### **Supplementary Methods**

# Study Participants

RAPPER (UKCRN1471; [1]) recruited participants enrolled in the RT01 (ISRCTN47772397 [2]) and CHHiP (ISRCTN97182923 [3, 4]) trials, and was approved by the Cambridge South Research Ethics Committee (05/Q0108/365). RADIOGEN recruited participants treated at the Clinical University Hospital of Santiago de Compostela, Spain and was approved by the Galician Ethical Committee [5]. Gene-PARE [6, 7] recruited participants treated at the Mount Sinai Hospital and was approved by the Mount Sinai Medical Center Institutional Review Board. UGhent recruited participants from the Ghent University Hospital [8] and was approved by the Ghent University Hospital ethics committee. CCI-BT and CCI-EBRT [9] were recruited from the Cross Cancer Institute and the Tom Baker Cancer Centre in Canada following approval by the Health Research Ethics Board of Alberta. PRRG participants were recruited at the Hospital of the National Institute of Radiological Sciences and received either external beam photon therapy (PRRG-photon) or carbon-ion therapy (PRRG-Cion). All the patients provided written informed consent to participate in the study between 2001 and 2010, which was approved by the Certified Review Board at the National Institute of Radiological Sciences (06-004) and by each collaborating institution (Tohoku University Hospital, Yokohama City University Hospital, Nagoya City University Hospital and Kyushu University Hospital). NTMC participants were recruited from the National Tokyo Medical Center and were treated with permanent seed brachytherapy with or without external beam photon therapy. All participants provided informed consent, and the study was approved by the local Institutional Review Board.

# Assessment of late radiotherapy toxicity

Toxicity was assessed using the following: the Late Effects in Normal Tissue [10], Royal Marsden Hospital [11], and Radiation Therapy Oncology Group [12] scales (RAPPER); the NCI CTCAE [13] (RADIOGEN, CCI-EBRT, UGhent); and the American Urological Association Symptom Score [14] and an institutional scale (GenePARE). UGhent and CCI-BT used a simple measure of presence or absence for rectal bleeding. Associations between pairs of toxicities were assessed by hazard ratios, considering each toxicity as a time-dependent covariate in a Cox model for each other toxicity, unadjusted for any other predictor. If the explanatory toxicity was censored before the dependent toxicity, the dependent toxicity was artificially censored at the same earlier time.

#### Genotype Imputation

Genetic data were imputed using, as reference haplotypes, the1000 Genomes Project Phase 3 (Haplotype release date October 2014) for chromosomes 1 to 22 and the 1000 Genomes Project Phase 1 (Haplotype ChrX release date Aug 2012) for chromosome X, since the phased data for Chr X from 1000GP Phase 3 was not available. A two-stage procedure used SHAPEIT (shapeit.v2.r790.Ubuntu\_12.04.4.static) to derive phased genotypes (default parameters with the following modifications: 10 burn-in iterations, 10 pruning iterations, and 50 iterations to compute transition probabilities) and IMPUTEv2 (impute\_v2.3.2\_x86\_64\_static) to perform imputation of the phased data (default parameters with the following modifications: 5Mb non-overlapping intervals, 800 reference haplotypes to use as templates when imputing missing genotypes, and 500kb buffer region). 1000 Genomes Project variants whose minor allele frequency in Europeans and East Asians was lower than 0.001 were excluded from imputation. All OncoArray datasets were imputed jointly; the Affymetrix and Illumina CytoSNP12 datasets were imputed separately following the same procedure.

#### Fine-scale mapping

Genomic regions were defined as the 1Mb interval surrounding each statistically significant independent association. We re-imputed genotypes for the non-directly-genotyped variants using IMPUTE2 [15] and a reference panel [16] using the standard IMPUTE2 MCMC algorithm for follow-up imputation (see [17] for detailed description of the parameters used) to improve accuracy at low frequency variants. Variants with imputation info score  $\geq$ 0.3 in all cohorts and MAF  $\geq$ 0.02 in at least one cohort were included. 4,190 variants across the chr1:230337180-231337180 region; 3,776 at chr5:156903410\_157903410 and 3,987 at chr9:30366808-31366808 were evaluated for hematuria, rectal bleeding or decreased urinary stream risk, respectively. For each cohort, we ran grouped relative risk models independently and meta-analyzed the results, using a fixed-effects meta-analysis (*meta*, https://mathgen.stats.ox.ac.uk/genetics\_software/meta/meta.html). Then, the most statistically significant variant (index variant at signal 1) was used to perform conditional analysis in each cohort independently. To define the cumulative posterior probability of the credible set, we estimated the empirical Bayes Factor [18].

The conditional results were meta-analyzed and the most significant variant (index variant at signal 2) selected. This loop continued until no variants at p-values of 10<sup>-4</sup> remained at the region. A preliminary set of credible causal variants (CCVs) was then determined among the variants within two orders of magnitude from the index variant for each signal. The most significant variant (final index variant) within the set was identified by adjusting the effect of each signal by the additional signals. The final credible set was redefined among the variants with p-values within two orders of magnitude smaller than the index variant after being conditioned by the additional index variant after being conditioned by the additional index variants at the region.

For each variant (i) we normalized its effect size  $(\hat{\beta}_i)$  and variance  $(\sigma_i)$  by its allele frequency  $(p_i)$  as follows

$$\beta_{Ni} = \hat{\beta}_i \sqrt{2p_i(1-p_i)}$$
  
$$\sigma_{Ni}^2 = \sigma_i^2 2p_i(1-p_i)$$

where  $p_i$  is the allele frequency for variant i in the OncoArray cohort, and estimated the prior variance ( $\omega$ ) using (Spencer et al., 2016) approach with normalized betas and normalized variance

$$\omega_N = \widehat{\beta_{N_{1\,30}}^2} - \sigma_{Nm}^2$$

We then estimated the cumulative posterior probability of the variants included in the credible set. For regions with more than one independent signal Bayes Factor was estimated using the summary statistics from the conditional analysis, after adjusting for other index variants at the region.

### Credible causal variant (CCV) annotation

Variants were annotated with Variant Effect Predictor [19] to determine their effect on genes, transcripts, and protein sequences. To evaluate whether CCVs were located at regulatory regions, we overlapped our CCVs with Encode enhancer-like and promoter-like regions for 73 tissues and cells (primary, immortalized, *in vitro* differentiated) with available data for both enhancer- and promoter-like regions ([19-21]

and DCC accession: ENCSR037HRJ; GEO accession: GSE30567). In order to evaluate whether the CCVs could drive the expression of local genes, we accessed the GTEx Portal on 04/19/2018 to retrieve the metasoft results for all tissues in the V7 release. LocusZoom [22] was used to visualize associations for regions containing CCVs. Linkage disequilibrium was estimated using as reference the European ancestry populations from the 1000 Genomes Project (Phase 3, release 20130502; [16]).

### Pathway Analysis

Gene- and pathway-based analysis was performed using *Pascal* (<u>Pa</u>thway <u>sc</u>oring <u>algorithm</u>) [23]. Gene-based scores were computed using the default "sum" option, which calculates the sum of chi-squared statistics and measures the strongest association signal per gene, respectively. SNPs were mapped to genes using a 100kb window surrounding each gene. Pathway-based scores are computed using a modified Fisher method, which improves statistical power compared with enrichment-based analysis while maintaining rigorous type I error control. The KEGG, Biocarta, and Reactome databases were queried for the pathway-based analysis.

#### Multivariable Modeling

Clinical variables were combined with genetic variants (identified via GWAS meta-analysis and from prior studies) using cohort-stratified grouped relative risk models, assuming an additive model for each variant, resulting in per allele hazard ratios (HR). Such grouped relative risk models estimate hazard ratios based on grouped survival data, assuming proportional hazards for the latent continuous survival times within each cohort stratum. The discrete monitoring times need not be equally spaced, nor need they be the same across cohort strata, but it is necessary that they be on the same temporal grid for all subjects within each cohort stratum. Confidence intervals and p-values were likelihood based and two-sided, with p-values  $\leq 0.05$  considered statistically significant.

Stepwise model selection was used to identify a parsimonious multivariable model for each toxicity outcome. For the multivariable cohort-stratified grouped relative risk models presented in Table 4, an alpha to enter of 0.10 and an alpha to stay of 0.05 were used as model selection parameters. Genetic variants were forced into the model in advance of the inclusion of any clinical variables. The large number of tied follow-up

times were handled using the exact method, equivalent to marginal likelihood (that Efron's method approximates) –not the discrete time method that assumes proportional odds rather than proportional hazards. The log<sub>2</sub> transformation was used to symmetrize the distribution of strongly positively skewed continuous variables, thus reducing the influence of the most extreme observed covariate values and resulting in a hazard ratio per doubling of the predictor. The functional form of each continuous variable was chosen via model selection from the following options: linear; piecewise constant histospline with knots at one or more quartiles, with the option to force the slope to be 0 to the left of the first knot (as a reference group, similar to that of a histospline).

Missing data were imputed within cohorts as follows. If a variable had ≤25% of values missing within a cohort, within-cohort mean imputation was used to impute the missing values. If a variable had >25% of values missing within a cohort, the variable was set to a constant of 0 within the cohort, allowing the hazard ratio for that variable to be estimated based only on cohorts with no more than 25% missing data, without requiring subjects missing data on a subset of variables within some cohorts to be excluded entirely from the analysis. This novel approach allowed us to at least partially adjust for variables that were available in some cohorts but not others, where the adjustment would be complete for variables that truly did not vary within cohorts in which they were missing –irrespective of the true constant value within each such cohort.

In addition to grouped relative risk regression, two other multivariable modeling methods were applied to derive separate predictive models for each of the four toxicity endpoints: Polygenic Risk Score (PRS) and a machine-learning method. In both methods, a "training" set was used to derive the model and a "test" set was used to evaluate model performance such that the training set was independent of the test set. The training set included a randomly selected 50% of the RAPPER study participants and all other cohorts; the testing set included the 50% of the RAPPER study participants not included in training data.

The first method, PRS, is a linear combination of risks of multiple SNPs identified by GWAS. The risk SNPs comprising the PRS were identified via GWAS meta-analysis of results from the cohorts comprising the training set for each of the four toxicity endpoints (rectal bleeding, increased urinary frequency, decreased urinary stream, and hematuria). On the training data, we tested several P<sub>meta</sub> thresholds (1E-1, 1E-2, ..., 1E-8) in selecting SNPs for PRS, followed by LD-pruned using 1000 Genomes Project EUR panel. Afterward, we

computed PRS for each individual of the testing data, and examine the association between PRS and toxicity endpoint.

The machine learning-based method, which was previously developed and applied to RNAseq-derived gene expression data [24], was used to derive multi-SNP models for predicting the toxicity outcomes considered in this study. Since this method was designed for binary classification tasks, we focused on the binarized versions of these endpoints (grade 2 or worse toxicity vs. grade 0 or 1 toxicity) in these experiments, as in the GWAS meta-analysis. We applied this method to the training set, with the constituent SNPs filtered at the same thresholds used for PGS, and evaluated the resultant models on the test set.

# Data Management and Analysis

Genomic data were formatted using R (version 3.2.2, R Foundation for Statistical Computing, Vienna, Austria). Analysis was carried out using ProbABEL [25], which employs the coxfit2 function in the R package survival. GWAS results were meta-analyzed using Stata (version 14.2, StataCorp LLC, College Station, TX). Multivariable modeling was done using SAS (version 9.4, SAS Institute, Cary, NC). Pascal was used to compute gene and pathway scores [23].

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# **Supplementary Notes**

# The PRACTICAL Consortium (<u>http://practical.icr.ac.uk/</u>):

Rosalind A. Eeles<sup>1,2</sup>, Brian E. Henderson<sup>3\*</sup>, Christopher A. Haiman<sup>3</sup>, ZSofia Kote-Jarai<sup>1</sup>, Fredrick R. Schumacher<sup>4,5</sup>, Ali Amin Al Olama<sup>6,7</sup>, Sara Benlloch<sup>6,1</sup>, Kenneth Muir<sup>8,9</sup>, Sonja I. Berndt<sup>10</sup>, David V. Conti<sup>3</sup>, Fredrik Wiklund<sup>11</sup>, Stephen Chanock<sup>10</sup>, Susan Gapstur<sup>12</sup>, Victoria L. Stevens<sup>12</sup>, Catherine M. Tangen<sup>13</sup>, Jyotsna Batra<sup>14,15</sup>, Judith Clements<sup>14,15</sup>, Australian Prostate Cancer BioResource (APCB)<sup>14</sup>, Henrik Gronberg<sup>11</sup>, Nora Pashayan<sup>16,17</sup>, Johanna Schleutker<sup>18,19</sup>, Demetrius Albanes<sup>10</sup>, Alicja Wolk<sup>20, 21</sup>, Catharine West<sup>22</sup>, Lorelei

Mucci<sup>23</sup>, Géraldine Cancel-Tassin<sup>24,25</sup>, Stella Koutros<sup>10</sup>, Karina Dalsgaard Sorensen<sup>26,27</sup>, Eli Marie Grindedal<sup>28</sup>, David E. Neal<sup>29,30,31</sup>, Freddie C. Hamdy<sup>31</sup>, Jenny L. Donovan<sup>32</sup>, Ruth C. Travis<sup>33</sup>, Robert J. Hamilton<sup>34</sup>, Sue Ann Ingles<sup>3</sup>, Barry S. Rosenstein<sup>35,36</sup>, Yong-Jie Lu<sup>37</sup>, Graham G. Giles<sup>38,39</sup>, Adam S. Kibel<sup>40</sup>, Ana Vega<sup>41</sup>, Manolis Kogevinas<sup>42,43,44,45</sup>, Kathryn L. Penney<sup>46</sup>, Jong Y. Park<sup>47</sup>, Janet L. Stanford<sup>48,49</sup>, Cezary Cybulski<sup>50</sup>, Børge G. Nordestgaard<sup>51,52</sup>, Hermann Brenner<sup>53,54,55</sup>, Christiane Maier<sup>56</sup>, Jeri Kim<sup>57</sup>, Esther M. John<sup>58,59</sup>, Manuel R. Teixeira<sup>60,61</sup>, Susan L. Neuhausen<sup>62</sup>, Kim De Ruyck<sup>63</sup>, Azad Razack<sup>64</sup>, Lisa F. Newcomb<sup>48,65</sup>, Davor Lessel<sup>66</sup>, Radka Kaneva<sup>67</sup>, Nawaid Usmani<sup>68,69</sup>, Frank Claessens<sup>70</sup>, Paul A. Townsend<sup>71</sup>, Manuela Gago-Dominguez<sup>72,73</sup>, Monique J. Roobol<sup>74</sup>, Florence Menegaux<sup>75</sup>, Kay-Tee Khaw<sup>76</sup>, Lisa Cannon-Albright<sup>77,78</sup>, Hardev Pandha<sup>79</sup>, Stephen N. Thibodeau<sup>80</sup>

<sup>\*</sup>In memorium

<sup>1</sup> The Institute of Cancer Research, London, UK.

<sup>2</sup> Royal Marsden NHS Foundation Trust, London, UK.

<sup>3</sup> Department of Preventive Medicine, Keck School of Medicine, University of Southern California/Norris Comprehensive Cancer Center, Los Angeles, CA, USA.

<sup>4</sup> Department of Epidemiology and Biostatistics, Case Western Reserve University, Cleveland, OH, USA. <sup>5</sup> Seidman Cancer Center, University Hospitals, Cleveland, OH, USA.

<sup>6</sup> Centre for Cancer Genetic Epidemiology, Department of Public Health and Primary Care, University of Cambridge, Strangeways Research Laboratory, Cambridge, UK.

<sup>7</sup> University of Cambridge, Department of Clinical Neurosciences, Cambridge, UK.

<sup>8</sup> Division of Population Health, Health Services Research and Primary Care, University of Manchester, Manchester, UK.

<sup>9</sup> Warwick Medical School, University of Warwick, Coventry, UK.

<sup>10</sup> Division of Cancer Epidemiology and Genetics, National Cancer Institute, NIH, Bethesda, MD, USA.

<sup>11</sup> Department of Medical Epidemiology and Biostatistics, Karolinska Institute, Stockholm, Sweden.

<sup>12</sup> Epidemiology Research Program, American Cancer Society, 250 Williams Street, Atlanta, GA, USA.

<sup>13</sup> SWOG Statistical Center, Fred Hutchinson Cancer Research Center, Seattle, WA, USA.

<sup>14</sup> Australian Prostate Cancer Research Centre-Qld, Institute of Health and Biomedical Innovation and School of Biomedical Science, Queensland University of Technology, Brisbane, Queensland, Australia.

<sup>15</sup> Translational Research Institute, Brisbane, Queensland, Australia.

<sup>16</sup> University College London, Department of Applied Health Research, London, UK.

<sup>17</sup> Centre for Cancer Genetic Epidemiology, Department of Oncology, University of Cambridge, Strangeways Laboratory, Cambridge, UK.

<sup>18</sup> Department of Medical Biochemistry and Genetics, Institute of Biomedicine, University of Turku, Finland.

<sup>19</sup> Tyks Microbiology and Genetics, Department of Medical Genetics, Turku University Hospital, Finland.

<sup>20</sup> Division of Nutritional Epidemiology, Institute of Environmental Medicine, Karolinska Institutet, Sweden.

<sup>21</sup> Department of Surgical Sciences, Uppsala University, Uppsala, Sweden.

<sup>22</sup> Division of Cancer Sciences, University of Manchester, Manchester Academic Health Science Centre, Radiotherapy Related Research, Manchester NIHR Biomedical Research Centre, The Christie Hospital NHS Foundation Trust, Manchester, UK.

<sup>23</sup> Department of Epidemiology, Harvard T.H Chan School of Public Health, Boston, MA, USA.

<sup>24</sup> CeRePP, Tenon Hospital, Paris, France.

<sup>25</sup> UPMC Sorbonne Universites, GRC N°5 ONCOTYPE-URO, Tenon Hospital, Paris, France.

<sup>26</sup> Department of Molecular Medicine, Aarhus University Hospital, Denmark.

- <sup>27</sup> Department of Clinical Medicine, Aarhus University, Denmark.
- <sup>28</sup> Department of Medical Genetics, Oslo University Hospital, Norway.

<sup>29</sup> University of Cambridge, Department of Oncology, Addenbrooke's Hospital, Cambridge, UK.

<sup>30</sup> Cancer Research UK Cambridge Research Institute, Li Ka Shing Centre, Cambridge, UK.

<sup>31</sup> Nuffield Department of Surgical Sciences, University of Oxford, Oxford, UK, Faculty of Medical Science, University of Oxford, John Radcliffe Hospital, Oxford, UK.

<sup>32</sup> School of Social and Community Medicine, University of Bristol, Bristol, UK.

<sup>33</sup> Cancer Epidemiology Unit, Nuffield Department of Population Health University of Oxford, Oxford, UK.

- <sup>34</sup> Dept. of Surgical Oncology, Princess Margaret Cancer Centre, Toronto, Canada.
- <sup>35</sup> Department of Radiation Oncology, Icahn School of Medicine at Mount Sinai, New York, NY, USA.

<sup>36</sup> Department of Genetics and Genomic Sciences, Icahn School of Medicine at Mount Sinai, New York, NY, USA.

<sup>37</sup> Centre for Molecular Oncology, Barts Cancer Institute, Queen Mary University of London, John Vane Science Centre, London, UK.

<sup>38</sup> Cancer Epidemiology & Intelligence Division, The Cancer Council Victoria, Melbourne, Victoria, Australia.
 <sup>39</sup> Centre for Epidemiology and Biostatistics, Melbourne School of Population and Global Health, The University of Melbourne, Melbourne, Australia.

<sup>40</sup> Division of Urologic Surgery, Brigham and Womens Hospital, Boston, MA, USA.

<sup>41</sup> Fundación Pública Galega de Medicina Xenómica-SERGAS, Grupo de Medicina Xenómica, CIBERER, IDIS, Santiago de Compostela, Spain.

<sup>42</sup> Centre for Research in Environmental Epidemiology (CREAL), Barcelona Institute for Global Health (ISGlobal), Barcelona, Spain.

<sup>43</sup> CIBER Epidemiología y Salud Pública (CIBERESP), Madrid, Spain.

<sup>44</sup> IMIM (Hospital del Mar Research Institute), Barcelona, Spain.

<sup>45</sup> Universitat Pompeu Fabra (UPF), Barcelona, Spain.

<sup>46</sup> Channing Division of Network Medicine, Department of Medicine, Brigham and Women's Hospital/Harvard Medical School, Boston, MA, USA.

<sup>47</sup> Department of Cancer Epidemiology, Moffitt Cancer Center, Tampa, USA.

<sup>48</sup> Division of Public Health Sciences, Fred Hutchinson Cancer Research Center, Seattle, Washington, USA.

<sup>49</sup> Department of Epidemiology, School of Public Health, University of Washington, Seattle, Washington, USA.

<sup>50</sup> International Hereditary Cancer Center, Department of Genetics and Pathology, Pomeranian Medical University, Szczecin, Poland.

<sup>51</sup> Faculty of Health and Medical Sciences, University of Copenhagen, Denmark.

<sup>52</sup> Department of Clinical Biochemistry, Herlev and Gentofte Hospital, Copenhagen University Hospital, Herlev, Denmark.

<sup>53</sup> Division of Clinical Epidemiology and Aging Research, German Cancer Research Center (DKFZ), Heidelberg, Germany.

<sup>54</sup> German Cancer Consortium (DKTK), German Cancer Research Center (DKFZ), Heidelberg, Germany.

<sup>55</sup> Division of Preventive Oncology, German Cancer Research Center (DKFZ) and National Center for Tumor Diseases (NCT), Heidelberg, Germany.

<sup>56</sup> Institute for Human Genetics, University Hospital Ulm, Ulm, Germany.

<sup>57</sup> The University of Texas MD Anderson Cancer Center, Department of Genitourinary Medical Oncology, Houston, TX, USA.

<sup>58</sup> Cancer Prevention Institute of California, Fremont, CA, USA.

<sup>59</sup> Department of Health Research & Policy (Epidemiology) and Stanford Cancer Institute, Stanford University School of Medicine, Stanford, CA, USA.

<sup>60</sup> Department of Genetics, Portuguese Oncology Institute of Porto, Porto, Portugal.

<sup>61</sup> Biomedical Sciences Institute (ICBAS), University of Porto, Porto, Portugal.

<sup>62</sup> Department of Population Sciences, Beckman Research Institute of the City of Hope, Duarte, CA, USA.

<sup>63</sup> Ghent University, Faculty of Medicine and Health Sciences, Basic Medical Sciences, Gent, Belgium.

<sup>64</sup> Department of Surgery, Faculty of Medicine, University of Malaya, Kuala Lumpur, Malaysia.

<sup>65</sup> Department of Urology, University of Washington, Seattle, WA, USA.

<sup>66</sup> Institute of Human Genetics, University Medical Center Hamburg-Eppendorf, Hamburg, Germany.

<sup>67</sup> Molecular Medicine Center, Department of Medical Chemistry and Biochemistry, Medical University, Sofia, Bulgaria.

<sup>68</sup> Department of Oncology, Cross Cancer Institute, University of Alberta, Edmonton, Alberta, Canada.

<sup>69</sup> Division of Radiation Oncology, Cross Cancer Institute, Edmonton, Alberta, Canada.

<sup>70</sup> Molecular Endocrinology Laboratory, Department of Cellular and Molecular Medicine, KU Leuven, Leuven, Belgium.

<sup>71</sup> Institute of Cancer Sciences, Manchester Cancer Research Centre, University of Manchester, Manchester Academic Health Science Centre, St Mary's Hospital, Manchester, UK.

<sup>72</sup> Genomic Medicine Group, Galician Foundation of Genomic Medicine, Instituto de Investigacion Sanitaria de Santiago de Compostela (IDIS), Complejo Hospitalario Universitario de Santiago, Servicio Galego de Saúde, SERGAS, Santiago De Compostela, Spain.

<sup>73</sup> University of California San Diego, Moores Cancer Center, La Jolla, CA, USA.

<sup>74</sup> Department of Urology, Erasmus University Medical Center, Rotterdam, the Netherlands.

<sup>75</sup> Cancer & Environment Group, Center for Research in Epidemiology and Population Health (CESP), INSERM, University Paris-Sud, University Paris-Saclay, Villejuif, France.

 <sup>76</sup> Clinical Gerontology Unit, University of Cambridge, Cambridge, UK.
 <sup>77</sup> Division of Genetic Epidemiology, Department of Medicine, University of Utah School of Medicine, Salt Lake City, Utah, USA. <sup>78</sup> George E. Wahlen Department of Veterans Affairs Medical Center, Salt Lake City, UT, USA.

<sup>79</sup> The University of Surrey, Guildford, Surrey, UK.

<sup>80</sup> Department of Laboratory Medicine and Pathology, Mayo Clinic, Rochester, MN, USA.

Supplementary Table 1. Definitions of toxicity grades. Semicolons indicate "or".

Toxicity endpoint by Study and toxicity grading tool	Grade definitions
Increased urinary frequency*	
RAPPER	0 = daytime frequency >4 hour intervals (LS-s) & no treatment (LS-m) & nocturia 0-1 times (RMH)
LENT-SOMA subjective [LS-s] and management [LS-m] scales; Royal Marsden Hospital [RMH] scale	<ul> <li>(RMII)</li> <li>1 = daytime frequency 3-4 hour intervals (LS-s) or 2-3 hour intervals (LS-s); alkalization (LS-m); nocturia 2-3 times (RMH)</li> <li>2 = daytime frequency 1-2 hour intervals (LS-s); anti-spasmotic (LS-m) OR regular narcotic (LS-m); nocturia 4-5 times (RMH)</li> <li>3 = daytime frequency hourly (LS-s); nocturia 6-8 times (RMH) or &gt;8 times (RMH)</li> <li>4 = cystectomy (LS-m)</li> </ul>
RADIOGEN	0 = No toxicity
CCI-EBRT UGhent <sup>†</sup>	1 = Increase in frequency or nocturia up to 2 x normal; enuresis
PRRG	2 = Frequency or nocturia >2 x normal but <hourly< td=""></hourly<>
CTCAEv3.0 - Urinary frequency/urgency	$3 =$ Frequency or nocturia $\geq 1 \text{ x/hr}$ ; urgency; catheter indicated
Gene-PARE	0 = Had to urinate again less than 2 hours after you have urinated 'not at all' or 'less than
NTMC	1 time in 5' & no nocturia or nocturia 1 time
American Urological Association Symptom Score Q2 <sup>‡</sup> and Q7 <sup>§</sup>	1 = Had to urinate again less than 2 hours after you have urinated 'less than $\frac{1}{2}$ the time' or 'about $\frac{1}{2}$ the time'; nocturia 2 times or 3 times
	<ul> <li>2 = Had to urinate again less than 2 hours after you have urinated 'more than ½ the time';</li> <li>nocturia 4 times or 5 times</li> </ul>
	$3 =$ Had to urinate again less than 2 hours after you have urinated 'more than $\frac{1}{2}$ the time' 'almost always'
Decreased urinary stream <sup>ll</sup>	
RAPPER LENT-SOMA subjective [LS-s] and management	0 = No toxicity (LS-s) & no treatment for decreased stream (LS-m) 1 = Occasionally weak (LS-s)
[LS-m] scales	2 = Intermittent (LS-s); < 1/day self-catheterization (LS-m)
	3 = Persistent but incomplete obstruction (LS-s); dilation or > 1/day self-catheterization (LS-m)
DADIOOFN	4 = complete obstruction (LS-s); permanent catheter or surgical intervention (LS-m)
RADIOGEN CCI-EBRT	0 = No toxicity 1 = Hesitancy or dribbling, no significant residual urine; retention occurring during the
PRRG	immediate postoperative period
UGhent	2 = Hesitancy requiring medication; or operative bladder atony requiring indwelling
CTCAEv3.0 - Urinary retention	catheter beyond immediate postoperative period but for <6 weeks
	3 = More than daily catheterization indicated; urological intervention indicated (e.g., TURP, suprapubic tube, urethrotomy)

Gene-PARE NTMC American Urological Association Symptom Score Q5 <sup>¶</sup> Hematuria <sup>#</sup>	<ul> <li>4 = Life-threatening consequences; organ failure (e.g., bladder rupture); operative intervention requiring organ resection indicated</li> <li>0 = Had a weak urinary stream 'not at all' or 'less than 1 time in 5'</li> <li>1 = Had a weak urinary stream 'less than ½ the time'</li> <li>2 = Had a weak urinary stream 'about ½ the time' or 'more than ½ the time'</li> <li>3 = Had a weak urinary stream 'almost always'</li> </ul>
RAPPER LENT-SOMA subjective [LS-s], objective [LS-o], and management [LS-m] scales; RTOG late effects [RTOG] scale	<ul> <li>0 = No toxicity (LS-s) &amp; no treatment for hematuria (LS-m) &amp; no toxicity (RTOG)</li> <li>1 = Microscopic, normal hemoglobin (LS-o)</li> <li>2 = Occasional or Intermittent (LS-s); Iron therapy or Occasional transfusion/single cauterization (LS-m); Intermittent macroscopic, &lt; 10% decrease in hemoglobin (LS-o); (RTOG)</li> <li>3 = Persistent with clot (LS-s); Persistent macroscopic, 10-20% decrease in hemoglobin (LS-o); Frequent transfusion or coagulation (LS-m); (RTOG)</li> <li>4 = Refractory (LS-s); Surgical intervention (LS-m); &gt; 20% decrease in hemoglobin (LS-o); (RTOG)</li> </ul>
RADIOGEN CCI-EBRT PRRG CTCAEv3.0 - Cystitis Gene-PARE	<ul> <li>0 = None</li> <li>1 = Asymptomatic</li> <li>2 = Frequency with dysuria; macroscopic hematuria</li> <li>3 = Transfusion; IV pain medications; bladder irrigation indicated</li> <li>4 = Catastrophic bleeding; major non-elective intervention indicated</li> <li>0 = None or microscopic hematuria</li> </ul>
Institutional scale Rectal bleeding <sup>**</sup> RAPPER LENT-SOMA objective (LS-o) and management (LS-m) scales; Royal Marsden Hospital (RMH) scale	<ul> <li>2 = Macroscopic hematuria</li> <li>0 = None or Occult (LS-o) &amp; None (LS-m) &amp; None (RMH)</li> <li>1 = Stool softener, iron therapy (LS-m); Occasional-no treatment (RMH)</li> <li>2 = Occasionally, &gt;2x/week (LS-o); Occasional transfusion (LS-m); Moderate-simple OPD treatment (RMH)</li> <li>3 = Severe (blood transfusion, surgery)</li> <li>4 = Life-threatening consequences; major urgent intervention indicated</li> </ul>
RADIOGEN CCI-EBRT CTCAEv3.0 - GI hemorrhage	<ul> <li>0 = None</li> <li>1 = Mild, intervention (other than iron supplements) not indicated</li> <li>2 = Symptomatic and medical intervention or minor cauterization indicated</li> <li>3 = Transfusion, interventional radiology, endoscopic, or operative intervention indicated;</li> <li>radiation therapy (i.e., haemostasis of bleeding site)</li> <li>4 = Life-threatening consequences; major urgent intervention indicated</li> </ul>
UGhent CCI-BT	0 = None 2 = Bleeding present
Institutional scale Increased urinary frequency was not analyzed in CCI-BT bec	ause pre-radiotherapy assessments were more than one year prior to starting radiotherapy for the

Increased urinary frequency was not analyzed in CCI-BT because pre-radiotherapy assessments were more than one year prior to starting majority of participants. Abbreviations: OPD, outpatient department; NA, grade not applicable <sup>†</sup> UGhent used an institutional-specific scaled based on CTCAEv3.0 <sup>‡</sup> During the past month or so, how often have you had to urinate again less than two hours after you finished urinating?

<sup>§</sup> Over the past month, how many times per night did you most typically get up to urinate from the time you went to bed at night until the time you got up in the morning?

Decreased urinary stream was not analyzed in CCI-BT because pre-radiotherapy assessments were more than one year prior to starting radiotherapy for the majority of participants.

<sup>¶</sup> During the past month, how often have you had a weak urinary stream?

<sup>#</sup> Endpoint not available in CCI-BT, UGhent or NTMC

\*\* Rectal bleeding was not assessed in NTMC or PRRG. Rectal bleeding was assigned a single grade in GenePARE using information across all follow up assessments, and so this endpoint was not available for analysis using Cox proportional hazards modeling.

**Supplementary Table 2.** Power<sup>\*</sup> to detect <u>statistically</u> significant associations among 3,871 radiotherapy patients

Per-allele Odds Ratio	Minor allele frequency, %									
	0.05	0.10	0.15	0.25	0.35	0.50				
1.15	0	0	0	0	0	0				
1.25	0	0	0	0	1	1				
1.50	0	2	7	26	43	52				
1.75	3	22	54	89	97	99				
2.00	12	65	93	100	100	100				
2.25	34	93	100	100	100	100				

\* Assuming a type I error rate of 5x10<sup>-8</sup> and number of toxicity cases and non-toxicity controls for the most rare toxicity, hematuria, included in the study. Power for more prevalent toxicities is greater than that presented in the table.

rsID	location	Toxicity Outcome	MAF <sup>*</sup>	Info <sup>†</sup>	HR <sup>‡</sup> (95% CI)	$P_{meta}^{§}$	<b>BFDP</b> <sup>  </sup>
Common SNPs with 5x10 <sup>-8</sup> <p<sub>meta&lt;5x10<sup>-7</sup></p<sub>	(chr_position_a1_a2)				<b>,</b>		
rs9644474	8 137163144 C T	Rectal bleeding	0.05	0.78	2.15 (1.64, 2.81) <sup>¶</sup>	7.9x10 <sup>-08</sup>	9.1
rs75759941	23 38151042 T C	STAT score	0.05	0.99	0.25 (0.16, 0.34) <sup>#</sup>	9.3x10 <sup>-8</sup>	5.8
rs11122572	1 230825427 T G	Hematuria	0.09	0.99	1.86 (1.48, 2.34)	9.8x10 <sup>-08</sup>	8.2
rs368141164	11_116445686_A_T	Increased urinary frequency	0.06	0.95	1.87 (1.48, 2.36)	1.8x10 <sup>-07</sup>	13.8
NA	18_57916552_A_C	STAT score	0.26	1.00	0.07 (0.04, 0.10) #	1.6x10 <sup>-7</sup>	23.5
rs2031925	10_30680024_T_C	Rectal bleeding	0.05	0.59	2.43 (1.73, 3.40)	$2.4 \times 10^{-07}$	33.1
rs17190422	2 56856260 A G	Hematuria	0.06	0.90	2.57 (1.79, 3.68)	3.1x10 <sup>-07</sup>	43.0
rs74346764	3 145230069 G A	Rectal bleeding	0.05	0.89	2.52 (1.77, 3.58)	3.1x10 <sup>-07</sup>	41.2
rs72993079	19_6573511_C_T	Hematuria	0.05	0.05	2.14 (1.60, 2.87)	$3.5 \times 10^{-07}$	30.3
rs9832989	3_133701823_A_G	Hematuria	0.05	0.83	4.45 (2.50, 7.92)	3.8x10 <sup>-07</sup>	88.9
rs60424486			0.08	0.83		$4.0 \times 10^{-07}$	
	7_15410311_G_INS	Decreased urinary stream			1.84 (1.45, 2.33)	$4.0010^{-07}$	24.1
rs11624322	14_37058609_C_T	Decreased urinary stream	0.37	0.99	1.51 (1.29, 1.77)	4.2X10	19.0
rs61871726	10_122293568_T_C	Rectal bleeding	0.07	0.92	1.98 (1.52, 2.59)	4.8x10 <sup>-07</sup>	31.2
rs2237706	7_107633171_C_T	Rectal bleeding	0.09	0.68	1.94 (1.50, 2.52)	$4.8 \times 10^{-07}$	30.0
rs6791846	3_177119317_G_A	Increased urinary frequency	0.45	0.90	1.47 (1.26, 1.71)	5.0x10 <sup>-07</sup>	21.3
Rare variants with P <sub>meta</sub> <5x10 <sup>-7</sup>					(	09	
NA	12_102080173_A_G	RecBld	0.03	0.82	3.57 (2.37, 5.39)	1.4x10 <sup>-09</sup>	2.6
rs180958289	19_1543771_G_A	DecStrm	0.02	0.65	7.78 (3.92, 15.5)	4.6x10 <sup>-09</sup>	72.4
rs13403657	2_241943450_A_G	Hematuria	0.03	0.82	3.26 (2.17, 4.90)	1.4x10 <sup>-08</sup>	11.8
rs139239158	3_59904329_C_G	DecStrm	0.02	0.80	3.69 (2.32, 5.87)	3.4x10 <sup>-08</sup>	34.0
rs191705561	4_184691564_G_T	UrineFreq	0.01	0.54	3.05 (2.05, 4.53)	3.7x10 <sup>-08</sup>	18.9
rs148048756	8_562963_A_G	RecBld	0.03	0.78	4.83 (2.75, 8.47)	4.0x10 <sup>-08</sup>	65.7
rs75988504	2_141730052_G_A	DecStrm	0.04	0.99	2.01 (1.56, 2.58)	$4.9 \times 10^{-08}$	5.5
rs139288166	8_705922_A_C	RecBld	0.03	0.85	4.30 (2.53, 7.31)	6.9x10 <sup>-08</sup>	64.4
rs139882217	3_54729912_C_T	Hematuria	0.04	0.74	4.78 (2.71, 8.44)	6.9x10 <sup>-08</sup>	73.8
NA	12_31100664_C_G	Hematuria	0.02	0.99	9.22 (4.06, 20.9)	1.1x10 <sup>-07</sup>	97.5
NA	12_33380941_C_T	STAT score	0.04	0.99	0.17 (0.11, 0.23) <sup>#</sup>	1.1x10 <sup>-07</sup>	8.0
rs112193369	1_7558251_A_INS	DecStrm	0.02	0.91	4.09 (2.43, 6.90)	1.2x10 <sup>-07</sup>	70.3
rs4688181	3_63989456_A_G	Hematuria	0.04	0.92	3.22 (2.09, 4.98)	1.3x10 <sup>-07</sup>	47.3
rs149176864	8_30559235_G_A	Rectal bleeding	0.02	0.73	2.99 (1.99, 4.49)	1.4x10 <sup>-07</sup>	41.4
rs3739643	9_4600633_C_T	Hematuria	0.04	0.89	3.44 (2.17, 5.47)	1.7x10 <sup>-07</sup>	60.3
rs73539559	6_112344737_C_T	Hematuria	0.02	0.85	2.86 (1.93, 4.26)	2.0x10 <sup>-07</sup>	44.4
rs190601686	9_86200809_C_T	Hematuria	0.01	0.79	9.13 (3.95, 21.1)	2.3x10 <sup>-07</sup>	98.3
rs149927798	3 26548947 C T	RecBld	0.02	0.55	3.98 (2.36, 6.71)	$2.3 \times 10^{-07}$	77.8
rs490393	6_114228700_C_A	Hematuria	0.03	0.82	2.96 (1.96, 4.47)	$2.4 \times 10^{-07}$	51.6
rs73712257	8_138319386_A_T	RecBld	0.03	0.49	4.72 (2.62, 8.50)	$2.4 \times 10^{-07}$	87.6
rs144214859	2_68624610_G_A	Hematuria	0.02	0.99	2.58 (1.80, 3.70)	$2.5 \times 10^{-07}$	39.3
rs61415111	11_17420841_A_C	UrineFreq	0.02	0.55	4.60 (2.57, 8.24)	3.0x10 <sup>-07</sup>	88.3
rs77581414	19_10934094_T_A	DecStrm	0.02	0.01	6.07 (3.04, 12.1)	3.1x10 <sup>-07</sup>	95.5
rs76661052	5 144759200 C T	Hematuria	0.04	0.93	2.33 (1.68, 3.22)	$3.3 \times 10^{-07}$	95.5 35.5
						$3.3 \times 10^{-07}$ 3.4x10 <sup>-07</sup>	
rs78166464	19_5200653_C_T	DecStrm	0.01	0.95	2.83 (1.90, 4.21)	3.4x10 3.4x10 <sup>-07</sup>	54.5
rs186353960	4_123920497_G_A	UrineFreq	0.04	0.66	2.44 (1.73, 3.44)	3.4X IU	40.5

Supplementary Table 3. Top SNPs that did not reach genome-wide significance and rare variants

rs11687040	2_238080051_G_A	DecStrm	0.01	0.99	3.37 (2.11, 5.40)	4.2x10 <sup>-07</sup>	74.5
rs73046248	3_21759604_T_C	DecStrm	0.04	0.85	2.40 (1.71, 3.36)	4.2x10 <sup>-07</sup>	43.3
rs192744896	15_27746339_T_C	RecBld	0.03	0.57	3.72 (2.23, 6.18)	4.3x10 <sup>-07</sup>	81.5
rs141203061	3_36244696_A_T	UrineFreq	0.03	0.86	2.71 (1.84, 4.00)	4.3x10 <sup>-07</sup>	55.7
rs113370662	4_3537697_C_T	DecStrm	0.04	0.81	2.78 (1.87, 4.15)	4.5x10 <sup>-07</sup>	59.3
rs138731641	9_83229027_GA_G	Hematuria	0.04	0.92	5.46 (2.82, 10.6)	4.8x10 <sup>-07</sup>	95.3
rs72915971	11_56925541_C_G	RecBld	0.02	0.95	2.16 (1.60, 2.91)	5.0x10 <sup>-07</sup>	38.1

<sup>\*</sup> Minor allele frequency is from PRACTICAL oncoarray samples of European ancestry. Abbreviations: SNP, single nucleotide polymorphism; MAF, minor allele frequency; HR, hazard ratio; CI, confidence interval; BFDP, Bayesian false discovery probability; NA, not available; INS, insertion.

<sup>†</sup> Imputation info score values are from the oncoarray.
 <sup>†</sup> Hazard ratio corresponds to the minor allele with the major allele treated as the reference group.
 <sup>§</sup> <u>Two-sided</u> P<sub>meta</sub> was calculated using a Wald test.
 <sup>II</sup> BFPD estimated assuming a prior variance, W = 0.32^2, and prior probability of a non-null association 0.0001
 <sup>II</sup> Hazard ratio is for the major allele with the minor allele treated as the reference group.

<sup>#</sup>Beta coefficient from linear regression of STAT score at 2 years after radiotherapy

		Rof	Effect						Gene PARE-II	RADIO	RAPPER-II	llGhant	CCI- ERBT		RAPPER-
rsid	Position <sup>*</sup>		Allele	Info <sup>†</sup>	Info <sup>‡</sup>	<b>Info</b> §	Info <sup>∥</sup>	EAF	EAF	GEN EAF	EAF	EAF	EAF	EAF	EAF
hr1:230337180-2	231337180, as	sociate	ed with h	ematur	ia										
Signal 1															
rs11122572	230825427	Т	G	0.96	0.96	0.95	0.93		0.09	0.10	0.08	0.09	0.09	0.07	0.09
rs4846866	230836065	Т	С	1.00	0.99	0.97	0.92		0.06	0.09	0.07	0.07	0.08	0.06	0.08
rs61762468	230836281	G	С	0.98	0.96	0.98	0.90		0.06	0.09	0.07	0.08	0.08	0.06	0.08
rs56117713	230836568	Т	С	0.99	1.00	0.99	0.92		0.06	0.08	0.07	0.07	0.08	0.06	0.08
rs16852352	230836786	Т	G	0.99	1.00	1.00	0.92		0.06	0.08	0.07	0.07	0.08	0.06	0.08
rs11122573	230837180	С	Т	0.99	1.00	1.00	0.92		0.06	0.08	0.07	0.07	0.08	0.06	0.08
rs10864770	230837437	G	А	0.99	0.97	1.00	0.92		0.06	0.08	0.07	0.07	0.08	0.06	0.08
rs10864771	230837672	Т	G	0.99	1.00	0.99	0.92		0.06	0.08	0.07	0.07	0.08	0.06	0.08
rs11122574	230837808	С	Т	0.97	0.96	0.96	0.86		0.04	0.06	0.05	0.05	0.07	0.04	0.05
rs1926723	230840096	Т	С	0.98	0.97	1.00	0.92		0.06	0.08	0.07	0.07	0.08	0.06	0.08
rs1926722	230840197	С	А	0.98	1.00	1.00	0.92		0.06	0.08	0.07	0.07	0.08	0.06	0.08
rs11122575	230840269	Α	G	0.98	1.00	1.00	0.92		0.06	0.08	0.07	0.07	0.08	0.06	0.08
rs11568056	230842497	С	Т	0.97	0.98	0.96	0.91		0.06	0.09	0.07	0.08	0.08	0.06	0.08
rs11122576	230846679	Т	С	1.00	1.00	0.98	0.95		0.06	0.09	0.07	0.08	0.08	0.06	0.08
rs11568028	230847244	С	Т	1.00	1.00	1.00	0.95		0.06	0.09	0.07	0.08	0.08	0.06	0.08
rs11122578	230847789	G	А	1.00	1.00	1.00	0.95		0.06	0.09	0.07	0.08	0.08	0.06	0.08
rs3789679	230849694	G	А	0.98	0.99	0.97	0.94		0.06	0.09	0.07	0.08	0.08	0.06	0.08
rs9804147	230853359	G	А	0.99	0.99	0.99	0.98		0.10	0.11	0.10	0.10	0.09	0.09	0.10
rs9804153	230853905	G	Т	0.99	1.00	0.99	0.98		0.10	0.11	0.10	0.10	0.09	0.09	0.10
rs4028824	230854141	G	А	0.99	1.00	1.00	0.98		0.10	0.11	0.10	0.10	0.09	0.09	0.10
rs10864773	230856996	С	Т	0.99	1.00	0.97	0.97		0.06	0.09	0.09	0.09	0.09	0.07	0.09
rs61762467	230836254-5	-	Т	0.99	1.00	0.99	0.92		0.06	0.08	0.07	0.07	0.08	0.06	0.08
Signal 2															
rs4846857	230436175	А	G	1.00	0.64	0.61	0.48		0.86	0.87	0.85	0.85	0.86	0.84	0.85
rs79434380	230451274	С	Т	0.68	0.52	0.57	0.45		0.03	0.02	0.01	0.01	0.02	0.03	0.02
rs564325629	230451573	G	-	0.71	0.52	0.53	0.45		0.03	0.01	0.01	0.01	0.01	0.03	0.01
rs147121532	230451849	т	С	0.72	0.52	0.53	0.45		0.03	0.01	0.01	0.01	0.01	0.02	0.01
rs28605378	230882070	G	A	0.96	0.95	0.78			0.08	0.07	0.06	0.08	0.06	0.09	0.06
rs12095859	230882213	Т	С	0.95	0.95	0.78	0.90		0.08	0.07	0.06	0.07	0.06	0.09	0.06
rs12091328	230884209	G	C	0.99	0.96	0.80	0.90		0.08	0.07	0.06	0.08	0.06	0.09	0.06
rs12059171	230885399	Т	G	0.99	0.96	0.81	0.91		0.08	0.07	0.06	0.07	0.06	0.09	0.06
rs12060898	230887539	A	G	0.98	0.96	0.82	0.90		0.08	0.07	0.06	0.07	0.06	0.09	0.06
rs75991123	230892946	C	A	0.96	0.97	0.89			0.06	0.06	0.05	0.06	0.05	0.08	0.06

Supplementary Table 4. Credible causal variants identified by fine-scale mapping.

chr5:156903410-1	57903410, as	sociate	d with	rectal b	leeding										
rs17055178	157403410	А	G	0.96	0.94		0.96	0.08		0.08	0.08	0.07	0.07		0.06
rs13180537	157419681	С	Т	0.90	0.81		0.88	0.07		0.07	0.07	0.06	0.05		0.05
rs78394554	157438561	А	С	0.93	0.91		0.94	0.08		0.08	0.08	0.07	0.06		0.06
rs34395161	157440433	С	А	0.90	0.80		0.89	0.07		0.07	0.07	0.06	0.05		0.05
rs35327501	157452625	G	А	0.94	0.91		0.94	0.08		0.08	0.08	0.07	0.06		0.06
rs4704767	157470956	С	Т	1.00	1.00		0.98	0.08		0.11	0.10	0.09	0.08		0.08
rs35929592	157471129	С	Т	1.00	1.00		0.98	0.08		0.11	0.10	0.09	0.08		0.08
rs35766682	157472745	С	Т	0.99	1.00		0.98	0.08		0.11	0.10	0.09	0.09		0.08
rs10515757	157473330	А	Т	1.00	1.00		0.98	0.08		0.11	0.10	0.09	0.08		0.08
rs35153425	157478391	А	G	1.00	1.00		0.98	0.08		0.11	0.10	0.09	0.08		0.08
rs17055241	157480829	G	А	1.00	1.00		0.98	0.08		0.11	0.10	0.09	0.08		0.08
rs1040926	157483536	А	G	1.00	1.00		0.98	0.09		0.11	0.10	0.09	0.08		0.08
rs13179825	157485371	С	Т	1.00	1.00		0.98	0.08		0.11	0.10	0.08	0.08		0.08
rs13184115	157485718	С	А	1.00	1.00		0.97	0.09		0.11	0.10	0.08	0.09		0.08
rs17229231	157486935	С	Т	0.99	1.00		0.98	0.08		0.11	0.10	0.09	0.08		0.08
chr9:30366808-31	366808, asso	ciated v	with de	ecrease	d urinar	y strear	n								
rs10969913	30866808	А	G	0.95	0.83	0.98	0.70		0.03	0.01	0.01		0.02	0.04	0.01
rs7868409	30868163	Т	С	0.95	1.00	0.96	0.72		0.03	0.01	0.01		0.02	0.04	0.01
rs10969915	30868871	А	С	0.95	0.88	0.98	0.72		0.03	0.01	0.01		0.01	0.04	0.01
rs10969916	30869372	С	G	0.95	0.86	0.98	0.72		0.03	0.01	0.01		0.01	0.04	0.01
rs1412406	30869687	С	Т	0.95	0.88	0.98	0.71		0.03	0.01	0.01		0.01	0.04	0.01
rs539024322	30873589	А	AT	0.94	0.87	0.97	0.69		0.03	0.01	0.01		0.02	0.04	0.02
rs10969918	30875780	Т	С	0.97	1.00	0.95	0.71		0.03	0.01	0.01		0.02	0.04	0.01
rs112134389	30876262	Т	С	0.97	0.92	0.99	0.71		0.03	0.01	0.01		0.02	0.04	0.01
rs111692482	30876567	С	Т	0.97	0.93	0.99	0.71		0.03	0.01	0.01		0.02	0.04	0.01
rs10969920	30876943	А	С	0.97	1.00	1.00	0.71		0.03	0.01	0.01		0.02	0.04	0.01
rs10969923	30877317	Т	С	0.96	0.88	0.97	0.68		0.03	0.01	0.01		0.02	0.04	0.02
rs73644367	30877456	А	С	0.98	0.92	0.98	0.71		0.03	0.01	0.01		0.02	0.04	0.01
rs73644368	30877580	А	G	0.97	0.91	0.97	0.71		0.03	0.01	0.01		0.02	0.04	0.01
rs10969926	30878999	Т	А	0.98	1.00	1.00	0.71		0.03	0.01	0.01		0.02	0.04	0.01
rs77191587	30866630-2	ATT	-	0.95	0.87	0.98	0.71		0.03	0.01	0.01		0.01	0.04	0.01
* O D O L O T //	A 1 1 1 1										-		-		

GRCh37/hg19, bp; Abbreviations: EAF, effect allele frequency; -- indicates that a value was not available. <sup>†</sup> Info score from imputation of SNP data generated via the Illumina Oncoarray <sup>‡</sup> Info score from imputation of SNP data generated in CCI-EBRT via the AffySNPv6.0 array <sup>§</sup> Info score from imputation of SNP data generated in GenePARE-I via the AffySNPv6.0 array <sup>II</sup> Info score from imputation of SNP data generated in RAPPER-I via the Illumina CytoSNP12 array

	Non-co	onditional	result	:s <sup>*</sup>		Co	onditional	results	6	
rsid	HR (95% CI)	$P^{\dagger}$	Q	ľ	${\pmb P_{het}}^\ddagger$	HR (95% CI)	$P^{\dagger}$	Q	l <sup>2</sup>	${\pmb P_{het}}^\ddagger$
chr1:230337180-2313	37180, associated with	hematuria	à							
Signal 1										
rs11122572	1.85 (1.47-2.32)	1.9E-07	8.0	24.7	0.24	1.82 (1.44-2.29)	3.9E-07	8.5	29.2	0.20
rs4846866	1.90 (1.50-2.40)	1.2E-07	8.5	29.0	0.21	1.87 (1.48-2.38)	2.4E-07	9.7	37.8	0.14
rs61762468	1.90 (1.50-2.41)	1.3E-07	8.1	25.9	0.23	1.87 (1.47-2.38)	2.8E-07	9.2	34.8	0.16
rs56117713	1.89 (1.49-2.40)	1.8E-07	7.8	23.6	0.25	1.86 (1.47-2.37)	3.7E-07	9.0	33.1	0.18
rs16852352	1.89 (1.49-2.40)	1.9E-07	7.9	23.9	0.25	1.86 (1.46-2.37)	3.9E-07	9.0	33.3	0.17
rs11122573	1.89 (1.49-2.40)	1.9E-07	7.8	23.2	0.25	1.86 (1.46-2.37)	3.9E-07	8.9	32.8	0.18
rs10864770	1.89 (1.49-2.40)	1.8E-07	7.8	22.8	0.26	1.86 (1.47-2.37)	3.7E-07	8.9	32.5	0.18
rs10864771	1.89 (1.49-2.40)	1.8E-07	7.8	22.8	0.26	1.86 (1.47-2.37)	3.7E-07	8.9	32.5	0.18
rs11122574	1.92 (1.43-2.57)	1.1E-05	9.3	35.7	0.16	1.90 (1.42-2.55)	1.6E-05	10.5	42.7	0.11
rs1926723	1.86 (1.46-2.37)	5.2E-07	6.9	13.5	0.33	1.83 (1.43-2.33)	1.1E-06	8.0	25.2	0.24
rs1926722	1.86 (1.46-2.37)	5.0E-07	6.9	13.6	0.33	1.83 (1.44-2.33)	1.1E-06	8.0	25.2	0.24
rs11122575	1.86 (1.46-2.37)	4.8E-07	7.0	13.9	0.32	1.83 (1.44-2.34)	1.0E-06	8.0	25.5	0.23
rs11568056	1.84 (1.45-2.34)	7.1E-07	7.8	23.6	0.25	1.82 (1.42-2.31)	1.4E-06	9.0	33.2	0.17
rs11122576	1.76 (1.39-2.23)	3.0E-06	7.9	24.0	0.25	1.74 (1.37-2.21)	5.4E-06	9.0	33.0	0.18
rs11568028	1.76 (1.39-2.23)	3.0E-06	7.9	23.9	0.25	1.74 (1.37-2.21)	5.4E-06	8.9	32.8	0.18
rs11122578	1.76 (1.39-2.24)	2.9E-06	7.9	24.3	0.24	1.74 (1.37-2.21)	5.3E-06	9.0	33.1	0.18
rs3789679	1.78 (1.40-2.26)	3.0E-06	7.1	15.3	0.31	1.75 (1.37-2.23)	5.9E-06	8.1	25.8	0.23
rs9804147	1.65 (1.32-2.05)	1.1E-05	6.4	6.6	0.38	1.63 (1.31-2.04)	1.7E-05	6.7	10.4	0.35
rs9804153	1.64 (1.31-2.05)	1.2E-05	6.4	6.1	0.38	1.62 (1.30-2.03)	1.8E-05	6.6	9.6	0.36
rs4028824	1.64 (1.32-2.05)	1.1E-05	6.2	3.8	0.40	1.63 (1.30-2.03)	1.7E-05	6.5	7.1	0.37
rs10864773	1.68 (1.33-2.12)	1.1E-05	6.1	1.9	0.41	1.67 (1.32-2.11)	1.8E-05	6.9	13.1	0.33
rs61762467	1.90 (1.50-2.41)	1.3E-07	8.0	25.1	0.24	1.87 (1.47-2.38)	2.8E-07	9.2	34.4	0.17
Signal 2										
rs4846857	0.66 (0.53-0.82)	1.8E-04	21.6	72.2	0.00	0.65 (0.52-0.81)	1.4E-04	22.8	73.7	0.00
rs79434380	4.02 (2.14-7.54)	1.5E-05	5.5	0.0	0.48	4.00 (2.12-7.54)	1.8E-05	5.0	0.0	0.55
rs564325629	4.29 (2.29-8.06)	5.8E-06	6.0	0.0	0.43	4.29 (2.27-8.08)	6.9E-06	6.3	4.4	0.39
rs147121532	4.43 (2.35-8.33)	3.9E-06	6.1	1.1	0.42	4.42 (2.34-8.34)	4.7E-06	6.4	5.7	0.38
rs28605378	1.61 (1.23-2.10)	4.9E-04	4.3	0.0	0.64	1.60 (1.22-2.09)	6.1E-04	4.3	0.0	0.64
rs12095859	1.63 (1.24-2.12)	3.6E-04	4.2	0.0	0.65	1.62 (1.24-2.11)	4.5E-04	4.2	0.0	0.65
rs12091328	1.62 (1.24-2.11)	3.4E-04	4.5	0.0	0.61	1.61 (1.23-2.10)	4.4E-04	4.5	0.0	0.61
rs12059171	1.64 (1.26-2.13)	2.5E-04	4.5	0.0	0.61	1.63 (1.25-2.12)	3.2E-04	4.5	0.0	0.60

Supplementary Table 5. Association results for credible causal variants identified by fine-scale mapping

rs12060898	1.65 (1.27-2.14)	1.8E-04	4.8	0.0	0.58	1.64 (1.26-2.13)	2.4E-04	4.8	0.0	0.58
rs75991123	1.72 (1.30-2.28)	1.3E-04	7.3	18.1	0.29	1.69 (1.28-2.24)	2.4E-04	7.2	17.1	0.30
chr5:156903410-157903	410, associated with	rectal blee	eding			, , , , , , , , , , , , , , , , , , ,				
rs17055178	1.88 (1.52-2.33)	4.9E-09	3.4	0.0	0.64					
rs13180537	1.91 (1.51-2.43)	8.1E-08	5.3	6.0	0.38					
rs78394554	1.85 (1.49-2.30)	2.7E-08	4.1	0.0	0.53					
rs34395161	1.89 (1.49-2.40)	1.4E-07	5.3	6.0	0.38					
rs35327501	1.82 (1.47-2.26)	5.9E-08	3.9	0.0	0.56					
rs4704767	1.68 (1.38-2.04)	2.4E-07	3.1	0.0	0.69					
rs35929592	1.68 (1.38-2.04)	2.4E-07	3.0	0.0	0.69					
rs35766682	1.66 (1.37-2.03)	4.2E-07	3.4	0.0	0.64					
rs10515757	1.68 (1.38-2.04)	2.4E-07	3.1	0.0	0.69					
rs35153425	1.68 (1.38-2.05)	2.3E-07	3.1	0.0	0.68					
rs17055241	1.68 (1.38-2.05)	2.3E-07	3.1	0.0	0.68					
rs1040926	1.68 (1.38-2.05)	2.4E-07	3.1	0.0	0.68					
rs13179825	1.68 (1.38-2.05)	2.2E-07	3.1	0.0	0.68					
rs13184115	1.67 (1.37-2.03)	3.5E-07	3.2	0.0	0.67					
rs17229231	1.67 (1.37-2.03)	3.1E-07	2.9	0.0	0.72					
chr9:30366808-3136680	8, associated with de	ecreased u	rinary	stream						
rs10969913	4.59 (2.76-7.63)	4.1E-09	11.3	55.6	0.05					
rs7868409	4.49 (2.75-7.35)	2.2E-09	8.6	42.0	0.13					
rs10969915	4.55 (2.74-7.54)	4.5E-09	9.9	49.3	0.08					
rs10969916	4.53 (2.72-7.54)	6.3E-09	9.8	48.9	0.08					
rs1412406	4.55 (2.74-7.54)	4.5E-09	9.9	49.4	0.08					
rs539024322	4.13 (2.50-6.79)	2.6E-08	10.3	51.2	0.07					
rs10969918	4.73 (2.90-7.74)	5.5E-10	8.7	42.8	0.12					
rs112134389	4.68 (2.84-7.72)	1.5E-09	8.7	42.3	0.12					
rs111692482	4.68 (2.84-7.72)	1.4E-09	8.6	42.1	0.12					
rs10969920	4.74 (2.90-7.74)	5.1E-10	8.7	42.8	0.12					
rs10969923	4.50 (2.72-7.46)	5.2E-09	10.5	52.4	0.06					
rs73644367	4.66 (2.81-7.74)	2.5E-09	8.5	52.8	0.08					
rs73644368	4.64 (2.80-7.69)	2.8E-09	8.9	44.0	0.11					
rs10969926	4.74 (2.91-7.75)	4.9E-10	8.5	52.7	0.08					

<sup>†</sup> Single variant summary statistics. Abbreviations: HR, hazard ratio; CI, confidence interval; -- indicates that a value was not available. <sup>†</sup>  $\frac{\text{Two-sided}}{\text{Two-sided}} P_{\text{meta}}$  was calculated using a Wald test. <sup>‡</sup>  $\frac{\text{Two-sided}}{\text{Two-sided}} P_{\text{meta}}$  was calculated using a chi-square test.

# Supplementary Table 6. ENCODE regulatory regions overlapping with credible causal variants.

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rsid	Encode ID	Regulatory region_Tissue
chr1:230337180-	231337180, associated with hematuria	
Signal 1		
rs11122572	ENCFF940MJZ	Enhancer_caudate nucleus
rs4846866	ENCFF940MJZ	Enhancer_caudate nucleus
rs61762468	ENCFF940MJZ	Enhancer_caudate nucleus
rs56117713	ENCFF940MJZ	Enhancer_caudate nucleus
rs16852352	ENCFF131AMT, ENCFF345YBN, ENCFF458GST, ENCFF940MJZ	Enhancer_HepG2, Enhancer_HepG2, Enhancer_HepG2, Enhancer_caudate nucleus
rs11122573	ENCFF131AMT, ENCFF345YBN, ENCFF458GST, ENCFF940MJZ	nucleus
rs10864770	ENCFF131AMT, ENCFF345YBN, ENCFF458GST, ENCFF940MJZ	nucleus
rs10864771	ENCFF131AMT, ENCFF345YBN, ENCFF458GST, ENCFF940MJZ	nucleus
rs11122574	ENCFF458GST	Enhancer_HepG2
rs1926723	ENCFF458GST	Enhancer_HepG2
rs1926722	ENCFF458GST	Enhancer_HepG2
rs11122575 rs11568056	ENCFF131AMT, ENCFF345YBN, ENCFF357FRW, ENCFF458GST, ENCFF768HKR, ENCFF940MJZ ENCFF458GST, ENCFF529FRB, ENCFF603HRN, ENCFF768HKR,	Enhancer_HepG2, Enhancer_HepG2, Enhancer_layer of hippocampus, Enhancer_HepG2, Promoter_HepG2, Enhancer_caudate nucleus Enhancer_HepG2, Promoter_caudate nucleus, Promoter_liver, Promoter_HepG2,
1511506050	ENCFF856JWA, ENCFF925ARX, ENCFF940MJZ	Enhancer_liver, Promoter_HepG2, Enhancer_caudate nucleus
rs11122576	ENCFF021ERU, ENCFF131AMT, ENCFF345YBN, ENCFF458GST, ENCFF529FRB, ENCFF603HRN, ENCFF662TKM, ENCFF768HKR, ENCFF856JWA, ENCFF925ARX, ENCFF940MJZ	Enhancer_HepG2, Promoter_caudate nucleus, Promoter_liver,Promoter_heart lef ventricle, Promoter_HepG2, Enhancer_liver, Promoter_HepG2, Enhancer_caudate nucleus
rs11568028	ENCFF021ERU, ENCFF131AMT, ENCFF345YBN, ENCFF357FRW, ENCFF458GST, ENCFF529FRB, ENCFF603HRN, ENCFF662TKM, ENCFF768HKR, ENCFF856JWA, ENCFF925ARX, ENCFF940MJZ	Enhancer_HepG2, Enhancer_middle frontal area 46, Enhancer_HepG2, Enhancer_layer of hippocampus, Enhancer_HepG2, Promoter_caudate nucleus, Promoter_liver,Promoter_heart left ventricle, Promoter_HepG2,Enhancer_liver, Promoter_HepG2, Enhancer_caudate nucleus
rs11122578	<ul> <li>ENCFF021ERU, ENCFF026HMJ, ENCFF084LBA, ENCFF131AMT, ENCFF138BOV, ENCFF144PJO, ENCFF154VRY, ENCFF169ILY, ENCFF232DYU, ENCFF283LOL, ENCFF305UOC, ENCFF345YBN, ENCFF357FRW, ENCFF407NSM, ENCFF458GST,</li> <li>ENCFF468LNN, ENCFF518VHQ, ENCFF524SKZ, ENCFF529FRB, ENCFF602NDV, ENCFF603HRN, ENCFF608IMQ, ENCFF621WVX, ENCFF662TKM, ENCFF608IMQ, ENCFF690YPD, ENCFF699XIX, ENCFF768HKR, ENCFF798VKK, ENCFF809ZVL, ENCFF826LIL, ENCFF856JWA, ENCFF920NZB, ENCFF925ARX, ENCFF937LPQ, ENCFF940MJZ, ENCFF952DET, ENCFF955CNN, ENCFF978UAS</li> </ul>	<ul> <li>Enhancer_HepG2, Enhancer_large intestine, Enhancer_middle frontal area 46, Enhancer_myotube, Enhancer_substantia nigra,Enhancer_small intestine,</li> <li>Enhancer_skeletal muscle tissue, Enhancer_pancreas, Enhancer_small intestine,</li> <li>Enhancer_temporal lobe, Promoter_skeletal muscle tissue, Enhancer_IMR-90,</li> <li>Enhancer_HepG2, Enhancer_layer of hippocampus, Promoter_large intestine,</li> <li>Enhancer_heart left ventricle,Promoter_caudate nucleus, Enhancer_psoas muscle</li> <li>Promoter_liver, Promoter_skeletal muscle of trunk, Enhancer_angular gyrus,</li> <li>Promoter_heart left ventricle, Enhancer_muscle of trunk, Enhancer_psoas muscle</li> <li>Enhancer_myotube, Promoter_HepG2, Enhancer_right cardiac atrium,</li> <li>Enhancer_muscle of leg, Promoter_HepG2, Enhancer_liver,</li> <li>Enhancer_muscle of leg, Promoter_HepG2, Enhancer_large intestine,</li> </ul>
rs3789679	ENCFF212AGQ	Enhancer_fibroblast of lung
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rs9804147

rs9804153		
rs4028824		
rs10864773	ENCFF940MJZ	Enhancer_caudate nucleus
rs61762467	ENCFF033LLU, ENCFF154VRY,ENCFF447BWN, ENCFF875CFO, ENCFF937LPQ, ENCFF978UAS	Enhancer_large intestine, Enhancer_mucosa of rectum, Enhancer_pancreas,Enhancer_B cell, Enhancer_B cell, Enhancer_large intestine
Signal 2		
rs4846857	ENCFF518VHQ	Enhancer_cingulate gyrus
rs79434380		
rs564325629		
rs147121532	ENCFF043YPD	Enhancer_mucosa of rectum
rs28605378	ENCFF043YPD, ENCFF715DPV, ENCFF892DRC	Enhancer_mucosa of rectum,Enhancer_stomach,Enhancer_stomach
rs12095859	ENCFF013DHK, ENCFF033LLU, ENCFF043YPD, ENCFF084LBA, ENCFF183FMS, ENCFF412OXP, ENCFF519JQV, ENCFF605WXQ, ENCFF715DPV, ENCFF741KDK, ENCFF892DRC	Enhancer_MCF-7, Enhancer_mucosa of rectum, Enhancer_mucosa of rectum, Enhancer_substantia nigra, Enhancer_stomach, Promoter_stomach, Enhancer_colonic mucosa, Enhancer_stomach, Enhancer_stomach, Enhancer_sigmoid colon, Enhancer_stomach
rs12091328	ENCFF013DHK, ENCFF021ERU, ENCFF033LLU, ENCFF043YPD, ENCFF084LBA, ENCFF357FRW, ENCFF518VHQ, ENCFF621WVX, ENCFF715DPV, ENCFF741KDK, ENCFF892DRC	Enhancer_MCF-7,Enhancer_middle frontal area 46 ,Enhancer_mucosa of rectum,Enhancer_mucosa of rectum,Enhancer_substantia nigra,Enhancer_layer of hippocampus,Enhancer_cingulate gyrus,Enhancer_angular gyrus,Enhancer_stomach,Enhancer_sigmoid colon,Enhancer_stomach
rs12059171	ENCFF013DHK,ENCFF021ERU,ENCFF033LLU,ENCFF043YPD,E NCFF059PMT,ENCFF084LBA,ENCFF131AMT,ENCFF138BOV,EN CFF183FMS,ENCFF232DYU,ENCFF345YBN,ENCFF357FRW,EN CFF458GST,ENCFF518VHQ,ENCFF519JQV,ENCFF605WXQ,EN CFF621WVX,ENCFF715DPV,ENCFF741KDK,ENCFF892DRC,EN CFF937LPQ,ENCFF940MJZ	Enhancer_HepG2,Enhancer_MCF-7,Enhancer_middle frontal area 46 ,Enhancer_mucosa of rectum,Enhancer_mucosa of rectum,Enhancer_lung,Enhancer_substantia nigra,Enhancer_small intestine,Enhancer_stomach,Enhancer_temporal lobe,Enhancer_HepG2,Enhancer_layer of hippocampus,Enhancer_HepG2,Enhancer_cingulate gyrus,Enhancer_colonic mucosa,Enhancer_stomach,Enhancer_angular gyrus,Enhancer_stomach,Enhancer_sigmoid colon,Enhancer_stomach,Enhancer_cingulate nucleus
rs12060898		
rs75991123		
chr5:156903410-1	57903410, associated with rectal bleeding	
rs17055178		
rs13180537		
rs78394554	ENCFF031NTY,ENCFF242UFI	Enhancer_mesendoderm, Enhancer_mesendoderm
rs34395161	ENCFF031NTY,ENCFF242UFI	Enhancer_mesendoderm, Enhancer_mesendoderm
rs35327501	ENCFF937LPQ,ENCFF952DET,ENCFF953YED,ENCFF978UAS	Enhancer_large intestine, Enhancer_large intestine, Enhancer_adrenal gland, Enhancer_adrenal gland
rs4704767		
rs35929592		
rs35766682	ENCFF446JOJ,ENCFF952DET,ENCFF953YED	Enhancer_esophagus, Enhancer_adrenal gland, Enhancer_adrenal gland
rs10515757	ENCFF952DET, ENCFF953YED	Enhancer_adrenal gland, Enhancer_adrenal gland
rs35153425		
rs17055241		
rs1040926	ENCFF210ALH,ENCFF595MLU,ENCFF733LQH	Enhancer_keratinocyte, Enhancer_foreskin keratinocyte, Enhancer_keratinocyte

Abbreviations: -- indicates that a value was not available.

# **Supplementary Table 7.** Top-ranking genes (p < 0.001) associated with each toxicity outcome.

Toxicity by Location	Gene ID	Gene symbol	Gene Type	No. of SNPs	p-value*
Rectal bleeding					
chr16	100130958	SYCE1L	protein coding	369	3.22x10-4
chr16	22879	MON1B	protein coding	330	3.40x10-4
chr3	55096	EBLN2	protein coding	261	5.45x10-4
chr4	100874374	ARHGEF38-IT1	protein coding	188	5.72x10-4
chr2	5498	KIDINS220	protein coding	251	6.38x10-4
chr4	54848	ARHGEF38	protein coding	244	6.83x10-4
chr5	2559	GABRA6	protein coding	248	7.55x10-4
chr2	151254	ALS2CR11	protein coding	274	7.94x10-4
chr6	168002	DACT2	protein coding	439	8.27x10-4
chr3	151987	PPP4R2	protein coding	482	8.30x10-4
chr7	79571	GCC1	protein coding	126	9.90x10-4
ncreased urinary frequency					
chr14	145567	TTC7B	ncRNA	1031	9.37x10-5
chr14	9623	TCL1B	protein coding	418	1.27x10-4
chr20	63935	PCIF1	protein coding	162	1.86x10-4
chr15	9836	LCMT2	protein coding	154	3.16x10-4
chr14	27004	TCL6	ncRNA	465	3.24x10-4
chr20	5360	PLTP	protein coding	193	3.50x10-4
chr15	161823	ADAL	protein coding	179	3.73x10-4
chr12	8738	CRADD	protein coding	34	4.56x10-4
chr15	116179	TGM7	protein coding	166	4.75x10-4
chr19	284382	ACTL9	protein coding	229	5.58x10-4
chr15	146050	ZSCAN29	protein coding	169	6.46x10-4
chr22	266697	POM121L4P	pseudo	228	6.51x10-4
chr13	196541	METTL21C	protein coding	309	6.91x10-4
chr20	5476	CTSA	protein coding	178	7.05x10-4
chr22	84222	TMEM191A	pseudo	222	7.24x10-4
chr22	5297	PI4KA	protein coding	500	7.90x10-4
chr22	3053	SERPIND1	protein coding	228	8.42x10-4
chr15	27229	TUBGCP4	protein coding	214	9.36x10-4
	100505576	LINC00672	ncRNA	175	9.42x10-4
chr17	6452	SH3BP2	protein coding	274	9.76x10-4
chr4	140825	NEURL2	protein coding	168	9.92x10-4
chr20	225	ABCD2	protein coding	6	9.92×10-4
chr12	225	ADCDZ	protein county	0	9.99810-2
Decreased urinary stream	54925	ZSCAN32	protein coding	360	1.14x10-5
chr16	54925 7727	ZSCAN32 ZNF174	protein coding protein coding	360 307	1.14x10-5 1.20x10-5
chr16					
chr14	26257	NKX2-8	protein coding	253 202	1.24x10-5
chr16	100463285	MTRNR2L4	protein coding	292	2.00x10-5
chr16	4993	OR2C1	ncRNA	286	2.29x10-5
chr16	79903	NAA60	protein coding	357	6.89x10-5
chr16	146434	ZNF597	pseudo	337	6.95x10-5
chr16	7627	ZNF75A	protein coding	243	7.06x10-5
chr9	100616351	MIR4670	protein coding	139	8.25x10-5
chr15	8924	HERC2	protein coding	104	8.41x10-5

chr9	2649	NR6A1	protein coding	170	8.72x10-5
chr9	1842	ECM2	protein coding	186	9.69x10-5
chr9	2516	NR5A1	protein coding	125	1.19x10-4
chr14	4140	MARK3	protein coding	471	1.24x10-4
chr14	7080	NKX2-1	protein coding	164	1.28x10-4
chr9	347088	GPR144	protein coding	149	1.59x10-4
chr11	100616311	MIR3973	ncRNA	157	1.65x10-4
chr14	5083	PAX9	protein coding	268	1.67x10-4
chr2	3635	INPP5D	protein coding	689	1.76x10-4
chr14	253970	SFTA3	protein coding	220	1.89x10-4
chr9	54829	ASPN	protein coding	138	2.54x10-4
chr9	401541	CENPP	protein coding	389	2.68x10-4
chr9	100379345	MIR181A2HG	ncRNA	48	2.76x10-4
chr9	406954	MIR181A2	ncRNA	39	3.15x10-4
chr9	406956	MIR181B2	ncRNA	39	3.19x10-4
chr9	23511	NUP188	protein coding	117	4.64x10-4
chr9	56904	SH3GLB2	protein coding	70	4.71x10-4
chr14	1152	CKB	protein coding	210	4.76x10-4
	22845	DOLK	protein coding	78	5.84x10-4
chr9	4958	OMD	protein coding	110	5.88x10-4
chr9	728752	LOC728752	ncRNA	230	6.31x10-4
chr19		PHYHD1			
chr9	254295		protein coding	90	6.47x10-4
chr9	169611	OLFML2A	protein coding	161	6.90x10-4
chr3	401097	C3orf80	protein coding	105	7.15x10-4
chr3	255758	TCTEX1D2	protein coding	300	8.92x10-4
chr18	54808	DYM	protein coding	973	9.18x10-4
chr9	4969	OGN	protein coding	115	9.29x10-4
chr9	56262	LRRC8A	protein coding	106	9.49x10-4
Hematuria					
chr6	94120	SYTL3	protein coding	595	2.82x10-5
chr6	101409257	EZR-AS1	ncRNA	309	5.86x10-5
chr1	183	AGT	protein coding	370	7.91x10-5
chr6	202459	OSTCP1	pseudo	301	8.65x10-5
chr6	7430	EZR	protein coding	463	9.48x10-5
chr4	80144	FRAS1	protein coding	1570	1.19x10-4
chr6	100500851	MIR3918	ncRNA	333	1.38x10-4
chr20	650	BMP2	protein coding	227	2.76x10-4
chr1	22796 63901	COG2	protein coding	425 106	2.98x10-4 3.58x10-4
chr11 chr5	23037	FAM111A PDZD2	protein coding protein coding	1522	3.56x10-4 3.61x10-4
chr15	283726	FAM154B	protein coding	145	4.05x10-4
chr7	100288524	LOC100288524	ncRNA	486	4.38x10-4
chr10	55130	ARMC4	protein coding	492	5.27x10-4
chr11	374393	FAM111B	protein coding	131	5.54x10-4
chr1	10753	CAPN9	protein coding	486	6.46x10-4
chr15	390660	LOC390660	pseudo	94	6.86x10-4
chr8	286097	MICU3	protein coding	463	7.01x10-4
chr17	6426	SRSF1	protein coding	91	7.34x10-4
chr17	1277	COL1A1	protein coding	204	7.36x10-4
chr15	79631	EFTUD1	protein coding	413	7.38x10-4
chr11	23220	DTX4	protein coding	174	7.99x10-4

chr12 chr8	6579 100874052	SLCO1A2 LINC00534	protein coding ncRNA	31 509	8.53x10-4 8.65x10-4
chr15	161742	SPRED1	protein coding	557	9.56x10-4
chr12	10599	SLCO1B1	protein coding	26	9.80x10-4

\* <u>Two-sided</u>  $P_{meta}$  was calculated using a Wald test.

**Supplementary Table 8.** Pathway scores for top-ranking pathways (chi-square p < 0.05) associated with each toxicity outcome.

Toxicity	Pathway Name	chi2 pvalue <sup>*</sup>	empirical pvalue <sup>†</sup>
Rectal bleeding	REACTOME_PEROXISOMAL_LIPID_METABOLISM	6.04x10-4	6.70x10-4
Ũ	BIOCARTA_PYK2_PATHWAY	0.006	0.005
	REACTOME_GABA_RECEPTOR_ACTIVATION	0.009	0.007
	REACTOME_TRNA_AMINOACYLATION	0.01	0.01
	KEGG PEROXISOME	0.01	0.01
	BIOCARTA_GPCR_PATHWAY	0.01	0.01
	REACTOME_INWARDLY_RECTIFYING_K_CHANNELS	0.01	0.009
	BIOCARTA_INTEGRIN_PATHWAY	0.01	0.01
	BIOCARTA_CHEMICAL_PATHWAY	0.01	0.01
	BIOCARTA FMLP PATHWAY	0.01	0.01
	REACTOME_TRANSCRIPTIONAL_REGULATION_OF_WHITE_ADIPOCYTE_DIFFERENTIATION*	0.01	0.02
	REACTOME_CA_DEPENDENT_EVENTS	0.01	0.01
	BIOCARTA_AT1R_PATHWAY	0.01	0.01
	BIOCARTA_BCR_PATHWAY	0.02	0.01
	REACTOME_A_TETRASACCHARIDE_LINKER_SEQUENCE_IS_REQUIRED_FOR_GAG_SYNTHESIS	0.02	0.01
	BIOCARTA_UCALPAIN_PATHWAY <sup>†</sup>	0.02	0.02
	KEGG_GLYCOSPHINGOLIPID_BIOSYNTHESIS_GLOBO_SERIES	0.02	0.02
	REACTOME_OPIOID_SIGNALLING	0.02	0.02
	BIOCARTA_CBL_PATHWAY	0.02	0.02
	BIOCARTA_TNFR1_PATHWAY	0.02	0.02
	BIOCARTA_FCER1_PATHWAY	0.02	0.02
	BIOCARTA ECM PATHWAY <sup>‡</sup>	0.03	0.03
	REACTOME_CIRCADIAN_REPRESSION_OF_EXPRESSION_BY_REV_ERBA	0.03	0.03
	REACTOME_ARMS_MEDIATED_ACTIVATION	0.03	0.04
	REACTOME_CYTOSOLIC_TRNA_AMINOACYLATION	0.03	0.03
	REACTOME_MEIOTIC_RECOMBINATION	0.03	0.03
	BIOCARTA FAS PATHWAY	0.03	0.03
	REACTOME_GABA_B_RECEPTOR_ACTIVATION	0.03	0.03
	BIOCARTA_IL3_PATHWAY	0.03	0.03
	KEGG_TYPE_II_DIABETES_MELLITUS <sup>†</sup>	0.03	0.03
	BIOCARTA_TCR_PATHWAY	0.03	0.03
	REACTOME_PLC_BETA_MEDIATED_EVENTS	0.03	0.03
	REACTOME_GLYCOLYSIS	0.03	0.03
	KEGG_NATURAL_KILLER_CELL_MEDIATED_CYTOTOXICITY	0.03	0.03
	BIOCARTA_AGR_PATHWAY	0.03	0.03
	BIOCARTA_NUCLEARRS_PATHWAY	0.04	0.03
	REACTOME_PROSTACYCLIN_SIGNALLING_THROUGH_PROSTACYCLIN_RECEPTOR	0.04	0.03
	KEGG_CARDIAC_MUSCLE_CONTRACTION	0.04	0.04
	KEGG_AMINOACYL_TRNA_BIOSYNTHESIS	0.04	0.04

Rectal bleeding	REACTOME_RORA_ACTIVATES_CIRCADIAN_EXPRESSION	0.04	0.05
	BIOCARTA_MCALPAIN_PATHWAY	0.04	0.04
	REACTOME_DAG_AND_IP3_SIGNALING	0.04	0.04
	KEGG_P53_SIGNALING_PATHWAY	0.04	0.04
	BIOCARTA_NDKDYNAMIN_PATHWAY <sup>‡</sup>	0.04	0.04
	REACTOME_ERKS_ARE_INACTIVATED	0.04	0.04
	REACTOME_FACILITATIVE_NA_INDEPENDENT_GLUCOSE_TRANSPORTERS	0.04	0.04
	BIOCARTA_NOS1_PATHWAY	0.04	0.04
	BIOCARTA_RACCYCD_PATHWAY	0.04	0.04
	BIOCARTA_GABA_PATHWAY	0.04	0.05
	REACTOME_INHIBITION_OF_VOLTAGE_GATED_CA2_CHANNELS_VIA_GBETA_GAMMA_SUBUNITS	0.04	0.04
	BIOCARTA_THELPER_PATHWAY	0.04	0.05
	BIOCARTA_DREAM_PATHWAY	0.05	0.04
	BIOCARTA_CERAMIDE_PATHWAY	0.05	0.05
	BIOCARTA_FREE_PATHWAY	0.05	0.04
	REACTOME_GLUCAGON_SIGNALING_IN_METABOLIC_REGULATION	0.05	0.05
	BIOCARTA_EGFR_SMRTE_PATHWAY	0.05	0.05
	REACTOME_BINDING_AND_ENTRY_OF_HIV_VIRION	0.05	0.05
	REACTOME_MITOCHONDRIAL_TRNA_AMINOACYLATION	0.05	0.05
	BIOCARTA_NFAT_PATHWAY <sup>†</sup>	0.05	0.05
Increased	REACTOME_PURINE_METABOLISM	0.001	0.001
urinary	REACTOME_BASIGIN_INTERACTIONS <sup>§</sup>	0.005	0.004
frequency	REACTOME_PURINE_CATABOLISM	0.006	0.005
	REACTOME_APOPTOSIS_INDUCED_DNA_FRAGMENTATION	0.009	0.009
	KEGG_SELENOAMINO_ACID_METABOLISM	0.01	0.01
	REACTOME_VOLTAGE_GATED_POTASSIUM_CHANNELS	0.01	0.01
	REACTOME_STRIATED_MUSCLE_CONTRACTION	0.02	0.01
	REACTOME_ABACAVIR_TRANSPORT_AND_METABOLISM	0.02	0.02
	REACTOME_TRANSCRIPTIONAL_REGULATION_OF_WHITE_ADIPOCYTE_DIFFERENTIATION*	0.02	0.02
	REACTOME_PURINE_SALVAGE	0.02	0.02
	REACTOME_CELL_SURFACE_INTERACTIONS_AT_THE_VASCULAR_WALL	0.02	0.02
	REACTOME_ETHANOL_OXIDATION	0.02	0.02
	REACTOME CD28 DEPENDENT VAV1 PATHWAY	0.02	0.02
	REACTOME_ALPHA_LINOLENIC_ACID_ALA_METABOLISM	0.02	0.03
	KEGG_PURINE_METABOLISM	0.03	0.02
	KEGG_INOSITOL_PHOSPHATE_METABOLISM	0.03	0.03
	KEGG VIRAL MYOCARDITIS	0.03	0.03
Increased	REACTOME_N_GLYCAN_TRIMMING_IN_THE_ER_AND_CALNEXIN_CALRETICULIN_CYCLE	0.03	0.03
urinary	REACTOME_SIGNAL_TRANSDUCTION_BY_L1	0.03	0.03
frequency	KEGG_RETINOL_METABOLISM	0.03	0.03
	REACTOME_CALNEXIN_CALRETICULIN_CYCLE	0.04	0.04
	REACTOME_HDL_MEDIATED_LIPID_TRANSPORT	0.04	0.04
	REACTOME CS DS DEGRADATION	0.04	0.03
			0.00

	DIGGADTA DNAFDAGNENT DATI WAANA	0.04	0.04
		0.04	0.04
		0.04	0.04
	REACTOME_COPI_MEDIATED_TRANSPORT	0.04	0.04
	BIOCARTA_CDK5_PATHWAY	0.04	0.05
	KEGG_RENIN_ANGIOTENSIN_SYSTEM	0.05	0.05
Decreased	REACTOME_CHROMOSOME_MAINTENANCE	0.005	0.005
urinary stream	REACTOME_CHOLESTEROL_BIOSYNTHESIS	0.006	0.009
	REACTOME_CASPASE_MEDIATED_CLEAVAGE_OF_CYTOSKELETAL_PROTEINS	0.01	0.01
	REACTOME_MITOTIC_PROMETAPHASE	0.01	0.009
	REACTOME_MEIOTIC_SYNAPSIS	0.01	0.02
	REACTOME_XENOBIOTICS	0.01	0.02
	BIOCARTA_CDMAC_PATHWAY	0.01	0.02
	REACTOME_MITOTIC_M_M_G1_PHASES	0.02	0.01
	BIOCARTA_BIOPEPTIDES_PATHWAY <sup>II</sup>	0.02	0.02
	REACTOME_CELL_CYCLE	0.02	0.02
	REACTOME_DNA_REPLICATION	0.02	0.02
	REACTOME_DEPOSITION_OF_NEW_CENPA_CONTAINING_NUCLEOSOMES_AT_THE_CENTROMERE	0.02	0.01
	REACTOME_OLFACTORY_SIGNALING_PATHWAY	0.02	0.01
	REACTOME_CELL_CYCLE_MITOTIC	0.03	0.03
	REACTOME_RAF_MAP_KINASE_CASCADE	0.03	0.04
	REACTOME SYNTHESIS OF PC	0.03	0.03
	BIOCARTA_ECM_PATHWAY <sup>‡</sup>	0.03	0.04
	KEGG FC EPSILON RI SIGNALING PATHWAY	0.04	0.03
	KEGG FC GAMMA R MEDIATED PHAGOCYTOSIS	0.04	0.03
	REACTOME_CYTOKINE_SIGNALING_IN_IMMUNE_SYSTEM <sup>II</sup>	0.04	0.04
	BIOCARTA_NDKDYNAMIN_PATHWAY <sup>‡</sup>	0.04	0.04
	KEGG_MELANOGENESIS	0.04	0.05
	REACTOME_SHC_RELATED_EVENTS	0.04	0.05
	REACTOME_TRANSPORT_OF_RIBONUCLEOPROTEINS_INTO_THE_HOST_NUCLEUS	0.05	0.04
	REACTOME_AMINE_LIGAND_BINDING_RECEPTORS	0.05	0.05
	REACTOME_SIGNALING_BY_ILS	0.05	0.05
	BIOCARTA_RHO_PATHWAY	0.05	0.05
	REACTOME_METABOLISM_OF_RNA	0.05	0.05
Hematuria			
Hematuna	REACTOME_PLATELET_ADHESION_TO_EXPOSED_COLLAGEN	0.002	0.002
	REACTOME_INTERFERON_ALPHA_BETA_SIGNALING	0.004	0.005
		0.007	0.005
	REACTOME_L1CAM_INTERACTIONS	0.008	0.005
	BIOCARTA_ALK_PATHWAY	0.008	0.008
	REACTOME_REGULATION_OF_IFNA_SIGNALING	0.009	0.009
	REACTOME_ACYL_CHAIN_REMODELLING_OF_PI	0.01	0.01
	BIOCARTA_UCALPAIN_PATHWAY <sup>†</sup>	0.01	0.007
	REACTOME_RECYCLING_PATHWAY_OF_L1	0.01	0.008
	BIOCARTA_ACE2_PATHWAY	0.02	0.01
	KEGG_TYPE_II_DIABETES_MELLITUS <sup>†</sup>	0.02	0.02

REACTOME_ACYL_CHAIN_REMODELLING_OF_PG	0.02	0.02
REACTOME_REGULATION_OF_BETA_CELL_DEVELOPMENT	0.02	0.02
REACTOME_ACTIVATION_OF_BH3_ONLY_PROTEINS	0.02	0.02
REACTOME_REGULATION_OF_IFNG_SIGNALING	0.02	0.02
KEGG_JAK_STAT_SIGNALING_PATHWAY	0.02	0.02
REACTOME_GROWTH_HORMONE_RECEPTOR_SIGNALING	0.02	0.02
REACTOME_ACTIVATED_NOTCH1_TRANSMITS_SIGNAL_TO_THE_NUCLEUS	0.03	0.02
BIOCARTA_NFAT_PATHWAY <sup>†</sup>	0.03	0.02
KEGG_VASCULAR_SMOOTH_MUSCLE_CONTRACTION	0.03	0.03
REACTOME_SIGNALING_BY_ACTIVATED_POINT_MUTANTS_OF_FGFR1	0.03	0.03
REACTOME_RNA_POL_I_TRANSCRIPTION	0.03	0.03
REACTOME_REGULATION_OF_GENE_EXPRESSION_IN_BETA_CELLS	0.04	0.04
REACTOME_SYNTHESIS_OF_PA	0.04	0.04
REACTOME_TRANSPORT_OF_ORGANIC_ANIONS	0.04	0.04
REACTOME_E2F_ENABLED_INHIBITION_OF_PRE_REPLICATION_COMPLEX_FORMA	TION 0.04	0.05
REACTOME_AXON_GUIDANCE	0.04	0.03
REACTOME_RNA_POL_I_TRANSCRIPTION_INITIATION	0.04	0.04
REACTOME_BASIGIN_INTERACTIONS <sup>§</sup>	0.04	0.04
BIOCARTA_CARDIACEGF_PATHWAY	0.05	0.03
REACTOME_SIGNALING_BY_BMP	0.05	0.04
BIOCARTA_PTC1_PATHWAY	0.05	0.05
REACTOME_ACYL_CHAIN_REMODELLING_OF_PS	0.05	0.05
REACTOME_INTRINSIC_PATHWAY_FOR_APOPTOSIS	0.05	0.05
REACTOME_CYTOKINE_SIGNALING_IN_IMMUNE_SYSTEM <sup>II</sup>	0.05	0.05
han anna an Iorda ta dhachan a shi' annana ta st		

 REACTOME\_CYTOKINE\_SIGNALING\_IN\_IMMUNE\_SYSTEM\*
 0.05

 Two-sided pvalue was calculated using a chi-square test.

 \* Following methods in [23], a Monte Carlo estimate of the p-value is obtained by sampling random gene sets of size and calculating the fraction of sets reaching a higher score than gene set of the given pathway.

 \* Pathway is associated with both rectal bleeding and increased urinary frequency

 § Pathway is associated with both rectal bleeding and decreased urinary stream

 II Pathway is associated with both increased urinary frequency and hematuria

 # Pathway is associated with both increased urinary frequency and hematuria

 # Pathway is associated with both increased urinary frequency and hematuria

Supplementary Table 9. Evaluation of radiotoxicity risk variants in Japanese cohorts treated with either external beam photon therapy (PRRGphoton), permanent seed brachytherapy with or without external beam photon therapy (NTMC) or carbon ion therapy (PRRG-Cion).

			Effect size (95% CI)				
SNP	Toxicity Outcome	Info <sup>*</sup>	In NTMC (N=254)	In PRRG-photon (N=170)	In PRRG-Cion (N=538)		
rs17055178 chr5:157,403,410 <sup>†</sup> MAF <sup>‡</sup> 0.07	Time to onset of grade 2+ rectal bleeding	0.98	NA <sup>§</sup>	NA <sup>§</sup>	NA <sup>§</sup>		
rs10969913 chr9:30,866,808 <sup>b</sup> MAF <sup>‡</sup> 0.37	Time to onset of grade 2+ decreased urinary stream <sup>II</sup>	0.98	HR = 1.15 (0.82 to 1.61)	HR = 1.83 (0.44 to 7.68)	HR = 0.90 (0.36 to 2.26)		
rs11122573 chr1:230,837,180 <sup>†</sup> MAF <sup>‡</sup> 0.31	Time to onset of grade 2+ hematuria <sup>¶</sup>	0.99	0.99 NA <sup>#</sup> NA <sup>**</sup>		HR = 1.18 (0.53 to 2.61)		
rs147121532 chr1: 230,451,849 <sup>†</sup> MAF <sup>‡</sup> 0.09	Time to onset of grade 2+ hematuria <sup>¶</sup>	0.85	NA <sup>#</sup>	NA	HR = 1.52 (0.48 to 4.84)		
rs17599026 chr5:137,763,798 <sup>†</sup> MAF <sup>‡</sup> 0.11	Presence of grade 1+ increased urinary frequency at 2 years after radiotherapy <sup>††</sup>	0.84	OR = 1.13 (0.49 to 2.64)	OR = 1.51 (0.56 to 4.05)	OR = 0.63 (0.27 to 1.49)		
rs7720298 chr5:13,858,328 <sup>†</sup> MAF <sup>‡</sup> 0.20	Presence of grade 1+ decreased urinary stream at 2 years after radiotherapy <sup>‡‡</sup>	0.97	OR = 1.42 (0.77 to 2.63)	NA <sup>§§</sup>	NA <sup>§§</sup>		
rs1801516 chr11:108,175,462 <sup>†</sup> MAF <sup>‡</sup> 0.00	Overall toxicity	NA <sup>IIII</sup>	NA <sup>IIII</sup>	NA <sup>IIII</sup>	NA <sup>IIII</sup>		
rs7582141 chr2:159,899,489 <sup>†</sup> MAF <sup>‡</sup> 0.12	Overall toxicity <sup>¶¶</sup>	0.99	OR = 1.07 (0.53 to 2.16)	OR = 0.97 (0.79 to 1.18)	OR = 0.95 (0.90 to 1.01)		

Imputation info score values are from the oncoarray. Abbreviations: SNP, single nucleotide polymorphism; CI, confidence interval; MAF, minor allele frequency; HR, hazard ratio; OR, odds ratio; NA, not analyzed.

<sup>†</sup> Base position according to Genome Reference Consortium Human Build 37 (hg19).

<sup>‡</sup> Minor allele frequency from PRACTICAL Oncoarray samples of Japanese ancestry

 <sup>§</sup> Rectal bleeding was not assessed in NTMC or PRRG
 <sup>II</sup> 82 out of 262 individuals in NTMC,5 out of 170 individuals in PRRG-photon, and 11 out of 538 individuals in PRRG-Cion developed grade 2+ decreased urinary stream.

<sup>1</sup> 16 out of 538 individuals in PRRG-Cion developed grade 2+ hematuria. <sup>#</sup> Hematuria was not assessed in NTMC.

No individuals in PRRG-photon reported grade 2 or worse toxicity and so this outcome was not analyzed.

<sup>++</sup> 75 out of 198 individuals in NTMC, 35 out of 98 individuals in PRRG-photon, and 49 out of 414 individuals in PRRG-Cion developed grade 1+ increased urinary frequency at 2 years after radiotherapy. <sup>#</sup> 58 out of 180 individuals in NTMC developed grade 1+ decreased urinary stream at 2 years after radiotherapy.

<sup>§§</sup> Only 2 individuals in PRRG-photon and no individuals in PRRG-Cion developed grade 1+ decreased urinary stream at 2 years after radiotherapy and so this outcome was not analyzed.

SNP is monomorphic in the Japanese population and was not analyzable in NTMC or PRRG.

<sup>¶</sup> Overall toxicity was measured using STAT score [26] and dichotomized by comparing those individuals with a STAT score greater than or equal to one standard deviation above the mean to individuals with a STAT score less than one standard deviation above the mean.

Supplementary Table 10. C-statistics representing model performance comparing models with only clinical

	C-statistic						
Toxicity outcome	Model only clinical factors	Model with clinical factors and SNP(s)					
Increased urinary frequency	0.563	0.567					
Decreased urinary stream	0.561	0.575					
Hematuria	0.578	0.617					
Rectal bleeding	0.603	0.621					

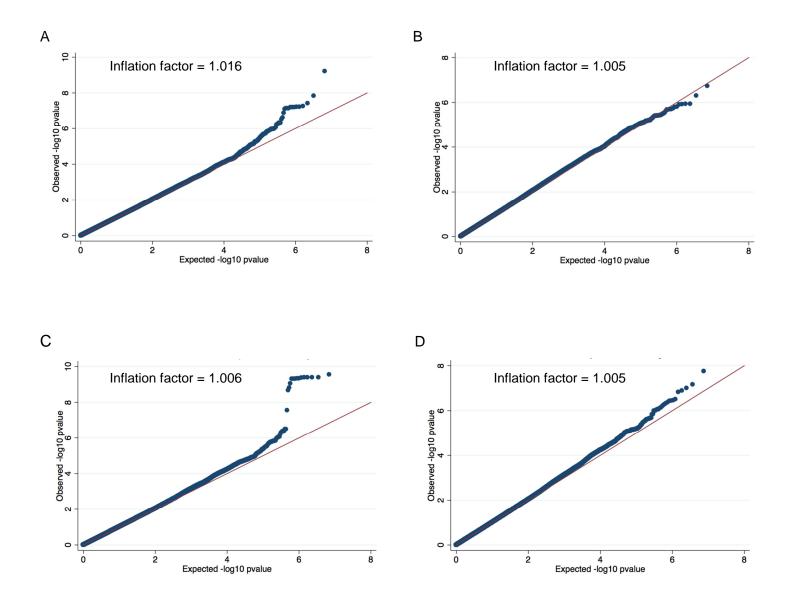
factors to models including clinical factors and SNPs

	RAPPER-I N=658	RAPPER-II N=1,947	RADIOGEN N=677	GenePARE-I N=382	GenePARE-II N=338	UGhent N=317	CCI-BT N=258	CCI-EBRT N=155	PRRG- photon N=172	PRRG- Cion N=545	NTMC N=256
Removed: poor quality DNA	12	35	0	30	22	0	0	1	1	7	1
Removed: included in batch I	NA	339	NA	NA	4	NA	NA	NA	NA	NA	NA
Removed: Ancestry <sup>*</sup>	39	55	1	91	62	1	3	3	0	0	0
Removed: lacking covariate data⁺ ✔	9	6	18	5	14	5	3	3	1	0	1
Included in analysis	N=598	N=1,512	N=658	N=256	N=236	N=311	N=252	N=148	N=170	N=538	N=254

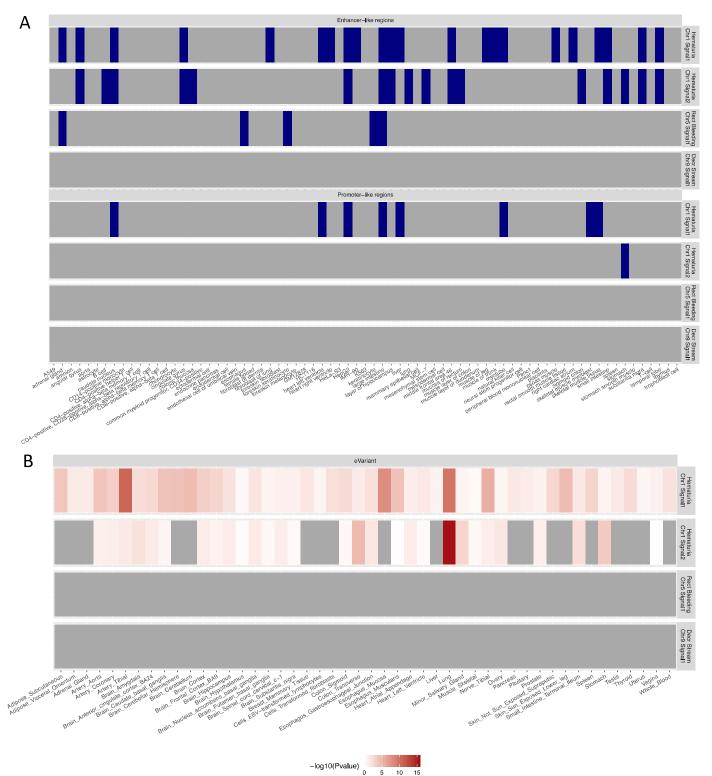
Supplementary Figure 1. Study participants. N=4,833 included in analysis (N=3,871 from European cohorts; N=962 from Japanese cohorts).

\*Non-European ancestry samples were removed from RAPPER, RADIOGEN, GenePARE, UGhent, CCI-BT and CCI-EBRT. Non-Japanese ancestry samples were removed from NTMC and PRRG. Abbreviations: NA, not applicable.

<sup>†</sup>Covariates included in the GWAS meta-analysis are age, total BED, prior prostatectomy, and receipt of androgen deprivation therapy.



**Supplementary Figure 2.** QQ plots. The plots show expected and observed p-values from GWAS metaanalysis of rectal bleeding (A), increased urinary frequency (B), decreased urinary stream (C), and hematuria (D).



**Supplementary Figure 3**. Mapping of credible causal variants (CCVs). In panel A, CCVs overlap with regulatory regions, enhancer- and promoter-like according to ENCODE. X axis: cell lines or tissues. Y axis: independent signals. Top graph shows enhancer-like regions. Bottom graph shows promoter-like regions. Blue: at least one CCV overlap a regulatory region active in the specific cell-line or tissue. Dark grey: any CCV overlap an active regulatory region. Panel B shows co-localization of CCVs with variants driving the expression of a particular transcript according to GTEX. X axis: tissues. Y axis: independent signals. Red, most significant expression p-value out of all CCVs at the signal and all evaluated transcripts for that tissue. Dark grey, no significant variants driving the expression of any transcript in the evaluated tissues.