An R-Loop-initiated CSB-RAD52-POLD3 Pathway Suppresses ROS-induced Telomeric DNA Breaks

Tan J. et.al



Supplementary Figure 1. TRF2 and TERRA, but not TRF1, is required for ROS induced R-loop formation at the telomere.

(a-b) The staining of XRCC1 (a), pAR (a) and S9.6 (b) with Abs at KR/RFP-TRF1 in U2OS cells after light exposure for 30 min and recovery for 30 min. (c) The staining of S9.6 at KR-TRF1 and RFP-TRF1 after exposing to light for 30 min and recovery for 30 min in BJ, MEF and HeLa cells. (d) The staining of S9.6 at KR-TRF1 after treating with DRB (20 μ M, 24 h) and α -amanitin (100 μ g/ml, 2 h) in U2OS cells overexpressing HA-RNaseH WT or D210N mutant. (e) The staining of S9.6 at KR-TRF1 and FOK1-TRF1 in U2OS cells after light treatment. (f) Northern dot blot after TERRA KD with LNA in U2OS cells. (g) The staining of S9.6 at KR-TRF1 in U2OS cells after treating with TRF2 knockdown by siRNA. (h) The staining of S9.6 at KR-TRF2 in U2OS after TRF1 knockdown by siRNA. For (a, c, h), Mean values with SD from 30 cells in three independent experiments are given. P-value is calculated by unpaired t-test.



Supplementary Figure 2. CSB knockout leads to telomere shortening. (a) Western blot confirmed the CSB KO and KD efficiency in U2OS and 293 cells. (b) TRF analysis of average telomere length in 293 CSB KO cells after culturing for more than three months. (c) SA- β -gal staining of KR-TRF1 transiently-expressing in U2OS and U2OS CSB KO cells. Number SA- β -gal positive cells of per 300 cells were obtained from three independent experiments. Error bars represent SD, **P < 0.01, ***P < 0.001. (d) KR-TRF1 foci in U2OS CSB KO cells after culturing for more than three months. Quantification of the intensity of KR-TRF1 foci. The mean values of over 100 RFP-TRF1 foci with SD are showed. (e-f) The staining of CSB at KR-TRF1 in U2OS cells after treating with DRB (20 μ M, 24 h) and α -amanitin (100 μ g/ml, 2 h) and TERRA knockdown by LNA (D), overexpressed with HA-RNaseH WT, D210N mutant). (g) The staining of S9.6 at KR-TRF1 in U2OS cells after treating with CSB knockdown by siRNA.



Supplementary Figure 3. R464 of CSB contributes to DNA: RNA hybrid binding. (a) The recruitment of CSB fragment to KR-TRF1/RFP-TRF1 after light activation. (b) GFP-AD foci colocalization with KR-TRF1 in U2OS cells treated with α -amanitin (100 µg/ml, two h), overexpressed with HA-RNaseH WT, D210N mutant or TERRA depleted with LNA. (c) The recruitment of GFP-AD mutant to KR-TRF1. (d) GFP-AD WT, ADN and ADC co-localization with KR-TRF1 after light activation. (e) The recruitment of ADC WT and ADC R464A to KR-TRF1. (f) Coomassie blue staining of GST-ADC WT and R464A.



Supplementary Figure 4. RAD52 is recruited to telomeric R-loop. (a) The clearance of γ -H2AX after RAD52 KD by siRNA. (b) The recruitment of RAD52 to RFP/KR/FOK1-TRF1after light activation or transfection. (c) The recruitment of RAD52 to KR-TRF1 after treating with DRB (20 μ M, 24 h) and α -amanitin (100 μ g/ml, 2 h), TERRA knockdown by LNA, or overexpressed with HA-RNaseH WT, D210N mutant. (d) TRF1 foci in U2OS RAD52 KO cells after culturing for more than three months. Quantification of the intensity of KR-TRF1 foci. Error bar represents over 100 KR-TRF1 foci. (e) SA- β -gal staining of KR-TRF1 transiently-expressing in U2OS and U2OS RAD52 KO cells. Number SA- β -gal positive cells (n = 300) were obtained from three independent experiments, Error bars represent SD, **P < 0.01, ***P < 0.001.



Supplementary Figure 5. The recruitment of RAD52 to damaged telomere is dependent on CSB and DNA: RNA binding. (a) The recruitment of RAD52 to damaged telomere in U2OS CSB KO cells after light exposure for 30 min and 60 min. (b) Polymer structure of RAD52. (c) U2OS cells expressing with GFP-RAD52 K141A or K144A were laser-micro-irradiated and monitored by live cell imaging. (d) ssRNA EMSA assay of purified RAD52 WT and RAD52 K144A protein.



Supplementary Figure 6. The recruitment of POLD3 to damaged telomere requires RAD52.

(a) The recruitment of RAD51 to FOK1-TRF1 after transfection. (b) The recruitment of RAD51 to KR-TRF1 after inducing damage for 30 min and recovery for 24 hours in U2OS cells with CSB or RAD52 knockdown by siRNA. (c) The recruitment of POLD3 to KR-TRF1 in U2OS RAD52 KO cells after transient expressing RAD52 mutants. (d) The recruitment of ERCC1 and XPF to KR-TRF1 after light activation.



Supplementary Figure 7. RAD52 is required for the loading of POLD3 to damaged telomeres. (a) The recruitment of POLD3 to KR-TRF1 in U2OS RAD52 KO cells after transient expressing RAD52 mutants. (b) The clearance of γ -H2AX after POLD3 knockdown by siRNA

Supplementary Table 1:

Sequence-Based Primer	
AD-R464A-Rev	CCTTAACGCCTGCTTATAATAATCTTCATC
AD-R464A-For	AAGCAGGCGTTAAGGAGATGGAATAAACTG
AD-RR466, 467AA-Rev	ATTCCAAGCGGCTAACCGCTGCTTATAATAATC
AD-RR466, 467AA-For	CGGTTAGCCGCTTGGAATAAACTGAGACTGCAG
AD-3RA-Rev	ATTCCAAGCGGCTAACGCCTGCTTATAATAATCTTCATC
AD-K470A-For	TGGAATGCGCTGAGACTGCAGGACAAAGAG
AD-R472A-Rev	CTGCAGAGCCAGTTTATTCCATCTCCTTAAC
AD-R472A-For	AAACTGGCTCTGCAGGACAAAGAGAAACGTC
RAD52 K141A-For	TTGGAGGCCGCAAGGAAGGAGGCGGTGACAGACGGGC
RAD52 K141A-Rev	CGCCTCCTTCCTTGCGGCCTCCAAAGATAAAGCCTTG
RAD52 K144A-For	TTGGAGAAGGCAAGGGCCGAGGCGGTGACAGACGGGC
RAD52 K144A-Rev	CGCCTCGGCCCTTGCCTTCTCCAAAGATAAAGCCTTG
RAD52 Y65A-For	TAAGTAGCGCCATGGCTGGCGGAGGCCAGA
RAD52 Y65A- Rev	GCCAGCCATGGCGCTACTTATGTATTCTGGGC
RAD52 K152A-For	ACGGGCTGGCGCGAGCCCTCAGGAGTTTT
RAD52 K152A-Rev	AGGGCTCGCGCCAGCCCGTCTGTCACC
RAD52 K153A-For	GGCTGAAGGCAGCCCTCAGGAGTTTTGGGAA
RAD52 K153A-Rev	CTGAGGGCTGCCTTCAGCCCGTCTGTCACC
RAD52 K156A-For	CGAGCCCTCGCGAGTTTTGGGAATGCACTTGG
RAD52 K156A-Rev	CCAAAACTCGCGAGGGCTCGCTTCAGCCCG

Supplementary Table 2:

Sequence-Based Reagents	Source
TERRA FISH probe: (TAACCC)7-Alexa488-30	Integrated DNA Technologies
LNA gapmer Scr: 5'-CACGTCTATACACCAC-3'	Exiqon
LNA gapmer TERRA: 5'-TAACCCTAACCCTAAC-3'	Exiqon
TERRA probe for Northern blot : 5'-TAACCCTAACCC TAACCCTAACCCTAACCC-3'	Integrated DNA Technologies
GAPDH probe for Northern blot : 5'-GTAGACCCACGACA TACTCAGCACCGGCCTCACCCCATT-3'	Integrated DNA Technologies
Oligo 1: 5'-ATCATCACCATAACGTCGATGTATCAA CTTCGATTAGTCACACCAATTAA-3'	Integrated DNA Technologies
Oligo 2: 5'-UUAAUUGGUGUGACUAAUCGAAGUUGAUA CAUCGACGUUAUGGUGAUGAU-3'	Integrated DNA Technologies