Immunity, Volume 52

Supplemental Information

Precursors for Nonlymphoid-Tissue Treg Cells

Reside in Secondary Lymphoid Organs and Are

Programmed by the Transcription Factor BATF

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Figure S1. ATAC-seq and RNA-seq of tissue-derived T cells, related to Figure 1. (A) Lungderived KIrg1⁻ST2⁻ Treg cells as well as lung KIrg1⁺ST2⁺ tisTregST2 cells were FACS-sorted and subjected to ATAC-sequencing. Samples were normalized and reads for each sample in the "core tisTregST2" peak set derived from Figure 1D were counted. Heatmap with unsupervised clustering shows normalized ATAC signal with color code high (red) vs low (blue) (n=3-4). (B) Batf1-3 gene expression in BM and spleen-derived Klrq1⁻ST2⁻ Treg (black) as well as lung, liver, visceral adipose tissue (VAT), and skin-derived Klrg1⁺ST2⁺ tisTregST2 (brown, green, orange and dark blue). Statistics based on Deseg2. Asterisks indicate statistical significance, with Benjamini-Hochberg correction, ***p<0.001, **p<0.01, *p<0.05 and ns p>0.05) (n=3-4). (C) Gata1-6 gene expression in BM and spleen-derived Klrg1⁻ST2⁻ Treg (black) as well as liver, lung, VAT, and skin-derived Klrg1⁺ST2⁺ tisTregST2 (brown, green, orange, and dark blue). Colors indicate cell type (n=3-4). (D) ATAC-seq data for the KIra1. Ppara. Mreg and Dusp26 gene loci from lymph node (LN)-derived CD25⁺Foxp3(GFP)⁺CD44⁻ naive Treg, LN-derived CD25⁺Foxp3(GFP)⁺CD44⁺ memory Treg as well as VAT, lung, skin and colon-derived CD25⁻Foxp3(GFP)⁻CD44⁺ memory Tconv or CD25⁺Foxp3(GFP)⁺CD44⁺Klrg1⁺ST2⁺ tisTregST2 (n=4). Y-axis ATAC signal intensity, x-axis gene structure, with exons indicated as heightened bars and introns as line, arrows indicate gene direction. All datasets group-normalized to maximum peak height indicated in brackets. Overall display length indicated on top in kilobases (kb). Yellow box indicates area of interest. Data representative of two or more independent experiments or cell sorts.



Figure S2. Nfil3-GFP reporter mouse, related to Figure 2. (A) Overview of insert into the BAC construct used for generating the Nfil3GFP reporter mouse. A fusion construct composed of the DNA for iCre (red), 2A (grey), and eGFP (green) was generated, and inserted at the start codon of the Nfil3 gene in the BAC RP23-227M5 using Escherichia coli DH10B. Top, gene structure and insertion site. Bottom, DNA code inserted into BAC. (B) Gating strategy used to identify Treg cells in Nfil3^{GFP} reporter or control mice (data derived from lymph node). G1: CD4⁺ T cells; G2: single cells; G3: CD8⁻CD19⁻MHCII⁻Dead⁻TCRbeta⁺ T cells; G4: CD4⁺CD8⁻ T cells; G5: CD4⁺CD25⁺ Treg cells. (C) Expression of Nfil3(GFP) in Klrg1⁺ST2⁺ tisTregST2 (G2) vs Klrg1⁻ST2⁻ Treg cells (G1) from various tissues (paired t test, ing LN = inguinal LN; mes LN = mesenteric LN. n=7). (D) Presence of Klrg1⁺Nfil3(GFP)⁺ Treg cells (red gate), Klrg1⁻Nfil3(GFP)⁺ Treg cells (light blue gate), and Klrg1-Nfil3(GFP)- Treg cells (black gate) in different cell populations. First row: CD4+CD25+ Trea cells: Second row: CD4⁺CD25⁻ Tconv cells: Third row: CD8⁺ cvtotoxic T cells: Fourth row: CD19⁺ B cells. Contour plots to the left show presence of populations in spleen or skin. Additional data points, tissues and statistical evaluation shown in the graphs to the right (one-way ANOVA with Tukey correction, n=7). (E) Memory phenotype of Klrg1⁻Nfil3(GFP)⁻ Treg cells (black), Klrg1⁻ Nfil3(GFP)⁺ Treg cells (blue) and Klrg1⁺Nfil3(GFP)⁺ Treg cells (red). Left, histograms illustrating expression of CD62L in spleen-derived Treg cell populations. Middle, additional data points, tissues and statistical evaluation (one-way ANOVA with Tukey correction, n=7). Right, CD44 MFI for Treg populations in spleen, mes LN and lung (one-way ANOVA with Tukey correction, n=7). Data representative of two or more independent experiments or cell sorts.



Figure S3. ScRNA-seq and scTCR-seq of tissue T cells, related to Figure 2+3. (A) t-Distributed Stochastic Neighbor Embedding (t-SNE) of single-cell RNA-sequencing data of spleen-derived Treg cell (CD4⁺TCRβ⁺CD25⁺) subpopulations: Klrg1⁻Nfil3(GFP)⁻ in black, Klrg1⁻Nfil3(GFP)⁺ in blue, and Klrg1⁺Nfil3(GFP)⁺ in red (n=5). (B) Uniform manifold approximation and projection (UMAP) of single-cell RNA-sequencing data of spleen-derived Treg cell (CD4⁺TCRβ⁺CD25⁺) subpopulations as in (A). Contour gates were drawn to include 70% of the parent population (Gate 1-3). Contribution of cell types to Gate1-3 shown in the pie charts to the right (n=5). (C) Violin plots illustrating the expression of Foxp3, Klrg1, Id2, Id3, Tnfrsf4, Tnfrsf9, Icos, and Maf in Klrg1-*Nfil3*(GFP)⁻ Treg, Klrg1⁻*Nfil3*(GFP)⁺ Treg, and Klrg1⁺*Nfil3*(GFP)⁺ Treg as in (F-G). Violin plots were scaled by width resulting in the same maximum width for all violins. Right, gene expression data of Klrg1-Nfil3(GFP)- Treg cells (black), Klrg1-Nfil3(GFP)+ Treg cells (blue) and Klrg1+Nfil3(GFP)+ Treg cells (red) for Foxp3, Klrg1, Id2, Id3, Tnfrsf4, Tnfrsf9, Icos, and Maf. Statistics based on Deseg2 (n=4). (D) Monocle plots derived from scRNA-seq data of spleen Klrg1-Nfil3(GFP)⁻ Treg, spleen Klrg1⁻Nfil3(GFP)⁺ Treg, spleen Klrg1⁺Nfil3(GFP)⁺ Treg as well as spleen, inguinal LN (ing LN), bone marrow, blood, VAT, skin, lung, and liver memory Treg (CD4⁺TCRβ⁺CD44⁺CD25⁺Foxp3(GFP)⁺). Color code indicates expression of Gata3. X-axis and y-axis indicate monocle dimension 1 and 3. Each dot represents a hexagonal bin, and each dot is colored by the mean expression value of Gata3 of the cells that are within the hexagonal bin. (E) Pseudotime plot as in (A) with monocle dimension 1 and dimension 2. (F) Data derived from scTCR-seq of spleen Klrg1 Nfil3(GFP)⁻ Treg, spleen Klrg1⁻Nfil3(GFP)⁺ Treg, spleen Klrg1⁺Nfil3(GFP)⁺ Treg as well as colon, skin, and VATderived KIrg1+Nfil3(GFP)+ tisTregST2 from two individual mice. Graphical representation of similarity coefficients between these samples based on Jaccard Index for both experiments. Color indicates similarity with low (blue) to high (red) (n=2). (G) Data derived from scTCR-seq of VAT, colon and skin-derived Klrg1⁺Nfil3(GFP)⁺ tisTregST2 as well as spleen Klrg1⁺Nfil3(GFP)⁺ Treg, spleen Klrg1-Nfil3(GFP)⁺ Treg, and spleen Klrg1-Nfil3(GFP)⁻ Treg from an individual mouse. Set size enumerates total number of successfully identified TCR α + β chains (left). To the right, individual clones and shared clones between all groups are displayed. On top, the total number of shared clones is displayed and numbered (n=2). Data representative of two or more independent experiments or cell sorts.



Klrg1

Figure S4. Adoptive transfer and development, related to Figure 4. (A) Gating strategy used to identify transferred and host Treg cells in DT-treated host animals. G1: Lymphocytes; G2: CD8-CD19⁻MHCII⁻Dead⁻TCRbeta⁺CD4⁺ T cells; G3: TCRbeta⁺CD4⁺CD25⁺ Treg cells; G4: CD45.1⁺CD45.2⁻ host Treg cells; G5: CD45.1⁻CD45.2⁺ transferred Treg cells. Two plots to the right illustrate expression of Klrg1 in CD45.1⁺CD45.2⁻ host Treg cells and CD45.1⁻CD45.2⁺ transferred Treg cells. (B) Identification of transferred Treg cells in tissues of recipient animals 10 days after transfer of CD45.2⁺ Klrg1⁻Nfil3(GFP)⁺ Treg cells. Contour plots illustrate expression of Klrg1 in transferred Treg (top) or host Treg (bottom). (C) Identification of transferred Treg cells in lung tissue of recipient animals 10 days after transfer of spleen-derived CD45.2⁺Klrg1⁻Nfil3(GFP)⁺ Treg cells. Contour plots illustrate expression of Klrg1 in transferred and host Treg. (D) Identification of transferred Treg cells in spleen, mesenteric LN (Mes) and lung tissue of recipient animals 10 days after transfer of lung-derived CD45.2⁺Klrg1⁻Nfil3(GFP)⁺ Treg cells. Contour plots illustrate expression of Klrg1 in transferred and host Treg. (E) Representative examples of spleen Klrg1⁻ Nfil3(GFP)⁺ and Klrg1⁺Nfil3(GFP)⁺ Treg cells 5d, 8d, 10d, 15d, 20d, 25d, and 70+d after birth. Pregate on CD8⁻CD19⁻MHCII⁻Dead⁻TCRbeta⁺CD4⁺CD25⁺ Treg cells. (F) Representative examples of skin, colon and liver Klrg1⁺Nfil3(GFP)⁺ Treg cells 5d, 10d, 12d, 15d, 20d, 25d, and 70+d after birth. Pre-gate on CD8⁻CD19⁻MHCII⁻Dead⁻TCRbeta⁺CD4⁺CD25⁺ Treg cells. For skin, lung and colon contour plots, flow cytometry data of several replicates were concatenated to increase visibility. Data representative of two or more independent experiments, n=3-19 per group for each experiment.



Figure S5. ATAC-seq of tissue T cells, related to Figure 5. (A) ATAC-seq data for parts of the Rora, Gpr55, Rad50, Bcl2, II1rl1, Klrg1, Mreg and Cish gene loci of spleen-derived Klrg1⁻ Nfil3(GFP)⁻ Treg cells (black), spleen Klrg1⁻Nfil3(GFP)⁺ Treg cells (light blue), spleen Klrg1+Nfil3(GFP)+ Treg cells (red) as well as lung, VAT. and skin-derived CD25⁺Foxp3(GFP)⁺CD44⁺Klrg1⁺ST2⁺ tisTregST2 (light brown, orange, dark blue). Y-axis ATAC signal intensity, x-axis gene structure, with exons indicated as heightened bars and introns as line, arrows indicate gene direction. All datasets group-normalized to maximum peak height indicated in brackets. Overall display length indicated on top in kilobases (kb) (n=4). (B) Left, heatmap illustrating number of open peaks from VAT tisTregST2 cells found in precursor/number of open peaks in VAT tissue for three groups spleen-derived Klrg1⁻Nfil3(GFP)⁻ Treg cells (black), spleen Klrg1⁻Nfil3(GFP)⁺ Treg cells (light blue), spleen Klrg1⁺Nfil3(GFP)⁺ Treg cells (red). X-axis sample type, v-axis gene name. To the right, ATAC-seg peaks of exclusively expressed genes from VAT tisTregST2 for three groups spleen-derived Klrg1⁻Nfil3(GFP)⁻ Treg cells (black), spleen Klrg1⁻ Nfil3(GFP)⁺ Treg cells (light blue) and spleen Klrg1⁺Nfil3(GFP)⁺ Treg cells (red). X-axis sample type, y-axis number of open peaks from tissue found in precursor/number of open peaks in tissue (n=4). (C) Heatmap illustrating number of open peaks from lung tisTregST2 found in precursor/number of open peaks in lung tissue for three groups as in (B) (n=4), (**D**) Heatmap illustrating number of open peaks from skin tisTregST2 found in precursor/number of open peaks in skin tissue for three groups as in (B) (n=4). (E) Correlation between common ATAC signature (Lung, skin and VAT Treg) and RNA expression (lung, skin and VAT Treg); 10kb distance to gene (n=4). Data representative of two or more independent experiments or cell sorts.



Figure S6. Batf and tissue Treg development, related to Figure 7. (A) Gating strategy used to identify Klrg1⁻Pd1⁻, Klrg1⁻Pd1⁺ or Klrg1⁺Pd1⁺ Treg cells in Batf^{-/-} vs Batf^{+/+} mice. G1: CD4⁺ T cells; G2: lymphocytes; G3: CD8⁻CD19⁻MHCII⁻Dead⁻TCRbeta⁺ T cells; G4: CD4⁺CD25⁺ Treg cells. From G4, Klrg1⁻Pd1⁻, Klrg1⁻Pd1⁺ or Klrg1⁺Pd1⁺ Treg cells can be identified. Expression of Batf in all three populations of Batf^{-/-} vs Batf^{+/+} mice is shown as histogram to the right. (B) Total numbers of KIrg1⁻ Pd1⁺ or Klrg1⁺Pd1⁺ Treg cells in spleens of Batf^{-/-} vs Batf^{+/+} mice (n=3-7, unpaired t-test). Right, total number of Klrg1⁺Pd1⁺ Treg cells per g VAT, per g lung or per whole colon (unpaired t-test, n=4-6). (C) Total numbers of Klrg1⁻Pd1⁺ or Klrg1⁺Pd1⁺ Treg cells in spleens of mixed bone marrow chimeras with 50% Batf^{+/+} and 50% Batf^{-/-} bone marrow six weeks after bone marrow transfer (unpaired t-test, n=5). (D) Gating strategy to identify transferred CD45.1 CD45.2⁺ Treg cells (mixed 50% CD90.1+CD90.2-Batf+/+ and 50% CD90.1-CD90.2+Batf-/-) in DT-treated CD45.1+CD45.2-Foxp3^{DTR} host animals two weeks after transfer. (E) Quality control of cells before transfer into congenic DT-treated recipient. (F) Presence of CD90.1⁺CD90.2⁻Batf^{+/+} vs CD90.1⁻CD90.2⁺Batf^{-/-} transferred Treg cells Klrg1⁻Pd1⁺ or Klrg1⁺Pd1⁺ Treg cells of spleens isolated from DT-treated host animals two weeks after transfer. Statistical verification across replicates to the right, gating (G6 and G7) derived from (E) (unpaired t test, n=4-6). (G) Presence of CD90.1⁺CD90.2⁻Batf^{+/+} vs CD90.1⁻CD90.2⁺Batf^{-/-} transferred Treg cells in lung or skin of DT-treated host animals two weeks after transfer with statistical verification across replicates (unpaired t test, n=4-6). (I) Gating strategy used to analyse Treg cells expanded with anti-CD3/28 microbeads and cytokines in-vitro. G1: lymphocytes; G2: CD4⁺TCRbeta⁺ T cells; G3: CD8⁻CD19⁻MHCII⁻Dead⁻CD4⁺ T cells. From G3, *Nfil3*(GFP)⁺ST2⁻ or *Nfil3*(GFP)⁺ST2⁺ Treg cells can be identified. Histogram to the right depicts Nfil3(GFP) expression in both groups. Below, expression of Nfil3(GFP) vs ST2 in expanded Treg cells treated with IL-2, IL-2/IL-4, IL-2/IL-33, or IL-2/IL-4/IL-33. To the right, expanded Treg cells with fixation and permeabilization to detect intracellular proteins. MFI of Gata3 or Batf was extracted (unpaired t test, n=4). Data representative of two or more independent experiments or cell sorts.



Figure S7. Batf and tissue Treg development, related to Figure 7. (A) Spleen Treg (CD4⁺CD25⁺Foxp3(GFP)⁺) or Tconv (CD4⁺CD25⁺Foxp3(GFP)⁺) cells were expanded with anti-CD3/28 microbeads and cytokines in-vitro for six days, followed by RNA isolation and cDNA synthesis. Nfil3 gene expression was measured by RT-PCR and normalized to a house keeping gene (Hprt). Induction was calculated based on baseline Nfil3 gene expression in untreated expanded Treg or Tconv cells and used to generate heatmap (colour code normalized to Treg and Tconv values) (n=1). (B) Cytokine titration of expanded Treg and Tconv as in (A), followed by measurement of Areg and II10 RNA and protein (n=1). (C) Time course experiment with expanded Treg cells and a fixed dose of 100 ng/mL IL-4 and 100 ng/mL IL-33 and four replicates (n=4). Cytokines and media were exchanged on day 7. On day 1, 3, 5, 7, 9, and 11, RNA was extracted and gene expression of II1Ir1, KIrg1, Areg and IL10 was measured by RT-PCR. Expression was normalized to control wells treated with IL-2 only (n=4). (D) Cytokine titration of expanded Treg cells as in (A-B), this time with IL-4 vs IFN-y, IL-4 vs IL5, IL-4 vs IL-9, and IL-4 vs IL-13. Gene expression of Nfil3 is shown (n=1). (E) ATAC-seq data for the Cd69, Mki67, Stat5a, Stat5b, and Tgfb1 gene and associated promoter region with LN-derived CD25⁺Foxp3(GFP)⁺CD44⁺ memory Treg (grey) as well as well as Batf^{-/-} or control Treg cells treated with either IL-2 or IL-2/IL-4/IL-33 for six days in-vitro (grev. blue, light brown). Y-axis ATAC signal intensity, x-axis gene structure, with exons indicated as heightened bars and introns as line, arrows indicate gene direction. All datasets groupnormalized to maximum peak height indicated in brackets. Overall display length indicated on top in kilobases (kb). Yellow box indicates area of interest (n=4). (F) ATAC-seg data for the Maf gene and associated promoter region as in (E), with top 4 lanes public dataset-derived Batf ChIP-seq data for CD4 or CD8 T cells including antibody control data (dark grey). Below, Batf^{-/-} or control Treg cells treated with either IL-2 or IL-2/IL-4/IL-33 for six days in-vitro (grey, blue, light brown), spleen-derived Klrg1-Nfil3(GFP)⁻ Treg cells (black), spleen Klrg1-Nfil3(GFP)⁺ Treg cells (light blue), spleen Klrg1+Nfil3(GFP)+ Treg cells (red) and VAT-derived tisTregST2 (orange) (n=4). Data representative of independent experiments or cell sorts.