

Supplementary Information

Table S1 Amino acid accession and database sequence information for cytochrome b sequences used in trait evolution analyses including the resistance matrix for CWD (R = resistant, S = susceptible, U = unknown) for each species. For this analysis humans are used as the reference sequence and are assumed to be resistant to CWD infection.

Family	Species	CWD	Accession	Database source	RefSeq accession	Citation
Antilocapridae	<i>Antilocapra americana</i>	U	YP_007625795.1	RefSeq	NC_020679.1	Hassanin et al. 2012
Bovidae	<i>Bison bison</i>	U	YP_002791041.1	RefSeq	NC_012346.1	Achilli et al. 2008
Bovidae	<i>Bos taurus</i>	R	YP_209217.1	RefSeq	NC_006853.1	Chung and Ha 2005
Bovidae	<i>Oreamnos americanus</i>	U	YP_007625158.1	RefSeq	NC_020630.1	Hassanin et al. 2009
Bovidae	<i>Ovis canadensis</i>	U	YP_004769596.1	RefSeq	NC_015889.1	Miller et al. 2011
Canidae	<i>Canis latrans</i>	R	YP_626727.1	RefSeq	NC_008093.1	Björnerfeldt et al. 2006
Cebidae	<i>Saimiri sciureus</i>	S	YP_002929489.1	RefSeq	NC_012775.1	Matsui et al. 2009
Cercopithecidae	<i>Macaca fascicularis</i>	R	YP_002884238.1	RefSeq	NC_012670.1	Yi et al. 2009
Cervidae	<i>Alces alces</i>	S	YP_007625769.1	RefSeq	NC_020677.1	Hassanin et al. 2012
Cervidae	<i>Alces alces alces</i>	S	O47922.1	UniProt	NA	Randi et al. 1998
Cervidae	<i>Cervus canadensis canadensis</i>	S	AAL17840.1	Protein	NA	Ludt et al. 2004
Cervidae	<i>Muntiacus reevesi</i>	S	NP_663804.1	RefSeq	NC_004069.1	Zhang et al. 2010
Cervidae	<i>Odocoileus hemionus</i>	S	YP_007626445.1	RefSeq	NC_020729.1	Hassanin et al. 2012
Cervidae	<i>Odocoileus virginianus</i>	S	YP_004300520.1	RefSeq	NC_015247.1	Seabury et al. 2011
Cervidae	<i>Rangifer tarandus</i>	S	YP_448945.1	RefSeq	NC_007703.1	Wada et al. 2007
Cervidae	<i>Rangifer tarandus granti</i>	S	AAV84418.1	Protein	NA	Cronin et al. 2005
Cricetidae	<i>Phodopus sungorus</i>	R	ACG76170.1	Protein	NA	Meshchersky and Feoktistova 2009
Felidae	<i>Felis catus</i>	R	NP_008263.1	RefSeq	NC_001700.1	Lopez et al. 1996
Hominidae	<i>Homo sapiens</i>	R	YP_003024038.1	RefSeq	RefSeq	Anderson et al. 1981
Mustelidae	<i>Mustela putorius</i>	R	YP_007625262.1	RefSeq	NC_020638.1	Yu et al. 2011

Mustelidae	<i>Mustela putorius furo</i>	R	BAB08027.1	Protein	NA	Kurose et al. 2000
Mustelidae	<i>Neovison vison</i>	R	YP_007625301.1	RefSeq	NC_020641.1	Kurose et al. 2000
Procyonidae	<i>Procyon lotor</i>	R	YP_001096027.1	RefSeq	NC_009126.1	Takada et al. 2008

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Figure S1. Maximum likelihood phylogram of cytochrome b used in ancestral and unknown state predictions. Branches with less than 50% bootstrap support were collapsed. Susceptible species indicated with white circles at branch tips, resistant species with black circles, and unknown with grey.

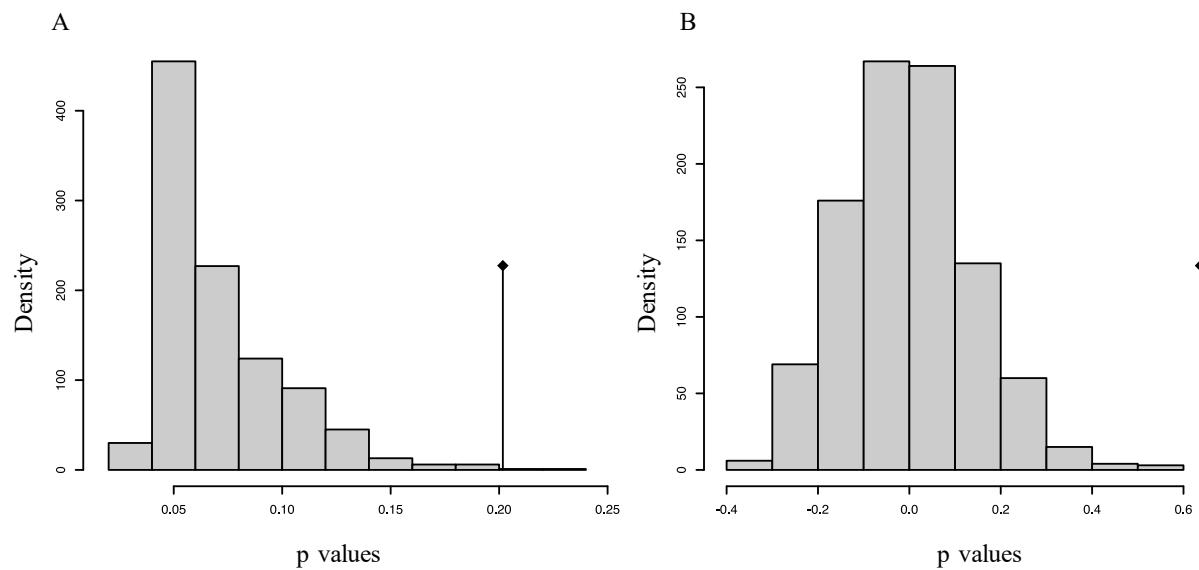


Figure S2. Susceptibility to CWD and phylogenetic signal are correlated with (A; $p = 0.003$) and without branch lengths (B; $p = 0.001$) as a factor. Significance of correlation tested using Moran's I with relationships among taxa inferred from the 19 species cytochrome b phylogeny (Fig. 2A) that does not have any missing infection data.

Additional File 1 - scripts

Model testing:

```
phyml -i infile.phy -d aa -m MTMAM -f m -v e -a e -c 4 -o tlr  
raxmlHPC-SSE3 -s infile.phy -m PROTGAMMAIMTMAM -n EXEC_NAME -p PARSIMONY_SEED  
raxml-ng --msa infile.phy --model MTMAM+I+G4  
paup -s infile.phy  
iqtree -s infile.phy -m MTMAM+I+G4
```

Running raxml:

```
raxmlHPC-HYBRID -T 4 -s infile -N automRE -n result -k -f a -p 12345 -x 12345 -m PROTCATMTMAMF
```

Adding BS values to raxml best tree:

```
raxmlHPC-AVX -f b -t RAxML_bestTree.result -z RAxML_bootstrap.result -m PROTCATMTMAMF -n BStree
```

Phylogenetic correlation

```
setwd("~/Documents/sidePieces/CWD/prnpsequencedata")  
library(adephylo)  
library(ape)  
library(phyllobase)  
prnpTre <- read.tree("RAxML_bipartitions.BStree")  
plot(prnpTre)  
suscpetMatr <- read.table("SusceptMatr.txt", header = T, sep = "\t", as.is = 1)  
suscpetMatr <- data.frame(suscpetMatr, row.names = 1)  
prnpTre$node.label<-c()  
prnpData <- phylo4d(prnpTre, suscpetMatr)  
table.phylo4d(prnpData)  
prnp.abTest <- abouheif.moran(prnpData)  
prnp.abTest  
plot(prnp.abTest)  
prnp.prox <- vcv.phylo(prnpTre)  
prnp.abTest2 <- abouheif.moran(prnpData, W=prnp.prox)  
prnp.abTest2
```