

Supporting Information

Room Temperature Acceptorless Alkane Dehydrogenation from Molecular σ -Alkane Complexes

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S.1. Experimental details

S.1.1. General Methods

All manipulations (unless otherwise stated) were performed under an atmosphere of argon, using standard Schlenk techniques on a dual vacuum/inlet manifold or by employment of an MBraun glovebox. Glassware was dried in an oven at 130°C overnight prior to use. Pentane, benzene and dichloromethane (abbreviated as CH₂Cl₂) were dried using an MBraun SPS-800 solvent purification system and degassed by three freeze-pump-thaw cycles. 1,2-F₂C₆H₄ (abbreviated as F₂C₆H₄) was stirred over Al₂O₃ for two hours then over CaH₂ overnight before being vacuum distilled and subsequently degassed by three freeze-pump-thaw cycles. d₂-dichloromethane (abbreviated to CD₂Cl₂) and d₃-acetonitrile (abbreviated to d₃-MeCN) was dried by stirring over CaH₂ overnight before being vacuum distilled onto 3 Å molecular sieves and subsequently degassed by three freeze-pump-thaw cycles. d₆-Benzene (abbreviated as C₆D₆) was stirred over Na pieces overnight before being vacuum distilled onto 3 Å molecular sieves and subsequently degassed by three freeze-pump-thaw cycles. Isobutene and Isobutane was purchased from CK Gases and used as received. All other chemicals were purchased from commercial vendors and used as received.

[Rh(Cy₂PCH₂CH₂PCy₂)(C₆H₄F₂)]^{S1} [1-C₆H₄F₂][BAR^F₄],
[Rh(Cy₂PCH₂CH₂PCy₂)(NBD)]^{S2} (NBD = norbornadiene) [1-NBD][BAR^F₄] and
[Rh(Cy₂PCH₂CH₂PCy₂)(NBA)]^{S2} (NBA = norborane) [1-NBA][BAR^F₄] were prepared by the literature procedures.

Solution NMR data were collected on either a Bruker AVD 500 MHz or a Bruker Ascend 400 MHz spectrometer at room temperature unless otherwise stated. Non-deuterated solvents were locked to standard external CD₂Cl₂ solutions. Residual protio solvent resonances were used as a reference for ¹H NMR spectra. ²H NMR spectra were referenced to CD₂Cl₂ (δ 5.32). ³¹P{¹H} NMR spectra were referenced externally to 85 % H₃PO₄ (D₂O). All chemical shifts (δ) are quoted in ppm and coupling constants in Hz.

Solid state NMR (SSNMR) samples were prepared packing powdered microcrystalline sample into either a 3.2 mm or 4 mm zirconia solid state rotor inside an argon filled glove box. SSNMR spectra were obtained on a Bruker Avance III HD

spectrometer equipped with a 9.4 Tesla magnet, operating at 100.6 MHz for ^{13}C and 162 MHz for ^{31}P , respectively, and a MAS rate of 10 kHz. Relaxation time for ^1H and contact time for $^{31}\text{P}\{^1\text{H}\}$ CP/MAS, $^{13}\text{C}\{^1\text{H}\}$ CP/MAS, $^{13}\text{C}\{^1\text{H}\}$ NQS and FSLG-HETCOR NMR experiments were optimized for each compound as appropriate. All $^{13}\text{C}\{^1\text{H}\}$ CP/MAS spectra were referenced to adamantane (up field methine resonance, δ 29.5)^{S3} on a scale where δ (TMS) = 0 as a secondary reference. The temperature for Variable Temperature (VT) NMR experiments at low temperatures was externally calibrated using lead nitrate (PbNO_3). The ^1H chemical shifts obtained from $^1\text{H}/^{13}\text{C}$ Frequency Switched Lee-Goldburg HETCOR SSNMR spectra were referenced internally to the *p*-ArH resonance of the $[\text{BAr}_4^{\text{F}}]^-$ (δ = 7.12).^{S4}

Gas phase ^1H NMR spectroscopy was carried out using a Bruker Ascend 400 MHz spectrometer. The spectrometer was pre-locked and shimmed to a separate CD_2Cl_2 sample in a similar bore tube. The T1 delay was set to 1 s, and this has been previously shown to allow for the accurate comparison of integrals.^{S5}

Electrospray ionization mass spectrometry (ESI-MS) was carried out using a Bruker MicrOTOF instrument directly connected to a modified Innovative Technology glovebox.^{S6} Typical acquisition parameters were used (sample flow rate: $4 \mu\text{L min}^{-1}$, nebulizer gas pressure: 0.4 bar, drying gas: Argon at 333 K flowing at 4 L min^{-1} , capillary voltage: 4.5 kV, exit voltage: 60 V). The spectrometer was calibrated using a mixture of tetraalkyl ammonium bromides $[\text{N}(\text{C}_n\text{H}_{2n+1})_4]\text{Br}$ ($n = 2-8, 12, 16$ and 18). Samples were diluted to a concentration of $1 \times 10^{-6} \text{ M}$ in the appropriate solvent before sampling by ESI-MS.

Gas Chromatography Electron Ionization-Mass Spectrometry (GC EI-MS) analyses were performed on an Agilent 7200 quadrupole time of flight (Q-ToF) instrument equipped with a 7890B gas chromatograph and a PAL auto-sampler fitted with a 2.5 mL headspaces syringe. Instrument control and data processing were performed using Agilent MassHunter software. The system was calibrated within 1 hour prior to the analysis and its mass accuracy with external calibration (as used for these experiments) is better than 5 ppm for 2 hours following calibration. Samples were prepared in 20 mL headspace vials. Vials were incubated at $35 \text{ }^\circ\text{C}$ for 30 seconds before a 2.5 mL sample of the headspace was taken and injected into the GC inlet (headspace syringe was held at 40°C). The GC-inlet was operated in split mode held

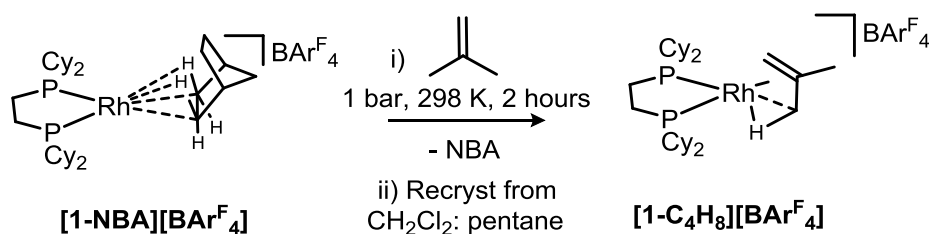
at 300 °C with a 2:1 split. The column was a Restek RT Q-Bond 30 m x 320 µm with at 10 µm film thickness. The oven was held at 40 °C for 4 min then ramped at 20 °C/min to 300 and held for 3 min. Column flow rate was 1 mL/min. Mass spectrometer was operated in EI mode and with the ionization energy set to 20 eV.

Elemental analyses were conducted by Mr. Stephan Boyer at London Metropolitan University.

S.2. Synthetic procedures

S.2.1. $[\text{Rh}(\text{Cy}_2\text{PCH}_2\text{CH}_2\text{PCy}_2)(\text{C}_4\text{H}_8)][\text{BAr}^{\text{F}}_4]$, $[\text{1-C}_4\text{H}_8][\text{BAr}^{\text{F}}_4]$

S.2.1.1. Synthesis of $[\text{1-C}_4\text{H}_8][\text{BAr}^{\text{F}}_4]$



To an orange sample of crystalline $[\text{1-NBA}][\text{BAr}^{\text{F}}_4]$ (200 mg, 135 μmol) in an evacuated ($< 3 \times 10^{-2}$ mbar) J Young flask (~ 50 mL) isobutene gas (1 bar, 298 K) was added and left to stand for 2 hours, to form $[\text{Rh}(\text{Cy}_2\text{PCH}_2\text{CH}_2\text{PCy}_2)(\text{C}_4\text{H}_8)][\text{BAr}^{\text{F}}_4]$, $[\text{1-C}_4\text{H}_8][\text{BAr}^{\text{F}}_4]$. After this time, working under an atmosphere of isobutene (1 bar) and at -78 $^\circ\text{C}$ in an acetone/dry ice bath, the sample was dissolved in CH_2Cl_2 (3 mL), filtered *via* cannula and layered with pentane (25 mL) at the same temperature. The solution was warmed to ambient temperature and after 3 days yielded orange block like crystals suitable for single crystal x-ray diffraction. Yield: 169 mg (86 %). The crystalline material is stored in a glove box in a freezer operating at -25 $^\circ\text{C}$.

Note: CH_2Cl_2 and pentane used in this preparation were freeze-pump-thawed degassed three times and then saturated and stored with isobutene gas (1 bar) prior to use. When solvents were left under an argon atmosphere, ~ 50 % decomposition of $[\text{1-C}_4\text{H}_8][\text{BAr}^{\text{F}}_4]$ to $[\text{1-BAr}^{\text{F}}_4]^{\text{S}2}$ occurs in 2 hours.

S.2.1.2. Characterization data for [1-C₄H₈][BAr^F₄]

¹H NMR (CD₂Cl₂, 298 K, 400 MHz): δ 7.72 (s, 8H, ortho-ArH), 7.55 (s, 4H, para-ArH), 2.09-1.64 (br m, 28H, overlapping aliphatic CH), 1.61-1.42 (br s, 8H, aliphatic CH), 1.40-1.096 (br m, 20H, overlapping aliphatic CH).

¹H NMR (CD₂Cl₂, 183 K, 400 MHz): δ 7.75 (s, 8H, ortho-ArH), 7.56 (s, 4H, para-ArH), 3.75 (br s, ~ 2H, alkene CH), 2.09-1.64 (br m, ~ 30H, overlapping aliphatic CH), 1.40-1.09 (br m, ~ 20H, overlapping aliphatic CH), -0.15 (br, CH₃).

Note: The low temperature limit was not reached at 183K in the ¹H NMR spectrum so integrated values are approximate. NMR samples prepared under argon and immediately run after preparation.

³¹P{¹H} NMR (CD₂Cl₂, 298 K, 162 MHz): δ 95.3 (d, J_{RhP} 179 Hz).

³¹P{¹H} NMR (CD₂Cl₂, 183 K, 162 MHz): δ 97.6 (dd., J_{RhP} 201 Hz, J_{PP} 26 Hz *trans to agostic interaction*), 93.6 (d. of d., J_{RhP} 158 Hz, J_{PP} 26 Hz *trans to alkene*).

¹⁹F{¹H} NMR (CD₂Cl₂, 298 K, 377 MHz): δ -62.9 (s).

¹³C{¹H} NMR (CD₂Cl₂, 298 K, 126 MHz): δ 161.1 (1:1:1:1 q, J_{CB} 50 Hz, ipso-ArC), 134.6 (s, ortho-ArC), 128.8 (q, J_{CF} 32 Hz, meta-ArC), 124.8 (q, J_{CF} 272 Hz, CF₃), 117.3 (s, para-ArC), 53.8 (qu. CD₂Cl₂), 37.3 (s, cyclohexyl-CH), 30.9 (s, cyclohexyl-CH₂), 29.9 (s, cyclohexyl-CH₂), 29.6 (s, cyclohexyl-CH₂), 29.2 (s, cyclohexyl-CH₂), 26.5 (cyclohexyl-CH₂), 25.7 (cyclohexyl-CH₂), 25.3 (cyclohexyl-CH₂), 22.1 (s, cyclohexyl-CH₂).

¹³C{¹H} NMR (CD₂Cl₂, 183 K, 126 MHz): δ 161.1 (1:1:1:1 q, J_{CB} 50 Hz, ipso-ArC), 134.6 (s, ortho-ArC), 128.8 (q, J_{CF} 32 Hz, meta-ArC), 124.8 (q, J_{CF} 272 Hz, CF₃), 117.3 (s, para-ArC), 111.5 (br s, alkene-CH), 72.6 (br s, alkene-CH), 53.8 (qu. CD₂Cl₂), 36.7 (br s.), 35.2 (br s.), 31.1 (s, cyclohexyl-CH₂), 29.4 (m, cyclohexyl-CH₂), 25.6 (s, cyclohexyl-CH₂), 24.9 (s, cyclohexyl-CH₂), 24.3 (s, cyclohexyl-CH₂), 22.2 (s, cyclohexyl-CH₂), 19.4 (br. s).

³¹P{¹H} SSNMR (162 MHz, 158 K, 10 kHz spin rate): δ 94.8 (br m, *coincident environments*).

$^{13}\text{C}\{^1\text{H}\}$ SSNMR (101 MHz, 158K, 10 kHz spin rate): δ 161.1 (ipso-ArC), 131.2 (ortho-ArC), 127.4 (meta-ArC), 122.9 (br, CF_3), 115.6 (para-ArC), 113.1 (para-ArC), 108.6 ($\text{C}=\text{CH}_2$), 70.6 ($\text{C}=\text{CH}_2$), 35.7, 34.3, 31.2, 29.8, 27.5, 24.1, 22.7, 21.9, 15.7.

Note: Signals at 108.6 and 70.6 disappeared upon addition of H_2 , to form **[1- C_4H_{10}][BAr^{F}_4]**. The agostic $-\text{CH}_3$ could not be unambiguously identified, see section S.2.2.

ESI-MS: Not stable under mass spectrometric conditions (20eV, 333K). Species with appropriate isotopic distributions at m/z found = 581.240, calculated to $[(\text{Cy}_2\text{PCH}_2\text{CH}_2\text{PCy}_2)\text{Rh}(\text{N}_2)_2]^+$ (581.242). There is no evidence for the nitrogen compound in bulk samples so it is assumed to form via an in-situ ESI-MS process.

Elemental analysis found (calc. for $\text{C}_{62}\text{H}_{68}\text{BF}_{24}\text{P}_2\text{Rh}$): C 52.39 (51.54), H 4.87 (4.74).

S.2.1.3. NMR spectra of [1-C₄H₈][BAr^F₄]

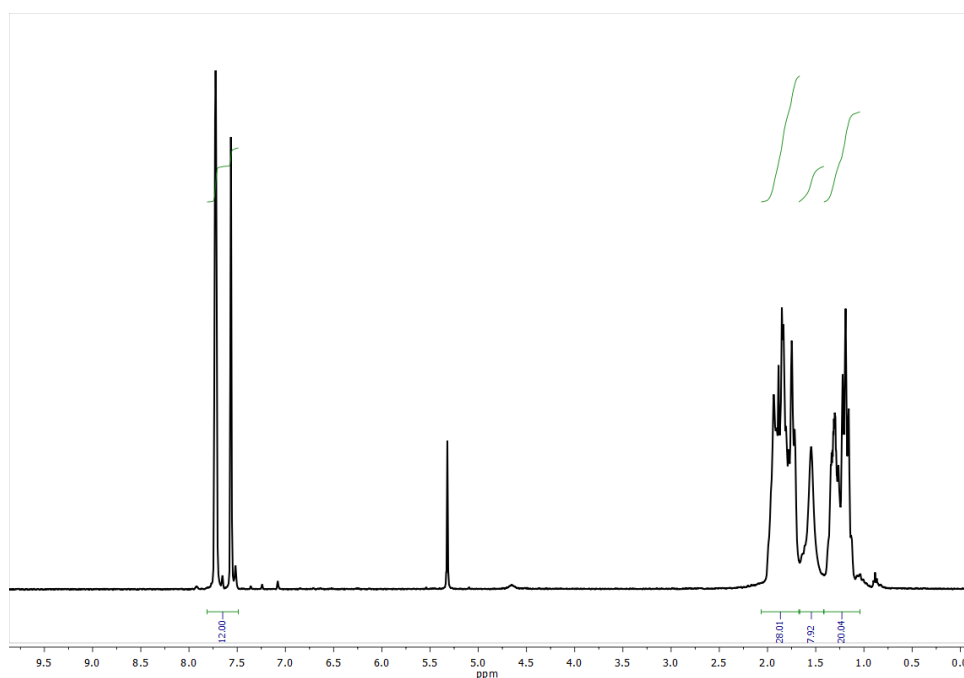


Figure S1: The solution ¹H NMR (CD₂Cl₂, 298 K, 400 MHz) spectrum of [1-C₄H₈][BAr^F₄].

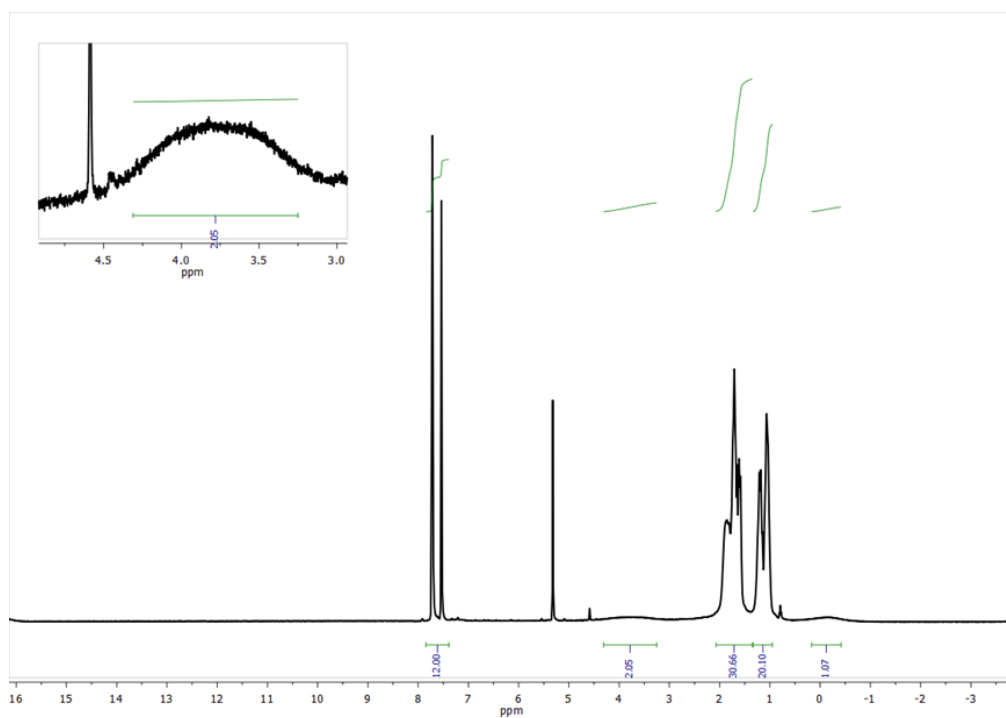


Figure S2: The solution ¹H NMR (CD₂Cl₂, 183 K, 400 MHz) spectrum of [1-C₄H₈][BAr^F₄]. The low temperature limit was not reached at 183K, so integrated values are approximate.

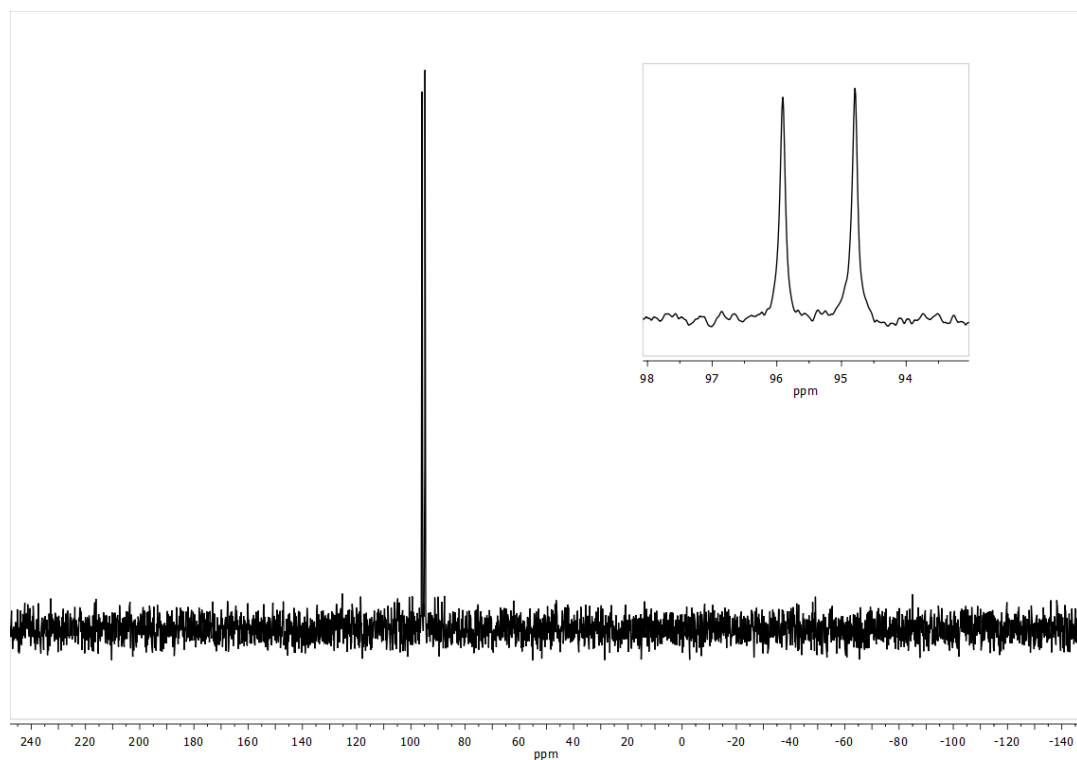


Figure S3: The solution $^{31}\text{P}\{^1\text{H}\}$ NMR (CD_2Cl_2 , 298 K, 162 MHz) spectrum of $[1\text{-C}_4\text{H}_8][\text{BARF}_4]$.

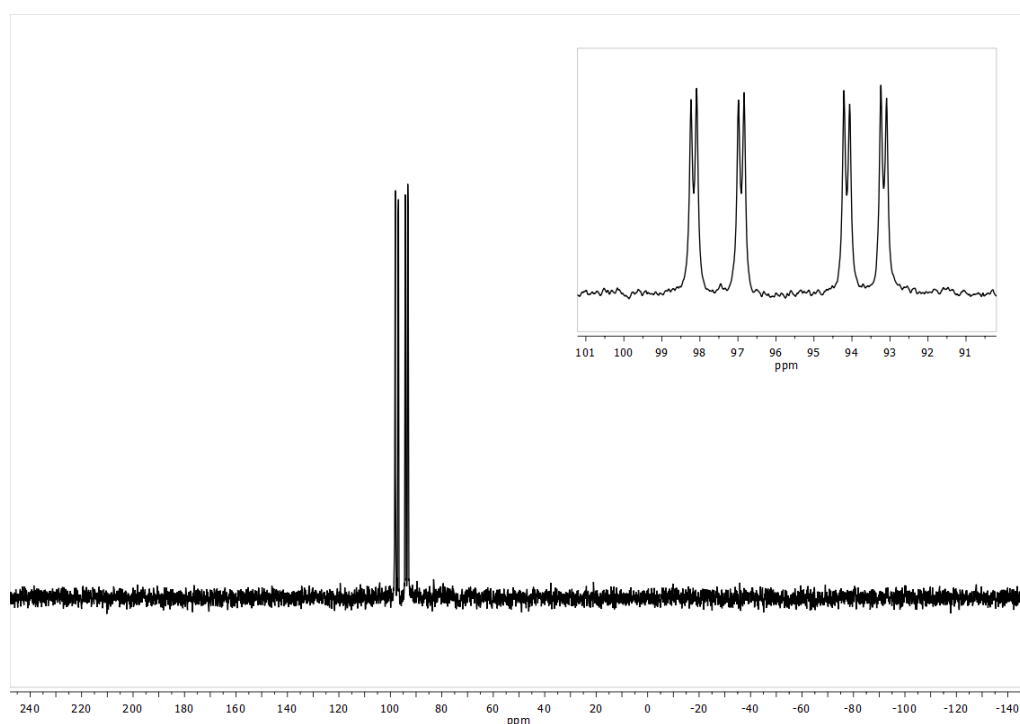


Figure S4: The solution $^{31}\text{P}\{^1\text{H}\}$ NMR (CD_2Cl_2 , 183 K, 162 MHz) spectrum of $[1\text{-C}_4\text{H}_8][\text{BARF}_4]$.

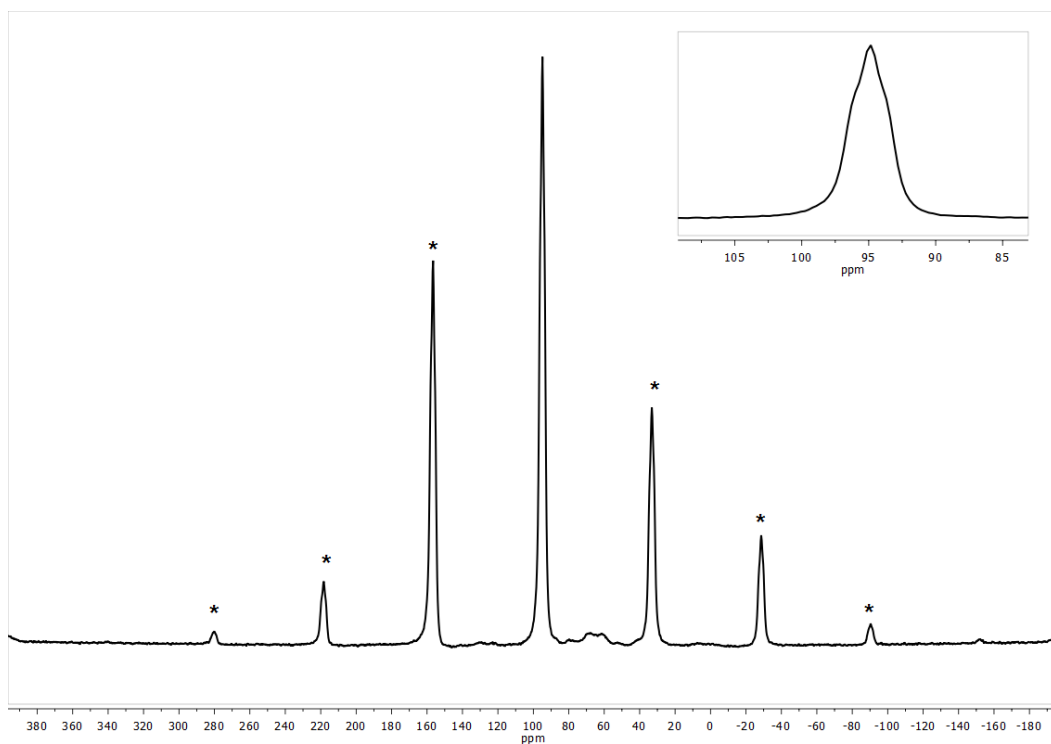


Figure S5: The $^{31}\text{P}\{^1\text{H}\}$ SSNMR (162 MHz, 158 K, 10 kHz spin rate) spectrum of **[1- C_4H_8][BAr^{F}_4]**. The resonances marked * are due to spinning sidebands.

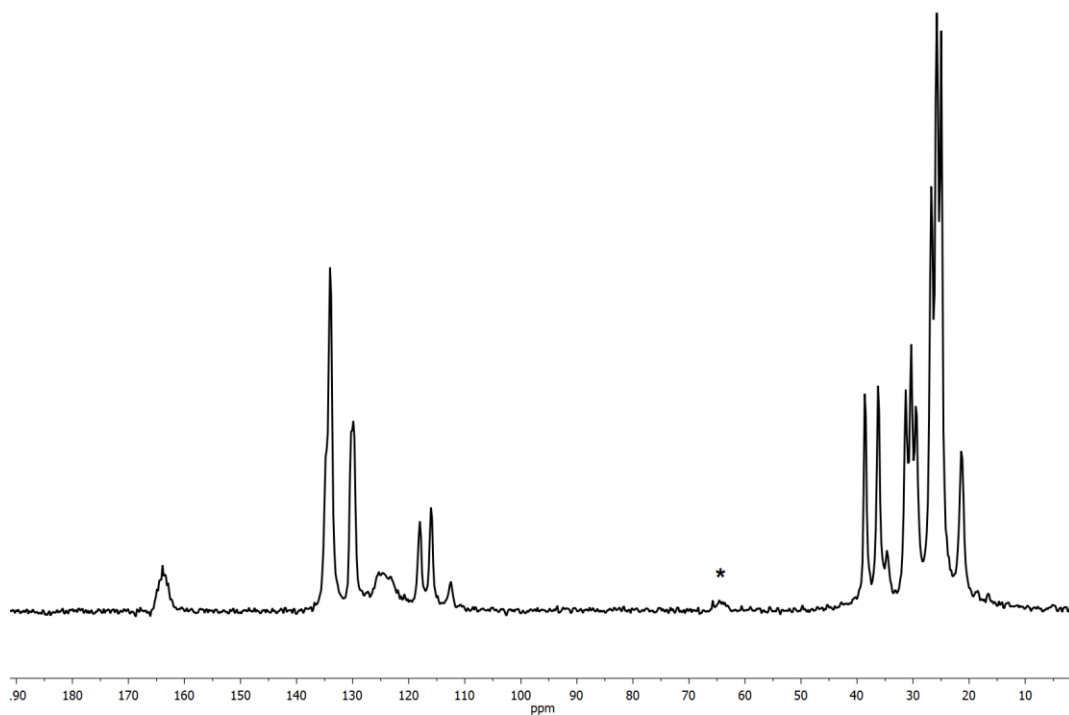


Figure S6: The $^{13}\text{C}\{^1\text{H}\}$ SSNMR (101 MHz, 298 K, 10 kHz spin rate) spectrum of **[1- C_4H_8][BAr^{F}_4]**. The resonances marked * are due to spinning sidebands.

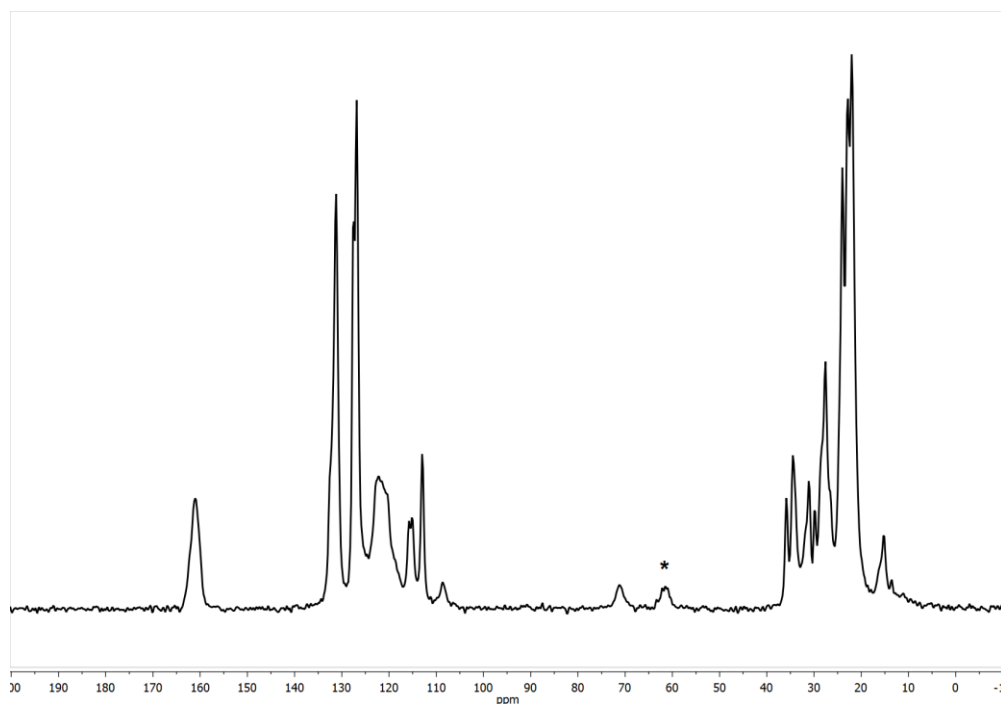


Figure S7: The $^{13}\text{C}\{^1\text{H}\}$ SSNMR (101 MHz, 158 K, 10 kHz spin rate) spectrum of **[1- C_4H_8][BARF_4]**. The resonances marked * are due to spinning sidebands.

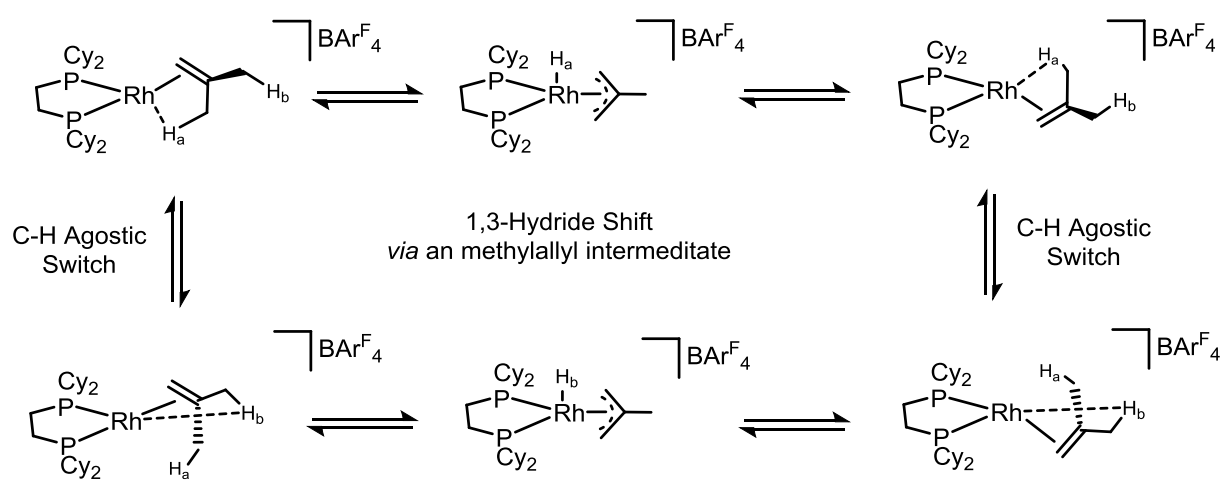
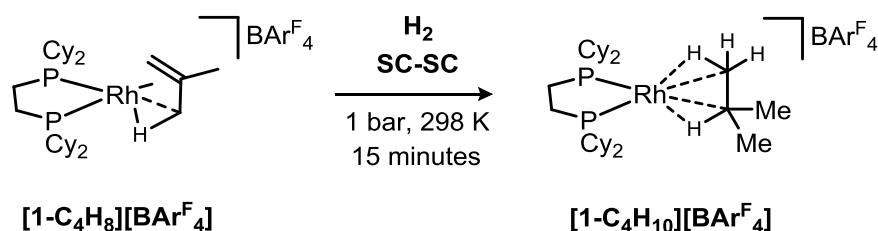


Figure S8: Proposed structural diagrams of the species and mechanism of the fluxional nature of the isobutene ligand of **[1- C_4H_8][BARF_4]**, in both solution and solid-states.

S.2.2. $[\text{Rh}(\text{Cy}_2\text{PCH}_2\text{CH}_2\text{PCy}_2)(\text{C}_4\text{H}_{10})][\text{BAr}^{\text{F}}_4]$, $[\text{1-C}_4\text{H}_{10}][\text{BAr}^{\text{F}}_4]$

S.2.2.1. Synthesis of $[\text{1-C}_4\text{H}_{10}][\text{BAr}^{\text{F}}_4]$ for an x-ray diffraction study



Orange crystals of $[\text{1-C}_4\text{H}_8][\text{BAr}^{\text{F}}_4]$ were treated with H_2 (1 bar, 298 K, 15 mins) inside a J Young flask. The color of the crystals immediately turned a dark red. The crystalline material was then coated with Fomblin® Y oil under an argon-flush; a suitable crystal was rapidly selected and then transferred to the Cryostream of a diffractometer and an x-ray diffraction study was undertaken. For further details of structural refinement please refer to section S.5.

S.2.2.2. Synthesis of $[\text{1-C}_4\text{H}_{10}][\text{BAr}^{\text{F}}_4]$ for SSNMR characterization

A powdered microcrystalline sample of $[\text{1-C}_4\text{H}_8][\text{BAr}^{\text{F}}_4]$ (35 mg) was packed in a 3.2 mm SSNMR rotor, inside an argon filled glove box. The rotor was then placed in a custom built glass J Young flask^{S7} and the sample was then exposed to H_2 (1 bar, 298 K). After 15 minutes, the rotor cap was fitted under a flush of H_2 . The sample was immediately transferred to the bore of a pre-cooled (158 K) SSNMR spectrometer (~ 10 mins) and analyzed $^{31}\text{P}\{^1\text{H}\}$ and $^{13}\text{C}\{^1\text{H}\}$ solid state NMR spectroscopy.

These conditions were optimized to form $[\text{1-C}_4\text{H}_{10}][\text{BAr}^{\text{F}}_4]$. At 158K, no onward reactivity with H_2 is observed. When H_2 replaced by argon at 298K, as so the rotor is packed under an argon atmosphere, dehydrogenation to re-form $[\text{1-C}_4\text{H}_8][\text{BAr}^{\text{F}}_4]$ is observed before cooling to 158K, see section S.4.1.1.

Further exposure to H_2 (90 mins) at 298 K results in complete decomposition to $[\text{1-BAr}^{\text{F}}_4]$, see section S.2.2.6.

S.2.2.3. Characterization data for [1-C₄H₁₀][BAr^F₄]

³¹P{¹H} SSNMR (162 MHz, 298K, 10 kHz spin rate): δ 106.4 (br m).

³¹P{¹H} SSNMR (162 MHz, 158 K, 10 kHz spin rate): δ 106.8 (br, environments overlapping)

¹³C{¹H} SSNMR (101 MHz, 158 K, 10 kHz spin rate): δ 160.5 (ArC), 132.9 (ArC), 127.1 (ArC), 121.6 (br, CF₃), 114.9 (ArC), 112.8 (ArC), 35.0, 32.1, 27.9, 22.8, 17.3, 15.9 (multiple aliphatic resonances).

¹H projection from ¹H/¹³C FSLG HETCOR SSNMR (10 kHz spin rate, 158 K): δ 9.81 (br), 7.12 (br), 0.7 (br), -3.4 (br).

S.2.2.4. Solid State NMR spectra of $[1-C_4H_{10}][BAr^F_4]$

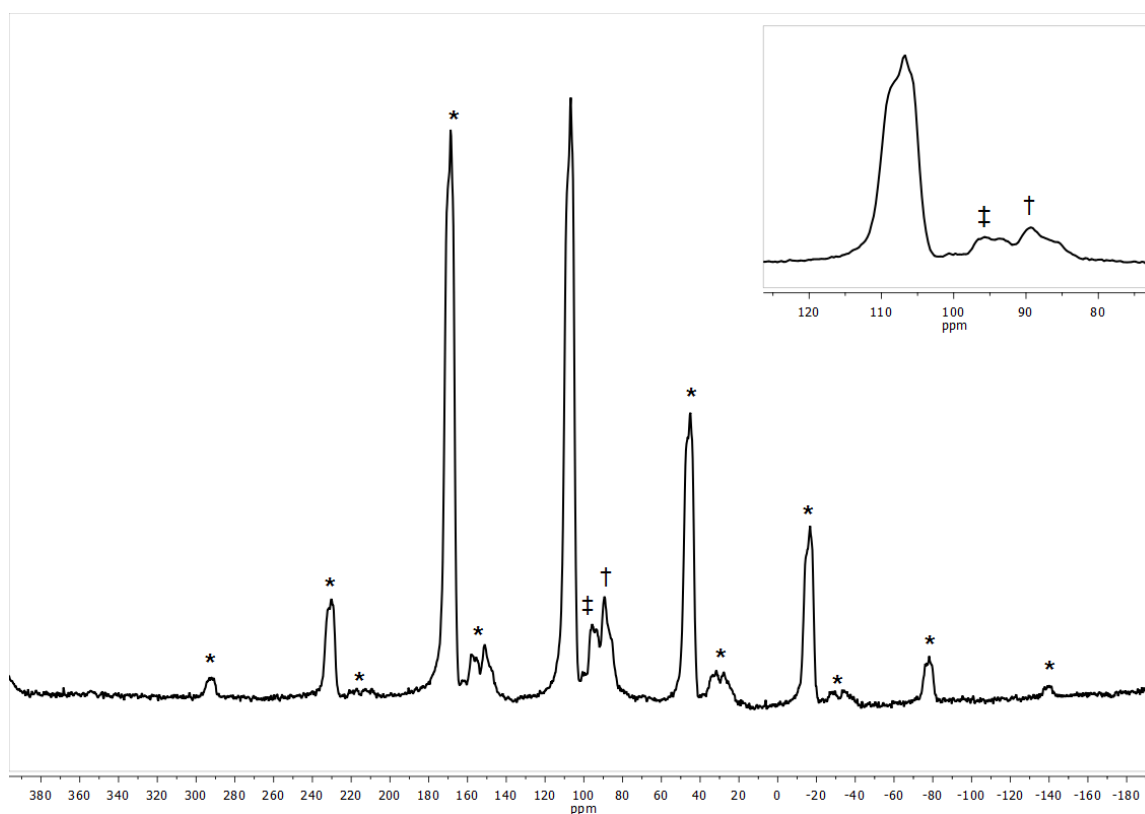


Figure S9: The $^{31}P\{^1H\}$ SSNMR (162 MHz, 158 K, 10 kHz spin rate) spectrum of $[1-C_4H_{10}][BAr^F_4]$ prepared in situ after 15 mins under H_2 at 298K.

The resonances marked ‡ are assigned to $[1-C_6H_8][BAr^F_4]$ from either incomplete hydrogenation or dehydrogenation of $[1-C_4H_{10}][BAr^F_4]$ at 298K. The resonances marked † are assigned to an $[1-BAr^F_4]$ decomposition product under H_2 at 298K. The resonances marked * are due to spinning sidebands. The inset is an enlargement of the central resonances.

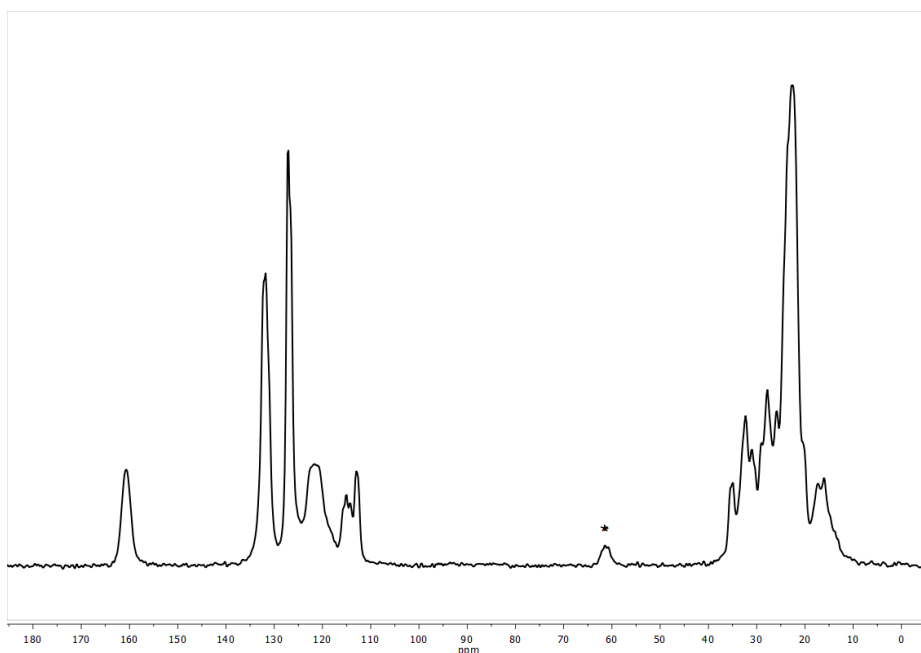


Figure S10: The $^{13}\text{C}\{^1\text{H}\}$ SSNMR (100 MHz, 158 K, under H_2 , 10 kHz spin rate) spectrum of $[\text{1-C}_4\text{H}_{10}][\text{BARF}_4]$ prepared in situ at time of measuring. The resonance marked * is due to a spinning sideband.

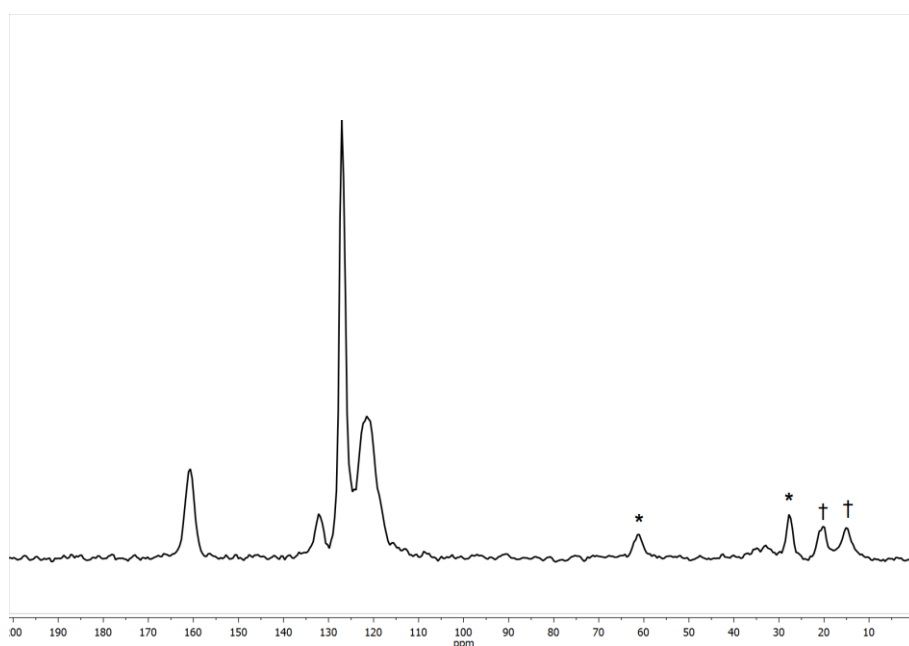


Figure S11: The $^{13}\text{C}\{^1\text{H}\}$ NQS SSNMR (100 MHz, 203 K, 10 kHz spin rate) spectrum of $[\text{1-C}_4\text{H}_{10}][\text{BARF}_4]$ prepared in situ. The resonances marked * are due to spinning sidebands. Peaks marked † at $\delta \sim 21$ and $\delta \sim 15$ are assigned to the C_4H_{10} fragment undergoing motion at 203 K.

Note: When $^{13}\text{C}\{^1\text{H}\}$ NQS SSNMR was ran at 158 K; the peaks marked † are absent, suggesting freezing out of any motion.

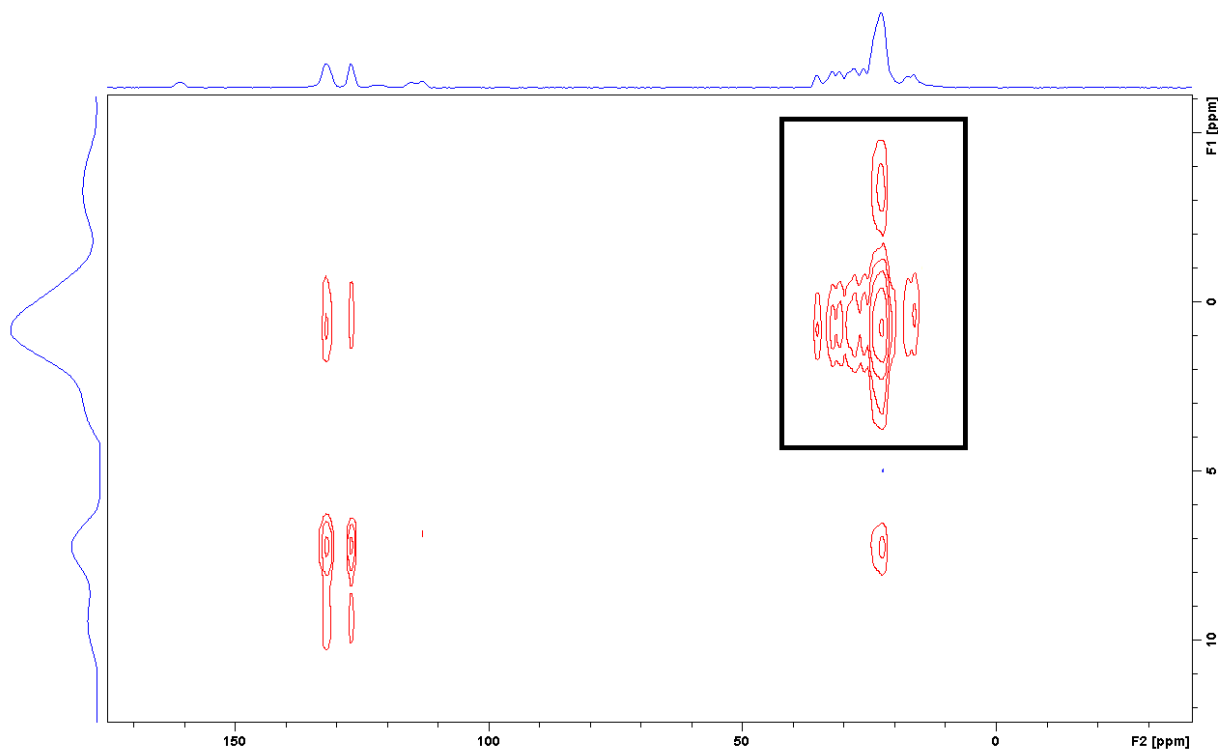


Figure S12: The $^1\text{H}/^{13}\text{C}$ FSLG HETCOR SSNMR (158 K, 10 kHz spin rate) full spectrum of $[1\text{-C}_4\text{H}_{10}][\text{BAr}^{\text{F}}_4]$. Enlargement of the alkyl region, shown by the black box, is shown below in Figure S13.

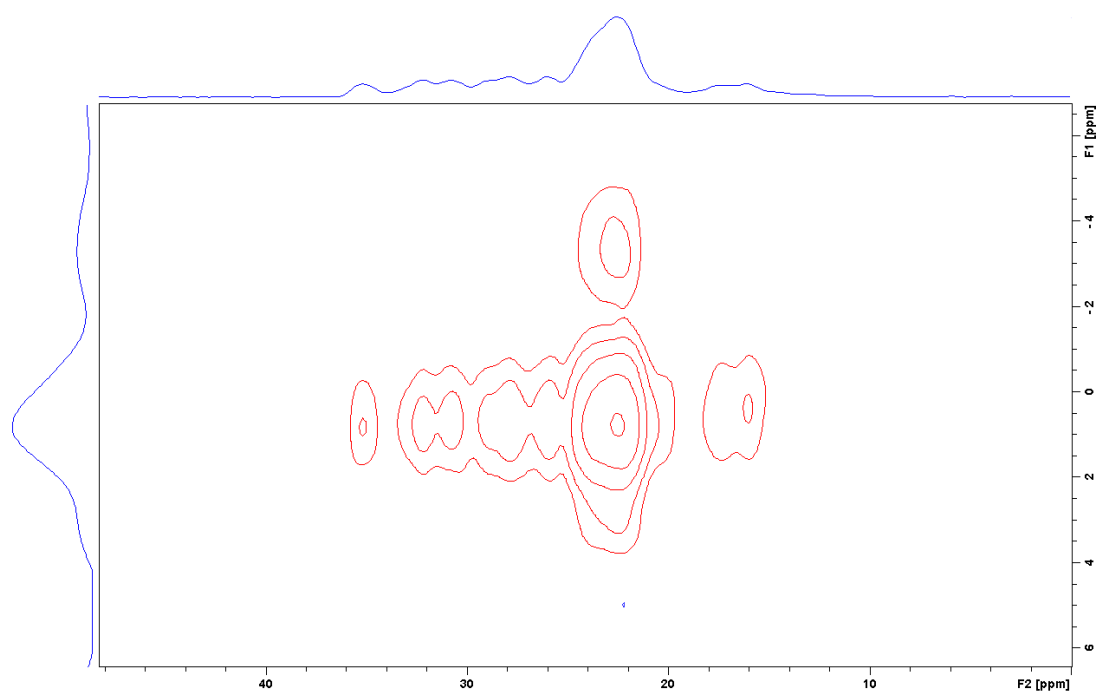


Figure S13: Enlargement of the $^1\text{H}/^{13}\text{C}$ FSLG HETCOR SSNMR (158 K, 10 kHz spin rate) spectrum of the alkyl region of $[1\text{-C}_4\text{H}_{10}][\text{BAr}^{\text{F}}_4]$.

S.2.2.5. Liberation of the bound isobutane from [1-C₄H₁₀][BAr^F₄].

Orange crystals of [1-C₄H₈][BAr^F₄] (20 mgs) were treated with H₂ (1 bar, 298 K, 15 mins) inside a J Young flask. The color of the crystals immediately turned a dark red, yielding [1-C₄H₁₀][BAr^F₄]. The solid was then cooled to 77 K under liquid nitrogen and the J Young flask placed under vacuum to remove any H₂. CO (1 bar, 298K) was added and solid left for 1 hour; to which a vibrant yellow solid material had formed. The volatiles excluded from the lattice could be distilled into a fresh NMR tube with d₃-MeCN and analyzed by ¹H NMR spectroscopy.

¹H NMR (d₃-MeCN, 400 MHz): δ 1.72 (decet, 1H, CH(CH₃)₃, J_{H-H} = 6.79 Hz), 0.89 (doublet, 9H, CH(CH₃)₃, J_{H-H} = 6.79 Hz).

The resultant solid was confirmed to be [Rh(dcpe)(CO)₂][BAr^F₄] by solution ³¹P{¹H} NMR spectroscopy (δ 85.22, d, J_{RhP} = 116 Hz).^{S7}

S.2.2.6. Solid state decomposition products of $[1\text{-C}_4\text{H}_{10}][\text{BAr}^{\text{F}}_4]$ under H_2

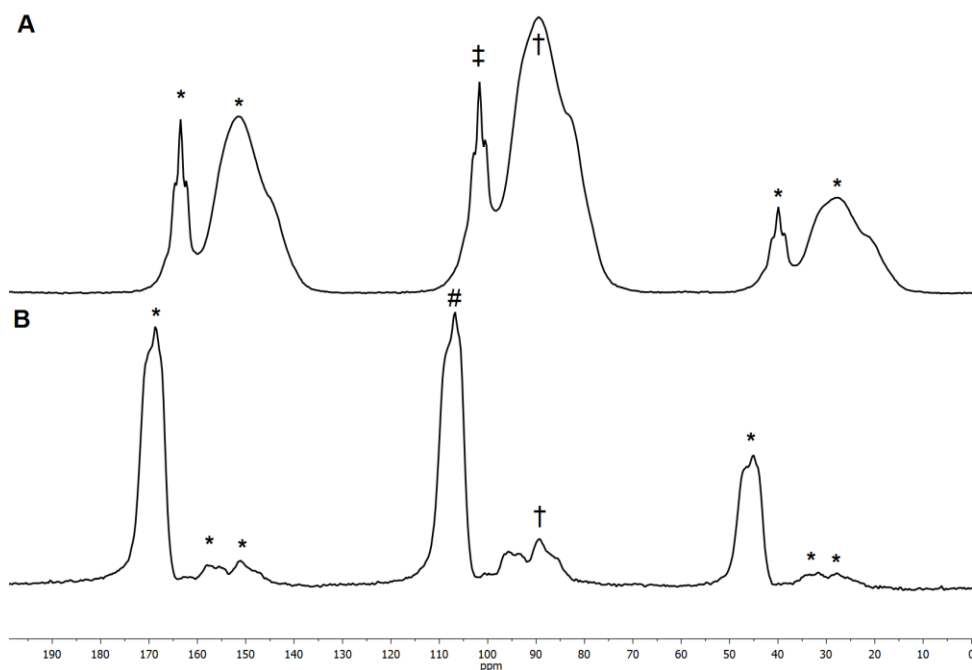
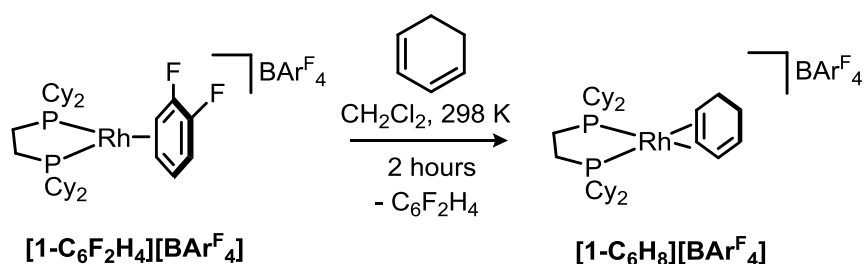


Figure S14: (A) The $^{31}\text{P}\{^1\text{H}\}$ SSNMR (162 MHz, 158 K, 10 kHz spin rate) spectrum of $[1\text{-C}_4\text{H}_{10}][\text{BAr}^{\text{F}}_4]$ left in situ under an H_2 atmosphere at 298K for 90 mins. The peak assigned to † is the main decomposition product under H_2 , $[\text{1-BAr}^{\text{F}}_4]^{\text{S}2}$ (approx. 70 %) and ‡ is an unidentified complex under H_2 (approx. 30 %).

(B) Repeat of Figure S9, showing the $^{31}\text{P}\{^1\text{H}\}$ SSNMR (162 MHz, 158 K, 10 kHz spin rate) spectrum, where # = $[1\text{-C}_4\text{H}_{10}][\text{BAr}^{\text{F}}_4]$, to show no signals in (A) relating to $[1\text{-C}_4\text{H}_{10}][\text{BAr}^{\text{F}}_4]$ are present after hydrogenation for 90 mins.

S.2.3. $[\text{Rh}(\text{Cy}_2\text{PCH}_2\text{CH}_2\text{PCy}_2)(\text{C}_6\text{H}_8)][\text{BAr}^{\text{F}}_4]$, $[\text{1-C}_6\text{H}_8][\text{BAr}^{\text{F}}_4]$

S.2.3.1. Synthesis of $[\text{1-C}_6\text{H}_8][\text{BAr}^{\text{F}}_4]$



A solution of $[\text{1-C}_6\text{H}_4\text{F}_2][\text{BAr}^{\text{F}}_4]$ (300 mg, 0.20 mmol) in CH_2Cl_2 (20 mL) was treated with 1,3-cyclohexadiene (200 μL) at ambient temperature. The resultant deep red solution was stirred at ambient temperature for 2 hours. Pentane (100 mL) was then added with vigorous stirring which resulted in the formation of a red precipitate. The solid was isolated by filtration, washed with pentane (2 \times 20 mL) and then dried under vacuum. The solid was then dissolved in CH_2Cl_2 (~ 8 mL) and filtered. The filtrate was then layered with pentane. Deep red crystals were obtained upon storage at 4 $^\circ\text{C}$ after 1 week. Yield: 204 mg (70 %).

Note: $[\text{1-C}_6\text{H}_8][\text{BAr}^{\text{F}}_4]$ displays poor stability in solution at room temperature, particularly at high concentrations. $[\text{1-C}_6\text{H}_6][\text{BAr}^{\text{F}}_4]$ was identified as the major decomposition product (~ 50 %) as well as several other, as yet, unidentified species are also present. These are assumed to be the products of C-Cl activation from the solvent, as identified by ESI-MS.

S.2.3.2. Characterization data for [1-C₆H₈][BAR^F₄]

¹H NMR (CD₂Cl₂, 298 K, 400 MHz): δ 7.73 (s, 8H, ortho-ArH), 7.57 (s, 4H, para-ArH), 5.36-5.32 (br m, 5H, alkene CH and CDHCl₂), 2.16 (br s, 2H, aliphatic CH), 1.94-1.70 (br m, 28H, overlapping aliphatic CH), 1.61-1.58 (br m, 2H, aliphatic CH), 1.38-1.08 (br m, 16H, overlapping aliphatic CH), 0.99-0.96 (br m, 4H, overlapping aliphatic CH).

¹H NMR (CD₂Cl₂, 183 K, 400 MHz): δ 7.75 (s, 8H, ortho-ArH), 7.56 (s, 4H, para-ArH), 5.30-5.23 (br m, 4H, alkene), 2.11 (br s, 2H, aliphatic CH), 1.89-1.42 (br m, 30H, overlapping aliphatic CH), 1.31-0.74 (br m, 20H, overlapping aliphatic CH).

³¹P{¹H} NMR (CD₂Cl₂, 298 K, 162 MHz): δ 78.7 (d, J_{RhP} 172 Hz).

³¹P{¹H} NMR (CD₂Cl₂, 183 K, 162 MHz): δ 78.6 (d, J_{RhP} 172 Hz).

¹⁹F{¹H} NMR (CD₂Cl₂, 298 K, 377 MHz): δ -62.9 (s).

¹³C{¹H} NMR (CD₂Cl₂, 298 K, 126 MHz): δ 162.2 (q, J_{CB} 50 Hz, ipso-ArC), 135.3 (s, ortho-ArC), 129.3 (q, J_{CF} 32 Hz, meta-ArC), 125.1 (q, J_{CF} 272 Hz, CF₃), 117.9 (s, para-ArC), 95.0 (d, J_{CRh} 4.4 Hz, alkene-CH), 82.3 (m, alkene-CH), 37.9 (m, cyclohexyl-CH), 31.3 (s, cyclohexadiene-CH₂), 29.6 (m, cyclohexyl-CH₂), 29.1 (s, cyclohexyl-CH₂), 26.3 (m, PCH₂), 22.5 (m, cyclohexyl-CH₂), 21.1 (s, cyclohexyl-CH₂).

³¹P{¹H} SSNMR (162 MHz, 294 K, 10 kHz spin rate): δ 78.6 (br m).

¹³C{¹H} SSNMR (101 MHz, 294 K, 10 kHz spin rate): δ 163.9 (ipso-ArC), 133.9 (ortho-ArC), 130.0 (meta-ArC), 124.7 (br, CF₃), 117.8 (para-ArC), 116.0 (para-ArC), 95.8 (C=C), 94.1 (C=C), 83.3 (C=C), 80.7 (C=C), 37.7 (CH), 36.0 (CH), 31.4 (CH₂), 30.5 (CH₂), 29.5 (CH₂), 26.6 (CH₂), 25.3 (CH₂), 22.0 (CH₂), 21.1 (CH₂), 19.7 (CH₂), 18.7 (CH₂).

¹H projection from ¹H/¹³C FSLG HETCOR SSNMR (10 kHz spin rate, 294 K): δ 7.1 (br), 3.9 (br), 0.5 (br).

ESI-MS found (calc. for [Rh(Cy₂PCH₂CH₂PCy₂)(C₆H₈)]⁺): m/z 605.29 (605.29).

Elemental analysis found (calc. for C₆₄H₆₈BF₂₄P₂Rh): C 52.32 (52.33), H 4.52 (4.67).

S.2.3.3. NMR spectra of [1-C₆H₈][BAr^F₄]

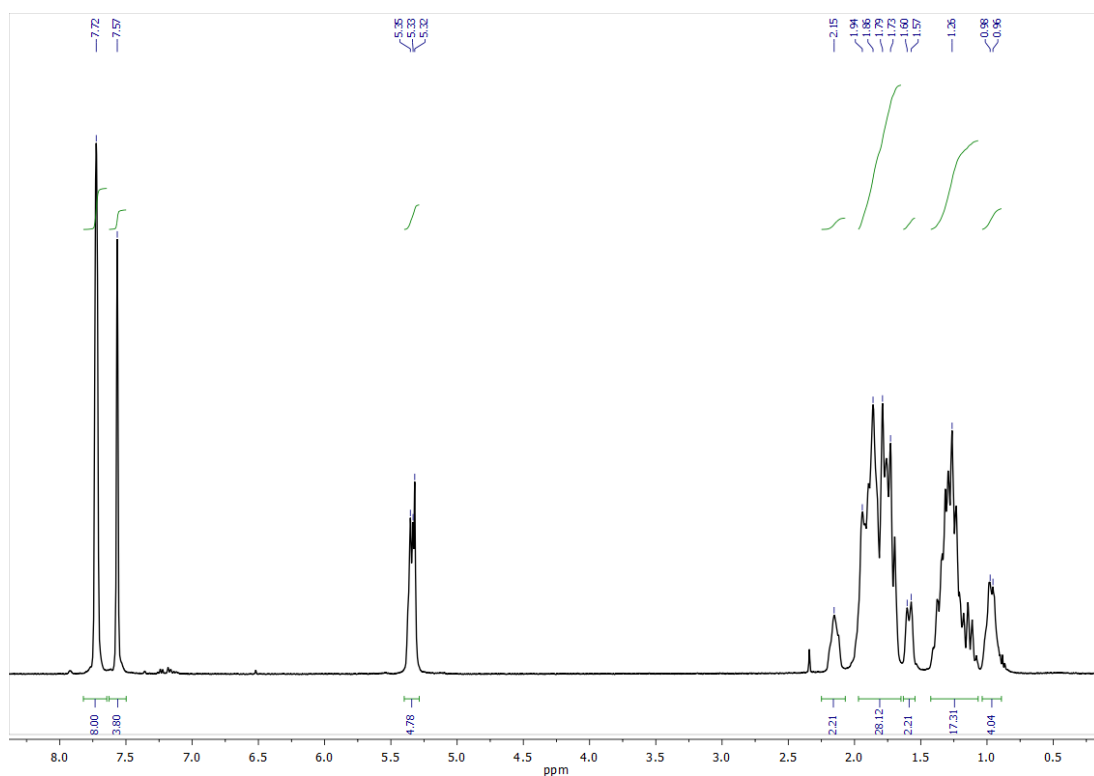


Figure S15: The solution ¹H NMR (CD₂Cl₂, 298 K, 400 MHz) spectrum of [1-C₆H₈][BAr^F₄].

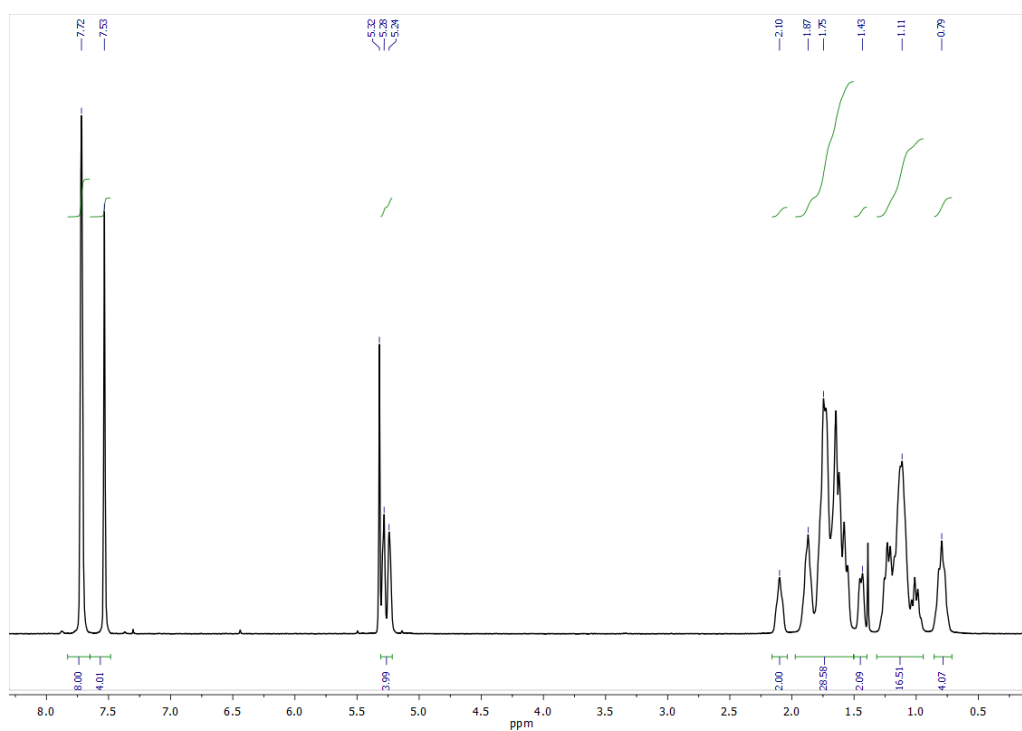


Figure S16: The solution ¹H NMR (CD₂Cl₂, 183 K, 400 MHz) spectrum of [1-C₆H₈][BAr^F₄].

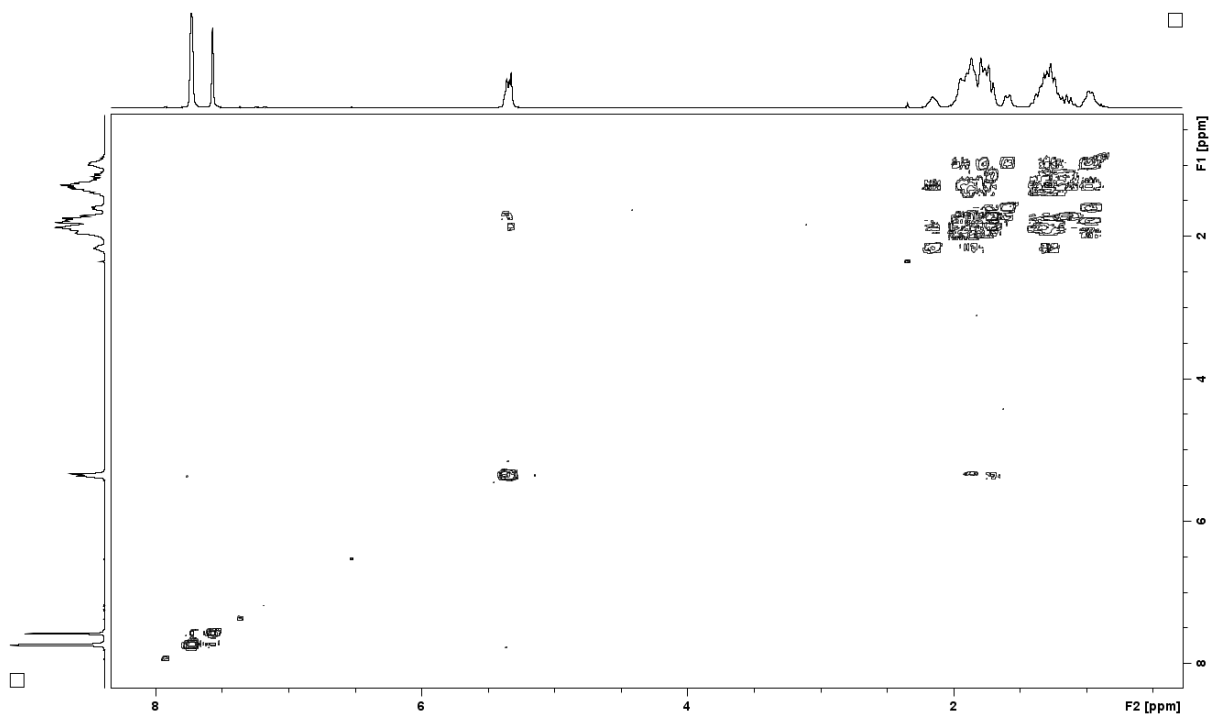


Figure S17: The solution $^1\text{H}/^1\text{H}$ COSY NMR (CD_2Cl_2 , 298 K, 400 MHz) spectrum of $[1\text{-C}_6\text{H}_8][\text{BARF}_4]$.

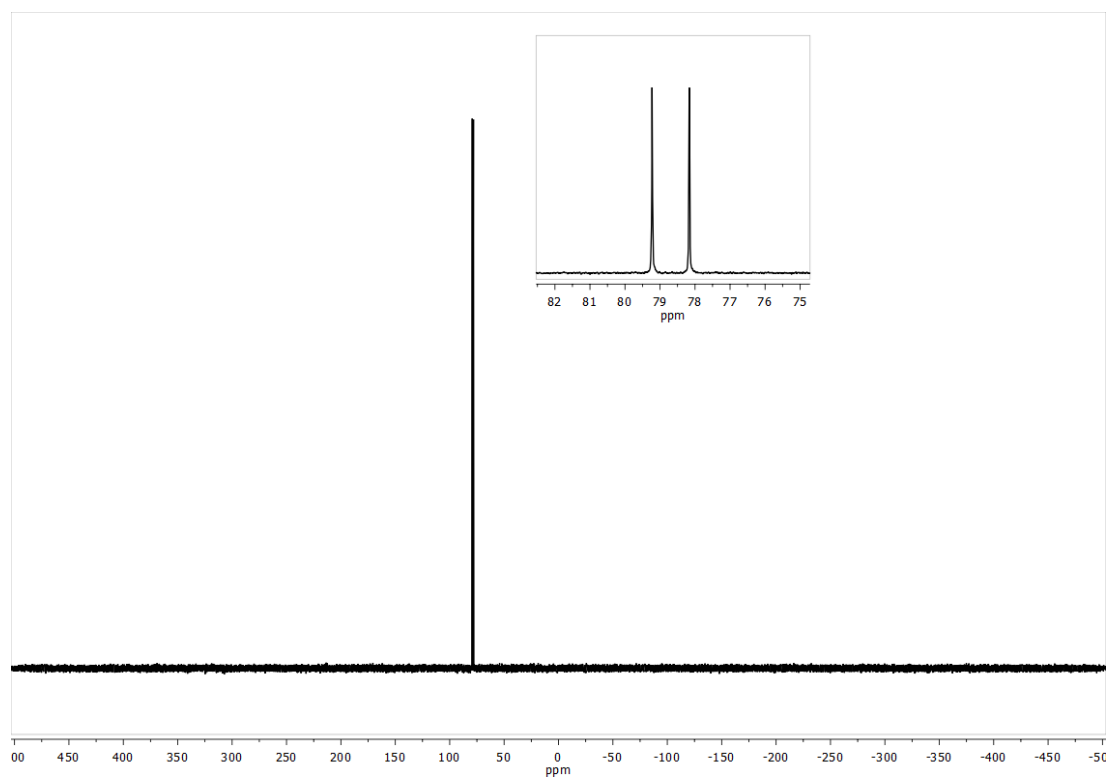


Figure S18: The solution $^{31}\text{P}\{^1\text{H}\}$ NMR (CD_2Cl_2 , 298 K, 162 MHz) spectrum of $[1\text{-C}_6\text{H}_8][\text{BARF}_4]$.

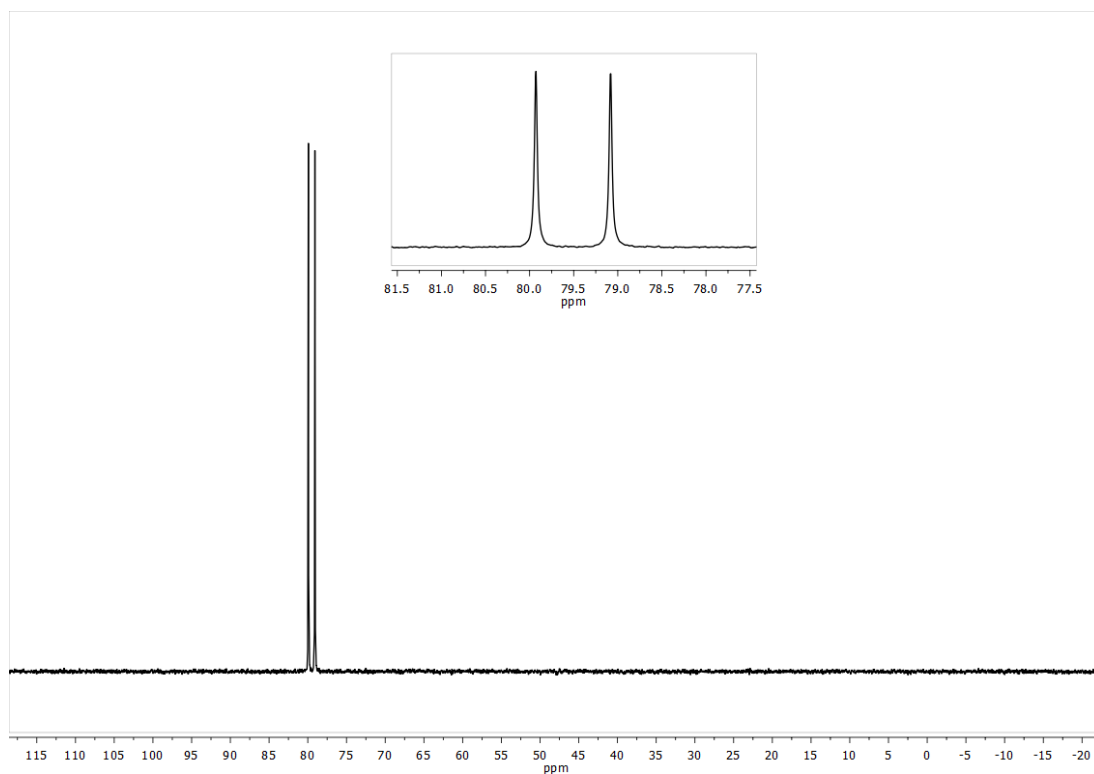


Figure S19: The solution $^{31}\text{P}\{^1\text{H}\}$ NMR (CD_2Cl_2 , 183 K, 162 MHz) spectrum of **[1- C_6H_8][BARF_4]**.

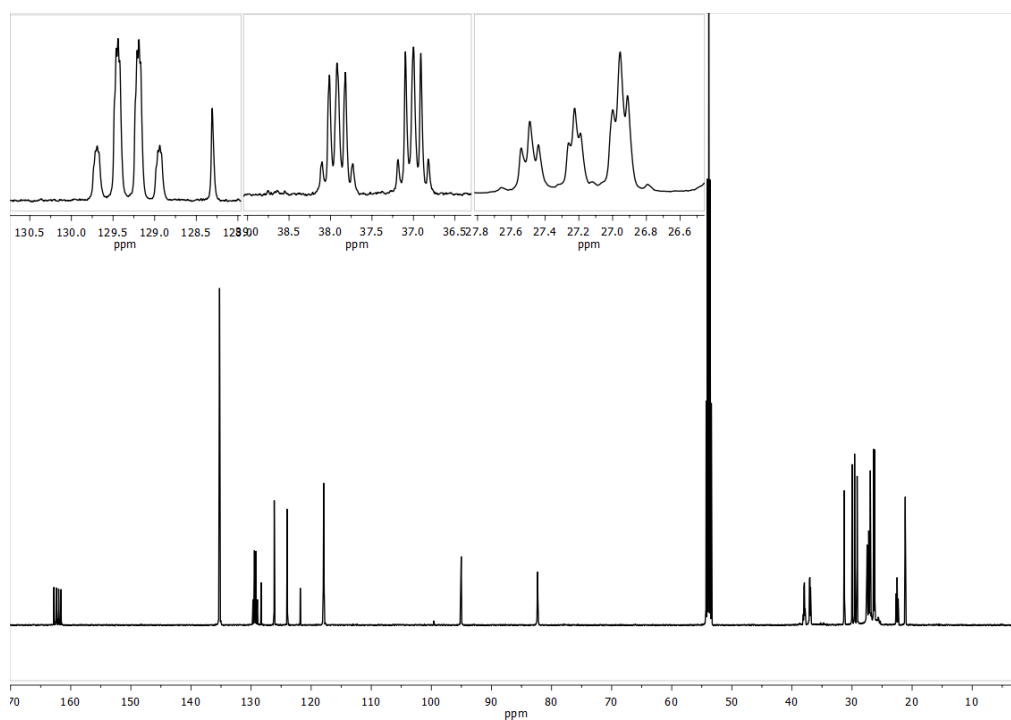


Figure S20: The solution $^{13}\text{C}\{^1\text{H}\}$ NMR (CD_2Cl_2 , 298 K, 126 MHz) spectrum of **[1- C_6H_8][BARF_4]**.

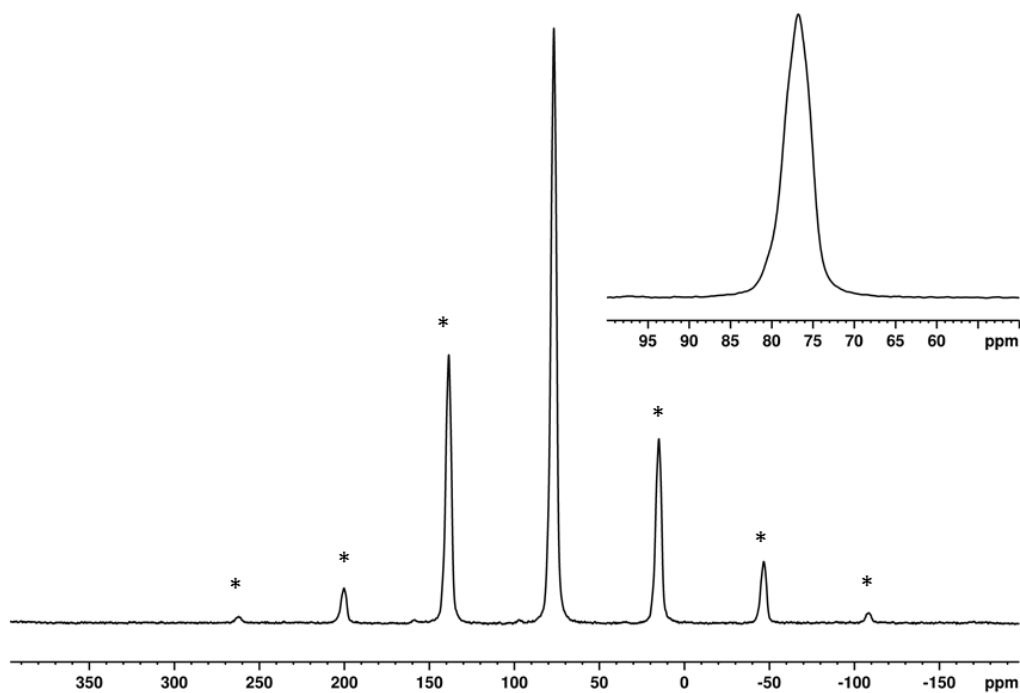


Figure S21: The $^{31}\text{P}\{^1\text{H}\}$ SSNMR (162 MHz, 294 K, 10 kHz spin rate) spectrum of $[\text{1-C}_6\text{H}_8][\text{BAR}^{\text{F}}_4]$. The resonances marked * are due to spinning sidebands. The inset is an enlargement of the central resonance.

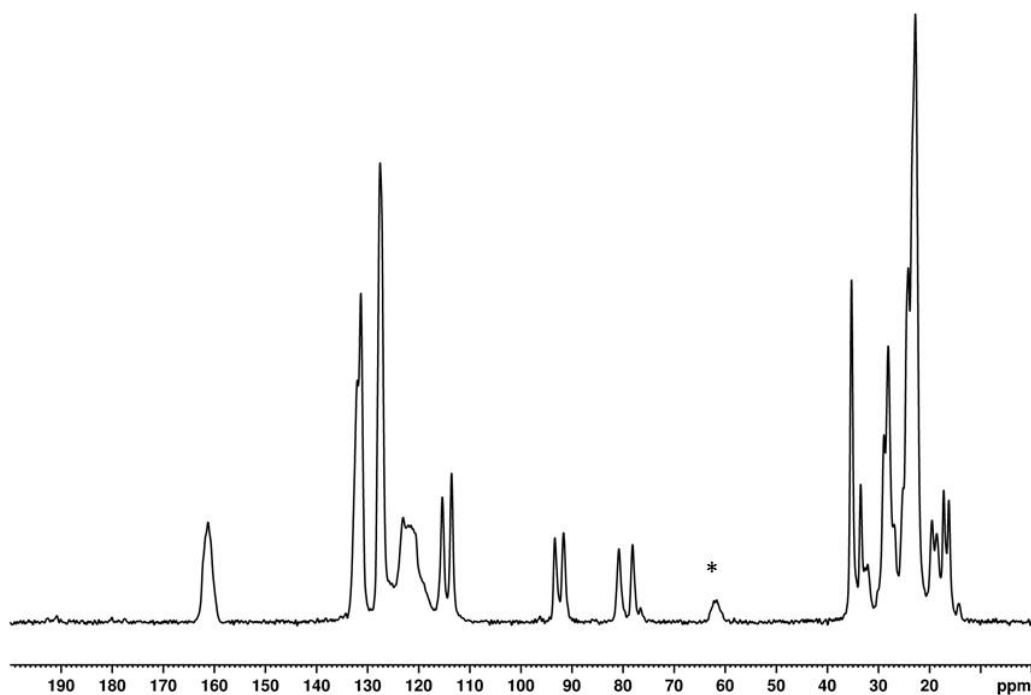
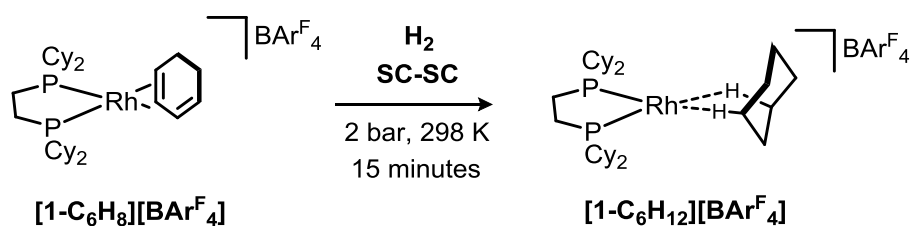


Figure S22: The $^{13}\text{C}\{^1\text{H}\}$ SSNMR (101 MHz, 294 K, 10 kHz spin rate) spectrum of $[\text{1-C}_6\text{H}_8][\text{BAR}^{\text{F}}_4]$. The resonance marked * is due to spinning sidebands.

S.2.4. $[\text{Rh}(\text{Cy}_2\text{PCH}_2\text{CH}_2\text{PCy}_2)(\text{C}_6\text{H}_{12})][\text{BAr}^{\text{F}}_4]$, $[\text{1-C}_6\text{H}_{12}][\text{BAr}^{\text{F}}_4]$

S.2.4.1. Synthesis of $[\text{1-C}_6\text{H}_{12}][\text{BAr}^{\text{F}}_4]$ for an x-ray diffraction study



Deep red crystals of $[\text{1-C}_6\text{H}_8][\text{BAr}^{\text{F}}_4]$ were treated with H_2 (2 bar, 298 K, 15 mins) inside a J Young flask. The colour of the crystals immediately darkens to a burgundy color. The crystalline material was then coated with Fomblin® Y oil under an argon flush, a suitable crystal was rapidly selected and then transferred to the Cryostream of a diffractometer and an x-ray diffraction study was undertaken. For further of the structural study, please refer to section S.5.

After 90 minutes under hydrogen, approx. 10 % of the sample has decomposed. This was shown by $^{31}\text{P}\{^1\text{H}\}$ SSNMR, see section S.2.4.6, figure S34.

S.2.4.2. Synthesis of [1-C₆H₁₂][BAr^F₄] for SSNMR characterization

A powdered microcrystalline sample of [1-C₆H₈][BAr^F₄] (35 mg) was packed in a 3.2 mm SSNMR rotor, inside an argon filled glove box. The rotor was then placed in a custom built glass J Young flask^{S7} and the sample was then exposed to H₂ (1 bar, 298 K). After 20 minutes, the rotor cap was fitted under a flush of H₂. The sample was immediately transferred to the bore of a pre-cooled (158 K) SSNMR spectrometer (~ 10 mins) and analyzed ³¹P{¹H} and ¹³C{¹H} solid state NMR spectroscopy.

These conditions were optimized to form [1-C₆H₁₂][BAr^F₄]. At 158K, no onward reactivity with H₂ is observed. When H₂ replaced by argon at 298K, as so the rotor is packed under an argon atmosphere, spontaneous dehydrogenation is observed before cooling to 158K, to form [1-C₆H₁₀][BAr^F₄], see section S.4.4.1.

S.2.4.3. Characterization data for [1-C₆H₁₂][BAr^F₄]

³¹P{¹H} SSNMR (162 MHz, 294 K, 10 kHz spin rate): δ 108.4 (two overlapping doublets, J_{RhP} = ~ 220 Hz).

³¹P{¹H} SSNMR (162 MHz, 158 K, 10 kHz spin rate): δ 107.1 (br), 104.5 (br).

¹³C{¹H} SSNMR (101 MHz, 198 K, 10 kHz spin rate): δ 161.1 (ArC), 131.8 (ArC), 127.2 (ArC), 121.8 (br, CF₃), 115.2 (ArC), 112.7 (ArC), 34.4, 32.4, 28.9, 27.8, 25.8, 24.2, 22.8, 22.3, 21.6, 17.2 (multiple aliphatic resonances).

¹³C{¹H} SSNMR (101 MHz, 158 K, 10 kHz spin rate): δ 161.1 (ArC), 131.8 (ArC), 127.1 (ArC), 121.6 (br, CF₃), 115.1 (ArC), 112.5 (ArC), 34.0, 32.1, 27.8, 24.1, 22.7, 19.7, 16.5 (multiple aliphatic resonances).

¹H projection from ¹H/¹³C FSLG HETCOR SSNMR (198 K, 10 kHz spin rate): δ 7.1 (br), 1.0 (br).

¹H projection from ¹H/¹³C FSLG HETCOR SSNMR (158 K, 10 kHz spin rate): δ 7.1 (br), 1.0 (br), -1.6 (br).

S.2.4.4. NMR spectra of $[1\text{-C}_6\text{H}_{12}][\text{BAr}^{\text{F}}_4]$

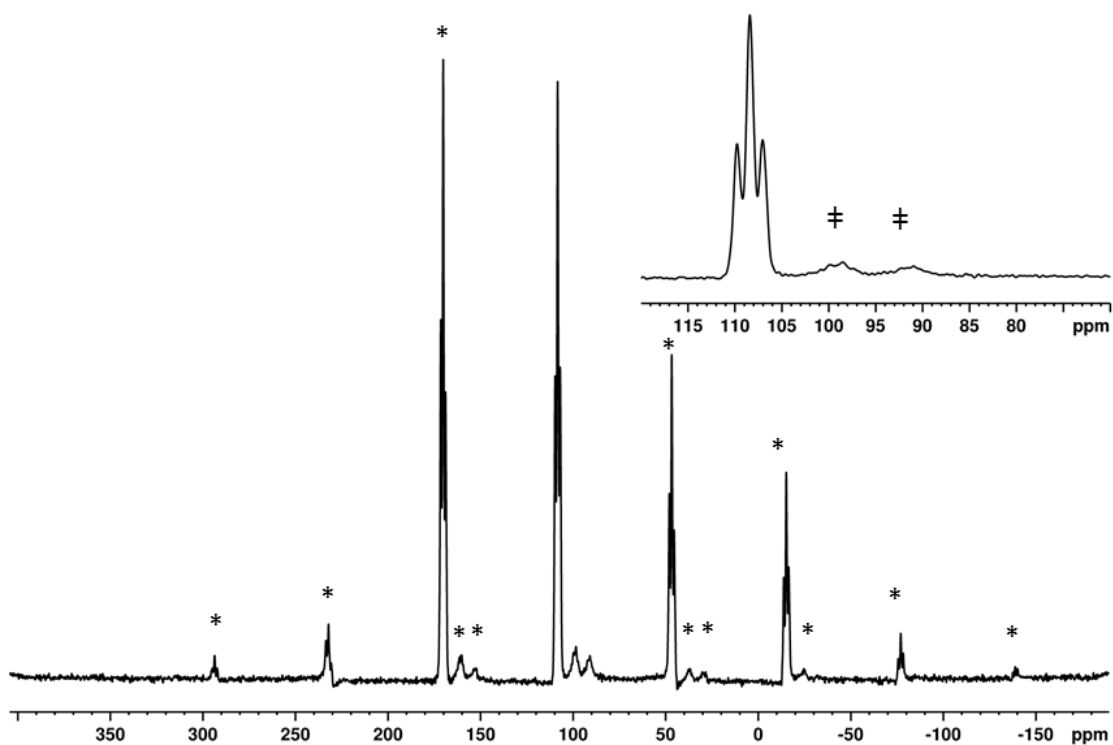


Figure S23: The $^{31}\text{P}\{^1\text{H}\}$ SSNMR (162 MHz, 298 K, 10 kHz spin rate) spectrum of $[1\text{-C}_6\text{H}_{12}][\text{BAr}^{\text{F}}_4]$ prepared in situ, packed under atmosphere of H_2 . The resonances marked ± are assigned to $[1\text{-C}_6\text{H}_{10}][\text{BAr}^{\text{F}}_4]$ which has formed by partial dehydrogenation, see section S.4.3. The resonances marked * are due to spinning sidebands. The inset is an enlargement of the central resonances.

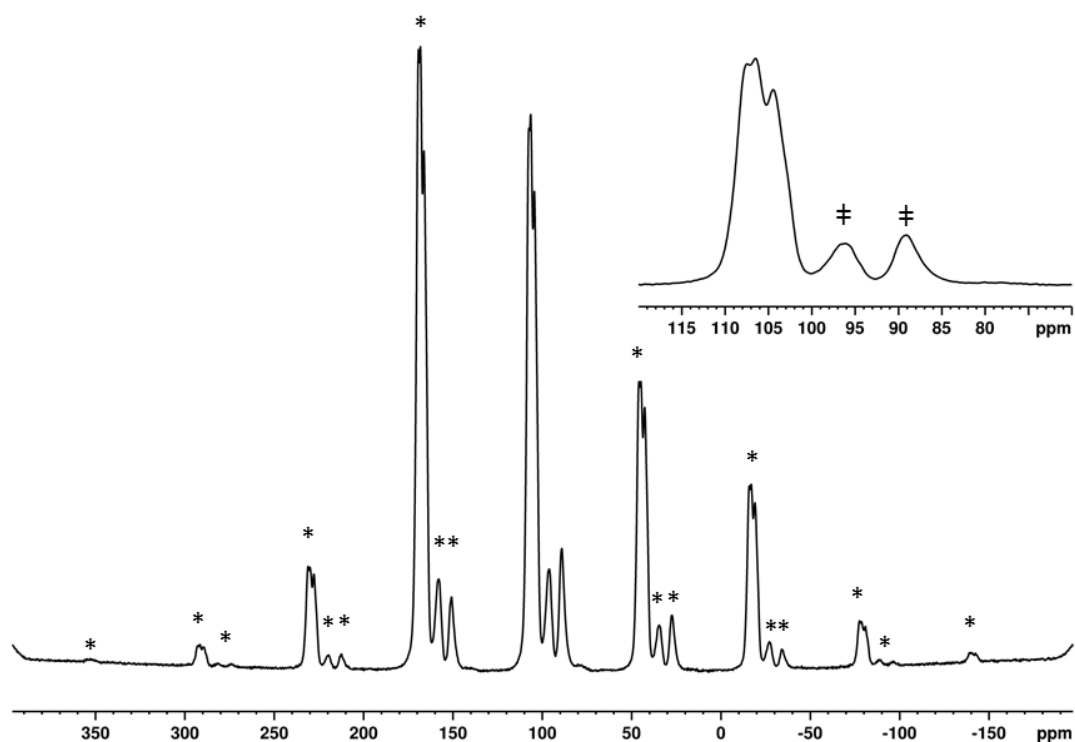


Figure S24: The $^{31}\text{P}\{^1\text{H}\}$ SSNMR (162 MHz, 158 K, 10 kHz spin rate) spectrum of $[\text{1-C}_6\text{H}_{12}][\text{BAR}^{\text{F}}_4]$ prepared in situ. The resonances marked $\oplus\ominus$ are assigned to $[\text{1-C}_6\text{H}_{10}][\text{BAR}^{\text{F}}_4]$ which has formed by partial dehydrogenation. The resonances marked * are due to spinning sidebands. The inset is an enlargement of the central resonances. This spectrum was obtained from a different sample to that above, resulting in a different degree of dehydrogenation.

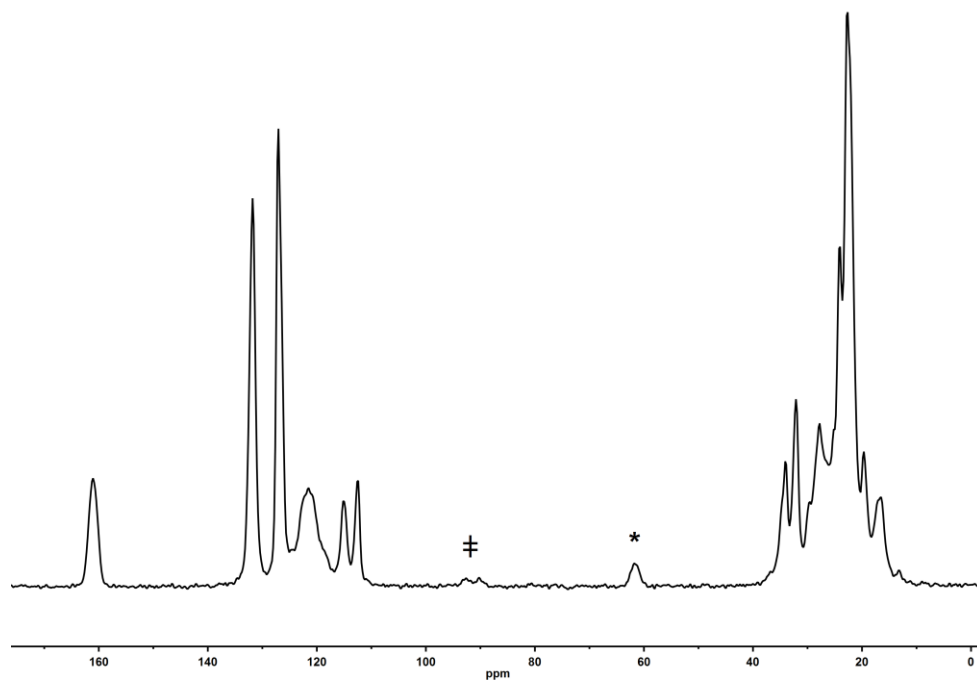


Figure S25: The $^{13}\text{C}\{^1\text{H}\}$ SSNMR (100 MHz, 158 K, 10 kHz spin rate) spectrum of $[\text{1-C}_6\text{H}_{12}][\text{BAR}^{\text{F}}_4]$ prepared in situ. The resonances marked \oplus are assigned to $[\text{1-C}_6\text{H}_{10}][\text{BAR}^{\text{F}}_4]$ which formed by partial dehydrogenation. The resonance marked * is due to a spinning sideband.

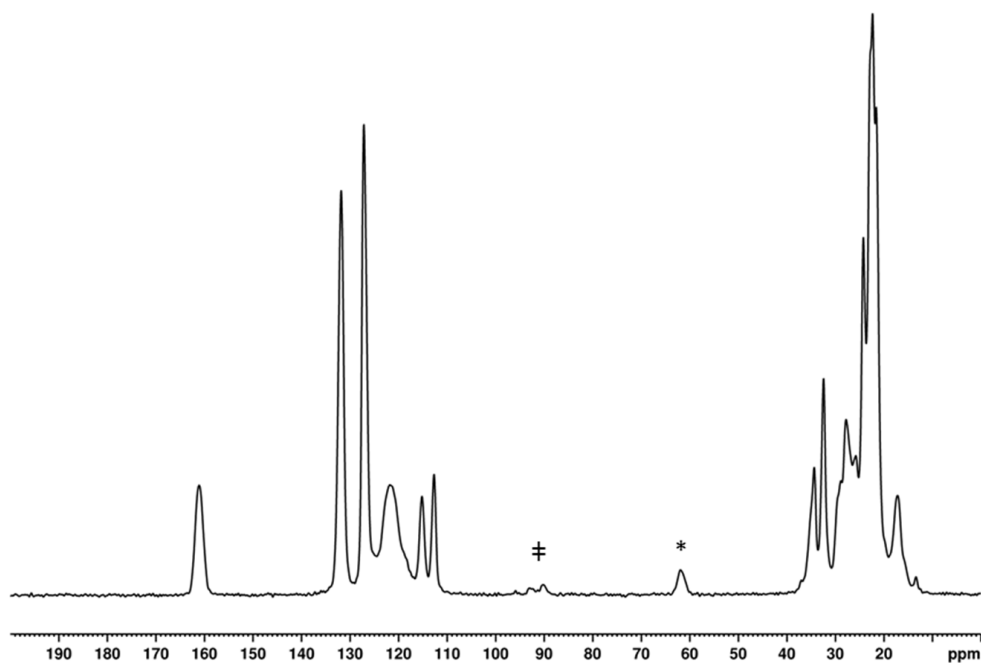


Figure S26: The $^{13}\text{C}\{^1\text{H}\}$ SSNMR (100 MHz, 198 K, 10 kHz spin rate) spectrum of $[\text{1-C}_6\text{H}_{12}][\text{BAR}^{\text{F}}_4]$ prepared in situ. The resonances marked \oplus are assigned to $[\text{1-C}_6\text{H}_{10}][\text{BAR}^{\text{F}}_4]$, which formed by partial dehydrogenation. The resonance marked * is due to a spinning sideband.

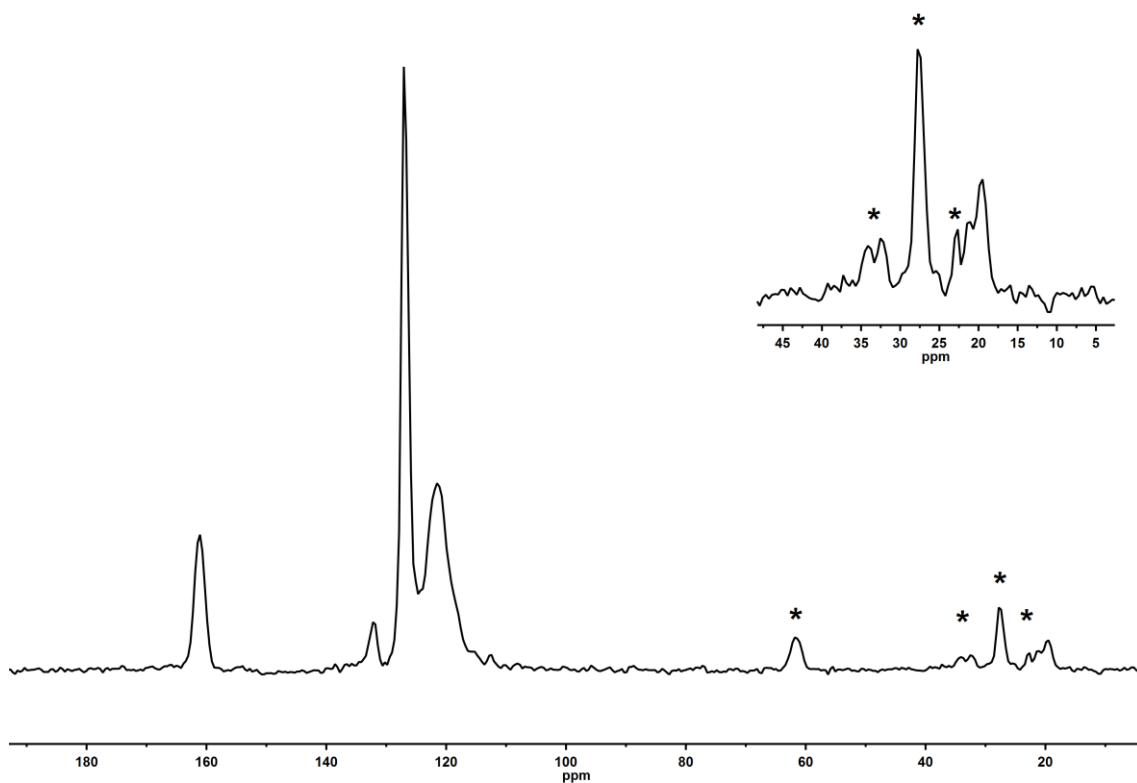


Figure S27: The $^{13}\text{C}\{^1\text{H}\}$ NQS SSNMR (100 MHz, 158 K, 10 kHz spin rate) spectrum of $[1\text{-C}_6\text{H}_{12}][\text{BArF}_4]$ prepared in situ. The resonances marked * are due to spinning sidebands.

The inset is an enlargement of the resonances between δ 45 to 5, showing two peaks at δ 19.7 and 21.4. These peaks did not shift position in the analogous 8 kHz NQS SSNMR spectrum, suggestive of real signals and not sidebands.

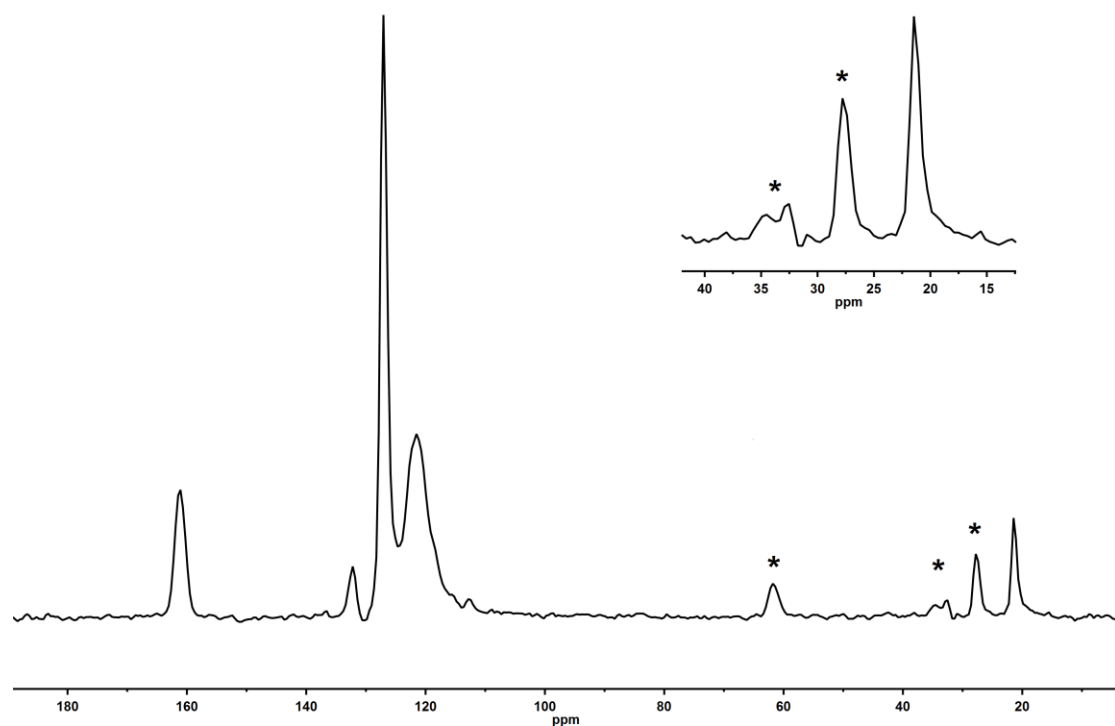


Figure S28: The $^{13}\text{C}\{^1\text{H}\}$ NQS SSNMR (100 MHz, 198 K, 10 kHz spin rate) spectrum of $[1\text{-C}_6\text{H}_{12}][\text{BAr}^{\text{F}}_4]$ prepared in situ. The resonances marked * are due to spinning sidebands.

The inset is an enlargement of the resonances at δ 45 to 15, showing only a single resonance at δ 21.4. This peak did not shift position in the analogous 8 kHz NQS SSNMR spectrum, suggestive of a real signal and not a sideband.

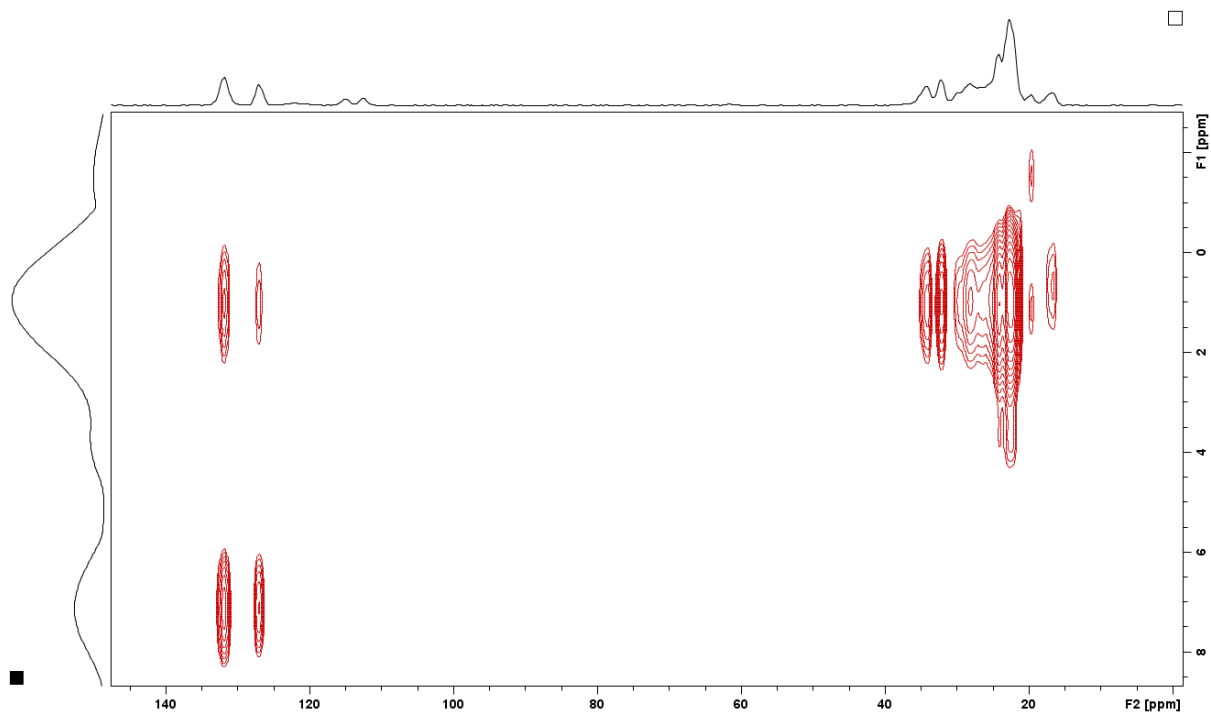


Figure S29: The $^1\text{H}/^{13}\text{C}$ FSLG HETCOR SSNMR (158 K, 10 kHz spin rate) spectrum of $[1\text{-C}_6\text{H}_{12}][\text{BAR}^{\text{F}}_4]$ prepared in situ.

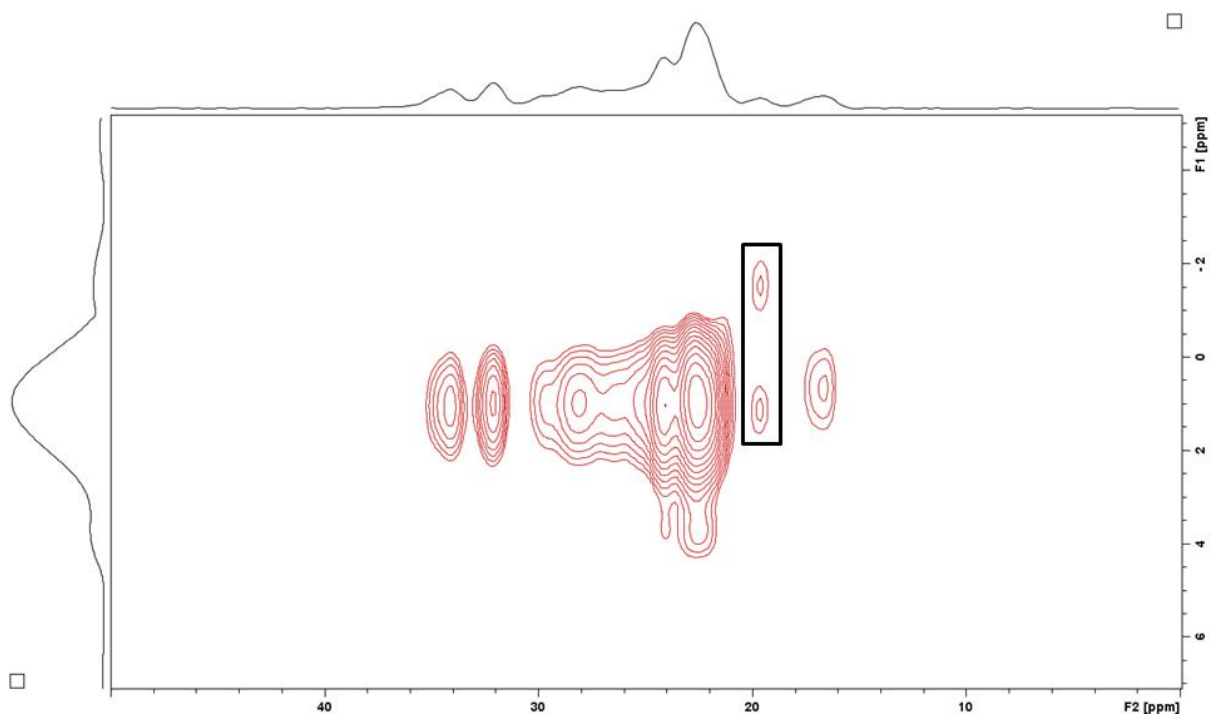


Figure S30: The $^1\text{H}/^{13}\text{C}$ FSLG HETCOR SSNMR (158 K, 10 kHz spin rate) spectrum of $[1\text{-C}_6\text{H}_{12}][\text{BAR}^{\text{F}}_4]$ prepared in situ (top). Enlargement of the alkyl region (bottom). Black box added to guide the eye.

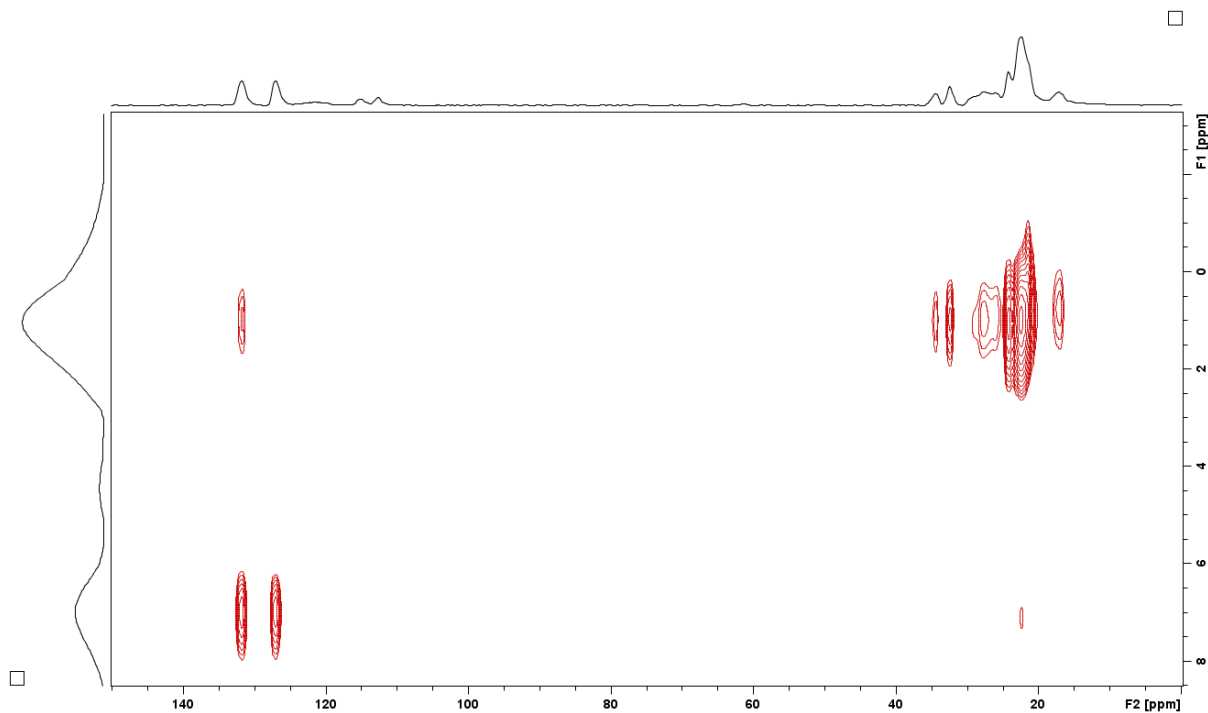


Figure S31: The $^1\text{H}/^{13}\text{C}$ FSLG HETCOR SSNMR (198 K, 10 kHz spin rate) spectrum of $[1\text{-C}_6\text{H}_{12}][\text{BAr}^{\text{F}}_4]$ prepared in situ.

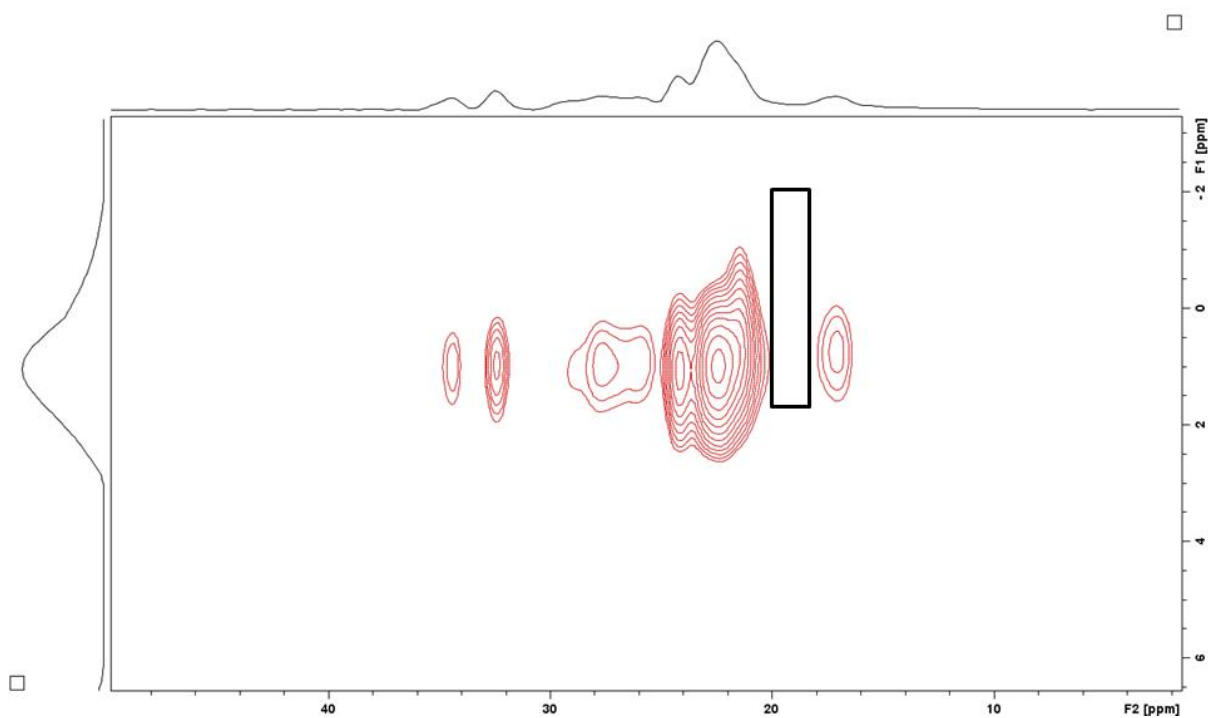


Figure S32: The $^1\text{H}/^{13}\text{C}$ FSLG HETCOR SSNMR (198 K, 10 kHz spin rate) spectrum of $[1\text{-C}_6\text{H}_{12}][\text{BAr}^{\text{F}}_4]$ prepared in situ (top). Enlargement of the alkyl region (bottom). Black box added to guide the eye.

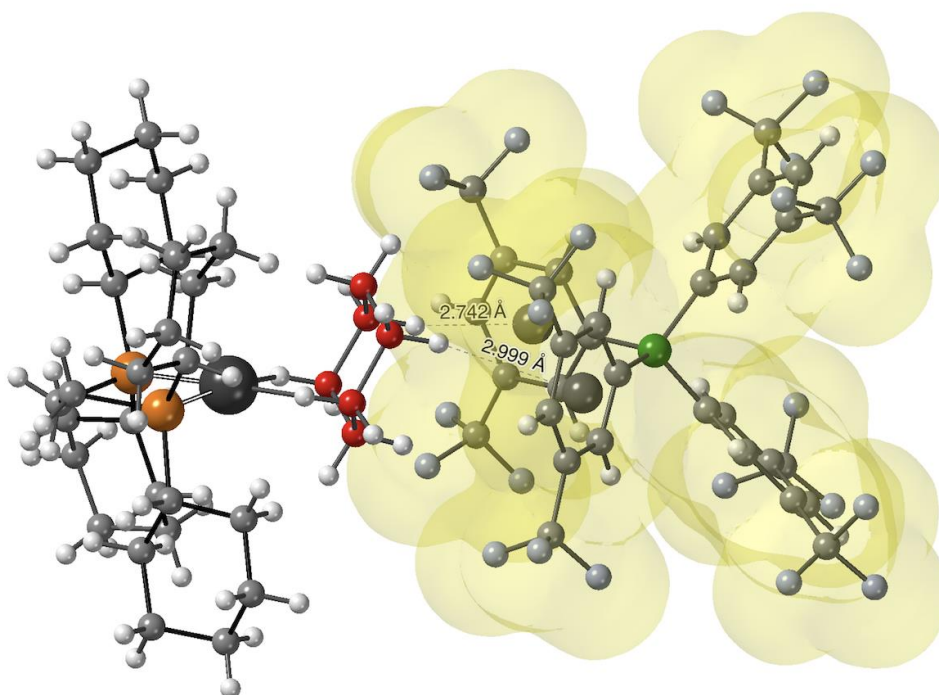


Figure S33: Molecular structure of $[1\text{-C}_6\text{H}_{12}][\text{BAR}^{\text{F}}_4]$ showing C-H...F-C contacts ($< 3.0\text{\AA}$) between the cyclohexane ligand and $[\text{BAR}^{\text{F}}_4]^-$ anion.

S.2.4.5. Liberation of the bound cyclohexane from $[1\text{-C}_6\text{H}_{12}][\text{BAR}^{\text{F}}_4]$.

Deep red crystals of $[1\text{-C}_6\text{H}_8][\text{BAR}^{\text{F}}_4]$ were treated with H_2 (2 bar, 298 K, 15 mins) inside a J Young flask. The colour of the crystals immediately darkens to a burgundy color, yielding $[1\text{-C}_6\text{H}_{12}][\text{BAR}^{\text{F}}_4]$. The solid was then cooled to 77 K under liquid nitrogen and the J Young flask placed under vacuum to remove any H_2 . CO (1 bar, 298K) was added and solid left for 1 hour; to which a vibrant yellow solid material had formed. The solid was then dissolved in $d_3\text{-MeCN}$, to which the volatile component was isolated by trap-to-trap distillation and analyzed by ^1H NMR spectroscopy.

^1H NMR ($d_3\text{-MeCN}$, 400 MHz): δ 1.44 (s, 12 H, alkane- CH_2 -)

The resultant solid was confirmed to be $[\text{Rh}(\text{dcpe})(\text{CO})_2][\text{BAR}^{\text{F}}]$ by solution NMR $^{31}\text{P}\{^1\text{H}\}$ NMR spectroscopy (δ 85.22, d, $J_{\text{RhP}} = 116$ Hz).^{S7}

S.2.4.6. Solid state decomposition products of $[1\text{-C}_6\text{H}_{12}][\text{BAr}^{\text{F}}_4]$ under H_2

A powdered microcrystalline sample of $[1\text{-C}_6\text{H}_8][\text{BAr}^{\text{F}}_4]$ (35 mg) was packed in a 3.2 mm SSNMR rotor, inside an argon filled glove box. The rotor was then placed in a custom built glass J Young flask^{S7} and the sample was then exposed to H_2 (1 bar, 298 K). After 80 minutes, the rotor cap was fitted under a flush of H_2 . The sample was immediately transferred to the bore of a pre-cooled (158 K) SSNMR spectrometer (~ 10 mins, total time = 90 mins) and analyzed $^{31}\text{P}\{^1\text{H}\}$ and $^{13}\text{C}\{^1\text{H}\}$ solid state NMR spectroscopy.

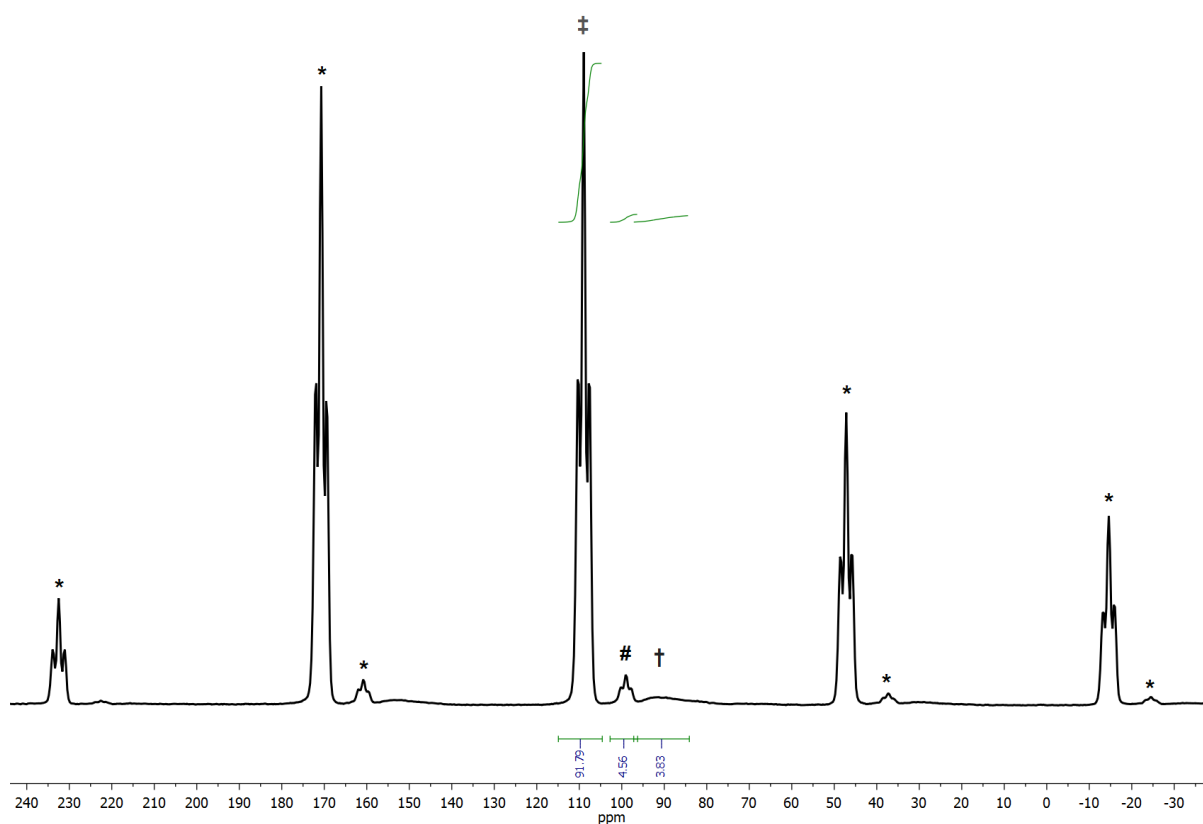
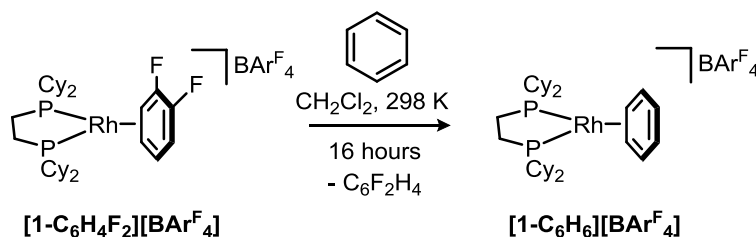


Figure S34: The $^{31}\text{P}\{^1\text{H}\}$ SSNMR (162 MHz, 295 K, 10 kHz spin rate) spectrum of the solid state decomposition products of $[1\text{-C}_6\text{H}_{12}][\text{BAr}^{\text{F}}_4]$ under H_2 after 90 minutes. The resonances \ddagger marked are assigned to $[1\text{-C}_6\text{H}_{12}][\text{BAr}^{\text{F}}_4]$ (approx. 90%) and the resonances marked \dagger are assigned to $[1\text{-BAr}^{\text{F}}_4]^{\text{S}2}$ (approx. 5%). The resonance marked $\#$ at δ 100 is unidentified complex under H_2 (approx. 5%), also seen in the decomposition of $[1\text{-C}_4\text{H}_{10}][\text{BAr}^{\text{F}}_4]$ also under hydrogen atmosphere (section S.2.2.6.). The resonances marked $*$ are due to spinning sidebands.

S.2.5. [Rh(Cy₂PCH₂CH₂PCy₂)(C₆H₆)](BARF₄), [1-C₆H₆](BARF₄)

S.2.5.1. Synthesis of [1-C₆H₆](BARF₄)



A solution of [1-C₆H₄F₂](BARF₄) (70 mg, 46.6 μmol) in CH₂Cl₂ (5 mL) was treated with C₆H₆ (100 μL) at ambient temperature. The resultant yellow solution was stirred at ambient temperature for 16 hours, and then filtered. Addition of pentane afforded a yellow powder, which was washed with further pentane and then dried under vacuum. Yield: 59 mg (86 %).

S.2.5.2. Characterization data for [1-C₆H₆](BARF₄)

¹H NMR (CD₂Cl₂, 298 K, 400 MHz): δ 7.72 (s, 8H, ortho-ArH), 7.57 (s, 4H, para-ArH), 6.52 (s, 6H, C₆H₆), 1.91-1.59 (br m, 28H, overlapping aliphatic CH), 1.37-1.14 (br m, 16H, aliphatic CH), 1.04-0.94 (br m, 4H, overlapping aliphatic CH).

¹H NMR (CD₂Cl₂, 183 K, 400 MHz): δ 7.72 (s, 8H, ortho-ArH), 7.54 (s, 4H, para-ArH), 6.44 (s, 6H, C₆H₆), 1.78-1.44 (br m, 28H, overlapping aliphatic CH), 1.26-0.96 (br m, 16H, aliphatic CH), 0.92-0.79 (br m, 4H, overlapping aliphatic CH).

³¹P{¹H} NMR (CD₂Cl₂, 298 K, 162 MHz): δ 99.7 (d, J_{RhP} 201 Hz).

³¹P{¹H} NMR (CD₂Cl₂, 183 K, 162 MHz): δ 99.4 (d, J_{RhP} 201 Hz).

¹⁹F{¹H} NMR (CD₂Cl₂, 298 K, 377 MHz): δ -62.9 (s).

¹³C{¹H} NMR (CD₂Cl₂, 298 K, 126 MHz): δ 162.2 (q, J_{CB} 50 Hz, ipso-ArC), 135.2 (s, ortho-ArC), 129.3 (q, J_{CF} 32 Hz, meta-ArC), 125.0 (q, J_{CF} 272 Hz, CF₃), 117.9 (m, para-ArC), 99.6 (d, J_{CRh} 1.9 Hz, C₆H₆), 38.6 (m, cyclohexyl-CH), 29.4 (m, cyclohexyl-CH₂), 27.1 (m, cyclohexyl-CH₂), 26.4 (s, cyclohexyl-CH₂), 22.9 (m, cyclohexyl-CH₂).

ESI-MS found (calc. for [Rh(Cy₂PCH₂CH₂PCy₂)(C₆H₆)]⁺): m/z 603.29 (603.28).

Elemental analysis found (calc. for C₆₄H₆₆BF₂₄P₂Rh): C 52.29 (52.40), H 4.35 (4.54).

S.2.5.3. Spectra of [1-C₆H₆][BAr^F₄]

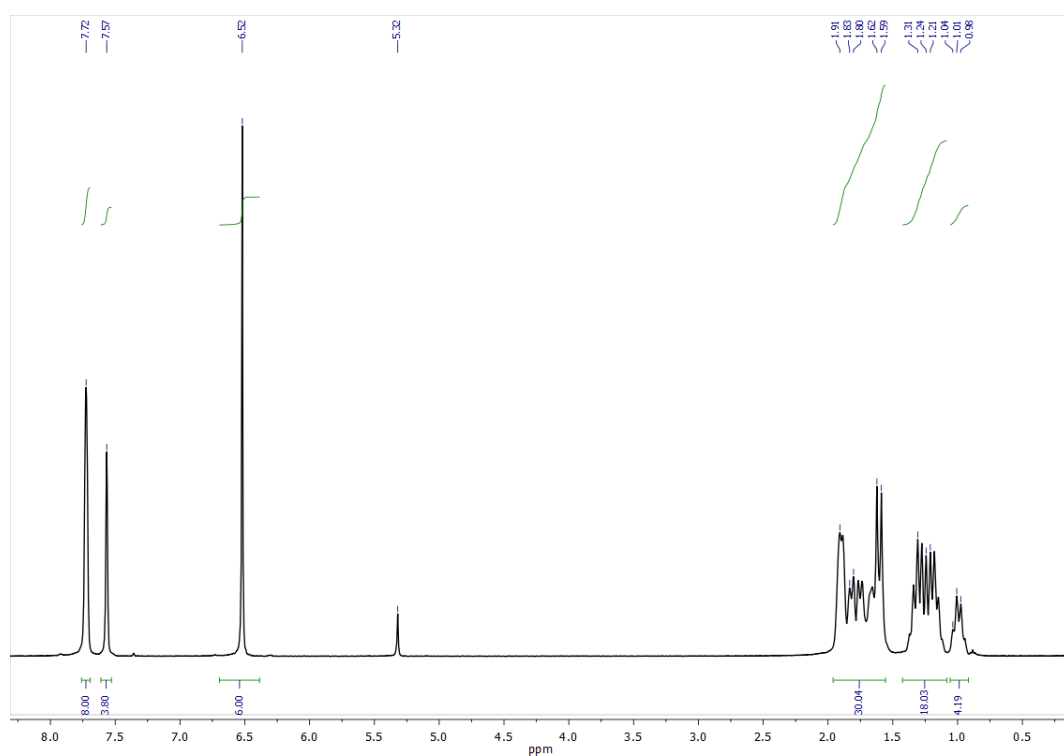


Figure S35: The solution ¹H NMR (CD₂Cl₂, 298 K, 400 MHz) spectrum of [1-C₆H₆][BAr^F₄].

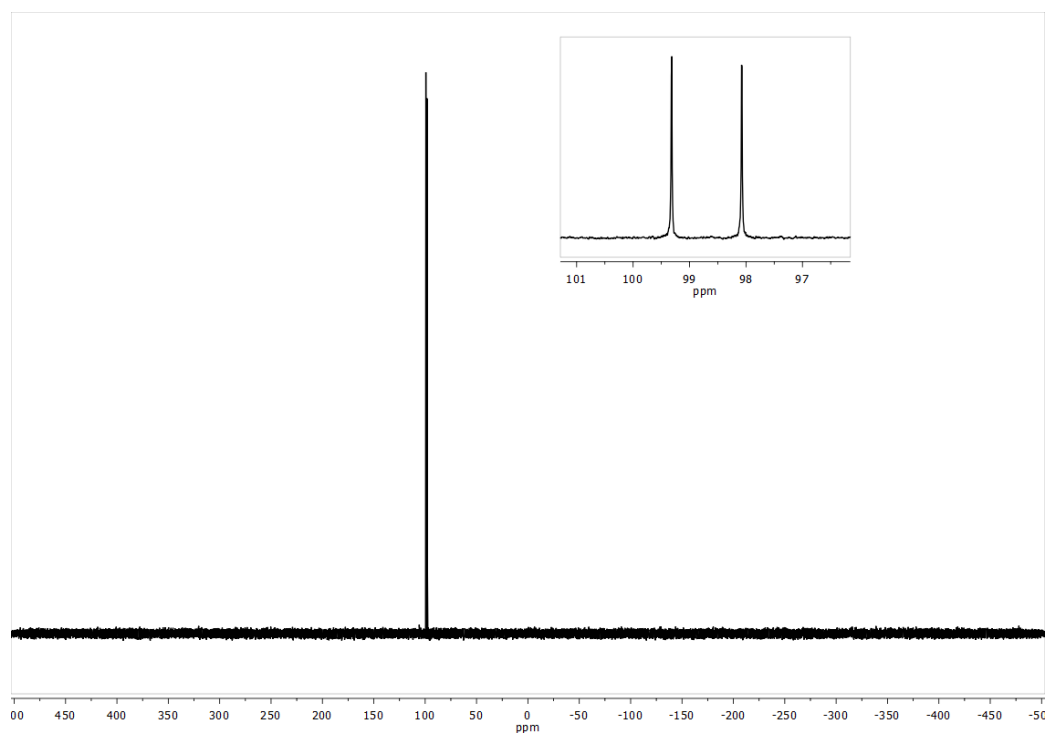


Figure S36: The solution ³¹P{¹H} NMR (CD₂Cl₂, 298 K, 162 MHz) spectrum of [1-C₆H₆][BAr^F₄].

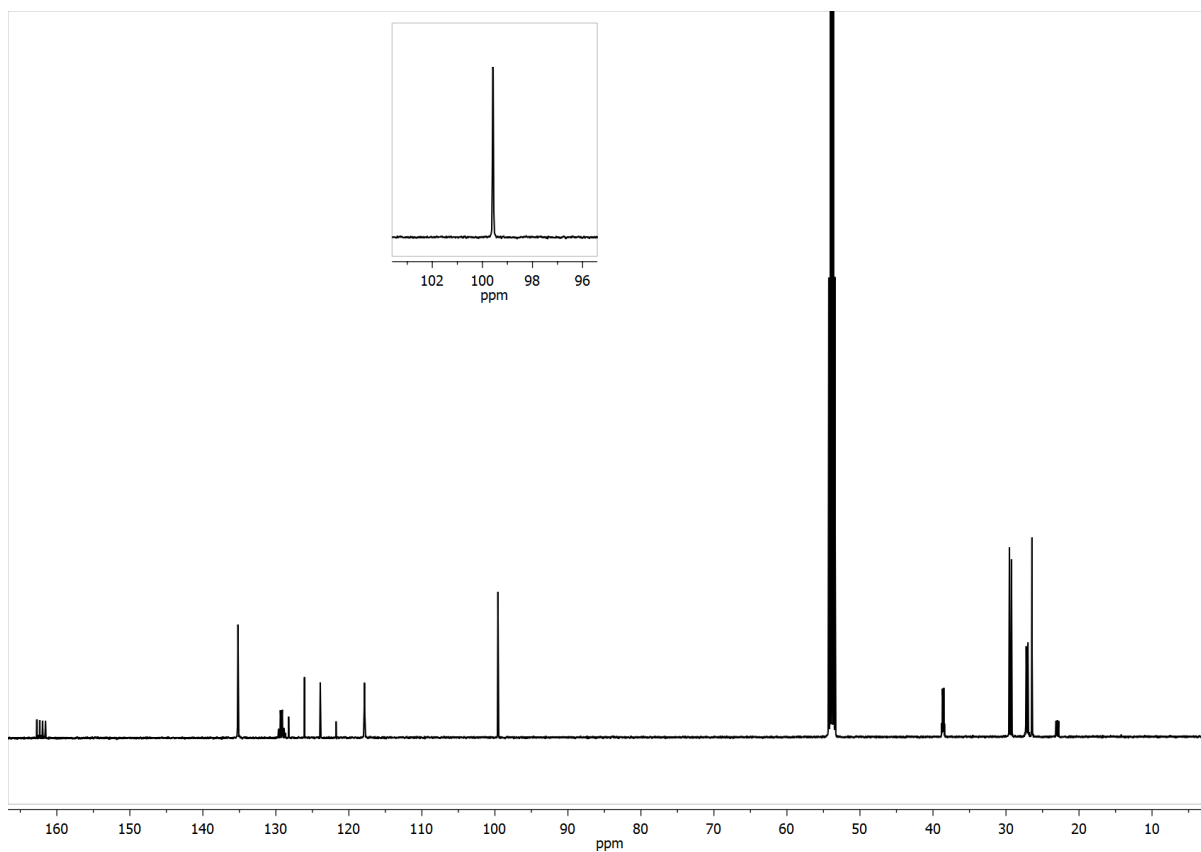


Figure S37: The solution $^{13}\text{C}\{^1\text{H}\}$ NMR (CD_2Cl_2 , 298 K, 126 MHz) spectrum of [1- C_6H_6][BAr^{F}_4]. The inset shows an enlargement of the benzene carbon signals at δ 99.6.

S.2.6. $[\text{Rh}(\text{Cy}_2\text{PCH}_2\text{CH}_2\text{PCy}_2)(\text{C}_6\text{D}_6)][\text{BAr}^{\text{F}}_4]$, $[\text{1-C}_6\text{D}_6][\text{BAr}^{\text{F}}_4]$

S.2.6.1. Synthesis of $[\text{1-C}_6\text{D}_6][\text{BAr}^{\text{F}}_4]$

A solution of $[\text{1-C}_6\text{H}_4\text{F}_2][\text{BAr}^{\text{F}}_4]$ (35 mg, 23.3 μmol) in CH_2Cl_2 (3 mL) was treated with C_6D_6 (50 μL) at ambient temperature. The resultant yellow solution was stirred at ambient temperature for 16 hours and then filtered. Addition of pentane afforded a yellow powder, which was washed with further pentane and then dried under vacuum. Yield: 17 mg (50 %).

S.2.6.2. Characterization data for $[\text{1-C}_6\text{D}_6][\text{BAr}^{\text{F}}_4]$

^1H NMR (CD_2Cl_2 , 298 K, 400 MHz): δ 7.72 (s, 8H, ortho-ArH), 7.56 (s, 4H, para-ArH), 1.91-1.59 (br m, 28H, overlapping aliphatic CH), 1.37-1.14 (br m, 16H, aliphatic CH), 1.04-0.94 (br m, 4H, overlapping aliphatic CH).

^1H NMR (CD_2Cl_2 , 183 K, 400 MHz): δ 7.72 (s, 8H, ortho-ArH), 7.54 (s, 4H, para-ArH), 1.78-1.44 (br m, 28H, overlapping aliphatic CH), 1.26-0.96 (br m, 16H, aliphatic CH), 0.92-0.79 (br m, 4H, overlapping aliphatic CH).

^2H NMR (CD_2Cl_2 , 298 K, 61 MHz): δ 6.56 (s, C_6D_6).

$^{31}\text{P}\{^1\text{H}\}$ NMR (CD_2Cl_2 , 298 K, 162 MHz): δ 98.7 (d, J_{RhP} 201 Hz).

$^{31}\text{P}\{^1\text{H}\}$ NMR (CD_2Cl_2 , 183 K, 162 MHz): δ 99.5 (d, J_{RhP} 201 Hz).

$^{19}\text{F}\{^1\text{H}\}$ NMR (CD_2Cl_2 , 298 K, 377 MHz): δ -62.9 (s).

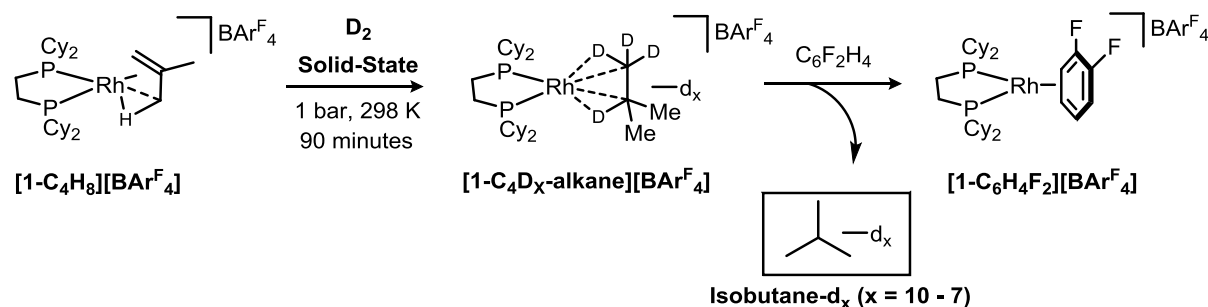
$^{13}\text{C}\{^1\text{H}\}$ NMR (CD_2Cl_2 , 298 K, 126 MHz): δ 162.2 (q, J_{CB} 50 Hz, ipso-ArC), 135.2 (s, ortho-ArC), 129.3 (q, J_{CF} 32 Hz, meta-ArC), 125.0 (q, J_{CF} 272 Hz, CF_3), 117.9 (m, para-ArC), 99.2 (t, J_{CD} 26.1 Hz, C_6H_6), 38.6 (m, cyclohexyl-CH), 29.5 (s, cyclohexyl- CH_2), 29.2 (s, cyclohexyl- CH_2), 27.2 (m, cyclohexyl- CH_2), 27.0 (m, cyclohexyl- CH_2), 26.4 (s, cyclohexyl- CH_2), 23.1 (m, cyclohexyl- CH_2), 23.0 (m, cyclohexyl- CH_2), 23.0 (br s, cyclohexyl- CH_2), 22.8 (m, cyclohexyl- CH_2).

ESI-MS found (calc. for $[\text{Rh}(\text{Cy}_2\text{PCH}_2\text{CH}_2\text{PCy}_2)(\text{C}_6\text{D}_6)]^+$): m/z 609.32 (609.31).

S.3. H/D Exchange Experiments

S.3.1. Solid-state H/D exchange on [1-C₄H₈][BAR^F₄]

S.3.1.1. General Experimental of H/D Exchange on [1-C₄H₈][BAR^F₄]



A J Young NMR tube (~ 2 mL) was charged with 25 mgs of an orange crystalline sample of [1-C₄H₈][BAR^F₄]. The tube was evacuated ($< 3 \times 10^{-2}$ mbar) and D₂ (1 bar, 298 K) was added. The color of the crystals immediately turned a dark red, yielding the deuterated analogue of [1-C₄H₁₀][BAR^F₄]; [1-C₄D_x-alkane][BAR^F₄].

The reaction was left for 90 minutes; after which the crystal color changed to a dark orange. After this time, it was found isobutane-d_x (x = 10 - 7) had been expelled from the crystal lattice, shown by gas phase ¹H and ²H NMR spectroscopy of the NMR tube headspace.^{S6} The volatiles were then distilled into a clean NMR tube with CD₂Cl₂ and analyzed by ¹H, ²H, ¹³C {¹H} NMR spectroscopy and GC EI-MS.

The remaining solid material was dissolved in C₆F₂H₄ to quantitatively yield [1-C₆F₂H₄][BAR^F₄] complex. No isobutane could be identified in the solution ¹H or ²H NMR spectrum, suggesting total expulsion of isobutane from the lattice by D₂ is complete after 90 minutes.

S.3.1.2. Characterization data for isobutane-d_x

¹H NMR (CD₂Cl₂, 400 MHz, 183 K): δ 0.86 (br m, CH₃, residual protio-isobutane)

²H NMR (CD₂Cl₂, 61 MHz, 183 K): δ 1.66 (br s, CD), 0.85 (br s, CD₃).

¹³C {¹H} NMR (CD₂Cl₂, 126 MHz, 183 K): δ 23.6 (septet, CD₃, J_{CD} = 18.8), 22.5 (triplet, CD, J_{CD} = 20.1), for C₄D₁₀, but also formed is C₄HD₉, C₄H₂D₈ and C₄H₃D₇.

GC EI-MS found (calc. for C₄D₉⁺): m/z 66.127 (66.127).

S.3.1.3. NMR Spectra of isobutane-d_x (x = 10 – 7)

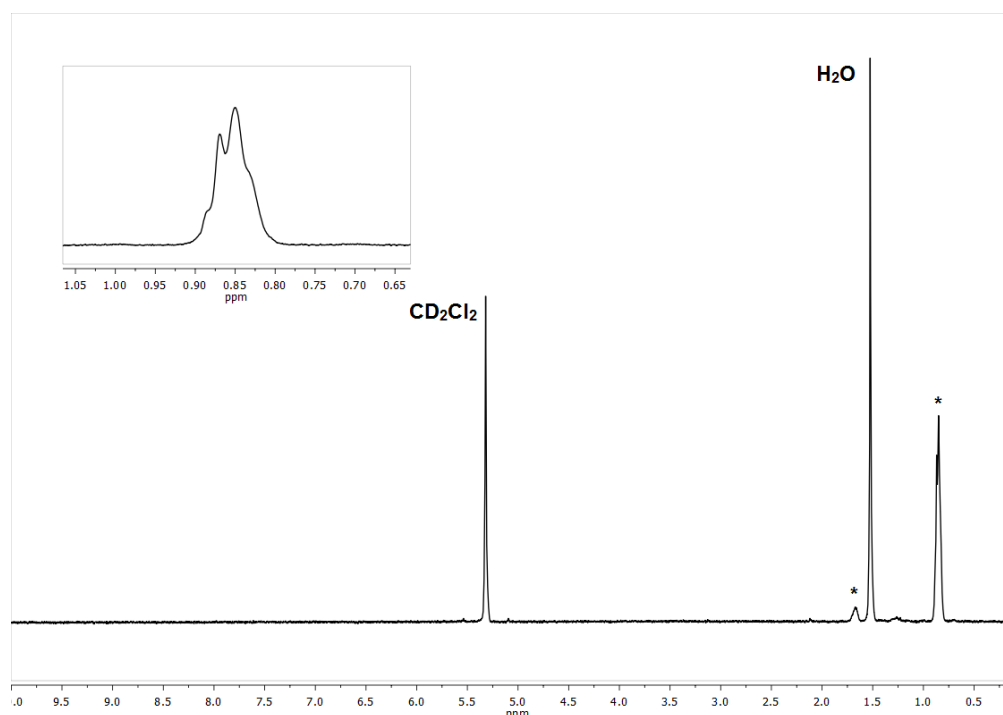


Figure S38: The solution ¹H NMR (CD₂Cl₂, 298 K, 400 MHz) spectrum of isolated volatiles. Residual protio-isobutane signals observed are marked with *.

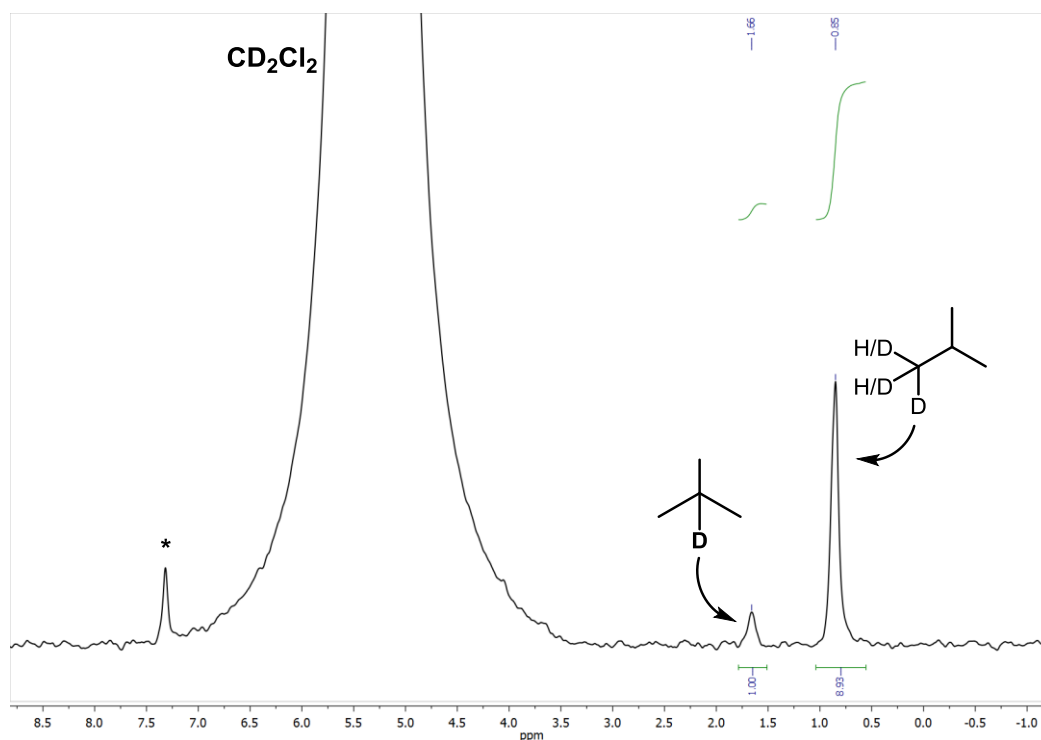


Figure S39: The solution ²H NMR (CD₂Cl₂, 298 K, 61 MHz) spectrum of isolated volatiles. The signal marked * is from a CDCl₃ impurity.

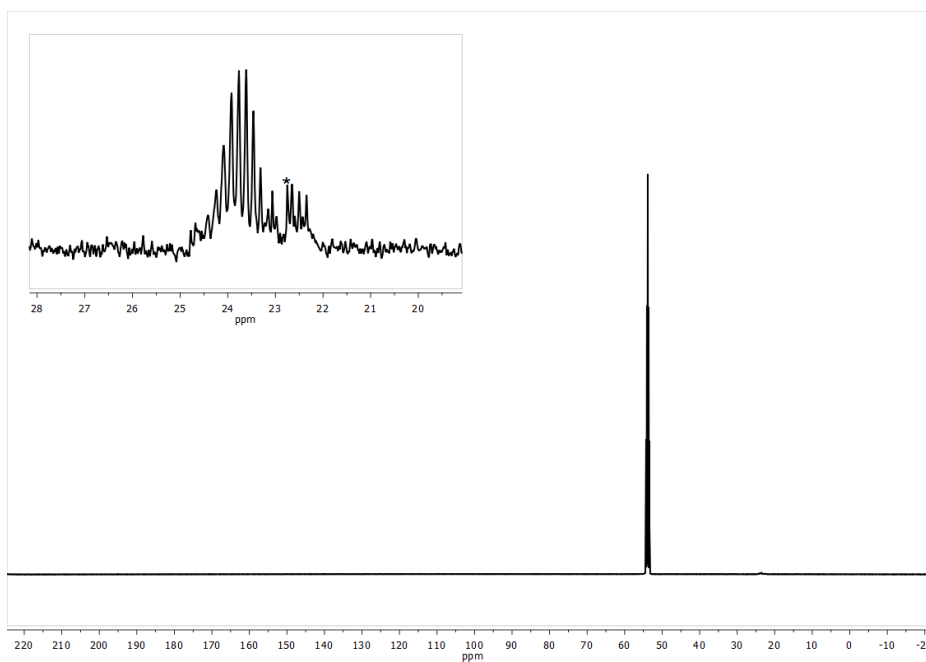


Figure S40: The solution $^{13}\text{C}\{^1\text{H}\}$ NMR (CD_2Cl_2 , 298 K, 126 MHz) spectrum of isolated volatiles. The inset is an enlargement of the resonances between δ 27 – 20, where the signal marked * is from a pentane impurity.

S.3.1.3. GC EI-MS Spectra of isobutane-d_x (x = 10 – 7) and isobutane-h₁₀

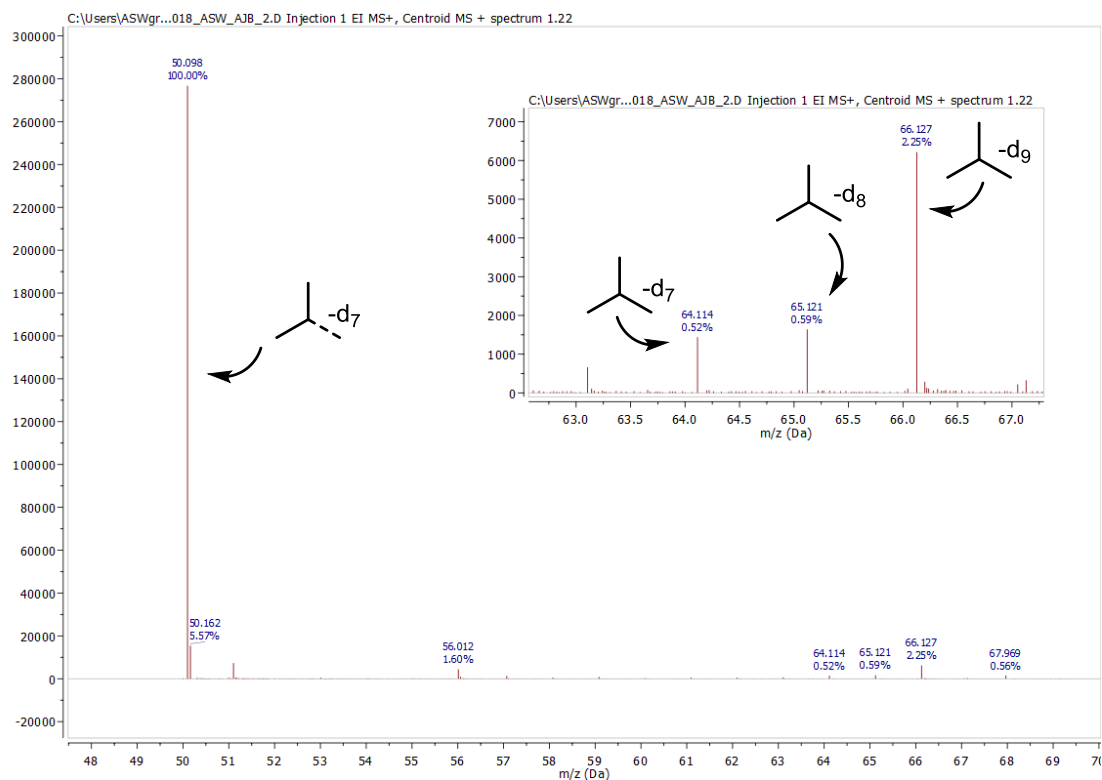


Figure S41: Mass spectrum of the isolated isobutane-d_x with enlargement of m/z = 62 – 69 region. Due to relative ease of fragmentation of isobutane,^{S8} the molecular ion peak is that of C₄D₉⁺, m/z = 66.127 (calc. 66.1269) with greatest intensity fragment of C₃D₇⁺, m/z = 50.098 (calc. = 50.098). Other main isotopologues are of C₄HD₈⁺, m/z = 65.121 (calc. 65.120), C₄H₂D₇⁺, m/z = 64.114 (calc. 64.114).

To further show the relative ease of fragmentation; commercially bought isobutane-H₁₀ was studied using GC EI-MS, shown in Figure S42.

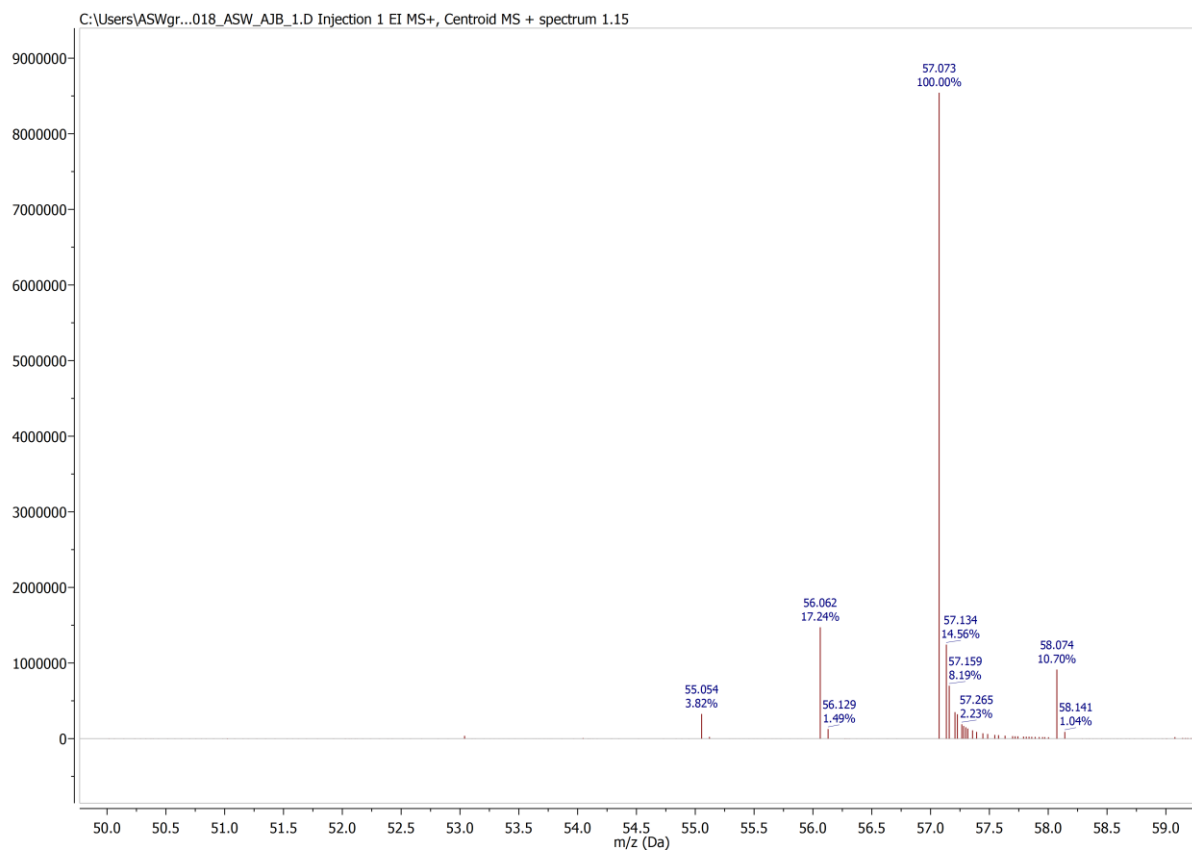


Figure S42: GC EI-Mass spectrum of commercially bought isobutane- H_{10} . Due to relative ease of fragmentation of isobutane,^{S8} the molecular ion peak is that of $C_4H_9^+$, $m/z = 57.073$ (calc. 57.078).

Although approx. 10 % $C_4H_{10}^+$, $m/z = 58.074$ (calc. 58.078) can be located, in the experimental condition presented above, the percentage of $C_4D_{10}^+$ in the isobutane- d_x sample is below the detection intensity of the equipment, and hence cannot be unequivocally identified.

S.3.1.4. Simulated solution $^{13}\text{C}\{^1\text{H}\}$ NMR spectra of isobutane- d_x ($x = 10 - 7$)

The solution $^{13}\text{C}\{^1\text{H}\}$ NMR spectrum of isolated isobutane- d_x ($x = 10 - 7$) (Figure S40) was de-convoluted using the Spin Simulation function on MestReNova, shown in Figure S43. The line width was kept at 6.0 Hz throughout, only changing the levels of population in each simulated environment.

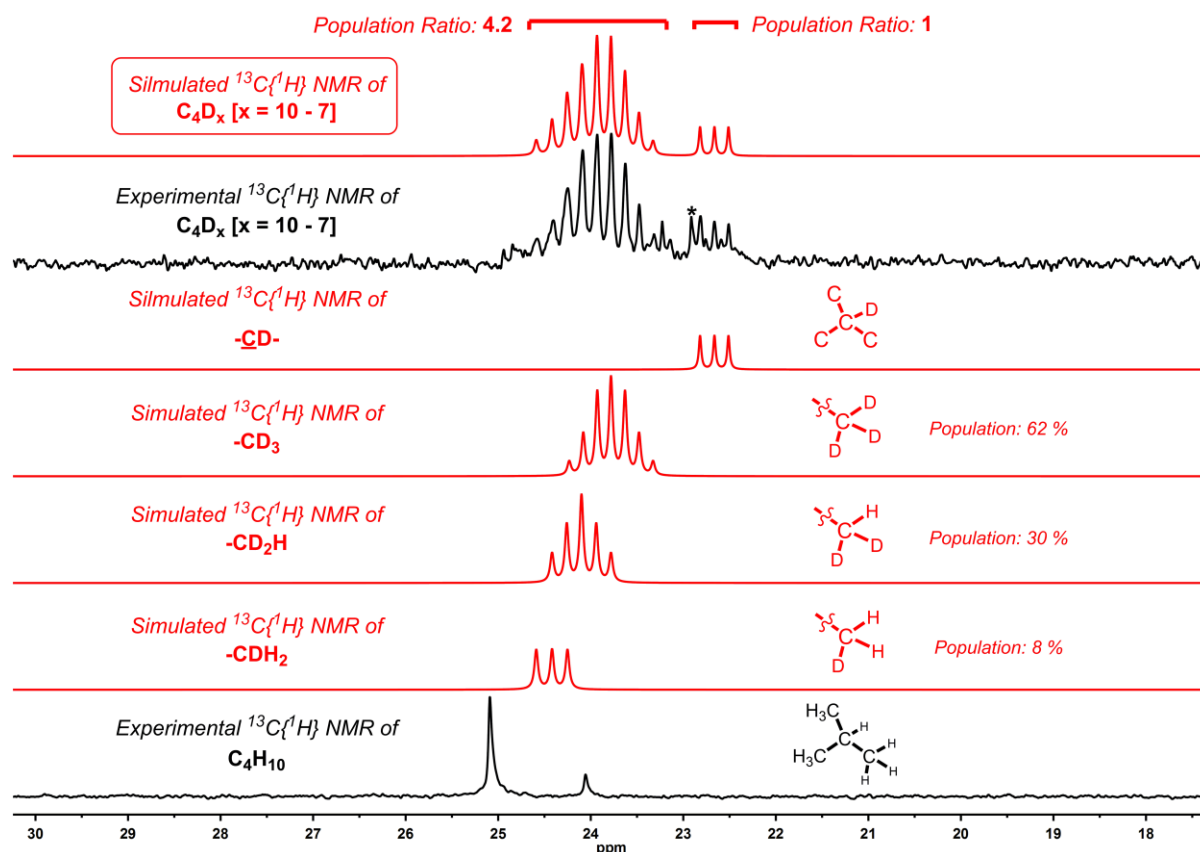


Figure S43: Simulated solution $^{13}\text{C}\{^1\text{H}\}$ NMR spectra of isobutane- d_x ($x = 10 - 7$), showing approximated populations of varying level of deuteration in the $-\text{CX}_3$ groups of isobutane. The signal marked by * is from a pentane impurity.

Three systems were used to recreate the multiplet at δ 23.85, modelling the ^{13}C methyl environments as $-\text{CD}_3$, $-\text{CD}_2\text{H}$ and $-\text{CDH}_2$. No evidence of $-\text{CH}_3$ could be seen in the experimental data so was not modelled. A separate fourth system was also modelled describing the triplet at δ 22.66; reported as the methine carbon of isobutane- d_x , split by a single bound deuterium. The proto-methine group could not be modelled, as signals obscured by multiplet at δ 23.85.

The simulation suggests that 62 % of the $-CX_3$ groups were of $-CD_3$, 30 % are $-CD_2H$ and 8 % are $-CDH_2$. This is consistent with the GC EI-MS shown in Figure S41, which showed a high proportion of $C_4D_9^+$ and $C_4D_8^+$ from the signals at 66.127 and 65.121 respectively as well as the molecular ion of 50.098^+ from $C_3D_7^+$.

The population ratio of each ^{13}C environment in the simulated spectra was calculated from the sum of each individual $-CD_xH_y$ component in each environment compared between the environments. These ratios suggest an approximate 1:4.2 ratio of carbon environments; albeit a little higher than predicted (1:3). This may be due to unmodelled $-C-H$ methine group's signals obscured by the multiplet at δ 23.85, affecting this ratio.

S.3.2. Solution-state deuteration of $[1-C_4H_8][BAr^F_4]$

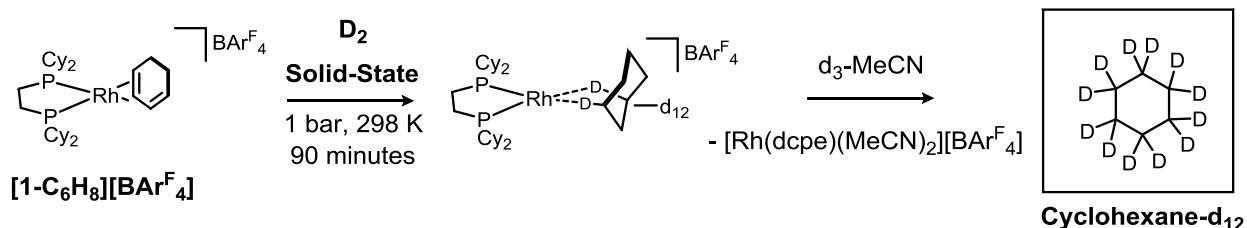
A solution of $[1-C_4H_8][BAr^F_4]$ (10 mg) in $C_6F_2H_4$ in a thick walled NMR tube was freeze-pump-thaw degassed three times and backfilled with D_2 (1 bar, 298 K). Upon agitation the color of the solution changed from red to yellow. Solution $^{31}P\{^1H\}$ NMR showed total conversion of the organometallic fragment to be $[1-C_6H_4F_2][BAr^F_4]$, where the volatile component was then isolated by trap-to-trap distillation. The products were identified as $C_4H_8D_2$ by 1H and 2H NMR spectroscopy which is the product of deuteration across the double bond only.

1H NMR (CD_2Cl_2 , 400 MHz, 183 K): δ 0.89 (br s, CH).

2H NMR (CD_2Cl_2 , 61 MHz, 183 K): δ 1.66 (br s, 1D, CD), 0.85 (br s, 1D, CD).

S.3.3. Solid-state H/D exchange on [1-C₆H₁₂][BAR^F₄]

S.3.3.1. General Experimental of H/D Exchange on [1-C₆H₁₂][BAR^F₄]



A thick walled NMR tube was charge with powdered microcrystalline [1-C₆H₈][BAR^F₄] (10 mg) in an argon filled glove box. The sample was then cooled to 77 K in liquid nitrogen, evacuated and backfilled with D₂ (1 bar, 298 K). After 30 min, sample was then cooled again to 77 K in liquid nitrogen, evacuated and backfilled with D₂ (1 bar, 298 K). This cycle was repeated once more, to give a total of 3 x 30 mins cycles, totaling 90 minutes under D₂. The reaction was then quenched by condensing d₃-MeCN (0.4 mL) into the NMR tube. The volatiles were then isolated by trap-to-trap distillation.

²H NMR (d₃-MeCN, 61 MHz): δ 1.37 (s).

¹³C{¹H} NMR (d₃-MeCN, 126 MHz): δ 25.3 (1:2:3:2:1 quintet, J_{CD} = 19 Hz).

S.3.3.2. NMR Spectra

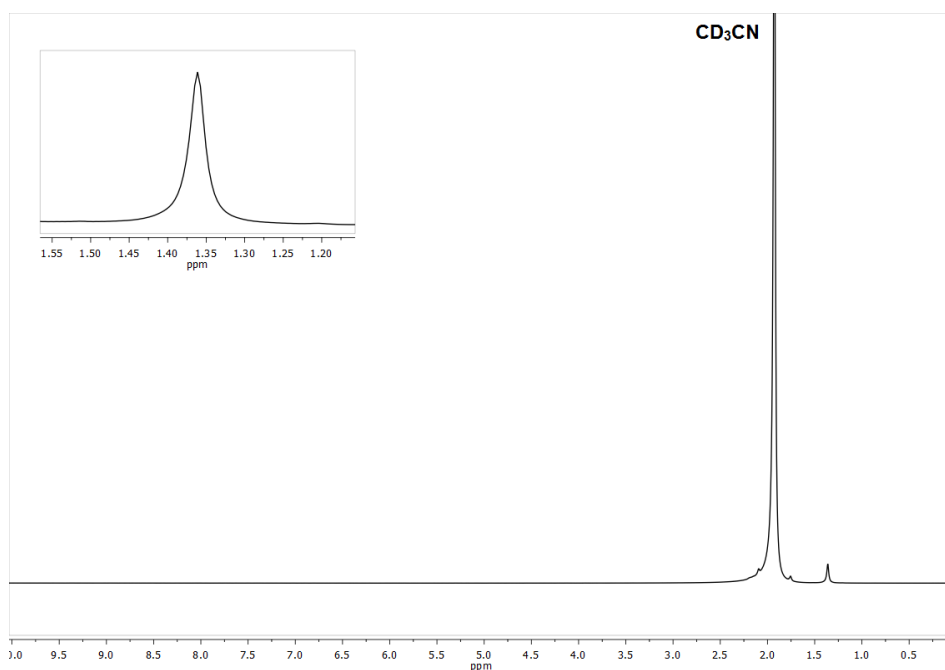


Figure S44: The solution ^2H NMR ($\text{d}_3\text{-MeCN}$, 298 K, 61 MHz) spectrum of cyclohexane- d_{12} .

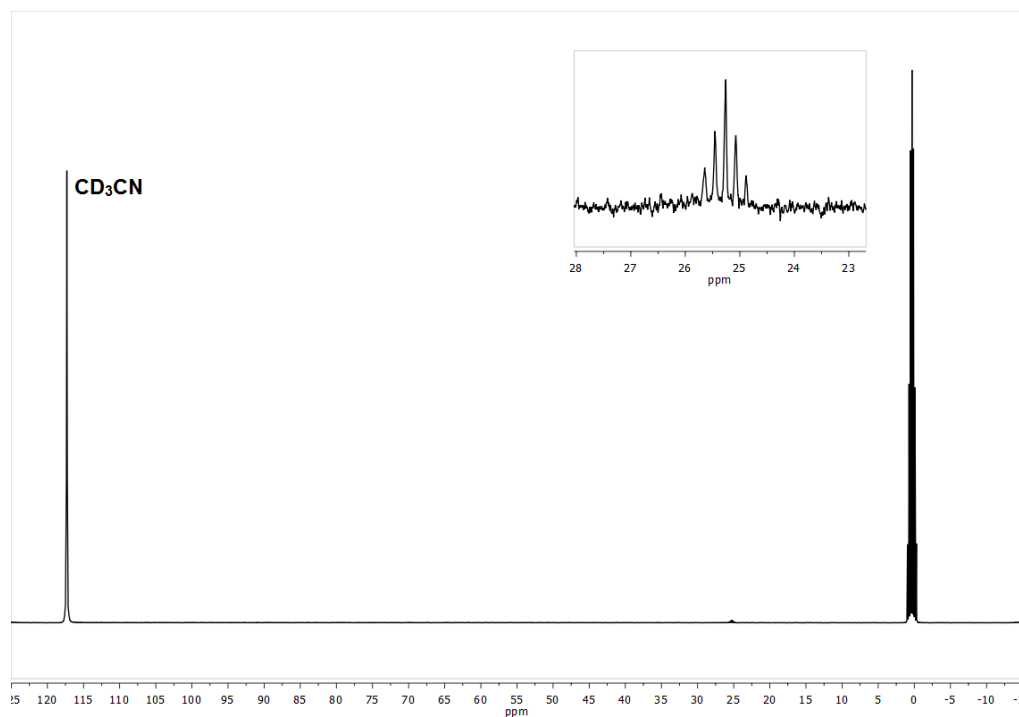


Figure S45: The solution $^{13}\text{C}\{^1\text{H}\}$ NMR ($\text{d}_3\text{-MeCN}$, 298 K, 100 MHz) spectrum of cyclohexane- d_{12} , δ 25.3 (1:2:3:2:1 quintet, $J_{\text{CD}} = 19$ Hz).

S.3.3.3. GC EI-Mass spectra

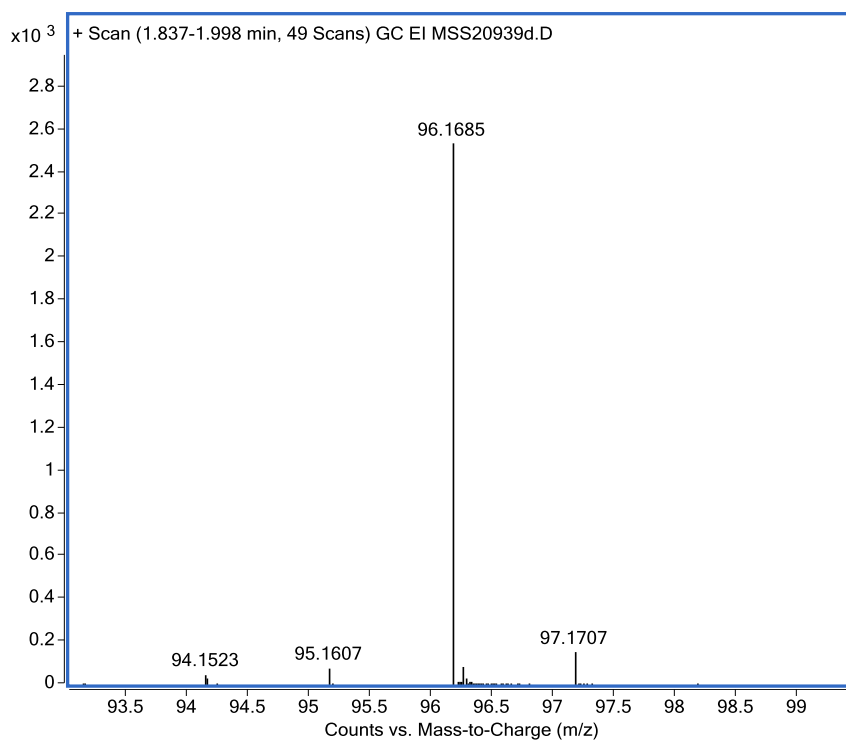
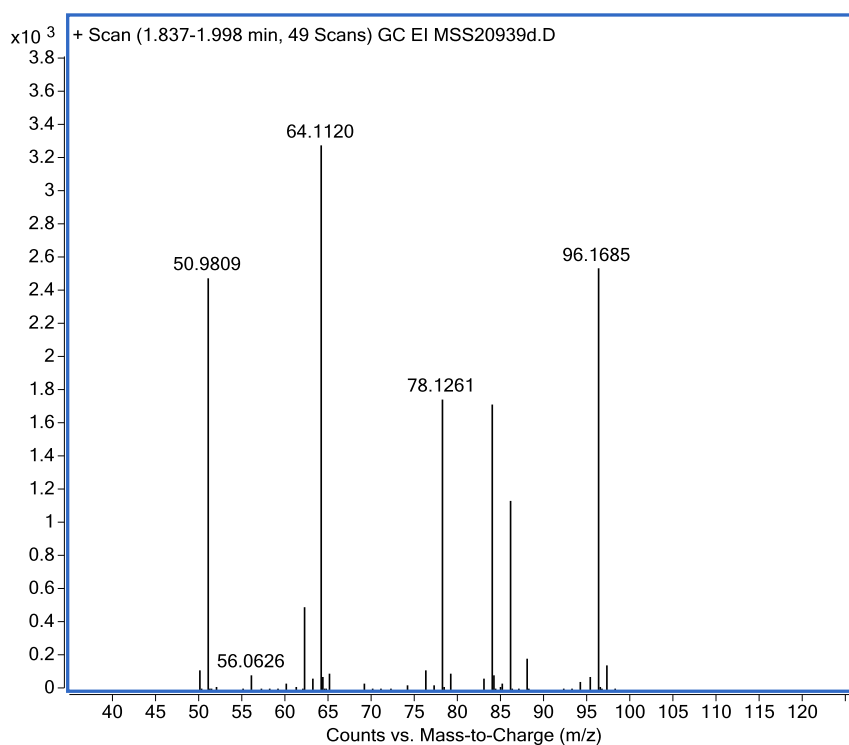


Figure S46: Mass spectrum of the isolated volatiles where the time of D₂ exposure was 30 minutes per cycle (top). Enlargement of m/z = 93 – 100 region (bottom). The signal at m/z = 96.1685 matches that of C₆D₁₂ (calc. 96.1687). Signals at m/z = 84 and 86 are due to CH₂Cl₂.

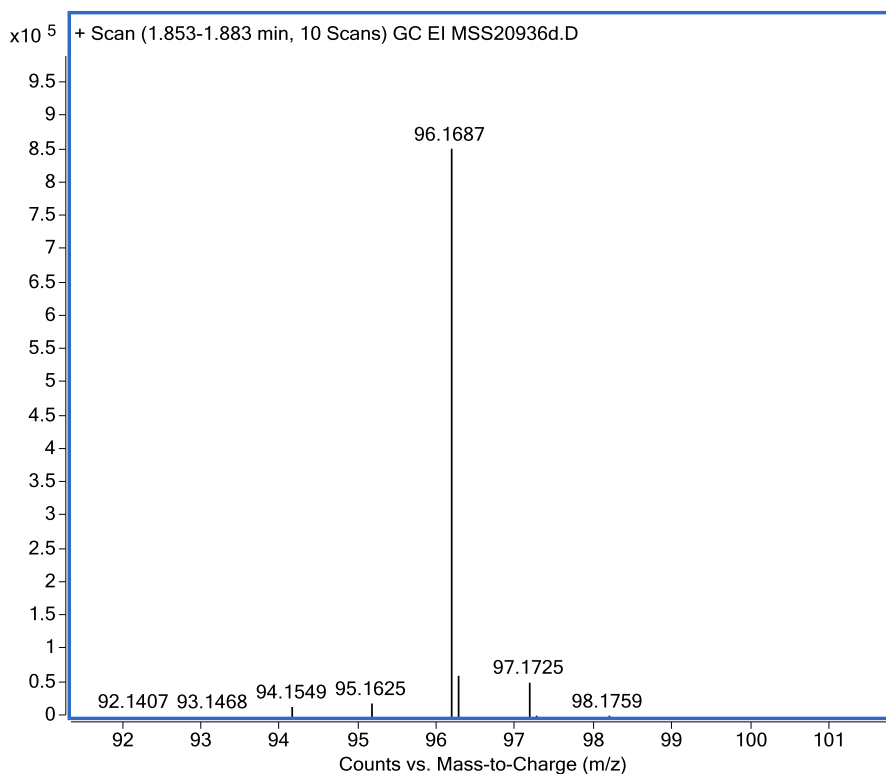
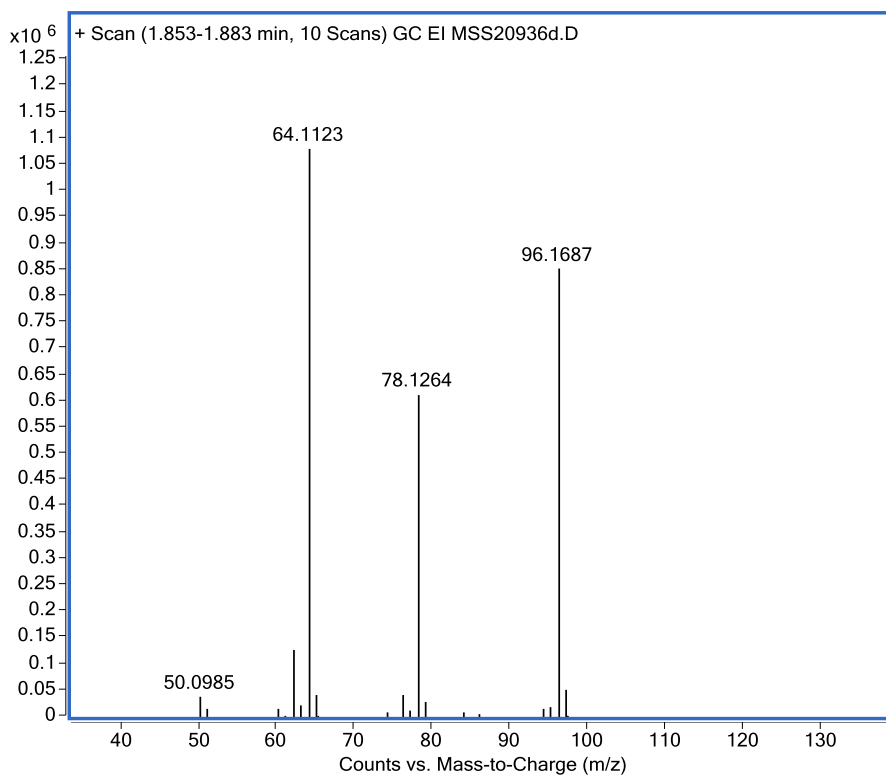


Figure S47: Mass spectrum of a commercially sourced sample of D₁₂-cyclohexane (top). Enlargement of m/z = 91 – 100 region (bottom).

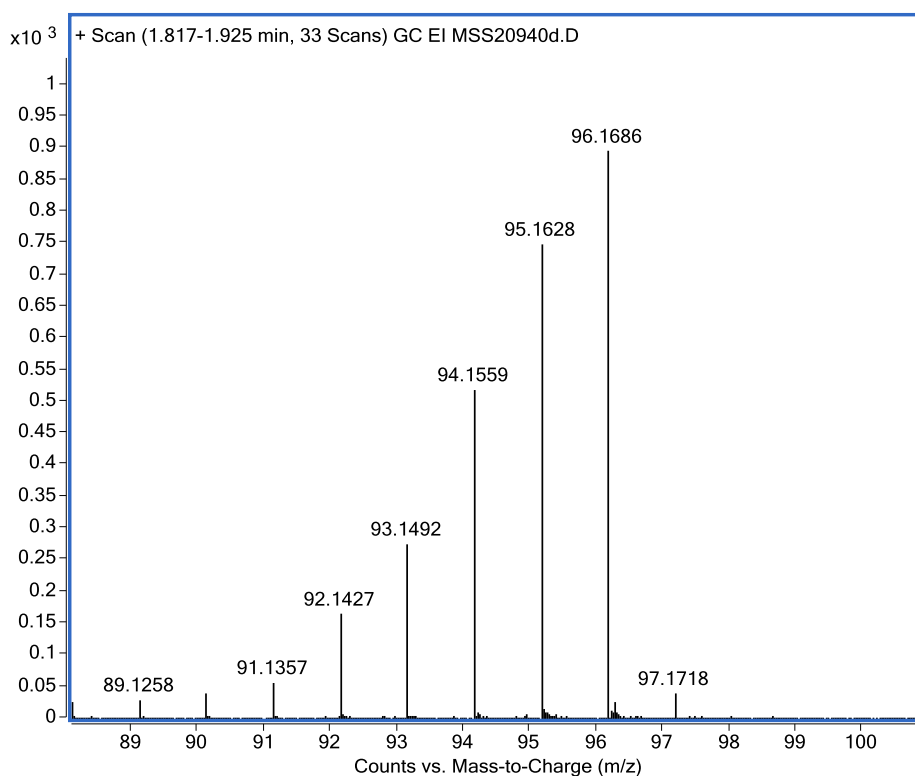
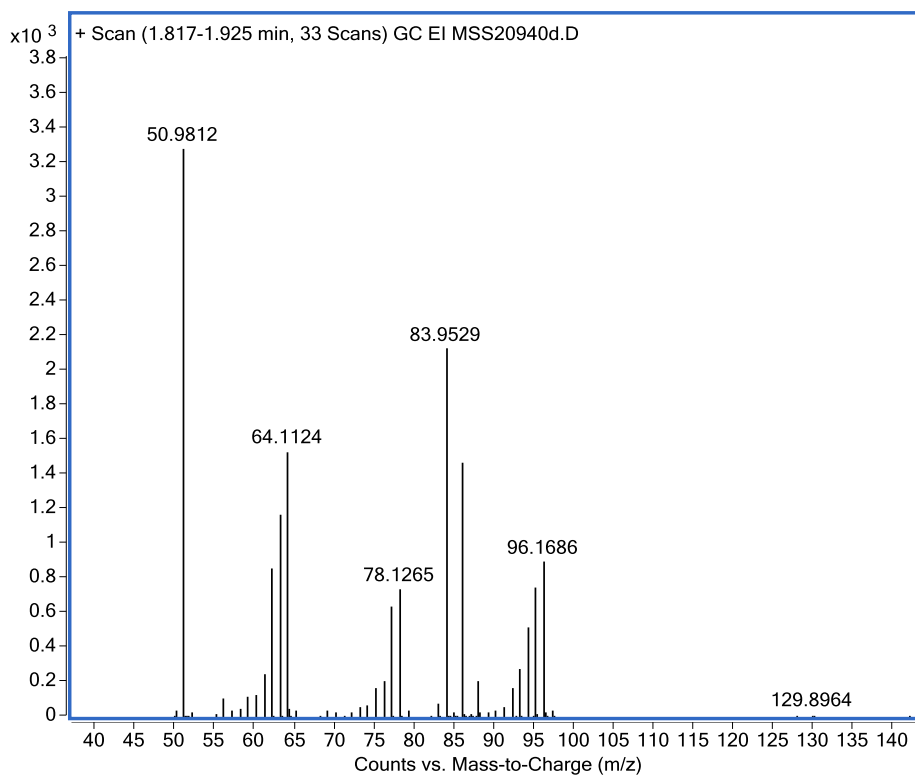


Figure S48: Mass spectrum of the isolated volatiles where the time of D₂ exposure was 5 minutes per cycle rather than 30 minutes (top). Enlargement of m/z = 88 – 100 region (bottom). Signals at m/z = 84 and 86 are due to CD₂Cl₂.

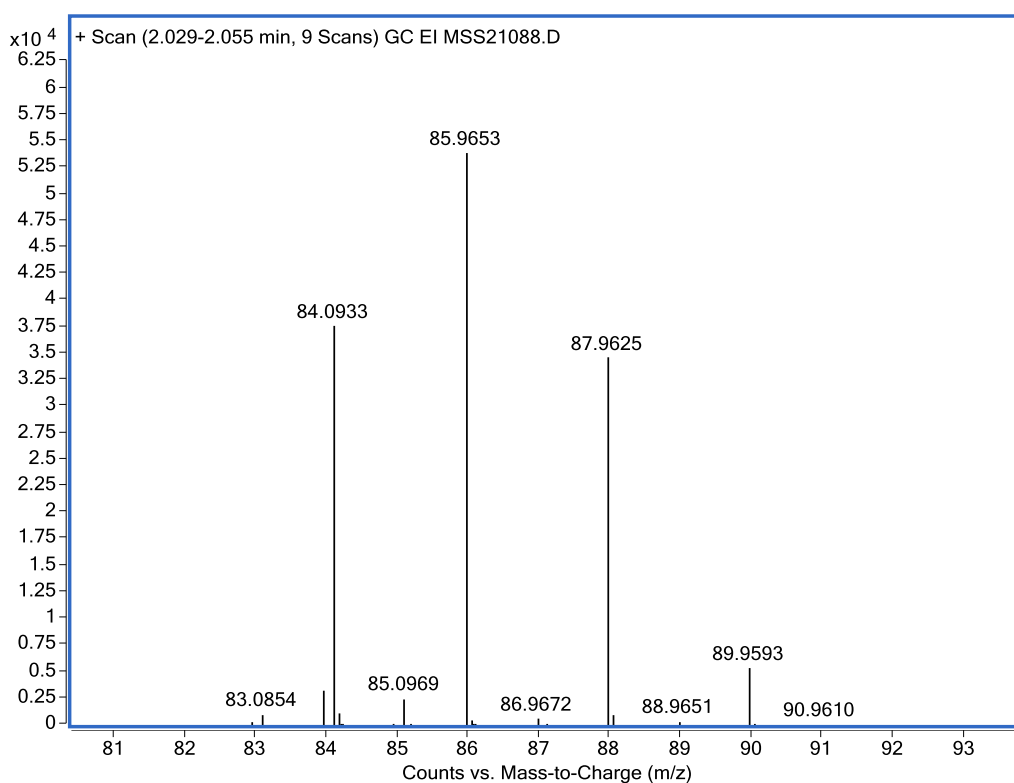
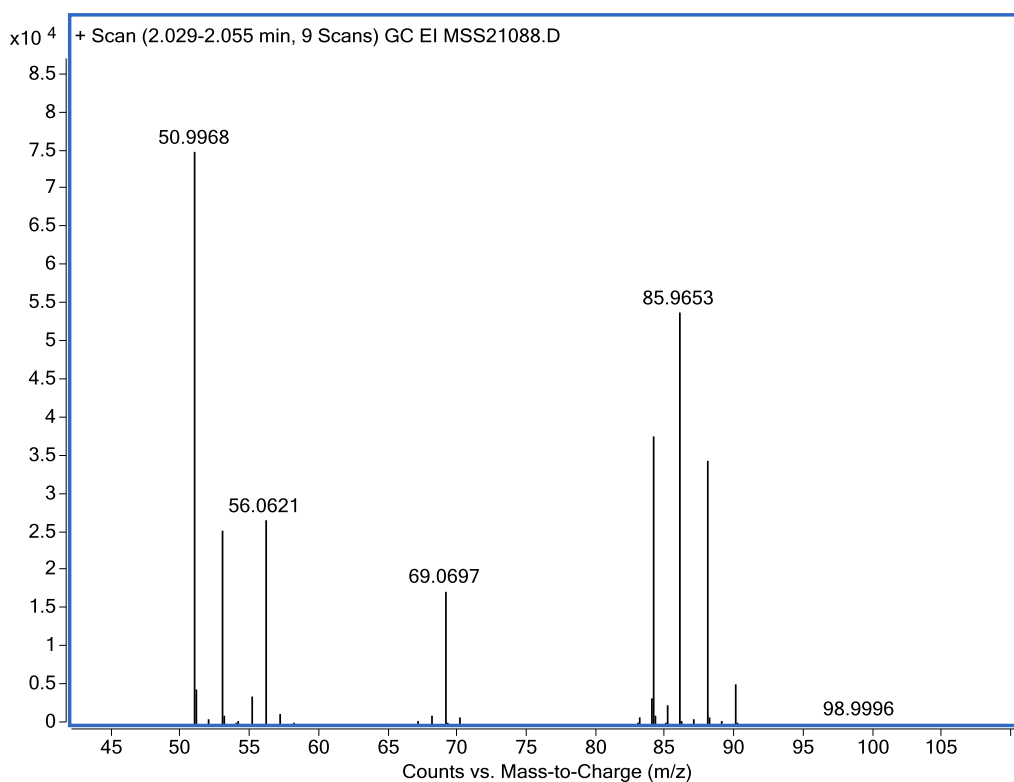


Figure S49: Mass spectrum of the isolated volatiles where the time of D₂ exposure was 0 minutes per cycle rather than 30 minutes (top), i.e., no D₂ exposure. Enlargement of m/z = 81 – 93 region (bottom). The signal at m/z = 84.0933 matches that of C₆H₁₂ (calc. 84.0933). Signals at m/z = 86 and 88 are due to CD₂Cl₂.

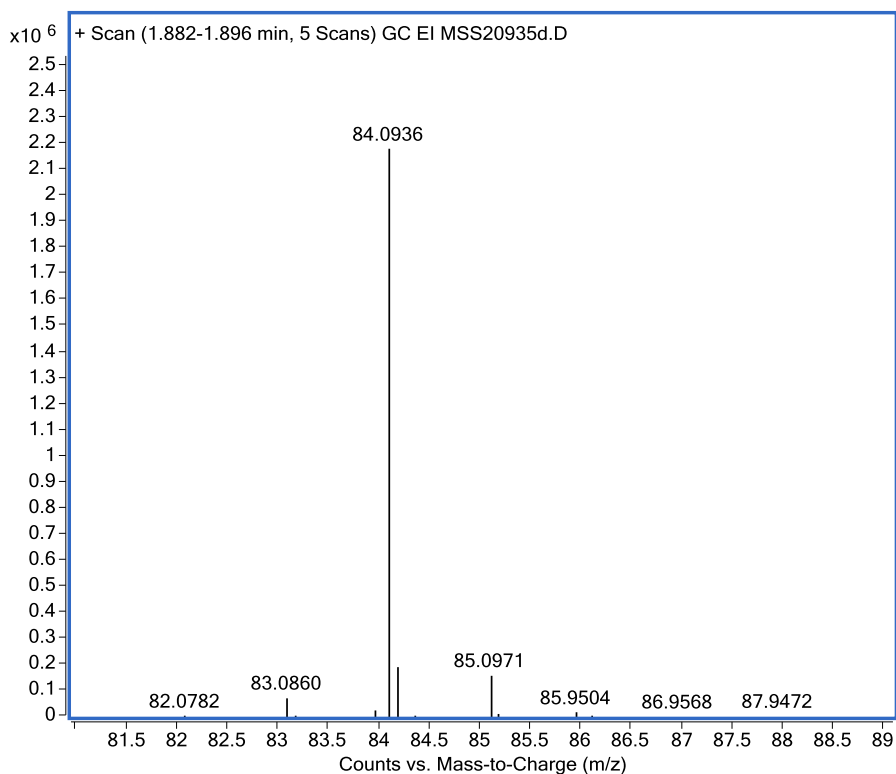
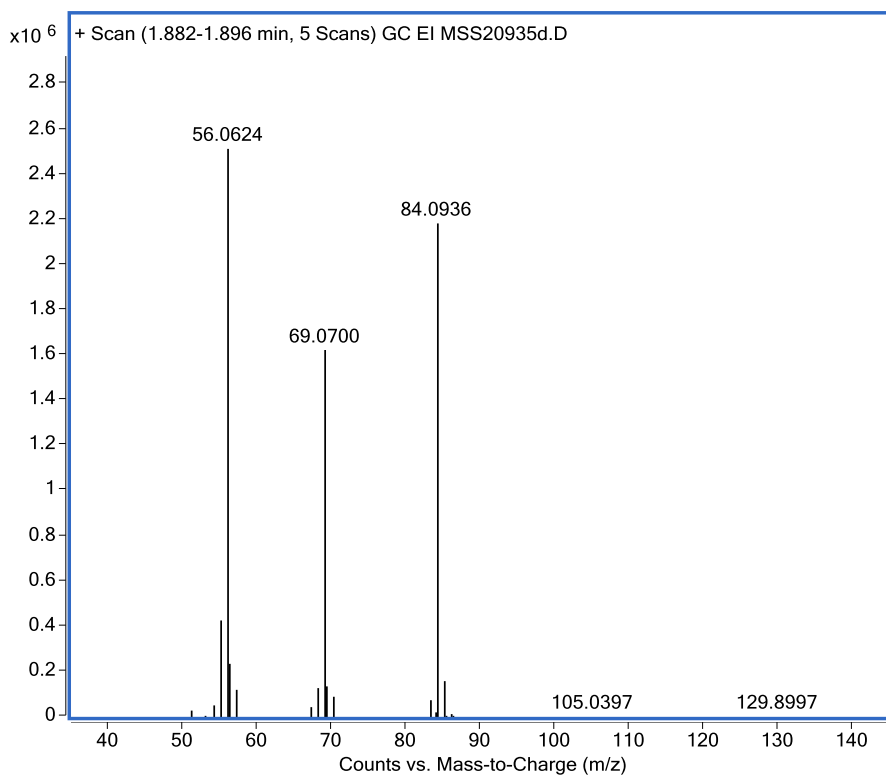


Figure S50: Mass spectrum of a commercially sourced sample of cyclohexane (top). Enlargement of $m/z = 81 - 89$ region (bottom).

S.3.4. Solution-state deuteration of [1-C₆H₈][BAr^F₄]

S.3.4.1. Procedure for the solution-state deuteration of [1-C₆H₈][BAr^F₄]

A solution of [1-C₆H₈][BAr^F₄] (10 mg) in CD₂Cl₂ in a thick walled NMR tube was freeze-pump-thaw degassed three times and backfilled with D₂ (1 bar, 298 K). Upon agitation the color of the solution changed from red to yellow. The volatile component was then isolated by trap-to-trap distillation. The product was identified as C₆H₈D₄, which is the product of deuteration across the double bonds only.

¹H NMR (CD₂Cl₂, 400 MHz, 183 K): δ 1.52 (br s, 4H, CH), 1.05 (br s, 4H, CH).

²H NMR (CD₂Cl₂, 61 MHz, 183 K): δ 1.54 (br s, 2D, CD), 1.08 (br s, 2D, CD).

S.3.4.2. GC EI-Mass Spectra

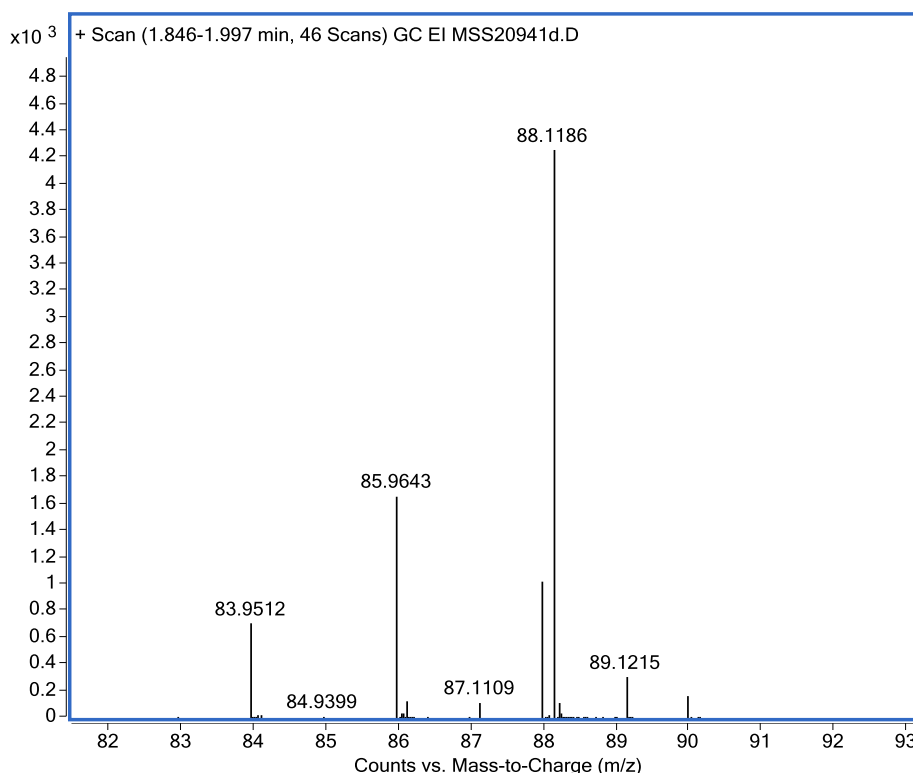
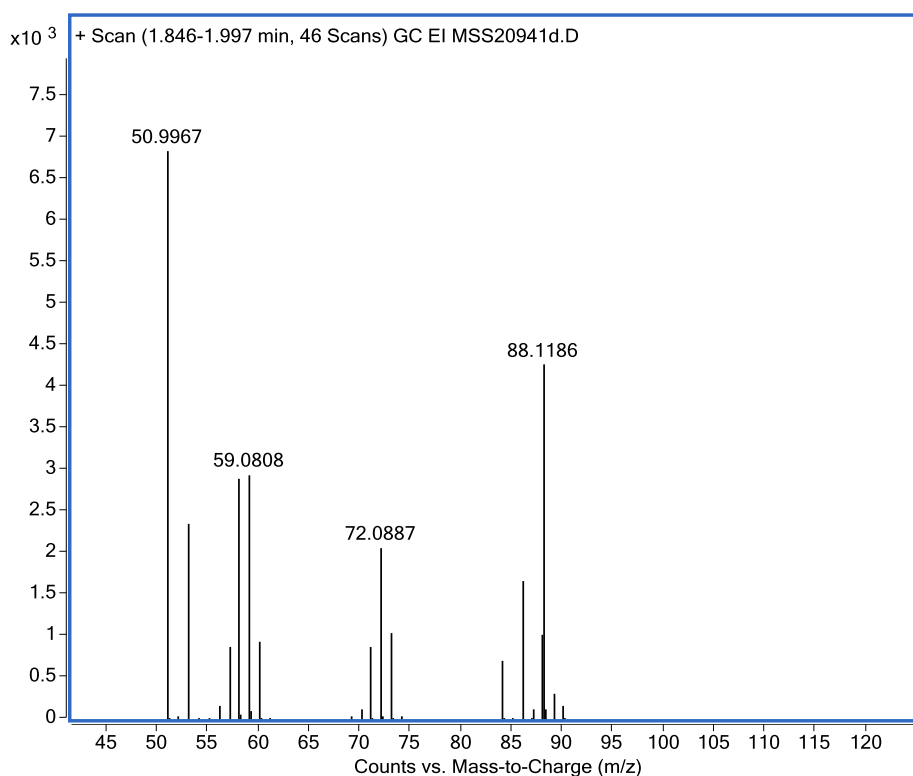
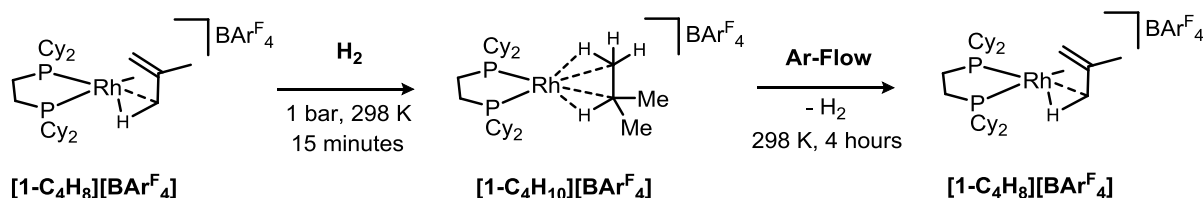


Figure S51: Mass spectrum of the isolated volatiles (top). Enlargement of $m/z = 81 - 89$ region (bottom). The signal at $m/z = 88.1186$ matches that of $C_6H_8D_4^+$ (calc. 88.1185). Signals at $m/z = 84$ and 86 are due to CH_2Cl_2 .

S.4. Alkane Dehydrogenation Experiments

S.4.1. Dehydrogenation of $[1\text{-C}_4\text{H}_{10}][\text{BAr}^{\text{F}}_4]$ to form $[1\text{-C}_4\text{H}_8][\text{BAr}^{\text{F}}_4]$

S.4.1.1. Solid-state dehydrogenation of $[1\text{-C}_4\text{H}_{10}][\text{BAr}^{\text{F}}_4]$ under *argon flow* to form $[1\text{-C}_4\text{H}_8][\text{BAr}^{\text{F}}_4]$



A J Young flask (ca. 50 mL) was charged with 50 mgs of block like crystalline material of $[1\text{-C}_4\text{H}_8][\text{BAr}^{\text{F}}_4]$. The flask was evacuated (1×10^{-2} mbar) and H_2 (1 bar, 298 K) was added for 15 minutes; to yield deep red coloured crystallites of $[1\text{-C}_4\text{H}_{10}][\text{BAr}^{\text{F}}_4]$.

A portion of this (approx. 10 mgs) was taken at this point, dissolved in CH_2Cl_2 and solution $^{31}\text{P}\{^1\text{H}\}$ NMR spectrum taken. No signals relating to $[1\text{-C}_4\text{H}_8][\text{BAr}^{\text{F}}_4]$ could be identified, only signals relating to $[1\text{-BAr}^{\text{F}}_4]$ and further solvent activated complexes – confirming the hydrogenation of total $[1\text{-C}_4\text{H}_8][\text{BAr}^{\text{F}}_4]$ had occurred.

The remaining solid was placed under an argon flow for 4 hours. This was done by placing an exit needle into the suba sealed J Young flask which was open to argon flow, with regulator pressure set at 0.5 bar. After 4 hours, the sample was sealed, transferred to a glove box and packed inside a solid state NMR rotor and analyzed $^{31}\text{P}\{^1\text{H}\}$ and $^{13}\text{C}\{^1\text{H}\}$ solid state NMR spectroscopy.

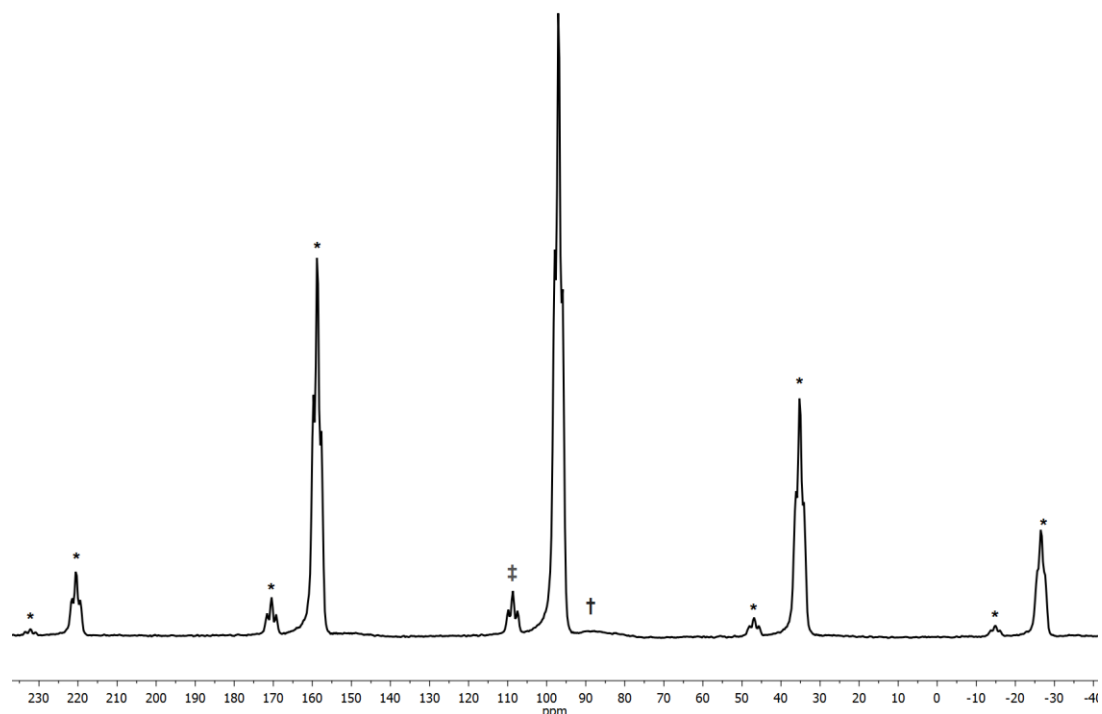
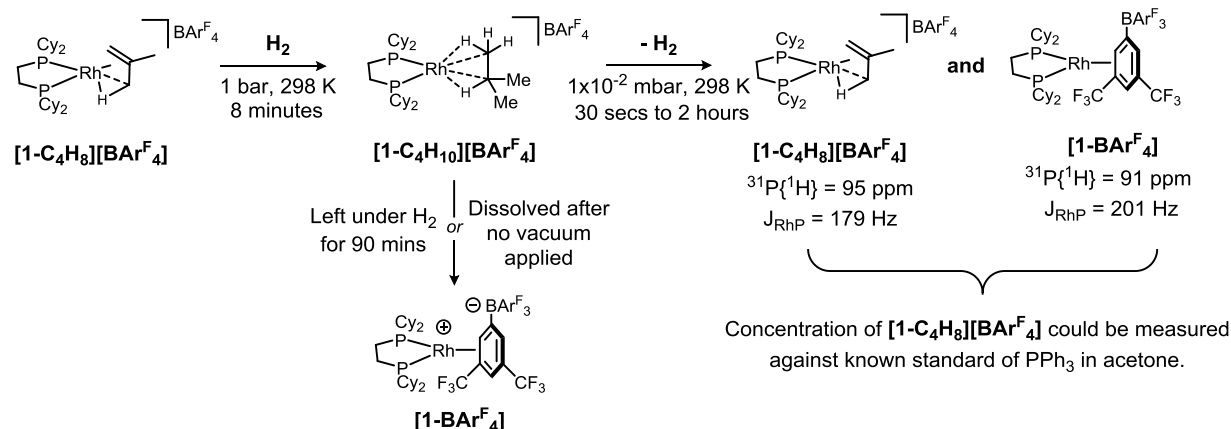


Figure S52: The $^{31}\text{P}\{^1\text{H}\}$ SSNMR (162 MHz, 295 K, 10 kHz spin rate) spectrum of the dehydrogenation of powdered microcrystalline $[\mathbf{1-C}_4\text{H}_{10}][\text{BAr}^{\text{F}}_4]$ under argon flow to form $[\mathbf{1-C}_4\text{H}_8][\text{BAr}^{\text{F}}_4]$ (approx. 90 %). The rotor is packed under an argon atmosphere. The resonances \ddagger marked are assigned to $[\mathbf{1-C}_4\text{H}_{10}][\text{BAr}^{\text{F}}_4]$ (approx. 7%), from incomplete dehydrogenation and the resonances marked \dagger are assigned to the decomposition product of $[\mathbf{1-BAr}^{\text{F}}_4]^{\text{S}2}$ (approx. 3 %), formed during the initial hydrogenation. The resonances marked * are due to spinning sidebands.

S.4.1.2. Kinetics of the solid-state [1-C₄H₁₀][BAR^F₄] dehydrogenation under vacuum to form [1-C₄H₈][BAR^F₄]



A J Young NMR tube (~ 2.1 mL) was charged with 10 mgs powdered crystalline material of [1-C₄H₈][BAR^F₄]. The tube was evacuated (1×10^{-2} mbar) and H₂ (1 bar, 298 K) was added for 8 minutes to yield deep red powder of [1-C₄H₁₀][BAR^F₄]. The samples were placed under vacuum (1×10^{-2} mbar) for varying times between 0 seconds and 2 hours, then dissolved in CD₂Cl₂ and analyzed by $^{31}P\{^1H\}$ NMR spectroscopy.

The above procedure was repeated on numerous occasions, varying the period of time the sample was exposed to vacuum at room temperature. When time = 0 seconds, no signals relating to [1-C₄H₈][BAR^F₄] could be identified, confirming the complete hydrogenation of the sample.

After dissolution in CD₂Cl₂, two doublets were observed in the $^{31}P\{^1H\}$ NMR spectra at 298 K; the dehydrogenation product [1-C₄H₈][BAR^F₄] (δ 95) and decomposition product [1-BAR^F₄]^{S2} (δ 91). The concentration of re-formed [1-C₄H₈][BAR^F₄] could be accurately measured by use of an separate, sealed capillary of PPh₃ in acetone.

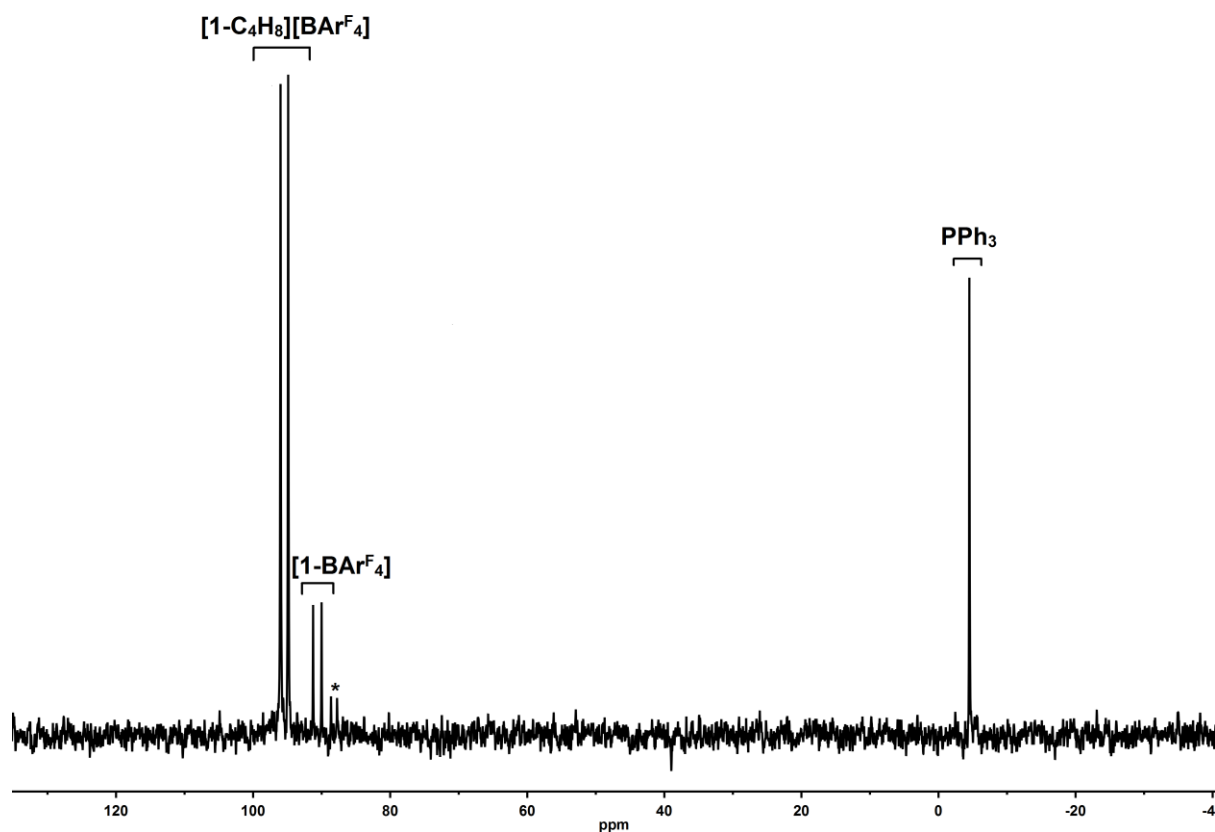


Figure S53: Example of a solution $^{31}\text{P}\{^1\text{H}\}$ NMR (CD_2Cl_2 , 298 K, 162 MHz) spectra of the dehydrogenation of $[\text{1-C}_4\text{H}_8][\text{BAr}^{\text{F}}_4]$ under vacuum for 1 hour. Signals can be observed from $[\text{1-C}_4\text{H}_8][\text{BAr}^{\text{F}}_4]$ (δ 95, d., 179 Hz) and $[\text{1-BAr}^{\text{F}}_4]$ (δ 91, d., $J_{\text{RHP}} = 201$ Hz) with PPh_3 (δ -6, in acetone within sealed capillary). * = a further, unidentified CH_2Cl_2 decomposition product of $[\text{1-BAr}^{\text{F}}_4]$.^{S2}

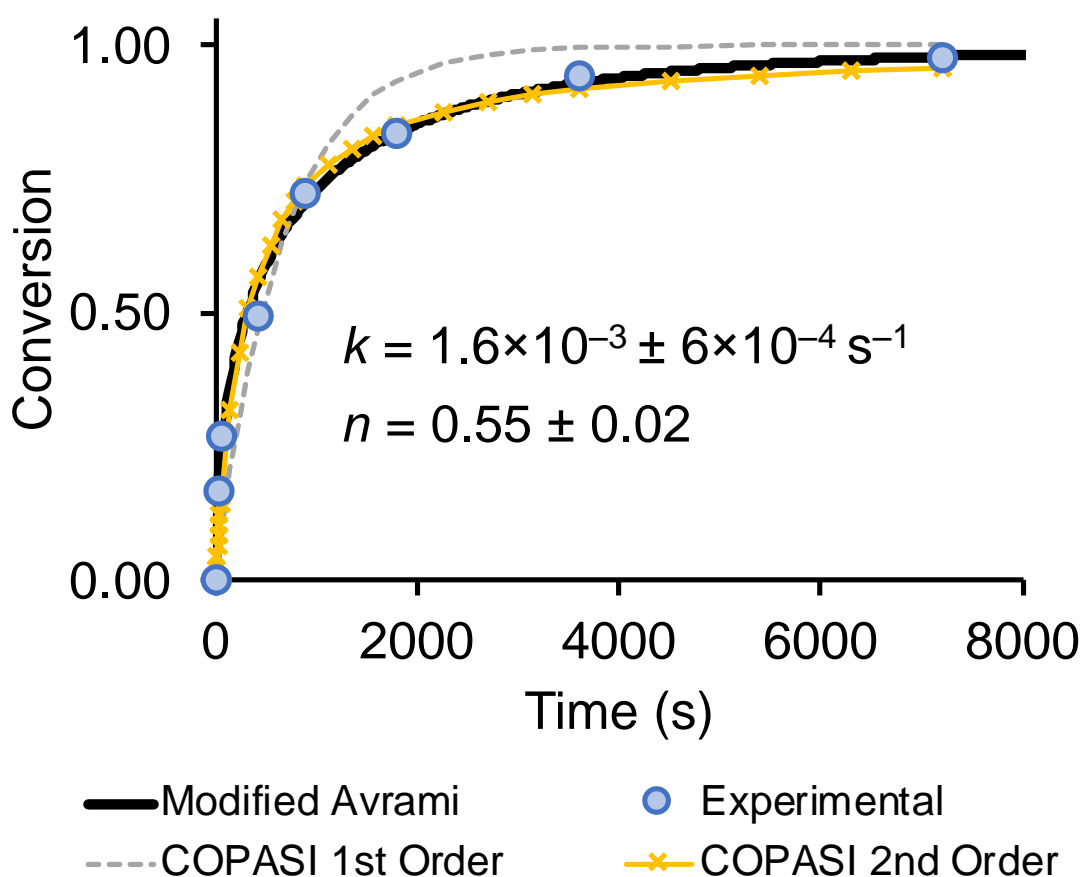


Figure S54: Modified JAMK Plot of Conversion^{S9} ($f = (1 - e^{-(kt)^n})$) versus time for the dehydrogenation of $[1\text{-C}_4\text{H}_{10}][\text{BAr}^{\text{F}_4}]$ to $[1\text{-C}_4\text{H}_8][\text{BAr}^{\text{F}_4}]$ in the solid-state. Where k = growth rate constant and n = Avrami exponent.

Shown with comparison to a COPASI^{S10} modelled first order process for dehydrogenation ($[1\text{-C}_4\text{H}_{10}][\text{BAr}^{\text{F}_4}] \rightarrow [1\text{-C}_4\text{H}_8][\text{BAr}^{\text{F}_4}]$, $k = 1.5(2) \times 10^{-3} \text{ s}^{-1}$) and a second order process ($2 \times [1\text{-C}_4\text{H}_{10}][\text{BAr}^{\text{F}_4}] \rightarrow [1\text{-C}_4\text{H}_8][\text{BAr}^{\text{F}_4}] + [1\text{-C}_4\text{H}_{10}][\text{BAr}^{\text{F}_4}]$, $k = 1.6(2) \times 10^{-4} \text{ M}^{-1}\text{s}^{-1}$). Initial concentration of dissolved samples = 19.8 mM (conversion = 0 %).

S.4.1.3. *Single-Crystal to Single-Crystal* dehydrogenation of [1-C₄H₁₀][BAr^F₄] under *vacuum* to form [1-C₄H₈][BAr^F₄] for an x-ray diffraction study

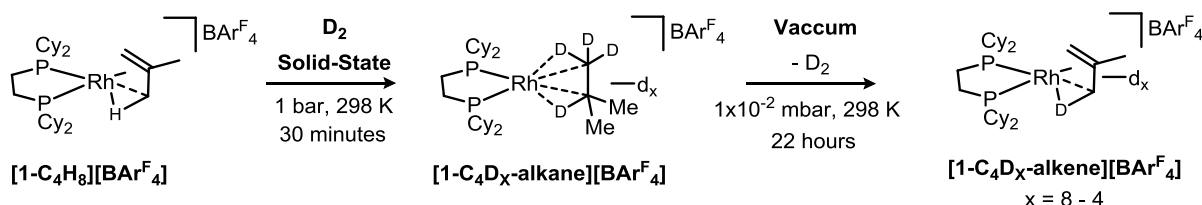
A J Young NMR tube (~ 2 mL) was charged with 10 mgs of orange coloured, block like crystalline material of [1-C₄H₈][BAr^F₄]. The flask was evacuated (1×10^{-2} mbar) and H₂ (1 bar, 298 K) was added for 15 minutes to yield deep red crystallites of [1-C₄H₁₀][BAr^F₄]. The sample was then placed under vacuum (2×10^{-2} mbar) for 4 hours, to which the colour of the crystals after this time had returned to orange.

The crystalline material was then coated with Fomblin® Y oil under an argon flush, a suitable crystal was rapid selected and then transferred to the cryostream of a diffractometer and an x-ray diffraction study was undertaken.

The crystals were of sufficient quality to be collected, and solved, as [1-C₄H₈][BAr^F₄]-SC-SC. For further details of the structural study, please refer to section S.5.

S.4.2. Dedeuteration of [1-C₄D_x-alkane][BAR^F₄] under vacuum to form [1-C₄D_x-alkene][BAR^F₄]

S.4.2.1. Solid state dedeuteration of [1-C₄D_x-alkane][BAR^F₄] under vacuum to form [1-C₄D_x-alkene][BAR^F₄]



A J Young NMR tube (~ 3 mL) was charged 30 mgs of block-like crystallites of [1-C₄H₈][BAR^F₄]. The flask was evacuated (1×10^{-2} mbar) and D₂ (1 bar, 298 K) was added to yield red crystalline powder of [1-C₄D_x-alkane][BAR^F₄], and left for 30 minutes. After this time, the sample was placed under vacuum at room temperature for 22 hours to yield [1-C₄D_x-alkene][BAR^F₄], where (x = 8 – 6).

S.4.2.2. Characterization data for [1-C₄D_x-alkene][BAR^F₄]

¹H NMR (CD₂Cl₂, 298 K, 400 MHz): δ 7.73 (s, 8H, ortho-ArH), 7.57 (s, 4H, para-ArH), 5.32 (CDHCl₂), 2.09-1.64 (br m, 24H, overlapping aliphatic CH), 1.40-1.0.96 (br m, 20H, overlapping aliphatic CH).

¹H NMR (CD₂Cl₂, 183 K, 400 MHz): δ 7.75 (s, 8H, ortho-ArH), 7.56 (s, 4H, para-ArH), 5.32 (CDHCl₂), 2.22-1.38 (br m, 24H, overlapping aliphatic CH), 1.38-0.63 (br m, 24H, overlapping aliphatic CH).

²H NMR (CD₂Cl₂, 298 K, 400 MHz): δ 1.55 (br. s)

²H NMR (CD₂Cl₂, 183 K, 400 MHz): δ 0.72 (br. m)

³¹P{¹H} NMR (CD₂Cl₂, 298 K, 162 MHz): δ 95.3 (d, J_{RhP} 179 Hz).

³¹P{¹H} NMR (CD₂Cl₂, 183 K, 162 MHz): δ 97.6 (d. of d., J_{RhP} 201 Hz, J_{PP} 26 Hz), 93.6 (d. of d., J_{RhP} 158 Hz, J_{PP} 26 Hz) .

ESI-MS: Not stable under mass spectrometric conditions (20eV, 333K). Species with appropriate isotopic distributions at m/z found = 581.240, calculated to [(Cy₂PCH₂CH₂PCy₂)Rh(N₂)₂]⁺ (581.242). There is no evidence for the nitrogen compound in bulk samples so it is assumed to form via an in-situ ESI-MS process.

S.4.2.3. NMR Spectra of [1-C₄D_x-alkene][BAr^F₄]

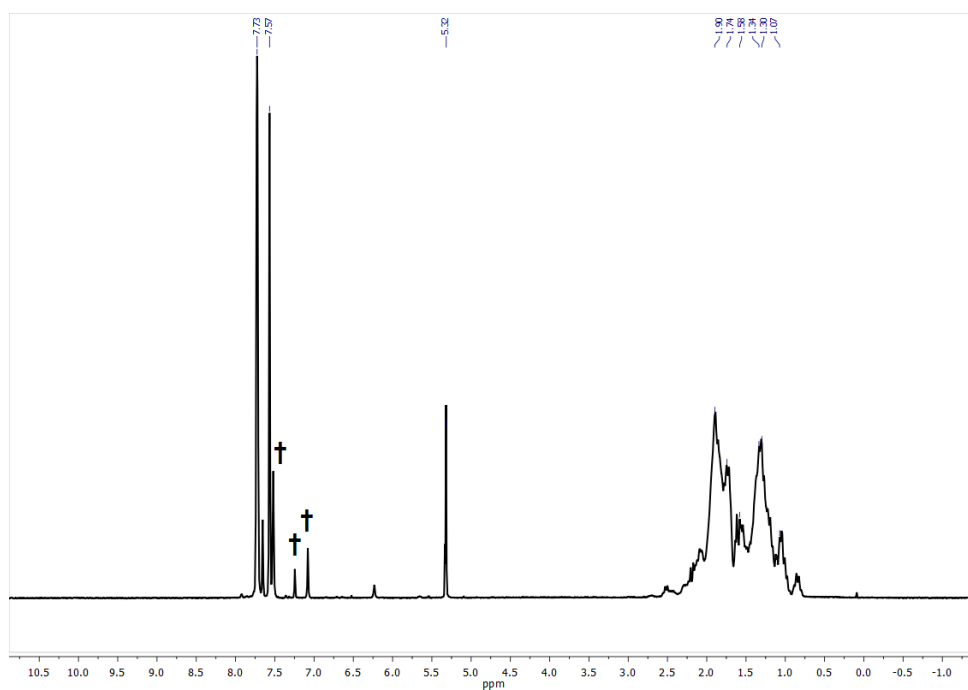


Figure S55: The solution ¹H NMR (CD₂Cl₂, 298 K, 400 MHz) spectrum of [1-C₄D_x-alkene][BAr^F₄]. Signals marked † are due to [1-BAr^F₄].

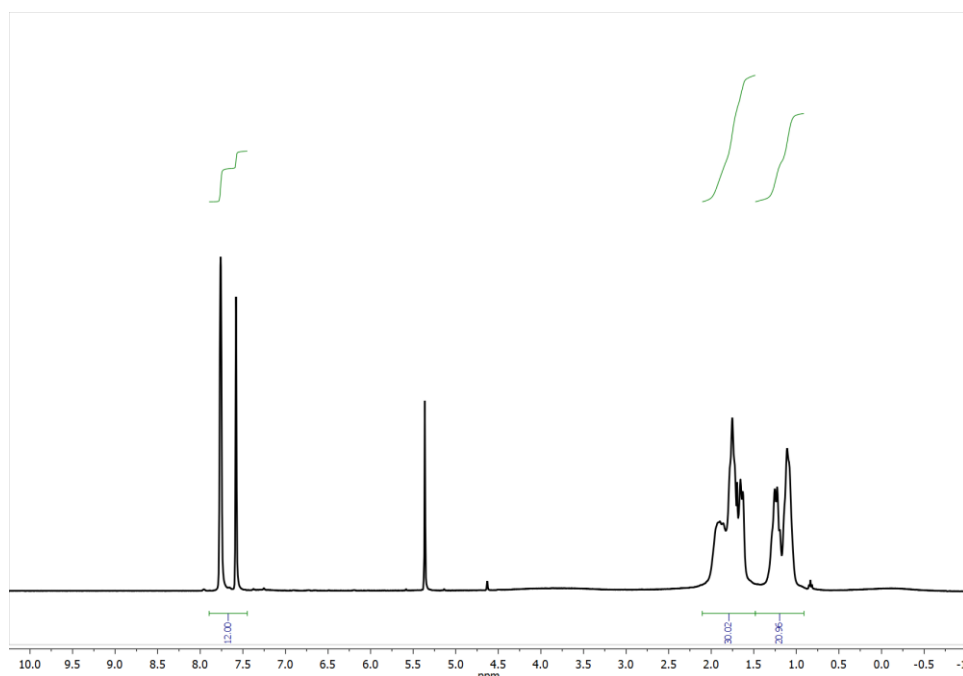


Figure S56: The solution ¹H NMR (CD₂Cl₂, 183 K, 400 MHz) spectrum of [1-C₄D_x-alkene][BAr^F₄]. Proton-alkene signals are not observed, unlike in Figure S2 of [1-C₄H₃][BAr^F₄] at 183 K, suggestive of isotopic substitution of these protons.

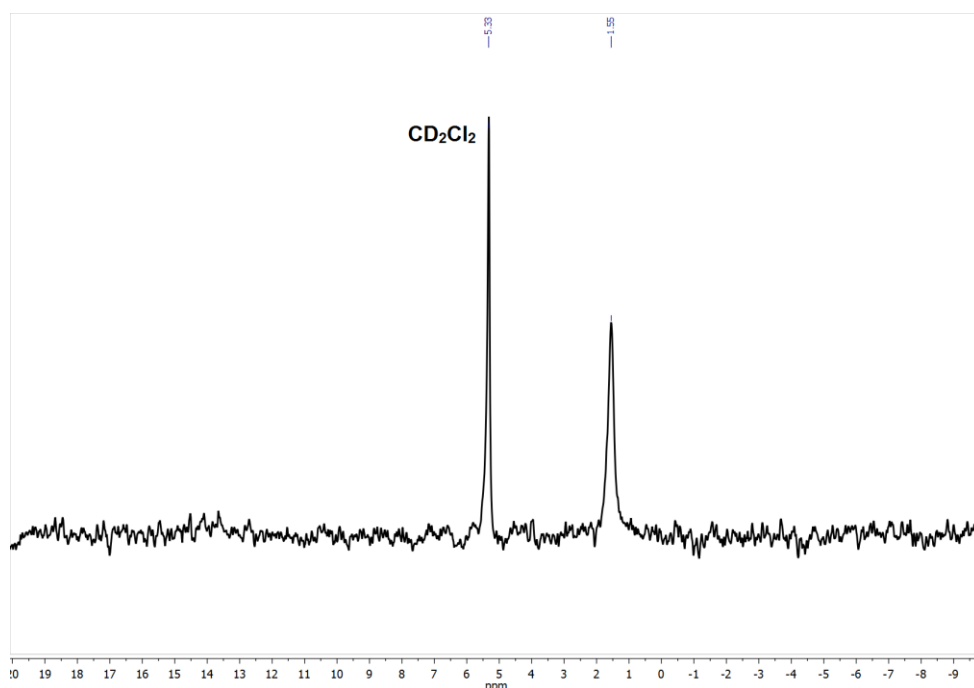


Figure S57: The solution ^2H NMR (CH_2Cl_2 , 298 K, 61 MHz) spectrum of **[1- C_4D_x -alkene][BAr^{F_4}]**.

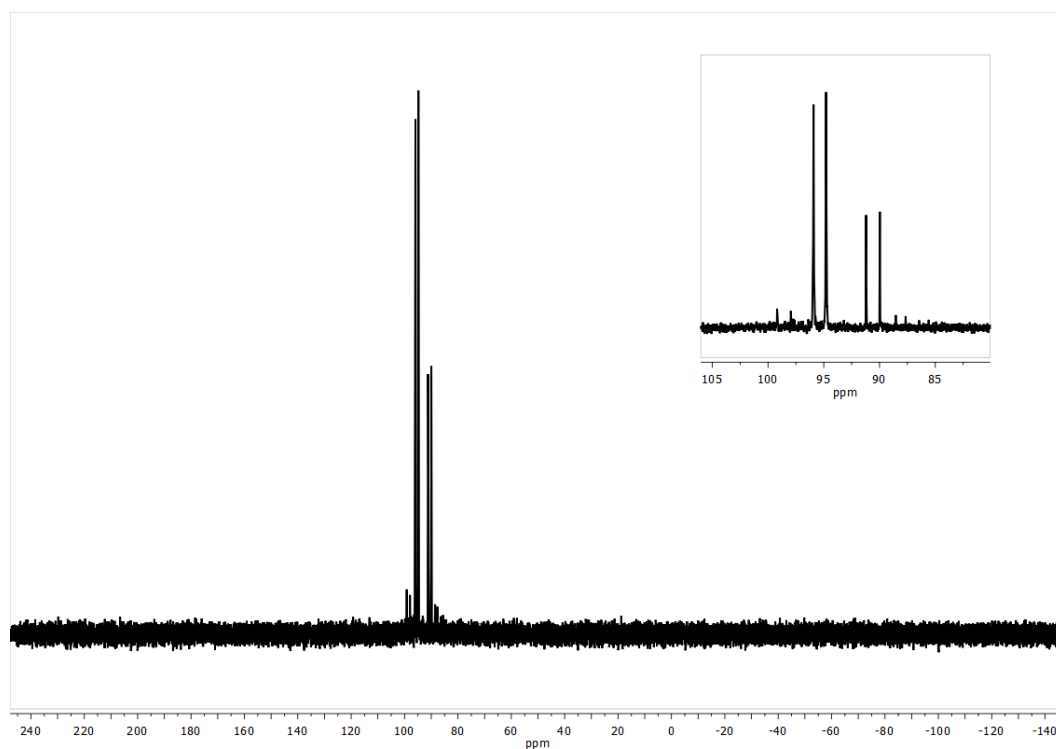


Figure S58: The solution ^{31}P $\{^1\text{H}\}$ NMR (CD_2Cl_2 , 298 K, 162 MHz) spectrum of **[1- C_4D_x -alkene][BAr^{F_4}]**, with $\sim 40\%$ decomposition to **[1- BAr^{F_4}]**. This decomposition to **[1- BAr^{F_4}]** has occurred after an extended time under D_2 , see section S.2.2.6.

S.4.2.4. Liberation of bound isobutene-d_x (x = 8 – 6)

After preparation of [1-C₄D_x-alkene][BAr^F₄] following the procedure in section S.4.2.1., [1-C₄D_x-alkene][BAr^F₄] was dissolved in CD₂Cl₂ in a J Young NMR tube and the solution was freeze pump thawed degassed three times. CO (1 bar, 298K) was added and after 2 minutes, a vibrant yellow solution had formed of [Rh(dcpe)(CO)₂][BAr^F₄], confirmed by solution ³¹P{¹H} NMR spectroscopy (δ 85.22, d, J_{RhP} = 116 Hz),^{S7} as well as liberated isobutene-d_x. The solvent and volatiles were distilled into a fresh NMR tube and analyzed by ¹H, ²H, ¹³C {¹H} NMR spectroscopy and GC EI-MS.

S.4.2.5. Characterization data for isobutene-d_x (x = 8 – 6)

¹H NMR (CD₂Cl₂, 400 MHz, 298 K): Residual protio Signals at δ 4.69 (s, C=CH₂), 1.71 br. m., CH₃)

²H NMR (CD₂Cl₂, 61 MHz, 298 K): δ 4.71 (br s, 2D, CD), 1.70 (br s, 5.6D, CD₃).

¹³C{¹H} NMR (CD₂Cl₂, 126 MHz, 298 K): δ 142.7 (s, C=CD₂), 110.1 (m, C=CD₂, J_{C-D} = 22.8), 23.4 (m, -CD₃, J_{C-D} = 20.1).

ESI-MS found (calc. for C₄D₈⁺): m/z 64.112 (64.1128).

S.4.2.6. Solution NMR for isobutene-d_x (x = 8 – 6)

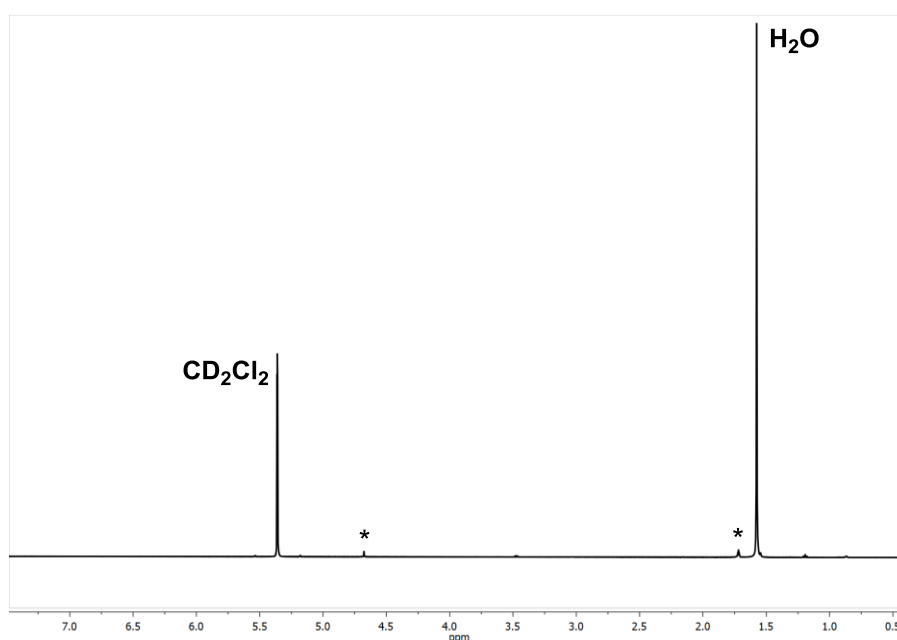


Figure S59: The solution ¹H NMR (C₆D₆, 298 K, 400 MHz) spectrum of isolated volatiles of isobutene-d_x. The signals marked * are from residual protio-isobutene.

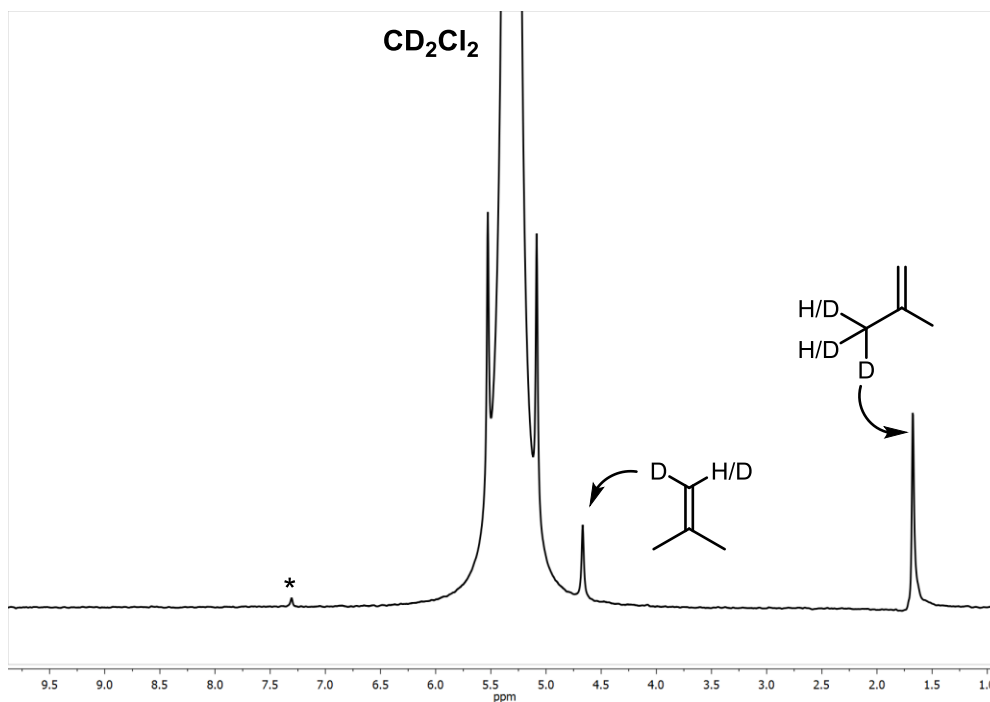


Figure S60: The solution ^2H NMR (CD_2Cl_2 , 298 K, 61 MHz) spectrum of isolated volatiles of isobutene- d_x . The signals marked * are from a CDCl_3 impurity.

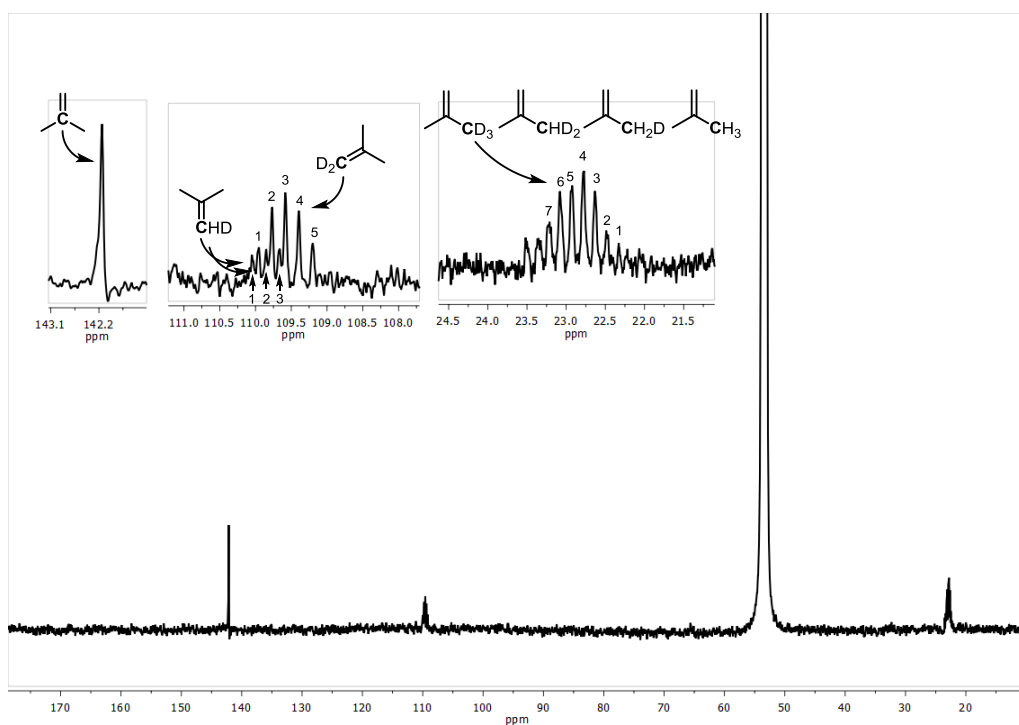


Figure S61: The solution $^{13}\text{C}\{^1\text{H}\}$ NMR (CD_2Cl_2 , 298 K, 126 MHz) spectrum of isolated volatiles of isobutene- d_x .

S.4.2.7. GC EI-MS Mass Spectra of isobutene-d_x (x = 8 – 6)

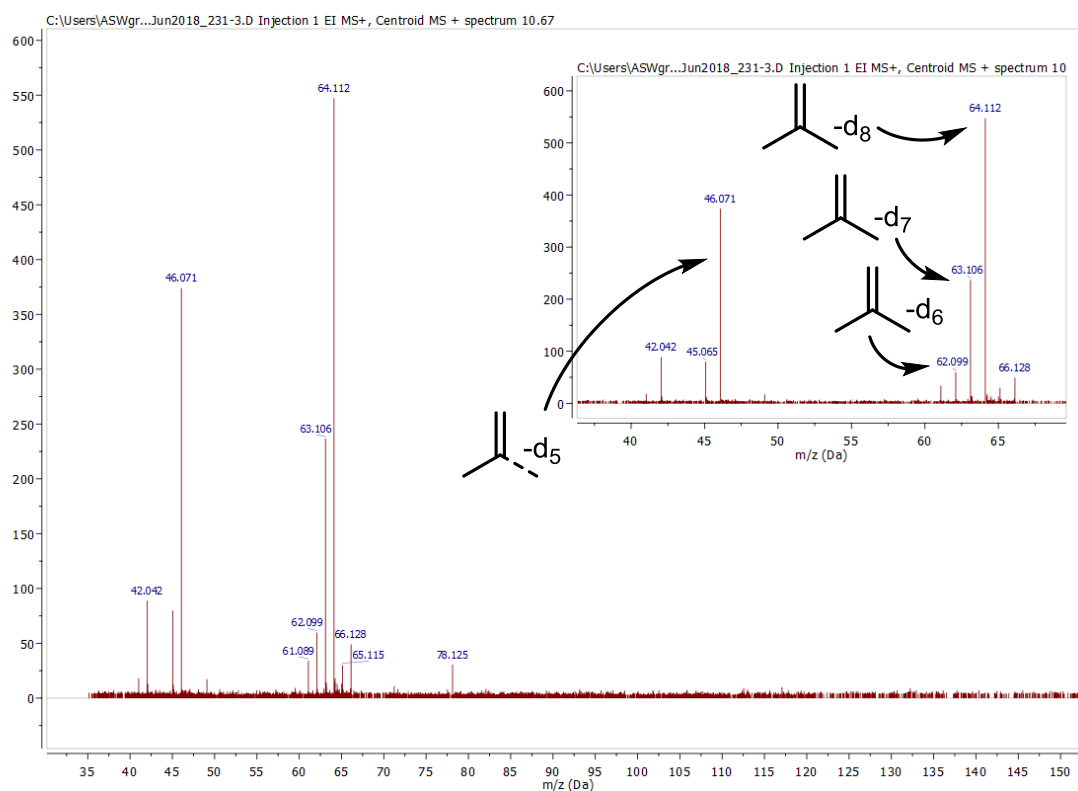


Figure S62: Mass spectrum of the isolated isobutene-d_x with enlargement of m/z = 35 – 70 region. The molecular ion peak is that of C₄D₈⁺, m/z = 64.112 (calc. 64.1128) with greatest intensity fragment of C₃D₅⁺, m/z = 46.071 (calc. = 46.0705). The 3 major isotopic contributions are that of C₄D₈⁺, C₄HD₇⁺ and C₄H₂D₆⁺, as shown by signals at 64.112, 63.106 and 62.099 respectively.

S.4.2.8. Simulated solution $^{13}\text{C}\{^1\text{H}\}$ NMR spectra of isobutene- d_x ($x = 8 - 6$)

As per section S.3.1.4., the solution $^{13}\text{C}\{^1\text{H}\}$ NMR of isolated isobutene- d_x ($x = 8 - 6$) (Figure S61) was de-convoluted using the Spin Simulation function on MestReNova, shown in Figure S63. Line width was kept at 4.5 Hz throughout, only changing levels of population in each simulated environment.

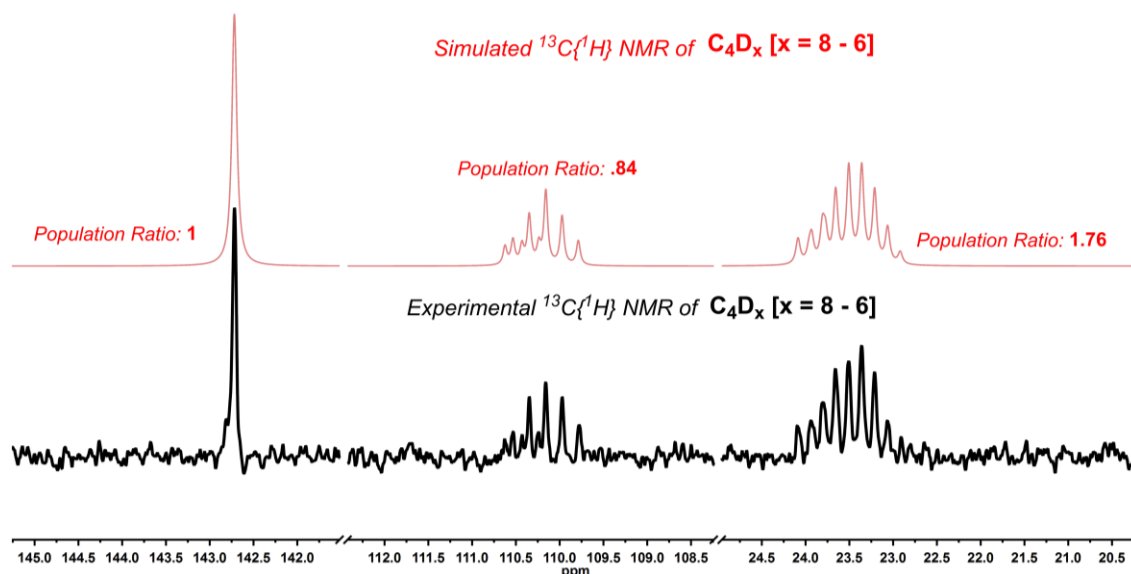


Figure S63: Simulated and partial experimental solution $^{13}\text{C}\{^1\text{H}\}$ NMR spectra of isobutene- d_x ($x = 8 - 6$). The three separate ^{13}C -environments are assigned to methyl ^{13}C ($\delta \sim 23$), methylene ^{13}C ($\delta \sim 110$) and quaternary ^{13}C ($\delta \sim 142$).

The population ratio of each ^{13}C environment in the simulated spectra was calculated from the sum of each individual $-\text{CD}_x\text{H}_y$ component in each environment compared between environments. These ratios suggest an approximate 1:1:2 ratios of carbon environments, in line with that of isobutene.

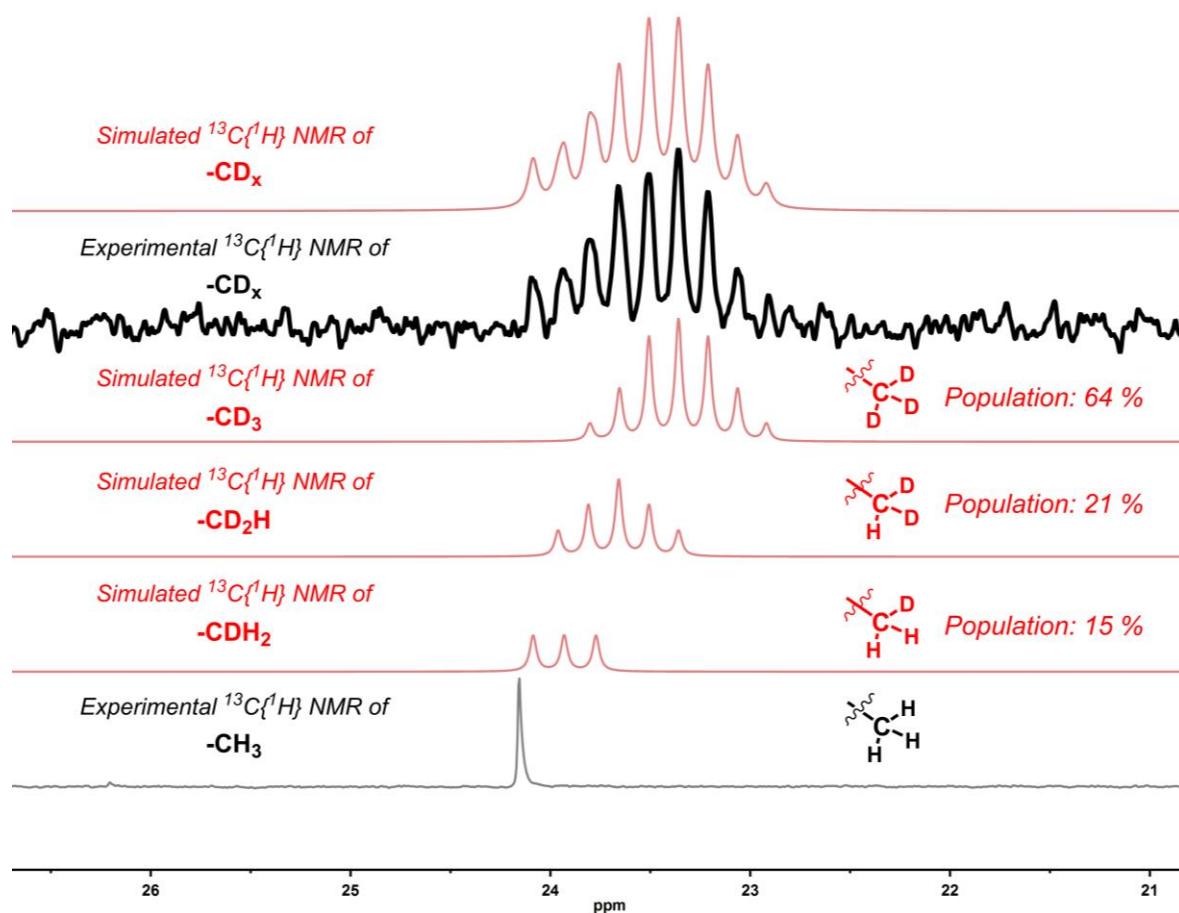


Figure S64: Three systems were used to recreate the multiplet at δ 23.5 from the methyl environment, modelling $-\text{CD}_3$, $-\text{CD}_2\text{H}$ and $-\text{CDH}_2$ groups. No evidence of $-\text{CH}_3$ could be seen in the experimental data so was not modelled. The simulation from this environments suggests that 64 % of $-\text{CX}_3$ groups are were of $-\text{CD}_3$ in nature, 21 % $-\text{CD}_2\text{H}$ and 15 % $-\text{CDH}_2$.

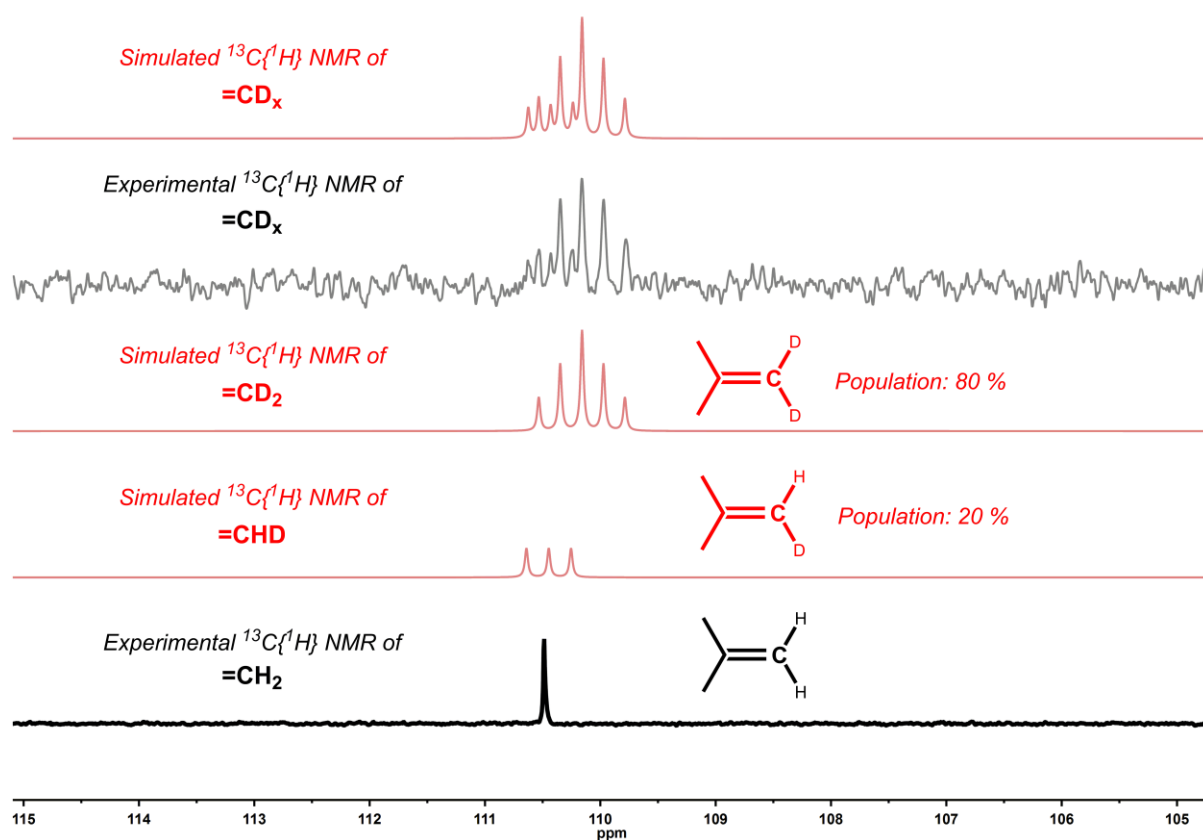


Figure S65: Two systems were used to recreate the multiplet at δ 110 from the methylene environment, modelling $=\text{CD}_2$ and $=\text{CDH}$ groups. No evidence of $=\text{CH}_2$ could be seen in the experimental data so was not modelled. The simulation from these environments suggests that 80 % of $=\text{CD}_x$ groups are $=\text{CD}_2$, and 20 % = $=\text{CDH}$.

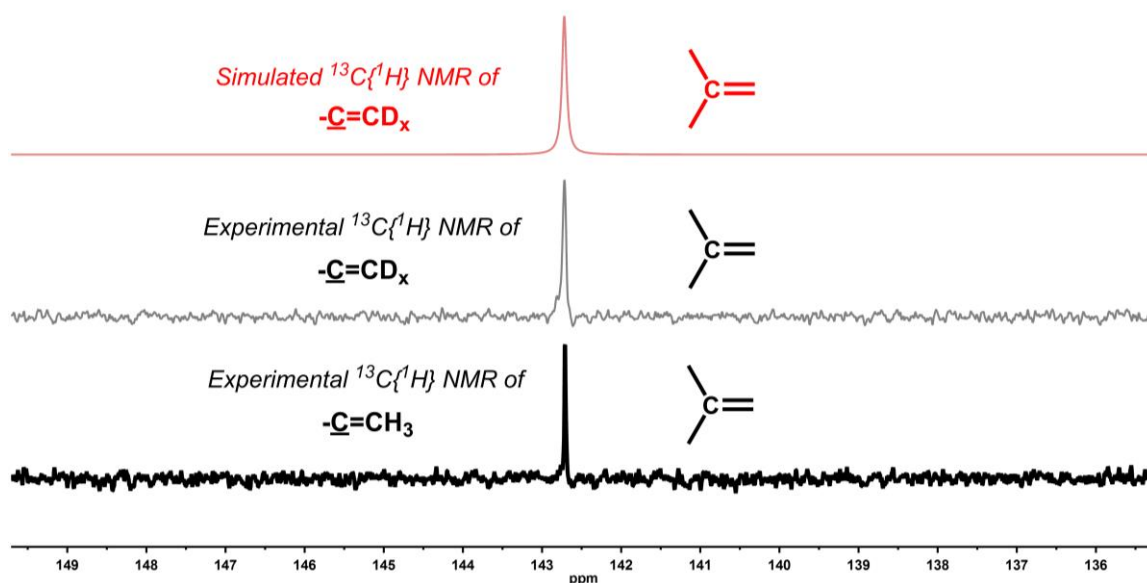


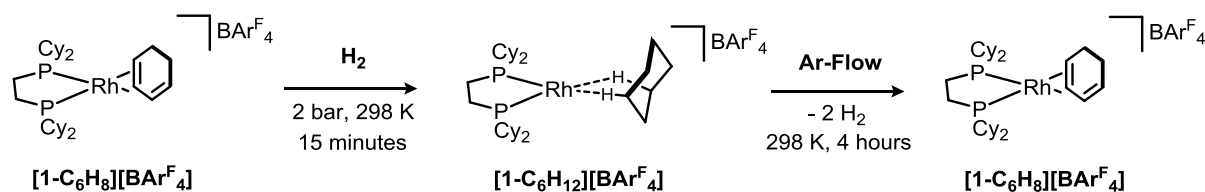
Figure S66: The singlet from the quaternary carbon at δ 142 was also modeled. This simulation was used as an approximate internal standard. The slight variation in chemical shift between the deuterated and proto isobutene ($\delta \sim .05$) we have assign to secondary isotope effects.

Overall, the three figures (S64 – S66) describe the full calculated breakdown of deuterated isobutene, confirming a high level of deuteration incorporation. This is consistent with the GC EI-MS shown in Figure S62, which showed a high amount of deuterated isobutene (C_4D_8^+ to C_4D_6^+).

However, overall, lower levels of deuteration are observed compared to isobutane- d_x (Section S.3.1.), as consistent with reduction of experimental time of $[\mathbf{1-C}_4\text{H}_8][\text{BAR}^{\text{F}}_4]$ under an D_2 atmosphere (30 vs 90 mins), see sections S.3.1. and S.4.2.

S.4.3. Solid-state dehydrogenation of [1-C₆H₁₂][BAr^F₄] to form [1-C₆H₈][BAr^F₄]

S.4.3.1. Solid-state dehydrogenation of [1-C₆H₁₂][BAr^F₄] under *argon flow* to form [1-C₆H₈][BAr^F₄]



A powdered microcrystalline sample of [1-C₆H₈][BAr^F₄] (35 mg) was packed in a 3.2 mm SSNMR rotor, inside an argon filled glove box. The rotor was then placed in a custom built glass J Young flask^{S7} and the sample was then exposed to H₂ (1 bar, 298 K) for 20 minutes, to yield deep red crystalline powder of [1-C₆H₁₂][BAr^F₄].

The sample was then placed under an argon flow for 18 hours. This was done by placing an exit needle into the suba sealed J Young flask which was open to argon flow, with regulator pressure set at 0.5 bar. After 18 hours, the rotor cap was fitted under a flush of Ar, and immediately transferred to the bore of a SSNMR spectrometer and analyzed ³¹P{¹H} and ¹³C{¹H} solid state NMR spectroscopy.

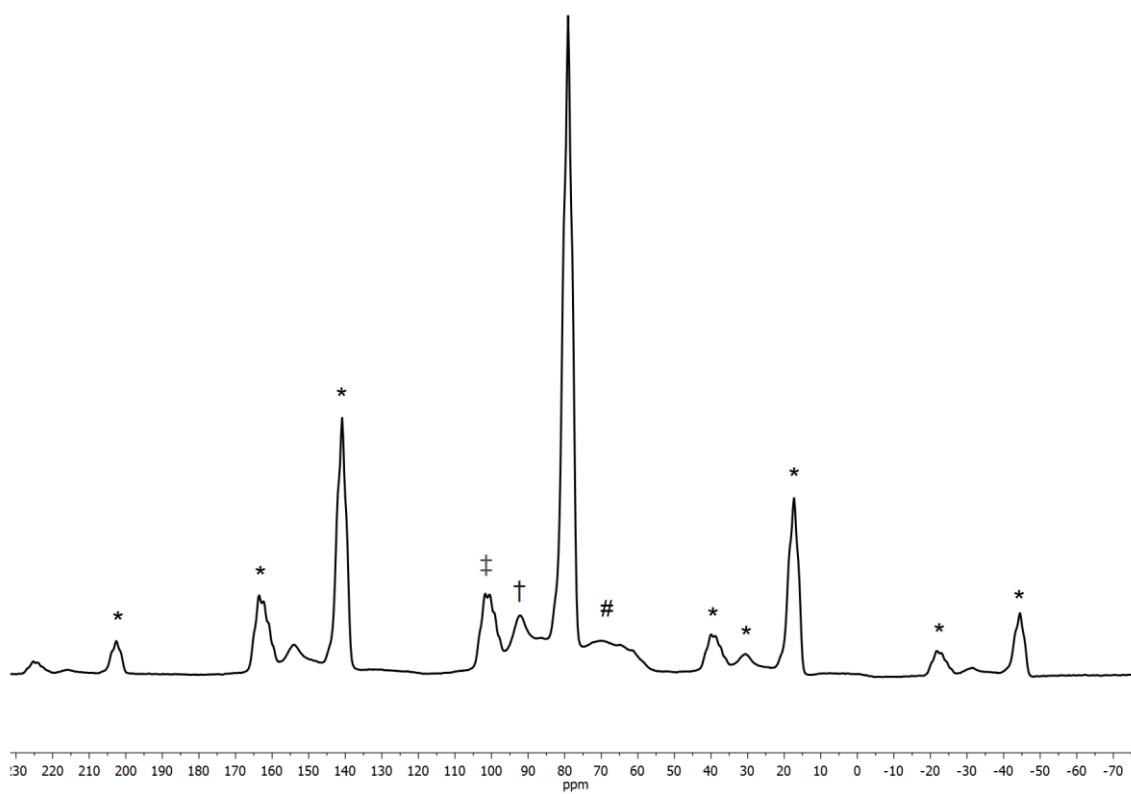


Figure S67: The $^{31}\text{P}\{^1\text{H}\}$ SSNMR (162 MHz, 295 K, 10 kHz spin rate) spectrum of the dehydrogenation of powdered microcrystalline $[\mathbf{1-C}_6\text{H}_{12}][\text{BAr}^{\text{F}}_4]$ under an argon flow. The resonance at δ 78 corresponds to $[\mathbf{1-C}_6\text{H}_8][\text{BAr}^{\text{F}}_4]$, in comparison to clean $[\mathbf{1-C}_6\text{H}_8][\text{BAr}^{\text{F}}_4]$ shown in Figure S21. The resonances ‡ marked are assigned to $[\mathbf{1-C}_6\text{H}_{10}][\text{BAr}^{\text{F}}_4]$, from incomplete dehydrogenation and † from $[\mathbf{1-BAr}^{\text{F}}_4]$, formed from a side reaction when under H_2 atmosphere. The resonances marked # are assigned to as yet unidentified decomposition products. The resonances marked * are due to spinning sidebands.

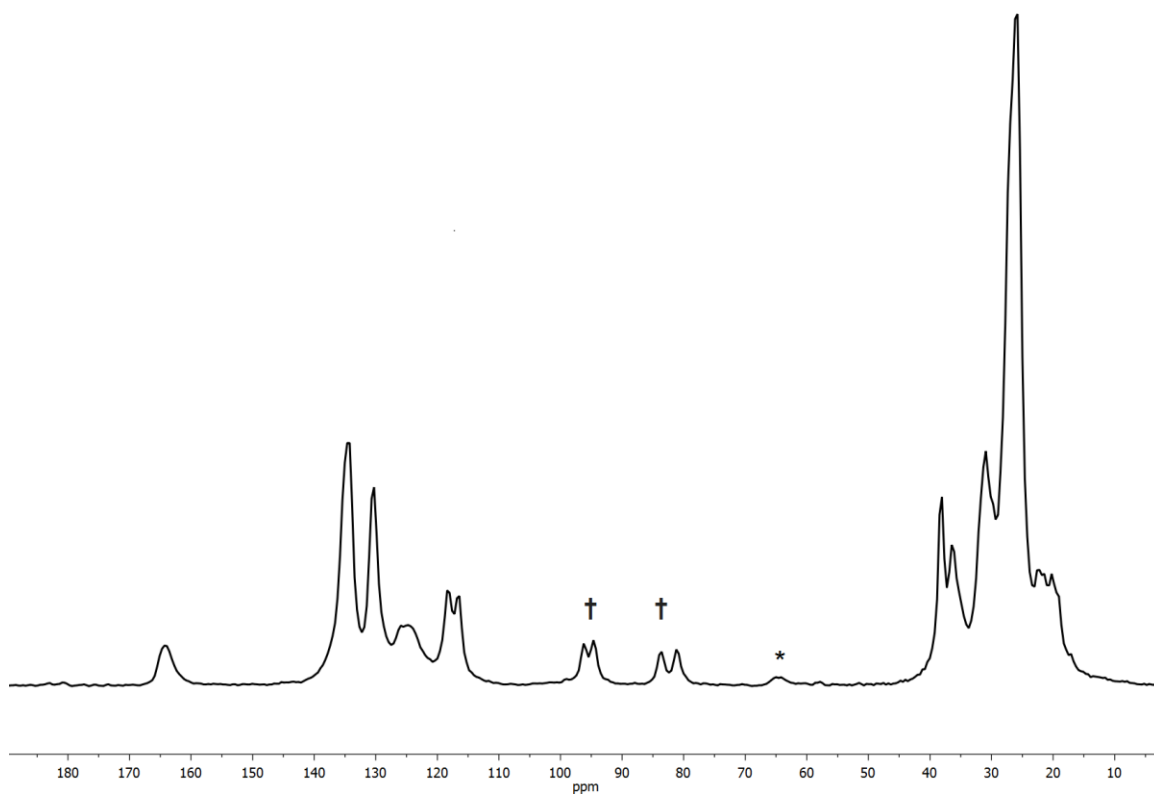


Figure S68: The $^{13}\text{C}\{^1\text{H}\}$ SSNMR (100 MHz, 295 K, 10 kHz spin rate) spectrum of the dehydrogenation of powdered microcrystalline $[1\text{-C}_6\text{H}_{12}][\text{BAr}^{\text{F}}_4]$ under an argon flow. The resonances marked † correspond to the alkene signals in $[1\text{-C}_6\text{H}_8][\text{BAr}^{\text{F}}_4]$, in comparison to clean $[1\text{-C}_6\text{H}_8][\text{BAr}^{\text{F}}_4]$ shown in Figure S22. The resonances marked * are due to spinning sidebands.

S.4.3.2. Solid-state dehydrogenation of $[1-\text{C}_6\text{H}_{12}][\text{BAr}^{\text{F}}_4]$ under vacuum to form $[1-\text{C}_6\text{H}_8][\text{BAr}^{\text{F}}_4]$

A J Young NMR tube was charged with powdered microcrystalline $[1-\text{C}_6\text{H}_8][\text{BAr}^{\text{F}}_4]$ (10 mgs) in an argon filled glove box. The sample was then evacuated (2×10^{-2} mbar) and backfilled with H_2 (1 bar, 298 K) for 15 minutes. The sample was then cooled to 77 K in liquid nitrogen and evacuated. After a steady vacuum was achieved (2×10^{-2} mbar) the sample was rapidly warmed to 298 K and a timer was simultaneously started. After a set period of time (5 mins to 960 mins), the sample was cooled to 77 K in liquid nitrogen and CH_2Cl_2 (0.40 mL) was condensed into the NMR tube. The sample was expediently thawed and rapidly transferred to the bore of a precooled (183 K) NMR spectrometer which was previously locked and shimmed to a sample of CD_2Cl_2 . A $^{31}\text{P}\{^1\text{H}\}$ NMR spectrum was then acquired.

The above procedure was repeated on numerous occasions, varying the period of time the sample was exposed to vacuum at room temperature.

Note: Vacuum pressure was used to normalize the conditions, opposed to an argon flow as per section S.4.3.1.

Three species are identified in the $^{31}\text{P}\{^1\text{H}\}$ solution NMR spectra;

$[1-(\text{CH}_2\text{Cl}_2)_n][\text{BAr}^{\text{F}}_4]$: δ 97.2 (br d, J_{RHP} 203 Hz).

$[1-\text{C}_6\text{H}_{10}][\text{BAr}^{\text{F}}_4]$: δ 96.9 (dd, J_{RHP} 207 Hz, J_{PP} 24 Hz), 90.4 (dd, J_{RHP} 159 Hz, J_{PP} 24 Hz).

$[1-\text{C}_6\text{H}_8][\text{BAr}^{\text{F}}_4]$: δ 78.3 (d, J_{RHP} 172 Hz).

Note: A powdered crystalline sample of $[1-\text{C}_6\text{H}_8][\text{BAr}^{\text{F}}_4]$ was placed under vacuum (1×10^{-2} mbar) for a period of 2 weeks. ^1H and $^{31}\text{P}\{^1\text{H}\}$ NMR spectra of the dissolved solid (CD_2Cl_2) showed no significant change, and no $[1-\text{C}_6\text{H}_6][\text{BAr}^{\text{F}}_4]$ was observed.

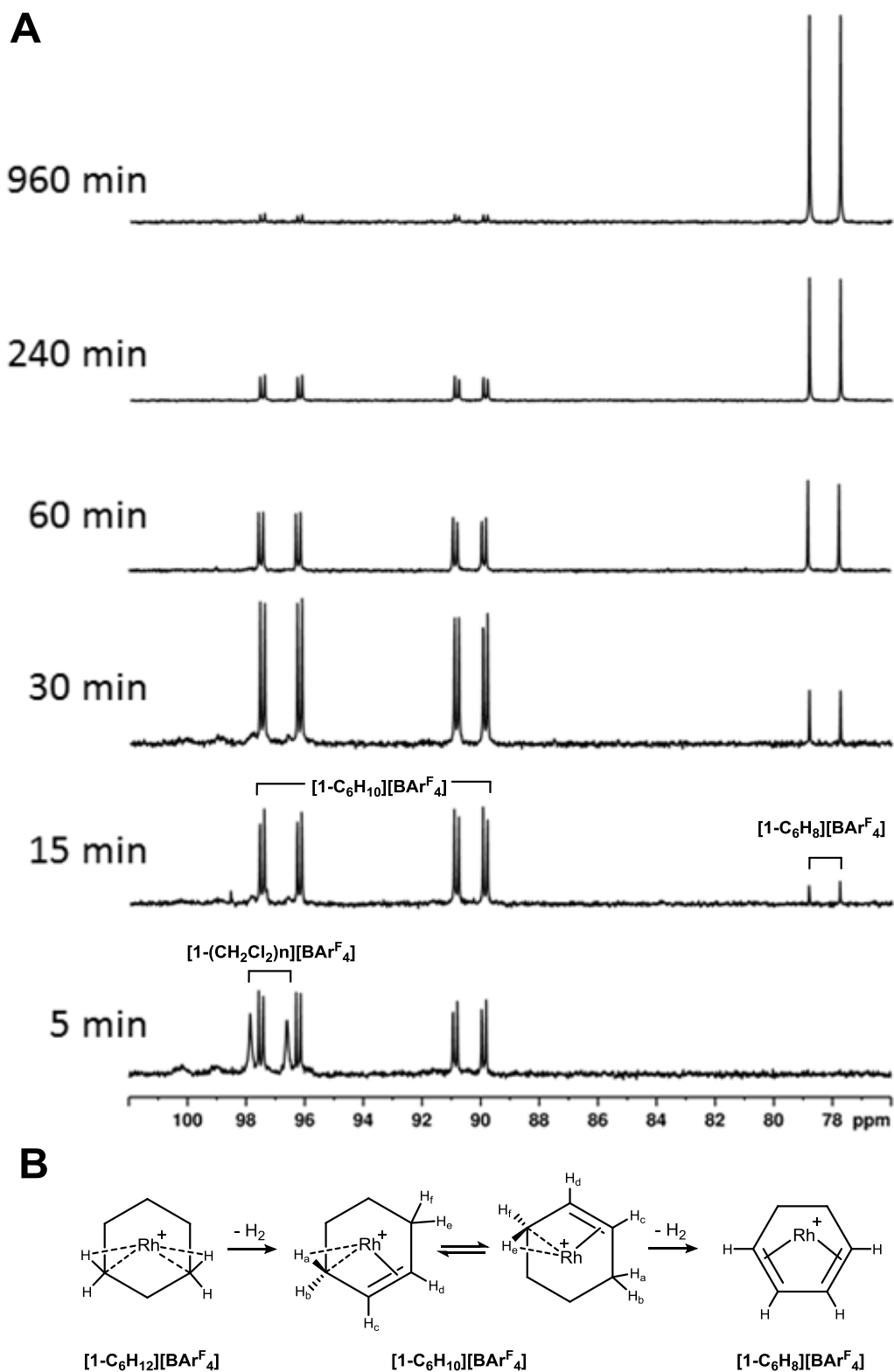


Figure S69: (A) Representative solution $^{31}P\{^1H\}$ NMR (CH_2Cl_2 , 183 K, 162 MHz) spectra of $[1-C_6H_{12}][BARF_4]$ exposed to vacuum for various periods of time in the solid-state then quenched by condensing CH_2Cl_2 onto the sample. **(B)** Structural diagrams of the species present during the dehydrogenation reactions; phosphine units and $[BARF_4]^-$ anions removed for clarity.

S.4.3.3. Kinetics of the solid-state $[1-C_6H_{12}][BAr^F_4]$ dehydrogenation *under vacuum* to form $[1-C_6H_8][BAr^F_4]$

In contrast to the dehydrogenation of $[1-C_4H_{10}][BAr^F_4]$, the dehydrogenation of $[1-C_6H_{12}][BAr^F_4]$ proceeds in two stages. Given the poor stability of the intermediate complex of $[1-C_6H_{10}][BAr^F_4]$ in solution, the $^{31}P\{^1H\}$ NMR spectra used in the kinetic study were collected at 183 K. As previously noted $[1-C_6H_{12}][BAr^F_4]$ imminently reacts with CH_2Cl_2 even at low temperatures to form several species two of which we have tentatively assigned as $[1-(CH_2Cl_2)_n][BAr^F_4]$ and $[1-BAr^F_4]$. At 183 K, the resonances of these species are typically very broad.

Dehydrogenation reactions were undertaken in J Young NMR tubes containing capillaries of a ^{31}P NMR standard (PPh_3 in d_6 -acetone), in which a known amount of powdered crystalline $[1-C_6H_8][BAr^F_4]$ was hydrogenated, then subjected to vacuum for a range of times from 15 mins to 960 mins, and worked up as per the procedure outlined in section S.4.3.3. The concentration of each capillary was calibrated to a sample of $Cy_2PCH_2CH_2PCy_2$ in CH_2Cl_2 at 183 K. The concentrations of the species involved were referenced against the internal standard.

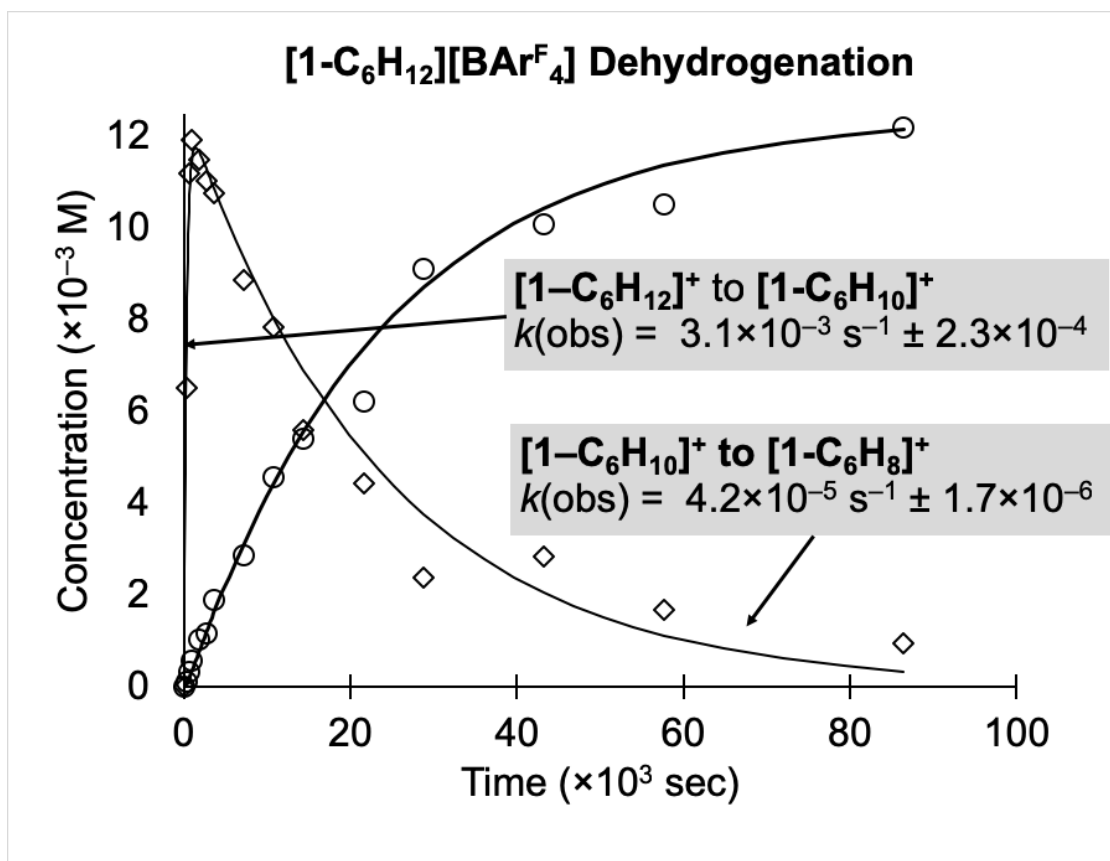


Figure S70: Temporal evolution of dehydrogenation of $[1\text{-C}_6\text{H}_{12}][\text{BAr}^{\text{F}}_4]$ under vacuum (1×10^{-2} mbar). Each data point comes from a separate experiment using 10 mg crystalline material, subjected to a vacuum for a set period of time and dissolved in cold CD_2Cl_2 (0.4 mL) that has been vacuum transferred into the sample.

This gives a $[\text{Rh}]_{\text{TOT}} = 13 \times 10^{-3}$ M. Integrals measured using quantitative $^{31}\text{P}\{^1\text{H}\}$ NMR spectroscopy at 183 K of the resulting Rh-complexes, $[1\text{-BAr}^{\text{F}}_4]$ (a proxy for $[1\text{-C}_6\text{H}_{12}][\text{BAr}^{\text{F}}_4]$), $[1\text{-C}_6\text{H}_{10}][\text{BAr}^{\text{F}}_4]$ and $[1\text{-C}_6\text{H}_8][\text{BAr}^{\text{F}}_4]$ which have been calibrated from use of an internal standard of PPh_3 in a sealed capillary.

Solid-lines are fits for COPASI^{S10} modelled processes for the sequential dehydrogenation ($[1\text{-C}_6\text{H}_{12}][\text{BAr}^{\text{F}}_4] \rightarrow [1\text{-C}_6\text{H}_{10}][\text{BAr}^{\text{F}}_4] \rightarrow [1\text{-C}_6\text{H}_8][\text{BAr}^{\text{F}}_4]$), where the *second* dehydrogenation is 1st order ($k = 3.1(2) \times 10^{-5} \text{ s}^{-1}$) and the *first* is either 1st or 2nd order. Initial concentration of dissolved samples = 12.5 mM (conversion = 0%).

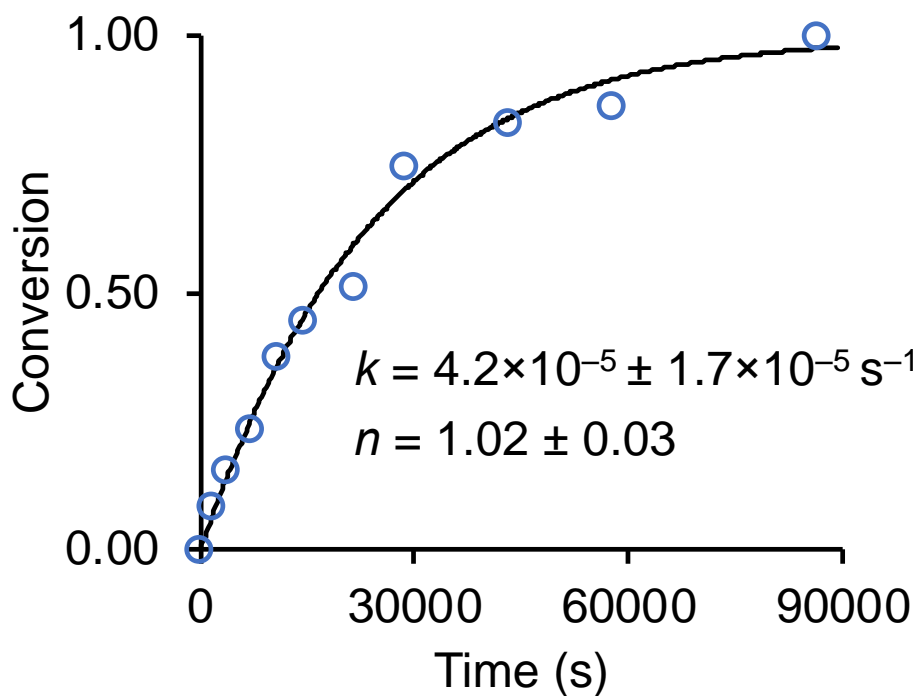
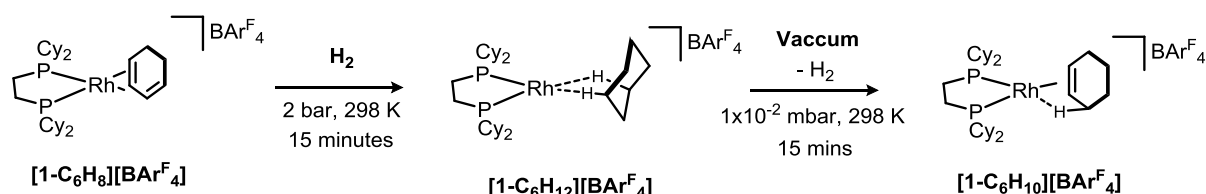


Figure S71: Modified JAMK Plot of conversion^{S9} ($f = (1 - e^{-(kt)^n})$) versus time for the dehydrogenation process $[1\text{-C}_6\text{H}_{10}][\text{BAr}^{\text{F}}_4]$ to $[1\text{-C}_6\text{H}_8][\text{BAr}^{\text{F}}_4]$ in the solid-state; which is the second dehydrogenation process of $[1\text{-C}_6\text{H}_{12}][\text{BAr}^{\text{F}}_4]$. Where k = growth rate constant and n = Avrami exponent.

S.4.4. Solid-state dehydrogenation of $[1\text{-C}_6\text{H}_{12}][\text{BAr}^{\text{F}}_4]$ to form $[1\text{-C}_6\text{H}_{10}][\text{BAr}^{\text{F}}_4]$

S.4.4.1. Solid-State Synthesis of $[1\text{-C}_6\text{H}_{10}][\text{BAr}^{\text{F}}_4]$



Attempts to trap $[1\text{-C}_6\text{H}_{10}][\text{BAr}^{\text{F}}_4]$ in the solid-state were conducted using the following method:

A powdered microcrystalline sample of $[1\text{-C}_6\text{H}_8][\text{BAr}^{\text{F}}_4]$ (35 mg) was packed in a 3.2 mm SSNMR rotor, inside an argon filled glove box. The rotor was then placed in a custom built glass J Young flask^{S7} and the sample was then exposed to H_2 (1 bar, 298 K) for 20 minutes, to yield deep red crystalline powder of $[1\text{-C}_6\text{H}_{12}][\text{BAr}^{\text{F}}_4]$. The sample was then placed under a steady vacuum (2×10^{-2} mbar) at 298 K. After 15 minutes, the rotor cap was fitted under a flush of Ar, and immediately transferred to the bore of a SSNMR spectrometer and analyzed by $^{31}\text{P}\{^1\text{H}\}$ and $^{13}\text{C}\{^1\text{H}\}$ solid state NMR spectroscopy.

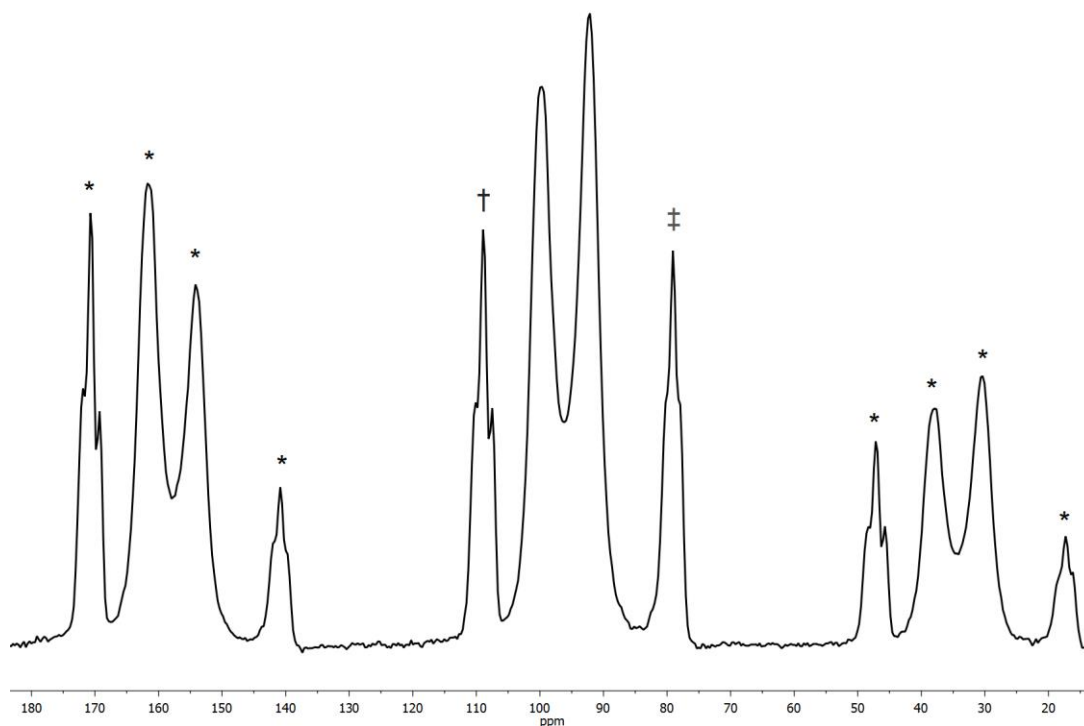


Figure S72: The $^{31}\text{P}\{^1\text{H}\}$ SSNMR (162 MHz, 295 K, 10 kHz spin rate) spectrum of the dehydrogenation of powdered microcrystalline $[\mathbf{1-C}_6\text{H}_{12}][\text{BAr}^{\text{F}}_4]$ under 15 minutes of vacuum. The resonances at δ 100 and δ 92 correspond to $[\mathbf{1-C}_6\text{H}_{10}][\text{BAr}^{\text{F}}_4]$ (~ 70 %), and the resonances † marked are assigned to $[\mathbf{1-C}_6\text{H}_{12}][\text{BAr}^{\text{F}}_4]$ (~ 15 %) and the resonances marked ‡ (~ 15 %) are assigned to $[\mathbf{1-C}_6\text{H}_8][\text{BAr}^{\text{F}}_4]$. The resonances marked * are due to spinning sidebands.

We postulate this mixture of species arises in the solid-state from non-uniform vacuum being applied throughout the densely packed solid-state rotor, leading to slightly different kinetics then observed in Section S.4.3.3.

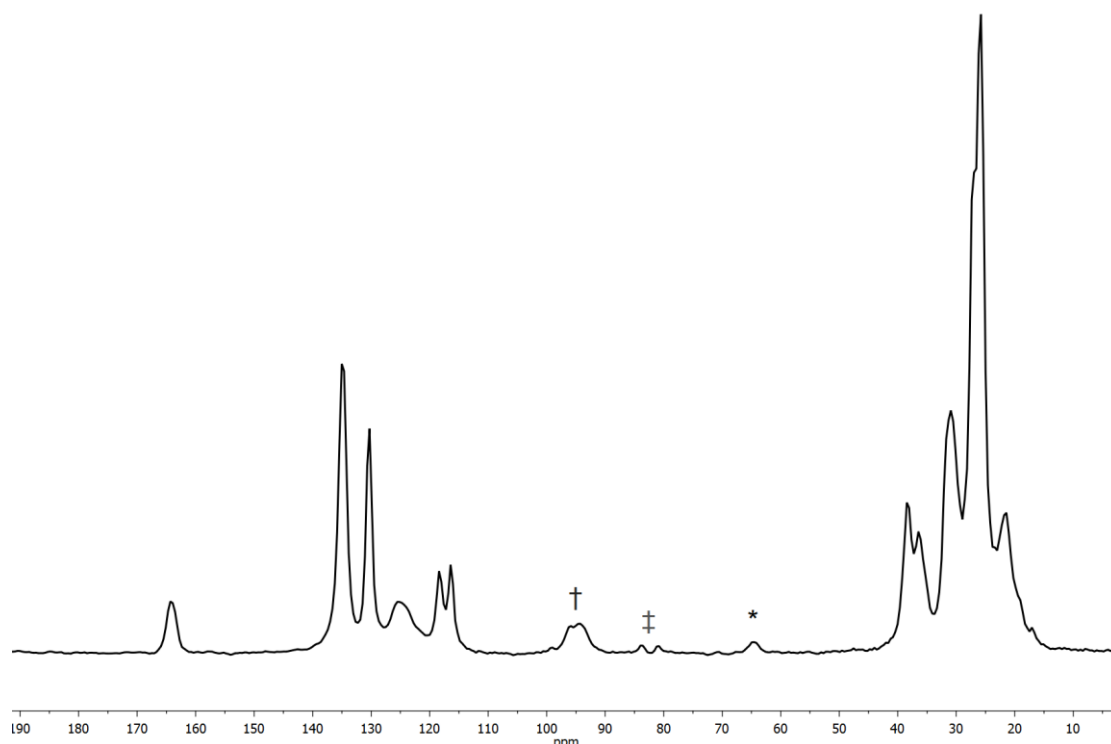


Figure S73: The $^{13}\text{C}\{^1\text{H}\}$ SSNMR (100 MHz, 295 K, 10 kHz spin rate) spectrum of the dehydrogenation of powdered microcrystalline $[1-\text{C}_6\text{H}_{12}][\text{BAr}^{\text{F}}_4]$ under 15 minutes of vacuum. The resonance marked † are from the carbons in alkene environments of $[1-\text{C}_6\text{H}_{10}][\text{BAr}^{\text{F}}_4]$, and resonances ‡ marked are assigned to $[1-\text{C}_6\text{H}_8][\text{BAr}^{\text{F}}_4]$, see Figure S22. The resonances marked * are due to spinning sidebands.

S.4.4.2. Isolation and solution-state characterization data for $[1-\text{C}_6\text{H}_{10}][\text{BAr}^{\text{F}}_4]$

Attempts to trap $[1-\text{C}_6\text{H}_{10}][\text{BAr}^{\text{F}}_4]$ in solution were conducted using the following method:

A J Young NMR tube was charge with powdered microcrystalline $[1-\text{C}_6\text{H}_8][\text{BAr}^{\text{F}}_4]$ (10 mg) in a glove box. The sample was then evacuated and backfilled with H_2 (2 bar, 298 K) for 15 minutes. The sample was then cooled to 77 K in liquid nitrogen and evacuated. After a steady vacuum was achieved (2×10^{-2} mbar) the sample was rapidly warmed to 298 K and a timer was simultaneously started. After 15 minutes, the sample was cooled to 77 K in liquid nitrogen and CH_2Cl_2 (0.40 mL) was condensed into the NMR tube. The sample was expediently thawed and rapidly transferred to the bore of a precooled (183 K) NMR spectrometer which was previously locked and shimmed to a sample of CD_2Cl_2 . A $^{31}\text{P}\{^1\text{H}\}$ NMR spectrum was then acquired.

^1H NMR (CD_2Cl_2 , 400 MHz, 183 K, selected resonances): δ 7.72 (s, 14H, ortho-ArH), 7.54 (s, 7H, para-ArH), 5.23 (s, \sim 2H, alkene-CH), 2.47 (br d, J_{HH} 14 Hz, cyclohexene-CH), 2.05-1.38 (br m, 73H, overlapping aliphatic CH), 1.30-0.98 (br m, 53H, overlapping aliphatic CH), -1.01 (br s, \sim 2H, agostic CH).

Note: The low temperature limit was not reached at 183K, so integrated values are approximate. As well, the integrals of the $[\text{BAr}^{\text{F}}_4]^-$ and alkyl resonances are higher than expected presumably due to contamination decomposition products of **[1-BAr^F₄]**, ^{S2} **[1-(CD₂Cl₂)_n][BAr^F₄]** ^{S1} and liberated cyclohexane.

$^{31}\text{P}\{^1\text{H}\}$ NMR (CD_2Cl_2 , 162 MHz, 183 K, selected resonances): δ 98.0 (dd, J_{RHP} 207 Hz, J_{PP} 24 Hz), 91.5 (dd, J_{RHP} 159 Hz, J_{PP} 24 Hz).

$^{19}\text{F}\{^1\text{H}\}$ NMR (CD_2Cl_2 , 377 MHz, 183 K, selected resonances): δ -62.9 (s).

Note: Samples rapidly decomposed in solution at room temperature. The decomposition products **[1-C₆H₈][BAr^F₄]**, **[1-C₆H₆][BAr^F₄]**, **[1-BAr^F₄]** and liberated cyclohexane were identified by ^1H and $^{31}\text{P}\{^1\text{H}\}$ NMR spectroscopy.

S.4.4.3. Solution NMR spectra of [1-C₆H₁₀][BAr^F₄]

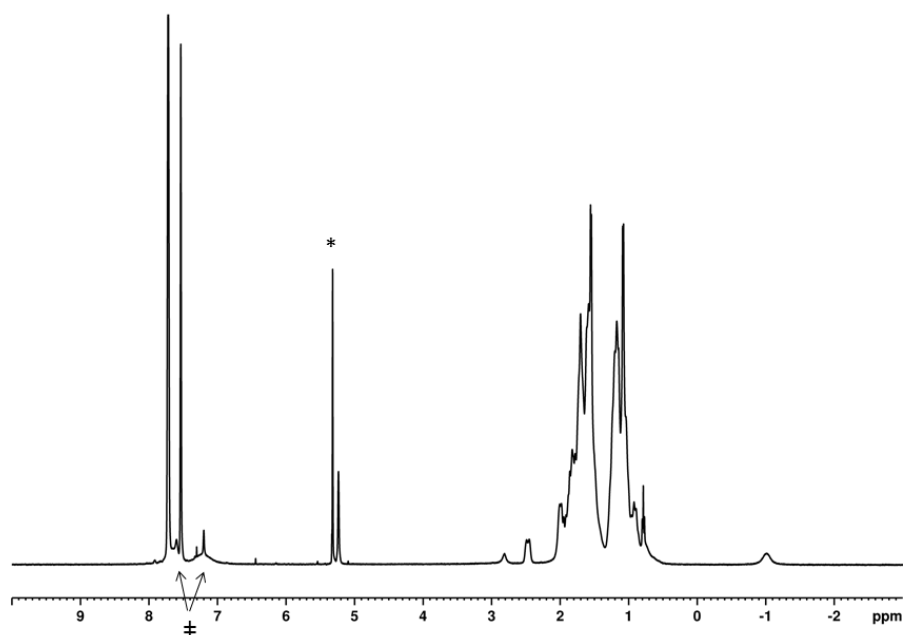


Figure S74: The solution ¹H NMR (CD₂Cl₂, 183 K, 400 MHz) spectrum of the intermediate stage of dehydrogenation. The resonance marked * is due to CHDCl₂. The resonances marked ⊕ are tentatively assigned to [1-BAr^F₄].

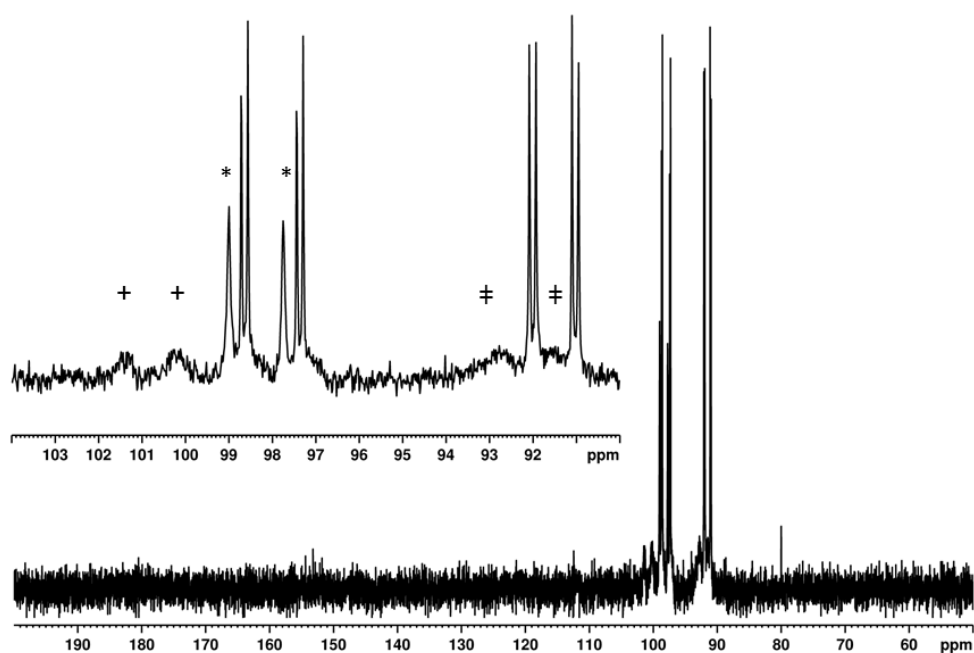


Figure S75: The ³¹P{¹H} NMR (CD₂Cl₂, 183 K, 162 MHz) spectrum of the intermediate stage of dehydrogenation. The resonances marked * are tentatively assigned to [1-(CD₂Cl₂)_n][BAr^F₄]. The resonances marked ⊕ are assigned to [1-BAr^F₄].^{S2} The resonances marked + could not be assigned.

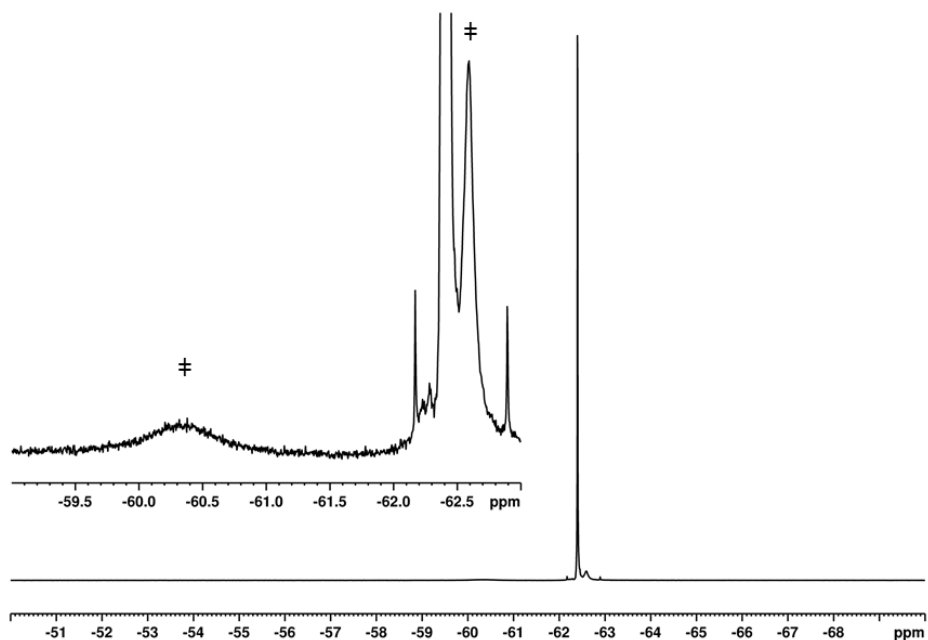


Figure S76: The $^{19}\text{F}\{^1\text{H}\}$ NMR (CD_2Cl_2 , 183 K, 377 MHz) spectrum of the intermediate stage of dehydrogenation. The inset is an enlargement of the baseline around the central resonance. The resonance marked \oplus is tentatively assigned to **[1-BAr $^{\text{F}}_4$]**.

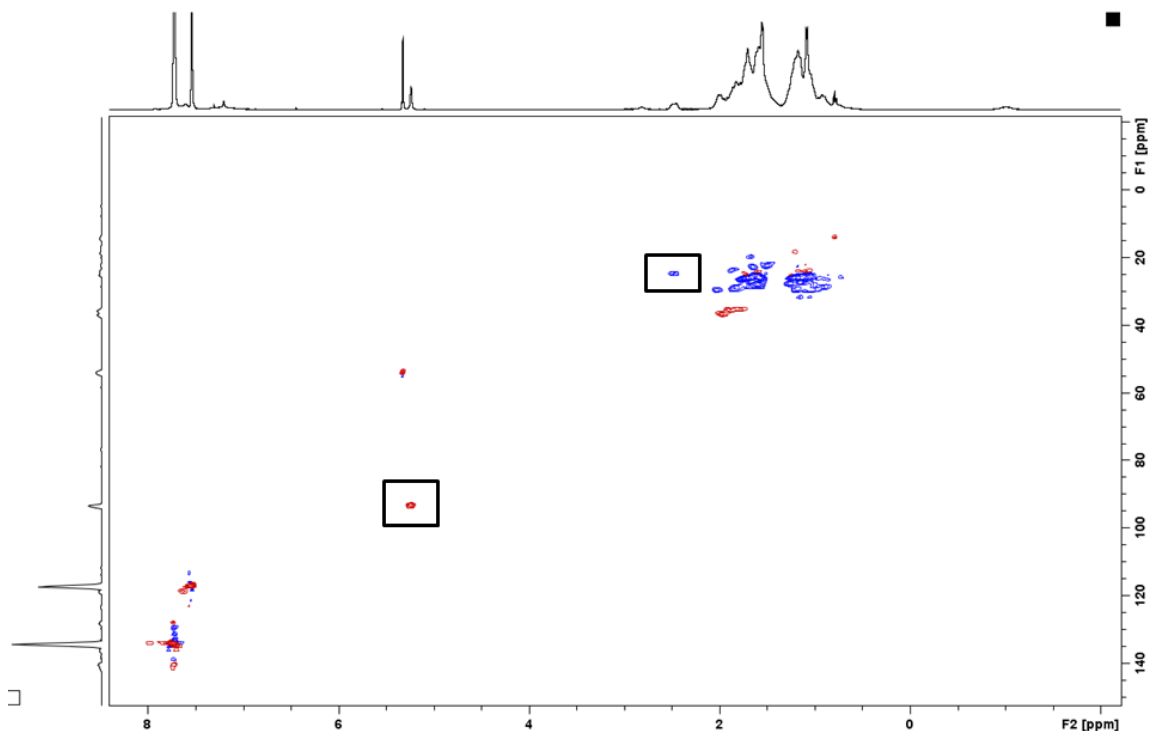


Figure S77: The solution $^1\text{H}/^{13}\text{C}$ HSQC NMR (CD_2Cl_2 , 183 K) spectrum of intermediate in the dehydrogenation. Black boxes added to guide the eye. A cross peak for the high field ^1H resonance was not unambiguously identified.

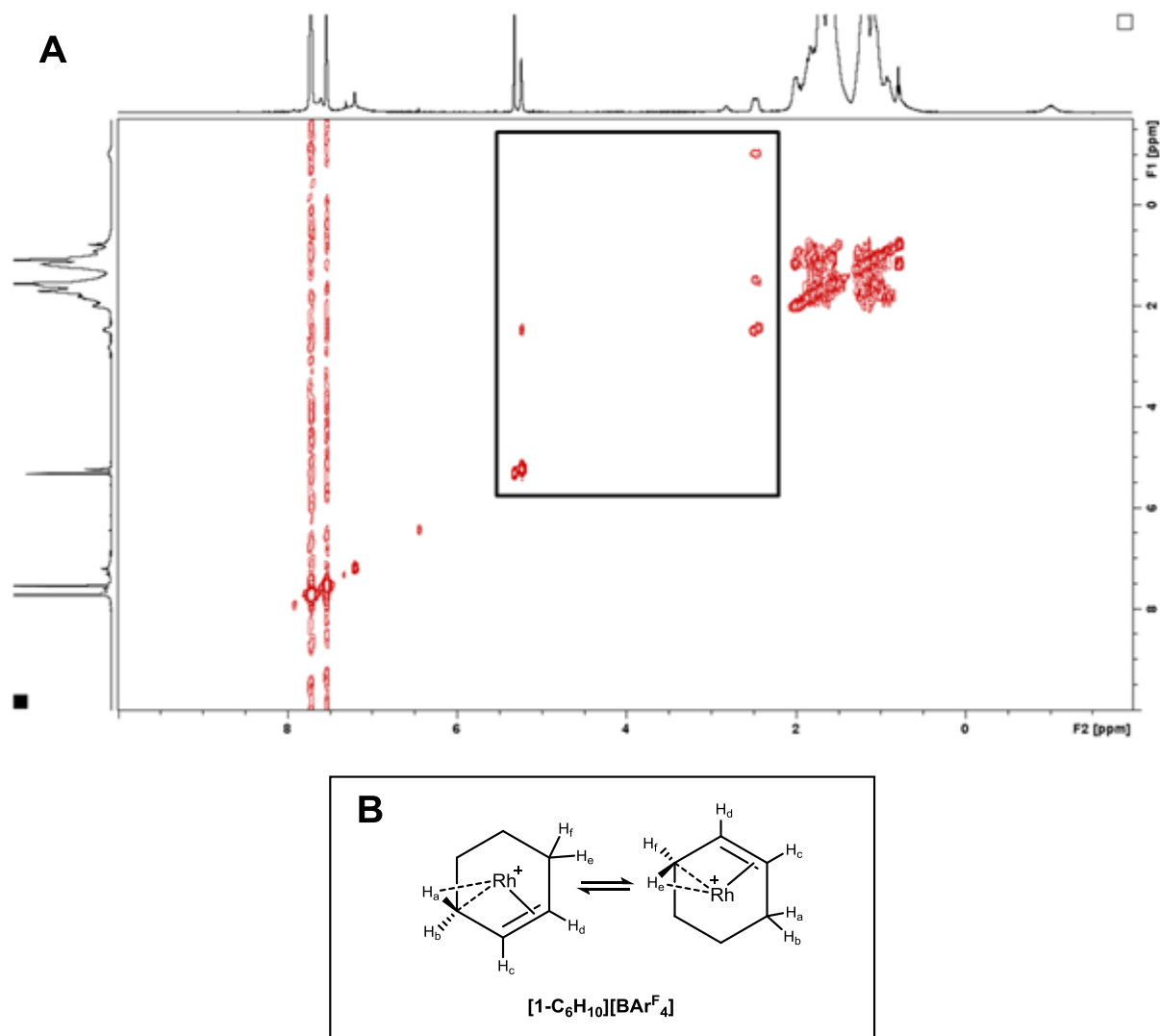


Figure S78: (A) The solution $^1\text{H}/^1\text{H}$ COSY NMR (CD_2Cl_2 , 183 K) spectrum of intermediate in the dehydrogenation. Black box added to guide the eye, showing $^1\text{H}/^1\text{H}$ correlation between agostic and alkene protons in $[1\text{-C}_6\text{H}_{10}][\text{BAR}^{\text{F}}_4]$. (B) Structural diagrams of the fluxionality at room temperature of $[1\text{-C}_6\text{H}_{10}][\text{BAR}^{\text{F}}_4]$; phosphine units and $[\text{BAR}^{\text{F}}_4]^-$ anions removed for clarity.

S.4.4.4. Liberation of the bound cyclohexene from [1-C₆H₁₀][BAr^F₄].

A J Young NMR tube was charge with powdered microcrystalline [1-C₆H₈][BAr^F₄] (10 mg) in a glove box. The sample was then evacuated and backfilled with H₂ (2 bar, 298 K). The sample was then cooled to 77 K in liquid nitrogen and evacuated. After a steady vacuum was achieved (2×10^{-2} mbar) the sample was rapidly warmed to 298 K and a timer was simultaneously started. After 15 mins, the sample was cooled to 77 K in liquid nitrogen and CH₂Cl₂ (0.40 mL) was condensed into the NMR tube. When thawing, care was taken such that the sample was not warmed above 233 K. The cool sample was then carefully refilled with CO (1 bar, 233 K). After 5 minutes, a vibrant yellow solution had formed of [Rh(dcpe)(CO)₂][BAr^F₄],^{S7} confirmed by solution ³¹P{¹H} NMR spectroscopy (δ 85.22, d, $J_{\text{RhP}} = 116$ Hz), as well as liberated cyclohexene. The volatile component was then isolated by trap-to-trap distillation and analyzed by ¹H NMR spectroscopy and GC EI-MS.

¹H NMR (CD₂Cl₂, 400 MHz): δ 5.66 (s, 2H, alkene-CH), 1.98 (br m, 4H, CH), 1.61 (m, 4H, CH).

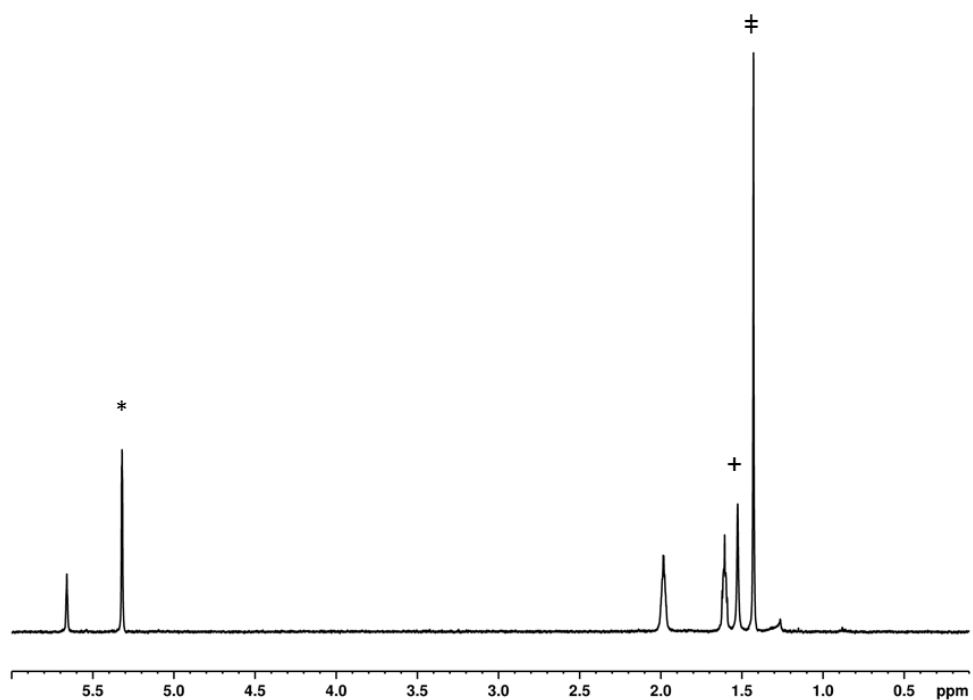


Figure S79: The ^1H NMR (CD_2Cl_2 , 298 K, 400 MHz) spectrum of volatiles isolated post carbonylation. The resonance marked * is due to CHDCl_2 . The resonance marked \oplus is assigned to cyclohexane. The resonance marked + is assigned to H_2O .

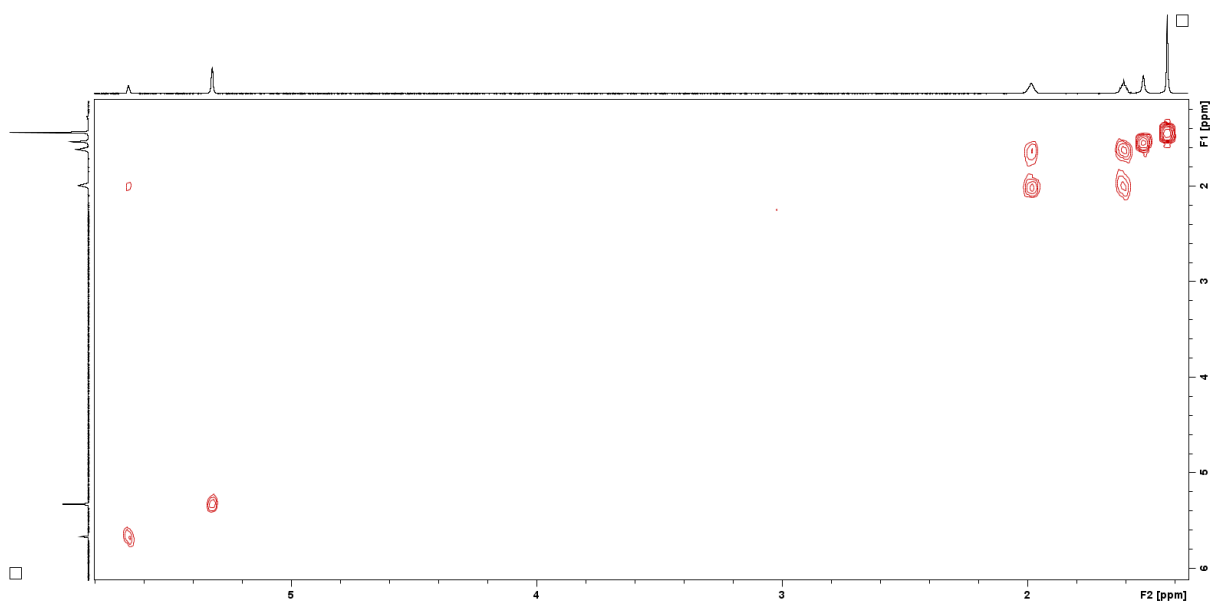


Figure S80: The $^1\text{H}/^1\text{H}$ COSY NMR (CD_2Cl_2 , 298 K, 400 MHz) spectrum of volatiles isolated post carbonylation.

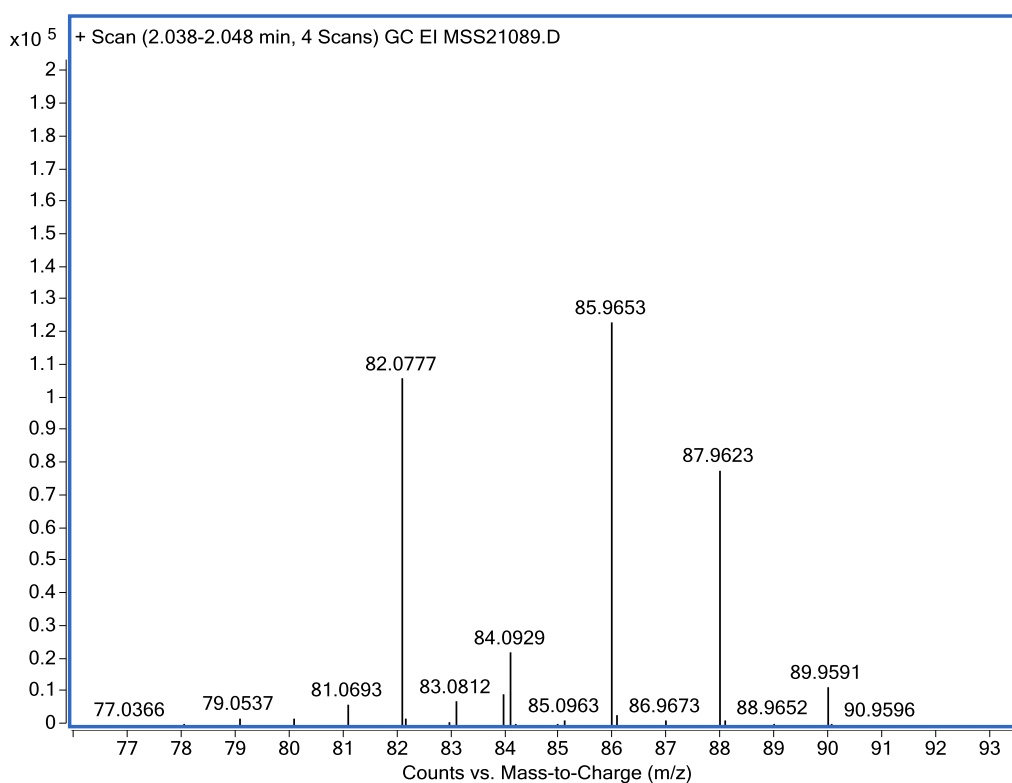
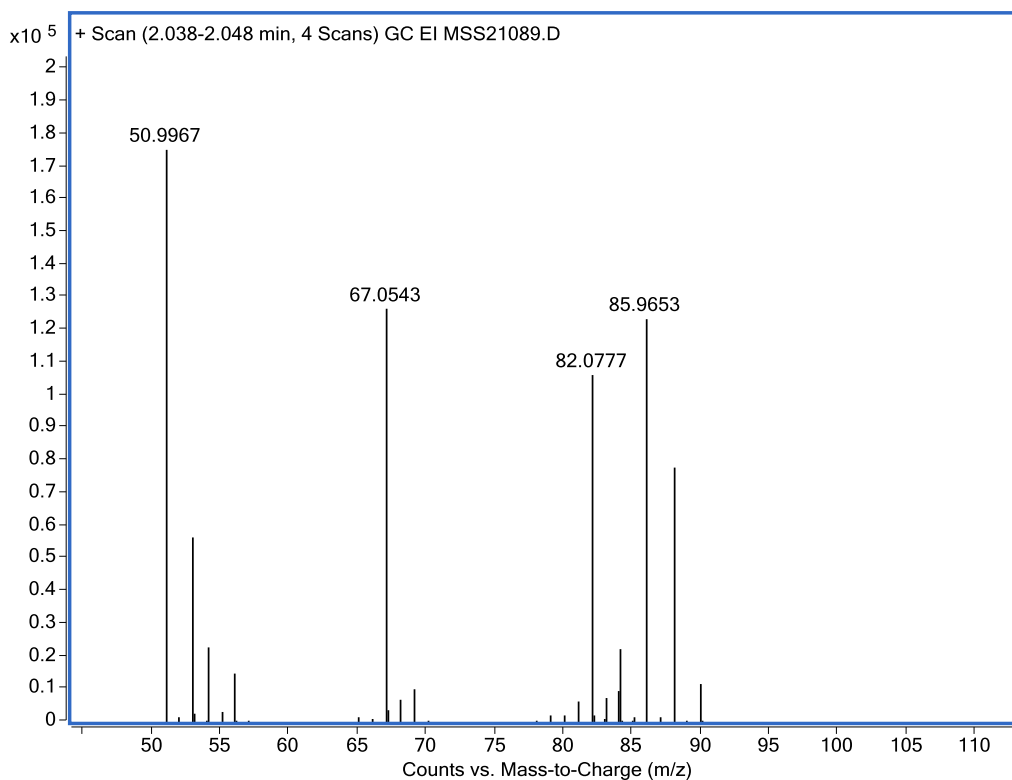


Figure S81: Mass spectrum of the isolated volatiles (top). Enlargement of $m/z = 77 - 93$ region (bottom). The signal at $m/z = 82.0777$ matches that of C_6H_{10} (calc. 82.0777). The signal at $m/z = 84.0929$ matches that of C_6H_{12} (calc. 84.0933). Signals at $m/z = 86$ and 88 are due to CD_2Cl_2 .

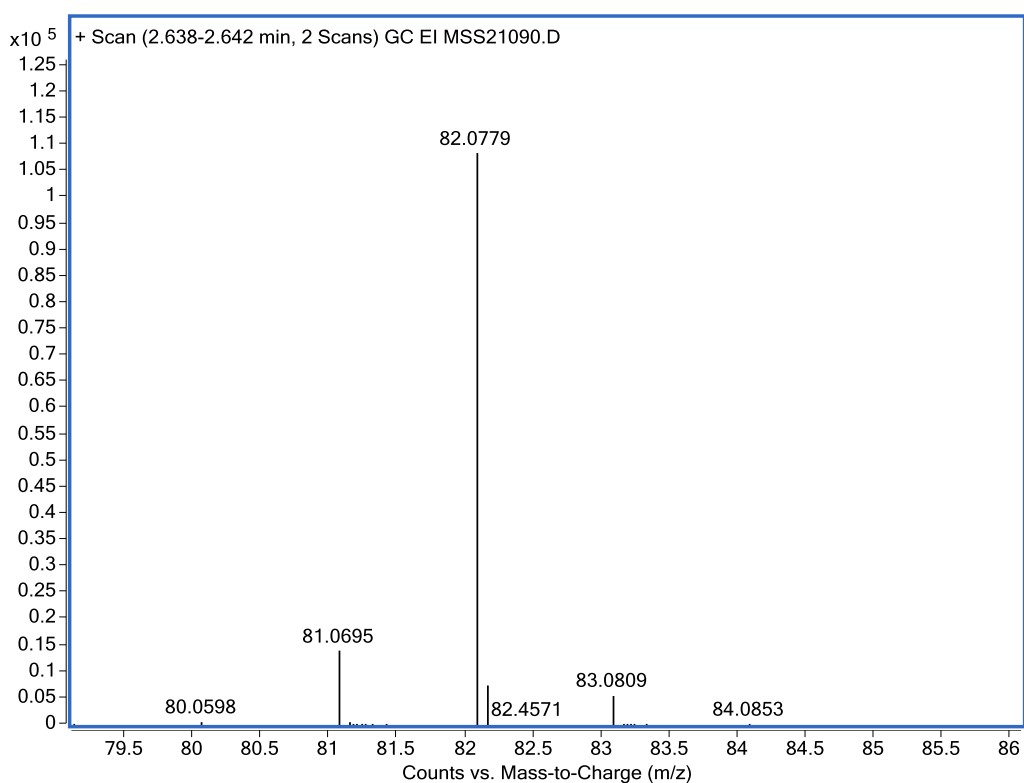
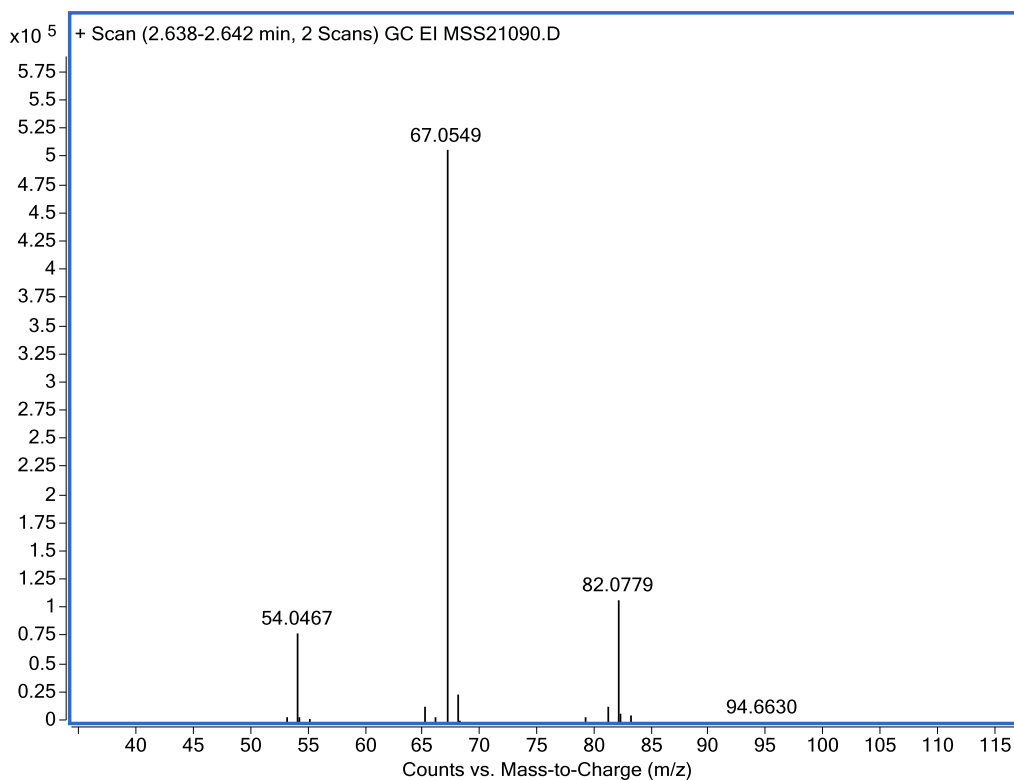
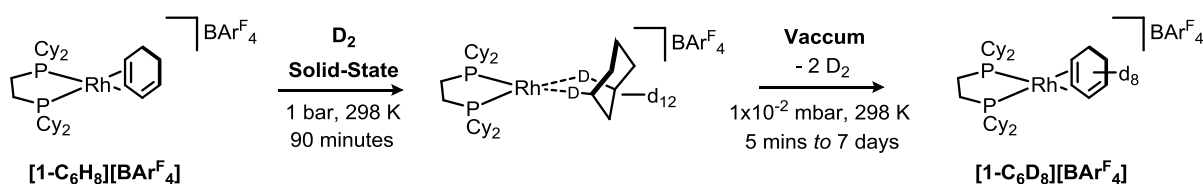


Figure S82: Mass spectrum of a commercially sourced sample of cyclohexene (top). Enlargement of $m/z = 79 - 86$ region (bottom).

S.4.5. Solid state dedeuteration of [1-C₆D₁₂][BARF₄] under vacuum to form [1-C₆D₈][BARF₄]



A thick walled NMR tube was charge with powdered microcrystalline [1-C₆H₈][BARF₄] (10 mg) and sample was cooled to 77 K in liquid nitrogen, evacuated and backfilled with D₂ (1 bar, 298 K). After 30 min, sample was then cooled to 77 K in liquid nitrogen, evacuated and backfilled with D₂ (1 bar, 298 K). This cycle was repeated once more, to give a total of 3 x 30 minute cycles, totaling 90 minutes under D₂. The sample was placed under a steady vacuum (2 × 10⁻² mbar), was rapidly warmed to 298 K and a timer was simultaneously started. After set periods of time (5 mins to approx. 3 days) the sample was cooled to 77 K in liquid nitrogen and CH₂Cl₂ (0.40 mL) was condensed into the NMR tube. The sample was expediently thawed and rapidly transferred to the bore of a precooled (183 K) NMR spectrometer which was previously locked and shimmed to a sample of CD₂Cl₂. A ³¹P{¹H} NMR spectrum was then acquired.

The above procedure was repeated on numerous occasions, varying the period of time the sample was exposed to vacuum at room temperature.

S.4.5.1. Kinetics of the solid state dedeuteration of $[1\text{-C}_6\text{D}_{12}][\text{BAr}^{\text{F}}_4]$ under vacuum to form $[1\text{-C}_6\text{D}_8][\text{BAr}^{\text{F}}_4]$

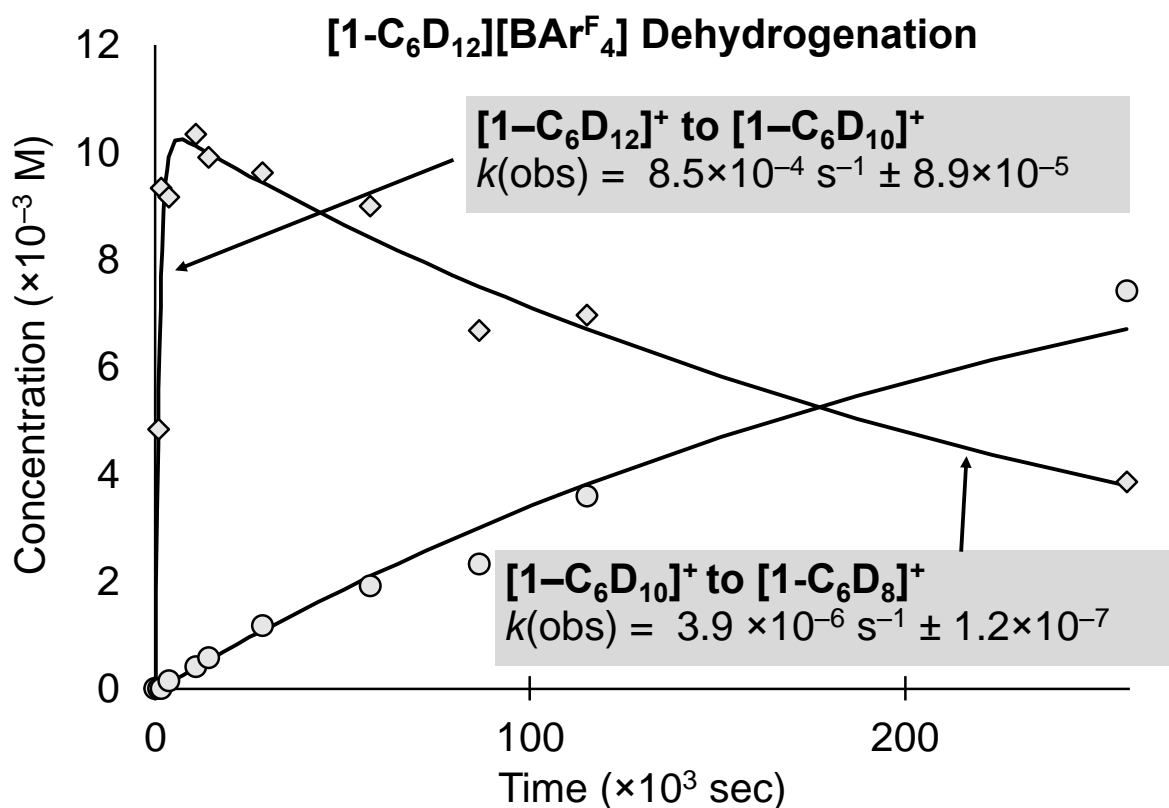


Figure S83: Temporal evolution of the dehydrogenation of $[1\text{-C}_6\text{D}_{12}][\text{BAr}^{\text{F}}_4]$ under vacuum (1×10^{-2} mbar). A sample of $[1\text{-C}_6\text{D}_{12}][\text{BAr}^{\text{F}}_4]$ was initially prepared by exposing samples of $[1\text{-C}_6\text{H}_{12}][\text{BAr}^{\text{F}}_4]$ to 3×30 minutes of D_2 (1 bar, 298 K). This prolonged exposure resulted in a small but significant systematic decomposition of $[1\text{-C}_6\text{H}_{12}][\text{BAr}^{\text{F}}_4]$. This shown by $^{13}\text{P}\{^1\text{H}\}$ SSNMR to be $\sim 10\%$ (Section S.2.4.6.). The best COPASI simulated fits (both first order) agree well with this attenuation in $[1\text{-C}_6\text{D}_{12}][\text{BAr}^{\text{F}}_4]$.

S.4.5.2. Characterization data for [1-C₆D₈][BAr^F₄]

¹H NMR (CD₂Cl₂, 298 K, 400 MHz): δ 7.73 (s, 8H, ortho-ArH), 7.57 (s, 4H, para-ArH), 2.15 (br s, 2H, aliphatic CH), 1.99-1.66 (br m, 24H, overlapping aliphatic CH), 1.61-1.58 (br m, 2H, aliphatic CH), 1.38-1.08 (br m, 16H, overlapping aliphatic CH), 0.99-0.96 (br m, 4H, overlapping aliphatic CH).

²H NMR (CH₂Cl₂, 298 K, 61 MHz): δ 5.30 (br s, 4D, alkene-CD), 1.76 (br s, 2D, aliphatic-CD), 1.62 (br s, 2D, aliphatic-CD).

³¹P{¹H} NMR (CD₂Cl₂, 298 K, 162 MHz): δ 78.7 (d, J_{RhP} 172 Hz).

¹⁹F{¹H} NMR (CD₂Cl₂, 298 K, 377 MHz): δ -62.9 (s).

¹³C{¹H} NMR (CD₂Cl₂, 298 K, 126 MHz): δ 162.2 (1:1:1:1 q, J_{CB} 50 Hz, ipso-ArC), 135.2 (s, ortho-ArC), 129.3 (q, J_{CF} 32 Hz, meta-ArC), 125.0 (q, J_{CF} 272 Hz, CF₃), 117.9 (s, para-ArC), 94.7 (1:1:1 t, J_{CD} 26 Hz, alkene-CD), 82.3 (1:1:1 t, J_{CD} 23 Hz, alkene-CD), 37.9 (dt, J_{CP} 24.4 Hz, J_{CRh} 11.5 Hz, cyclohexyl-CH), 37.0 (dt, J_{CP} 23.3 Hz, J_{CRh} 11.4 Hz, cyclohexyl-CH), 31.2 (s, cyclohexyl-CH₂), 30.0 (s, cyclohexyl-CH₂), 29.5 (s, cyclohexyl-CH₂), 29.1 (s, cyclohexyl-CH₂), 27.5 (t, J_{CP} 6.4 Hz, cyclohexyl-CH₂), 27.2 (t, J_{CP} 4.3 Hz, cyclohexyl-CH₂), 27.0 (t, J_{CP} 5.8 Hz, cyclohexyl-CH₂), 26.3 (d, J_{CP} 18.6 Hz, PCH₂), 22.6 (d, J_{CP} 3.4 Hz, cyclohexyl-CH₂), 22.5 (br s, cyclohexyl-CH₂), 22.3 (d, J_{CP} 3.2 Hz, cyclohexyl-CH₂), 21.3 (1:2:3:2:1 quintet, J_{CD} = 20 Hz, CD₂).

ESI-MS found (calc. for [Rh(Cy₂PCH₂CH₂PCy₂)(C₆D₈)]⁺): m/z 613.34 (613.34).

S.4.5.3. Solution NMR and mass spectra of [1-C₆D₈][BAr^F₄]

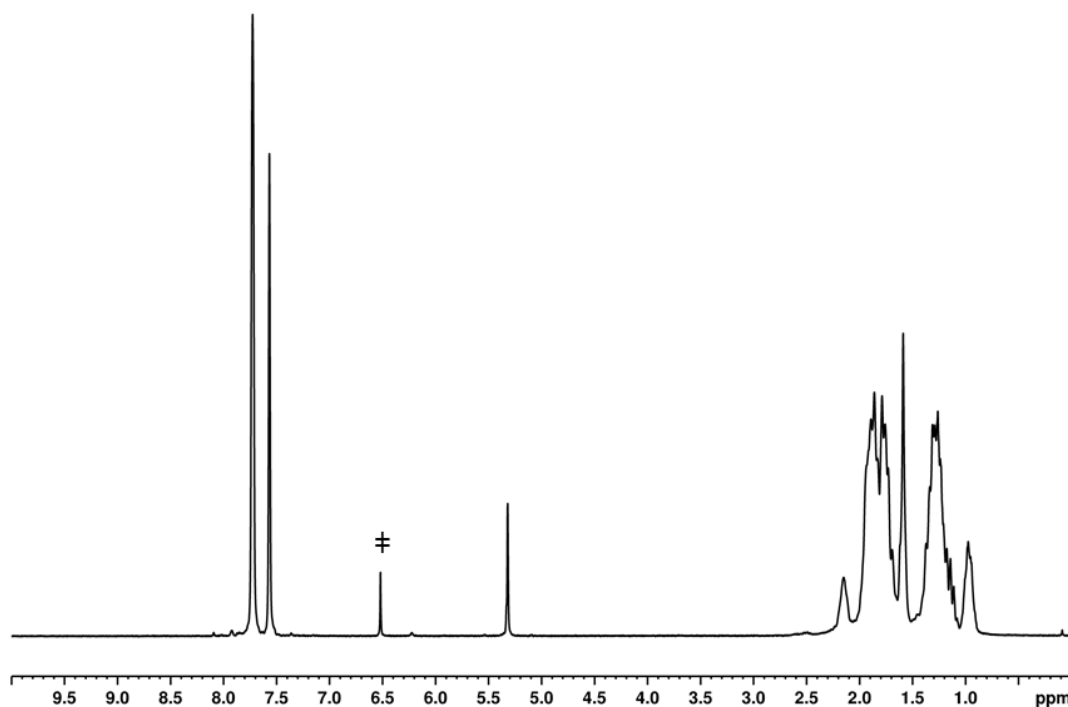
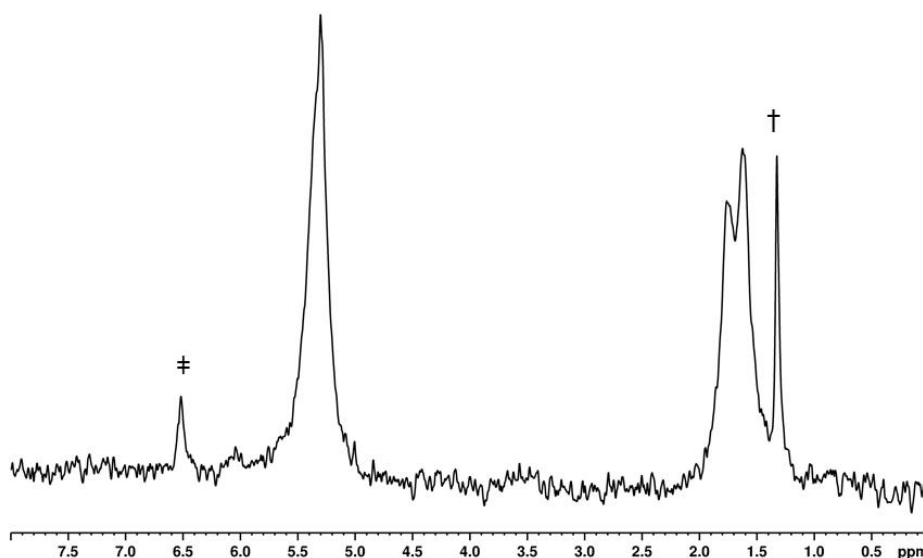


Figure S84: The solution ¹H NMR (CD₂Cl₂, 298 K, 400 MHz) spectrum of [1-C₆D₈][BAr^F₄]. The resonance marked ‡ is due to [1-C₆H₆][BAr^F₄] which is present as a minor (~ 5 %) contaminant in the [1-C₆H₈][BAr^F₄] starting material in this



experiment.

Figure S85: The solution ²H NMR (CH₂Cl₂, 298 K, 61 MHz) spectrum of [1-C₆D₈][BAr^F₄]. The resonances marked ‡ and † are due to [1-C₆D₆][BAr^F₄] and liberated C₆D₁₂ respectively, which arise due to the solution decomposition of [1-C₆D₈][BAr^F₄] and incomplete dehydrogenation.

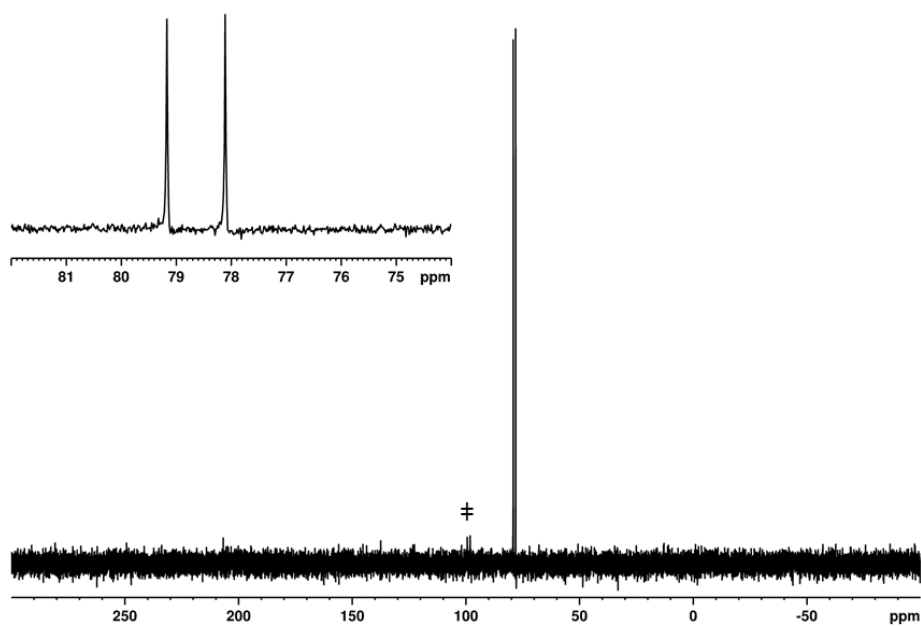


Figure S86: The solution $^{31}\text{P}\{^1\text{H}\}$ NMR (CD_2Cl_2 , 298 K, 162 MHz) spectrum of $[1\text{-C}_6\text{D}_8][\text{BAr}^{\text{F}}_4]$. The resonance marked ‡ is due to $[1\text{-C}_6\text{H}_6][\text{BAr}^{\text{F}}_4]$.

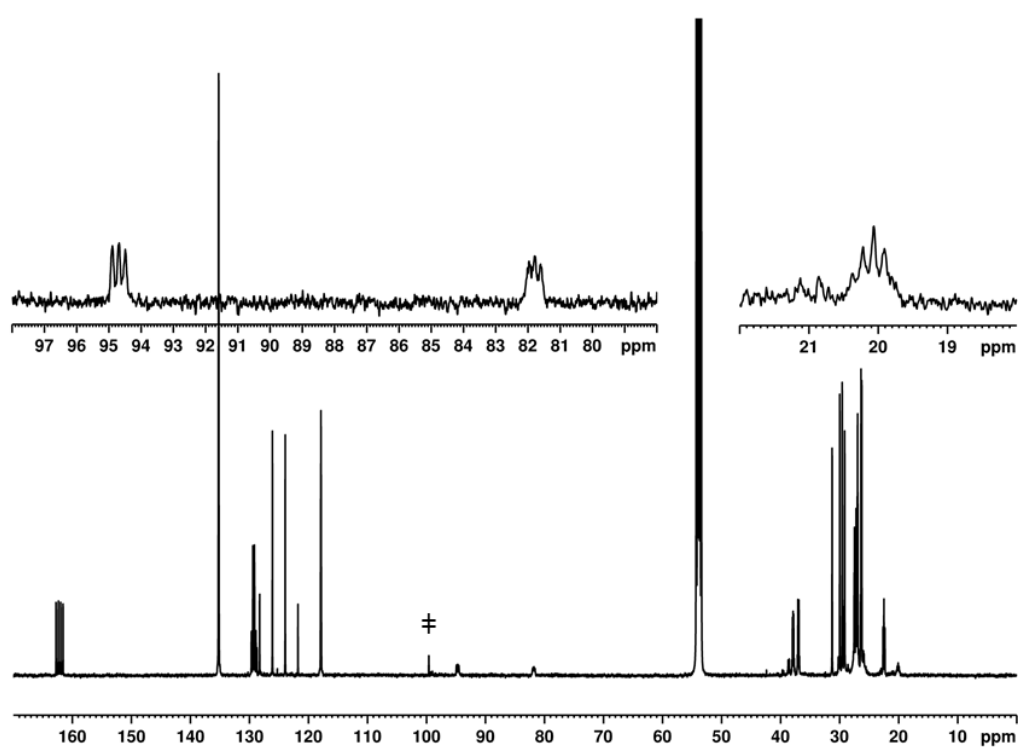


Figure S87: The solution $^{13}\text{C}\{^1\text{H}\}$ NMR (CD_2Cl_2 , 298 K, 126 MHz) spectrum of $[1\text{-C}_6\text{D}_8][\text{BAr}^{\text{F}}_4]$. The resonance marked ‡ is due to $[1\text{-C}_6\text{H}_6][\text{BAr}^{\text{F}}_4]$.

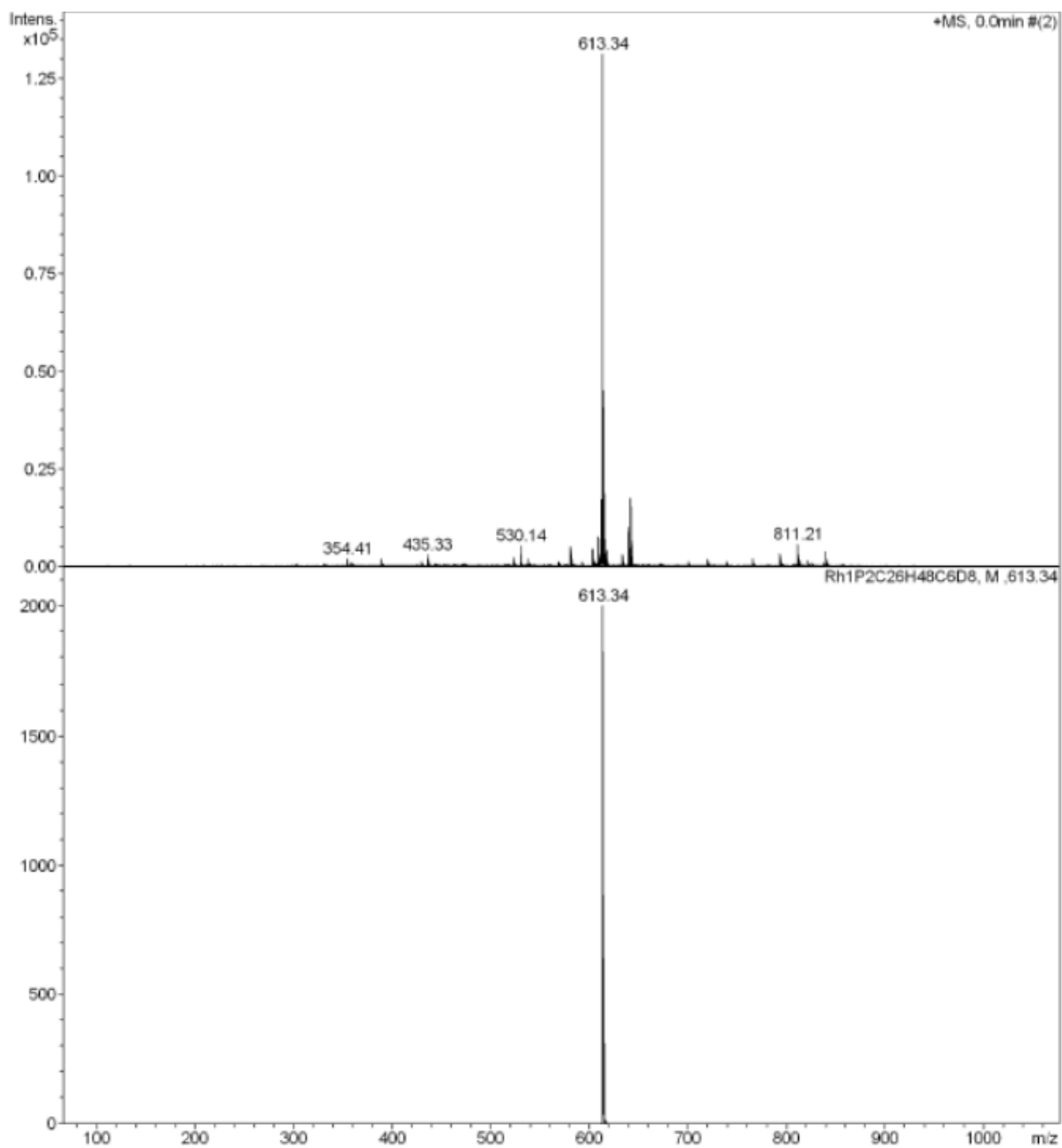


Figure S88: The experimental (top) and simulated (bottom) ESI mass spectra of [1-C₆D₈][BAr^F₄].

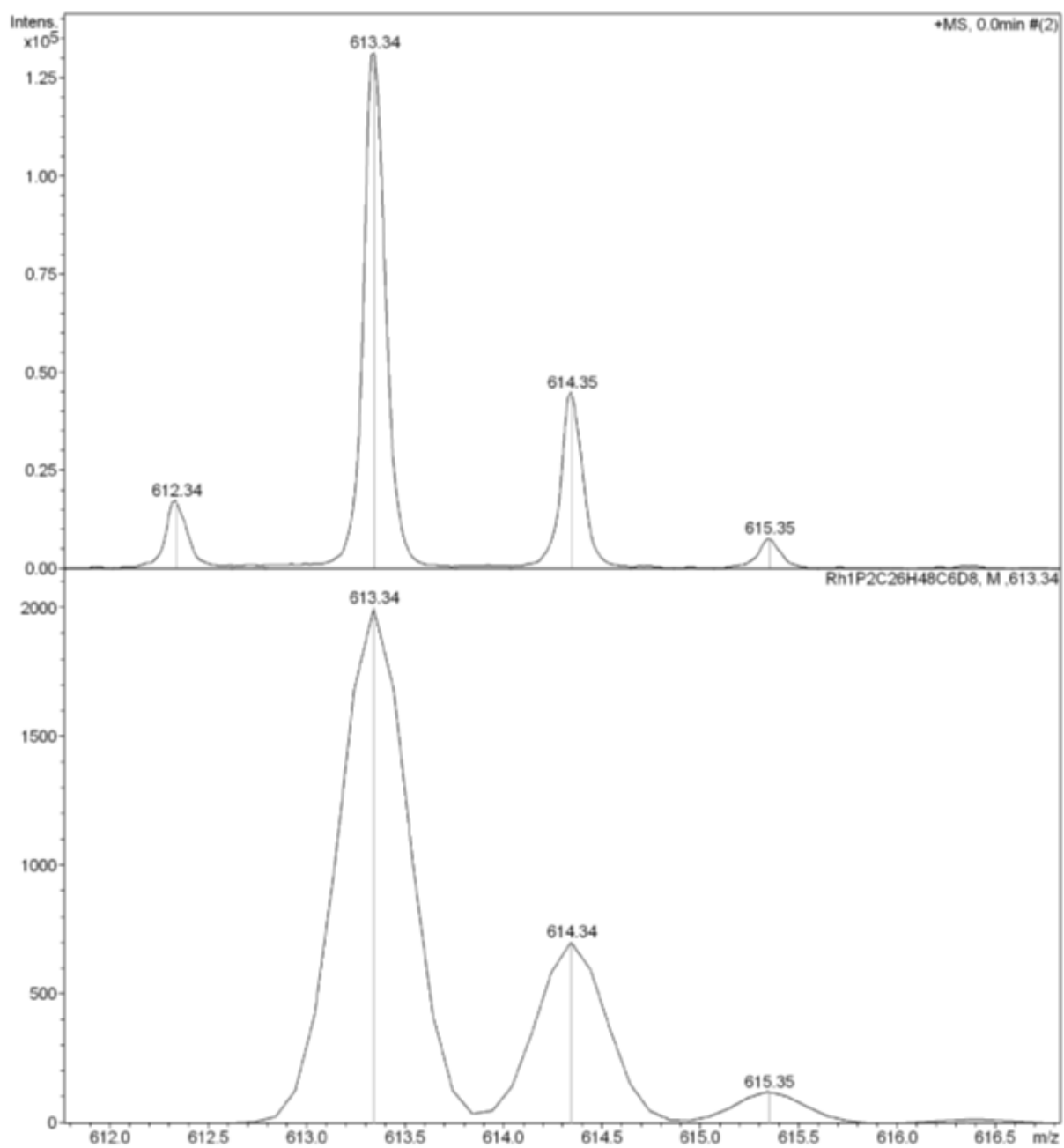


Figure S89: An enlargement of the ESI mass spectra of $[1\text{-C}_6\text{D}_8][\text{BAr}^{\text{F}}_4]$ showing the matching isotope patterns. The small peak at $m/z = 612.34$ is $[1\text{-C}_6\text{HD}_7]^+$ (calc. 612.34), which is presumably due to trace incomplete H/D exchange.

S.4.5.4. Liberation of cyclohexadiene-d₈

A CD₂Cl₂ or CH₂Cl₂ (0.4 mL) solution of [1-C₆D₈][BAr^F₄] in a J Young NMR tube was freeze-pump-thaw degassed three times. The sample was then carefully refilled with CO (1 bar, 298 K). Brief agitation of the sample resulted in a rapid change in the color of the solution from orange to yellow. The volatile component was then isolated by trap-to-trap distillation. The resultant solid was confirmed to be [Rh(dcpe)(CO)₂][BAr^F₄] by solution ³¹P{¹H} NMR spectroscopy (δ 85.22, d, J_{RhP} = 116 Hz).^{S7}

²H NMR (CH₂Cl₂, 298 K, 61 MHz): δ 5.91 (s, 2D), 5.81 (s, 2D), 2.08 (s, 4D).

¹³C{¹H} NMR (CD₂Cl₂, 298 K, 126 MHz): δ 126.1 (1:1:1 t, J_{CD} 24.5 Hz, alkene CD), 124.1 (1:1:1 t, J_{CD} 24.3 Hz, alkene CD), 21.3 (1:2:3:2:1 quintet, J_{CD} = 20 Hz, CD₂).

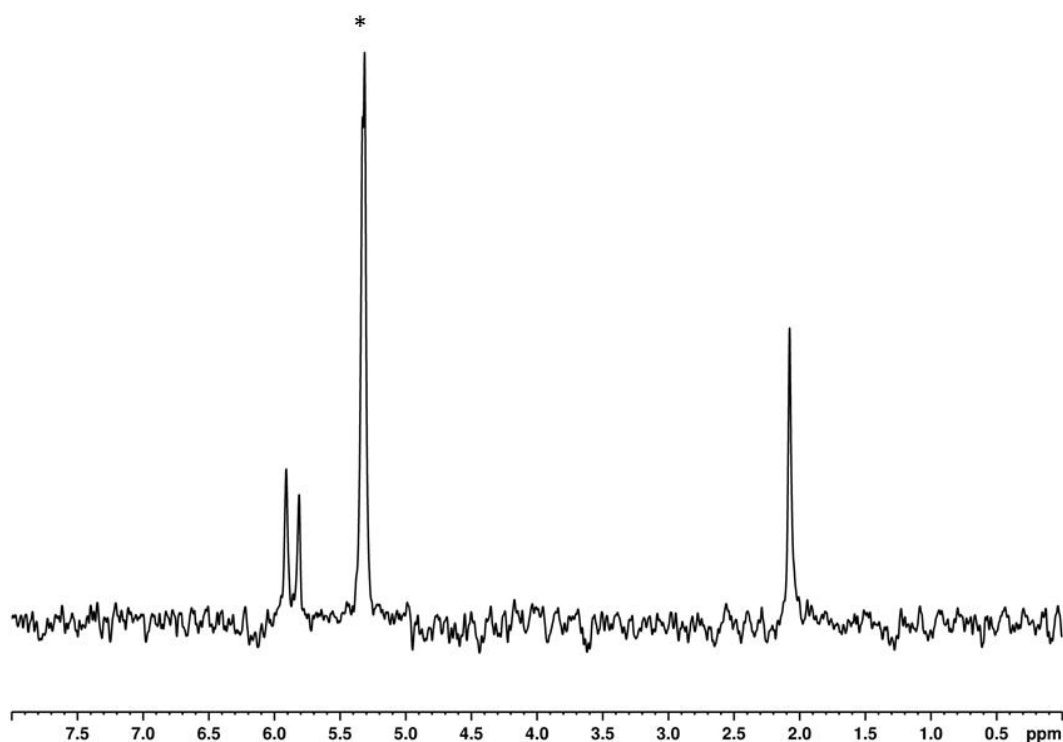


Figure S90: The solution ²H NMR (CH₂Cl₂, 298 K, 61 MHz) spectrum of liberated alkene. The resonance marked * is due to CHDCl₂.

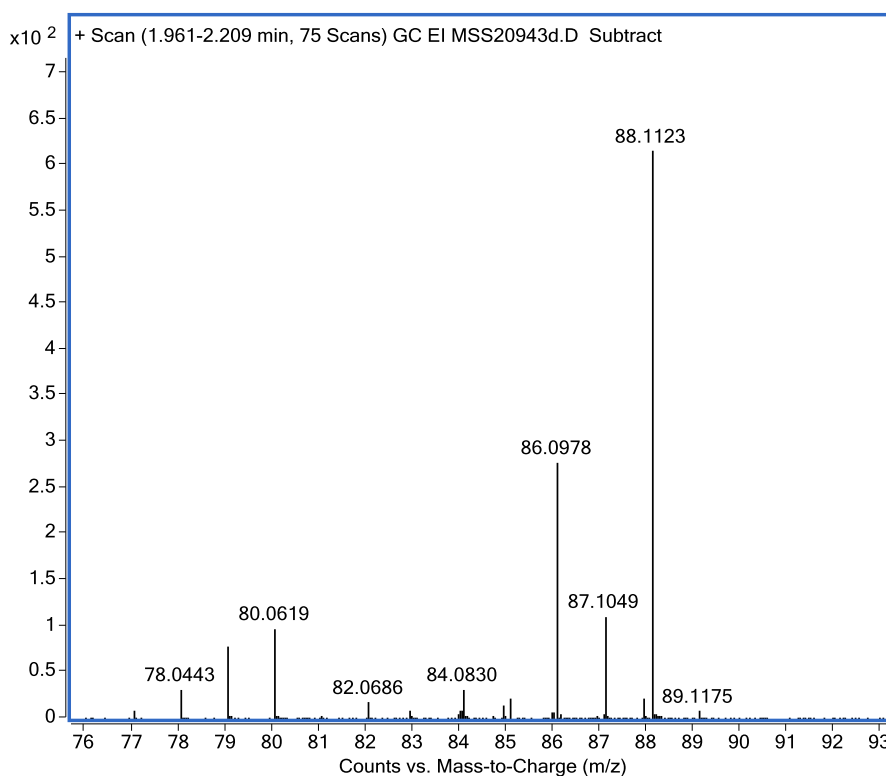
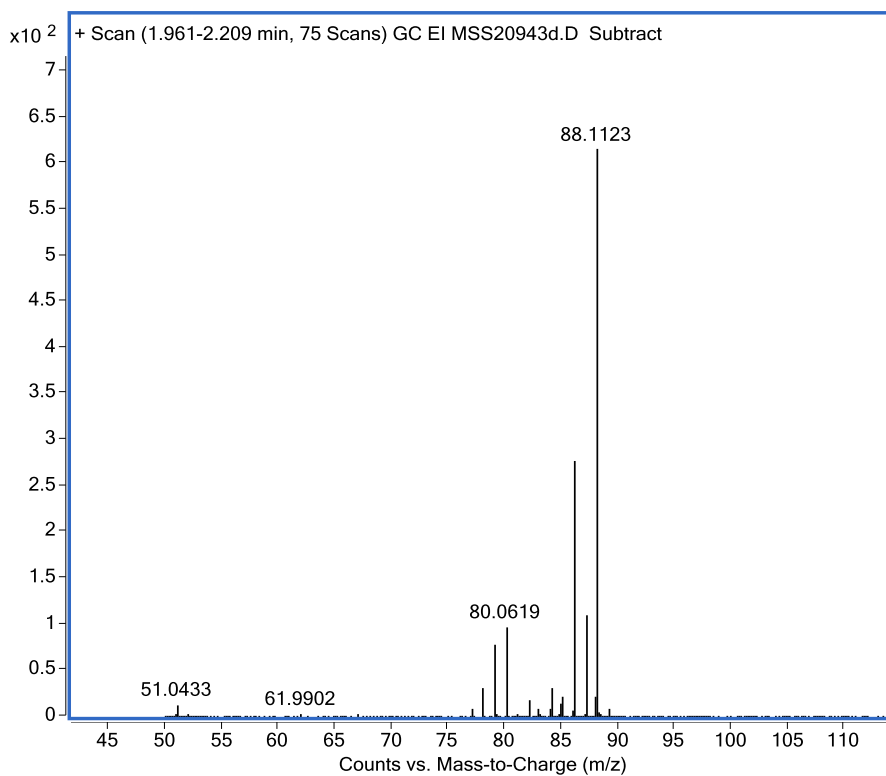


Figure S91: Mass spectrum of the isolated volatiles (top). Enlargement of $m/z = 76 - 93$ region (bottom). The signal at $m/z = 88.1123$ matches that of C_6D_8 (calc. 88.1123). The peak at $m/z = 86.0978$ represents that of C_6D_8-D (calc. 86.0987).

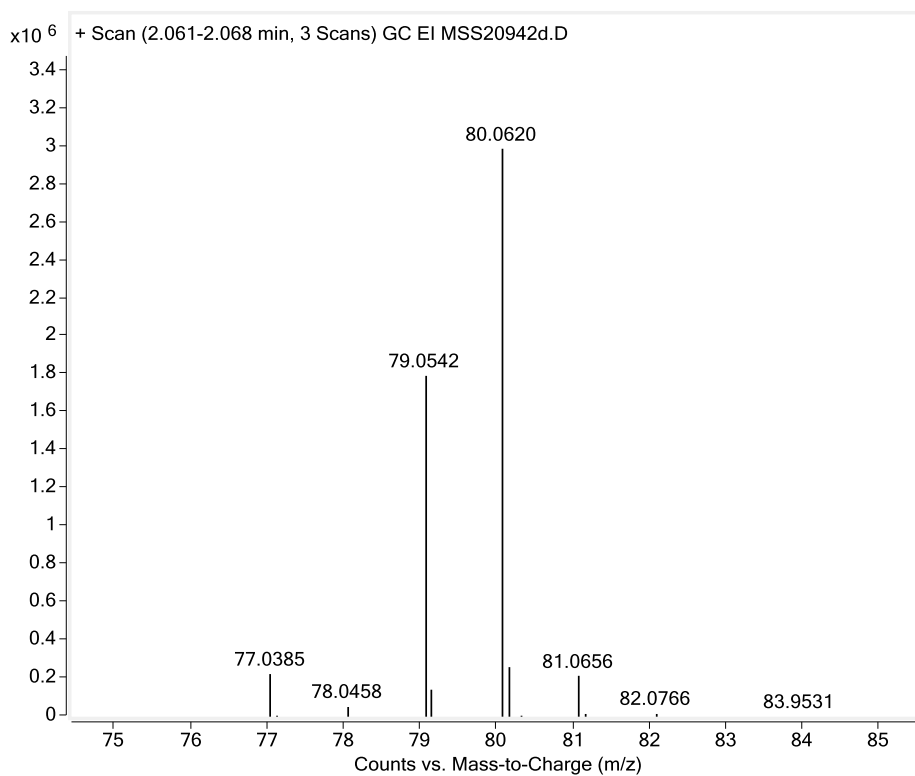
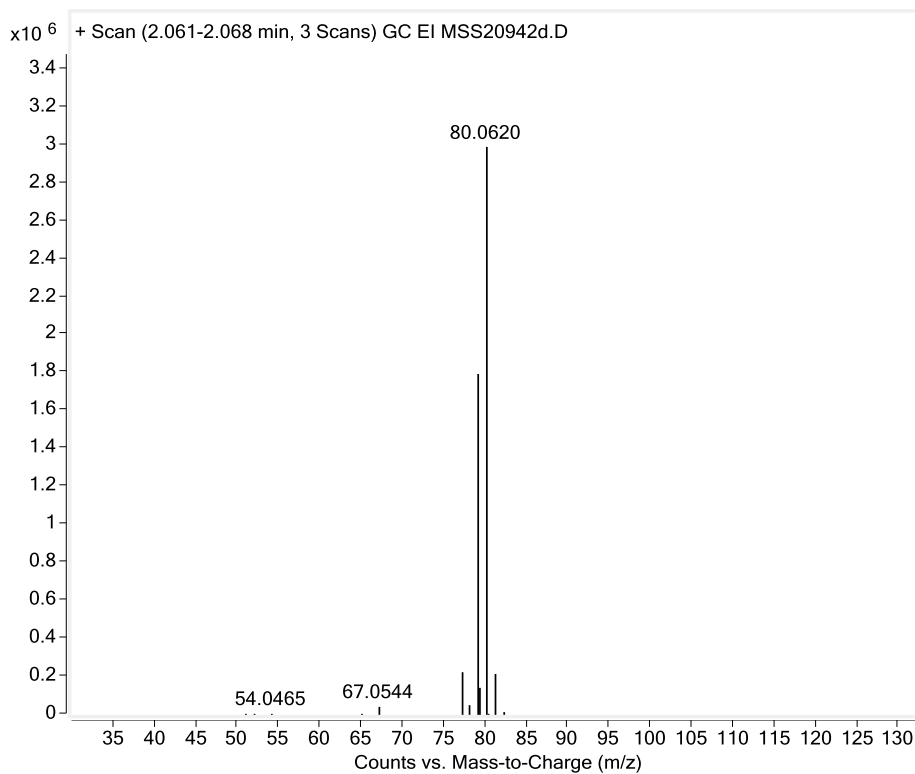


Figure S92: Mass spectrum of a commercially sourced sample of 1,3-cyclohexadiene (top). Enlargement of $m/z = 75 - 85$ region (bottom). The signal at $m/z = 80.0620$ matches that of $C_6H_8^+$ (calc. 80.0626). The peak at $m/z = 79.0542$ represents the $C_6H_7^+$ (calc. 79.0548).

S.5. Crystallographic and refinement data

S.5.1. Crystal structure determinations

Single crystal X-ray diffraction data for all samples were collected as follows: a typical crystal was mounted on a MiTeGen Micromounts using perfluoropolyether oil and cooled rapidly to the collection temperature in a stream of nitrogen gas using an Oxford Cryosystems Cryostream unit.^{S11} The structures of **[1-C₄H₈][BAr^F₄]** (150 K), **[1-C₄H₁₀][BAr^F₄]** (100 K) & **[1-C₆H₁₂][BAr^F₄]** (150 K) were collected at the Oxford Chemical Crystallography Service from the University of Oxford, with an Agilent SuperNova diffractometer (Cu K_α radiation, λ = 1.54180 Å). **[1-C₆H₈][BAr^F₄]** (250 K) was collected at the National Crystallography Service, University of Southampton, Southampton, UK, using a Rigaku 007HF diffractometer equipped with Varimax confocal mirrors (Cu K_α radiation, 1.54184 Å), an AFC11 goniometer, and HyPix 6000 detector.

Raw frame data were reduced using CrysAlisPro.^{S12} The structures were solved using SHELXT^{S13} and refined using full-matrix least squares refinement on all *F*² data using the SHELXL-18^{S14} using the interface OLEX2.^{S15} All hydrogen atoms were placed in calculated positions (riding model). Disorder of the -CF₃ groups was treated by introducing a split site model and restraining geometries and displacement parameters.

S.5.2. Additional comments crystal structures and refinement data

[1-C₄H₁₀][BAr^F₄]:

In reply to A-Level Alert [PLAT971_ALERT_2_A]: The origin of this residual electron density relates to a positional disorder affecting a dicyclohexylphosphino group from the 1,2-Bis(dicyclohexylphosphino)ethane phosphine ligand from the asymmetric unit. This is too small contribution to be satisfactorily modelled. As such it was deemed better to retain this electron density with a low R-factor.

In reply to B-Level Alert 3 [PLAT910_ALERT_3_B]: The sequential single-crystal to single-crystal transformation caused a loss in data quality at high angles. Because of

this data set was collected to satisfy a $\sin(\theta_{\max})/\lambda$ 0.82 giving an acceptable R-factor.

In reply to B-Level Alerts 2 [PLAT971_ALERT_2_B], [PLAT971_ALERT_2_B] and [PLAT973_ALERT_2_B]: The origin of this residual electron density relates to a positional disorder affecting a dicyclohexylphosphino group from the 1,2-Bis(dicyclohexylphosphino)ethane phosphine ligand from the asymmetric unit. This is too small contribution to be satisfactorily modelled. As such it was deemed better to retain this electron density with a low R-factor.

[1-C₄H₈][BAr^F₄]-SC-SC:

In reply to A-Level Alert [PLAT029_ALERT_3_A]: The sequential two single-crystal to single-crystal transformations caused a loss in data quality. Because of this, this data set was collected to satisfy an acceptable atom connectivity model only.

In reply to B-Level Alert [THETM01_ALERT_3_B]: The sequential two single-crystal to single-crystal transformations caused a loss in data quality at high angles. Because of this, this data set was collected to satisfy a $\sin(\theta_{\max})/\lambda$ of 0.56 giving a model for atom connectivity with an acceptable R-factor.

In reply to B-Level Alert [PLAT023_ALERT_3_B]: The sequential two single-crystal to single-crystal transformations caused a loss in data quality. Because of this, this data set was collected to satisfy an acceptable atom connectivity model only.

In reply to B-Level Alert [PLAT088_ALERT_3_B]: The sequential two single-crystal to single-crystal transformations caused a loss in data quality. Because of this, the data/parameter ratio is acceptable.

In reply to B-Level Alert [PLAT911_ALERT_3_B]: The sequential two single-crystal to single-crystal transformations caused a loss in data quality. Because of this, this data set was collected to satisfy an acceptable atom connectivity model only.

[1-C₆H₈][BAr^F₄]

Crystals of this material undergo a phase change upon slow cooling to 100 K. For this reason, the presented data was measured at 250 K.

[1-C₆H₁₂][BAr^F₄]

In reply to B-Level Alert [PLAT088_ALERT_3_B]: The single-crystal to single-crystal transformation caused a loss in data quality. Because of this, the data/parameter ratio is acceptable.

In reply to B-Level Alert [PLAT971_ALERT_2_B and [PLAT972_ALERT_2_B]: The origin of this residual electron density relates to a positional disorder affecting the Rh metal centre. This is too small contribution to be satisfactorily modelled. As such it was deemed better to retain this electron density with a low R-factor.

In reply to CB-Level Alert [PLAT971_ALERT_2_C] and [PLAT972_ALERT_2_C]: The origin of this residual electron density relates to a positional disorder affecting the Rh metal centre. This is too small contribution to be satisfactorily modelled. As such it was deemed better to retain this electron density with a low R-factor.

	[1-C ₄ H ₈][BAR ^F ₄]	[1-C ₄ H ₁₀][BAR ^F ₄]	[1-C ₄ H ₈][BAR ^F ₄]*	[1-C ₆ H ₈][BAR ^F ₄]	[1-C ₆ H ₁₂][BAR ^F ₄]
Chemical formula	C ₆₂ H ₆₈ BF ₂₄ P ₂ Rh	C ₆₂ H ₇₀ BF ₂₄ P ₂ Rh	C ₆₂ H ₆₈ BF ₂₄ P ₂ Rh	C ₆₄ H ₆₈ BF ₂₄ P ₂ Rh	C ₆₄ H ₇₂ BF ₂₄ P ₂ Rh
Formula weight	1444.82	1446.84	1444.82	1468.84	1472.87
Temperature (K)	150	100	150	250	150
Crystal system	triclinic	monoclinic	triclinic	monoclinic	monoclinic
Space group	P-1	P2 ₁ /n	P-1	C2/c	C2/c
a (Å)	12.7477(3)	19.1238(3)	12.7582(16)	19.2893(2)	19.1736(8)
b (Å)	13.0092(3)	34.6204(5)	12.9595(15)	17.2828(1)	17.6512(8)
c (Å)	19.8574(4)	19.5735(3)	19.8097(8)	20.2622(1)	19.7601(12)
α (deg)	92.3462(17)	90	92.028(6)	90	90
β (deg)	90.9972(16)	90.624(2)	90.685(7)	91.1490(1)	91.086(5)
γ (deg)	96.6033(17)	90	96.2300(10)	90	90
V (Å ³)	3267.70(13)	12958.3(3)	3253.5(6)	6753.52(9)	6686.3(6)
Z	2	8	2	4	4
ρ(calcd) (g cm ⁻³)	1.468	1.483	1.475	1.445	1.463
μ (mm ⁻¹)	3.530	3.561	3.545	3.426	3.461
Reflections collected	77320	160559	11700	32330	36704
Unique reflections	12411	24598	11700	6335	6351
Restraints Parameters	/ 791/1014	1706/1939	1171/1029	192/555	2524/806
R _{int}	0.0406	0.0943	0.0555	0.0349	0.0563
R ₁ [<i>I</i> > 2σ(<i>I</i>)]	0.0511	0.0949	0.1095	0.0548	0.1031
wR ₂ [all data]	0.1509	0.2543	0.2956	0.1709	0.2141
GooF	1.096	1.059	1.139	1.098	1.082
Residual electron density (e Å ⁻³)	2.38/-1.12	3.42/-1.76	1.99/-1.41	0.97/-0.44	3.45/-2.31
CCDC no.	1916756	1916755	N / A	1916748	1916750

Table S1: Selected crystallographic and refinement data. * is from the dehydrogenation of [1-C₄H₁₀][BAR^F₄], see section S.4.1.3.

S.5.3. Single Crystal X-ray Diffraction Images

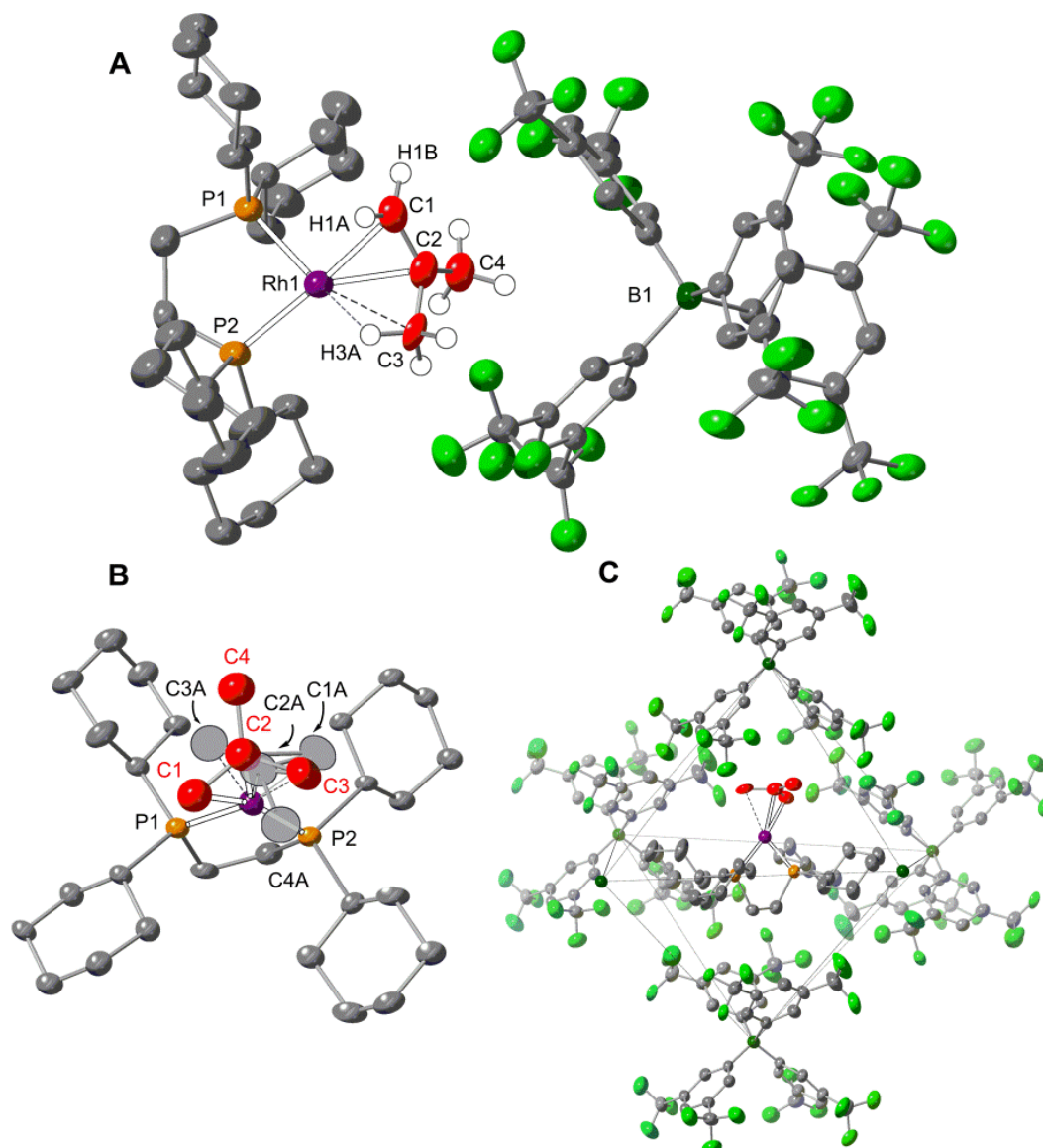


Figure S93: (A) Molecular structure of [1-C₄H₈][BArF₄]. (B) Cationic fragment of [1-C₄H₈][BArF₄] showing major disorder components, where the ratio of disorder components is 0.71: 0.29 for major: minor. (C) Structure displaying O_h anion network of the [BArF₄]⁻ ion, with cationic fragment sat within the cavity; -(ArF₄) removed from 2 boron atoms for clarity. Thermal ellipsoids set at 30 % and hydrogen from phosphine ligand and [BArF₄]⁻ anion removed for clarity in all parts.

Selected bond lengths (Å): Rh1-P1 2.2238(9), Rh1-P2 2.2400(9), Rh1-C1 2.262(6), Rh1-C2 2.136(8), Rh1-C3 2.368(9), Rh1-H3A 1.848, C1-C2 1.320(12), C2-C3 1.474(13), C2-C4 1.513(13).

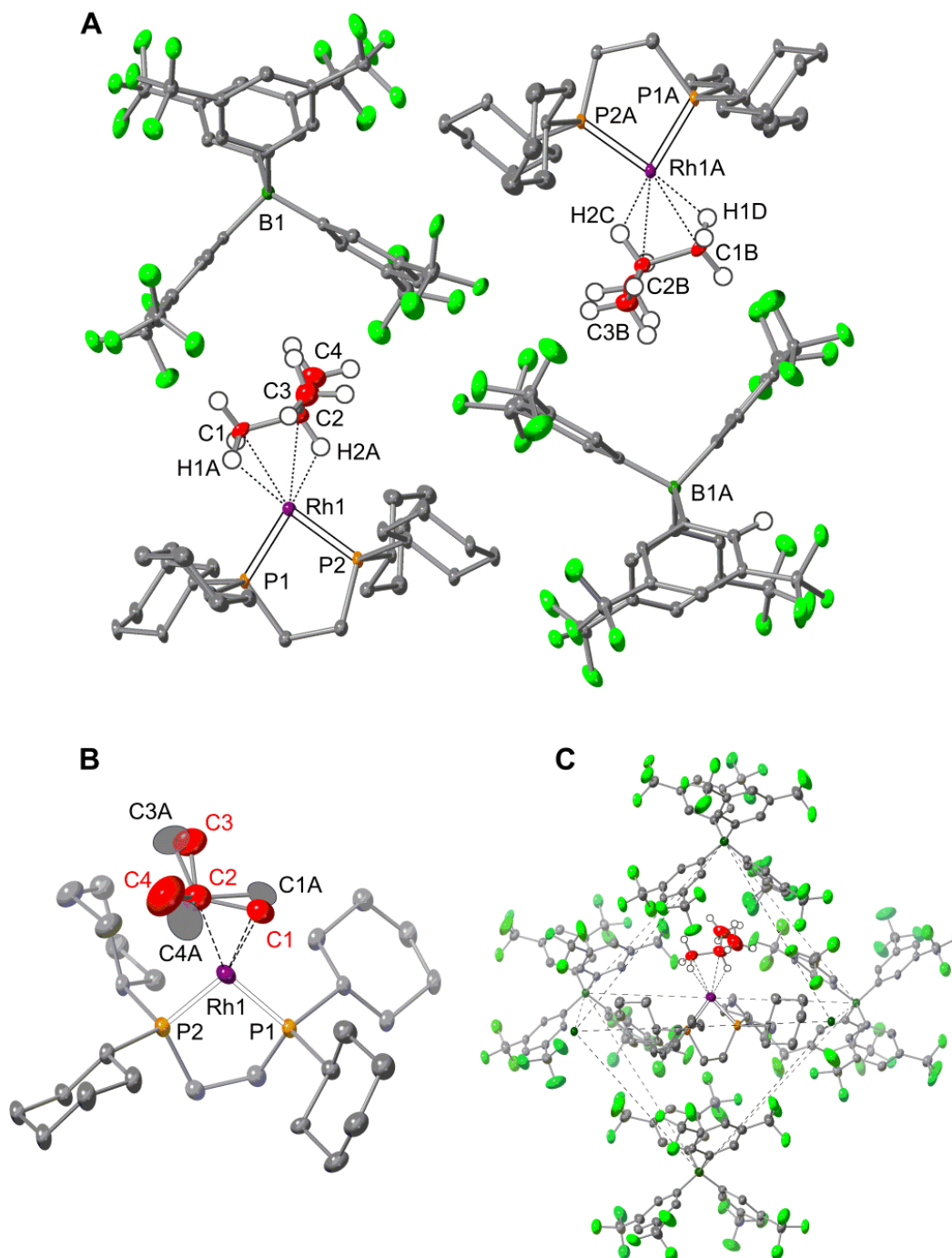


Figure S94: (A) Molecular structure of $[1-C_4H_{10}][BARF_4]$, showing only the major disorder components for both independent molecules in the asymmetric unit cell.

(B) Cationic fragment of the first molecule of $[1-C_4H_{10}][BARF_4]$ in the asymmetric cell, showing both disorder components (0.67: 0.33 for major (CX) and minor (CXA)). No disorder is present on C2, and is shared between the two disorder components. Not shown is the disorder in the second molecule in the asymmetric unit, however is

visually indistinguishable from that shown in **(B)**. The disorder ratio for the second molecule is 0.56: 0.44 for major: minor.

(C) Structure displaying O_h anion network of the $[\text{BAr}^{\text{F}}_4]^-$ ion, with cationic fragment sat within the cavity. $-(\text{Ar}^{\text{F}}_4)$ removed from 2 boron atoms for clarity. Thermal ellipsoids set at 30 % and hydrogen from phosphine ligand and $[\text{BAr}^{\text{F}}_4]^-$ anion removed for clarity in all parts.

Selected bond lengths for the 1st molecule (Å): Rh1-P1 2.1830(14), Rh1-P2 2.1914(14), Rh1-C1 2.362(14), Rh1-H1A 1.920, Rh1-C2 2.442(7), Rh1-H2A 1.8055, C1-C2 1.551(13), C2-C3 1.528(13), C2-C4 1.516(13).

Selected bond lengths for the 2nd molecule (Å): Rh1A-P1A 2.1841(14), Rh1A-P2A 2.1887(15), Rh1A-C1B 2.390(14), Rh1A-H1D 1.980, Rh1A-C2A 2.438(8), Rh1A-H2C 1.7978, C1B-C2B 1.544(16), C2B-C3B 1.500(19).

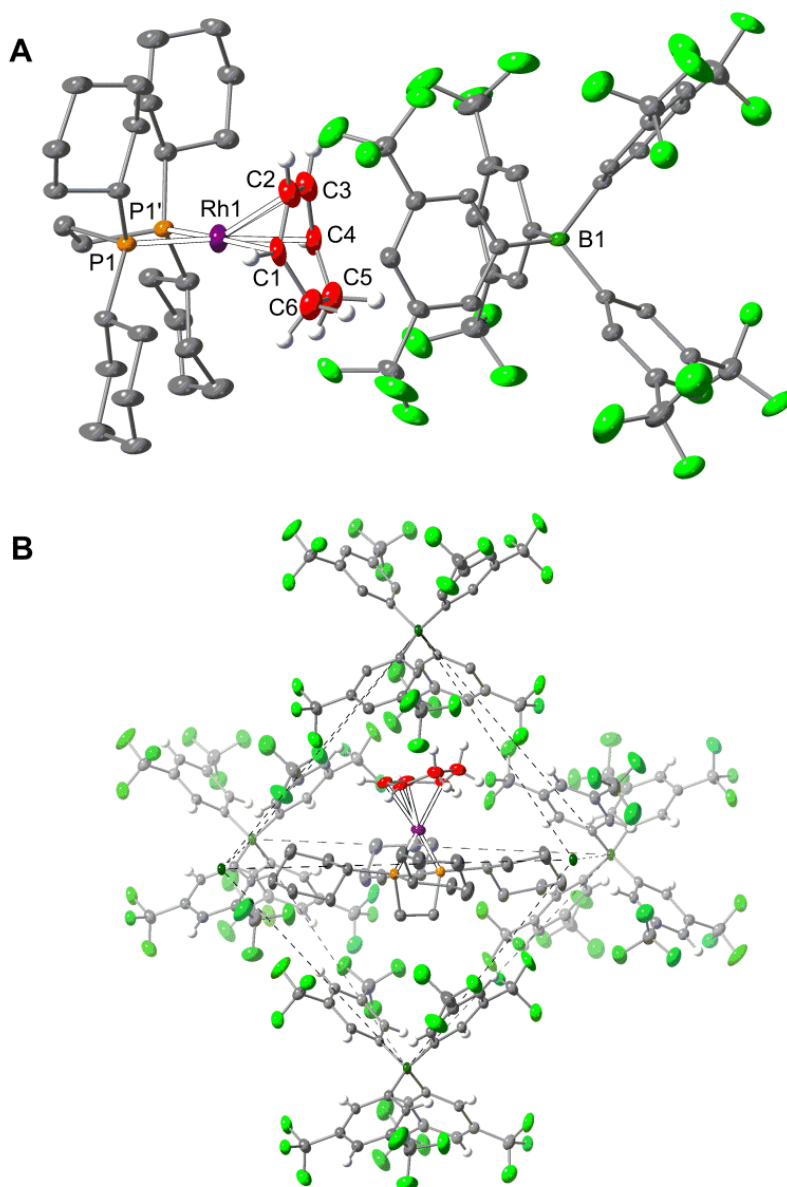


Figure S95: (A) Molecular structure of $[1-C_6H_8][BARF_4]$ displaying major disorder component only. Operation used to generate symmetry related atoms (P1 & P1') $1-x, y, \frac{1}{2}-z$. **(B)** Structure displaying O_h anion network of the $[BARF_4]^-$ ion, with cationic fragment sat within the cavity. $-(ArF_4)$ removed from 2 boron atoms for clarity. Thermal ellipsoids set at 30 % and hydrogens from phosphine ligand and $[BARF_4]^-$ anion removed for clarity in all parts.

Selected bond lengths (Å): Rh1-P1 2.2715(8), Rh1-P1' 2.2715(8), Rh1-C1 2.20(2), Rh1-C2 2.270(9), Rh1-C3 2.284(9), Rh1-C4 2.277(10), C1-C2 1.37(3), C2-C3 1.499(18), C3-C4 1.35(3), C4-C5 1.50(3), C5-C6 1.397(19), C6-C1 1.57(3).

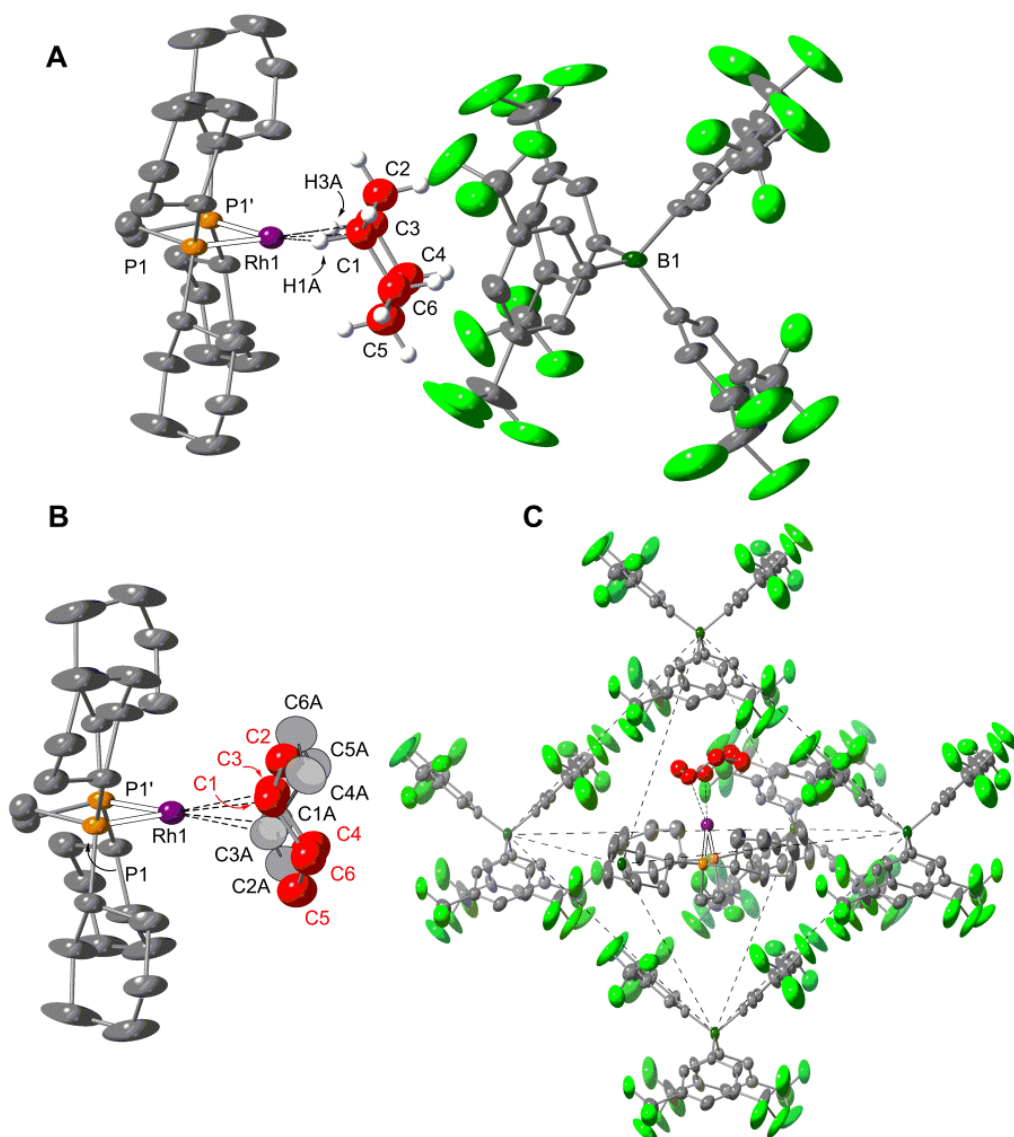


Figure S96: (A) Molecular structure of [1-C₆H₁₂][BARF₄]. (B) Cationic fragment of [1-C₆H₁₂][BARF₄] showing major disorder components (50%). Operation used to generate symmetry related atoms (P1 & P1') 1-x,y,1/2-z. (C) Structure displaying O_h anion network of the [BARF₄] ion, with cationic fragment sat within the cavity. -(ArF₄) removed from 1 boron atoms for clarity. Thermal ellipsoids set at 30 % and hydrogens from phosphine ligand and [BARF₄] anion removed for clarity in all parts.

Selected bond lengths (Å): Rh1-P1 2.191(2), Rh1-P1' 2.191(2), Rh1-C1 2.62(2), Rh1-H1A 1.9065, Rh1-C3 2.53(2), Rh1-H3A 1.7777, C1-C2 1.529(15), C2-C3 1.531(15), C3-C4 1.517(15), C4-C5 1.516(16), C5-C6 1.510(15), C6-C1 1.510(15).

S.6. Computational Methods

S.6.1. Solid-State Calculations

All static Kohn-Sham DFT calculations were performed on periodic models of the studied rhodium complexes, employing the Gaussian Plane Wave (GPW) formalism as implemented in the QUICKSTEP^{S16} module within the CP2K program suite (Version 5.0).^{S17} Molecularly optimized basis sets of double- ζ quality plus polarization in their short-range variant (DZVP-MOLOPT-SR-GTH)^{S18} were used on all atomic species. The interaction between the core electrons and the valence shell (Rh: 17, B: 3, C: 4, P: 5, F: 7, H: 1 electrons) was described by Goedecker-Teter-Hutter (GTH) pseudo potentials.^{S19-21} The generalized gradient approximation (GGA) to the exchange-correlation functional according to Perdew-Burke-Ernzerhof (PBE)^{S22} was used in combination with Grimme's D3-correction for dispersion interactions.^{S23} The auxiliary plane wave basis set was truncated at a cutoff of 500 Ry. The maximum force convergence criterion was set to 10^{-4} Eh-Bohr⁻¹, whilst default values were used for the remaining criteria. The convergence criterion for the self-consistent field (SCF) accuracy was set to 10^{-7} Eh and 10^{-8} Eh for geometry optimizations and vibrational analysis, respectively.

The Brillouin zone was sampled using the Γ -point. Initial coordinates for **[1-C₆H₁₂][BAr^F₄]**, **[1-C₆H₈][BAr^F₄]**, **[1-C₄H₁₀][BAr^F₄]** and **[1-C₄H₈][BAr^F₄]** were obtained from the experimental crystallographic data, with the hydrogen positions normalised where possible with Mercury.^{S50} Periodic boundary conditions (PBC) were applied throughout in combination with fixed unit cell parameters obtained from experiment. All geometries were first partially relaxed, keeping the heavy atoms (non-H, F) fixed, then fully relaxed without imposing any constraints, whilst keeping unit cell parameters constant in both cases. **[1-C₆H₁₂][BAr^F₄]** displays two crystallographically equivalent components. For **[1-C₄H₁₀][BAr^F₄]** two inequivalent disordered components are observed and all calculations were performed on the major component. Both independent cations were assessed in electronic structure and NMR calculations and gave similar results.

The structure of **[1-C₆H₁₀][BAr^F₄]** was constructed by removing appropriate hydrogen atoms from the experimental structure of **[1-C₆H₁₂][BAr^F₄]** under the constraint of retaining the crystal symmetry and fully optimising the remaining atomic

positions. This generated two possible structures depending on which C–C bond was dehydrogenated and the more stable form was used in the main paper.

For reactivity studies different reaction pathways were initially explored using an isolated rhodium molecular cation model with the Gaussian suite of programs (see details below). Transition states located in this way provided the basis for transition state searches in the solid state, with pre-optimisations in the solid state run by fixing the key reacting atoms at one of the Rh-centres. A partial vibrational analysis was then used to identify the corresponding imaginary mode. This pre-optimized TS structure was then refined using the dimer method^{S24} with the tighter convergence criteria detailed above. For challenging fluxional processes the climbing image nudged elastic band (CI-NEB) method,^{S25} using 8 or 12 images, was used to obtain candidate transition states that were then optimised using the dimer method as above. All optimized stationary points were characterized by analysis of their numerical second derivatives with a displacement of 0.01 Bohr. Minima and transition states have no or exactly one imaginary eigenvalue, respectively. All transition states were further analysed using an “IRC-like” approach, whereby transition state geometries were displaced along the negative mode in both directions and then fully optimising the two resulting structures. Further details on this protocol have been reported elsewhere.^{S26} A spurious imaginary frequency ($i = 6 \text{ cm}^{-1}$) was found in one of the stationary points (**IX**), which appears to be due to numerical inaccuracies in the vibrational analysis, which is a familiar issue in these type of calculations.^{S27-28}

Gibbs free energies for structures computed in the solid state were calculated using the TAMkin software toolkit.^{S29}

All computed structures are available as a separate file of Cartesian coordinates.

S.6.2. Molecular Calculations

Molecular calculations employed the GAUSSIAN 09 (revision D.01) program package^{S30} and employed the BP86 GGA functional.^{S31-32} Stuttgart-Dresden (SDD)^{S33} relativistic effective core potentials (ECP) in combination with the associated basis sets were utilized to describe Rh and P, with polarization functions added for P ($\zeta = 0.387$).^{S34} The 6-31G(d,p) basis sets^{S35-36} were used on remaining atoms.

Electronic structure analyses were performed on the geometries of the Rh cations extracted from the CP2K-optimised structures with the heavy atoms fixed at the experimental positions. An electron density file suitable for further analysis was generated from a single-point calculation.

The topology of the electron density was analysed by means of QTAIM (Quantum Theory of Atoms in Molecules),^{S37} as implemented in the AIMALL package.^{S38} Inner shell electrons on Rh and P modelled by ECPs were represented by core density functions (extended wavefunction format).

NBO calculations were performed using the NBO 6.0 program^{S39}, using the same geometries as for the QTAIM calculations above.

NCI calculations were performed using the NCIPLOT program^{S40-41}, using the nearest neighbour ion-pair molecular structures extracted from CP2K optimised geometries. The promolecular electron density was employed.

Orbital plots were created with Chemcraft^{S42} with an outer contour value of 0.0625.

S.6.3. NMR Calculations

Isotropic ^{13}C and ^1H magnetic shielding constants (σ_{iso}) were generated using the GIPAW method^{S43-44} as implemented in CASTEP 17.21.^{S45} For **[1-C₆H₁₂][BAR^F₄]** three approaches were assessed: Model 1 based on the fully optimised geometry in the solid-state; Model 2 based on the experimental structure with only the positions of H and F atoms being optimised and; an ion-pair model based on the nearest neighbour ion-pair extracted from the fully optimised geometry used in Model 1. Use of the ion-pair model gave comparable results. For **[1-C₄H₁₀][BAR^F₄]** the size of the unit cell precluded calculations on the full unit cell and so the ion-pair model was adopted. The plane wave basis set was truncated at 80 Ry, while the Brillouin zone was sampled using a Monkhorst-Pack grid^{S46} with a k-point spacing of 0.04 Å⁻¹. These settings were shown to yield converged NMR parameters on related systems.^{S26} The single-point calculations were performed using the PBE GGA functional,^{S22} using Ultra-soft pseudopotentials generated on-the-fly to represent the core electron shells. Scalar-relativistic effects were incorporated through the zeroth-order regular approximation (ZORA).^{S47-48} The ultrafine SCF convergence threshold of 10⁻⁸ eV atom⁻¹ was used throughout. Computed ^{13}C isotropic shielding constants were then converted into chemical shift values (δ) using the linear regression fit^{S49}

from the previously studied pentane system.^{S26} As previously relative isotropic proton (¹H) chemical shifts were obtained by referencing computed shielding constants against those of the standard tetramethylsilane (TMS) according to $\delta_{\text{iso}}(^1\text{H}) = \sigma_{\text{ref}}(\text{TMS}) - \sigma_{\text{iso}}$, where $\sigma_{\text{ref}}(\text{TMS})$ was calculated as δ 31.15 (cubic supercell with length 20 Å).^{S26}

S.7. Computed Reactions Profiles

S.7.1. Dehydrogenation of the Isolated $[1-C_6H_{12}]^+$ Cation

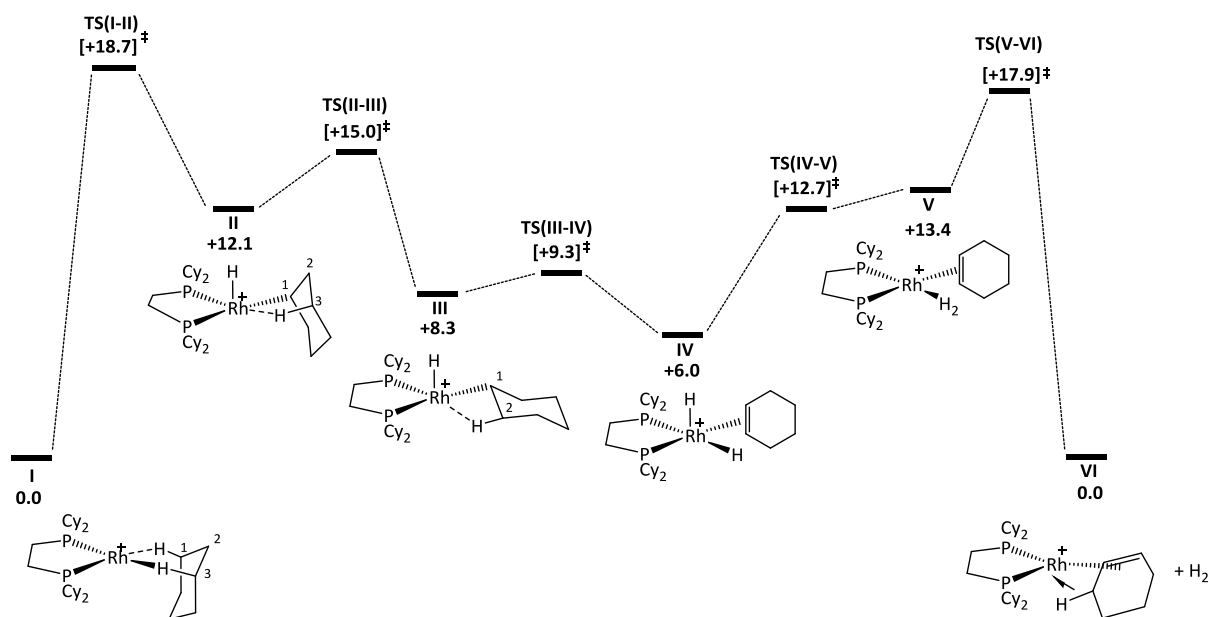


Figure S97: Computed free energy reaction profile (kcal/mol) for dehydrogenation of the isolated $[1-C_6H_{12}]^+$ cation via initial oxidative cleavage of the C-H¹ bond.

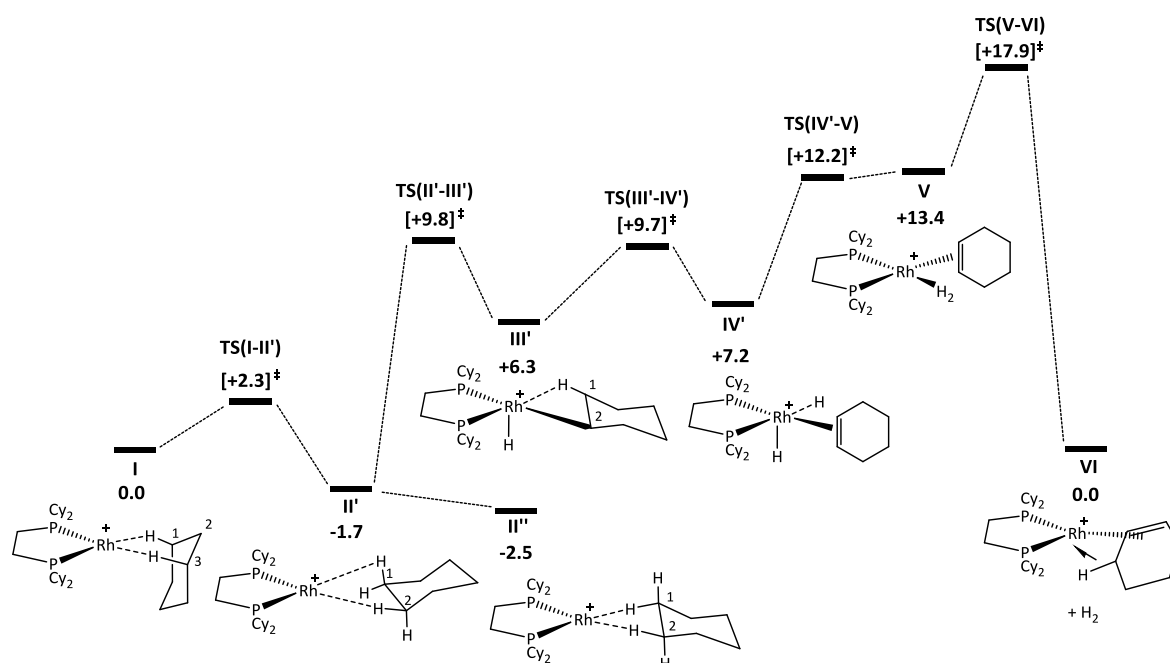


Figure S98: Computed free energy reaction profile (kcal/mol) for dehydrogenation of the isolated $[1-C_6H_{12}]^+$ cation via an initial rearrangement to a 1,2 binding mode followed by oxidative cleavage of the C-H² bond.

Isolated Cation	SCF Energy (a. u.)	Free Energy @ 298.15 K (a. u.)
I	-1379.280683	-1378.505513
TS (I – II)	-1379.248161	-1378.475735
II	-1379.259755	-1378.486258
TS (II – III)	-1379.255972	-1378.481609
III	-1379.265112	-1378.492212
TS (III – IV)	-1379.260011	-1378.49068
IV	-1379.267128	-1378.495987
TS (IV – V)	-1379.254042	-1378.485205
V	-1379.255114	-1378.484236
TS (V – VI)	-1379.245969	-1378.477052
VI	-1379.258177	-1378.505514
TS (I – II')	-1379.27733	-1378.501771
II'	-1379.28219	-1378.508175
II''	-1379.283105	-1378.509544
TS (II' – III')	-1379.260945	-1378.489911
III'	-1379.266156	-1378.495414
TS (III' – IV')	-1379.259012	-1378.490066
IV'	-1379.264991	-1378.493974
TS (IV' – V)	-1379.254484	-1378.486136

Table S2: Computed energies of stationary points along the pathways shown in Figures S97 and S98.

S.7.2. Dehydrogenation of the Isolated $[1\text{-C}_6\text{H}_{10}]^+$ Cation

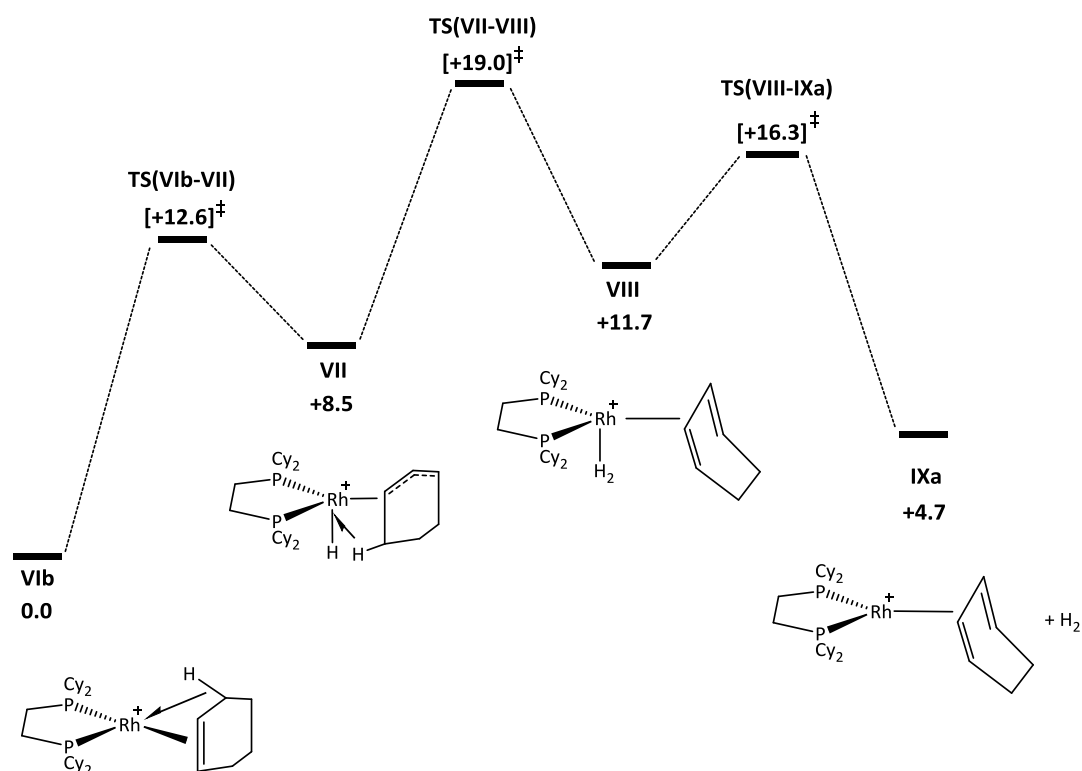


Figure S99: Computed free energy reaction profile (kcal/mol) for dehydrogenation of the isolated $[1\text{-C}_6\text{H}_{10}]^+$ cation via initial oxidative cleavage to form an exo-allyl hydride intermediate.

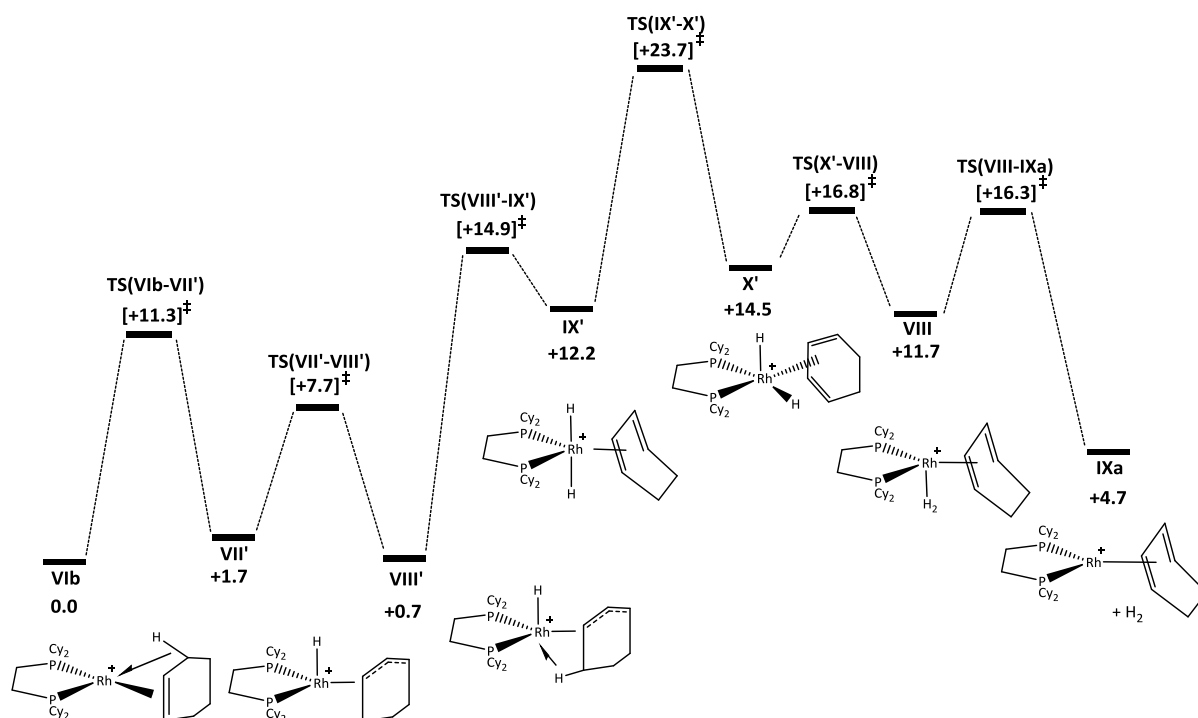


Figure S100: Computed free energy reaction profile (kcal/mol) for dehydrogenation of the isolated $[1\text{-C}_6\text{H}_{10}]^+$ cation via initial oxidative cleavage of the agostic C-H bond to form an endo-allyl hydride intermediate.

Isolated Cation	SCF Energy (a. u.)	Free Energy @ 298.16 K (a. u.)
VIb	-1378.081611	-1377.327262
TS (VIb – VII)	-1378.0601	-1377.307252
VII	-1378.067685	-1377.313707
TS (VII – VIII)	-1378.047275	-1377.296996
VIII	-1378.058924	-1377.308609
TS (VIII – IXa)	-1378.048014	-1377.301305
IXa	-1378.055202	-1377.319806
TS (VIb – VII')	-1378.061473	-1377.309208
VII'	-1378.077601	-1377.324588
TS (VII' – VIII')	-1378.068109	-1377.314998
VIII'	-1378.08005	-1377.326224
TS (VIII' – IX')	-1378.05293	-1377.303595
IX'	-1378.056535	-1377.307883
TS (IX' – X')	-1378.038878	-1377.289551
X'	-1378.055905	-1377.304155
TS (X' – VIII)	-1378.0513	-1377.300499

Table S3: Computed energies of stationary points along the pathways shown in Figures S99 and S100.

S.7.3. Dehydrogenation of [1-C₆H₁₂][BAR^F₄] in the Solid-State

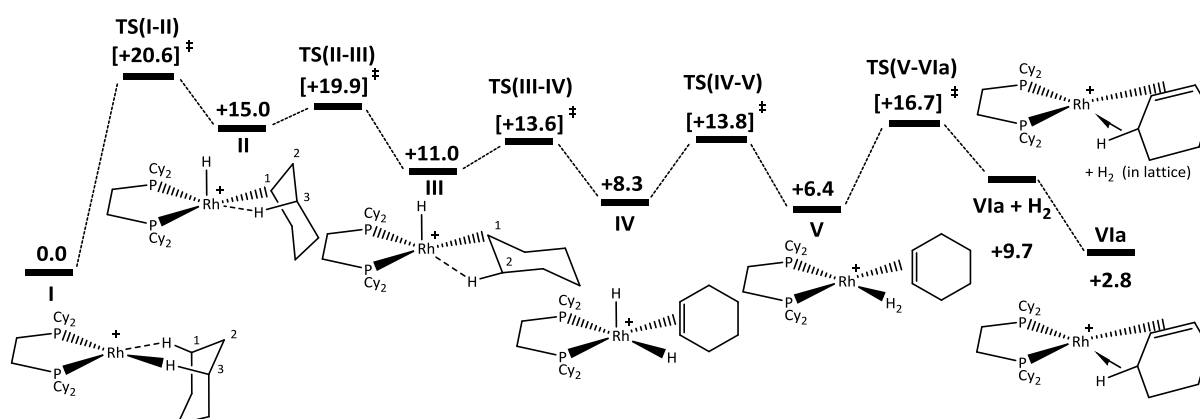


Figure S101: Computed free energy reaction profile (kcal/mol) for dehydrogenation of the [1-C₆H₁₂][BAR^F₄] system via initial oxidative cleavage of the C-H¹ bond. Only the reacting Rh complex is shown with [BAR^F₄]⁻ anions and spectator Rh cations omitted for clarity.

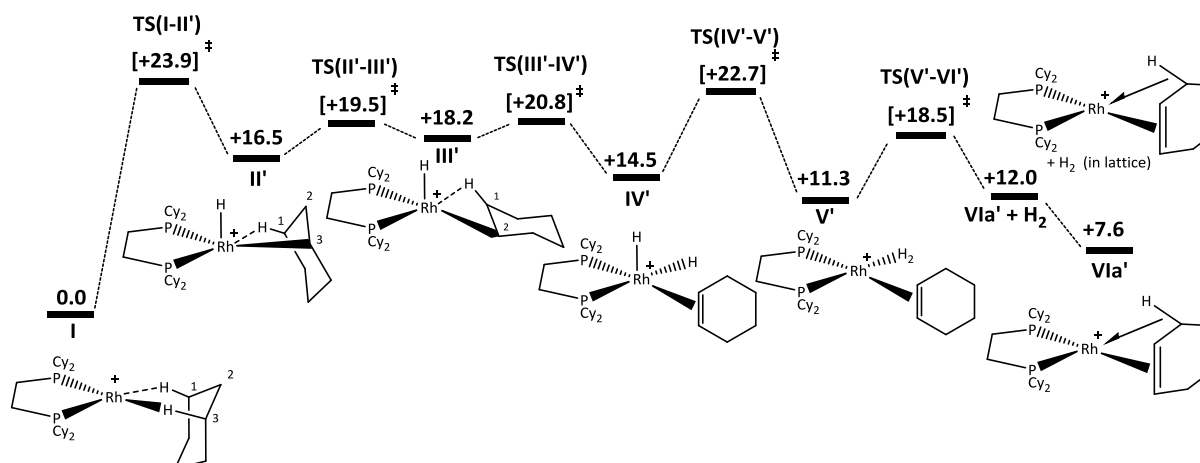


Figure S102: Computed free energy reaction profile (kcal/mol) for dehydrogenation of the [1-C₆H₁₂][BAR^F₄] system via initial oxidative cleavage of the C-H³ bond. Only the reacting Rh complex is shown with [BAR^F₄]⁻ anions and spectator Rh cations omitted for clarity.

Solid State	SCF Energy (a. u.)	SCF Energy Diff. (kcal / mol)
I	-4462.088766	0.0
TS (I – II)	-4462.053131	22.3609
II	-4462.063259	16.0057
TS (II – III)	-4462.056574	20.2009
III	-4462.061968	16.8156
TS (III – IV)	-4462.055429	20.9190
IV	-4462.065180	14.8002
TS (IV – V)	-4462.053137	22.3571
V	-4462.065450	14.6306
TS (V – VIa)	-4462.044764	27.6116
VIa + H ₂	-4462.056994	19.9371
VIa	-4460.891269	22.3211
TS (I – II')	-4462.048997	24.9551
II'	-4462.060729	17.5930
TS (II' – III')	-4462.056793	20.0633
III'	-4462.062665	16.3784
TS (III' – IV')	-4462.055583	20.8225
IV'	-4462.064813	15.0303
TS (IV' – V')	-4462.051139	23.6108
V')	-4462.064886	14.9847
TS (V' – VIa')	-4462.049781	24.4632
VIa' + H ₂	-4462.056830	20.0397
VIa'	-4460.891062	22.4514

Table S4: Computed energies of stationary points along the pathways shown in Figures S101 and S102.

S.7.4. Dehydrogenation of [1-C₆H₁₀][BAR^F₄] in the Solid-State

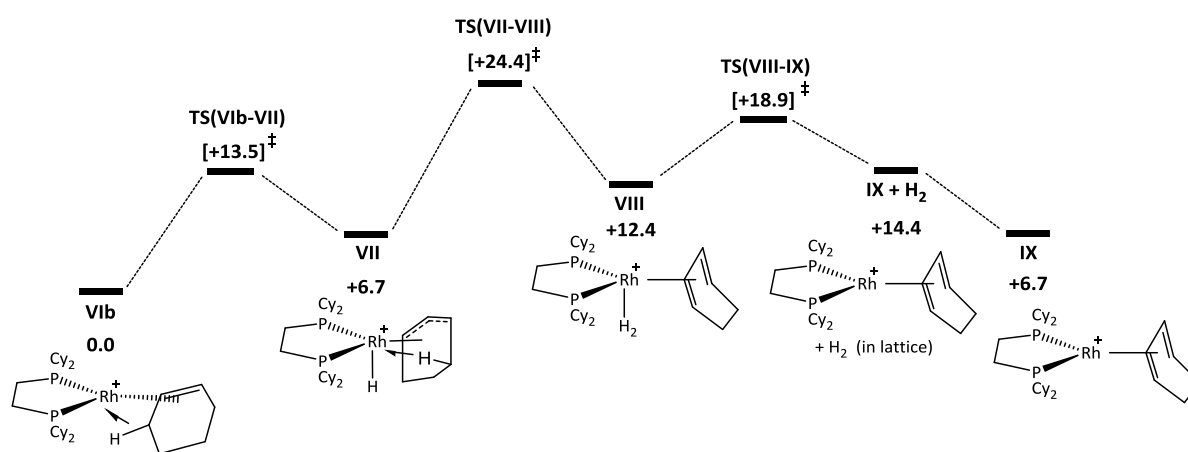


Figure S103: Computed free energy reaction profile (kcal/mol) for dehydrogenation of the [1-C₆H₁₀][BAR^F₄] system via initial oxidative cleavage of the left agostic C-H bond, forming an exo-allyl hydride. Only the reacting Rh complex is shown with [BAR^F₄]⁻ anions and spectator Rh cations omitted for clarity.

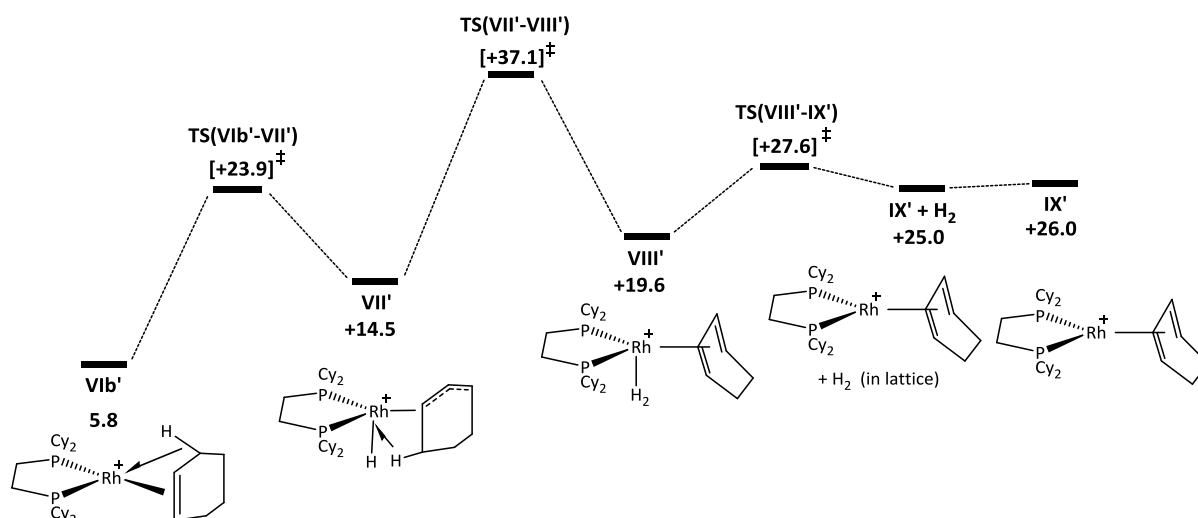


Figure S104: Computed SCF energy reaction profile (kcal/mol) for dehydrogenation of the [1-C₆H₁₀][BARF₄] system via initial oxidative cleavage of the right agostic C-H bond, forming an endo-allyl hydride. Only the reacting Rh complex is shown with [BARF₄]⁻ anions and spectator Rh cations omitted for clarity.

Solid State	SCF Energy (a. u.)	SCF Energy Diff. (kcal / mol)
Vib	-4457.310242	0.0
TS (Vib – VII)	-4457.281984	17.7317
VII	-4457.295942	8.9730
TS (VII – VIII)	-4457.262857	29.7343
VIII	-4457.284235	16.3195
TS (VIII – IX)	-4457.270651	24.8434
IX + H ₂	-4457.275759	21.6380
IX	-4456.111506	23.0988
Vib'	-4457.300921	5.8487
TS (Vib' – VII')	-4457.272084	23.9444
VII'	-4457.287112	14.5142
TS (VII' – VIII')	-4457.251144	37.0842
VIII'	-4457.279026	19.5859
TS (VIII – IX')	-4457.266255	27.6021
IX' + H ₂	-4457.270451	24.9687
IX'	-4456.106834	26.0306

Table S5: Computed energies of stationary points along the pathways shown in Figures S103 and S104.

S.7.5. Cyclohexane rearrangement processes of $[1\text{-C}_6\text{H}_{12}][\text{BAR}^{\text{F}}_4]$ in the Solid State

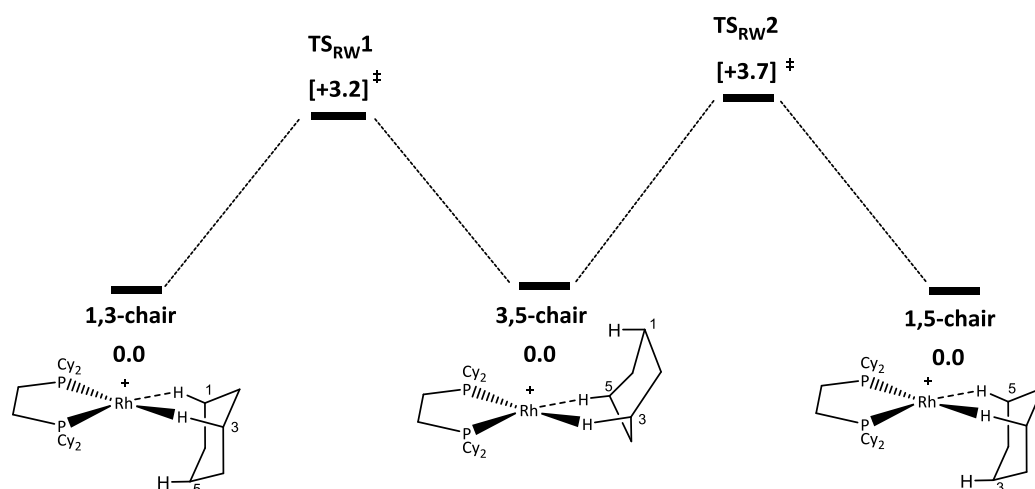


Figure S105: Computed free energy reaction profile (kcal/mol) for the 1,3,5 ring walk in $[1\text{-C}_6\text{H}_{12}][\text{BAR}^{\text{F}}_4]$. Only the reacting Rh complex is shown with $[\text{BAR}^{\text{F}}_4]^-$ anions and spectator Rh cations omitted for clarity.

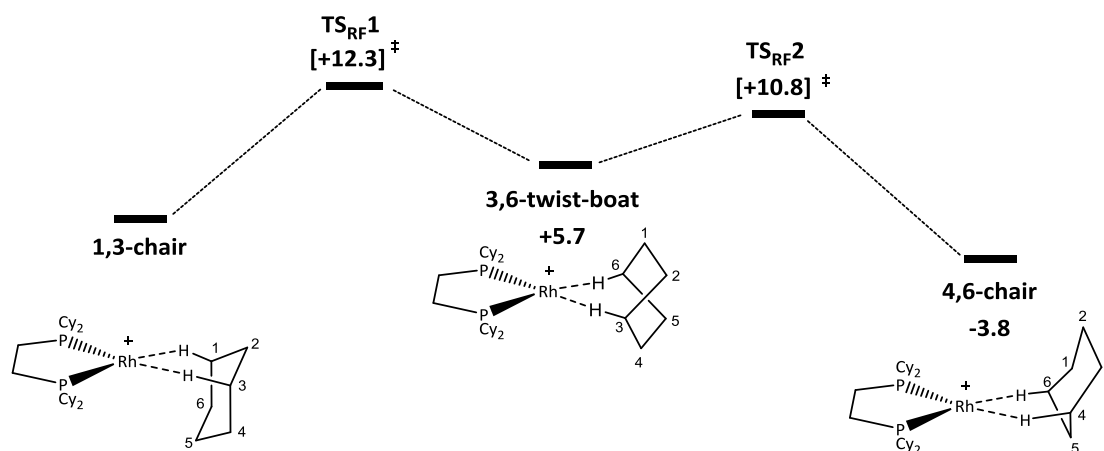


Figure S106: Computed free energy reaction profile (kcal/mol) for the ring flip of the $[1\text{-C}_6\text{H}_{12}][\text{BAR}^{\text{F}}_4]$ system. Only the reacting Rh complex is shown with $[\text{BAR}^{\text{F}}_4]^-$ anions and spectator Rh cations omitted for clarity.

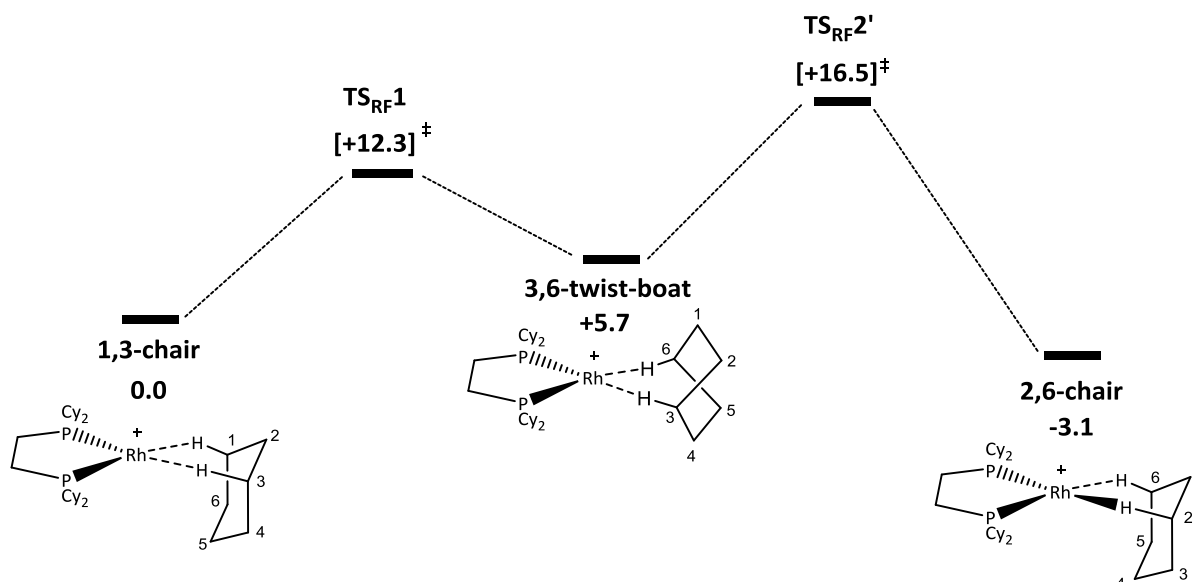


Figure S107: Computed free energy reaction profile (kcal/mol) for an alternative ring flip of the $[1\text{-C}_6\text{H}_{12}][\text{BAR}^{\text{F}}_4]$ system. Only the reacting Rh complex is shown with $[\text{BAR}^{\text{F}}_4]^-$ anions and spectator Rh cations omitted for clarity.

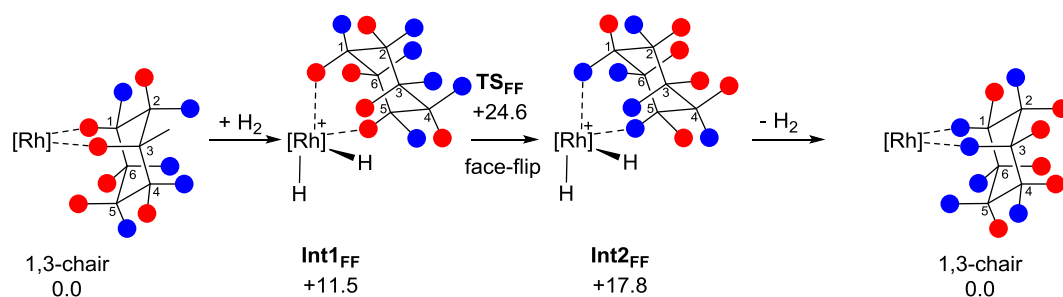


Figure S108: Computed free energies (kcal/mol) of key intermediates involved in the cyclohexane face flip in the $[1\text{-C}_6\text{H}_{12}][\text{BAR}^{\text{F}}_4]$ system, highlighting the movement of the H atoms involved. Only the reacting Rh complex is shown with $[\text{BAR}^{\text{F}}_4]^-$ anions and spectator Rh cations omitted for clarity.

Solid State	SCF Energy (a. u.)	SCF Energy Diff. (kcal / mol)
1,3-chair	-4462.088766	0.0
TS _{RW1}	-4462.086084	1.6825
3,5-chair	-4462.088565	0.1261
TS _{RW2}	-4462.085353	2.1414
1,5-chair	-4462.088762	0.0022
TS _{RF1}	-4462.068397	12.7814
3,6-twist-boat	-4462.080250	5.3436
TS _{RF2}	-4462.067364	13.4299
4,6-chair	-4462.085361	2.1363
TS _{RF2'}	-4462.065488	14.6071
2,6-chair	-4462.087509	0.7886
Int1 _{FF}	-4463.251501	-0.5076
TS1 _{FF}	-4463.231182	12.2425
Int2 _{FF}	-4463.242107	5.3869

Table S6: Computed energies of stationary points along the fluxional processes shown in Figures S105-S108.

S.8. Computed NMR Chemical Shifts

S.8.1. $[1-C_4H_{10}][BAr^F_4]$

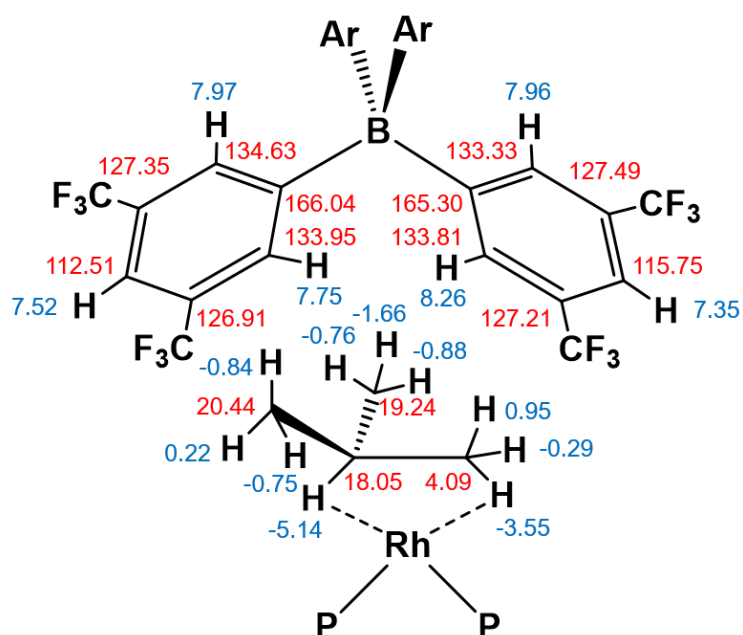


Figure S109: Computed NMR chemical shifts (ppm) for $[1-C_4H_{10}][BAr^F_4]$ (cation 1).

1H chemical shifts are shown in blue and ^{13}C shifts in red.

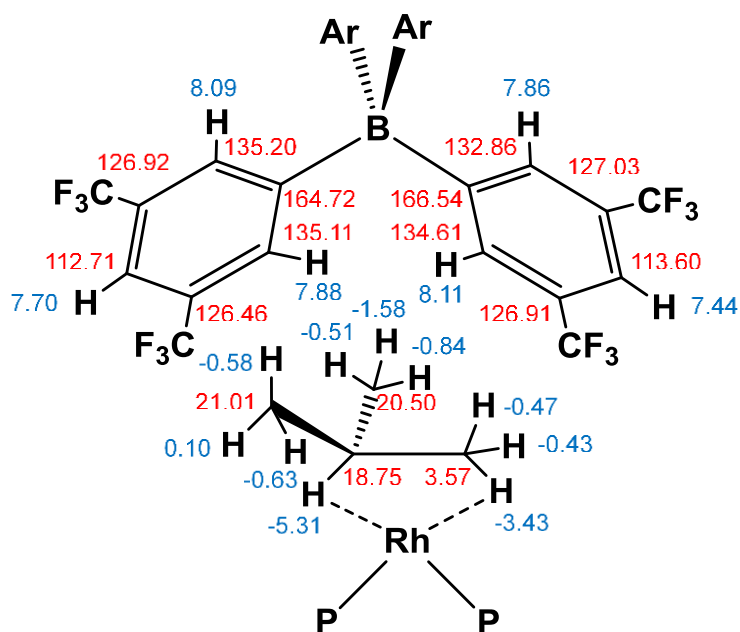


Figure S110: Computed NMR chemical shifts (ppm) for $[1-C_4H_{10}][BAr^F_4]$ (cation 2).

1H chemical shifts are shown in blue and ^{13}C shifts in red.

S.8.2. [1-C₆H₁₂][BAR^F₄]

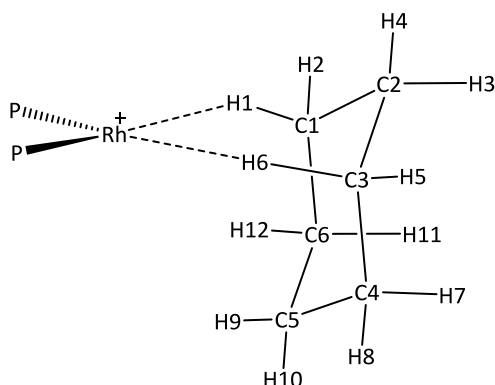


Figure S111: Labelling scheme employed.

Nucleus	Model 1			Model 2			Ion Pair		
	σ^{iso}	δ^1H	$\delta^{13}C$	σ^{iso}	δ^1H	$\delta^{13}C$	σ^{iso}	δ^1H	$\delta^{13}C$
H1	37.1528	-6.00		36.1609	-5.01		37.6566	-6.51	
H2	30.0546	1.10		30.4775	0.67		30.5041	0.65	
H3	30.7537	0.40		31.0223	0.13		31.1643	-0.01	
H4	29.3258	1.82		28.8933	2.26		30.0916	1.06	
H5	29.7757	1.37		29.9983	1.15		30.3096	0.84	
H6	36.7497	-5.60		35.7647	-4.61		37.2708	-6.12	
H7	32.8162	-1.67		32.8452	-1.70		33.0983	-1.95	
H8	30.2027	0.95		30.0231	1.13		30.8968	0.25	
H9	30.7822	0.37		31.0827	0.07		31.4869	-0.34	
H10	29.4862	1.66		29.0109	2.14		30.2264	0.92	
H11	32.9866	-1.84		32.3977	-1.25		33.2908	-2.14	
H12	30.0397	1.11		29.5275	1.62		30.7145	0.44	
C1	148.0118		23.74	157.3883		15.60	147.6427		24.10
C2	149.5209		22.29	148.8721		23.89	151.8025		20.08
C3	149.3763		22.43	153.8161		19.08	148.929		22.86
C4	145.8353		25.85	144.1107		28.53	146.2955		25.40
C5	144.7839		26.86	144.4966		28.16	144.7953		26.85
C6	145.1803		26.48	143.0411		29.58	145.8146		25.87

Table S7: [1-C₆H₁₂][BAR^F₄] computed isotropic chemical shifts (σ^{iso}) and relative chemical shifts (ppm) using the labelling scheme shown in Figure S108.^a

^a Model 1 data are based on the fully optimised geometry of [1-C₆H₁₂][BAR^F₄] in the solid-state; Model 2: data are based on the experimental structure with only the positions of H and F atoms being optimised; ion-pair model based on the nearest neighbour ion-pair extracted from the fully optimised geometry in Model 1.

S.9. Electronic Structure Analyses

S.9.1. [1-C₄H₁₀][BAr^F₄]

Note: Analyses were performed for both crystallographically independent cations contained in the unit cell.

Distance (Å)	Fully Optimized	Heavy Atom Fixed ^a	Experiment
Rh – P2	2.200	2.183	2.183(1)
Rh – P3	2.207	2.191	2.191(1)
Rh – H34	1.923	1.933	1.920
Rh – H37	1.899	1.873	1.806
Rh – C4	2.361	2.363	2.363(14)
Rh – C5	2.480	2.442	2.442(7)
C4 – H34	1.150	1.148	0.981
C4 – H35	1.099	1.099	0.980
C4 – H36	1.096	1.094	0.980
C5 – H37	1.159	1.160	1.000
Bite Angle (°)	87.401	89.322	76.96

Table S8: Key bond distances (Å) for 1st Cation from fully optimised and heavy atoms fixed geometries of [1-C₄H₁₀][BAr^F₄], with experimental data for comparison.^{S31}

^a structure is based on the experimental structure with only the positions of H and F atoms being optimised.

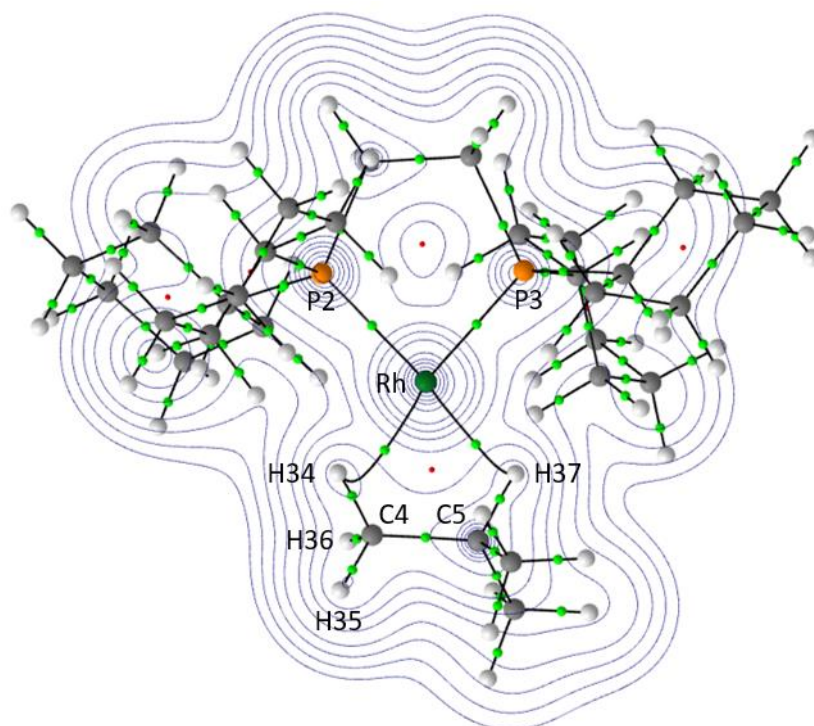


Figure S112: QTAIM molecular graph for 1st cation of **[1-C₄H₁₀][BAR^F₄]** in with density contours plotted in the Rh-H34-H37 plane.^{S30} Bond critical points (BCPs) are indicated in green and ring critical points (RCPs) are indicated in red.

Bond	$\rho(r) \text{ le}\text{\AA}^{-3}$	$\nabla^2 \rho(r) \text{ le}\text{\AA}^{-5}$	$H(r)$
Rh – P2	0.124	0.066	-0.065
Rh – P3	0.123	0.068	-0.063
Rh – H34	0.056	0.247	-0.004
Rh – H37	0.060	0.241	-0.009
C4 – H34	0.236	-0.643	-0.209
C4 – H35	0.271	-0.911	-0.272
C4 – H36	0.277	-0.953	-0.280
C5 – H37	0.232	-0.601	-0.200

Table S9: Key BCP metrics for the 1st cation of **[1-C₄H₁₀][BAR^F₄]**.

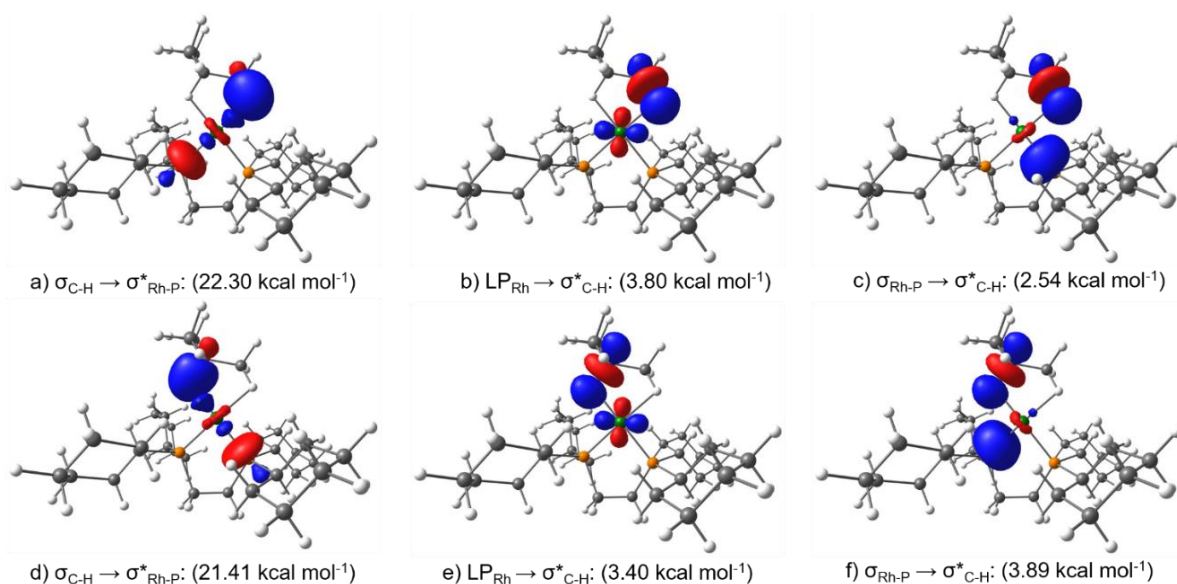


Figure S113: NBO donor-acceptor pairs for the 1st cation of **[1-C₄H₁₀][BAR^F₄]** with NBO occupancies as indicated.

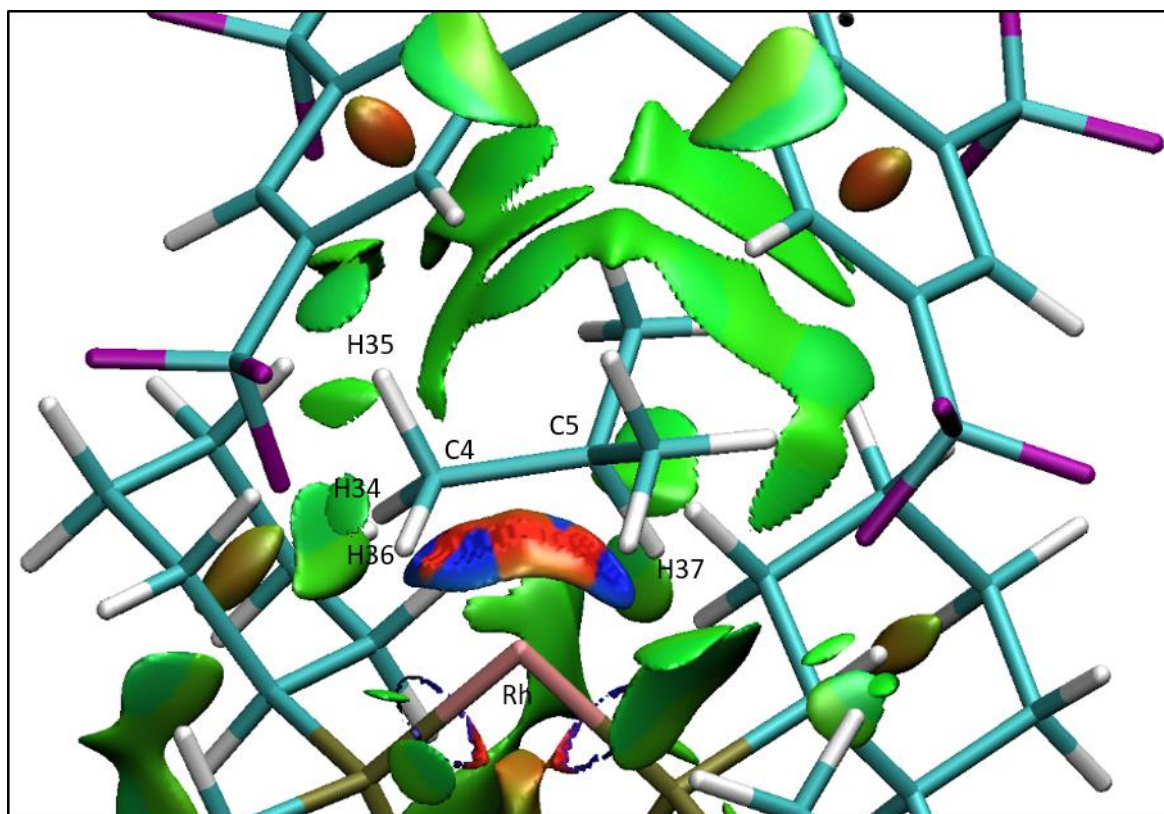


Figure S114: NCI plot of the 1st cation of **[1-C₄H₁₀][BAR^F₄]** ion-pair. Isosurfaces generated for $s = 0.3$ au and $-0.07 < \rho < 0.07$ au.

Distance (Å)	Fully Optimized	Heavy Atom Fixed ^a	Experiment
Rh – P2	2.203	2.184	2.184(1)
Rh – P3	2.206	2.189	2.189(1)
Rh – H34	1.934	1.955	1.981
Rh – H37	1.885	1.868	1.798
Rh – C4	2.383	2.390	2.390(14)
Rh – C5	2.480	2.438	2.438(8)
C4 – H34	1.147	1.145	1.098
C4 – H35	1.100	1.099	1.098
C4 – H36	1.096	1.094	1.098
C5 – H37	1.160	1.159	1.001
Bite Angle (°)	87.283	89.324	76.70

Table S10: Key bond distances (Å) for the **2nd cation** of **[1-C₄H₁₀][BAr^F₄]** from fully optimised and heavy atoms fixed geometries of the **1st cation** of **[1-C₄H₁₀][BAr^F₄]**, with experimental data for comparison.^{S31}

^astructure is based on the experimental structure with only the positions of H and F atoms being optimised.

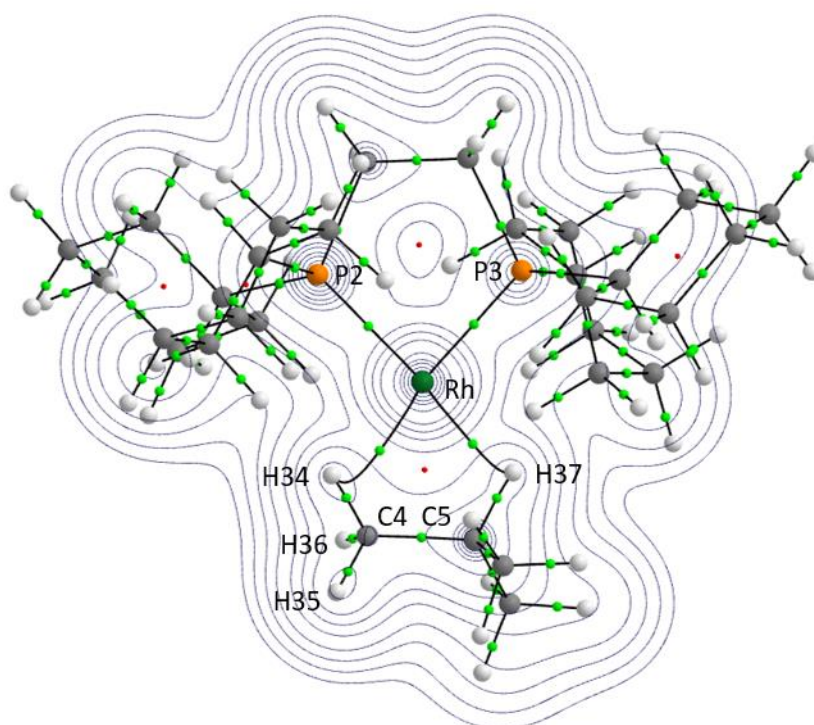


Figure S115: QTAIM molecular graph for the **2nd cation** of **[1-C₄H₁₀][BAr^F₄]**, with density contours plotted in the Rh-H34-H37 plane.^{S30} Bond critical points (BCPs) are indicated in green and ring critical points (RCPs) are indicated in red.

Bond	$\rho(r) / e\text{\AA}^{-3}$	$\nabla^2 \rho(r) / e\text{\AA}^{-5}$	$H(r)$
Rh – P2	0.124	0.070	-0.064
Rh – P3	0.124	0.061	-0.064
Rh – H34	0.053	0.235	-0.003
Rh – H37	0.061	0.245	-0.009
C4 – H34	0.239	-0.659	-0.213
C4 – H35	0.271	-0.919	-0.272
C4 – H36	0.277	-0.954	-0.281
C5 – H37	0.232	-0.608	-0.201

Table S11: Key BCP metrics for the 2nd cation of [1-C₄H₁₀][BAR^F₄].

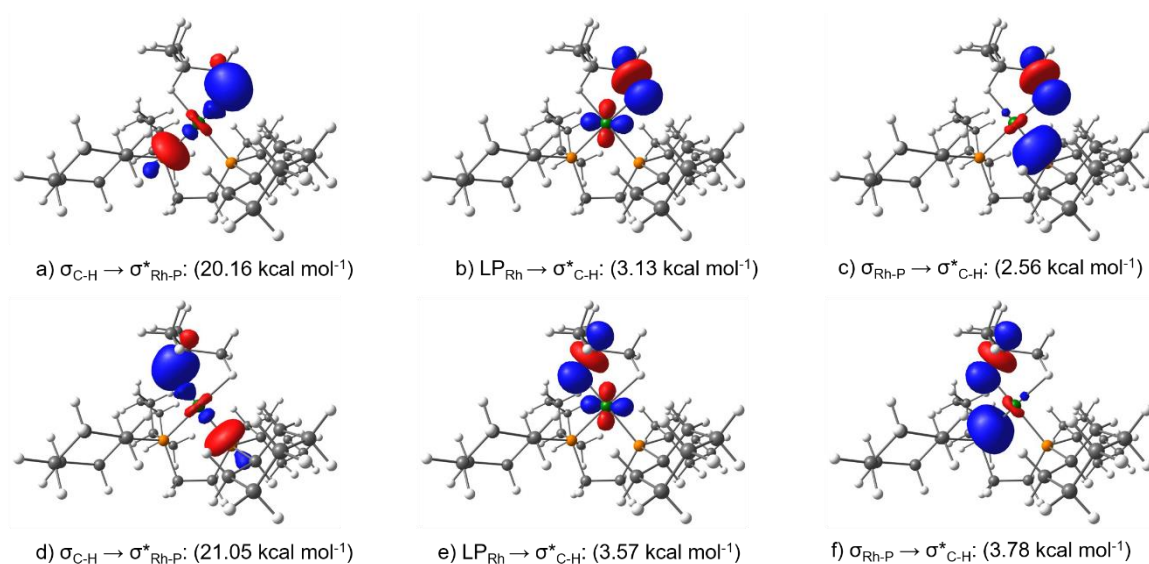


Figure S116: NBO donor-acceptor pairs for the 2nd cation of [1-C₄H₁₀][BAR^F₄] with NBO occupancies as indicated.

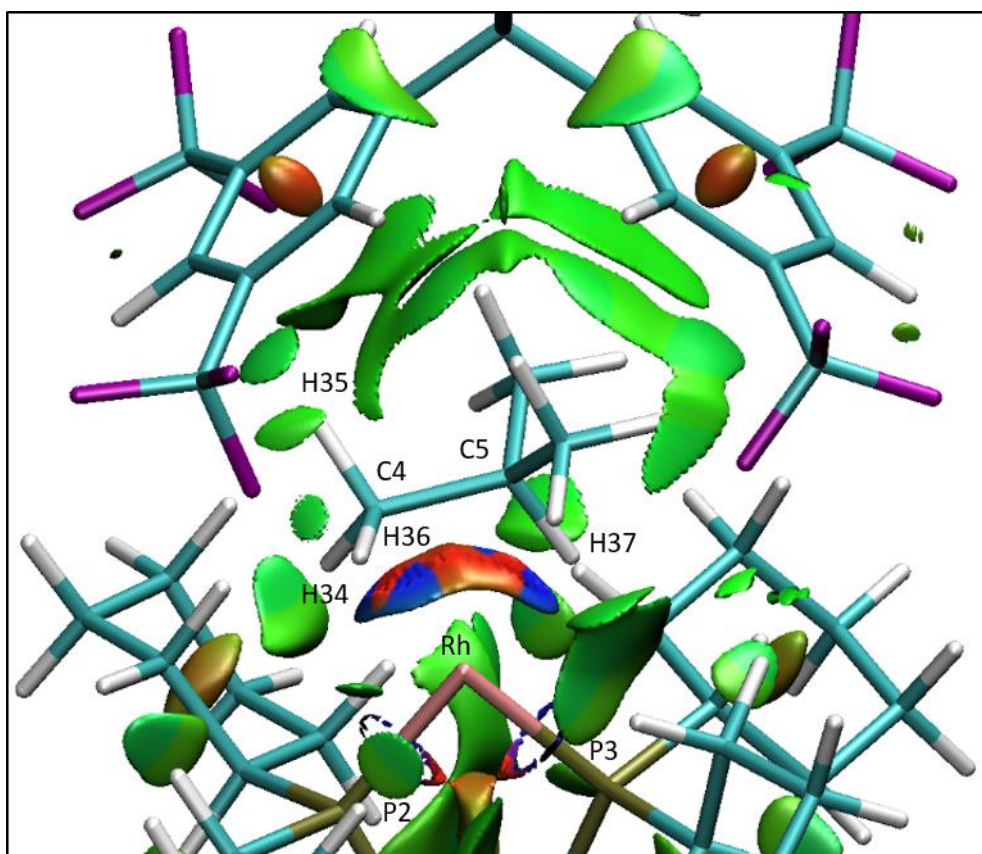


Figure S117: NCI plot for the 2nd cation of [1-C₄H₁₀][BARF₄] ion-pair. Isosurfaces generated for $s = 0.3$ au and $-0.07 < \rho < 0.07$ au.

S.9.2. [1-C₆H₁₂][BAR^F₄]

Distance (Å)	Fully Optimised	Heavy Atom Fixed ^a	Experiment
Rh – P1	2.201	2.192	2.192(2)
Rh – P2	2.195	2.192	2.192(2)
Rh – C1	2.679	2.530	2.530(2)
Rh – C3	2.681	2.642	2.642(2)
Rh – H11	1.928	1.852	1.777
Rh – H31	1.923	1.933	1.931
Rh – H51	2.677	2.668	2.721
C1 – H11	1.140	1.141	0.970
C1 – H12	1.101	1.100	0.970
C3 – H31	1.139	1.137	0.970
C3 – H32	1.100	1.099	0.971
C5 – H44	1.109	1.109	0.970
C5 – H45	1.098	1.094	0.970

Table S12: Key bond distances (Å) for fully optimised and heavy atoms fixed geometries, compared with experimental data.^{S31}

^a struture is based on the experimental structure with only the positions of H and F atoms being optimised.

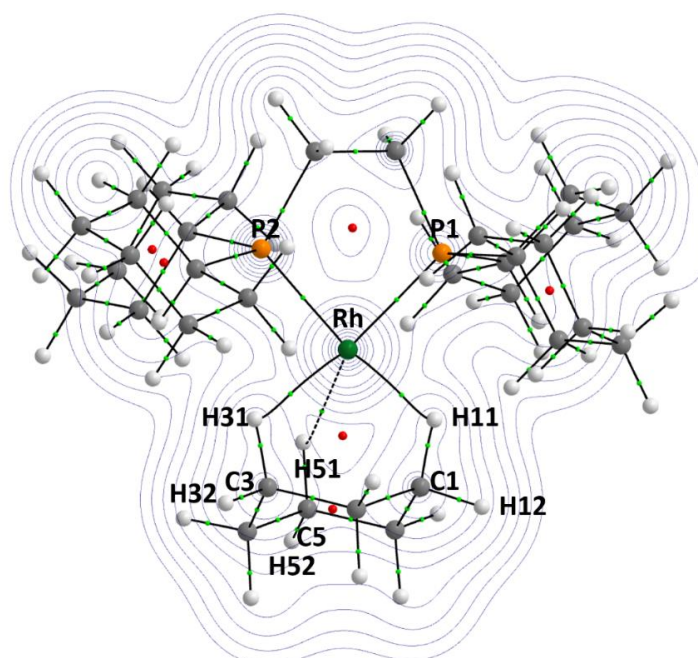


Figure S118: QTAIM molecular graph with density contours plotted in the Rh-H11-H31 plane. Bond critical points (BCPs) are indicated in green and ring critical points (RCPs) are indicated in red.

Bond	$\rho(r) / e\text{\AA}^{-3}$	$\nabla^2 \rho(r) / e\text{\AA}^{-5}$	H(r)
Rh – P1	0.123	0.065	-0.063
Rh – P2	0.123	0.070	-0.063
Rh – H11	0.061	0.226	-0.011
Rh – H31	0.050	0.181	-0.008
Rh – H51	0.015	0.041	0.000
C1 – H11	0.239	-0.648	-0.214
C1 – H12	0.274	-0.920	-0.274
C3 – H31	0.242	-0.671	-0.219
C3 – H32	0.274	-0.922	-0.274
C5 – H51	0.266	-0.853	-0.259
C5 – H52	0.276	-0.932	-0.279

Table S13. Key BCP metrics of **[1-C₆H₁₂][BAr^F₄]**

Donor	Acceptor	E(2) (kcal / mol)
Rh LP	σ^* C1 – H11	4.26
σ Rh – P1	σ^* C1 – H11	5.99
σ Rh – P2	σ^* C1 – H11	1.63
σ C1 – H11	σ^* Rh – P2	20.03
Rh LP	σ^* C3 – H31	2.60
σ Rh – P1	σ^* C3 – H31	1.76
σ Rh – P2	σ^* C3 – H31	5.45
σ C3 – H31	σ^* Rh – P1	15.13

Table S14: Key donor-acceptor interactions from the NBO second order perturbation analysis of **[1-C₆H₁₂][BAr^F₄]**.

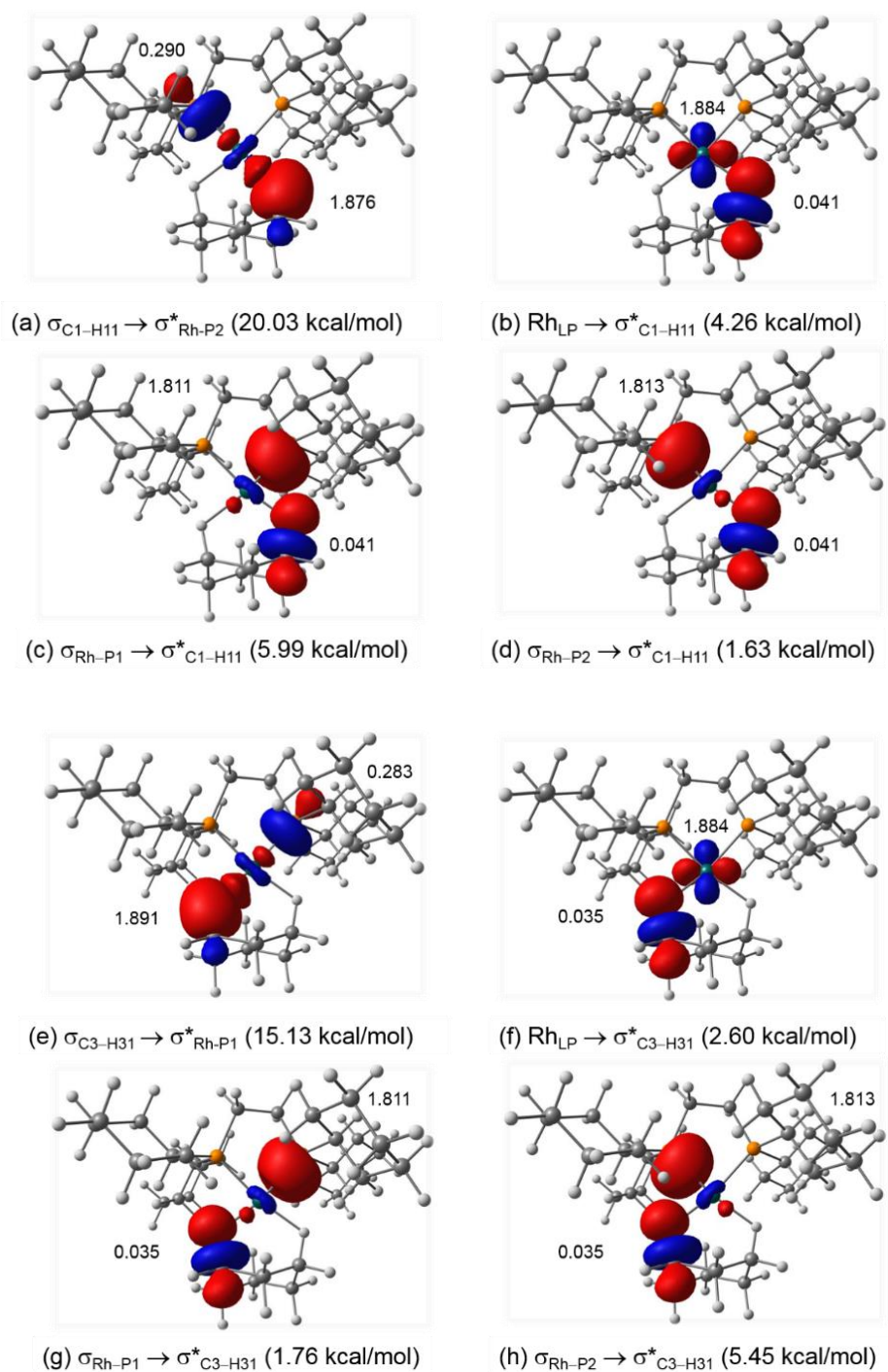


Figure S119: NBO donor-acceptor pairs for $[1-C_6H_{12}][BARF_4]$ with NBO occupancies as indicated.

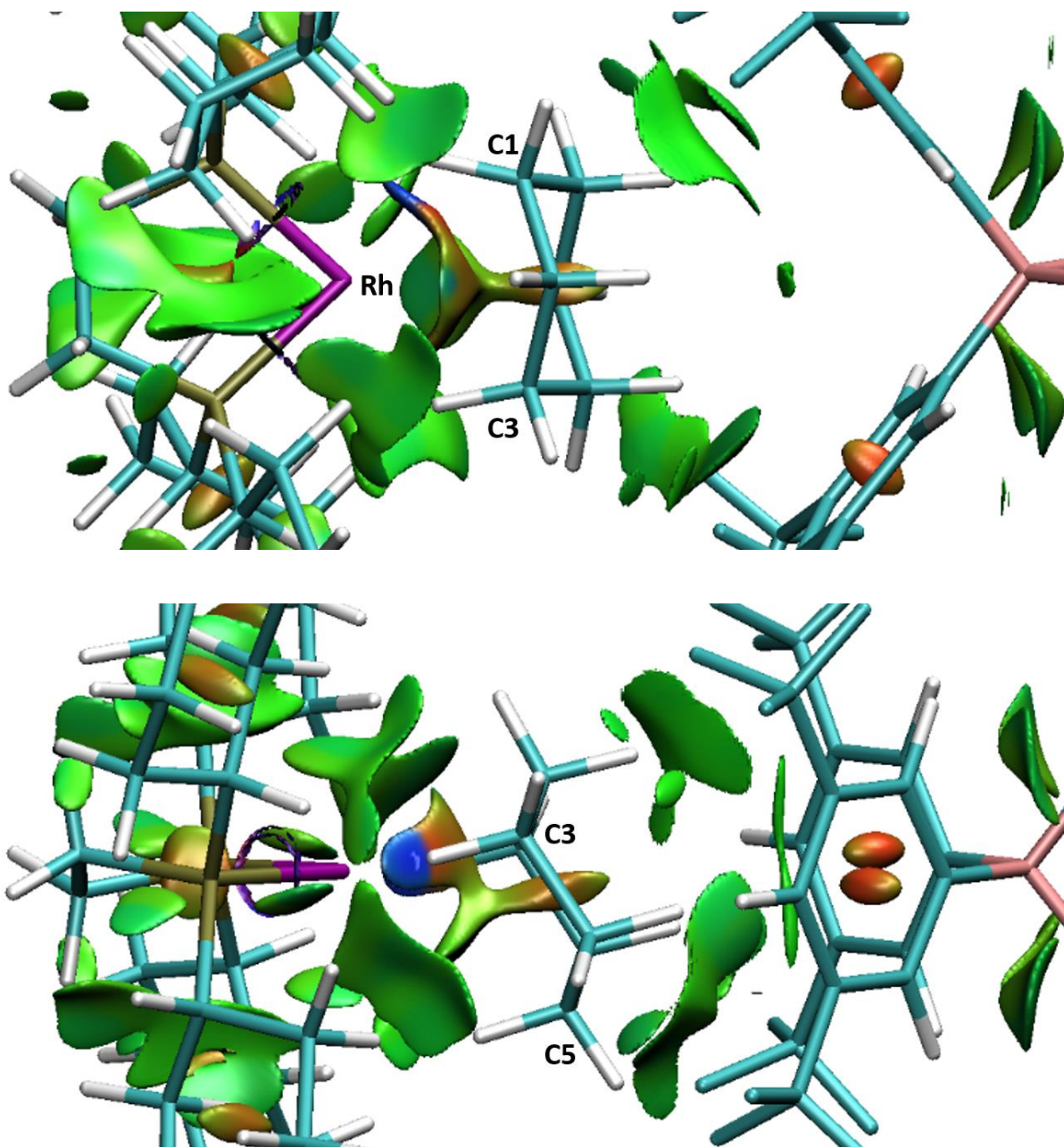


Figure S120: Two views of the NCI plots of the [1-C₆H₁₂][BARF₄] ion-pair. Isosurfaces generated for $s = 0.3$ au and $-0.07 < \rho$ Isosurface

S.10. References

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