

Meroterpene-like α -glucosidase inhibitors based on biomimic reactions staring from β -caryophyllene

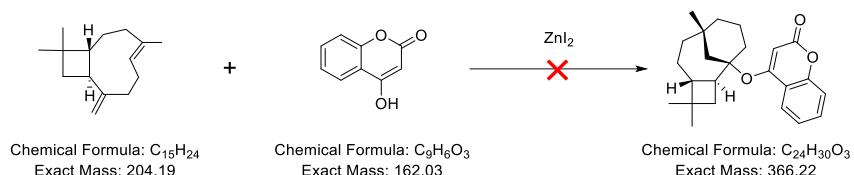
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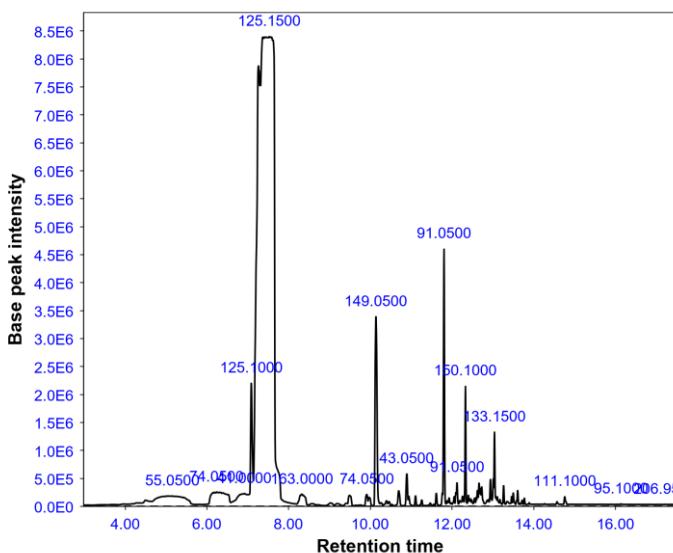
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Figure S1. GC-EI MS (a) and LC-ESI-MS/MS analysis

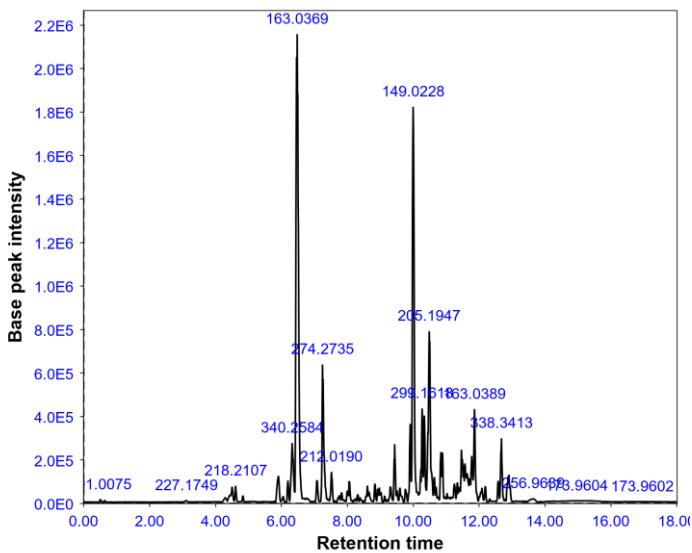


(a) GC-EI-MS analysis



The crude product was dissolved in MeOH (200 µg/ml) and was analyzed on a Shimadzu GCMS-QP2010 Ultra instrument equipped with a capillary column of RxI® -5Sil MS (30 m × 0.25 mm i.d., 0.25 µm film thickness). Helium (He) gas was used as a mobile phase at a constant flow of 1.3 mL/min. The temperature was held at 150 °C for 5 min, then increased to 300 at 4 °C/min, maintaining the last temperature for 5 min. The temperature of the ion source was 250 °C. EI-MS spectra (m/z 35 ~ 500) were recorded with a scan event time of 0.3 s. Electron energy was 70 eV. The raw data was saved into netCDF format in the GCMS solution software and then was converted into mzXML format by ProteoWizard MSConvert 3. The TIC chromatography was extracted using MZMine2.

(b) LC-QTOF-MS/MS analysis

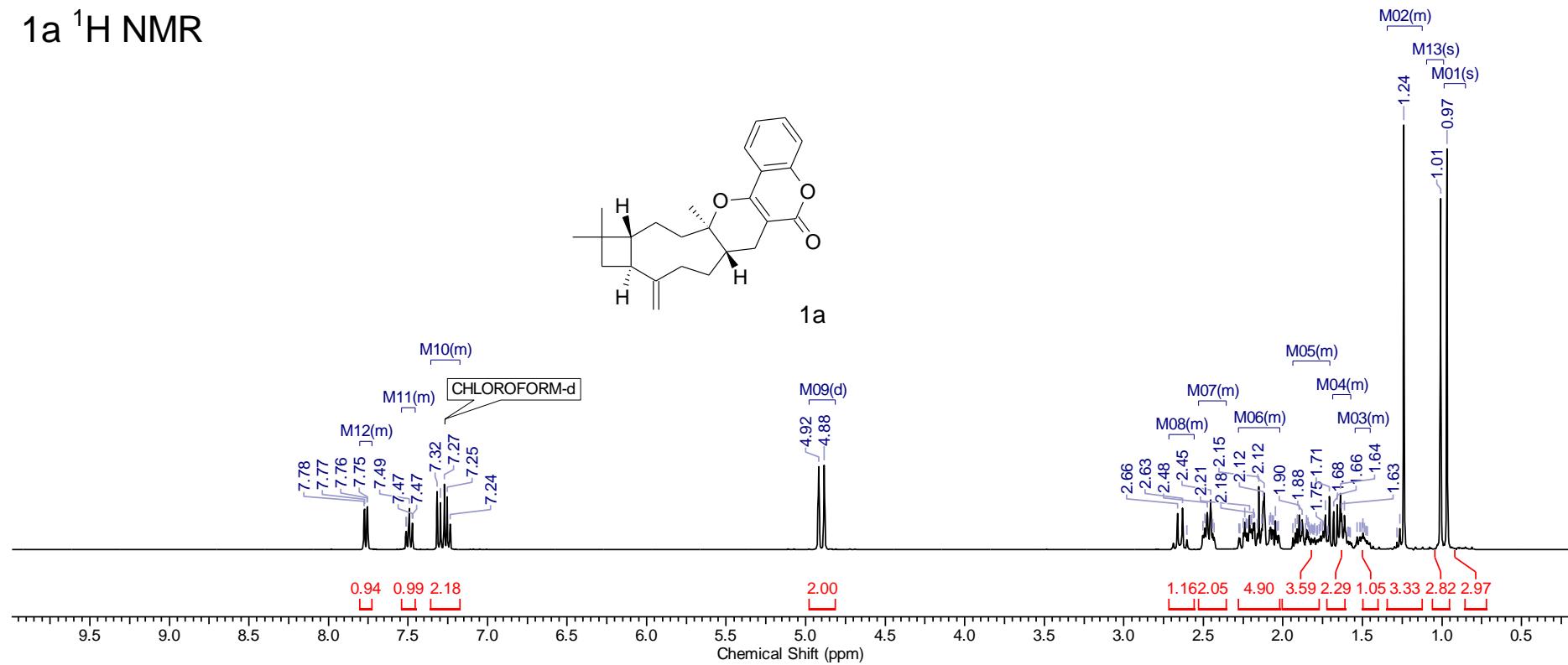


The crude product was analyzed by AB SCIEX TripleTOF5600+ equipped with a Shimadzu UPLC (LC-30A) Shim-pack XR-ODS (100 mm × 2.1 mm, 1.7 μ m) column. The temperature was maintained at 30 ° C. Mixtures of H₂O (A), and MeOH (B) were eluted at the flow rate of 0.3 mL/min with a linear gradient of 10 ~ 100% B (0 – 17 min). The samples (200 μ g/mL in MeOH, 2 μ L portions were injected) were analyzed in data-dependent acquisition (DDA) mode. The full MS survey scan was performed for 150 ms in the range of *m/z* 100 ~ 2000; The collision energy was set as 40 eV. The raw data were converted to the mzXML format with MSConvert 3.0. The TIC chromatography was extracted using MZMine2 with *m/z* range 100~1500.

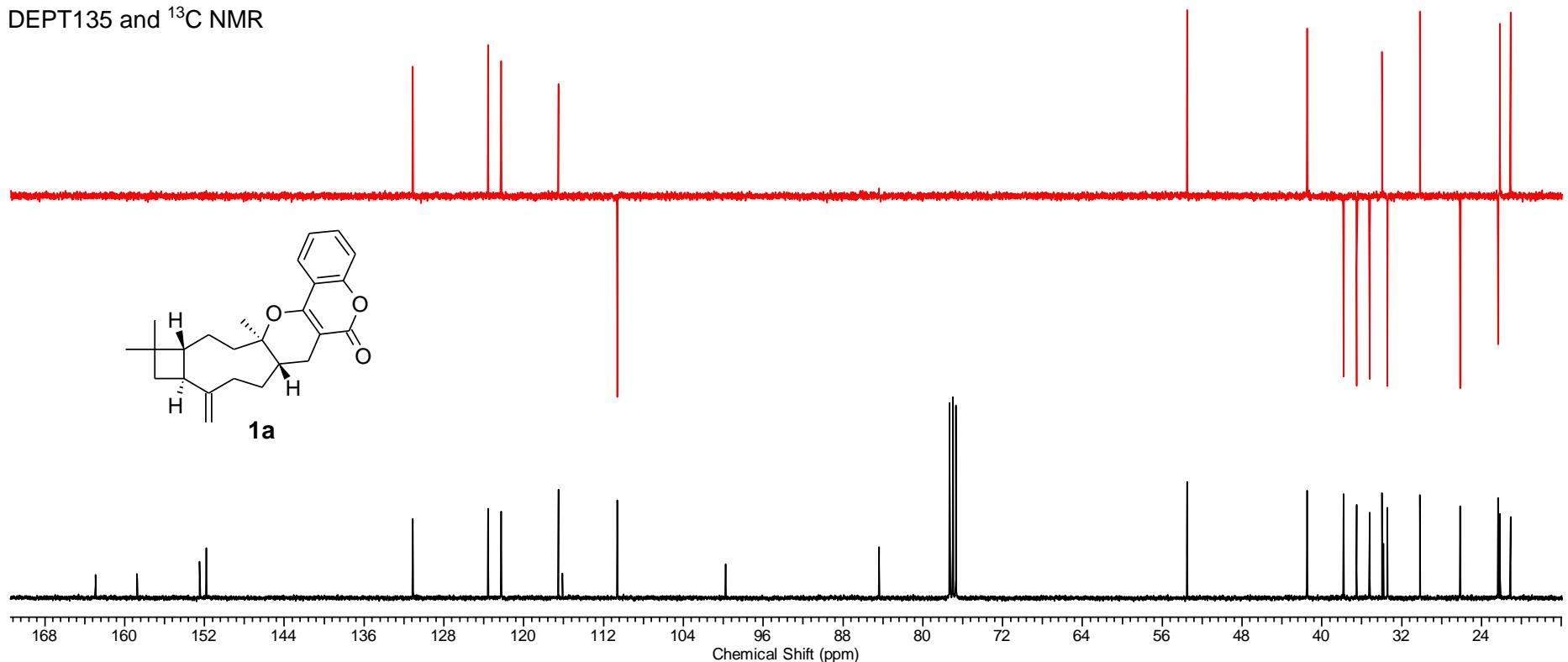
Figure S2: NMR and HR-MS spectra

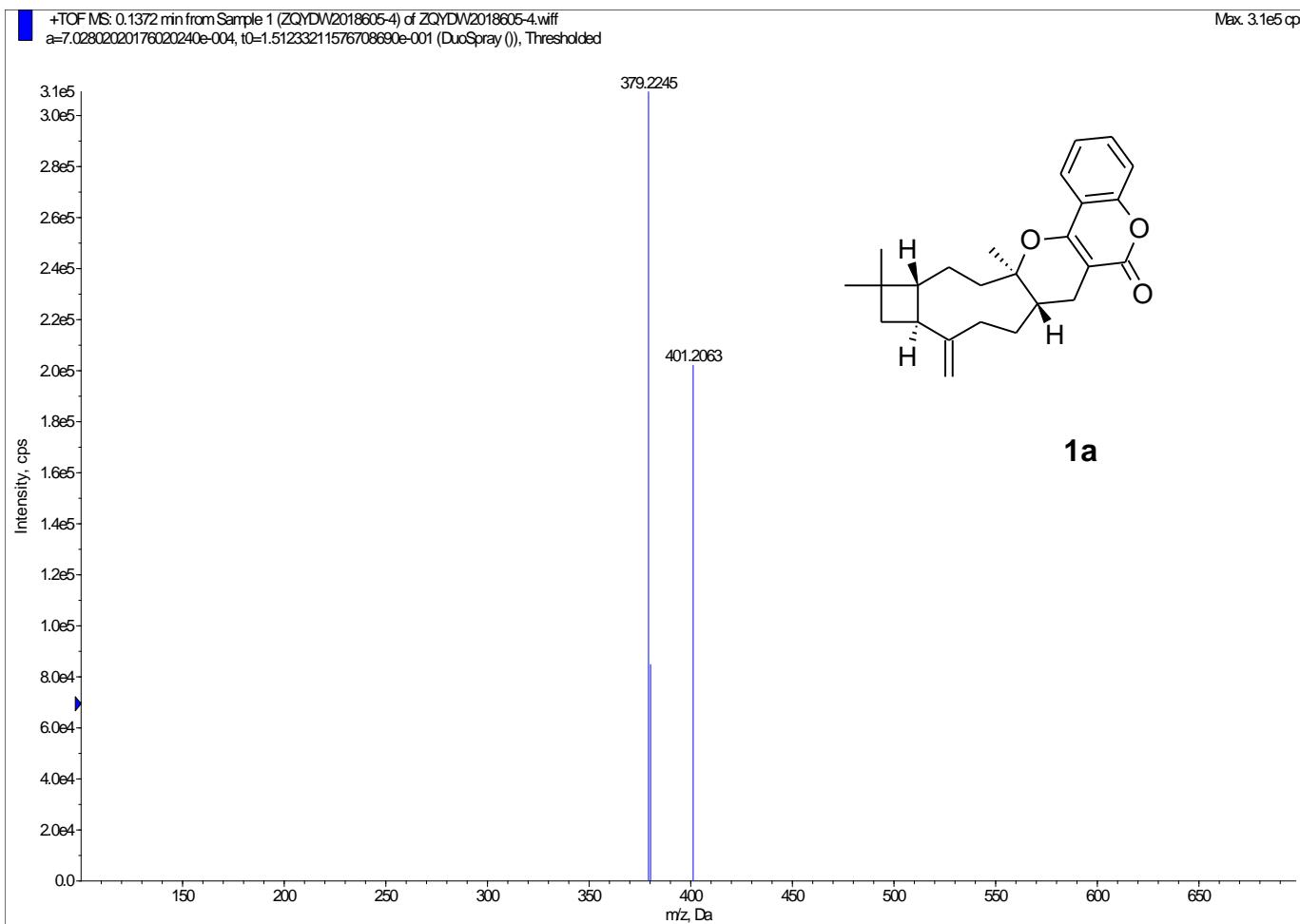
Compound 1a

1a ^1H NMR



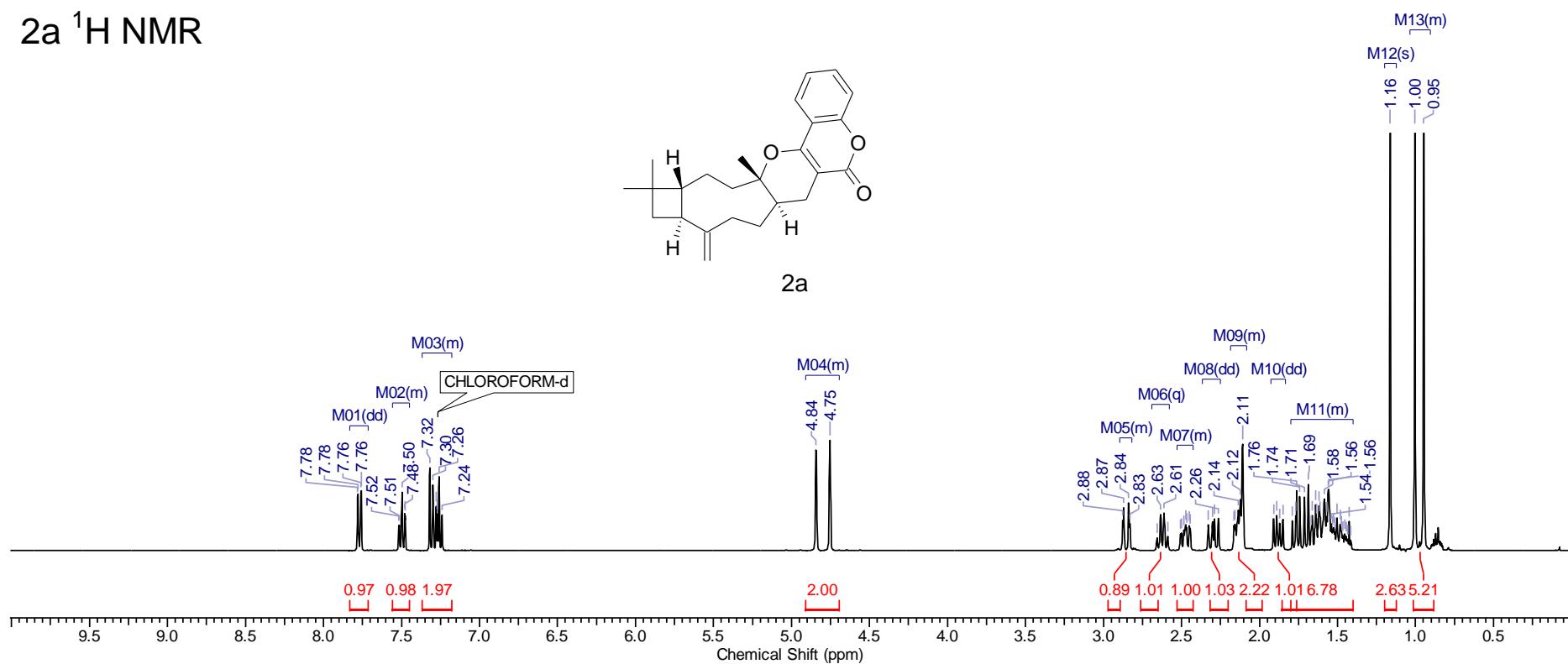
DEPT135 and ^{13}C NMR



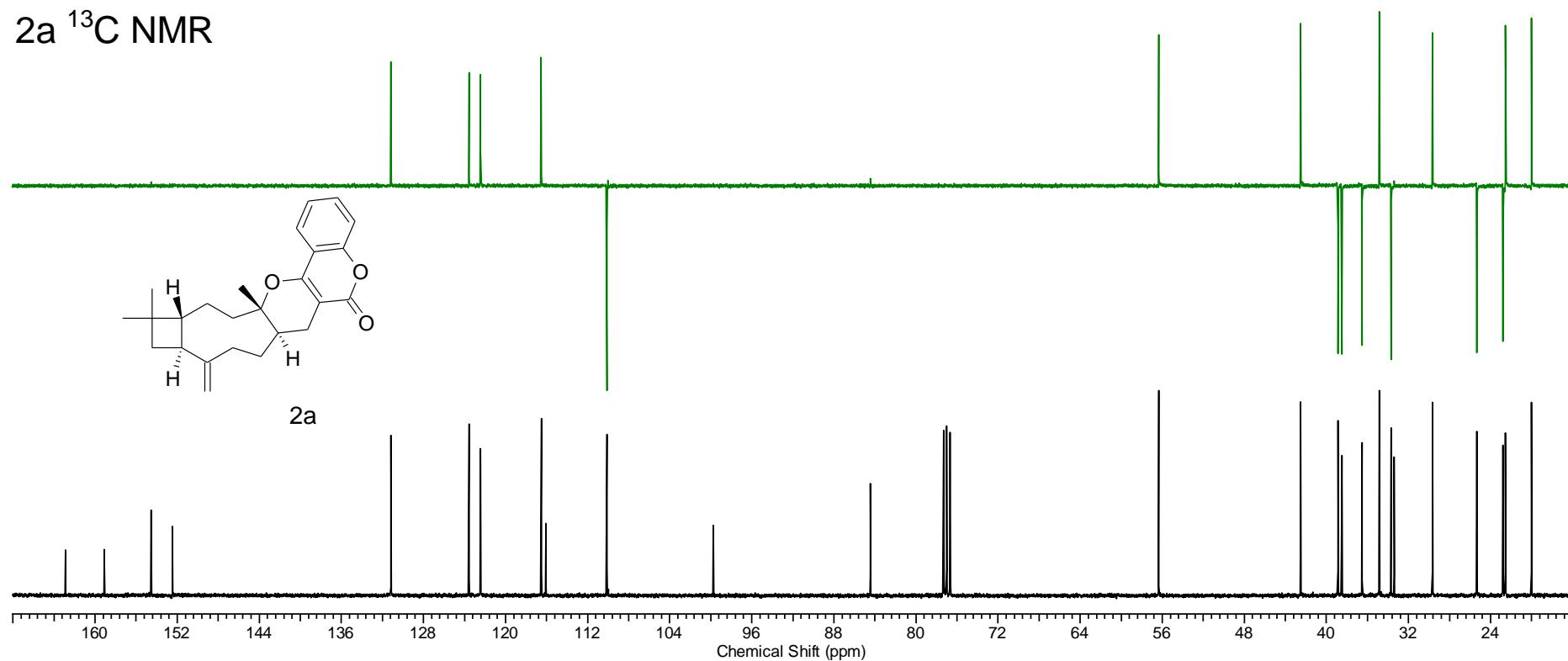


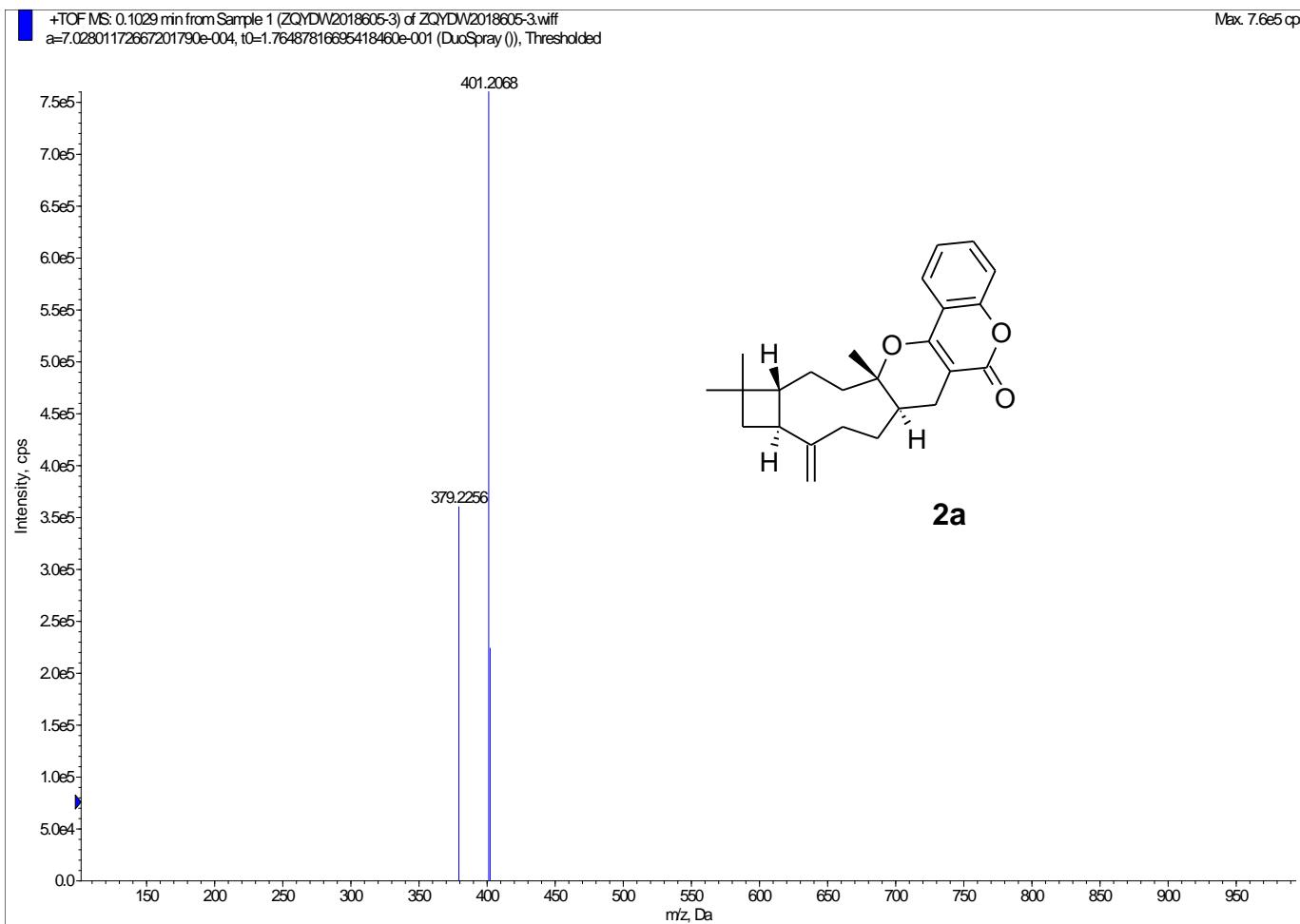
Compound 2a

2a ^1H NMR



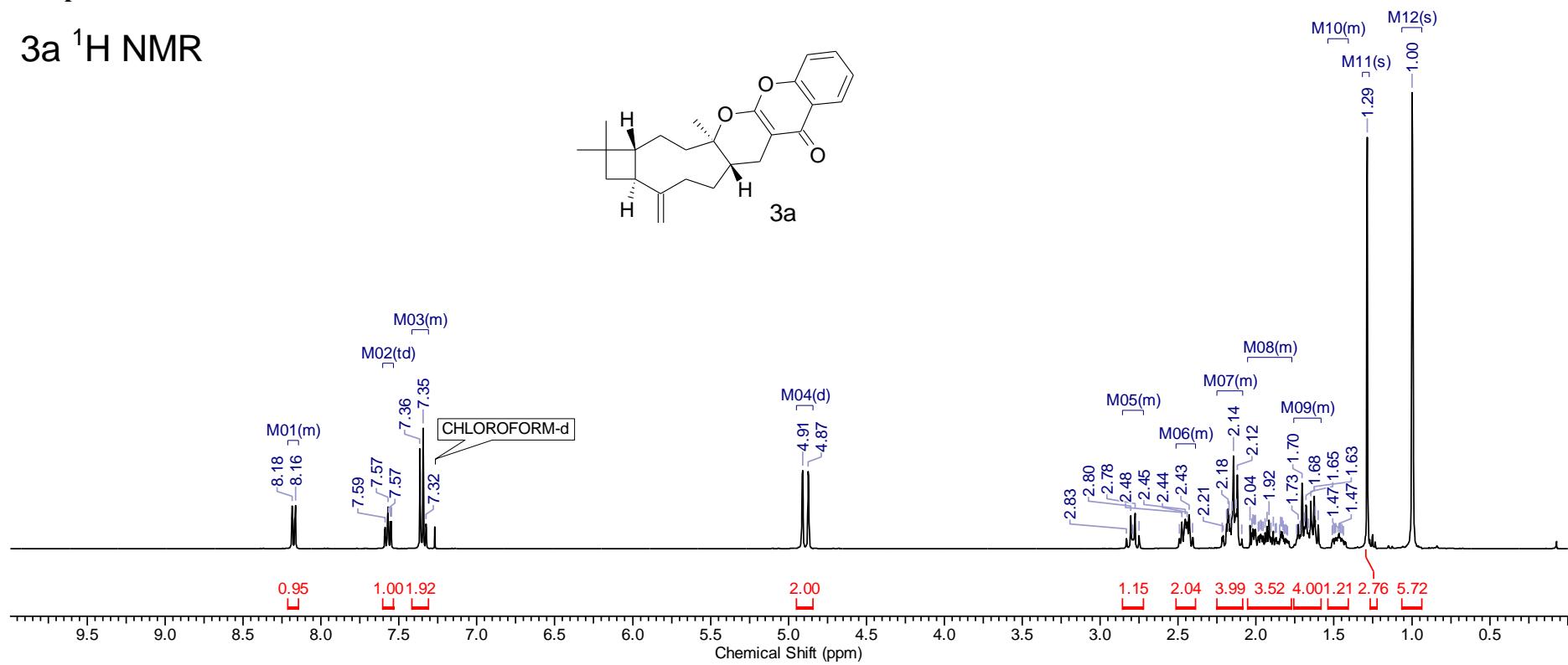
2a ^{13}C NMR



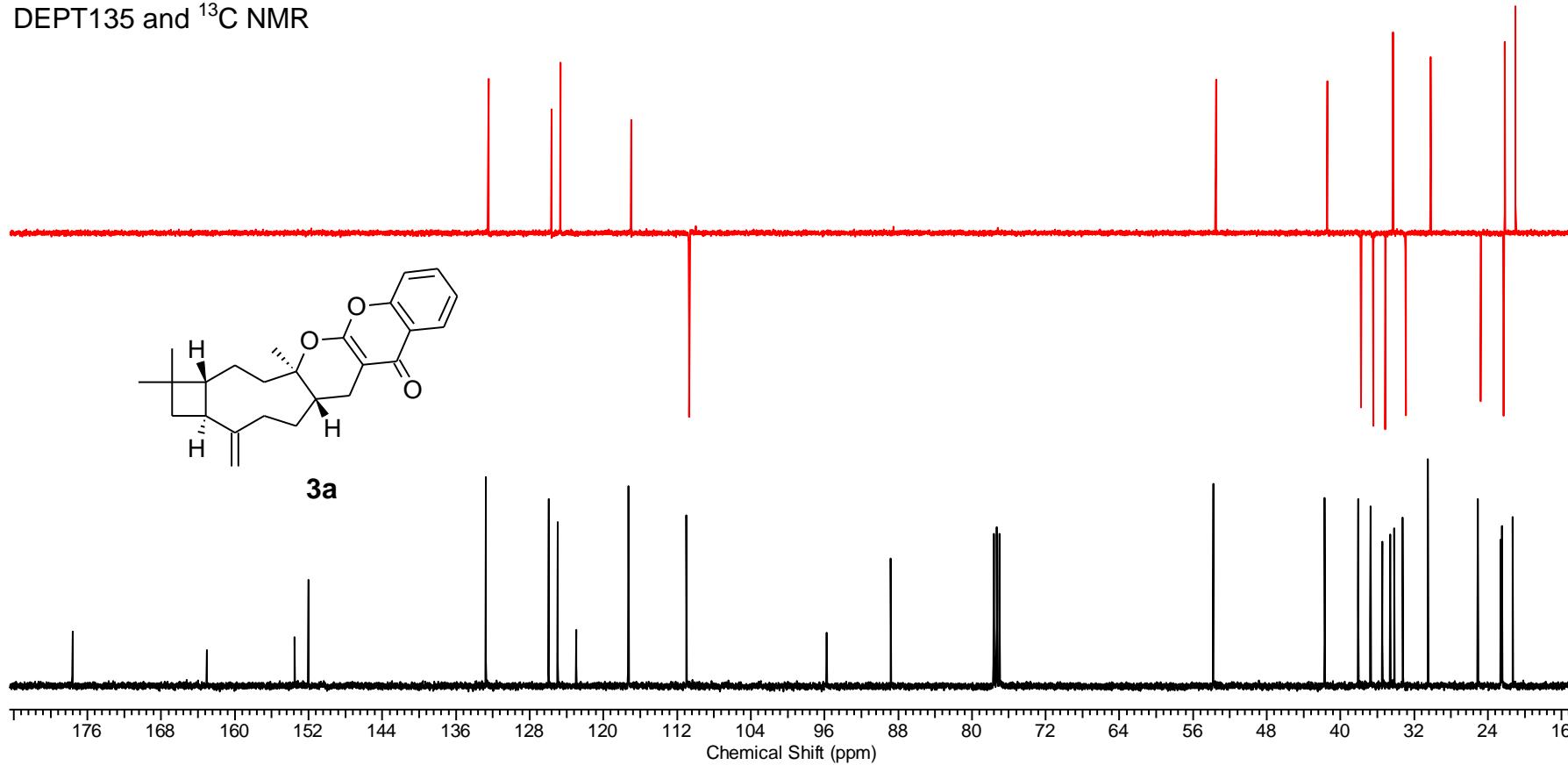


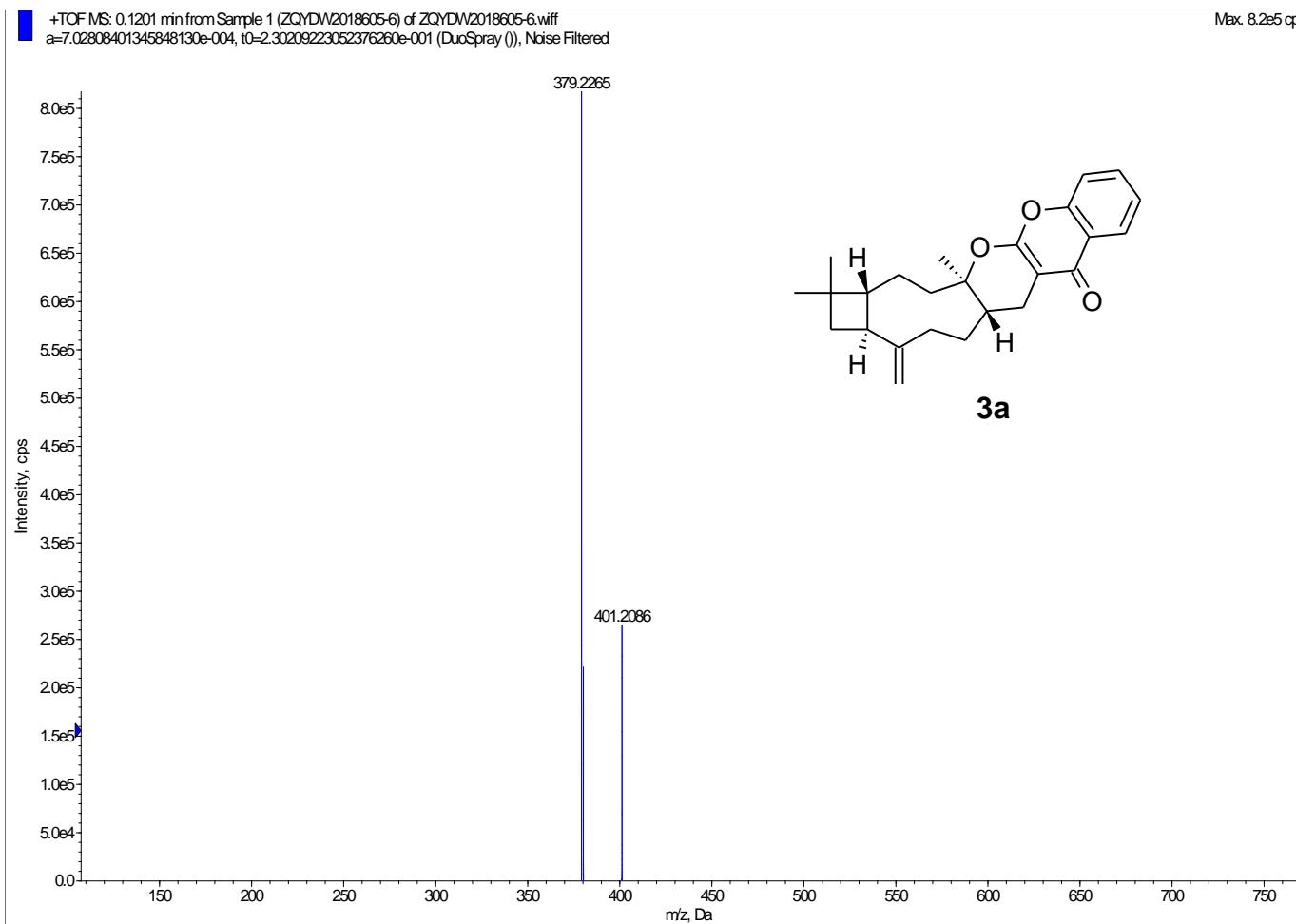
Compound 3a

3a ^1H NMR



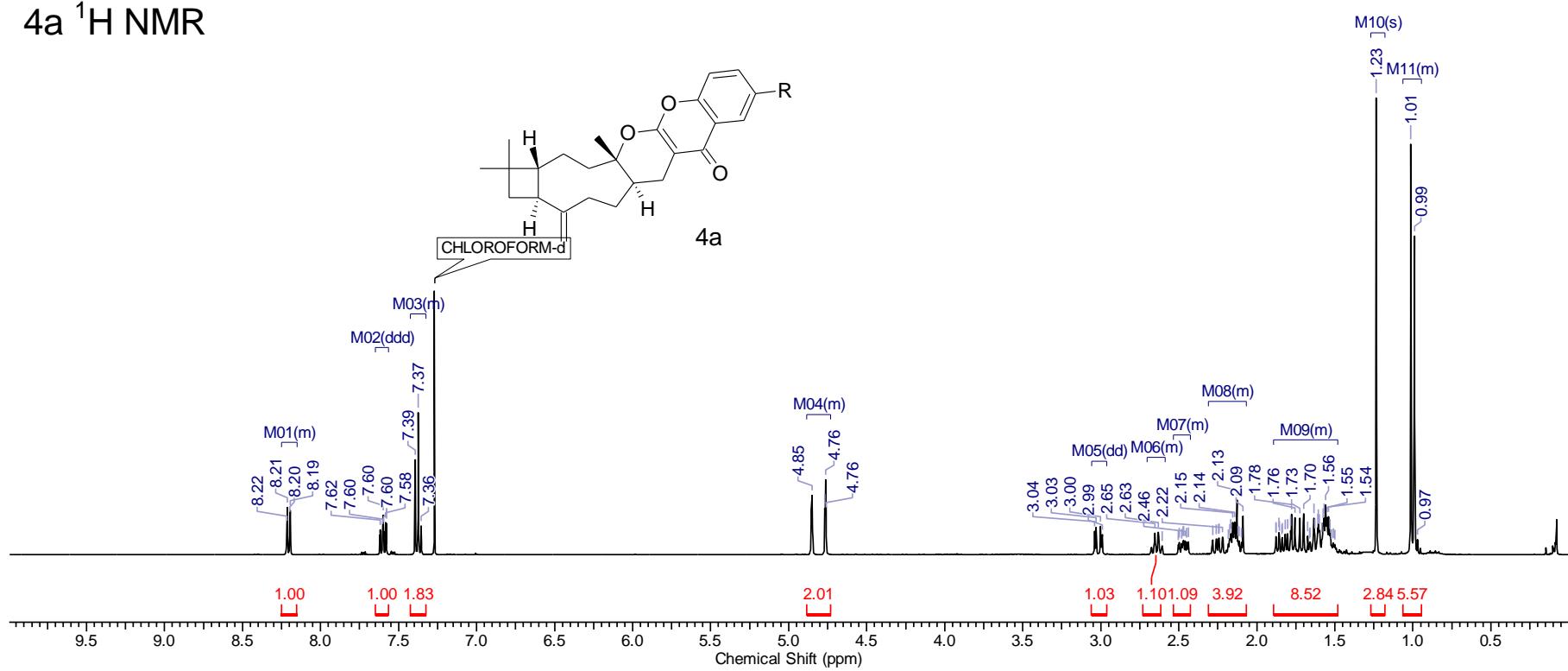
DEPT135 and ^{13}C NMR



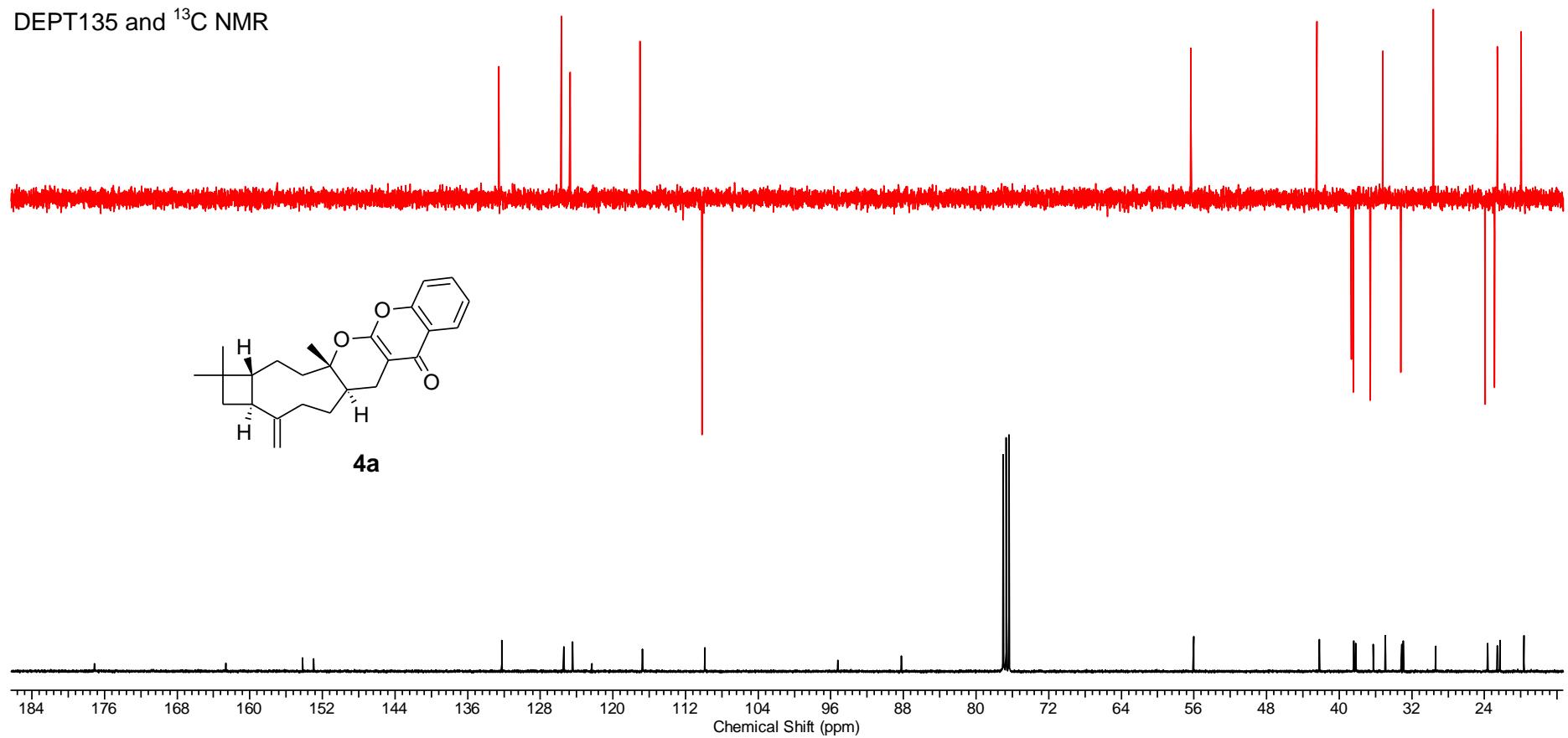


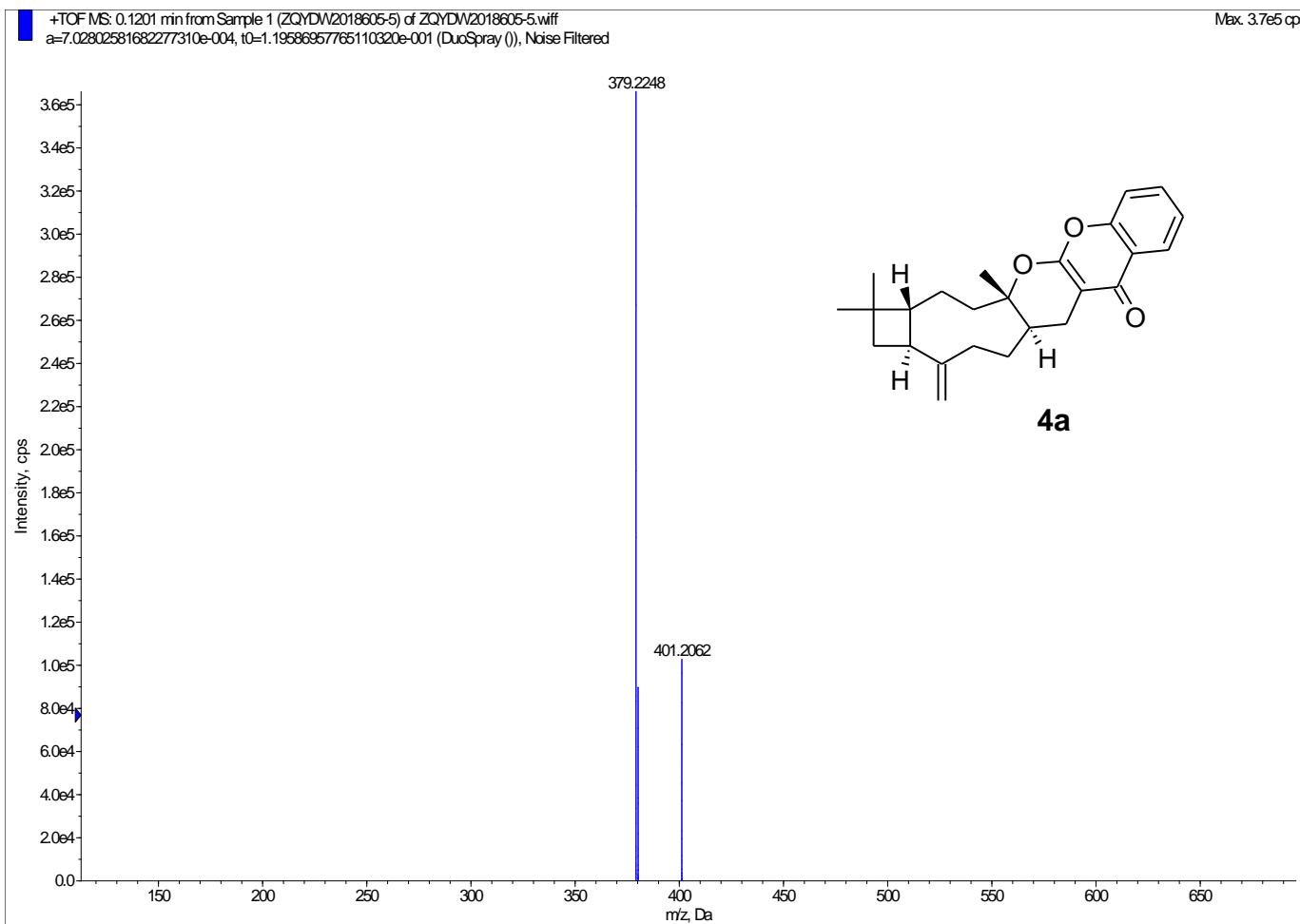
Compound 4a

4a ^1H NMR



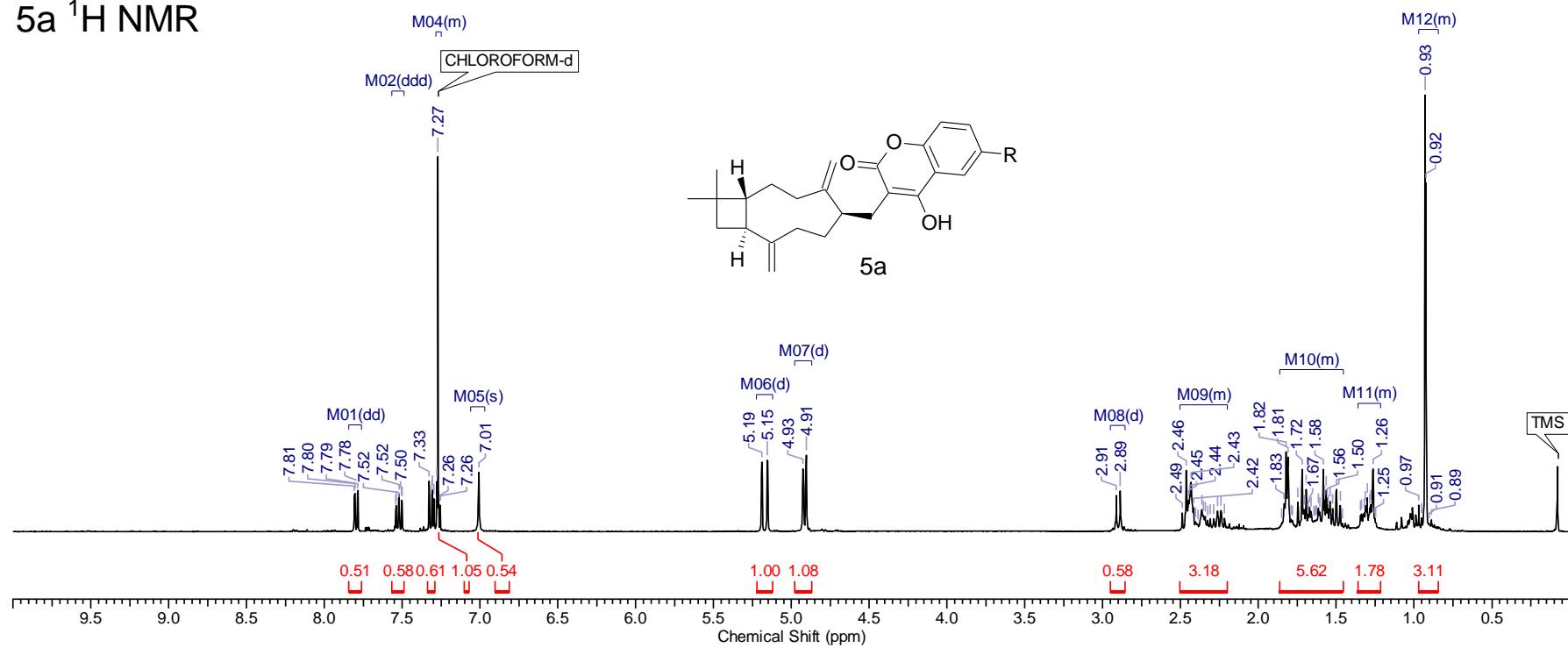
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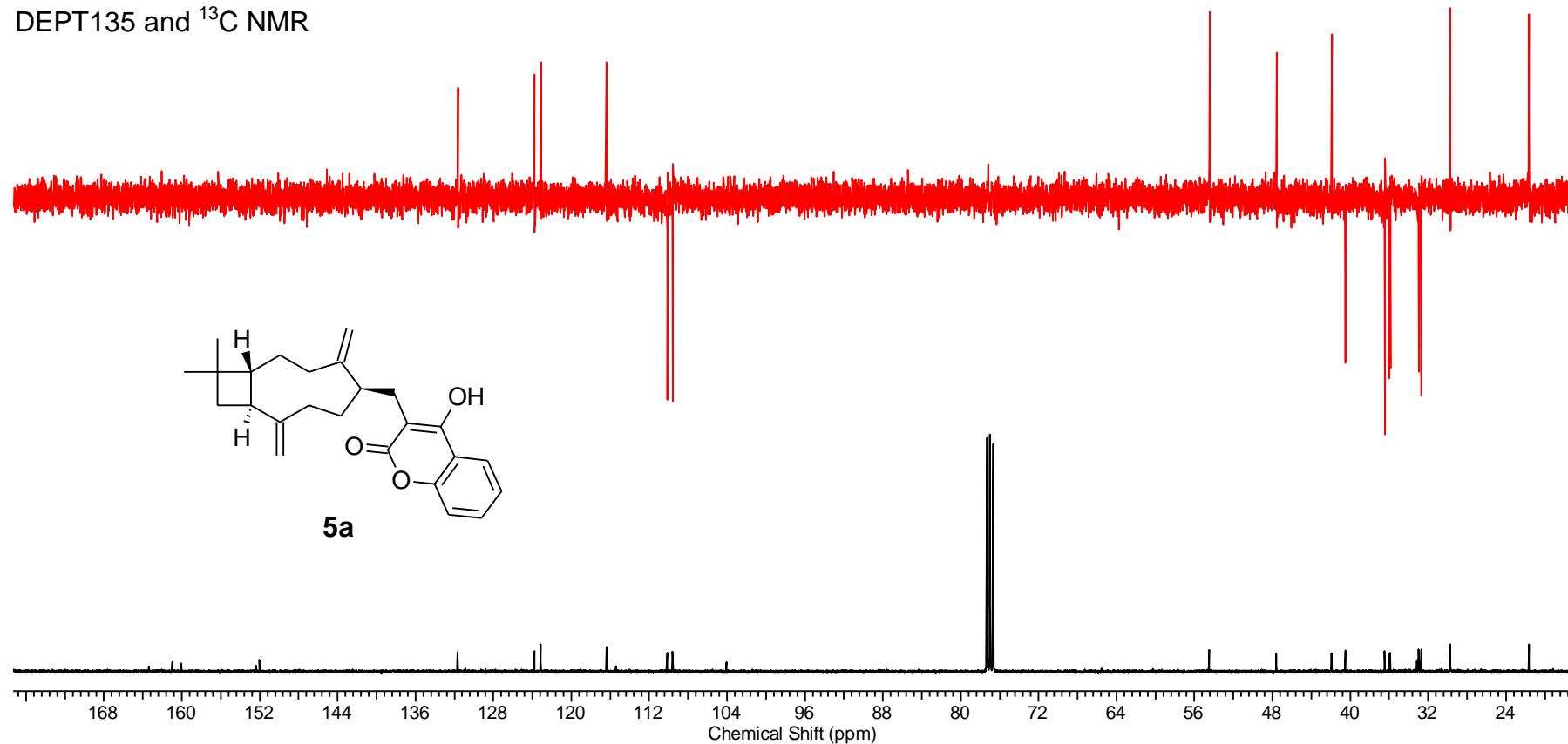


Compound 5a

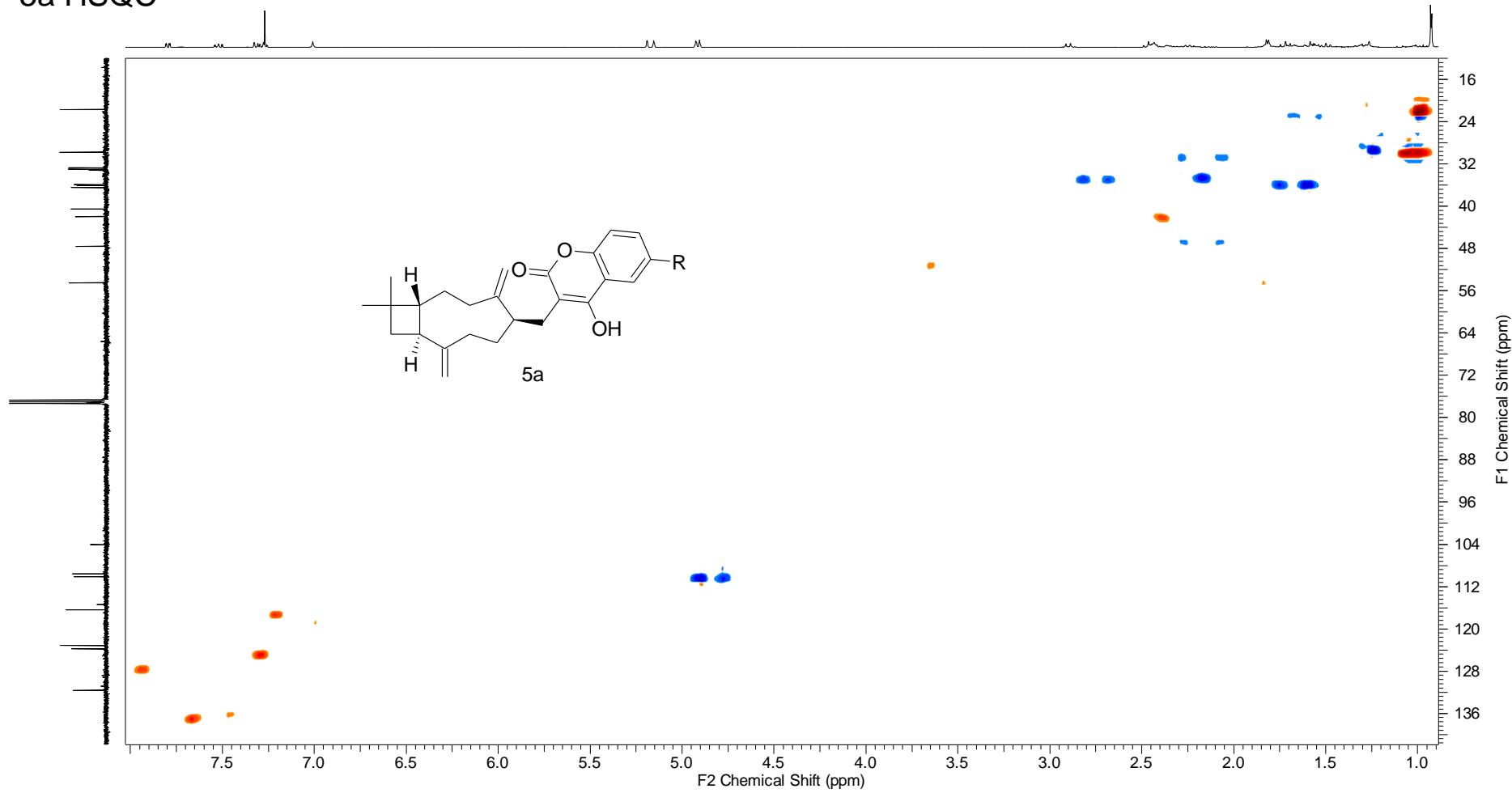
5a ^1H NMR



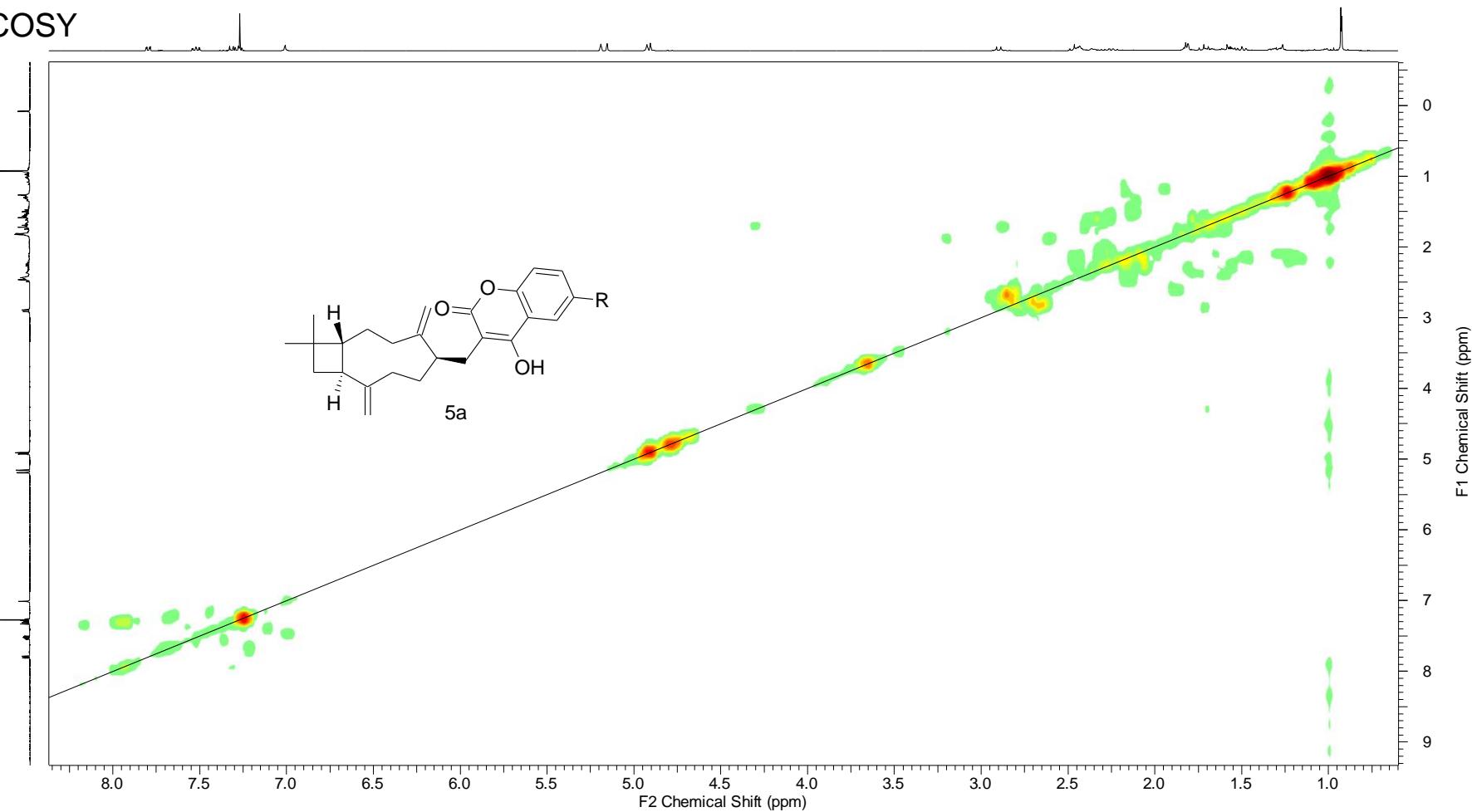
DEPT135 and ^{13}C NMR



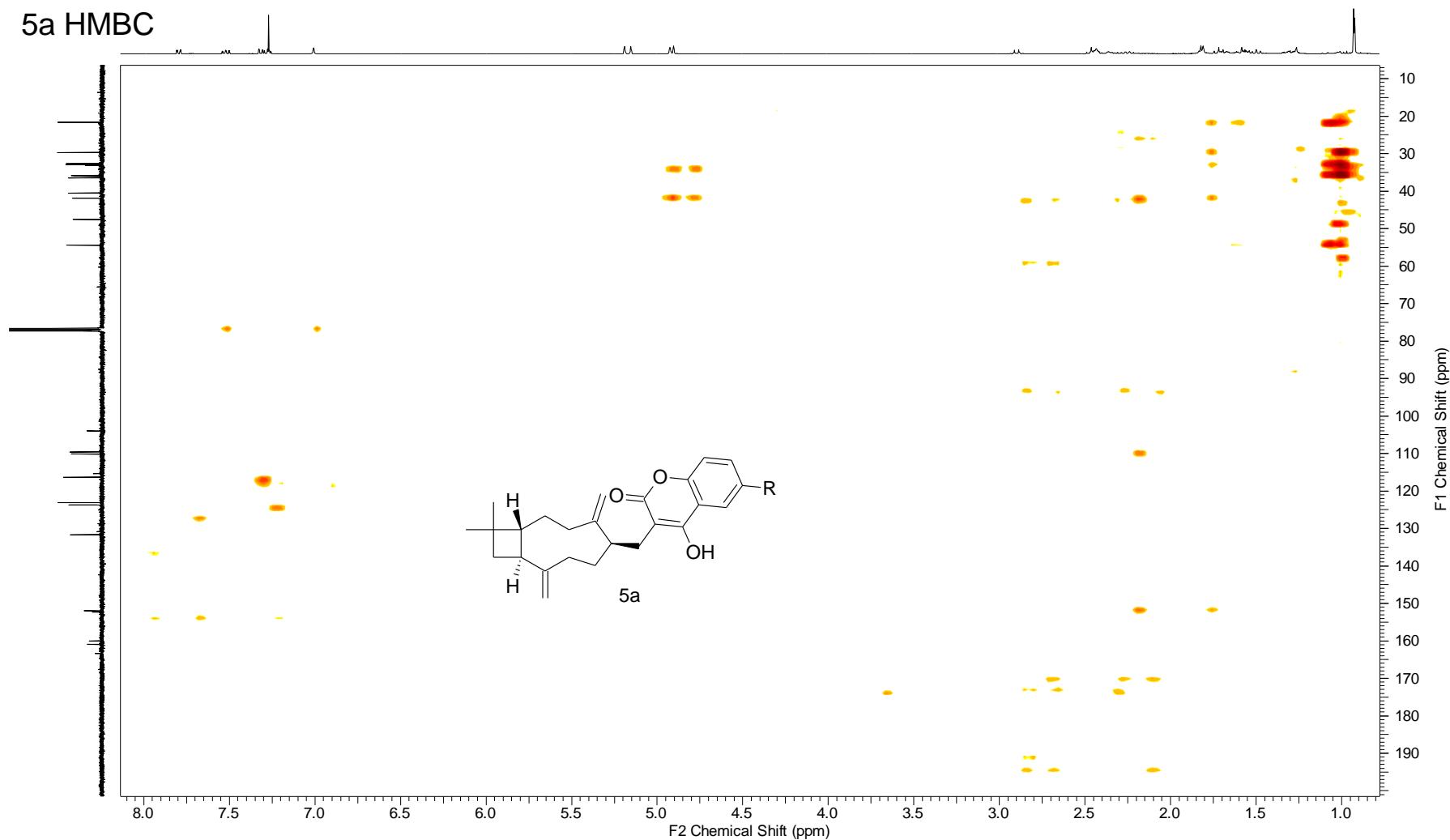
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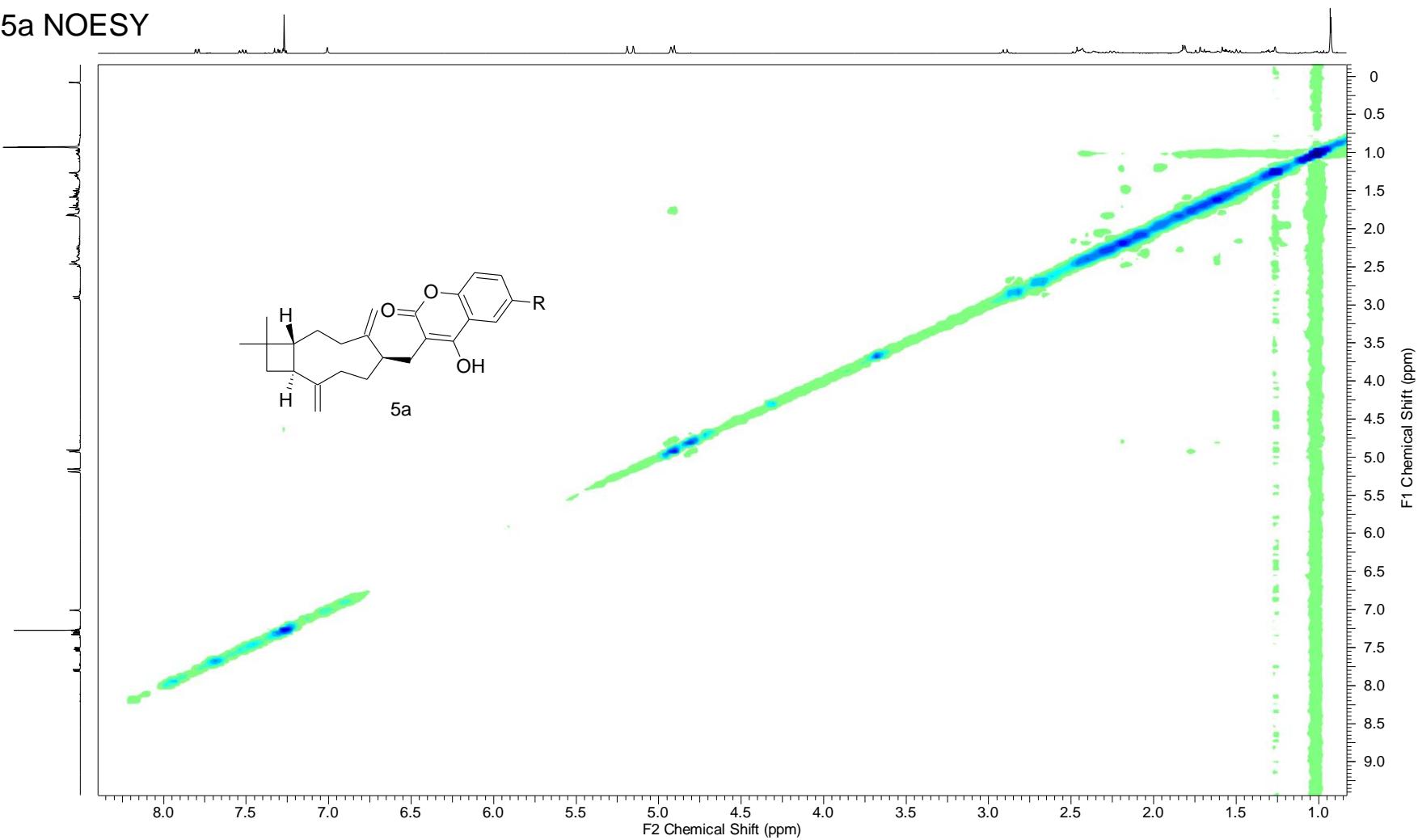
5a COSY

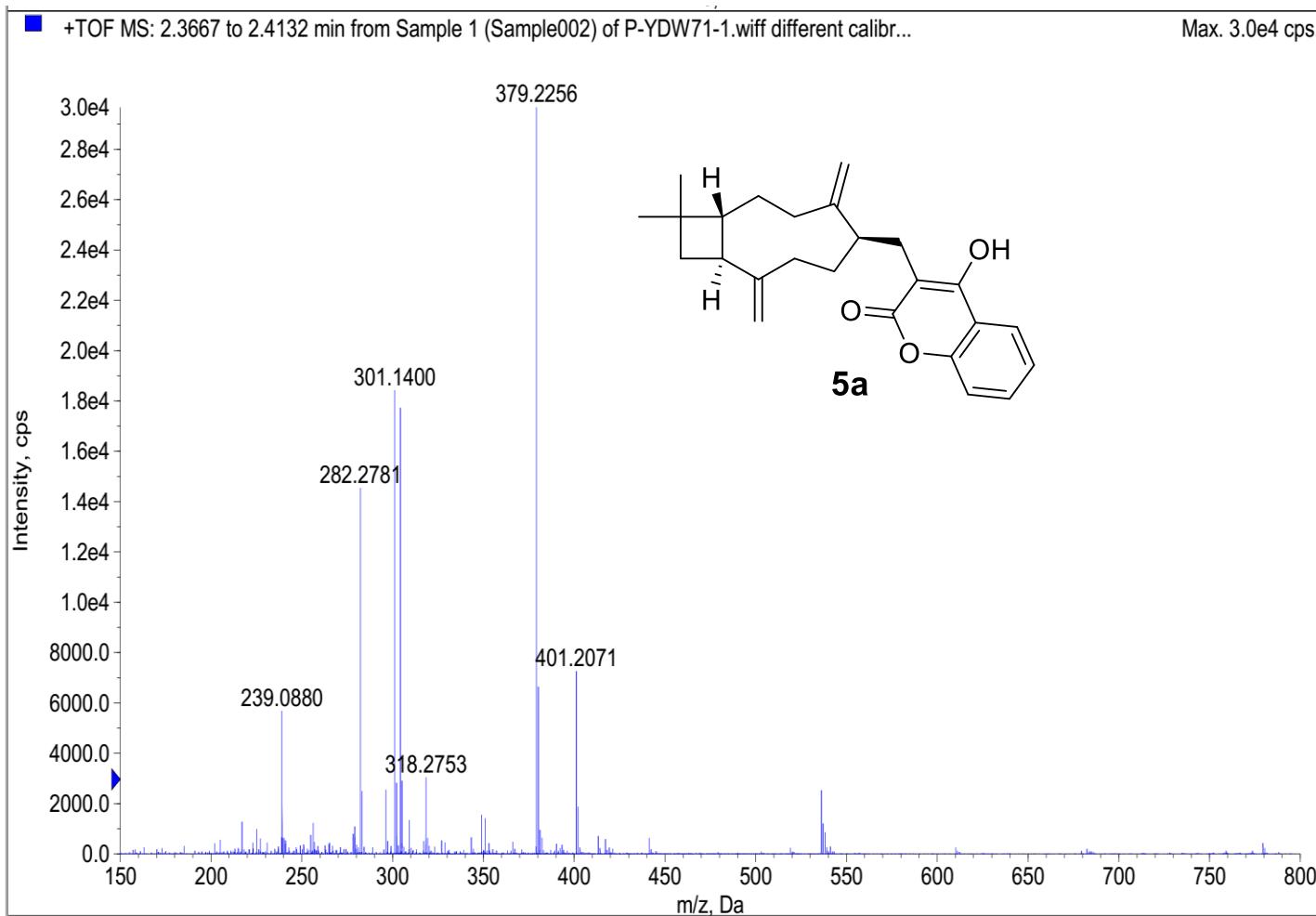


5a HMBC



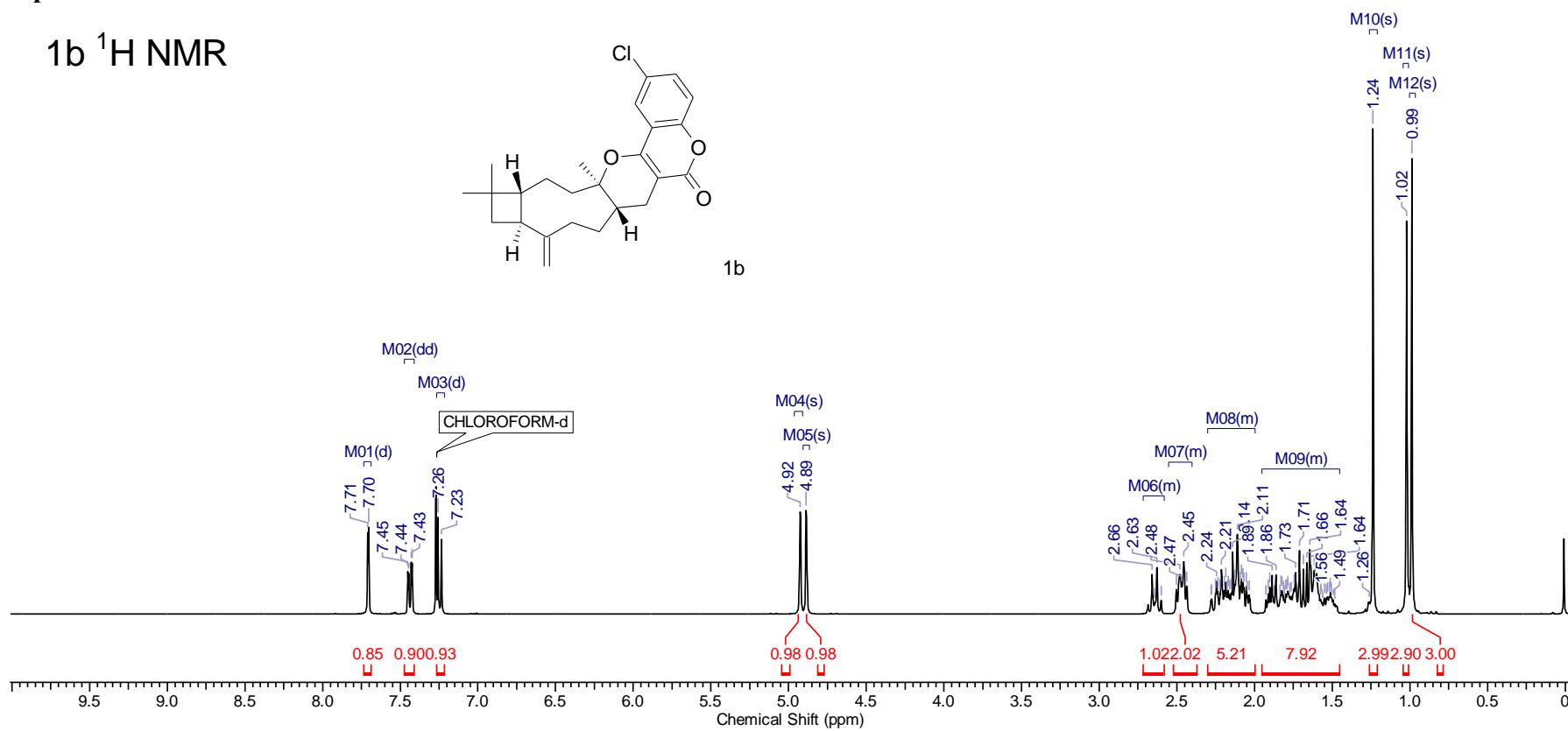
5a NOESY



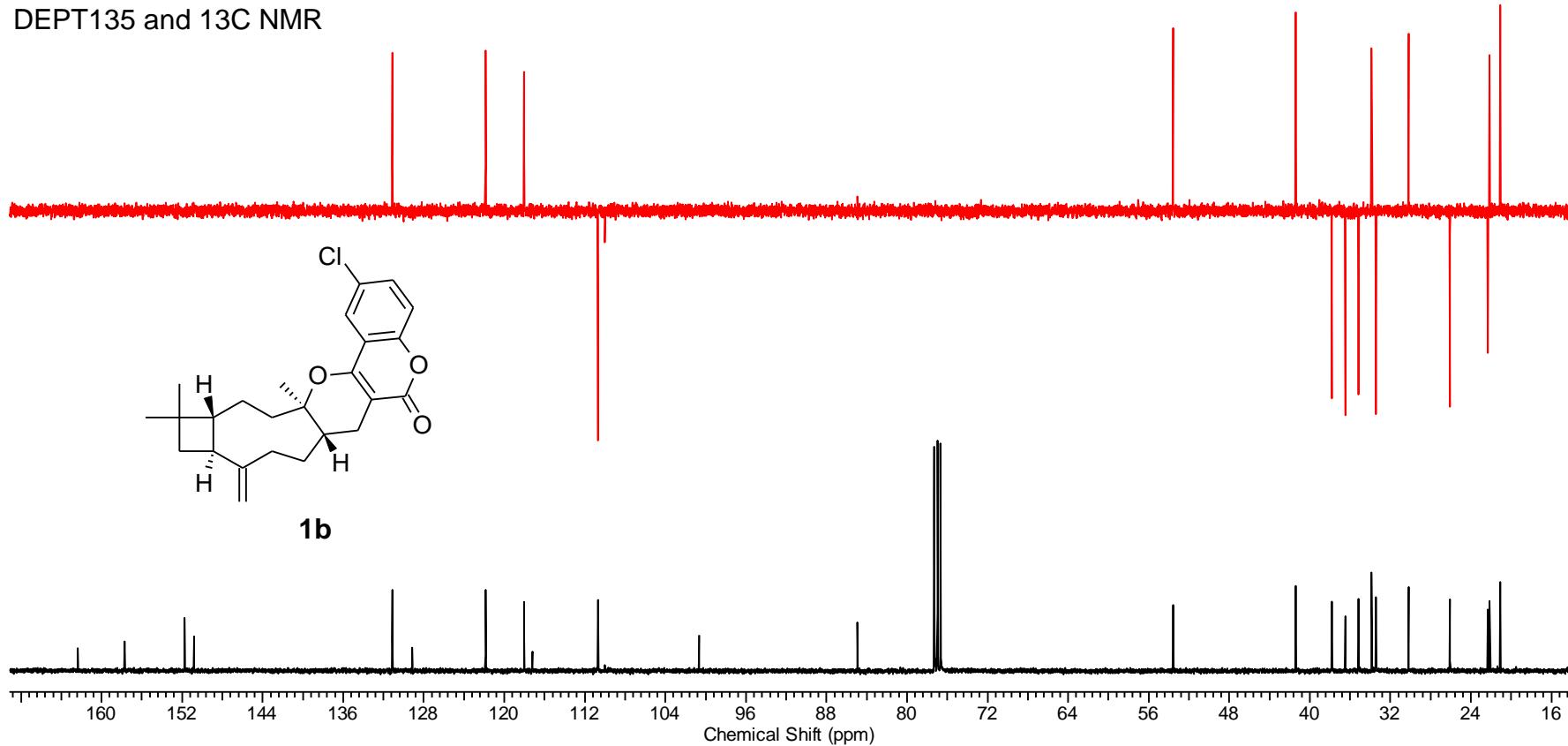


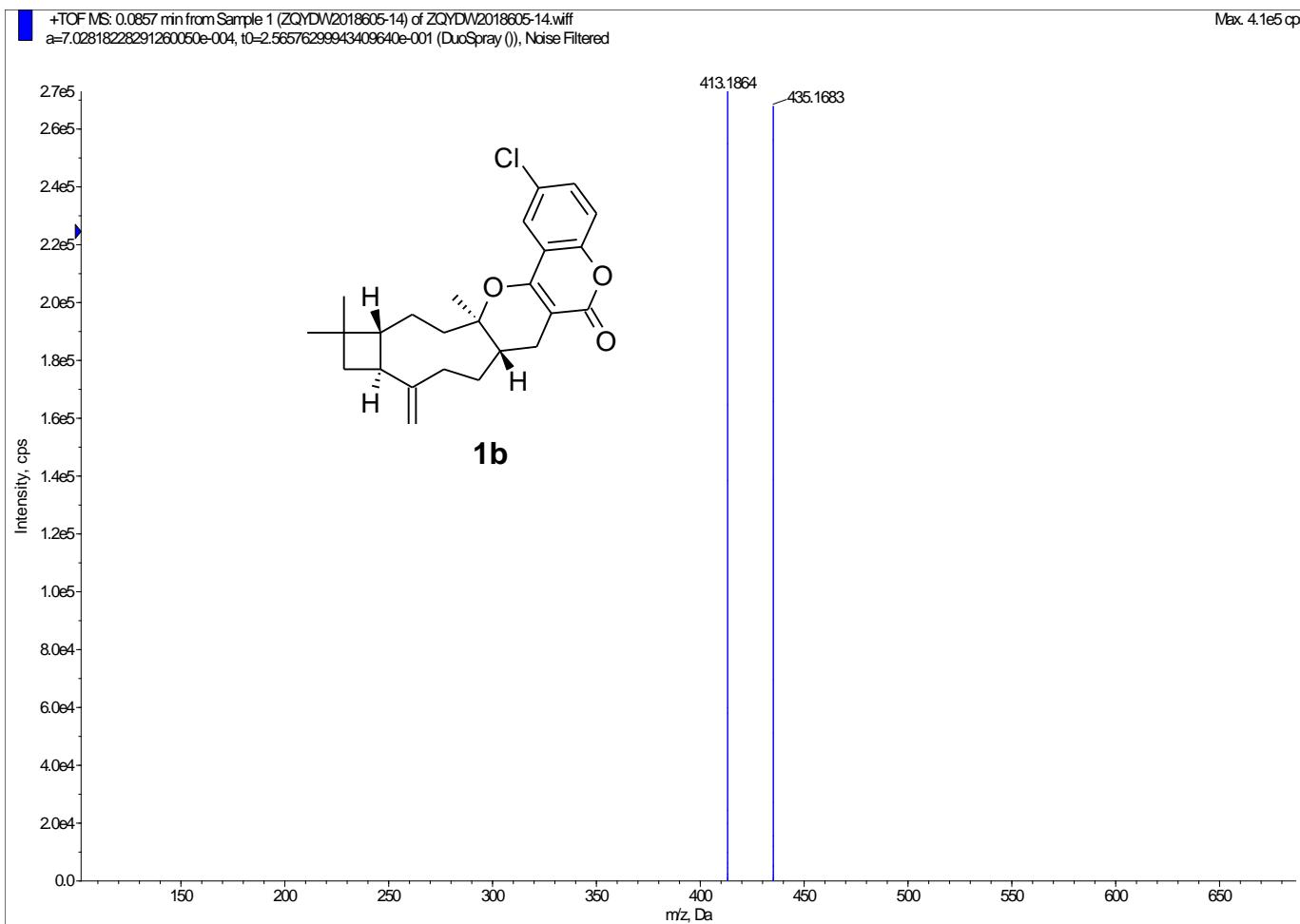
Compound 1b

1b ^1H NMR



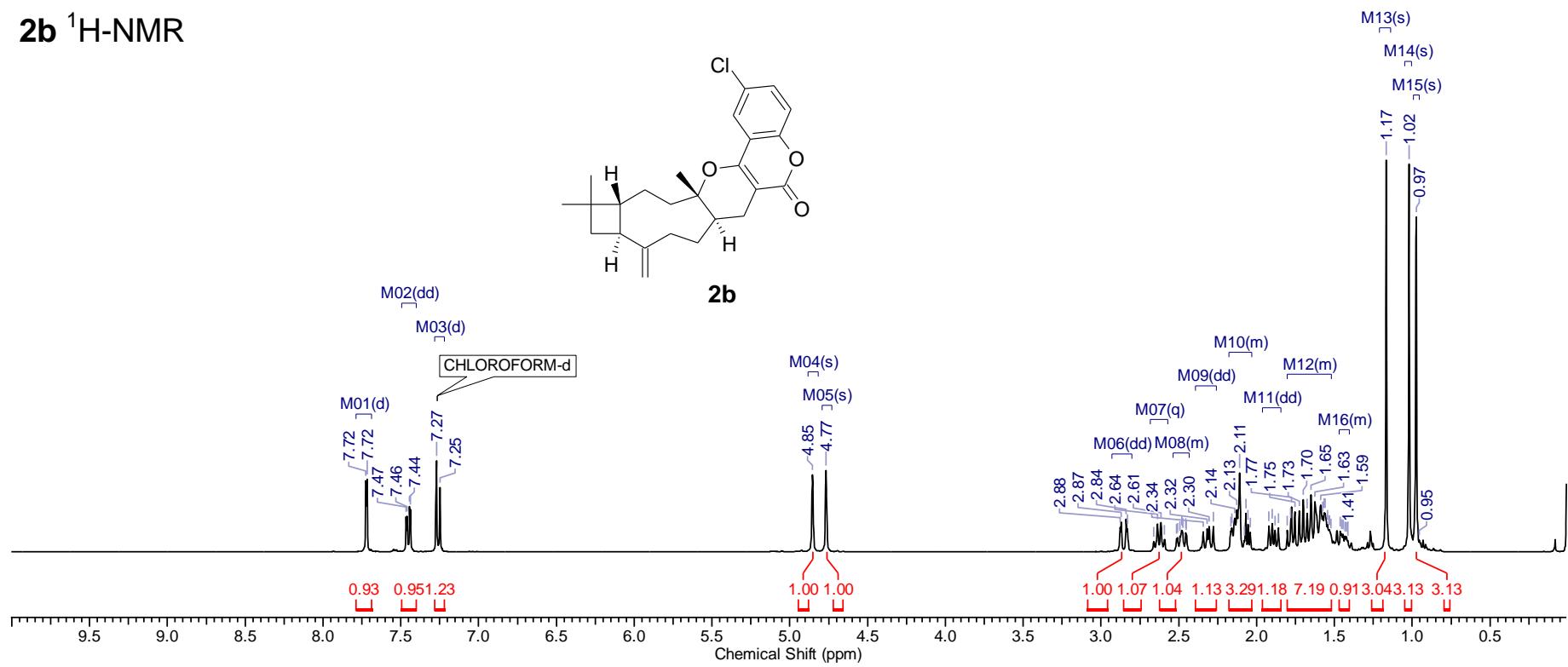
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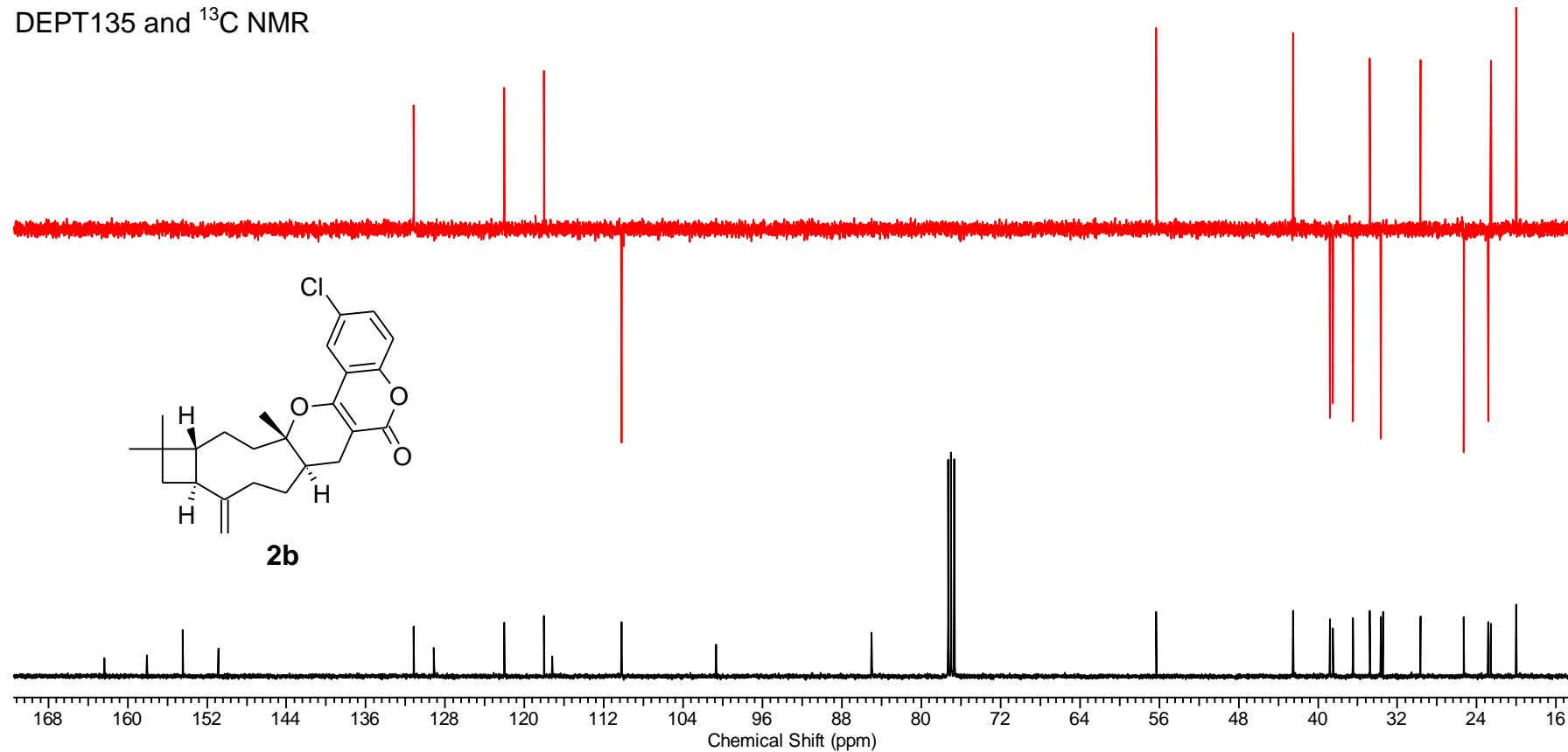


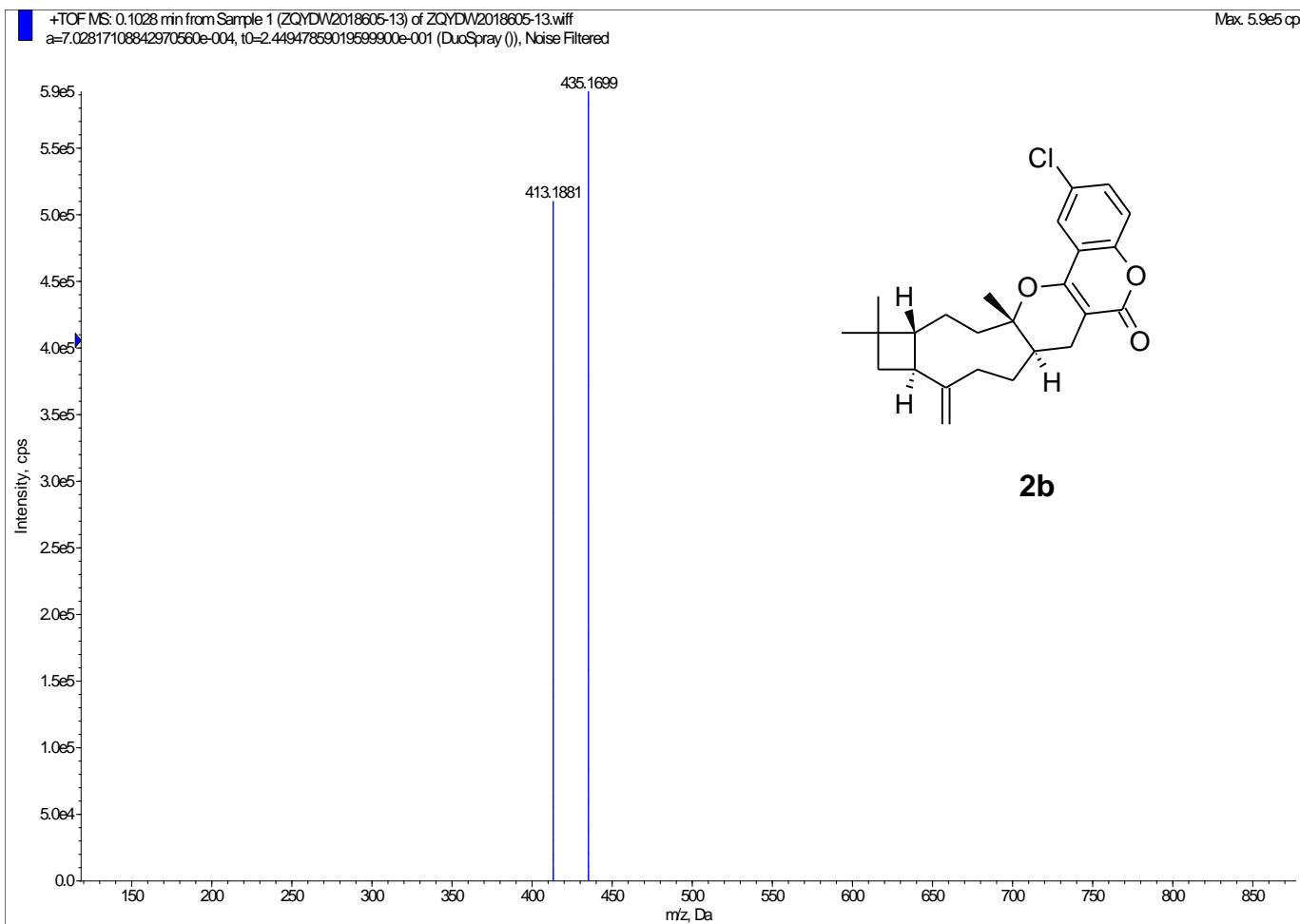
Compound 2b

2b ^1H -NMR



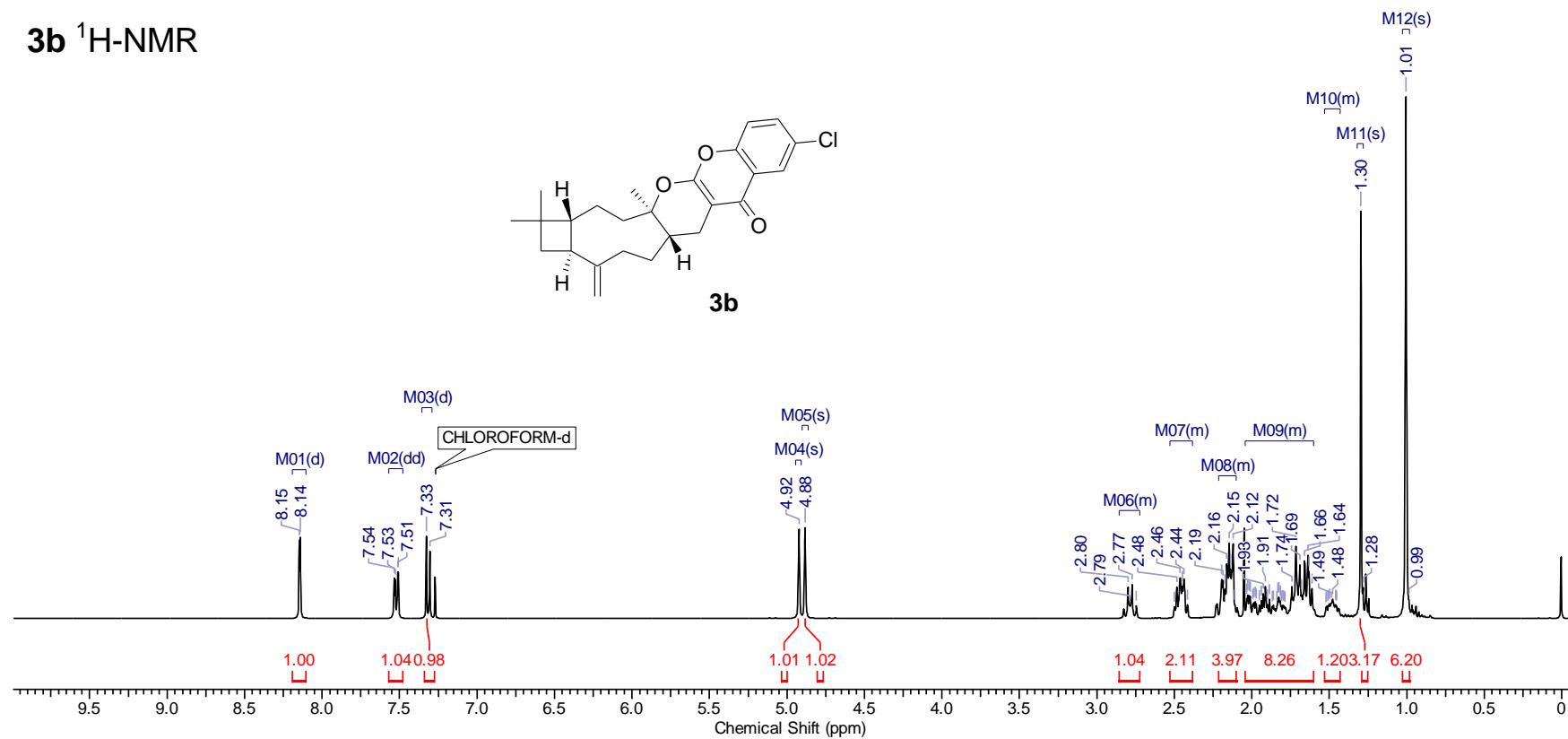
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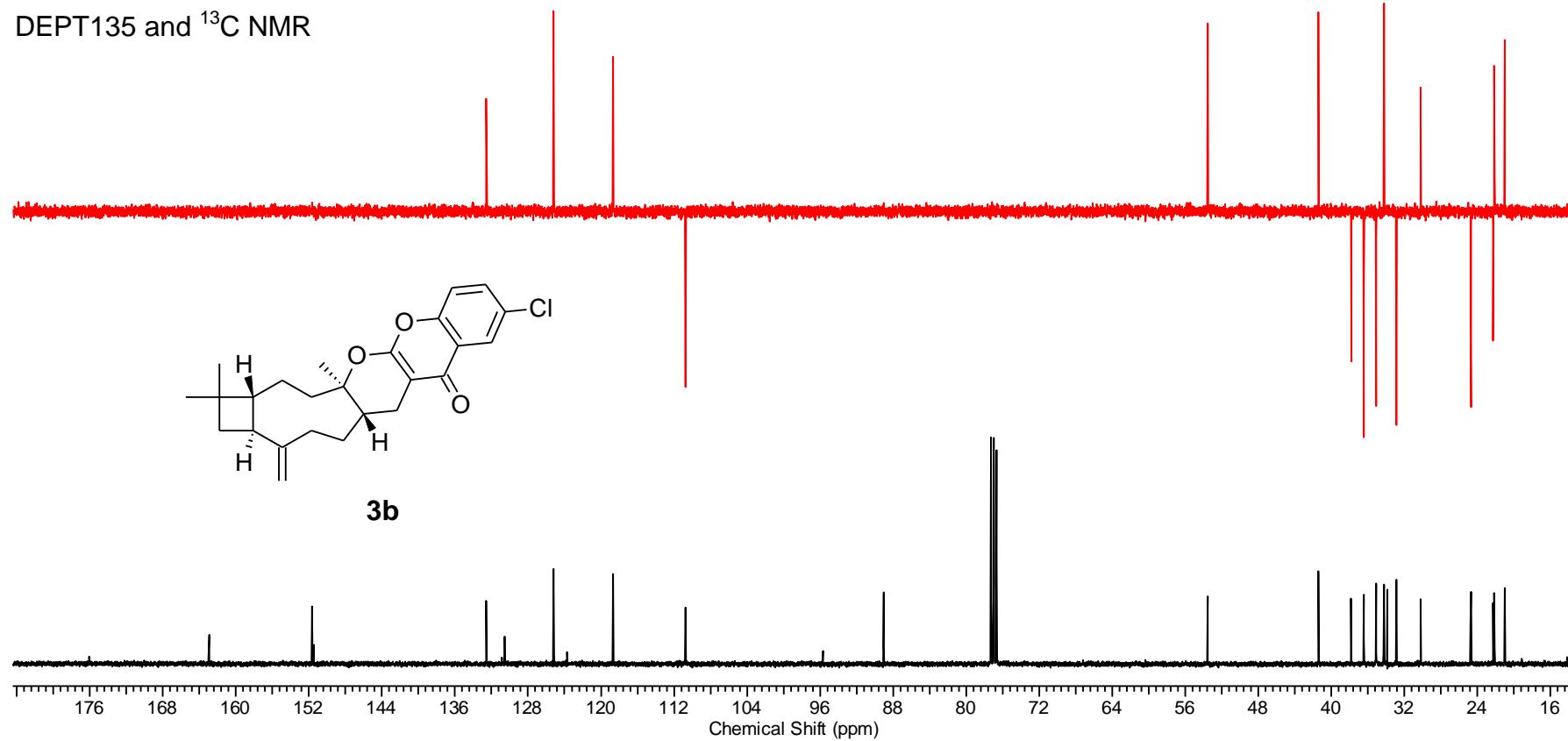


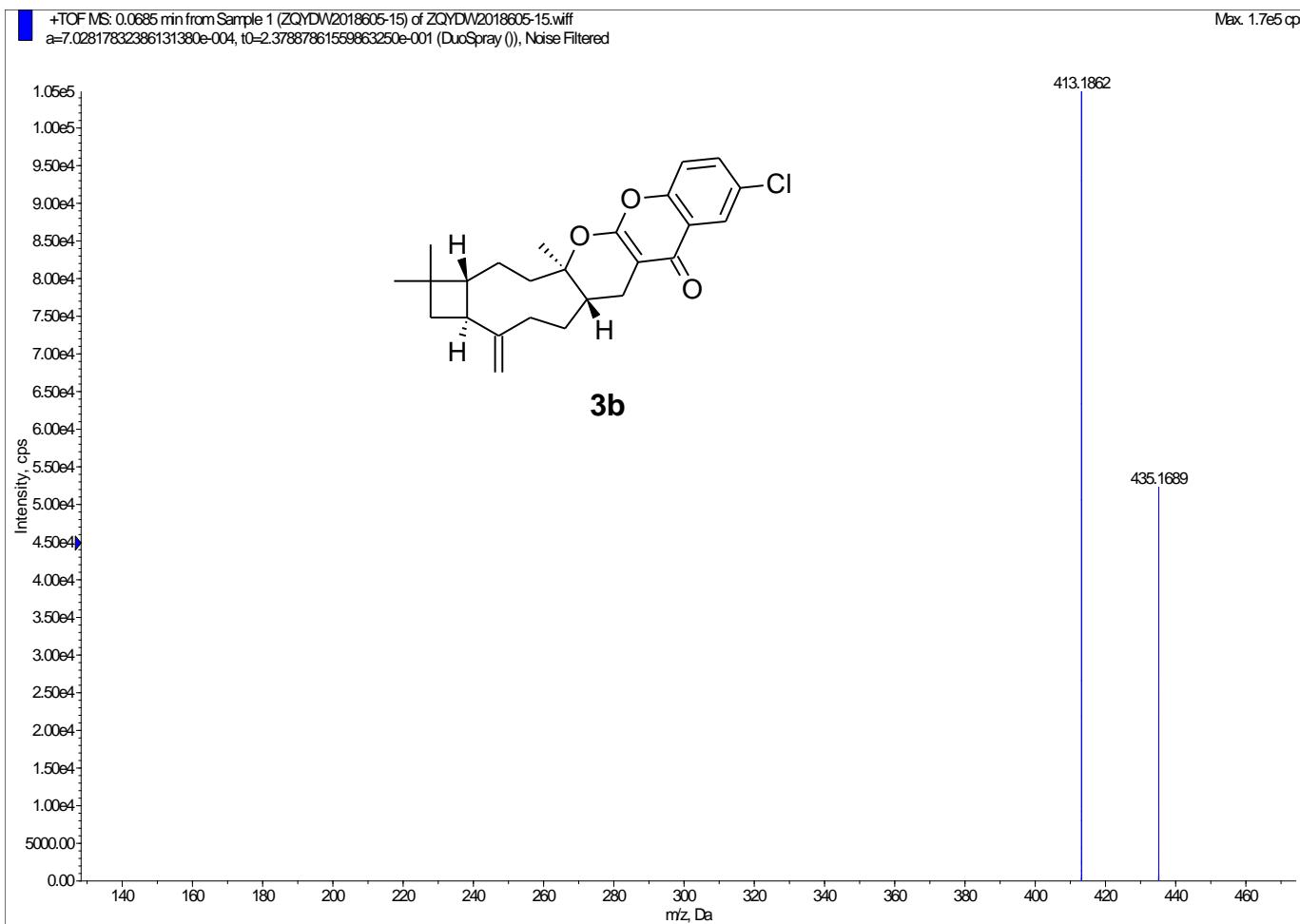
Compound 3b

3b $^1\text{H-NMR}$



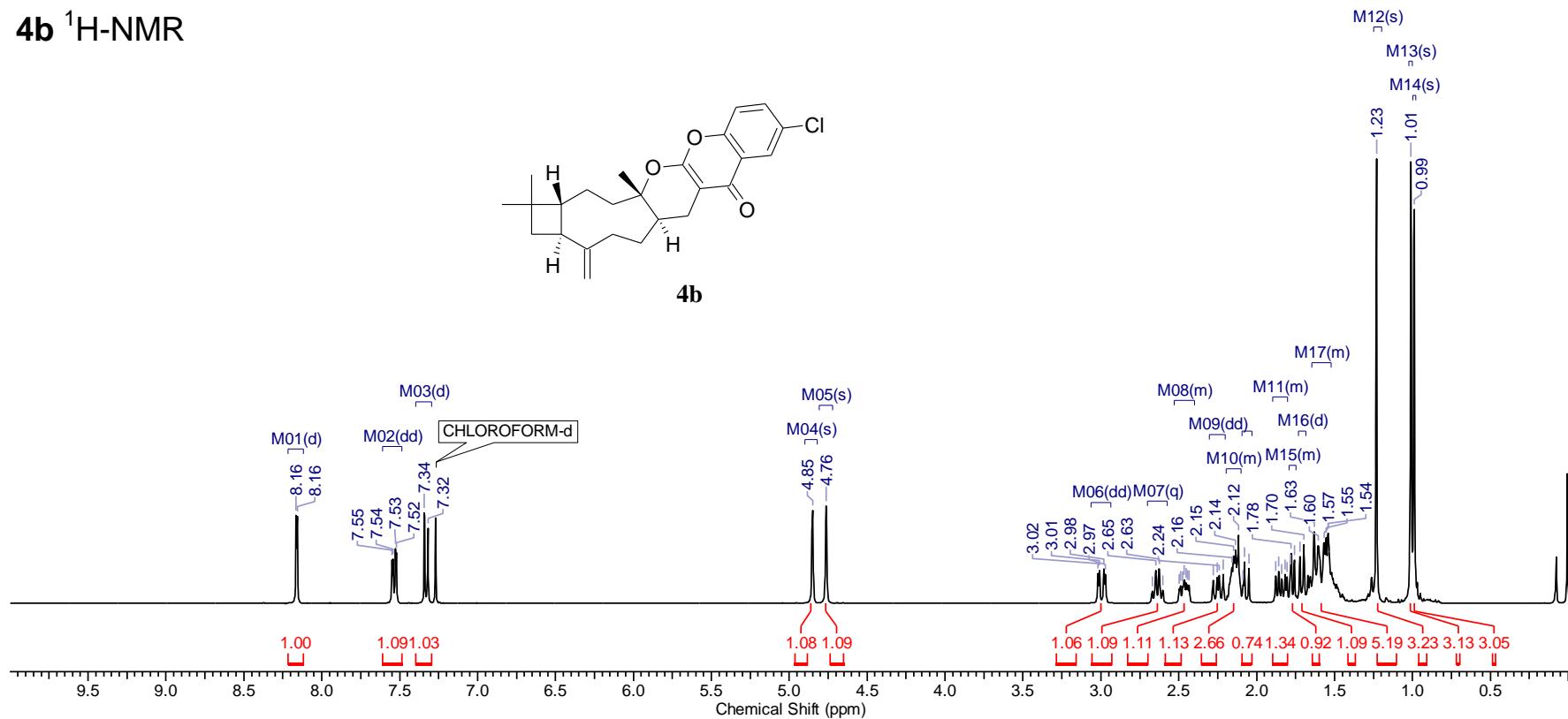
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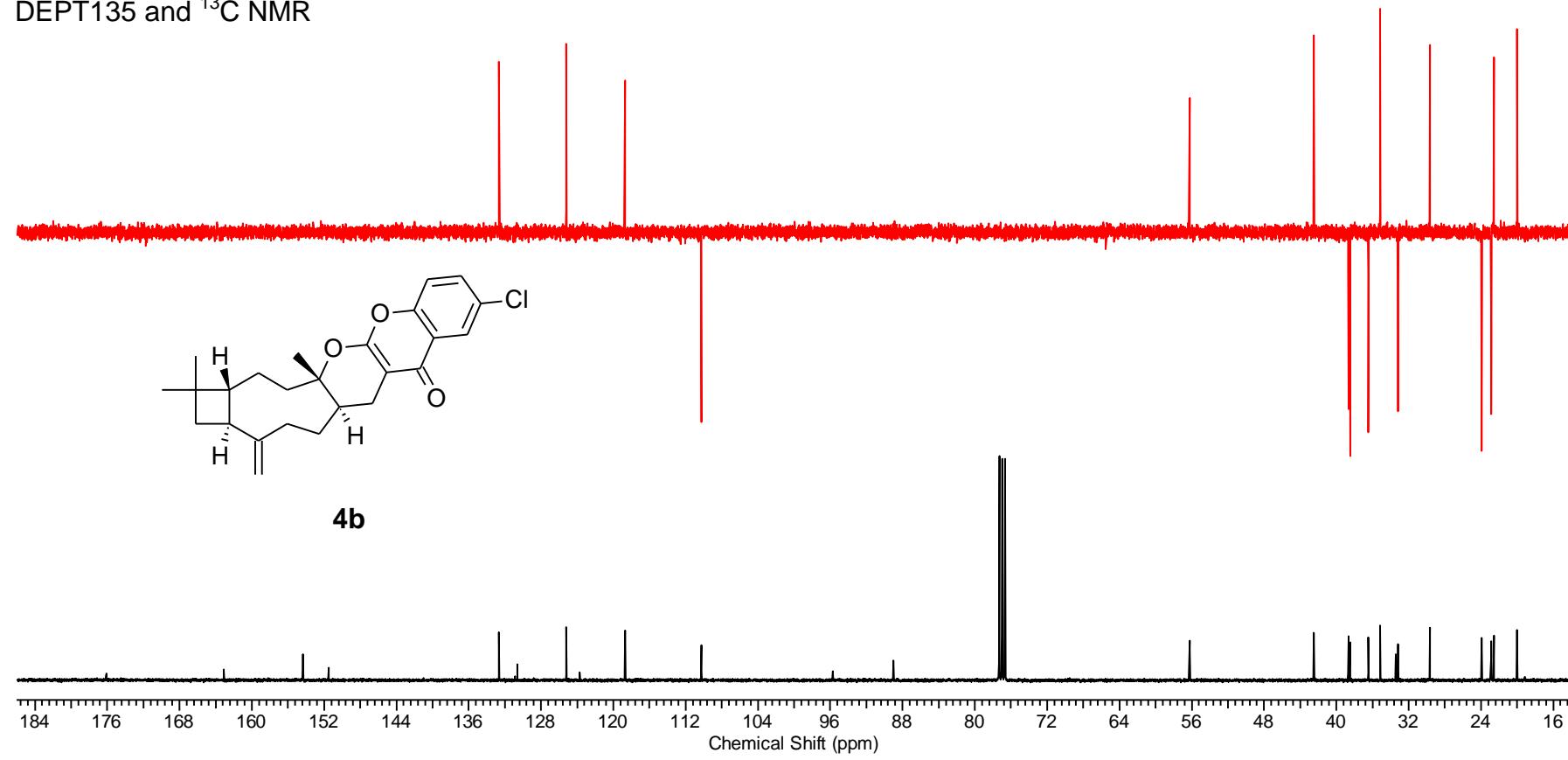


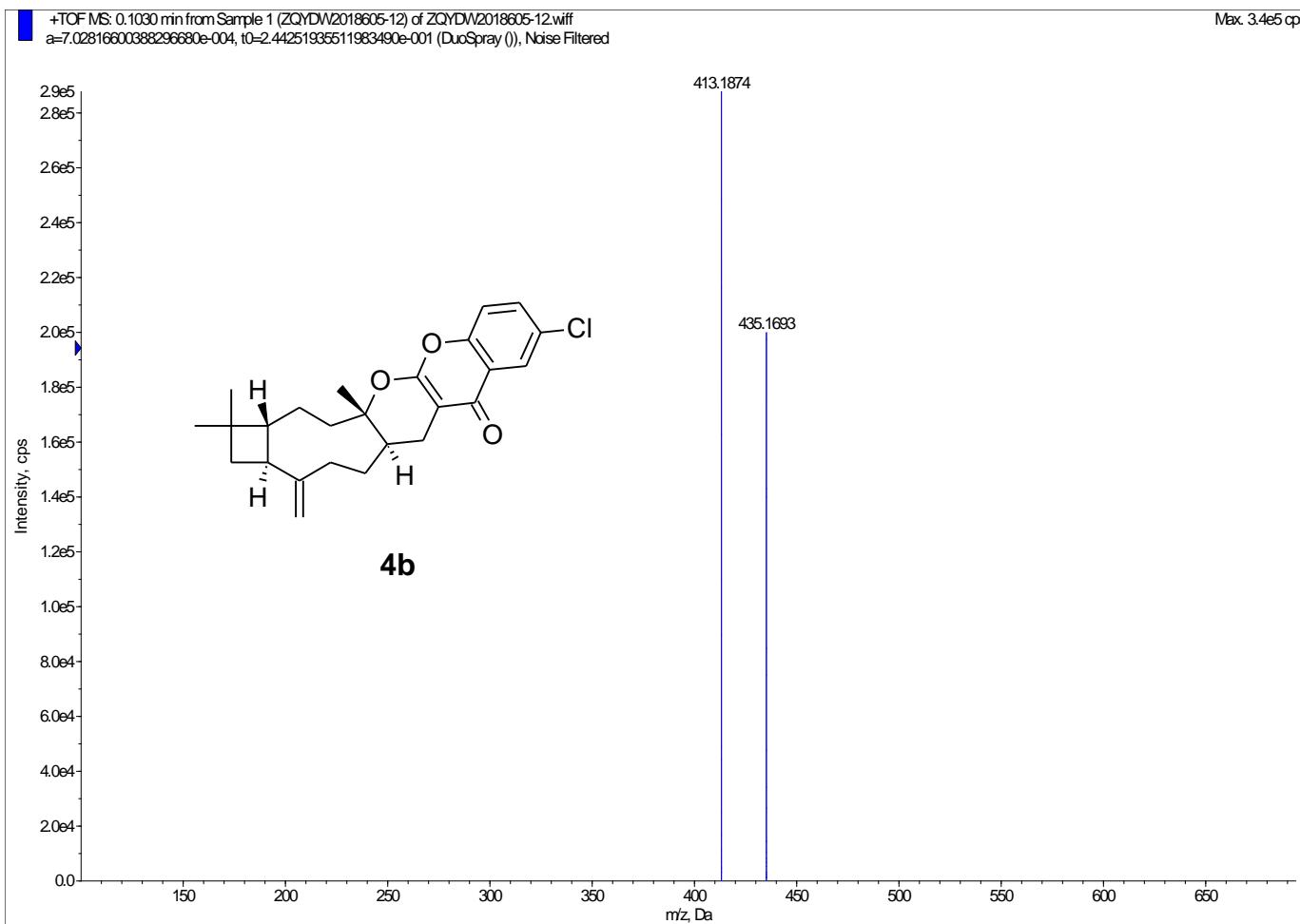
Compound 4b

4b ^1H -NMR



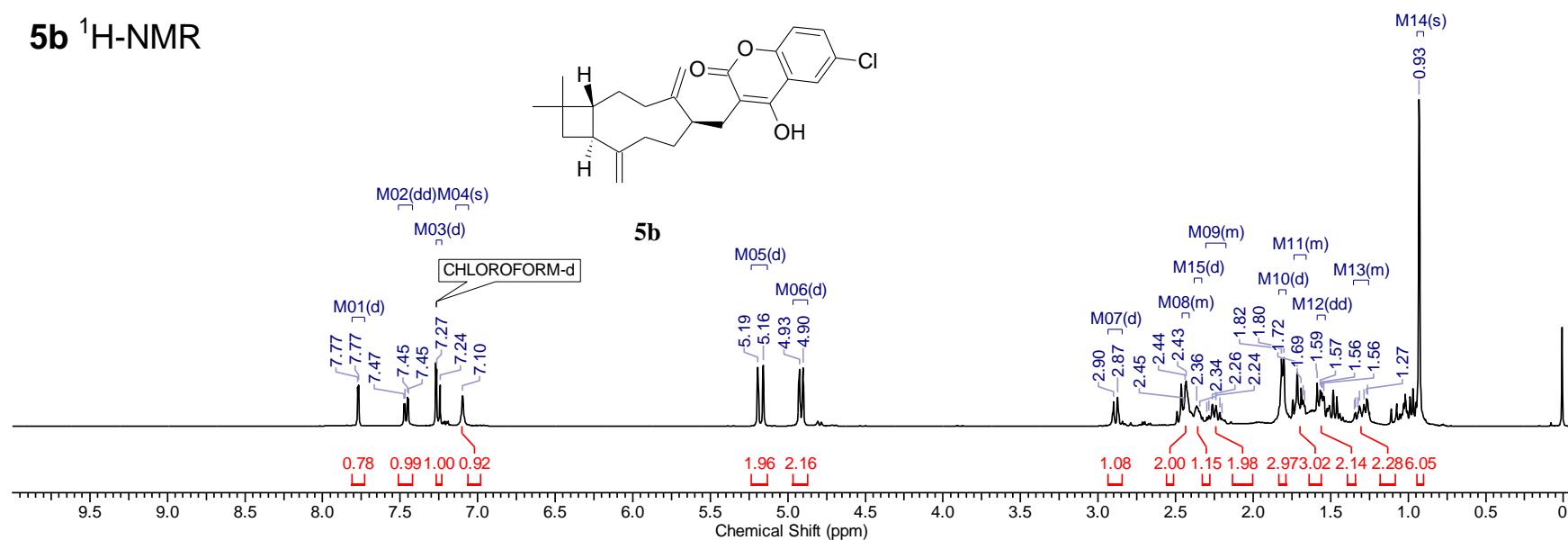
DEPT135 and ^{13}C NMR



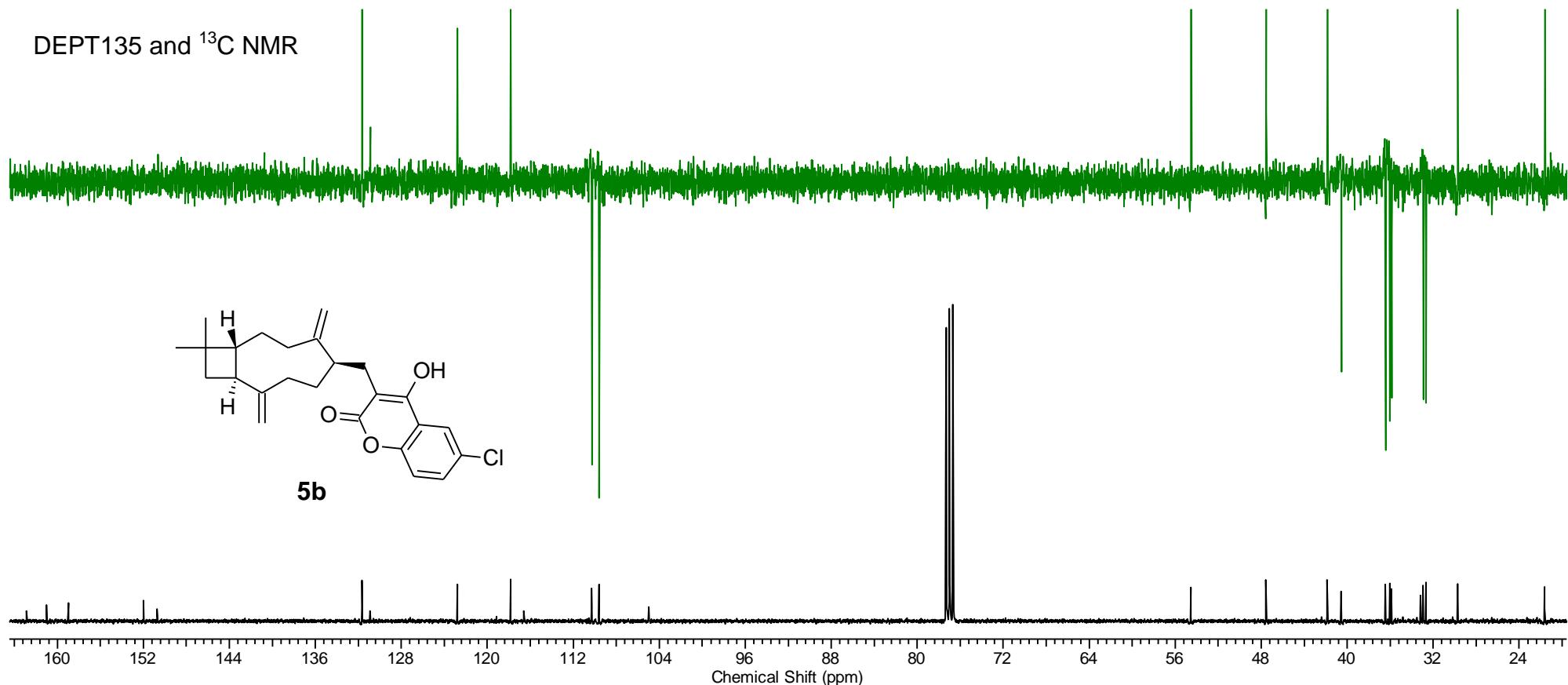
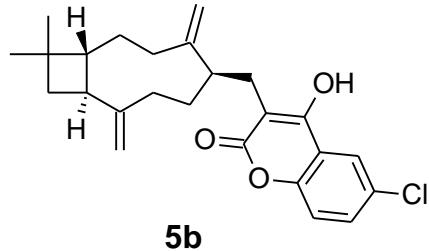


Compound 5b

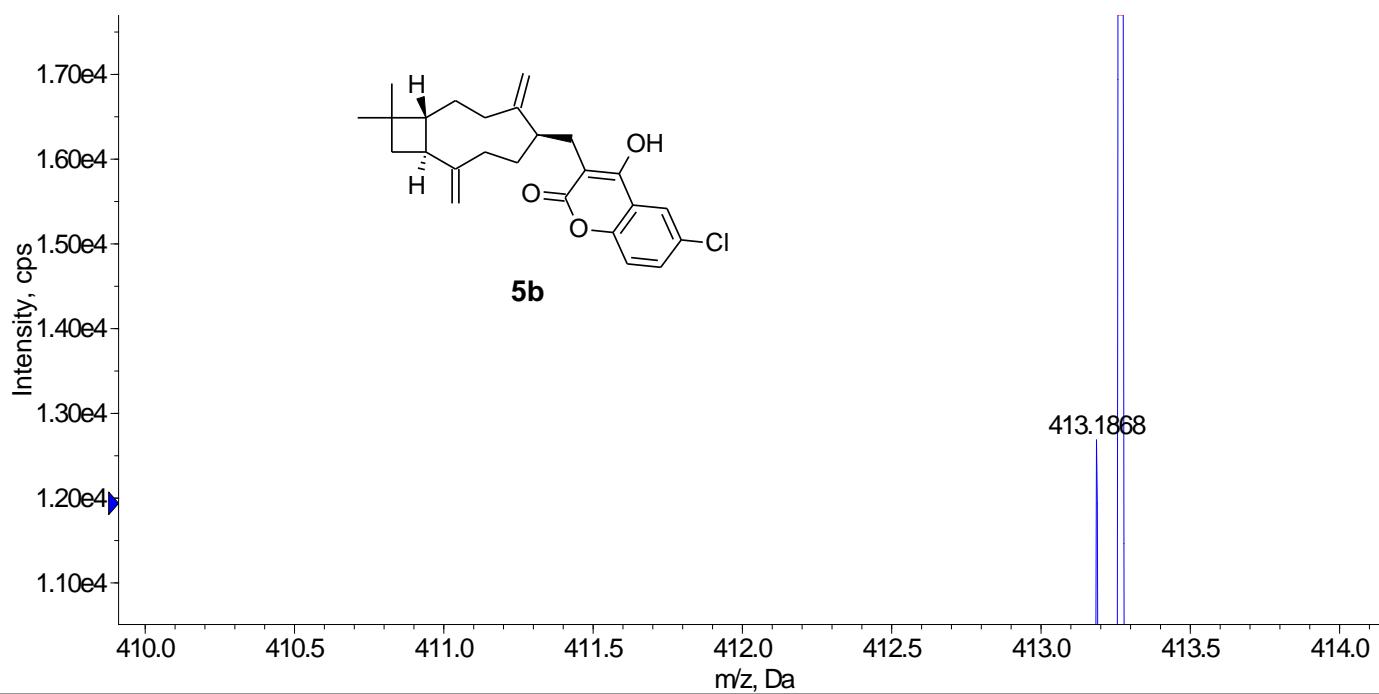
5b ^1H -NMR



DEPT135 and ^{13}C NMR



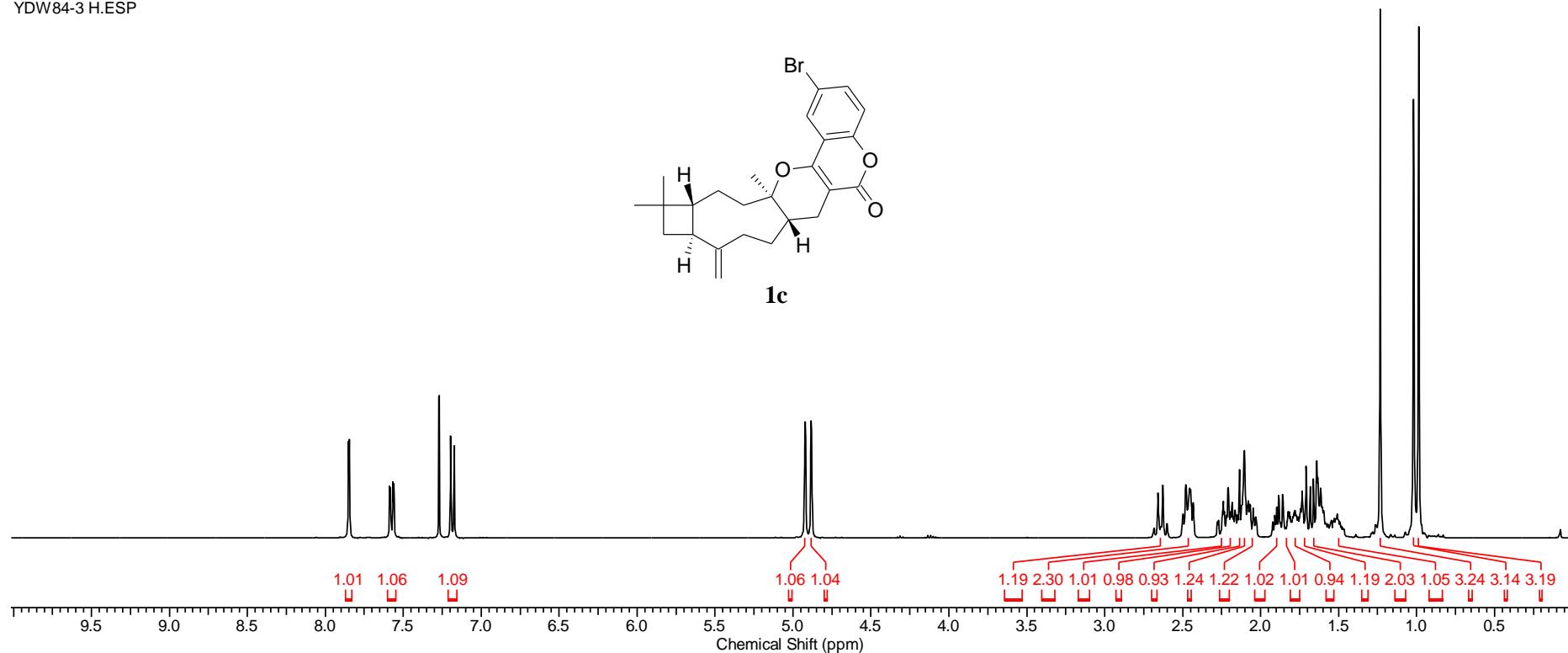
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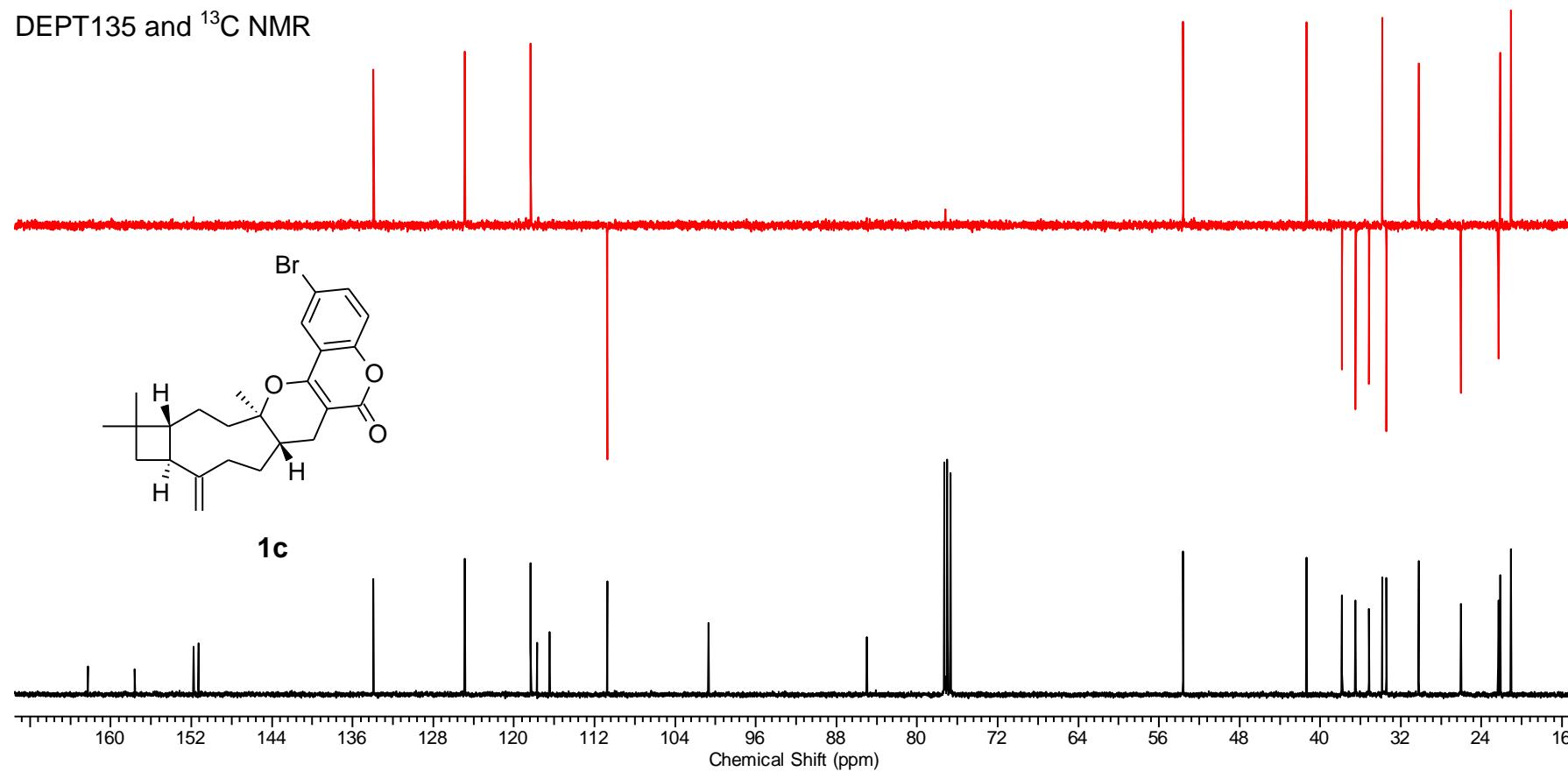
Compound 1c

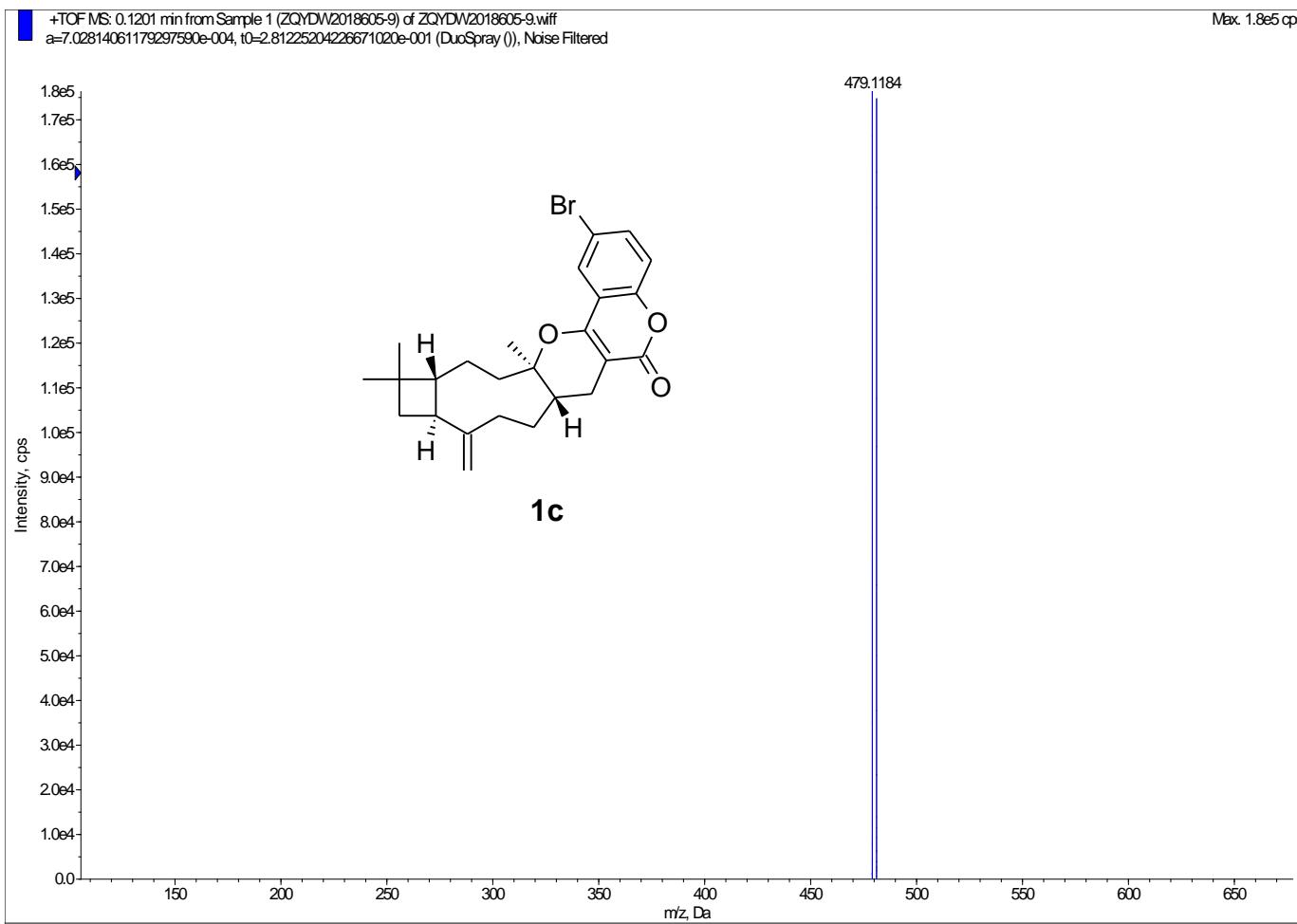
1c ^1H -NMR

YDW84-3 H.ESP



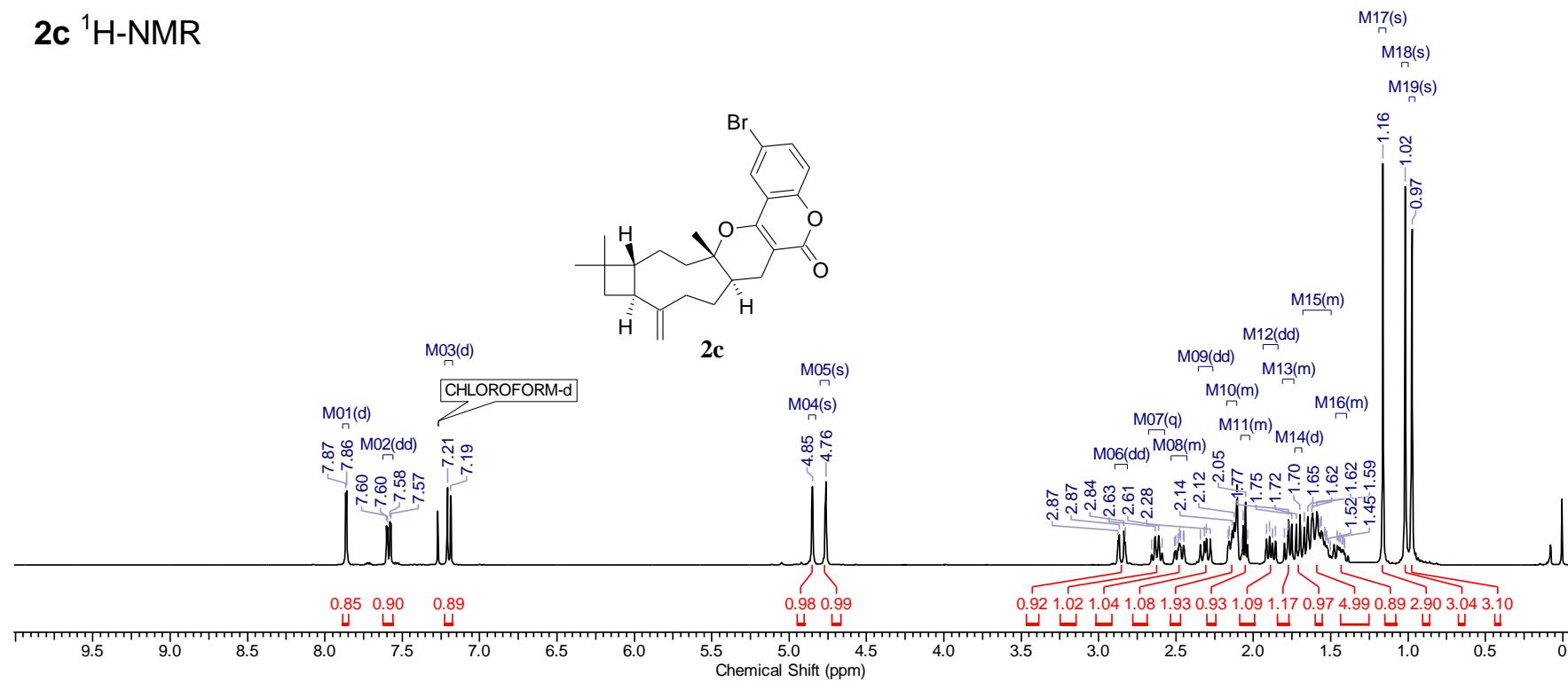
DEPT135 and ^{13}C NMR



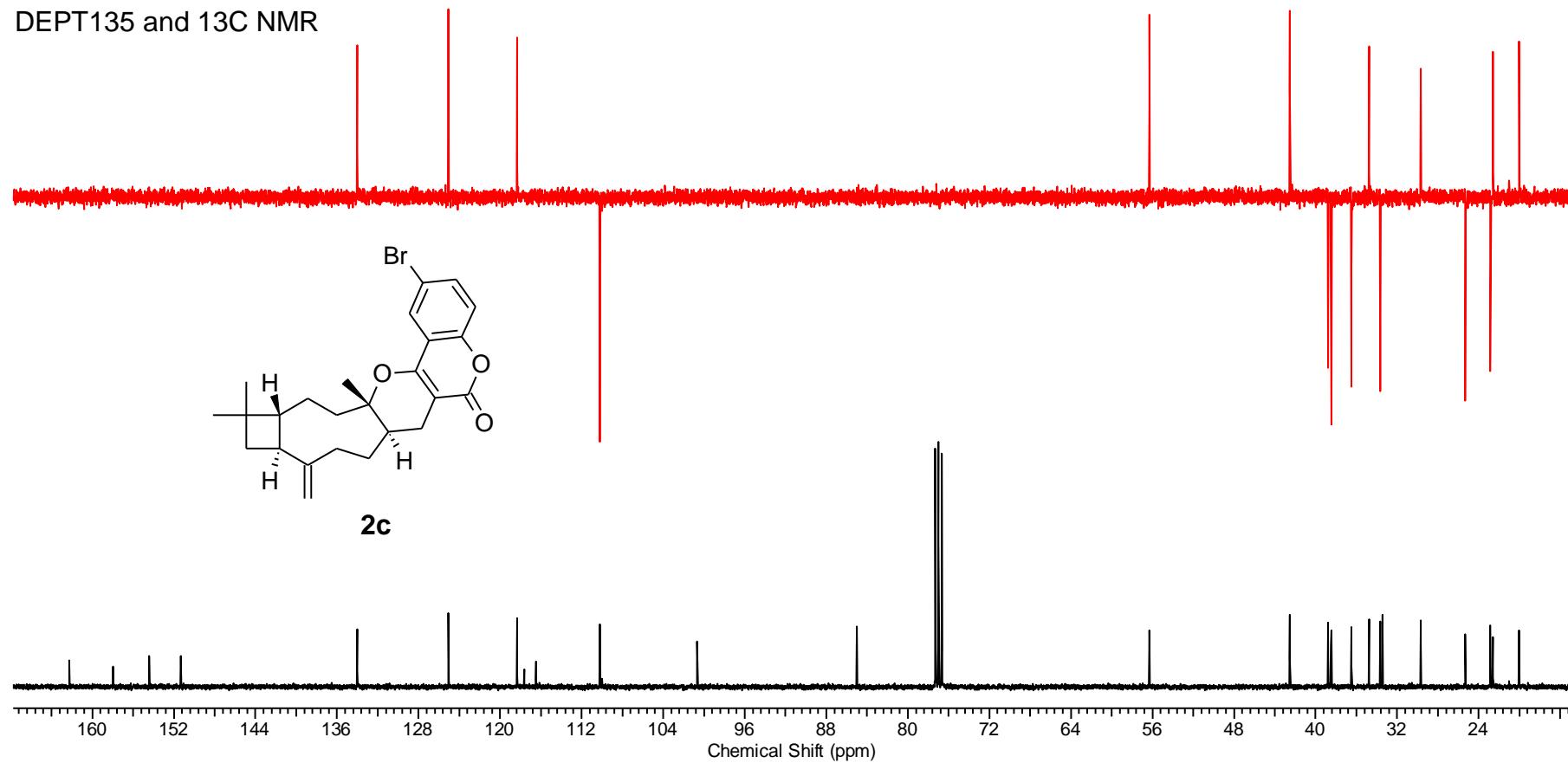


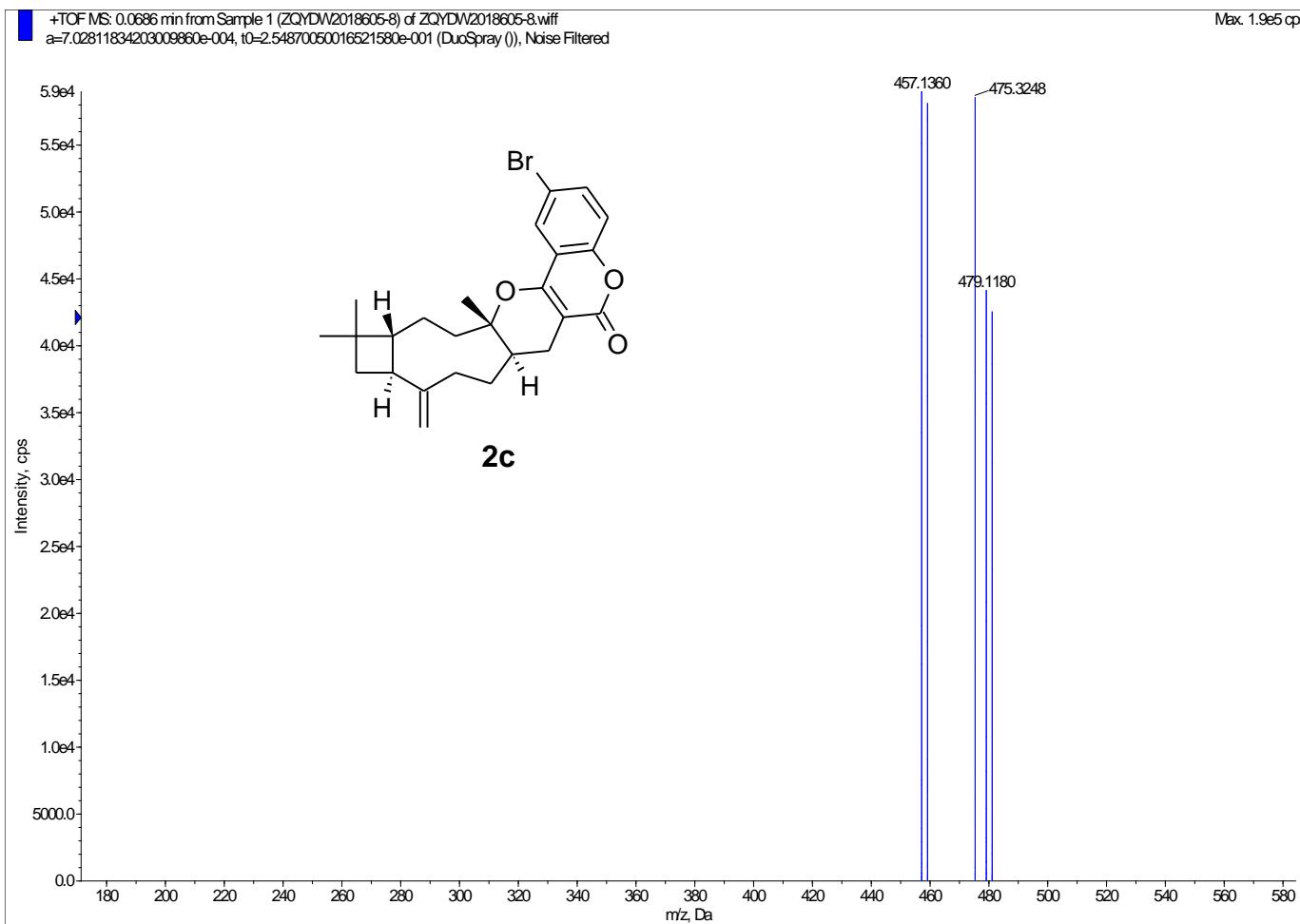
Compound 2c

2c $^1\text{H-NMR}$



DEPT135 and ^{13}C NMR

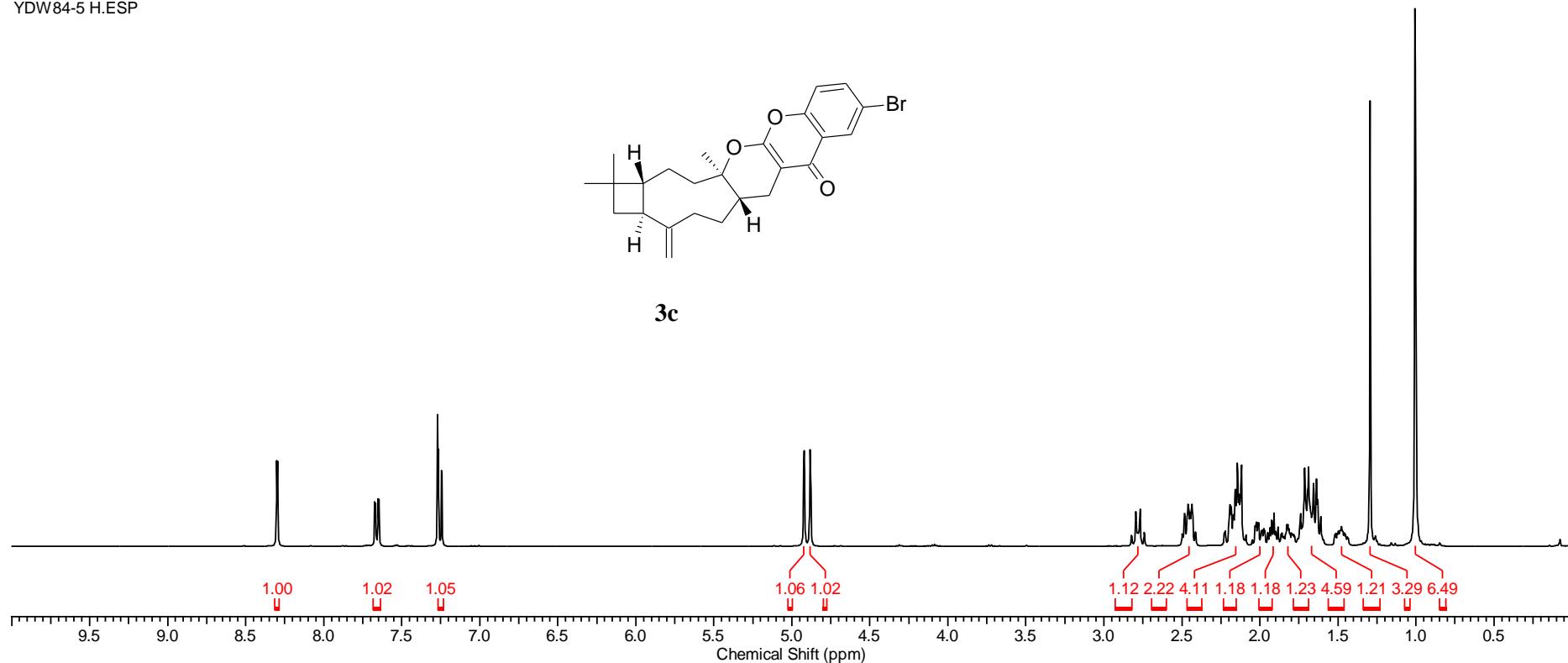




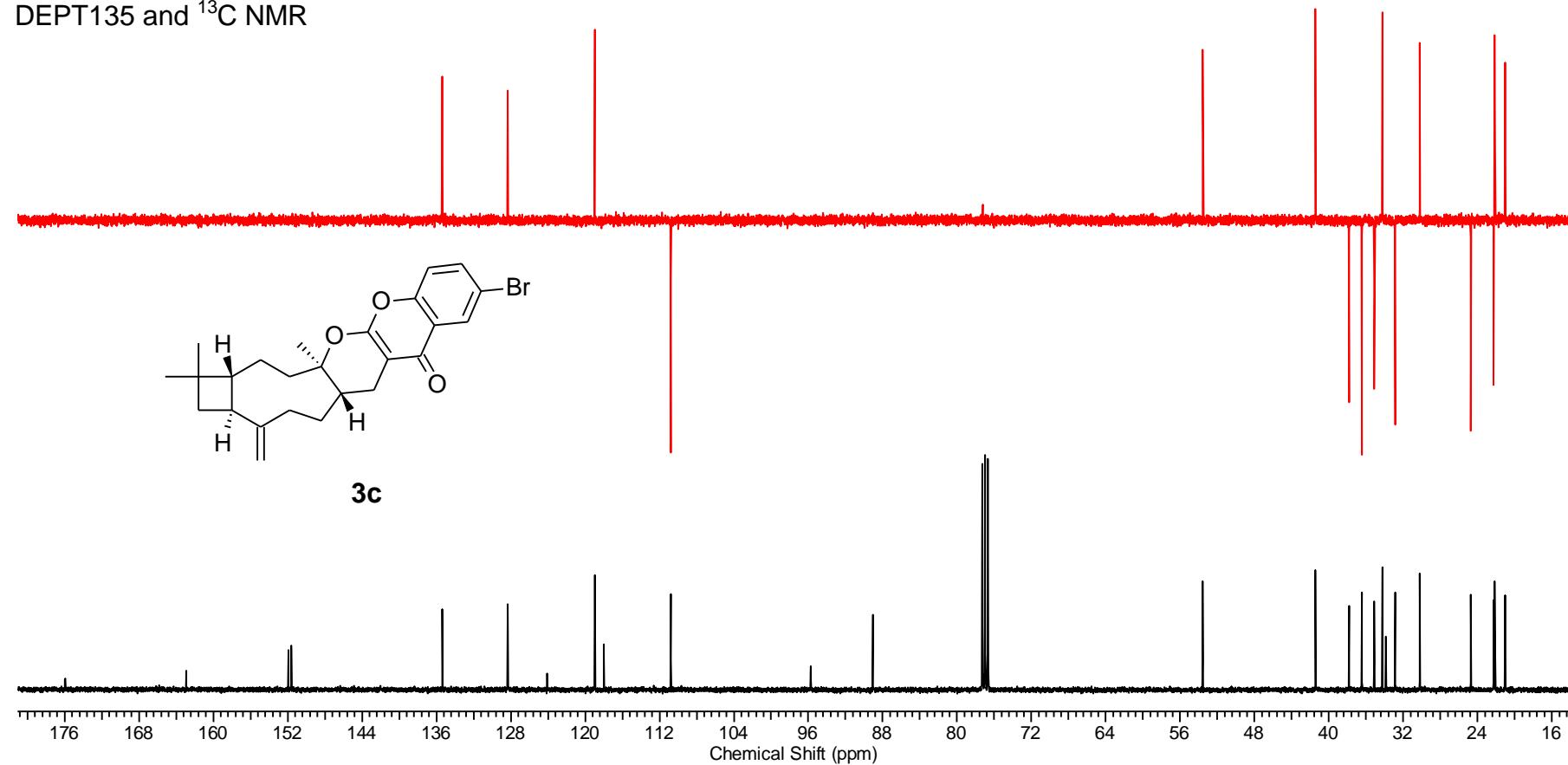
Compound 3c

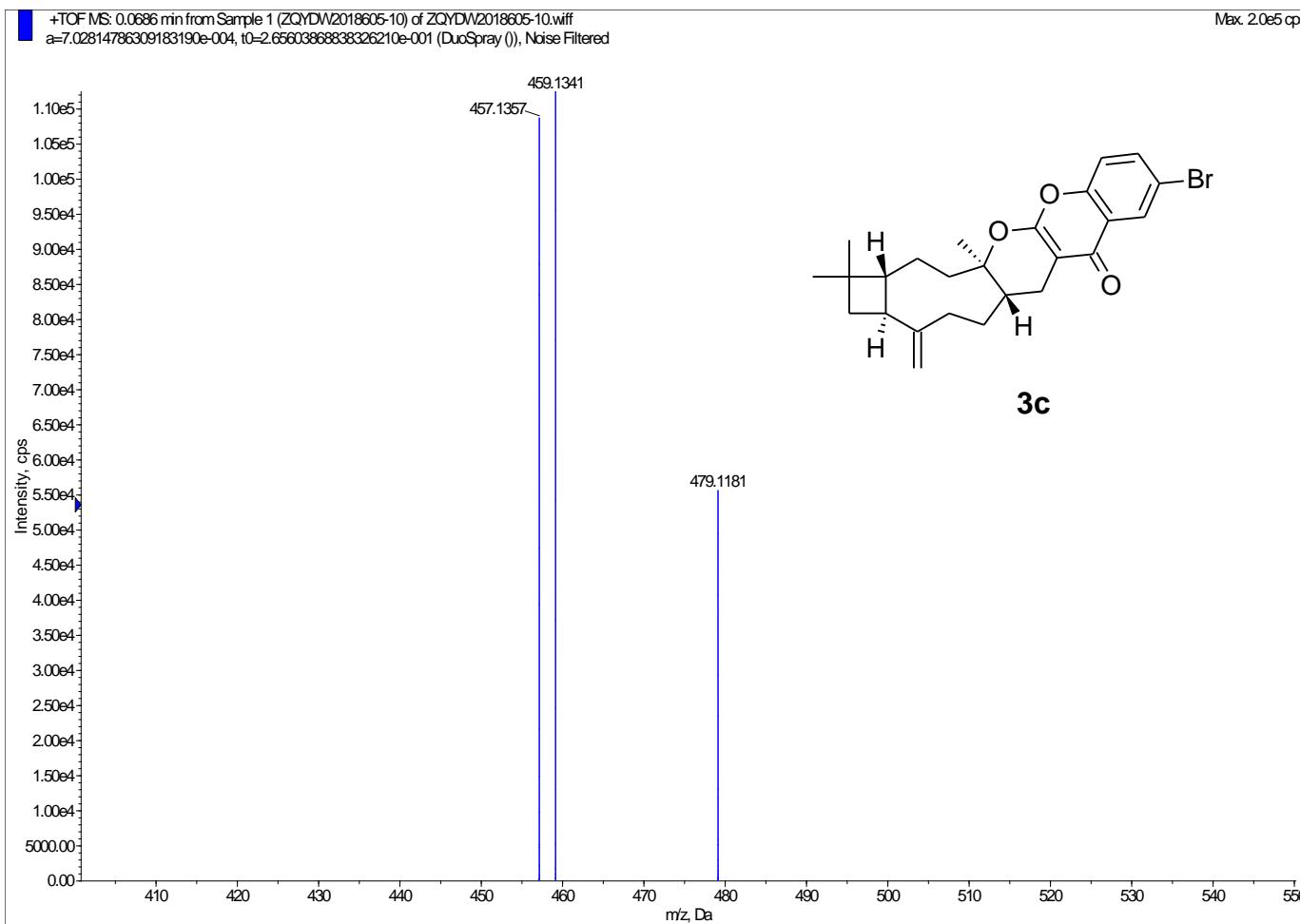
3c ^1H -NMR

YDW84-5 H.ESP



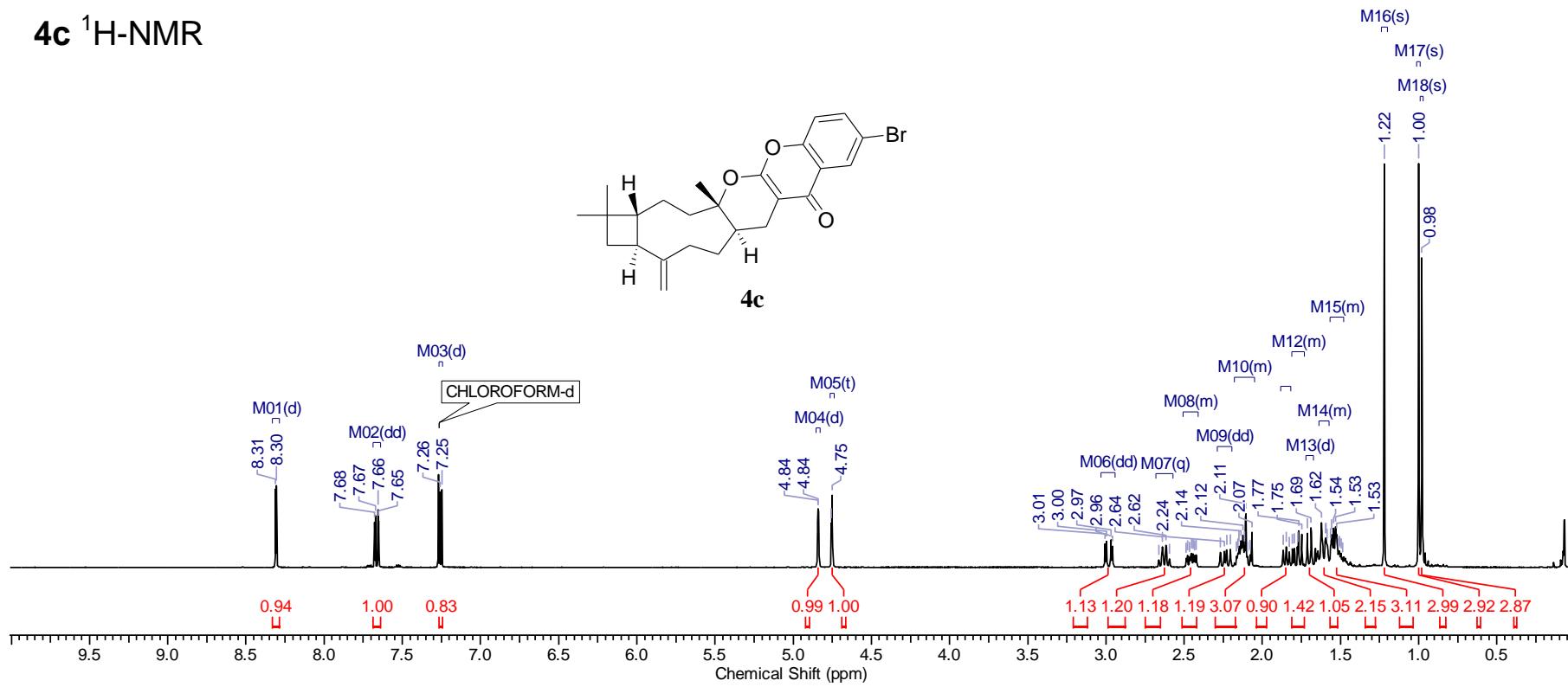
DEPT135 and ^{13}C NMR



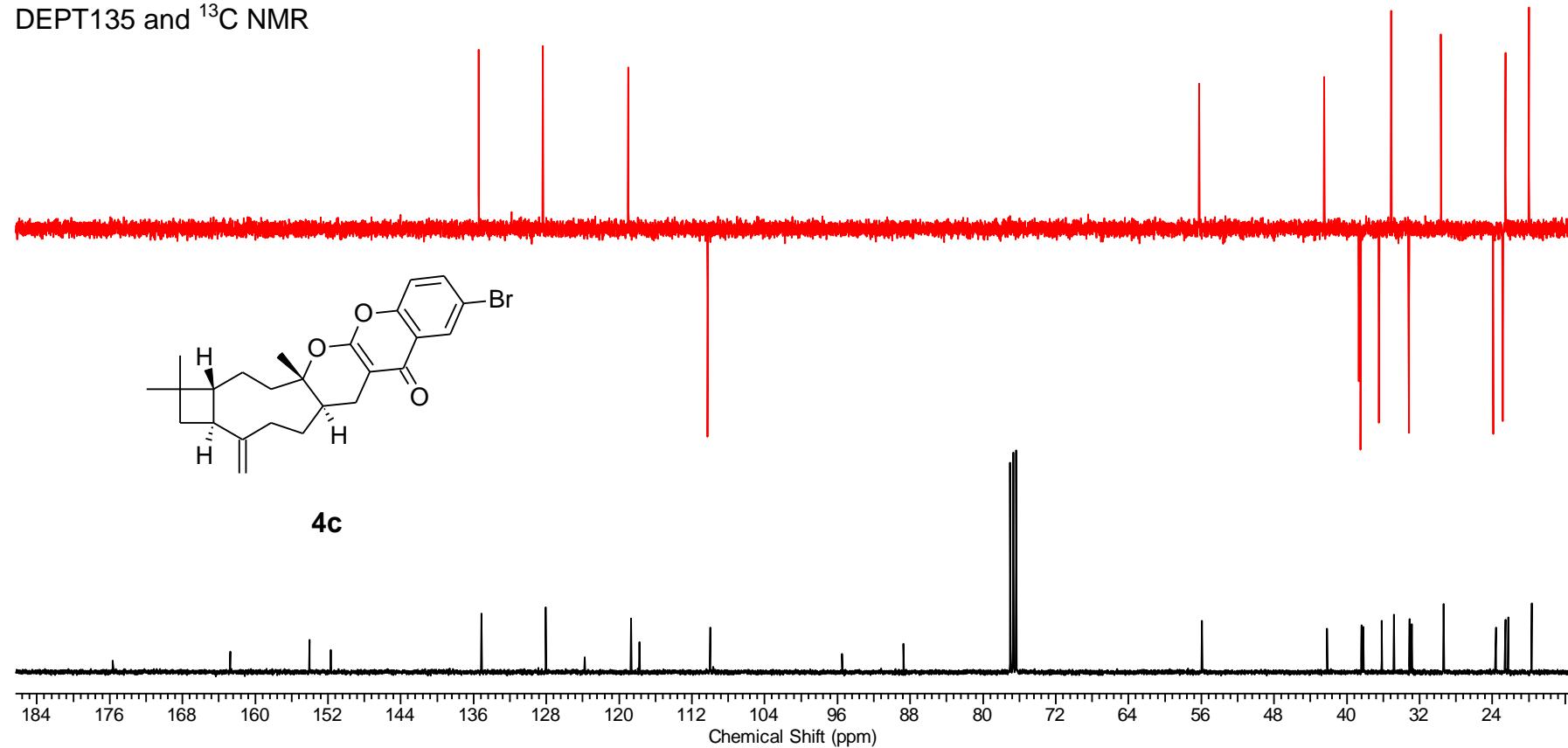


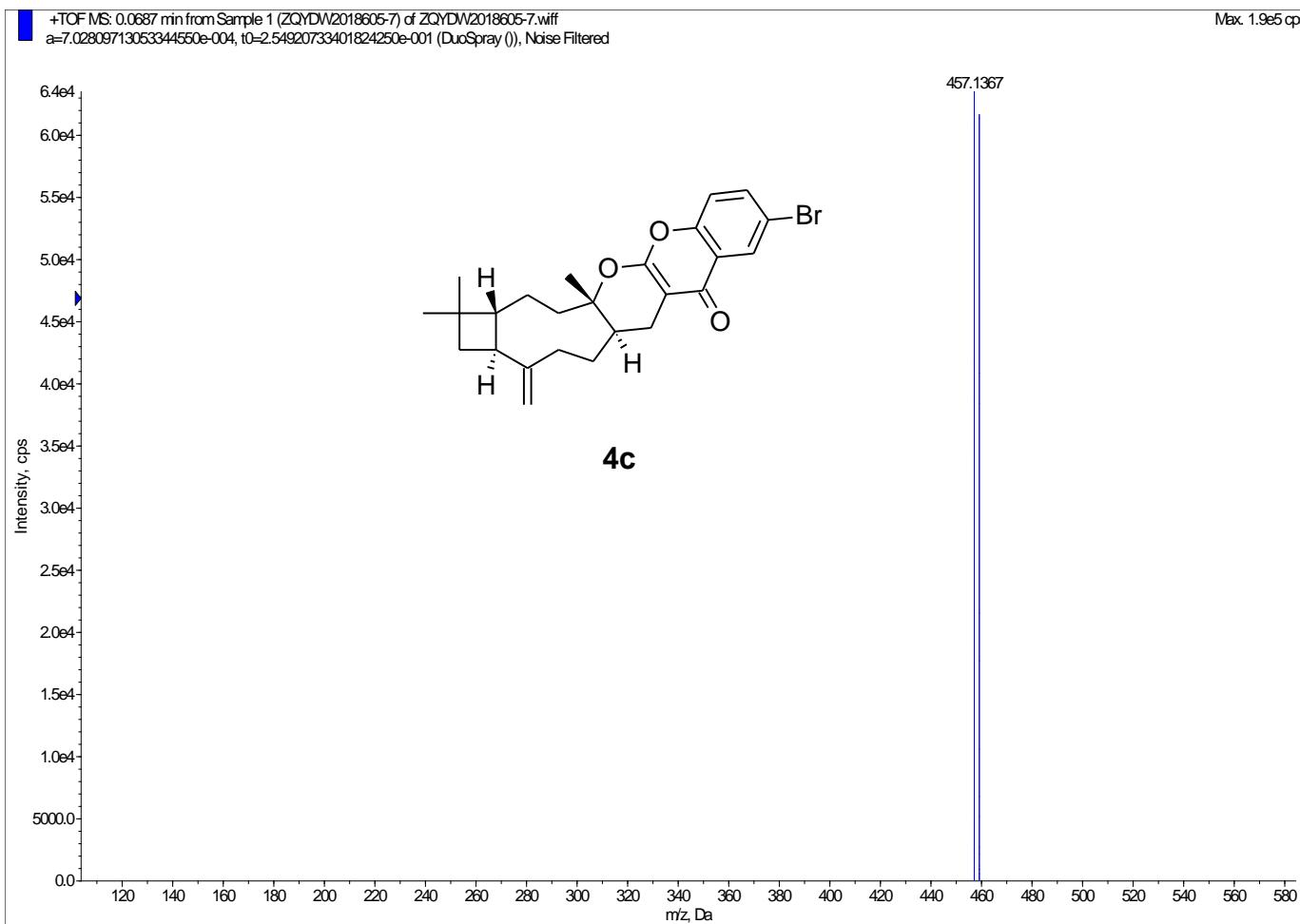
Compound 4c

4c ^1H -NMR



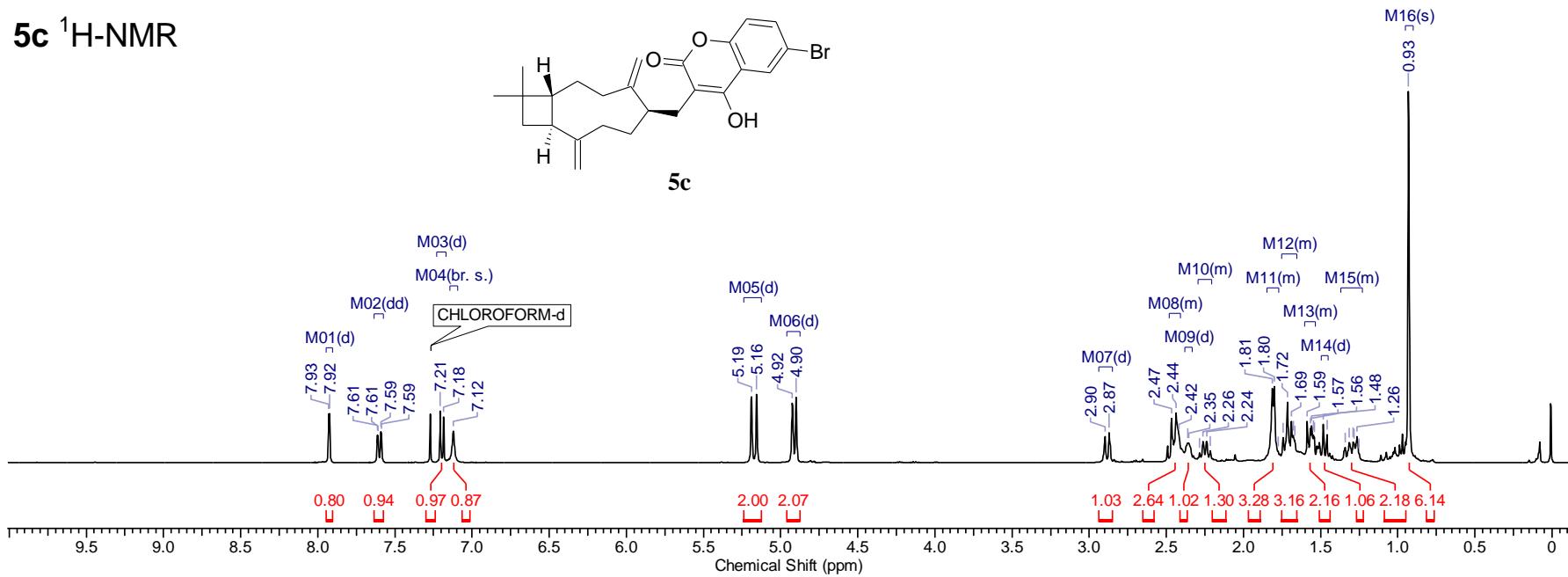
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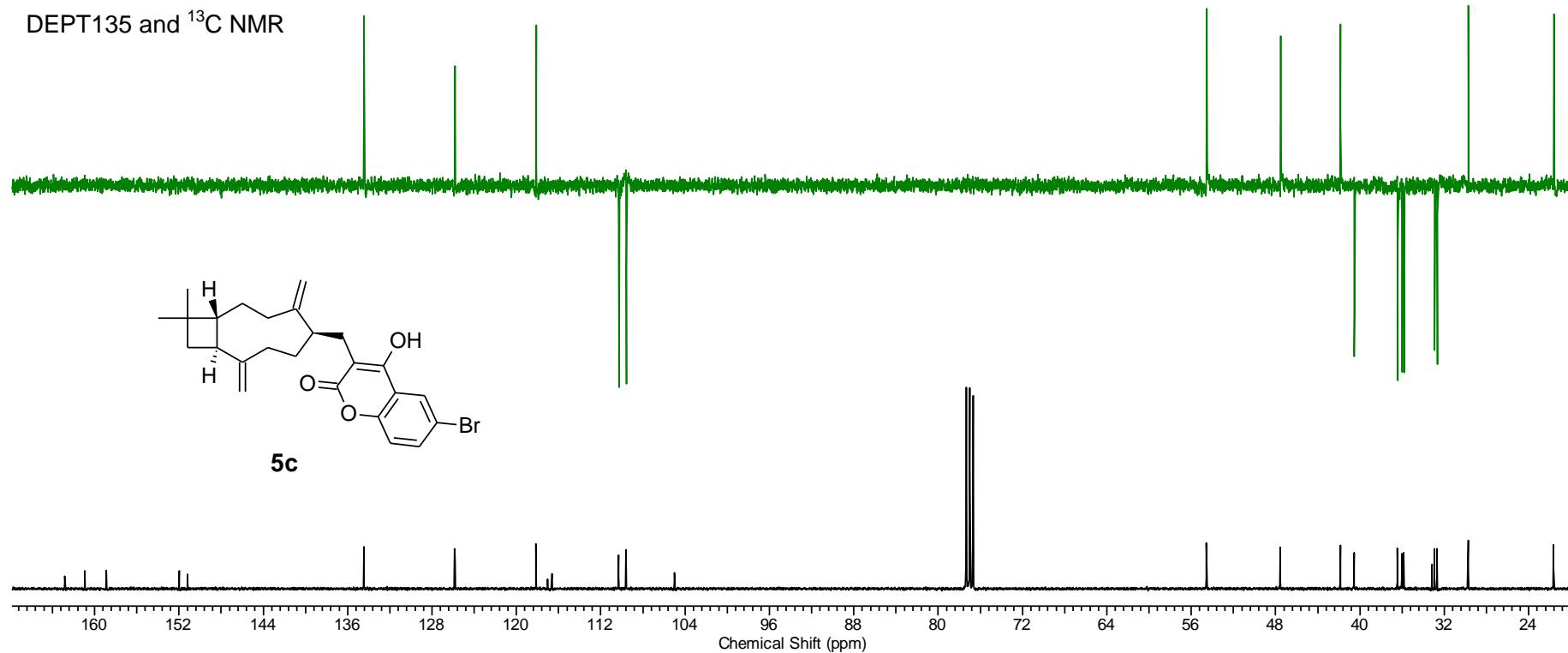


Compound 5c

5c ^1H -NMR

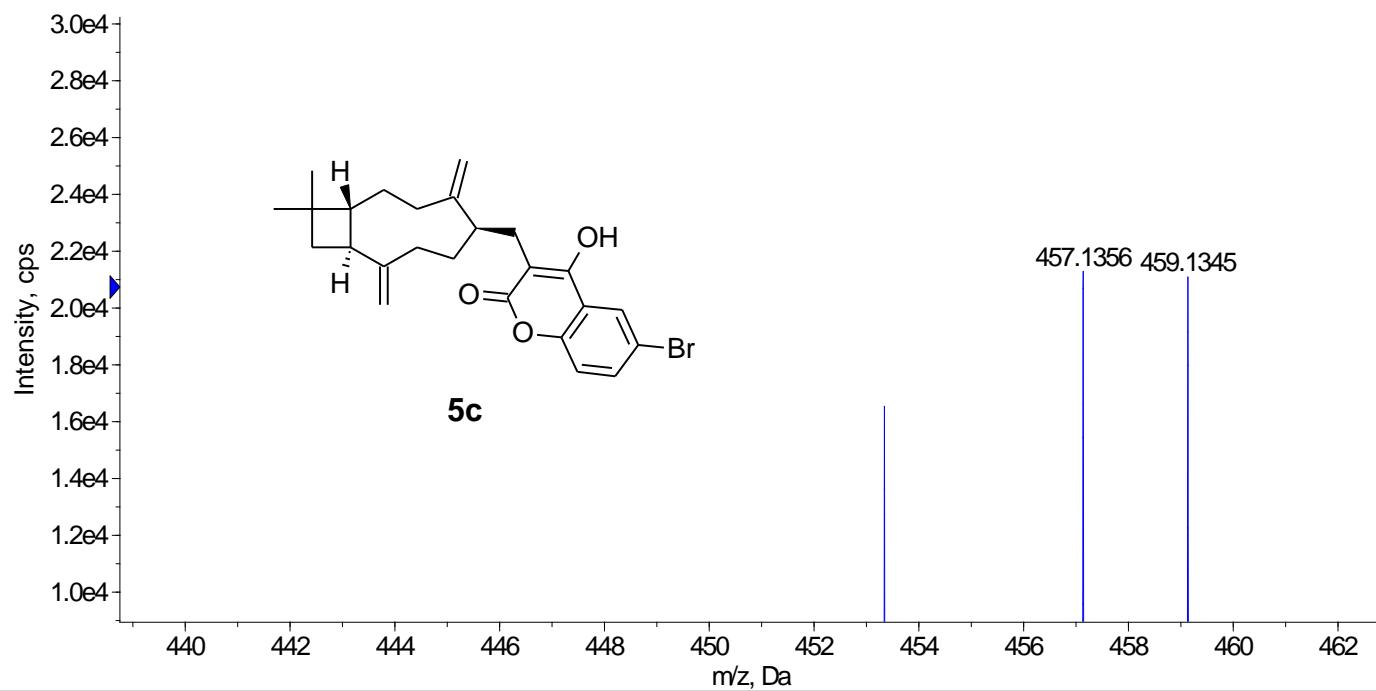


DEPT135 and ^{13}C NMR



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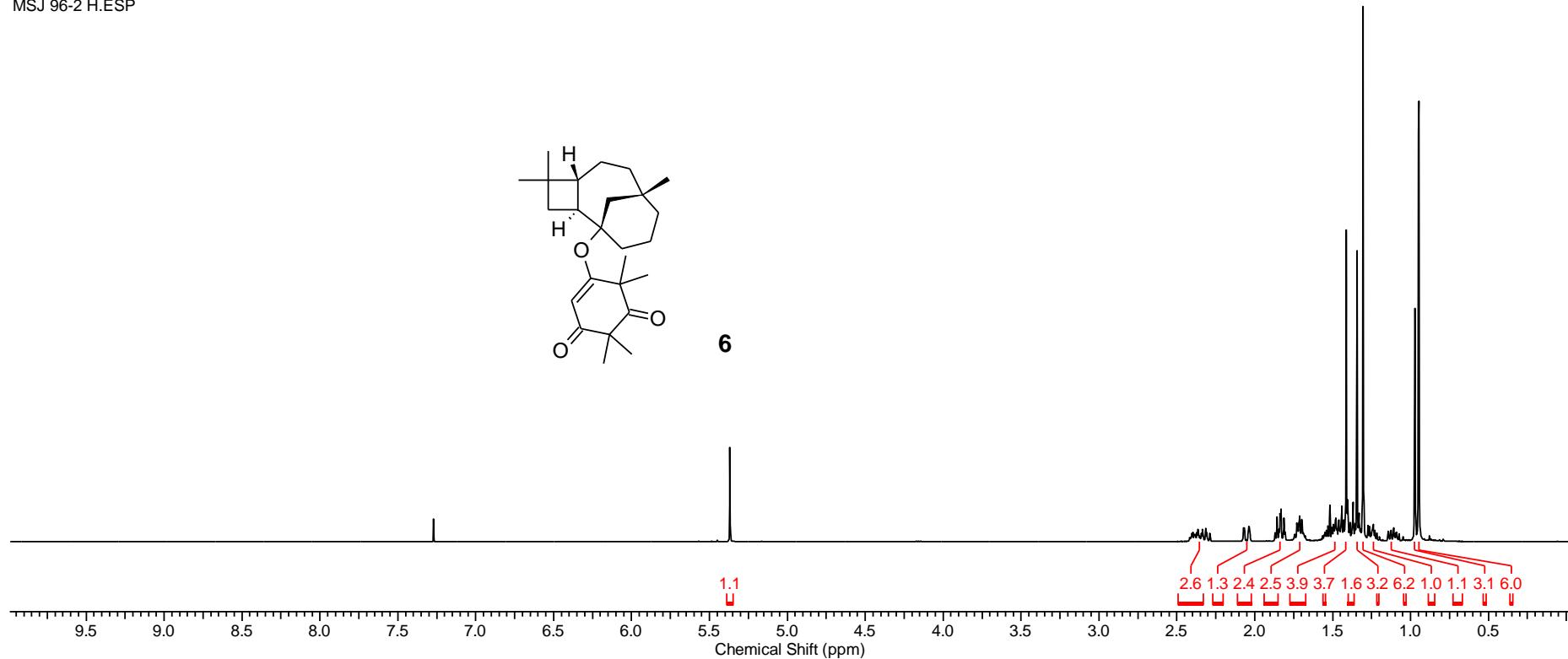
Max. 6.2e5 cps.



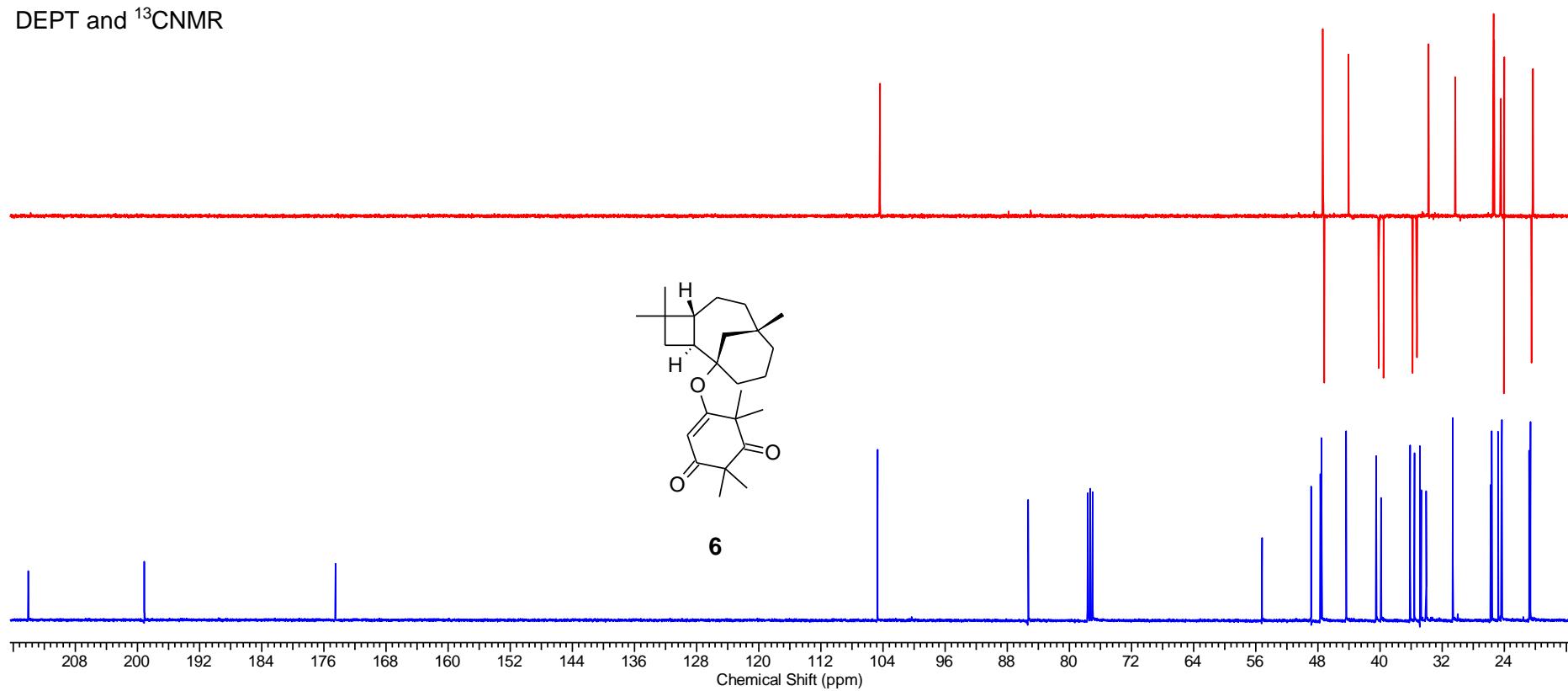
Compound 6

6 ^1H -NMR

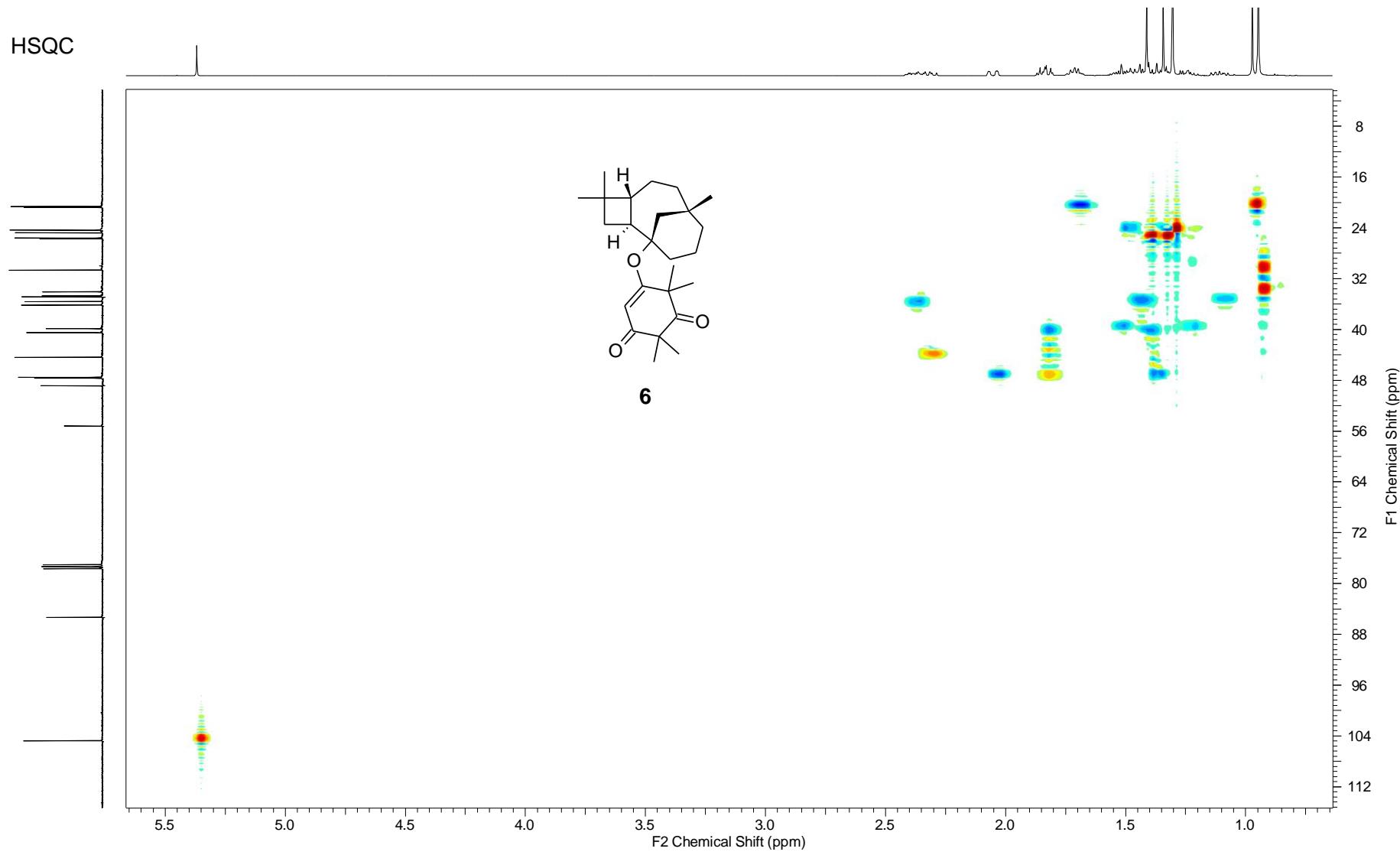
MSJ 96-2 H.ESP



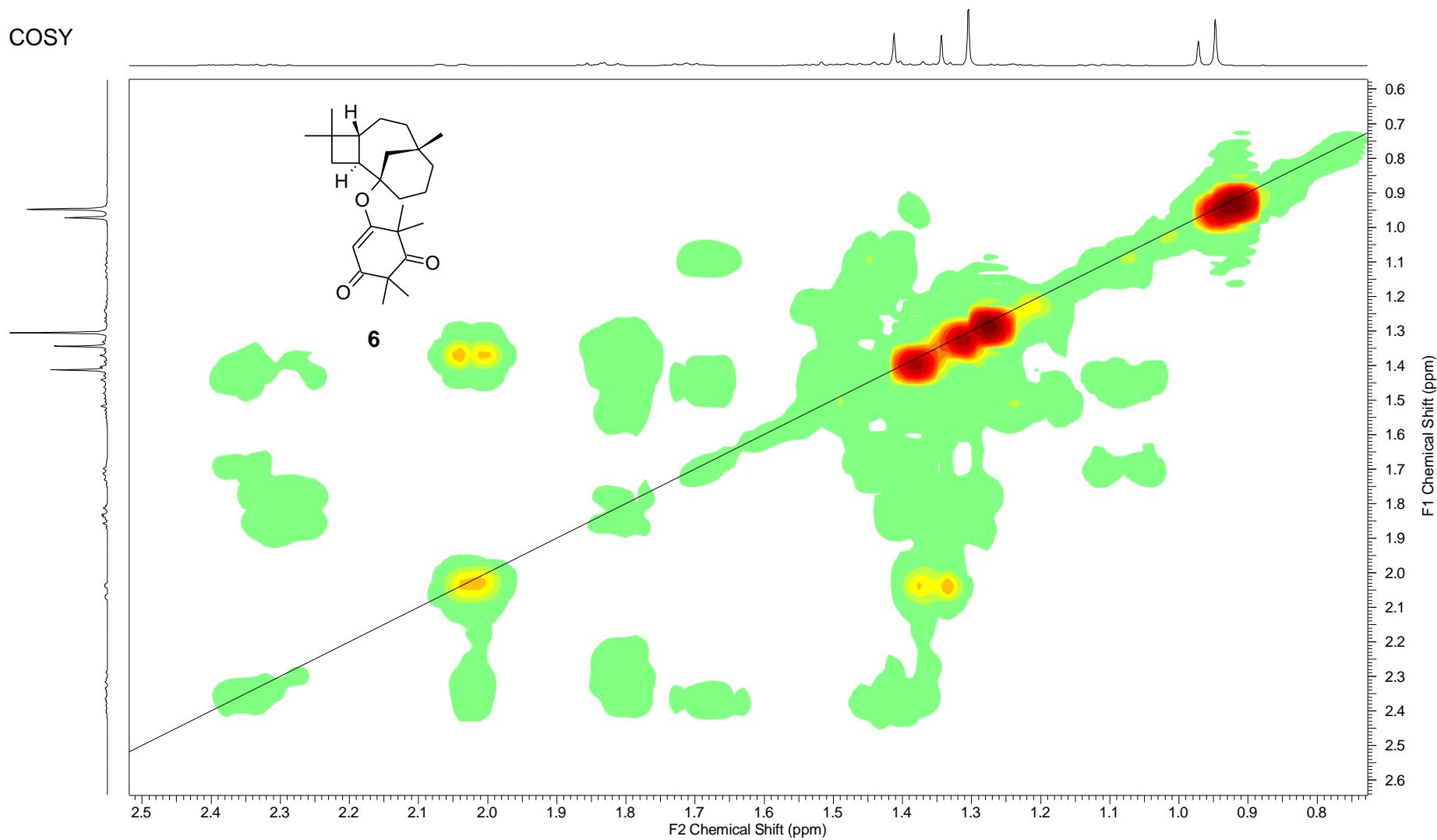
DEPT and ^{13}C NMR



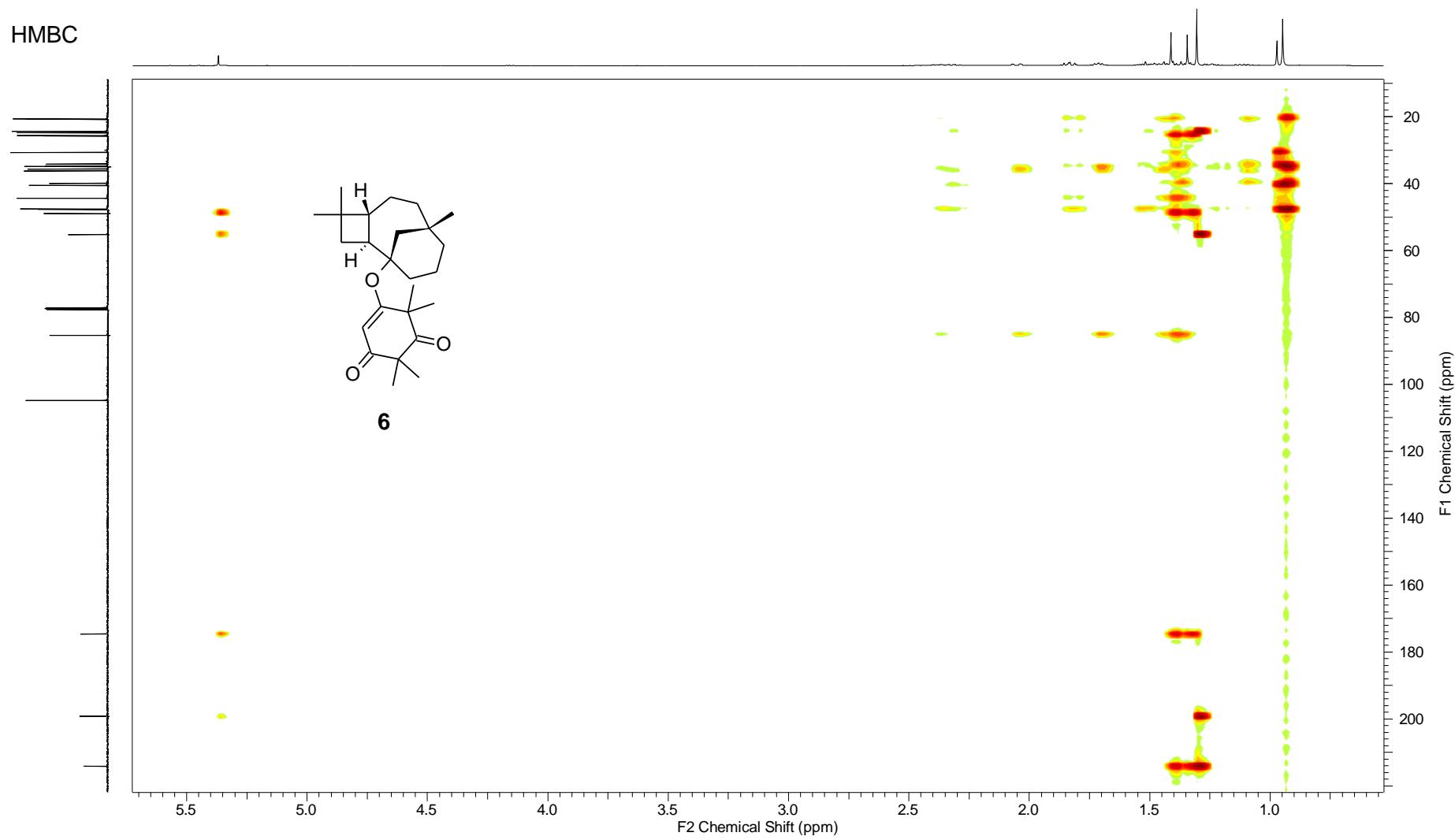
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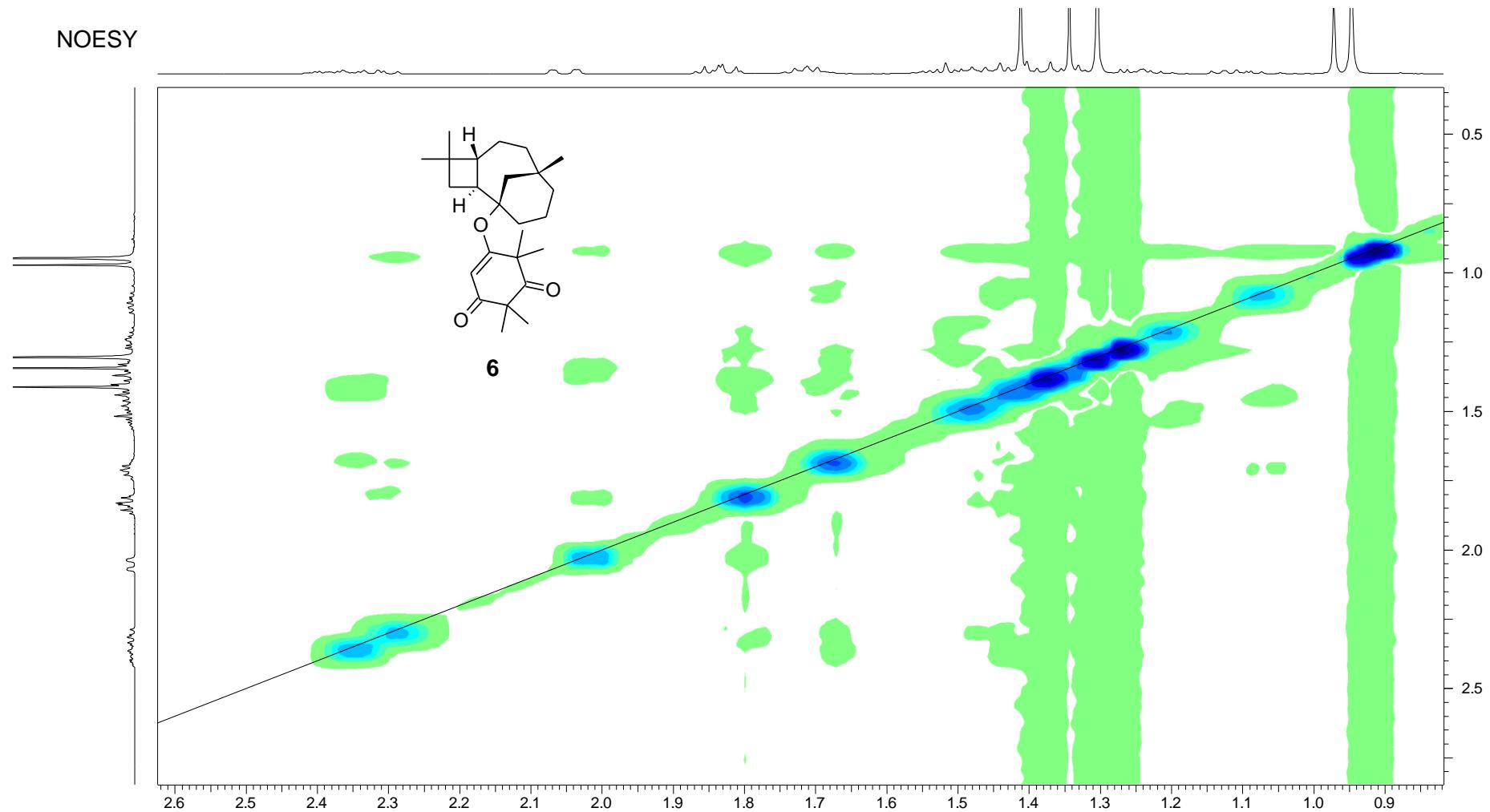
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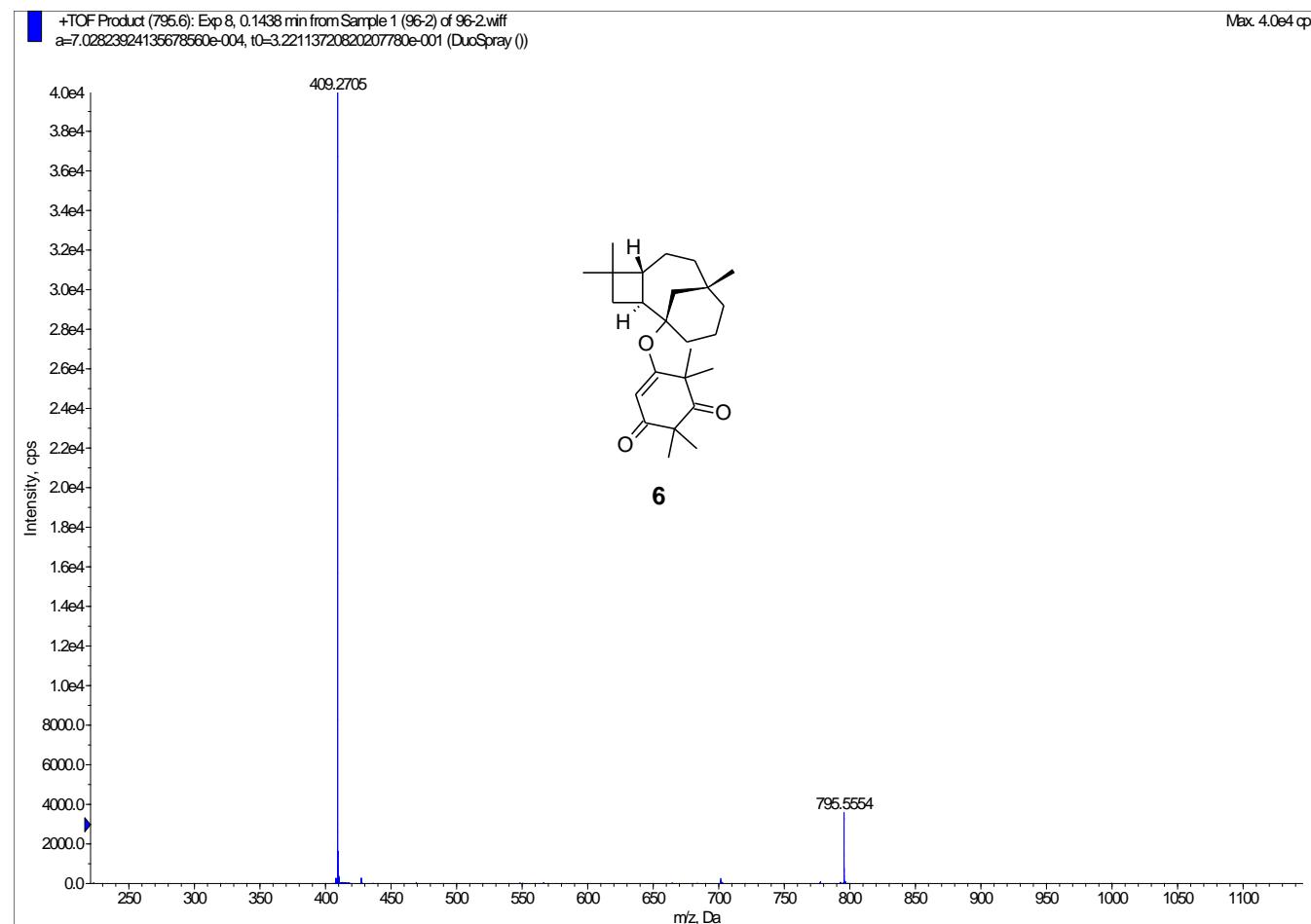
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NOESY

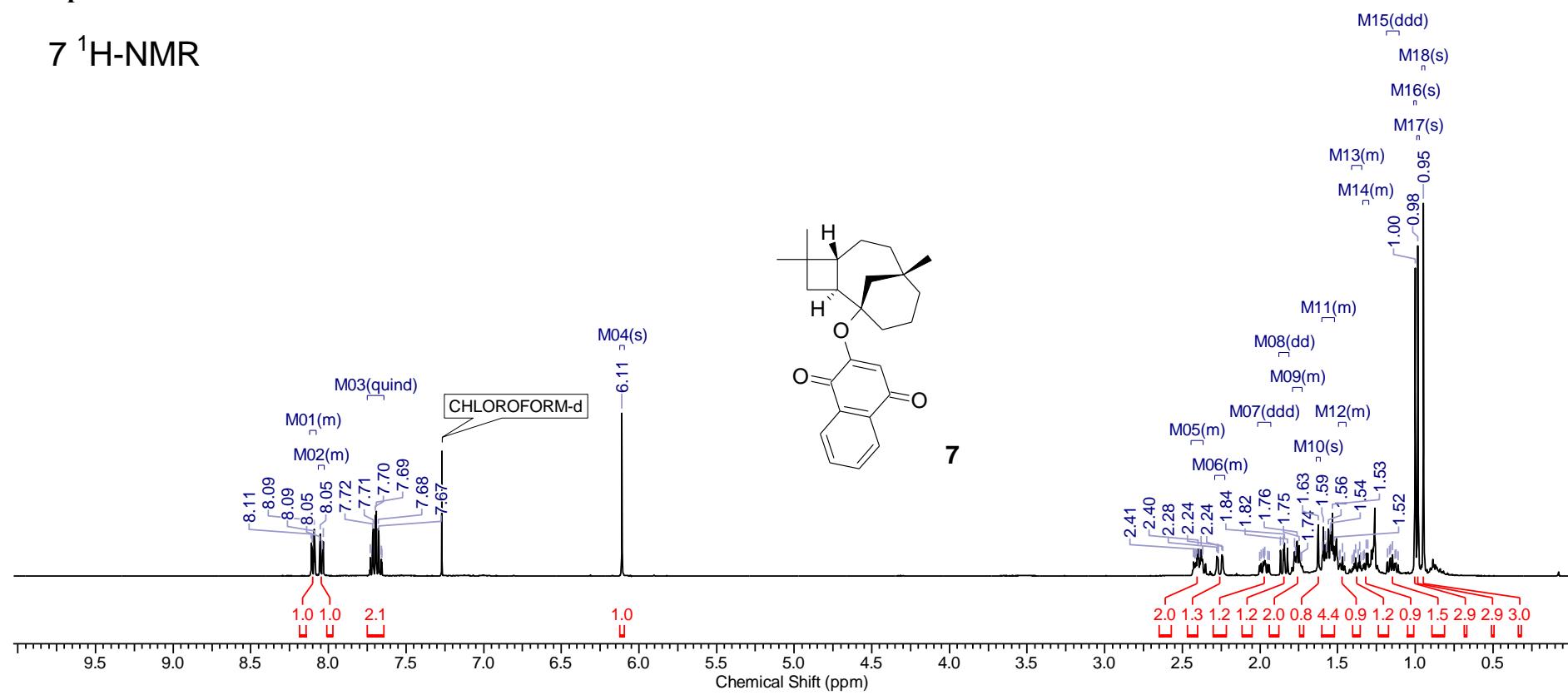


HR-ESI-MS

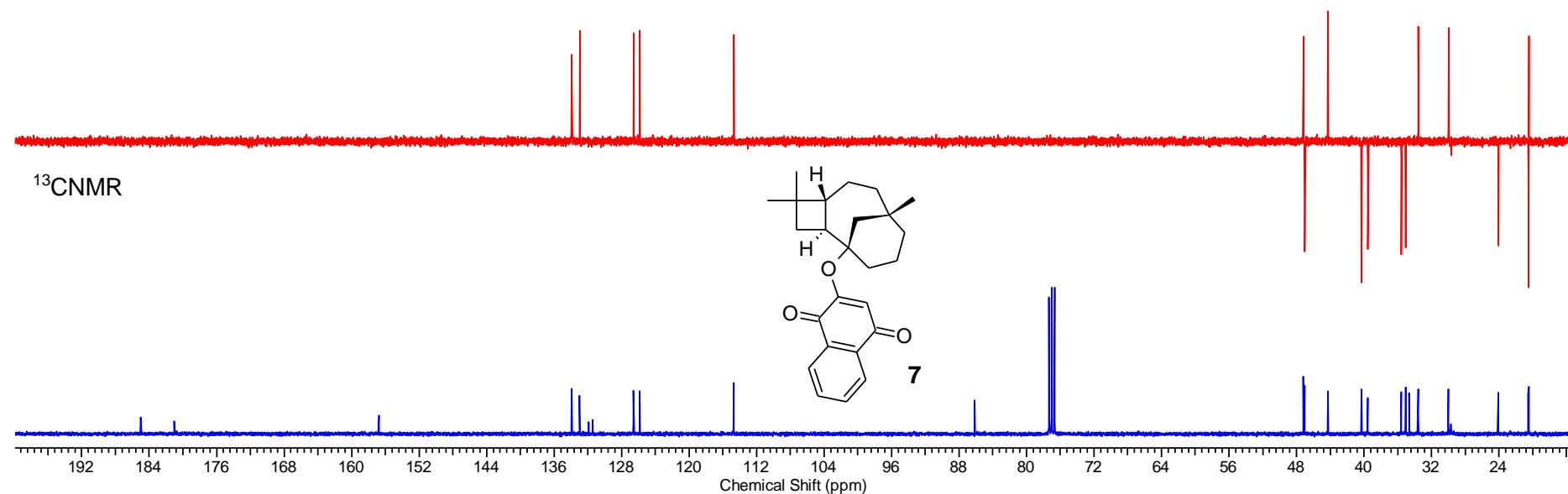


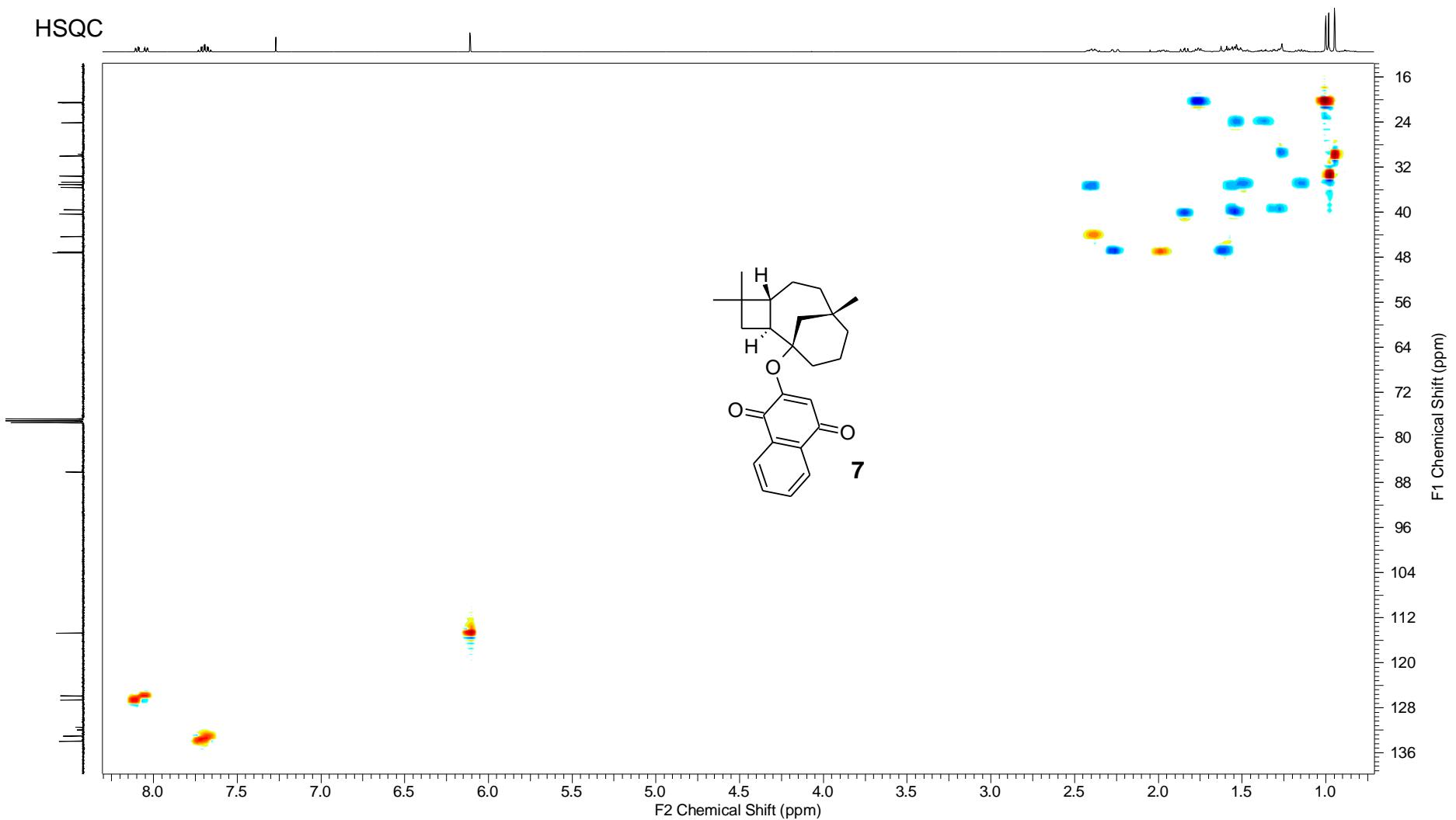
Compound 7

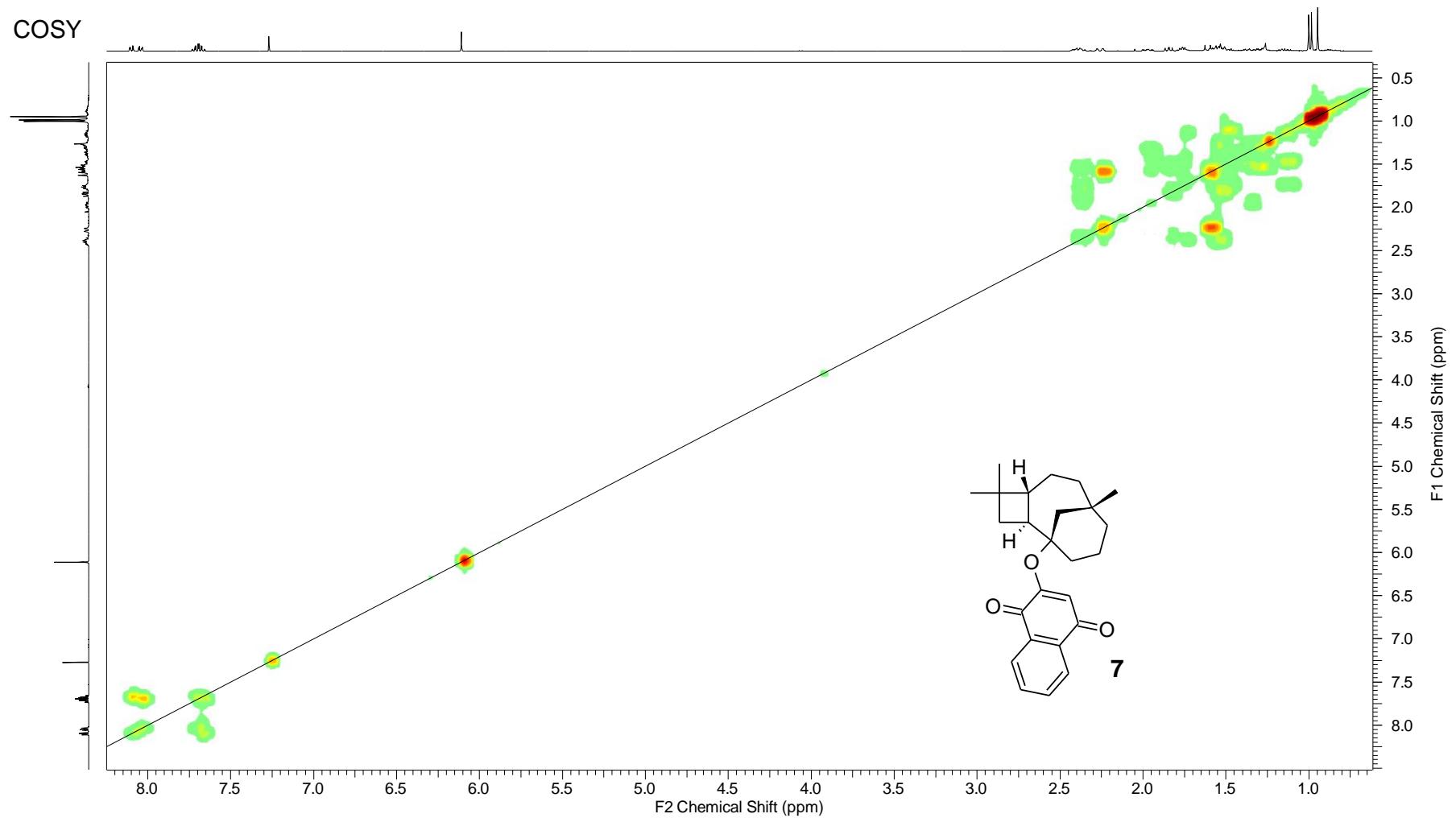
7 $^1\text{H-NMR}$

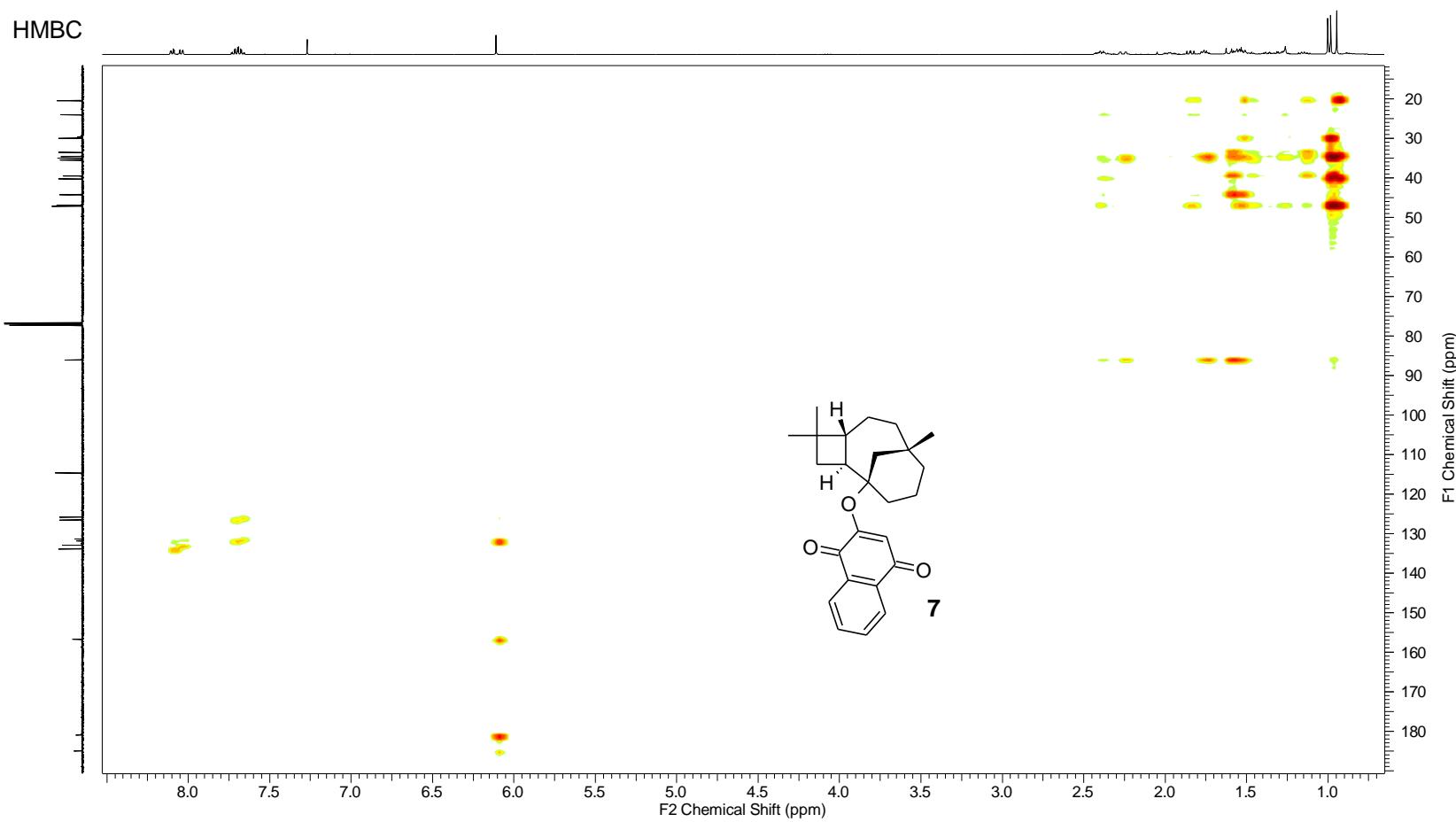


¹³C-NMR and DEPT135

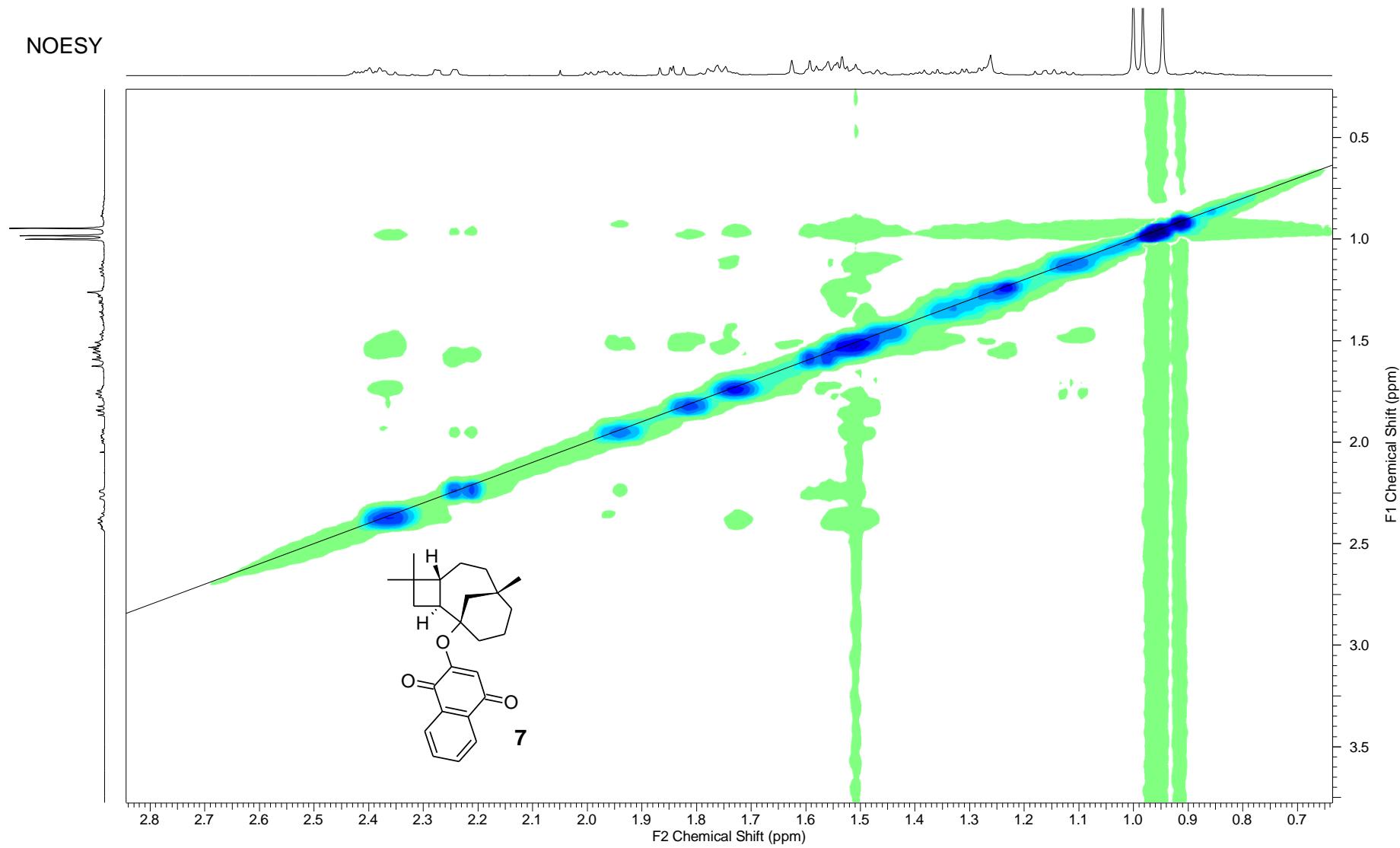








NOESY



HR-ESI-MS

