

## Supplemental data

**Supplemental Table 1.** List of primer sequences used for vector construction.

Transgenic line	Primer	Primer sequence (5' -> 3')
<b>PGD:OsEXPA7</b>	XhoI_PGD_F	TTCTCGAG TAGATATGCCGAACATGAC
	XhoI_PGD_R	AACTCGAG TCTGCCTGCAGATAGATG
<b>35S:OsEXPA7</b>	Xba_5UTR_OsEXPA7_F	TCTAGA TCGCACTCACAATGTCG
	SacI_OsEXPA7_R	GAGCTC TCAGACCCGGAAGTTCTTG
<b>OsEXPA7:GUS</b>	EXPA7promoter_B1_F	AAAAAGCAGGCTTACCAAAATTTGCCTATGAT
	EXPA7promoter_B2_R	AGAAAGCTGGGTTGTGAGTGCGAGTGGC

F, forward primer; R, reverse primer.

**Supplement Table 2.** List of primer sequences used for RT-PCR analysis.

Name	Primer sequence (5' -> 3')
<i>OsACT11-F_qRT</i>	CAGCCACACTGTCCCCATCTA
<i>OsACT11-R_qRT</i>	AGCAAGGTCGAGACGAAGGA
<i>OsEXPA7-F_qRT</i>	GCATCGTCCCCGTCTCCTA
<i>OsEXPA7-R_qRT</i>	GTTCGTGATCAGCACCAGGTT
<i>OsSOD1-F_qRT</i>	CACCACCAATGGTTGCATGT
<i>OsSOD1-R_qRT</i>	TGGCGGGTCTCATCTTCTG
<i>OsAPX1-F_qRT</i>	GGGTTCTGACCACCTAAGG
<i>OsAPX1-R_qRT</i>	CCAGAGAGGGCAACAATGT
<i>OsAPX2-F_qRT</i>	AGCTGCCAAGTGACAAAGC
<i>OsAPX2-R_qRT</i>	CCTCATCCGCAGCATATTTTC
<i>OsPOX8-F_qRT</i>	TCTCAGGTGCGCACACGAT
<i>OsPOX8-R_qRT</i>	GCGTCGATGTTGGTCTCGTT
<i>OsPOD1-F_qRT</i>	GTCCGCAAGATCAGCGACTT
<i>OsPOD1-R_qRT</i>	CGGAGTGGAAGGTGACGAT
<i>OsP5CS1-F_qRT</i>	AAGGTGGGCACTGCAGTTGT
<i>OsP5CS1-R_qRT</i>	TCCTTAACCTGCTCGCACAGA
<i>OsNHX4-F_qRT</i>	TCGCGAATCCTGAGGTT
<i>OsNHX4-R_qRT</i>	ACCTGGAAGCCTGCATTG
<i>OsSOS1-F_qRT</i>	TCAGAATGGTGCCAGCTT
<i>OsSOS1-R_qRT</i>	CGTACATGCTCAGCTGCAAT
<i>OsARF23-F_qRT</i>	CCAAATGGTACGCAAAGGAC
<i>OsARF23-R_qRT</i>	TCTAACTCGCTGCTATTGCC
<i>OsWAK1-F_qRT</i>	GAGATGTCAAGCCAGCCAAC
<i>OsWAK1-R_qRT</i>	GCCTAACTTGGAGGACCCAA
<i>OsSAPK2-F_qRT</i>	TGCCGTCAAGTTCATCGA
<i>OsSAPK2-R_qRT</i>	GCCTCAGTGACCTGTGGTT
<i>OsWRKY71-F_qRT</i>	GATGGGTACCAATGGAGGAA
<i>OsWRKY71-R_qRT</i>	GCGGGAGCAAATGAGCAT

F, forward primer; R, reverse primer.

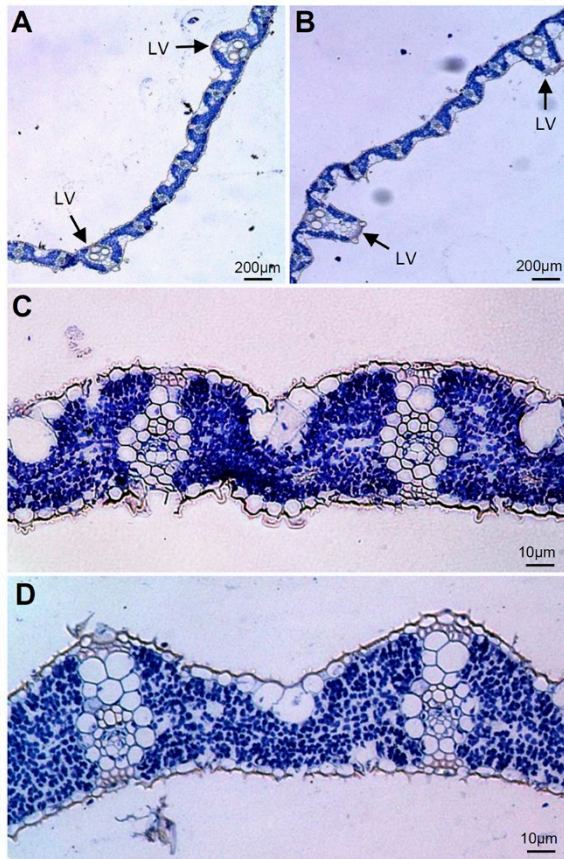
**Supplemental Table 3.** Total gene ontology (GO) terms for the DEGs in WT and *OsEXAP7*-OX plants under salt stress.

GO TYPE	GO NAME	UP	DOWN
<b>Biological Process</b>	metabolic process	66 26.9%	89 29.7%
	cellular process	59 24.1%	82 27.3%
	biological regulation	20 8.2%	25 8.3%
	Localization	19 7.8%	10 3.3%
	regulation of biological process	18 7.3%	23 7.7%
	response to stimulus	12 4.9%	20 6.7%
	Signaling	8 3.3%	6 2.0%
	cellular component organization or biogenesis	8 3.3%	12 4.0%
	multicellular organismal process	8 3.3%	6 2.0%
	developmental process	7 2.9%	6 2.0%
	reproductive process	5 2.0%	3 1.0%
	Reproduction	5 2.0%	3 1.0%
	multi-organism process	4 1.6%	4 1.3%
	rhythmic process	2 0.8%	0 0.0%
	negative regulation of biological process	1 0.4%	1 0.3%
	cell population proliferation	1 0.4%	0 0.0%
	positive regulation of biological process	1 0.4%	4 1.3%
	Growth	1 0.4%	0 0.0%
	immune system process	0 0.0%	4 1.3%
	Detoxification	0 0.0%	1 0.3%
carbon utilization	0 0.0%	1 0.3%	
<b>Molecular Function</b>	Binding	68 45.3%	77 43.8%
	catalytic activity	63 42.0%	79 44.9%
	transporter activity	7 4.7%	4 2.3%
	transcription regulator activity	6 4.0%	9 5.1%
	molecular function regulator	2 1.3%	1 0.6%
	molecular transducer activity	2 1.3%	1 0.6%
	structural molecule activity	1 0.7%	2 1.1%
	nutrient reservoir activity	1 0.7%	0 0.0%
	translation regulator activity	0 0.0%	1 0.6%
	antioxidant activity	0 0.0%	1 0.6%
	molecular transducer activity	0 0.0%	1 0.6%
	<b>Cellular component</b>	Cell	59 19.0%
cell part		58 18.7%	68 22.9%
Membrane		52 16.8%	35 11.8%
membrane part		46 14.8%	29 9.8%
Organelle		44 14.2%	43 14.5%
protein-containing complex		21 6.8%	27 9.1%
organelle part		19 6.1%	17 5.7%
extracellular region		4 1.3%	4 1.3%
membrane-enclosed lumen		2 0.6%	2 0.7%
extracellular region part		2 0.6%	0 0.0%

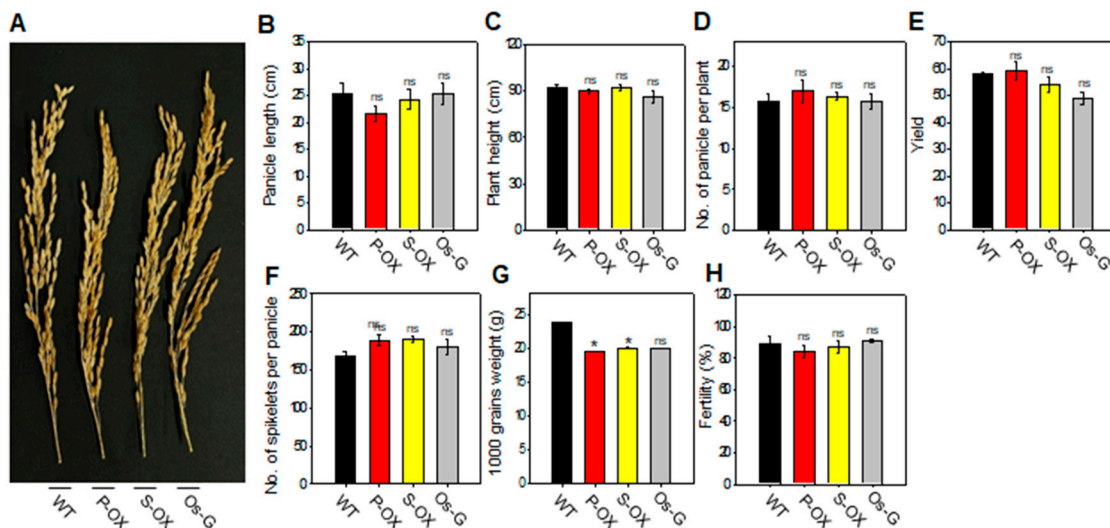
supramolecular complex	1	0.3%	0	0.0%
cell junction	1	0.3%	2	0.7%
Symplast	1	0.3%	2	0.7%
Total	705	1478	773	

**Supplemental Table 4.** List of candidate genes for salt stress tolerance among the DEGs, detected by RNA-sequencing (lower than log<sub>2</sub> scale-fold).

Gene name	Gene ID	Up or down ratio (Log <sub>2</sub> FoldChange)	Function
<i>OsNHX4</i>	LOC_Os06g21360.1	up (0.32)	Response to salt stress;regulation of intracellular pH;potassium ion homeostasis
<i>OsGAP1</i>	LOC_Os02g22130.1	up (0.62)	response to wounding;response to salt stress;abscisic acid-activated signaling pathway
<i>OsCPK13</i>	LOC_Os04g49510.1	up (0.42)	abscisic acid-activated signaling pathway
<i>OsSalT</i>	LOC_Os01g24710.1	up (1.47)	salt stress-induced protein
<i>OsTPP1</i>	LOC_Os02g44230.1	up(0.91)	response to salt stress
<i>OsANN4</i>	LOC_Os05g31760.1	up(1.81)	response to salt stress
<i>OsMAPK5</i>	LOC_Os03g17700.1	up (0.37)	stress-responsive MAPK gene



**Supplemental Figure 1.** Longitudinal section of leaf blades in WT (A, C) and *OsEXPA7-OX* plants (B, D). Large and small veins in WT (A) and *OsEXPA7-OX* (B), and enlarged small veins of WT (C) and *OsEXPA7-OX* (D). LV, long vein.



**Supplemental Figure 2.** Comparison of yield and yield components among three kinds of transgenic plants. Panicle phenotype (A), length of panicle (B), plant height (C), number of panicles per plant (D),

yield (E), number of spikelets per panicle (F), 1000 grains weight (G), and fertility (H). WT, wildtype; P-OX, PGD:*OsEXPA7*, S-OX, 35S:*OsEXPA7*; Os-G, OsEXPA7:*GUS*. Asterisks indicate significant difference examined by Student's t-test. Statistical significance was shown with \* $p < 0.05$ .