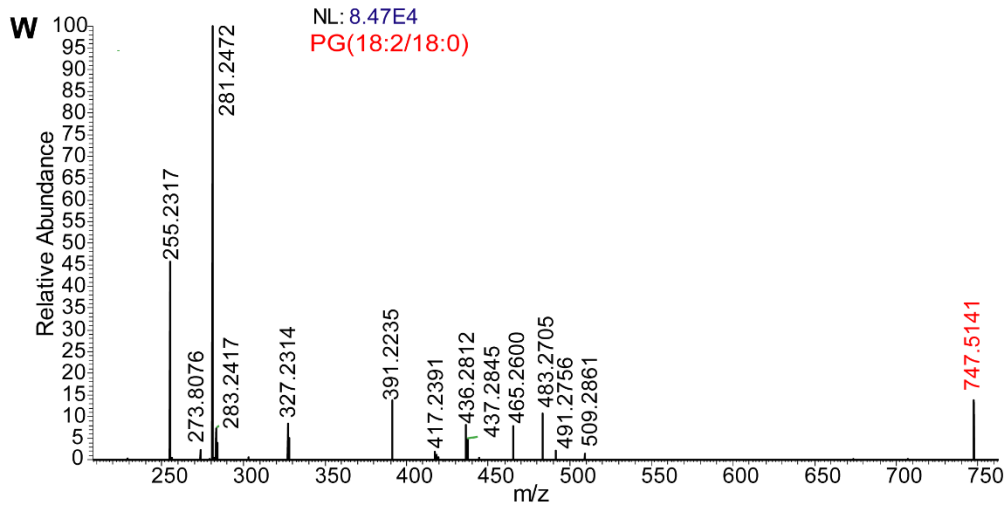
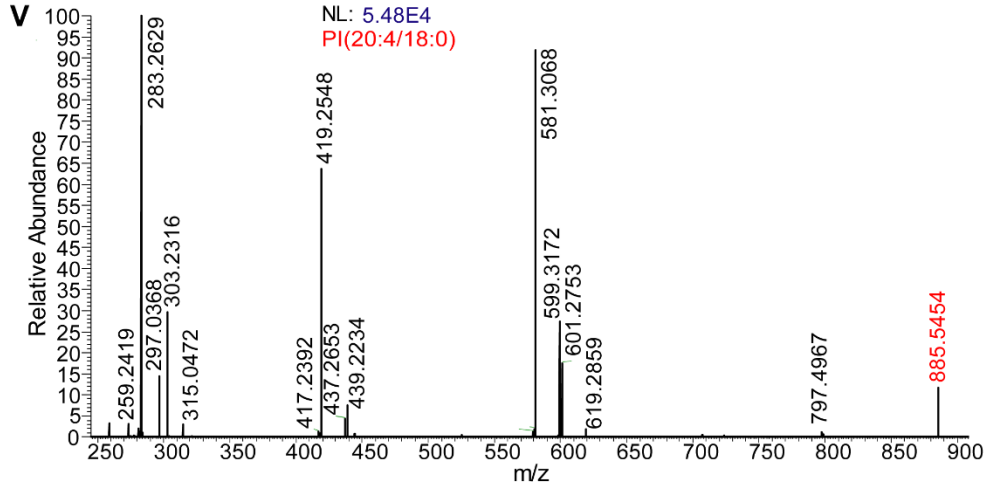
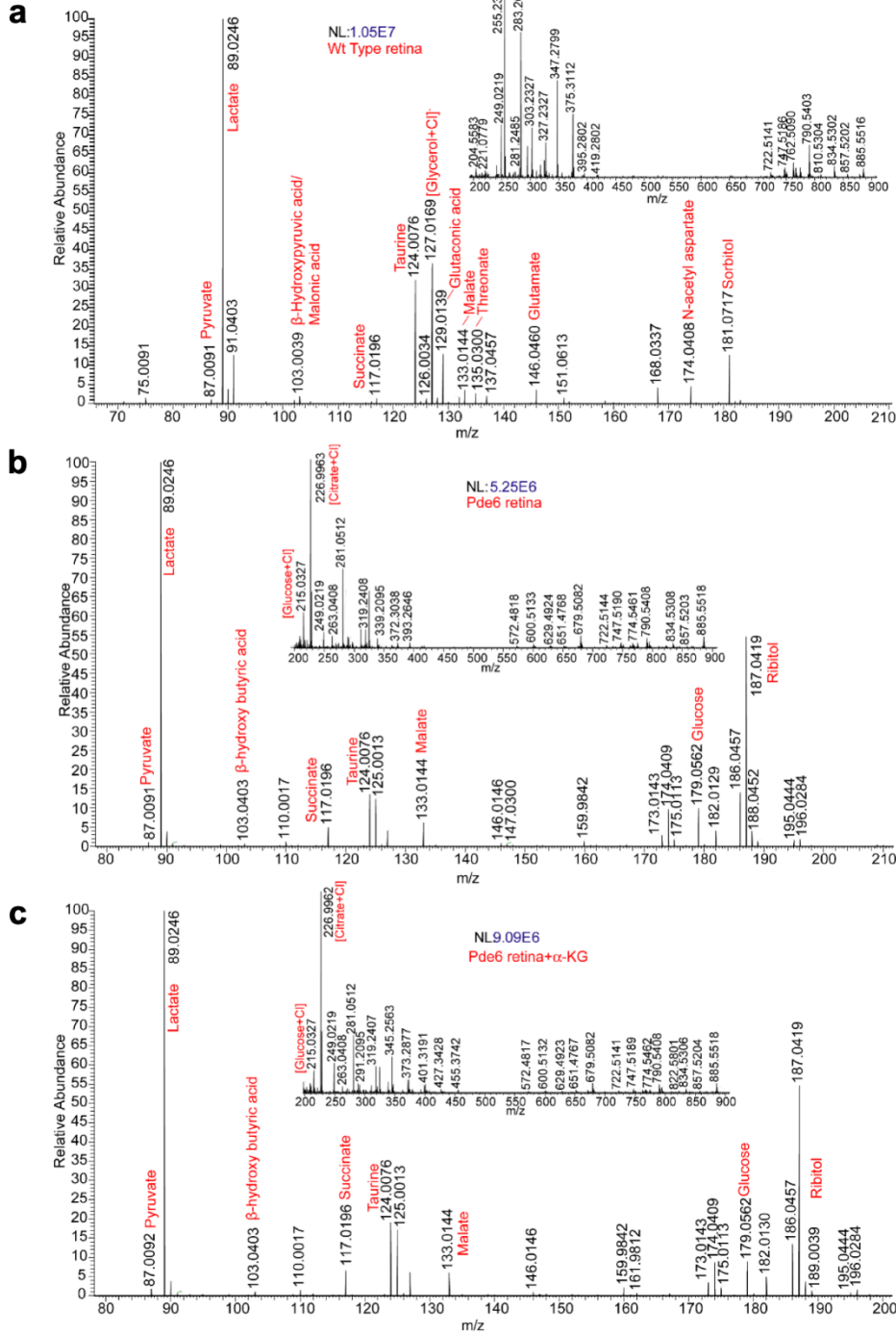


Fig. S8



Supplementary Figure Legends

Fig. S1. Gene ontology (GO) distributions of retina samples. Identified retinal proteins from the wild-type and *Pde6a*^{D670G} mice at P15, P28, and P90. Gene ontology analysis categorized each protein group by biological process, molecular function, and cellular compartment.

Fig. S2. Gene ontology (GO) distributions of vitreous samples. Identified vitreous proteins from the wild-type and *Pde6a*^{D670G} mice at P15, P28, and P90. Gene ontology analysis categorized each protein group by biological process, molecular function, and cellular compartment.

Fig. S3. Gene ontology (GO) distributions of candidate vitreous biomarkers. Gene ontology (GO) categorization of the 446 protein biomarkers by **(a)** cellular compartment and **(b)** biological process.

Fig. S4. DESI-MSI chemical maps of retina. The 2D-chemical maps of individual metabolites are shown for **(a)** wild-type **(b)** arRP and **(c)** α -KG treated mice. PA, phosphatidic acid; PE, phosphatidylethanolamine; PG, phosphatidylglycerol; PI, phosphatidylinositol; PS, phosphatidylserine; DHA, docosahexaenoic acid, CI, collision induced.

Fig. S5. DESI-MSI chemical maps of whole eye. The respective raw MS spectrum across all the pixels in a row and the 2D-chemical maps of individual metabolites is shown for **(a)** wild-type **(b)** arRP and **(c)** α -KG treated mice. NL, normalization level.

Fig. S6. Tandem-MS analysis of extracted metabolites from the retinal tissue: Collision induced dissociation analysis (CID), is shown for small metabolites, fatty acids, and lipids to identify molecular species based on fragmentation pattern: **(a)** Taurine; **(b)** Glutamate; **(c)** Malate; **(d)** Norepinephrine; **(e)** N-acetyl aspartate; **(f)** Sorbitol; **(g)** [Sorbitol + collision-induced (CI)] adduct; **(h)** Malate; **(i)** Glutaconic acid; **(j)** Thymidine; **(k)** Uridine; **(l)** Dihydro-uridine; **(m)** Deoxy-uridine; **(n)** Taurine (dimer); **(o)** Arachidonic acid; **(p)** Glutathione; **(q)** Phytanic acid. **(r)** Docosahexaenoic acid (DHA); **(s)** Phosphatidylethanolamine (PE, 18:0/22:6). **(t)** Phosphatidylserine (PS, 22:6/18:0). **(u)** Phosphatidylinositol (PI, 20:4/16:0). **(v)** Phosphatidylinositol (PI, 20:4/18:0). **(w)** Phosphatidylglycerol (PG, 18:2/18:0). **(x)** Phosphatidic acid (PA, 20:4/19:0). **(y)** 5-L-Glutamyl-L-alanine. **(z)** Phosphatidylserine (PS, 18:0/20:4). NL, normalization level.

Fig. S7. DESI-MS analysis of extracted metabolites from the retinal tissue: Electrospray ionization is performed on the retina tissue extraction in solvents in the mass range m/z 50-1000. Red labels denote peaks assigned to identified metabolites and lipids. NL, normalization level. Results are displayed as the relative intensity of the corresponding metabolite normalized by the total ion current (sum of all intensity values from the detected ions).

Fig. S8. Distribution of α -KG across wild-type, arRP, and α -KG treated mice: Levels of α -KG (m/z 145.01) are not significantly different between arRP and treated mice (fold change 1.26, false discovery rate < 0.05). The supplemental α -KG in treated state is consumed and reflected in the increased intensity of other metabolites in TCA and glutamate-glutamine conversion pathways.