

# PNAS

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Supplementary Information for

## **Intestinal microbes influence development of thymic lymphocytes in early life**

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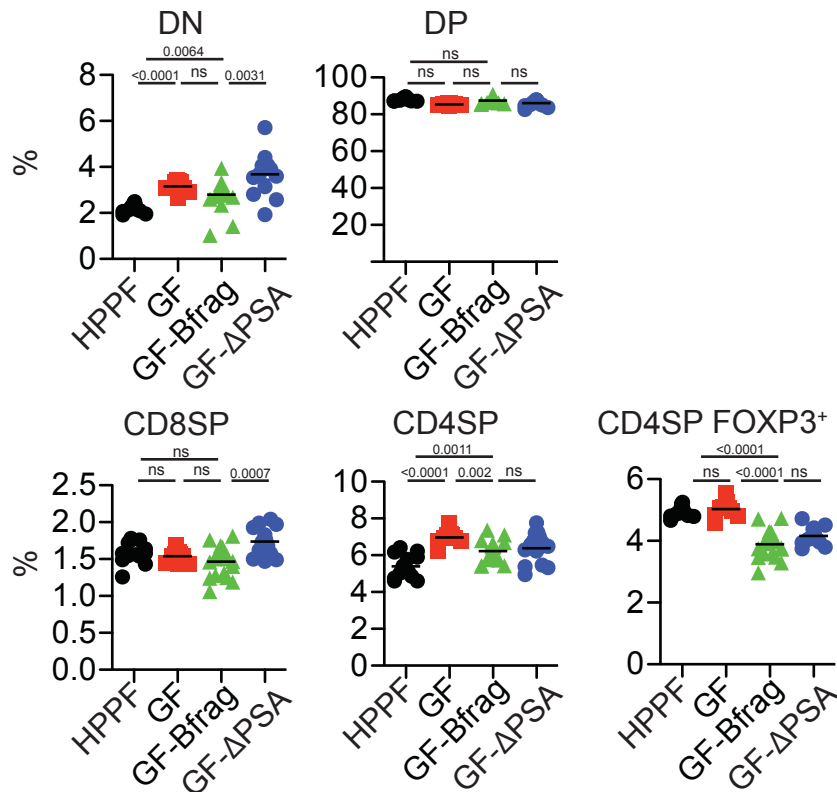
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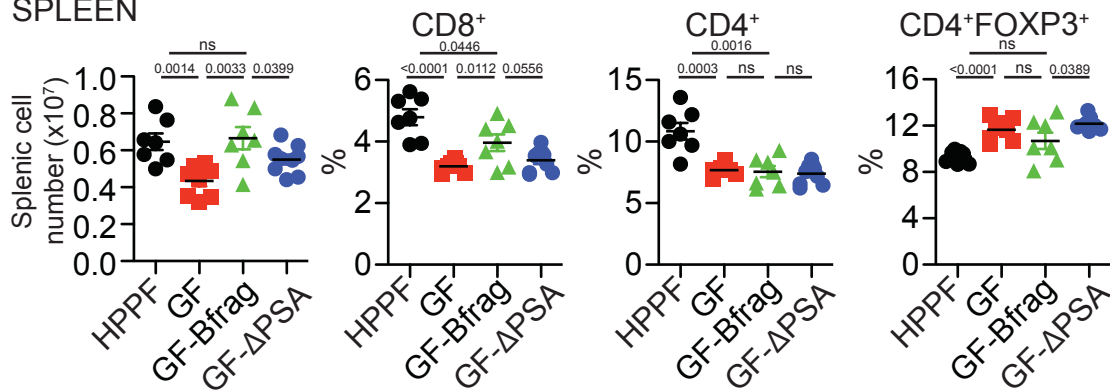
### **This PDF file includes:**

Supplementary text  
Figures S1 to S14

## A THYMUS



## B SPLEEN

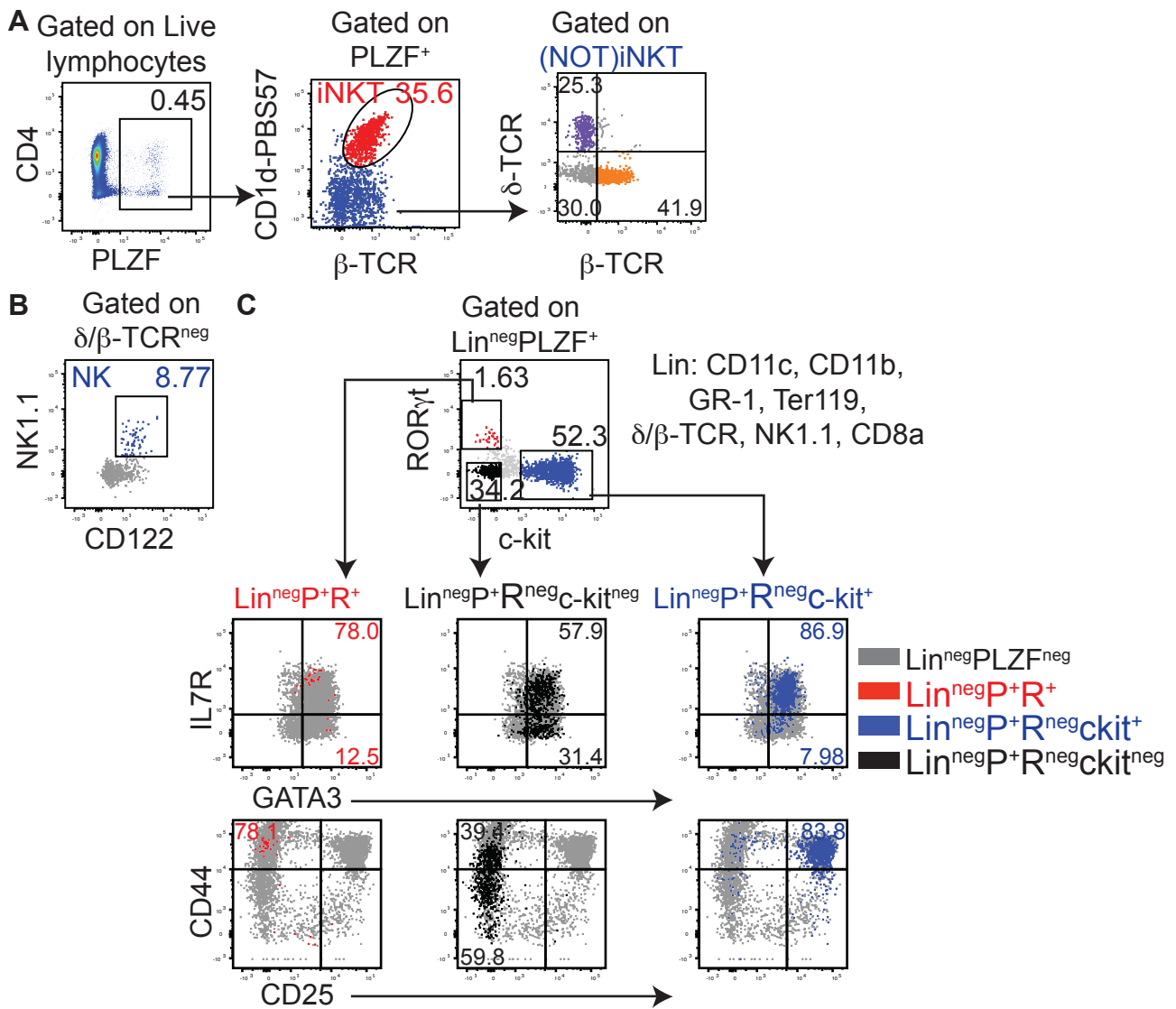


**Figure S1: Distribution of major T cell subsets in thymus and spleen of HPPF, GF, GF-Bfrag and GF- $\Delta$ PSA pups.**

**A.** Frequency of thymic DN (CD4<sup>neg</sup>CD8a<sup>neg</sup>), DP (CD4<sup>+</sup>CD8a<sup>+</sup>), CD8SP (CD4<sup>neg</sup>CD8<sup>+</sup>), CD4SP (CD4<sup>+</sup>CD8a<sup>neg</sup>) and CD4SP FOXP3<sup>+</sup> cells (gated on CD4SP) (HPPF n= 12; GF n=8; GF-Bfrag n=18; GF- $\Delta$ PSA n=14).

**B.** Total splenic cellularity and frequency of CD8<sup>+</sup>, CD4<sup>+</sup> (gated on Live) and CD4<sup>+</sup>FOXP3<sup>+</sup> cells (gated on CD4<sup>+</sup> T cells) (HPPF n= 7; GF n=8; GF-Bfrag n=7; GF- $\Delta$ PSA n=9).

Data are from 2-3 independent experiments for each group. Bars are Mean  $\pm$  SEM.



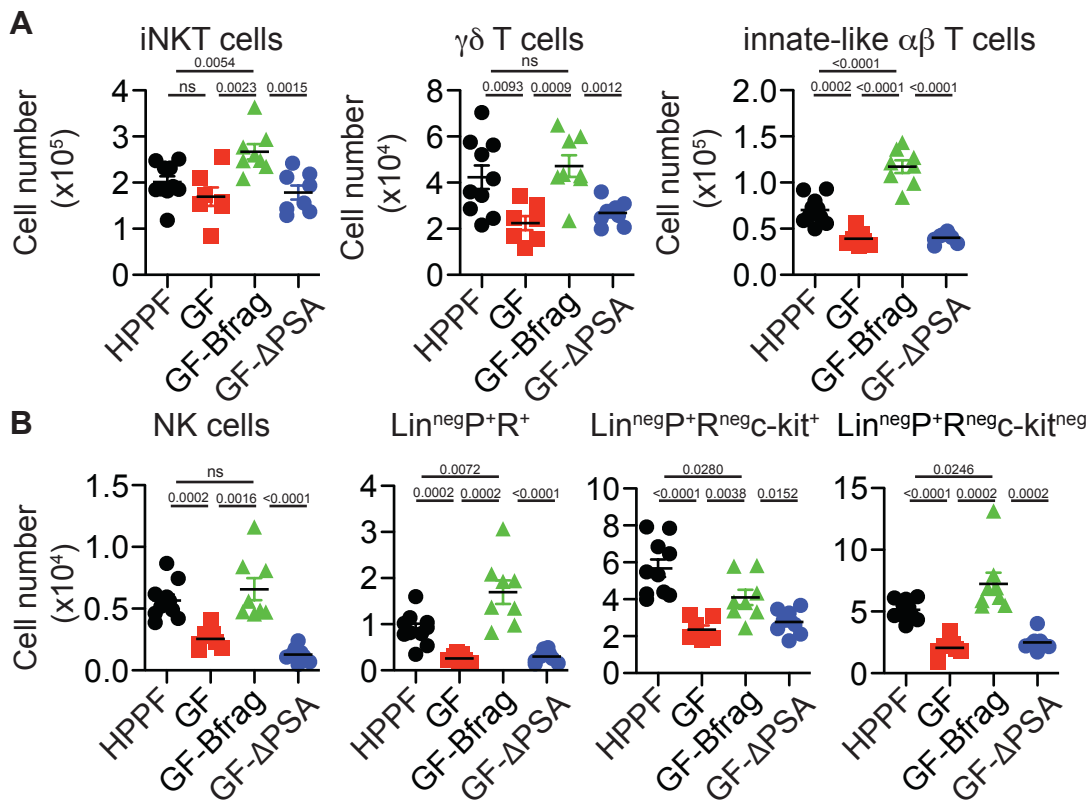
**Figure S2: Identification of PLZF expressing cell subsets by flow cytometry**

Gating strategy to identify PLZF expressing cell subsets in the thymus of 14 days old C57BL/6 pups. Live and single lymphocytes were analyzed as follows:

**A.** (Left) Representative flow cytometry plot showing expression of PLZF (y-axis) and CD4 (x-axis) to identify total PLZF<sup>+</sup> cells. (Middle) PLZF<sup>+</sup> cells were next analyzed for β-TCR (x-axis) and mCD1d-PBS57 tetramer (y-axis) expression to identify mCD1d-PBS57<sup>+</sup>TCRβ<sup>+</sup> iNKT cells. (Right) A (NOT)iNKT gate was used to determine expression of β-TCR (x-axis) and δ-TCR (y-axis) on mCD1d-PBS57<sup>neg</sup> cells to identify PLZF<sup>+</sup>γδ T cells and PLZF<sup>+</sup> innate-like αβ-T cells.

**B.** PLZF<sup>+</sup> cells were gated on β/δ-TCR<sup>neg</sup> cells and analyzed for expression of CD122 (x-axis) and NK1.1 (y-axis) to identify NK cells.

**C.** TCR<sup>neg</sup>NK1.1<sup>neg</sup>PLZF<sup>+</sup> cells that were also Lin<sup>neg</sup> (Lin: CD11c, CD11b, Ter119, CD19, GR-1 and CD8a) were analyzed for expression of c-kit (x-axis) and RORγt (y-axis). Cells identified as Lin<sup>neg</sup>PLZF<sup>+</sup>RORγt<sup>+</sup> (Lin<sup>neg</sup>P<sup>+</sup>R<sup>+</sup> in red), Lin<sup>neg</sup>PLZF<sup>+</sup>c-kit<sup>+</sup> (Lin<sup>neg</sup>P<sup>+</sup>c-kit<sup>+</sup> in blue) and Lin<sup>neg</sup>PLZF<sup>+</sup>c-kit<sup>neg</sup> (Lin<sup>neg</sup>P<sup>+</sup>c-kit<sup>neg</sup> in black) cells were further analyzed for expression of (top row) GATA3 (x-axis) and IL7R (y-axis) and (bottom row) CD25 (x-axis) and CD44 (y-axis).



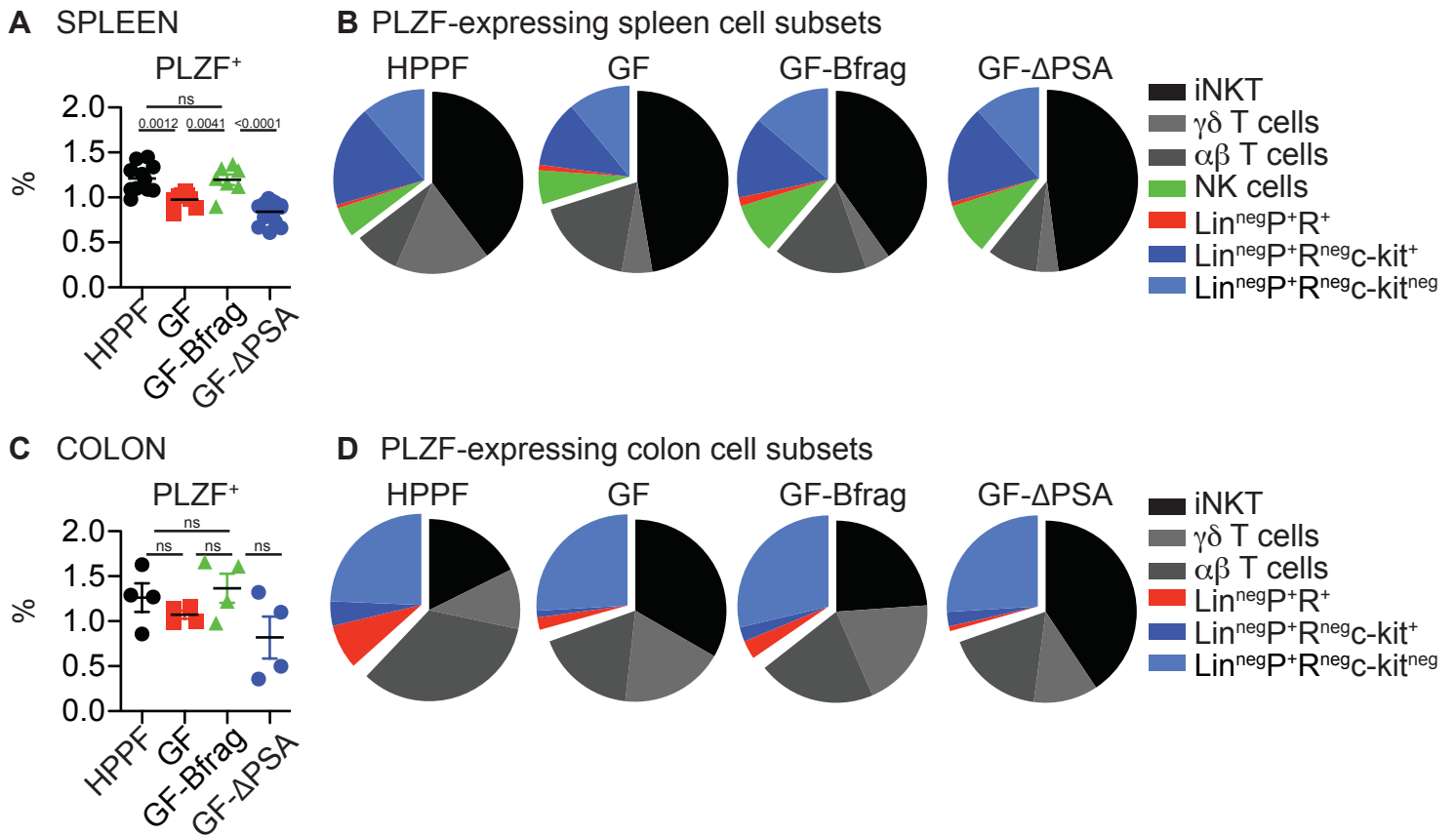
**Figure S3: Cell numbers of PLZF-expressing subsets in thymus of HPPF, GF, GF-Bfrag and GF- $\Delta$ PSA pups.**

**A.** Total numbers of PLZF<sup>+</sup> iNKT cells, PLZF<sup>+</sup>  $\gamma\delta$  T cells and PLZF<sup>+</sup> innate-like  $\alpha\beta$ -T cells (HPPF n= 10; GF n=7; GF-Bfrag n=8; GF- $\Delta$ PSA n=8).

**B.** Total numbers of PLZF<sup>+</sup> NK cells,  $\text{Lin}^{\text{neg}}\text{PLZF}^+\text{ROR}\gamma\text{t}^+$  cells,  $\text{Lin}^{\text{neg}}\text{PLZF}^+\text{ROR}\gamma\text{t}^{\text{neg}}\text{c-kit}^+$  cells and  $\text{Lin}^{\text{neg}}\text{PLZF}^+\text{ROR}\gamma\text{t}^{\text{neg}}\text{c-kit}^{\text{neg}}$  cells (HPPF n= 10; GF n=7; GF-Bfrag n=8; GF- $\Delta$ PSA n=8).

Data are representative of 2-3 experiments for each group. Bars are Mean  $\pm$  SEM.





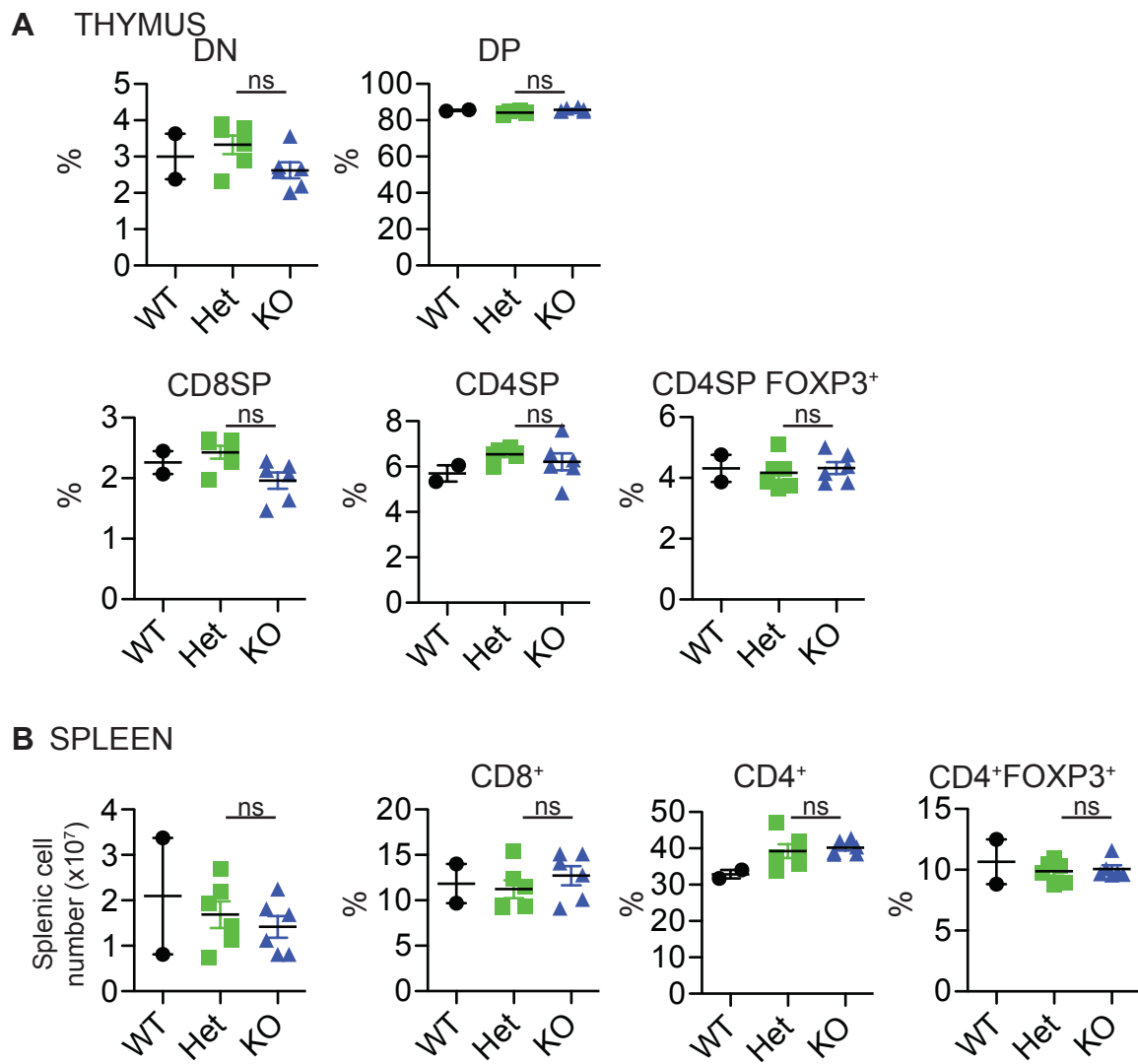
**Figure S4: Frequency of PLZF<sup>+</sup> cells in spleen and colon of HPPF, GF, GF-Bfrag and GF-ΔPSA pups.**

**A.** Frequency of PLZF<sup>+</sup> cells in spleen (HPPF n= 11; GF n=8; GF-Bfrag n=7; GF-ΔPSA n=17). Data are from of 2-3 experiments in each group.

**B.** Pie graphs showing distribution of indicated PLZF expressing cell subsets in spleen.

**C.** Frequency of PLZF<sup>+</sup> cells in colon (HPPF n= 4; GF n=4; GF-Bfrag n=4; GF-ΔPSA n=4). Data are representative of 2 independent experiments.

**D.** Pie graphs showing distribution of indicated PLZF expressing cell subsets in colon. Bars are Mean  $\pm$  SEM.

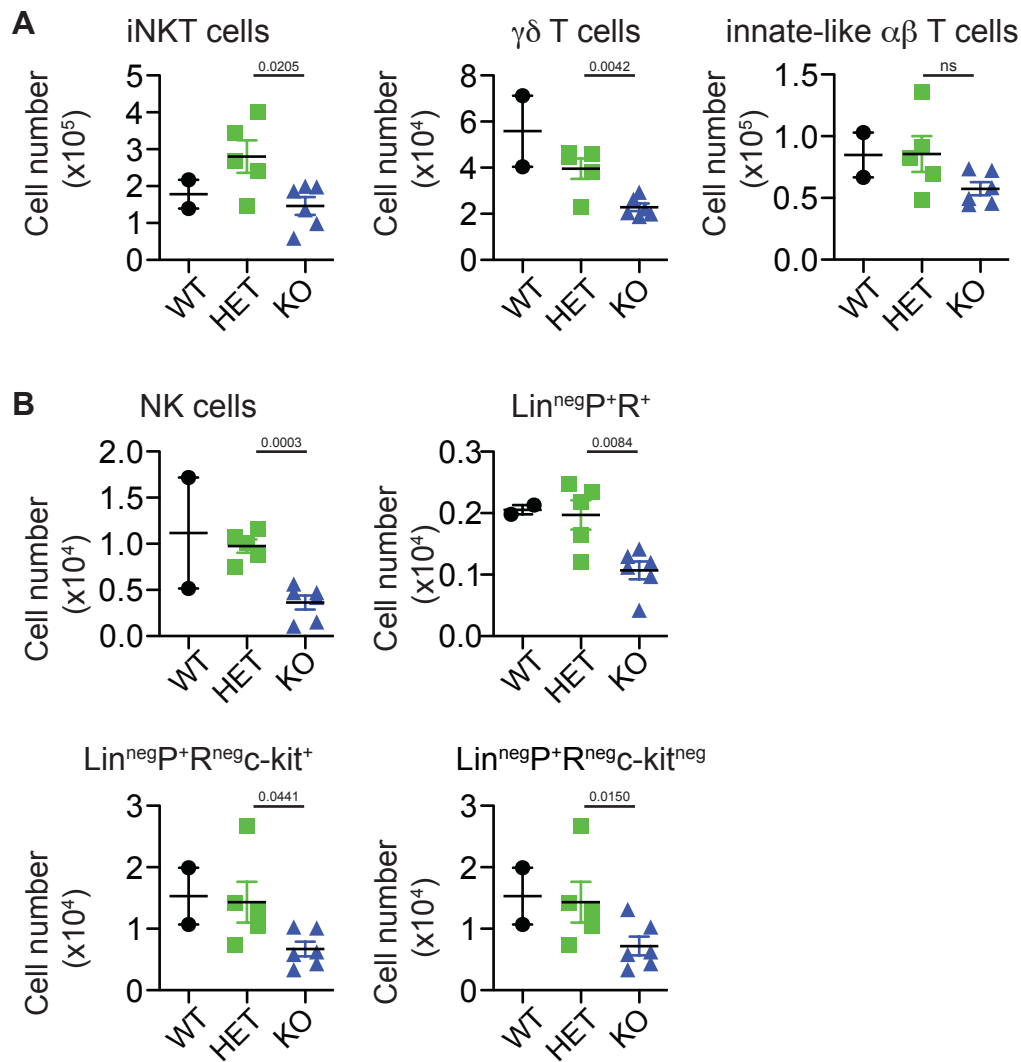


**Figure S5: Distribution of major T cell subsets in thymus and spleen of infant *Tlr2*<sup>-/-</sup> mice**

**A.** Frequency of thymic DN, DP, CD8SP, CD4SP and CD4SP FOXP3<sup>+</sup> cells (WT n= 2; HET n=6; KO n=6).

**B.** Total splenic cellularity and frequency of CD8<sup>+</sup>, CD4<sup>+</sup> and CD4<sup>+</sup>FOXP3<sup>+</sup> cells (WT n= 2; HET n=6; KO n=6).

Data are from 2 experiments. Bars are Mean  $\pm$  SEM.

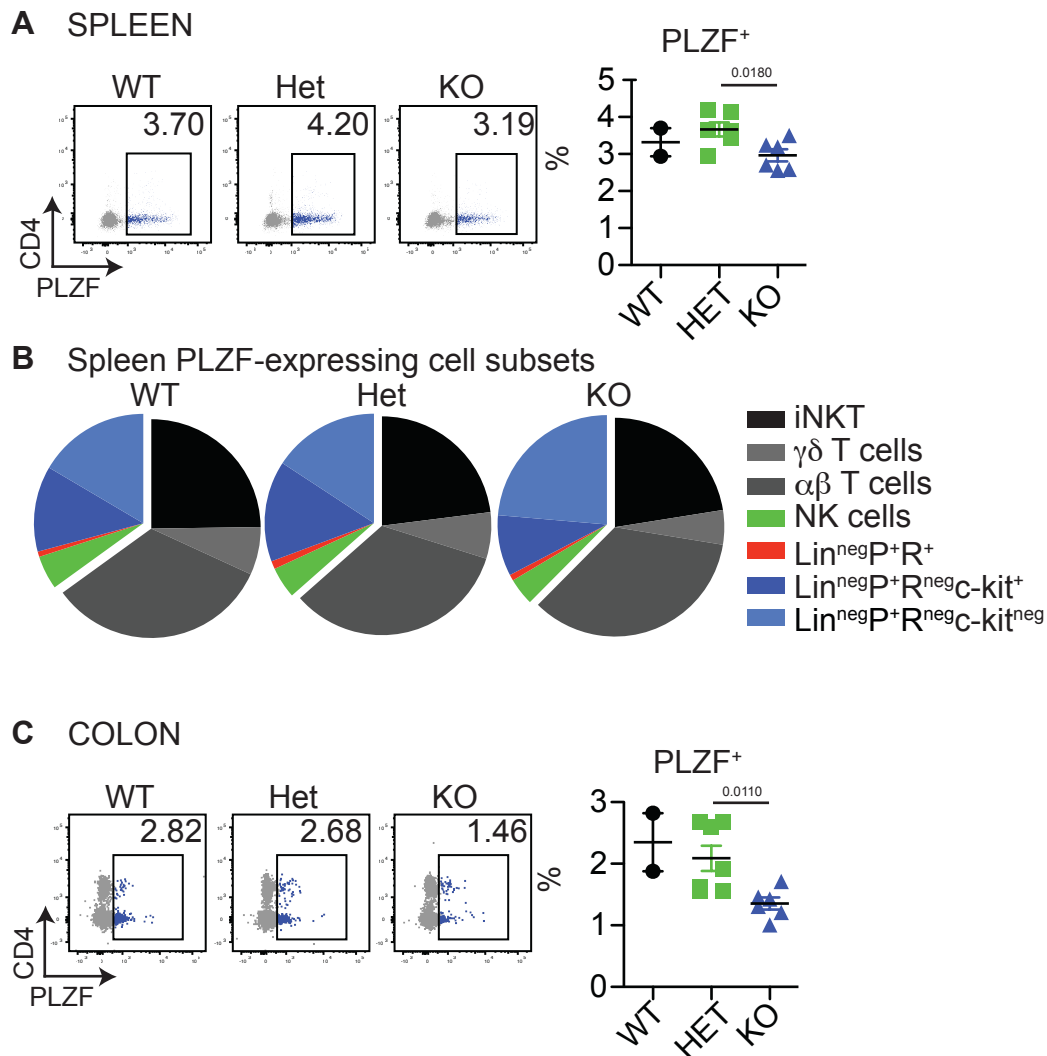


**Figure S6: Cell numbers of PLZF-expressing cell subsets in thymus of infant *Tlr2*<sup>-/-</sup> mice.**

**A.** Total numbers of PLZF<sup>+</sup> iNKT cells, PLZF<sup>+</sup>  $\gamma\delta$  T cells and PLZF<sup>+</sup> innate-like  $\alpha\beta$ -T cells (WT n= 2; HET n=5; KO n=6).

**B.** Total numbers of PLZF<sup>+</sup> NK cells,  $\text{Lin}^{\text{neg}}\text{PLZF}^+\text{ROR}\gamma\text{t}^+$  cells,  $\text{Lin}^{\text{neg}}\text{PLZF}^+\text{ROR}\gamma\text{t}^{\text{neg}}\text{c-kit}^+$  cells and  $\text{Lin}^{\text{neg}}\text{PLZF}^+\text{ROR}\gamma\text{t}^{\text{neg}}\text{c-kit}^{\text{neg}}$  cells (WT n= 2; HET n=5; KO n=6).

Data are from 2 experiments. Bars are Mean  $\pm$  SEM.



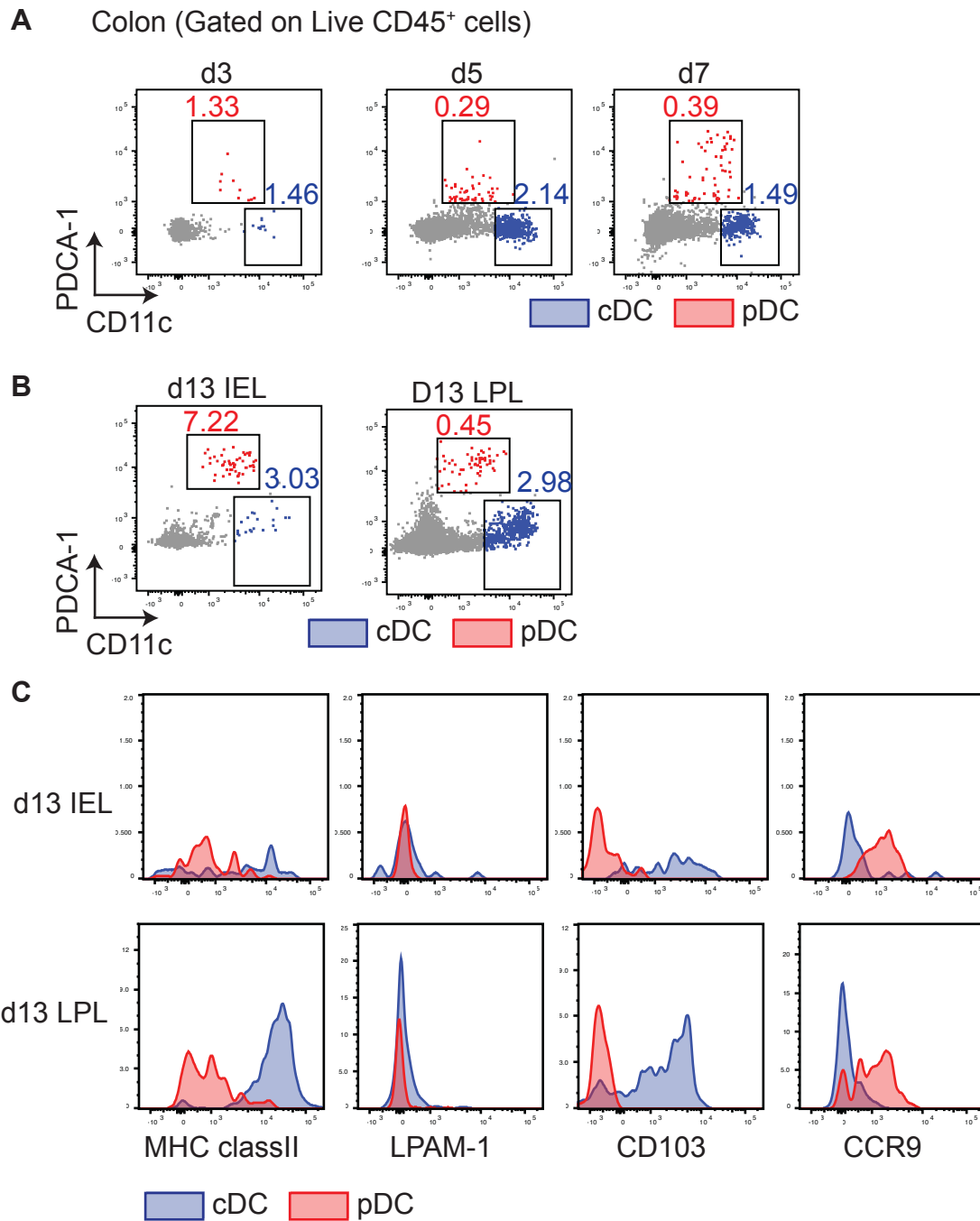
**Figure S7: Frequency of PLZF expressing subsets in spleen and colon of infant *Tlr2*<sup>-/-</sup> mice.**

**A.** Frequency of PLZF<sup>+</sup> cells in spleen (WT n= 2; HET n=6; KO n=6).

**B.** Pie graphs showing distribution of indicated PLZF expressing cell subsets in spleen.

**C.** Frequency of PLZF<sup>+</sup> cells in colon (WT n= 2; HET n=5; KO n=6).

Data in A-C are from 2 independent experiments. Bars are Mean  $\pm$  SEM.



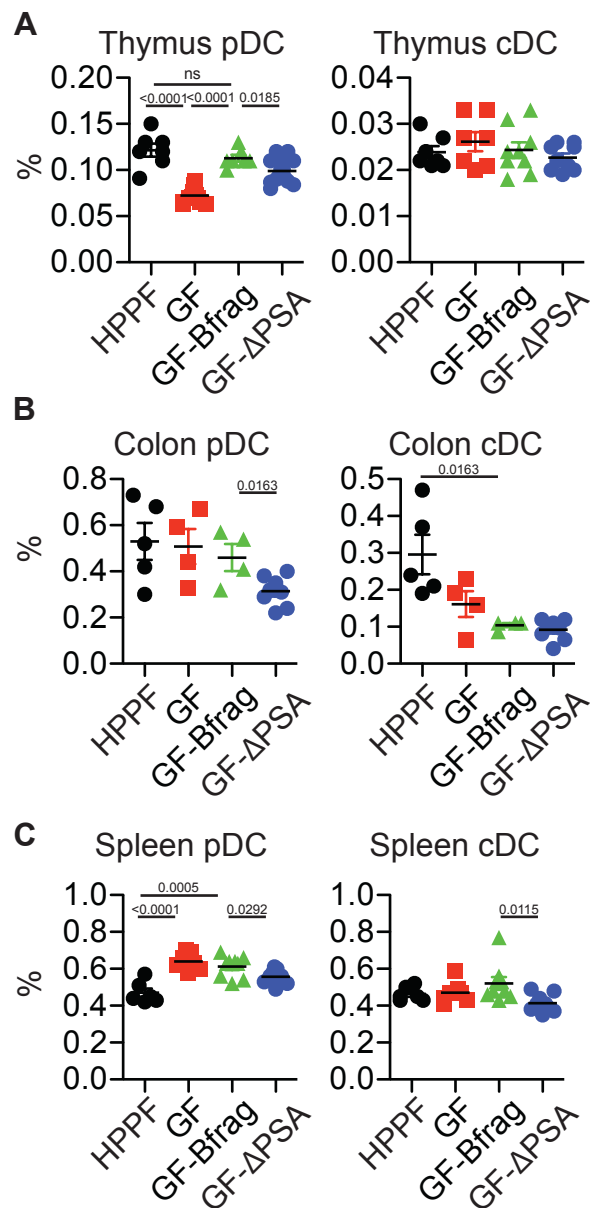
**Figure S8: Colonic pDCs in conventionally-housed infant C57BL/6 mice.**

**A.** Representative flow cytometry dot plots showing expression of CD11c (x-axis) and PDCA-1 (y-axis) in colonic lymphocyte preparations from 3, 5 and 7 days old C57BL/6 pups. Frequency of pDCs (CD11<sup>+</sup>PDCA-1<sup>+</sup>) and cDCs (CD11c<sup>hi</sup>PDCA-1<sup>neg</sup>) is shown.

**B.** Representative flow cytometry dot plots showing expression of CD11c (x-axis) and PDCA-1 (y-axis) in colonic intra-epithelial lymphocyte (IEL) and lamina propria lymphocyte (LPL) preparations from 13 days old C57BL/6 pups. Frequency of pDCs and cDCs is shown.

**C.** Histogram overlays of cDCs (blue) and pDCs (red) in IEL and LPL fractions showing expression of MHC class II, LPAM-1, CD103 and CCR9.

Data are representative of 2 independent experiments.



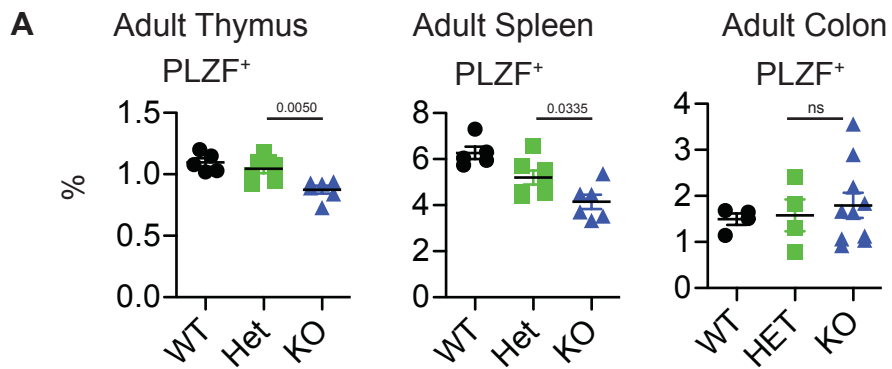
**Figure S9: Distribution of pDCs and cDCs in colon, spleen and thymus of d14 GF and monocolonized mice**

**A-C.** Frequency of pDCs and cDCs in the thymus (HPPF  $n=7$ ; GF  $n=12$ ; GF-Bfrag  $n=7$ ; GF-ΔPSA  $n=15$ ), colon (HPPF  $n=5$ ; GF  $n=4$ ; GF-Bfrag  $n=4$ ; GF-ΔPSA  $n=8$ ) and spleen (HPPF  $n=6$ ; GF  $n=7$ ; GF-Bfrag  $n=9$ ; GF-ΔPSA  $n=9$ ) is shown.

Data are representative of at least 2 independent experiments. Bars are Mean  $\pm$  SEM.

Figure S10: RNA-seq analysis of thymic pDCs from <i>B. fragilis</i> (GF-Bfrag) or PSA mutant <i>B. fragilis</i> (GF-ΔPSA) monoclonalized GF mice.															
	a1	a2	a3	a4	a5	a6	a7	a8	a9	a10	a11	a12	a13	a14	a15
1	Gm37347	Fam102a	Cyb5r1	Gpr174	Vcam1	Ccr7	C3	Cmah	Xpnp3	Smco4	Zfp324			Xist	Spata2l
2	Gm13919	Ormdl1	Stxbp5	Mir378b	Zfp46	Gm14636	Csf1r	Gm38014	Sic28a2	Cd151				Pla2g2d	Rpp25f
3	Emid1	A530099J19Rik	H2-Ob	Tmem2	Rpain	Cas21	Arhgef2	Pou2af1		Dnajc11				Oprd1	Stard4
4	Gm8369	Fasikd5	Gm16982	Bend4	Nod1	Zc2hc1a	Sepp1	RP24-351O18.4		Tnfrsf21				Tnfrsf11a	Gm16337
5	Gm15903	Mcm8	Ndrp1	Gm13528	Meaf6	Timm22	Col27a1	Gm16310		Wdfy3				Vdr41	Ydjc
6	Gm38387	RP23-21416.2	Ppap2a	Sic30a1	Nav1		Psen2	Sic43a2		Rab31				Taf5l	Il18
7	Sic36a3	Acot6	Aftph	Igkc	Atxn1		Xdh	Gm11131		Vps37c				Adam22	Oip5
8	Gdpp1		E130311K13Rik		Ccdc71		Phf11c	Pkd1l3		Fmn1				RP23-231L15.4	RP23-132D7.2
9	Gm24452		Fasn		Tiam1		Igsf3	B230317F23Rik		Ms4a6b				Clec4n	Adora2a
10	A630072L19Rik		Aire		Papp2		Rps13-ps1	Zbtb11os1		Gpnmb				Rnf145	Pla2g12a
11	Gf3c5		Zak		Cdc3		Plekhh2	Exd2		Cistn1				Sic12a2	Cd99l2
12			Ptpn22		Arb1		Plekhn1	Tmem140		Il18bp				Tin	Gm14730
13			Pygl		Anxa4		Bcor	Ppp1r9a		Rtp4				Nid1	
14			Bsdcl1		Lrp5		Siglec1	RP23-158A20.7		Clec7a				Fam26f	
15			Itprnl2		Chst2		Itga9	Ccdc93		Med4				Rbbp8	
16			Matr3		Zdhhc24		Egr1	2210016L21Rik		Adck5				Oas2	
17			Tmem236		Gfra2		Ppm1g	Sic16a5		Bcl9				Tmprss6	
18			Rras2		Lpl		Hist1h2bl	1700001D01Rik		Gaint10				Anxa3	
19			F2r2		Neur1b		Taf11	Gm17146		Ostm1				Igf2r	
20			Gm4876		Polr3e		Iah1	Ldoc1l		Spg21				Adamdec1	
21			Lcorl		Scamp1		Furin	Zfp948		Rnf144b				Fgl2	
22			Klf8		Gca		Ar4c	RP23-454A14.10		Agm				Xcr1	
23			Gm34727		Nrip1b		Myo6	Wbscr16		Top3a				Rab31f	
24			Tmem164		Gla		Kenn4	Ccdc88c		Olfir433				Mex3b	
25			Disc1		Mcm10		Abcc5	Elmod3		Pame3				Sic37a3	
26			Gm37699		Hnmp1		Gm14446	Cbr2		Asah2				Epb4.1i3	
27			Adap1		Sic7a8		Cyp4v3	Atxn7		Pex5				Brap	
28			Taf4b		Ddx60		Strp2	9230114K14Rik		Lyz2				Sxbp1	
29			Nccrp1		Hif1an		Cdc20	Gm14212		Ptms				Anpep	
30			Rarg		Igfa6		Tbxas1	Malsu1		Tmem26				Ccl12	
31			Pet100		2900026A02Rik		Ptgs1	Msh5		Selo				Dusp16	
32			Phf19		Tut1		Gins2	Ifo1						Il6st	
33			Apol7c		Focad		Ifi204	Bsn						Ifnlr1	
34			Zfp277		Ttc39b		Osbpl3	Vsig10						Arhgef40	
35			Ptpn11		Pid1		Gm28933	Pias2						Ccdc91	
36			1700056E22Rik		Mllt6		RP23-2C16.2	Fsd1l						Dusp19	
37			Sic25a30				Tyh3	Gm38147						9330175E14Rik	
38			Pabpc4				Zfp949	Rabl2						Gm37570	
39			F830112A20Rik				Mogs	L2hgdh						RP24-176P19.6	
40			H2-Dmb2				Rplf2	I7Rn6						Scaf1	
41			Mboat1				Ddx49	Strbp						Asb2	
42			Tgfr2				C1qc	Prdm11						Nab2	
43							Mtf1	Gm22639						9130221H12Rik	
44							Cisd1	Gm37368						Zfp366	
45							Kctd6	Adam33						Acp5	
46							Phtf2	Bcl7a						Sic36a1	
47							Zfp54	Rbx2						Trm61a	
48							A730011C13Rik	Gm37873						Utp14b	
49							Al846148	Ubxn11						Sigmar1	
50							Clec4a1	Zfp524						Zfp180	
51							A430033K04Rik	Sic25a23						Gm15965	
52							Wdr78	Nudt7						Serpinf1	
53							Map4k5	Csrp2						Sic41a3	
54							Pck2	Ccl2						G77080	
55							Zfp874a	RP23-422D12.3						Nfkb	
56							Gnal	Dok3						Doc2g	
57							Zfp395	Cyp26c1						Ggh	
58							Dnajc17	Impa1						Gid8	
59							Gbp2	Gm4258						Dnajb2	
60							Cdkn1a	Pla2g4f						Repin1	
61							Rpl36a	Nanos1						Timd4	
62							Fn1	Gm16675						Gas21	
63							Anxa1							Spred3	
64							Hmox1							Fosl2	
65							Pgap1							Loxl3	
66							Klrn2							Lfng	
67							Abcc3							Haus4	
68							Timm23							Gm37590	
69							Trim23							Arntl	
70							Zcchc9							Rhobtb1	
71							Myof							2610012C04Rik	
72							Gm28466							Vps18	
73							Gcfc2							Gm29151	
74							Kcnj10							Acot11	
75							Rab9							Msh2	
76							Inip							Pld1	
77							Lig3							2810429I04Rik	
78							Fcgr1							Gm22733	
79														Jazf1	
80														Vipr1	
81														Fzd5	
82														Lnc20	
83														Phldb1	
84														Aif1	
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86														Ms4a7	
87														Orn3	
88														Serf2	
89														Vdr34	
90														Prdm1	
91														Ect2	
92														Rgs3	
93														Stap2	
94														Pitpnm1	
95														Oas1b	
96														Lpcat4	
97														Chmp7	
98														Mettl13	
99														Mical2	
100														Gp49a	
101														Gm13710	
102														Olf99	

**Figure S10: RNA-seq analysis of thymic pDCs from GF, GF-Bfrag and GF-ΔPSA monoclonalized mice**  
 Genes in each numbered section (superscript) of the Venn diagram in **Figure 3F**.

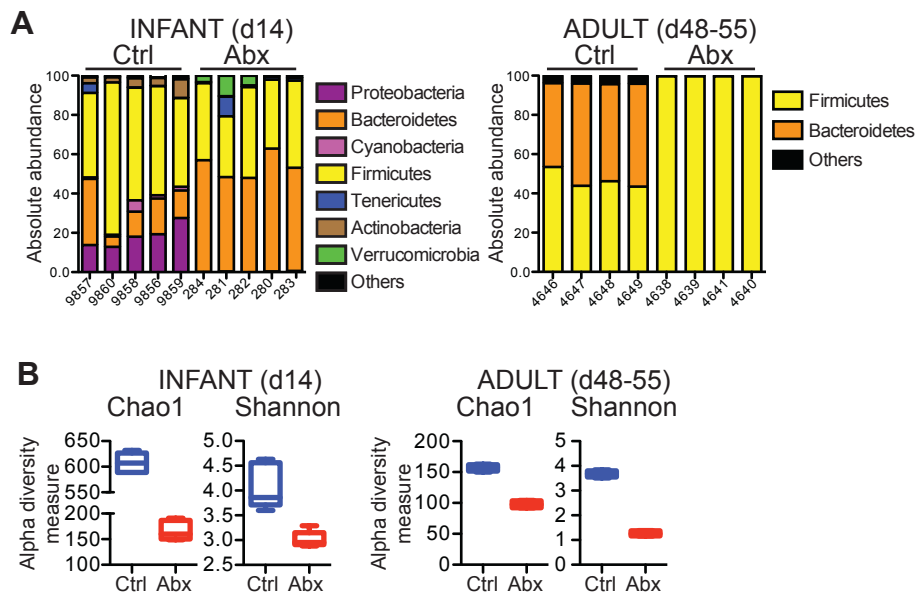


**Figure S11: Frequency of PLZF expressing subsets in thymus, spleen and colon of adult *Tlr2*<sup>-/-</sup> mice.**

**A.** Frequency of PLZF<sup>+</sup> cells in thymus, spleen and colon of adult (d48-55) mice (Thymus, Spleen: WT n= 5; HET n=7; KO n=6; Colon: WT n= 4; HET n=4; KO n=10).

Bars are Mean  $\pm$  SEM.

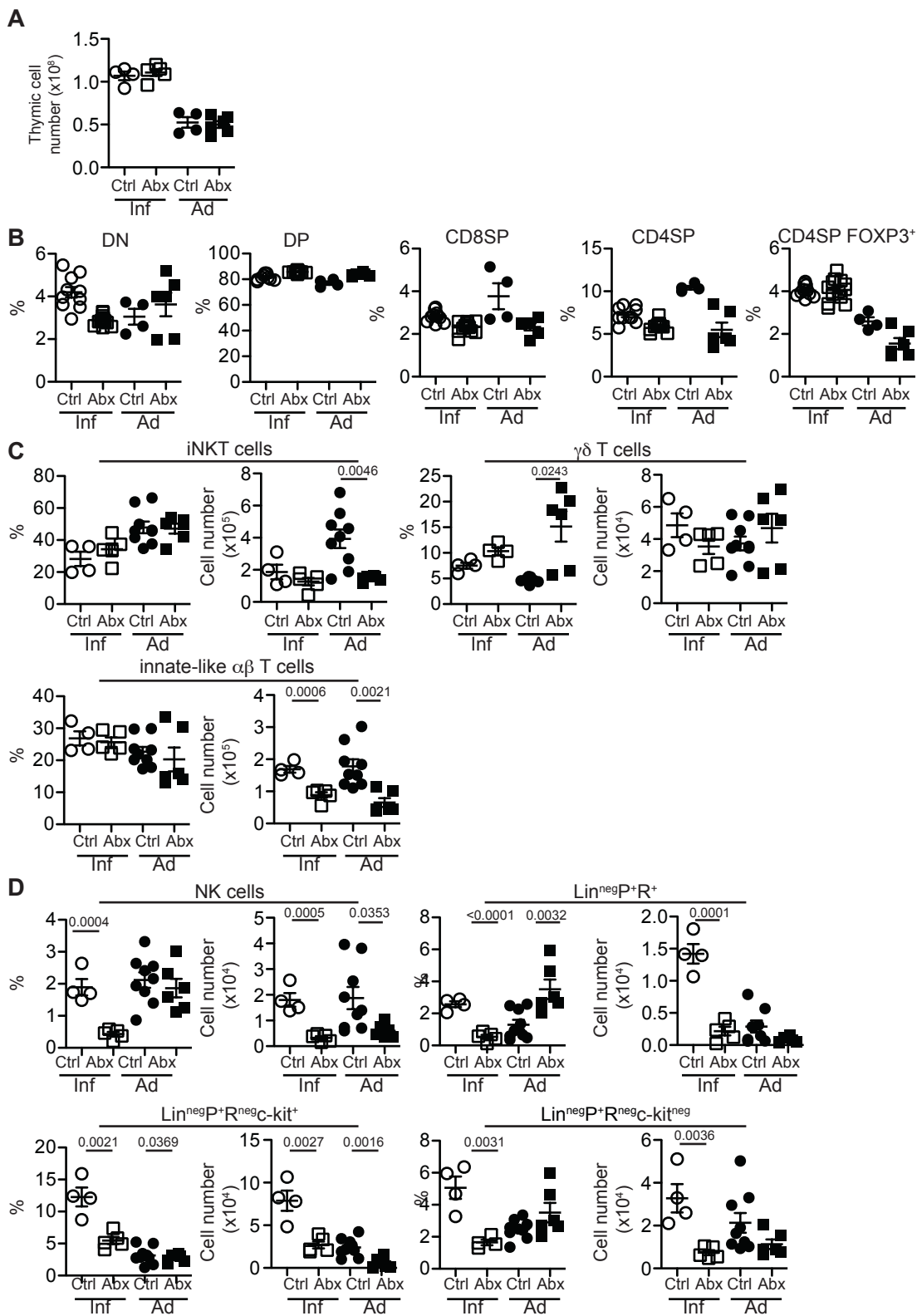




**Figure S12: Colonic microbial diversity of antibiotic treated infant and adult mice**

**A.** 16S sequencing of colonic content for bacterial diversity from infant (Ctrl n=5; Abx n=5) and adult mice that were treated with antibiotics in early life (Ctrl n=4; Abx n=4) was performed. Absolute abundance at phylum level is shown.

**B.** Chao1 and Shannon Index showing alpha diversity for complexity of microbiota within infant and adult groups.



**Figure S13: Distribution of major T cell subsets and PLZF expressing cell subsets in thymus of antibiotics treated infant and adult mice.**

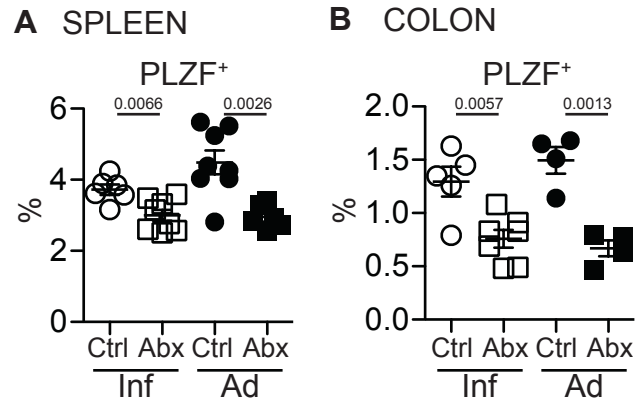
**A.** Total thymic cellularity (Inf-Ctrl n=4; Inf-Abx n=5; Ad-Ctrl n=4; Ad-Abx n=6).

**B.** Frequency of thymic DN, DP, CD8SP, CD4SP and CD4SP FOXP3<sup>+</sup> cells (Inf-Ctrl n=10; Inf-Abx n=13; Ad-Ctrl n=4; Ad-Abx n=6).

**C.** Frequency and numbers of PLZF<sup>+</sup> iNKT cells, PLZF<sup>+</sup>  $\gamma\delta$  T cells and PLZF<sup>+</sup> innate-like  $\alpha\beta$ -T cells (Inf-Ctrl n=4; Inf-Abx n=5; Ad-Ctrl n=9; Ad-Abx n=6).

**D.** Frequency and numbers of PLZF<sup>+</sup> NK cells,  $\text{Lin}^{\text{neg}}\text{PLZF}^+\text{ROR}\gamma\text{t}^+$  cells,  $\text{Lin}^{\text{neg}}\text{PLZF}^+\text{ROR}\gamma\text{t}^{\text{neg}}\text{-kit}^+$  cells and  $\text{Lin}^{\text{neg}}\text{PLZF}^+\text{ROR}\gamma\text{t}^{\text{neg}}\text{-kit}^{\text{neg}}$  cells in the thymus (Inf-Ctrl n=4; Inf-Abx n=5; Ad-Ctrl n=9; Ad-Abx n=6).

Data are from 2 independent experiments for each group. Bars are Mean  $\pm$  SEM.



**Fig. S14: Frequency of PLZF<sup>+</sup> cells in the spleen and colon of antibiotic treated infant and adult mice.**

**A.** Frequency of PLZF<sup>+</sup> cells in spleen (Inf-Ctrl n=6; Inf-Abx n=8; Ad-Ctrl n=8; Ad-Abx n=6).

**B.** Frequency of PLZF<sup>+</sup> cells in colon (Inf-Ctrl n=5; Inf-Abx n=7; Ad-Ctrl n=4; Ad-Abx n=4).

Data are representative of 2 independent experiments. Bars are Mean  $\pm$  SEM.