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Supplementary Materials for

Marked structural rearrangement of mannose 6-phosphate/IGF2 receptor at different pH environments

Rong Wang, Xiaofeng Qi, Philip Schmiege, Elias Coutavas, Xiaochun Li*

*Corresponding author. Email: xiaochun.li@utsouthwestern.edu

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Fig. S1. Sequence alignment of different domains in IGF2R. The sequence of domain 13 is listed without FNII domain. Strictly conserved residues are highlighted in shaded red boxes and conserved residues in open red boxes. The secondary structures of a domain are placed on the top of the alignments. The pairs of disulfide bonds are indicated by lines.



Fig. S2. Biochemical characterization of apo state IGF2R and IGF2-bound IGF2R. (A)

Size-exclusion chromatography of apo state IGF2R at pH 4.5. (**B**) Size-exclusion chromatography of IGF2-bound IGF2R at pH 7.4. The elution peak is analyzed by SDS-PAGE and Coomassie staining. Molecular standards are indicated on the left side of the SDS-PAGE.



Fig. S3. Data processing and model quality assessment of apo state IGF2R. (A) The data processing work-flow for apo state IGF2R. (B) A representative electron micrograph at -2.0 μ m defocus. (C) 2D classification for apo state IGF2R. (D) Fourier shell correlation (FSC) curve of the structure with FSC as a function of resolution using RELION-3 output. (E) FSC curves for cross-validation between the models and the maps. Curves for the final refined model versus the reconstruction from all particles in blue (sum), for the model refined against the reconstruction from only half of the particles versus the same reconstruction in red (work), and for the same model versus the reconstruction from the other half of the particles in green (free). (F) Density maps of apo state IGF2R colored by local resolution estimation using RELION-3. Each domain is separated by green dashed lines.



Fig. S4. Cryo-EM map of apo state IGF2R. β 3- β 6 and β 7- β 11 of each domain are shown. The

color scheme of the map is the same as in Fig. 1A.



Fig. S5. Structural comparison for seven subgroups of apo state IGF2R in two different

views. The seven subgroups are shown in different colors as noted in the table on the right side.



Fig. S6. Data processing for ligand-free IGF2R. (A) Negative staining image for ligand-free IGF2R at -2.0 μ m defocus. The zoomed in images show the extended morphology of IGF2R. The domains are marked with white arrows. (B) The data processing work-flow for ligand-free IGF2R with a representative electron micrograph at -2.0 μ m defocus.



Fig. S7. Data processing and model quality assessment of IGF2-bound IGF2R. (A) The data processing work-flow for IGF2-bound IGF2R. (B) A representative electron micrograph at -2.0 μ m defocus. (C) 2D classification for IGF2-bound IGF2R. (D) Fourier shell correlation (FSC) curve of the structure with FSC as a function of resolution using RELION-3 output. (E) FSC curves for cross-validation between the models and the maps. Curves for the final refined model versus the reconstruction from all particles in blue (sum), for the model refined against the

reconstruction from only half of the particles versus the same reconstruction in red (work), and for the same model versus the reconstruction from the other half of the particles in green (free). (**F**) Density maps of IGF2-bound IGF2R colored by local resolution estimation using RELION-3. Each domain is separated by green dashed lines.



Fig. S8. Cryo-EM map of IGF2-bound IGF2R. (A) β 3- β 6 and β 7- β 11 of the domains 4-14 of IGF2-bound IGF2R. The color scheme of the map is the same as in Fig. 1A. (B) α 1- α 3 of IGF2



Fig. S9. Structural comparison of CD-MPR and domains of IGF2R for Man6P. (A) Structural comparison of the lumenal domain of the CD-MPR in different pH. The proteins in pH 4.8 (PDB: 2RL7) and pH 6.5 (PDB: 2RL8) are colored in salmon and cyan, respectively. The bound Man6P and Mn^{2+} are shown with a stick model and a gray sphere, respectively. (B) Domain 5 and domain 9 of IGF2R in different pH. Domain 5 and domain 9 at pH 4.5 are colored the same as in Fig. 1A, while domain 5 and domain 9 at pH 7.4 are shown in pink and wheat. The conformational changes of the $\beta 10-\beta 11$ loops in different pH are indicated with dashed lines.

	IGF2R (pH 4.5)	IGF2R-IGF2 complex (pH7.4)
	(EMD-20815)	(EMD-20816)
	(PDB 6UM1)	(PDB 6UM2)
Data collection and processing		
Magnification	50,000	58,140
Voltage (kV)	300	300
Electron exposure $(e - / Å^2)$	80	100
Defocus range (µm)	-1.0 to -2.0	-1.0 to -2.4
Pixel size (Å)	1	0.86
Symmetry imposed	C1	C1
Initial particle images (no.)	314,735	447,586
Final particle images (no.)	128,789	75,821
Map resolution (Å)	3.46	4.32
FSC threshold 0.143		
Map resolution range (Å)	3.2-5.9	3.7-6.4
Refinement		
Initial model used (PDB code)	Domains 11-14 (2V5O)	IGF2R (pH 4.5)
	Domains 1-3 (1Q25)	Domains 11-14 (2V5O)
	Domain 5 (2VKA)	Domains 11-13-IGF2 (2V5P)
Model resolution (Å)	3.68	4.40
FSC threshold 0.5		4 40 202 7
Model resolution range (A)	3.68-280.0	4.40-302.7
Map sharpening <i>B</i> factor (A^-)	-105.2	-131.20
Model composition	17104	12004
Non-hydrogen atoms.	17134	13094
Protein residues	2208	1686
Ligands	BMA: I NAG: /	NAG: /
B factors (A ⁻)	22.26	20.00
Protein	23.26	20.80
Ligand	17.01	26.65
R.m.s. deviations	0.014	0.012
Bond lengths (A)	0.014	0.013
Bond angles (°)	1.841	1.783
Validation	1.05	1.07
MolProbity score	1.95	1.97
Clashscore	9.44	9.90
Poor rotamers (%)	0.95	0.62
Kamachandran plot	02.01	02.00
Favored (%)	92.91	92.90
Allowed (%)	6.86	6./4
Disallowed (%)	0.23	0.36

Table S1. Cryo-EM data collection, refinement, and validation statistics.