Supplementary Materials

to the manuscript "Class A GPCRs increase their sensitivity and selectivity by harnessing the energy of membrane potential" by D. N. Shalaeva, D. A. Cherepanov, M. Y. Galperin, G. Vriend, and A. Y. Mulkidjanian

Table S1. Experimental data on voltage-sensitive activation of GPCRs.

Receptor	Agonist	Agonist characterization	Voltage effect	Method of activity	Reference
			on activation	measurement	
M2 muscarinic receptor	acetylcholine	full agonist (endogenous)	enhanced	GIRK currents	[1, 2]
M2 muscarinic receptor	oxotremorine	full/strong agonist	enhanced		
M1 muscarinic receptor	acetylcholine	full agonist (endogenous)	decreased		
M2 muscarinic receptor	acetylcholine	full agonist (endogenous)	enhanced	GIRK currents	[3]
M2 muscarinic receptor	acetylcholine	full agonist (endogenous)	enhanced	ACh-activated K ⁺ current	[2, 4]
M2 muscarinic receptor	pilocarpine	partial agonist	decreased		
M2 muscarinic receptor	acetylcholine	full agonist (endogenous)	enhanced	ACh-activated K ⁺ current	[2, 5] [6]
M2 muscarinic receptor	pilocarpine	partial agonist	decreased		
M2 muscarinic receptor	bethanechol	Agonist (low affinity)	no effect		
M1 muscarinic receptor	carbachol	full/strong agonist	decreased	FRET-based assays	[2, 7]
M3 muscarinic receptor	carbachol	full/strong agonist	enhanced		
mGluR3 glutamate receptor	glutamate	full agonist (endogenous)	enhanced	K ⁺ and Cl ⁻ currents	[8]
mGluR1a glutamate	glutamate	full agonist (endogenous)	decreased	1	
receptor					
α _{2A} -AR adrenergic receptor	noradrenaline	full agonist (endogenous)	enhanced	FRET-based assays	[9]

β ₁ -AR adrenergic receptor	isoprenaline	full agonist	enhanced	FRET-based assays	[10, 11]
β ₁ -AR adrenergic receptor	adrenaline	full agonist (endogenous)	enhanced		
dopamine D2L receptor	dopamine	full agonist (endogenous)	enhanced	GIRK currents	[12, 13]
dopamine D2L receptor	quinpirole	full agonist	enhanced		
dopamine D2S receptor	dopamine	full agonist (endogenous)		GIRK currents	[14]
dopamine D2S receptor	dopamine	full agonist (endogenous)	enhanced	GIRK currents	[12, 15]
dopamine D2S receptor	<i>p</i> -tyramine	partial agonist	decreased		
dopamine D2S receptor	<i>m</i> -tyramine	partial agonist	decreased		
dopamine D2S receptor	phenylamine	partial agonist	decreased		
dopamine D2S receptor	S-5-OH-DPAT	full agonist	no effect		
dopamine D2S receptor	R-5-OH-DPAT	full/strong agonist[16]	no effect		
dopamine D2S receptor	R-7-OH-DPAT	full/strong agonist	no effect		
dopamine D2S receptor	RIS-OH-DPAT	full/strong agonist[12]	no effect]	

The effectiveness of a signaling molecule in the receptor activation does not necessarily reflect the affinity of that molecule to the receptor. Some partial agonists have a high binding affinity but induce lower receptor activity even when added in saturating amounts [2, 15]. Such molecules are believed to stabilize the receptor in an intermediate state and/or stimulate the transition to the active state with lower probability than full agonists.

1 Table S2. Conservation of functionally important residues in Class A GPCRs.

Residue in M ₂ muscarinic acetylcholine receptor (PDB: 3UON)	Generi c number	Most common residue		Second most common residue		Residue type, %		
	c number	AA	%	AA	%			
	Na ⁺ coordination							
Asn41	1.50	N	98	S	1	Polar, 100		
Asp69	2.50	D	92	N	3	Polar, 98		
Ser110	3.39	S	72	T	8	Polar, 83		
Trp400	6.48	W	68	F	16	Aromatic, 87		
Asn432	7.45	N	67	S	11	Polar, 93		
Ser433	7.46	S	64	С	13	Polar, 72		
	CWxP motif							
Thr399	6.47	С	71	S	10	Small, 86		
Trp400	6.48	W	68	F	16	Aromatic, 87		
Pro402	6.50	P	99	N/	N/A	Helix kink, 99		
	Hydrophobic shell around the Na ⁺ pocket							
Leu65	2.46	L	90	M	4	Hydrophobic, 99		
Val111	3.40	I	40	V	24	Hydrophobic, 88		
Leu114	3.43	L	73	I	10	Hydrophobic, 98		
Ile117	3.46	I	56	L	16	Hydrophobic, 99		
Ile392	6.40	V	37	I	28	Hydrophobic, 93		
Leu393	6.41	V	41	L	20	Hydrophobic, 91		
Phe396	6.44	F	75	V	4	Hydrophobic, 92		
	Second hydrophobic shell							
Val44	1.53	V	65	A	14	Hydrophobic, 92		
Ile62	2.43	L	36	I	35	Hydrophobic, 97		
Trp148	4.50	W	96	F	1	Hydrophobic, 99		
Pro198	5.50	P	79	V	5	Hydrophobic, 95		
Met202	5.54	I	33	M	30	Hydrophobic, 90		
Tyr206	5.58	Y	72	S	5	Hydrophobic, 86		
Ile389	6.37	L	38	V	21	Hydrophobic, 91		
	NPxxY motif							
Asn436	7.49	N	72	D	20	Polar, 98		
Pro437	7.50	P	94	A	2	Hydrophobic, 98		
Tyr440	7.53	Y	89	F	4	Aromatic,93		
	DRY motif (ionic lock)							
Asp120	3.49	D	64	Е	21	Polar, 97		
Arg121	3.50	R	95	Н	1	Polar, 98		
Tyr122	3.51	Y	66	F	10	Hydrophobic, 87		
Glu382	6.30	Е	26	K	15	Polar, 79		

4 the protein structure are shown in Fig. 1.

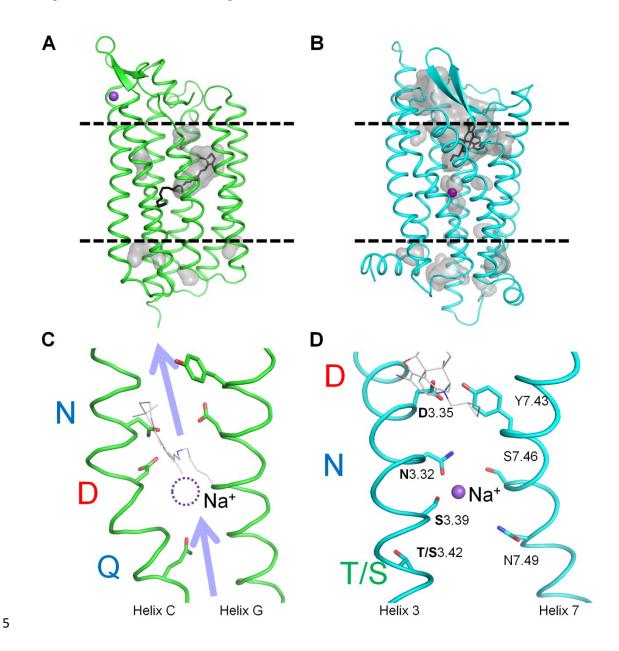


Figure S1. Structural and functional similarities between microbial rhodopsins (MRs) and GPCRs. A. Structure of sodium-translocating rhodopsin KR2 (PDB 6RF6 [18]); the protein is shown as a cartoon, Lys255 and retinal are shown as black sticks, cavities inside the protein are shown as gray volumes. B. Structure of the human δ-opioid receptor δ-OR (PDB 4N6H [19]); the protein is shown as a cartoon, antagonist (naltrindole) is shown as black sticks, cavities inside the protein are shown as gray volumes.. C. Signature residues involved in cation translocation by Na⁺-transporting rhodopsin KR2 (panel A). Signature residues of the NDQ motif are shown as sticks; Lys255 and retinal are shown in gray. Predicted Na⁺-binding site is indicated by a dotted circle. D. Corresponding residues in helices TM3 and TM7 in δ-OR (panel B), antagonist (naltrindole) shown in gray, residue numbering is according to Ballesteros-Weinstein [20].

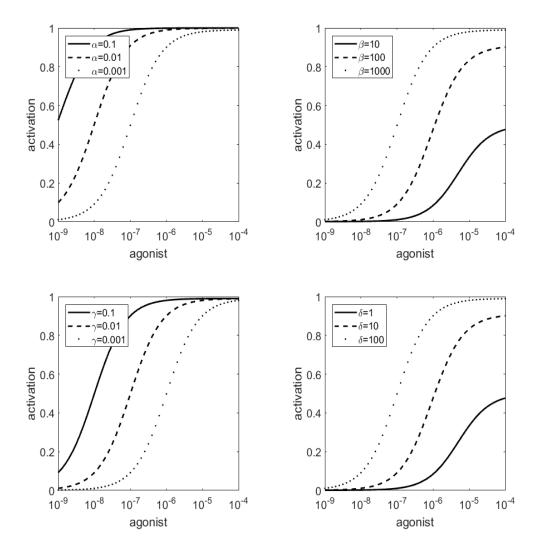


Figure S2. Effects of allosteric parameters α, β, γ and δ on the activation curves. Dependence of the GPCR activation on the coefficient α , intrinsic efficacy of sodium (A); coefficient β , intrinsic efficacy of the agonist (B); coefficient γ , binding cooperativity between sodium and the agonist (C), and coefficient δ , activation cooperativity between sodium and the agonist (D). In each panel, one of parameters was varied while the values of other parameters were taken from Table 2. The impact of the membrane voltage was not taken into account in the calculations.

A. Mode 1: carrier-on

B. Mode 2: carrier-off

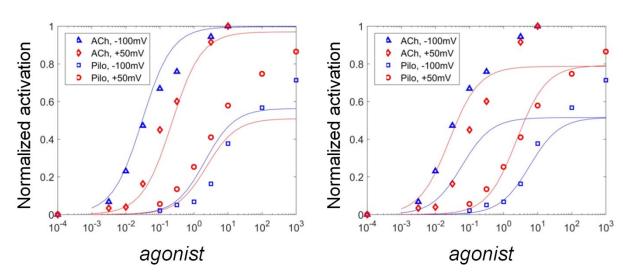


Figure S3. Fitting experimental data of voltage-sensitive muscarinic acetylcholine receptor M₂ activation. Concentration-response curves, as obtained for the full endogenous agonist acetylcholine (ACh) and partial synthetic agonist pilocarpine (Pilo), measured at different magnitudes of membrane potential were fitted for the carrier-on mode (A) and carrier-off mode (B). Experimental data from [4].

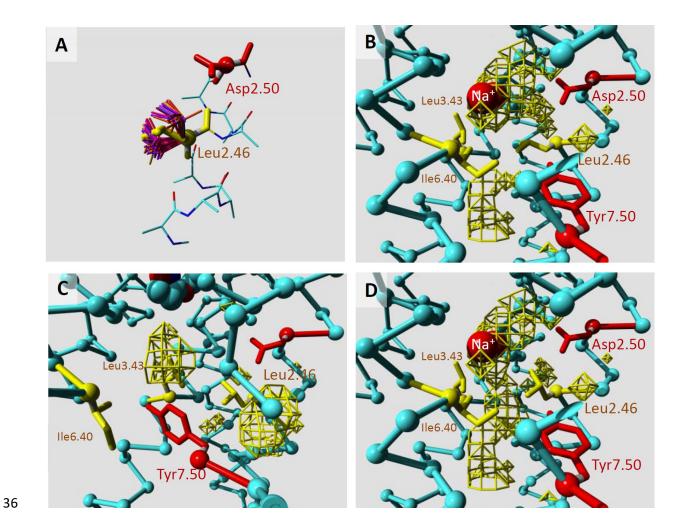


Figure S4. Rotamers of Leu2.46 in the inactive and inactive conformations of muscarinic acetylcholine receptor M2. A, Analysis of Leu2.46 rotamers, shown on the example of PDB 3UON. The section of Helix 2 is shown coloured by atom type. The Asp2.50 side chain is shown in red. Leu2.46 as in the 3UON structure is shown in yellow. The preferable rotamers for leucine at position 2.46 are shown in different shades of orange, blue, and red. **B-D**, The Na⁺ binding site in the structure of the inactive receptor, PDB 3UON (**B**); the active receptor (PDB 4MQT) (**C**), and the inactive receptor (PDB 3UON) with Leu2.46 moved into upper rotamer (**D**). C-alpha traces are shown in light-blue (some residues in the front that obscured the view have been removed from the plots). Side chains of Asp2.50 and Tyr7.50 are shown as red stick models. Cavities large enough to hold water molecules are represented by yellow mesh. The side chains of the three aliphatic residues Leu2.46, Leu3.43, and Ile6.40, are shown as yellow stick models.

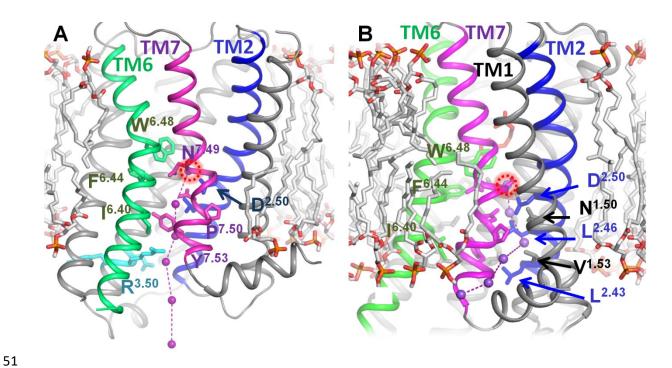
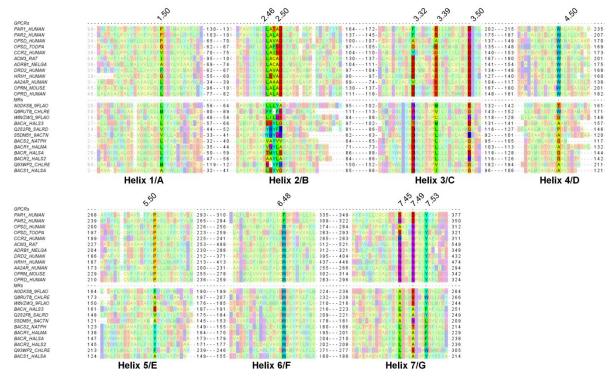


Figure S5. Suggested pathways of Na⁺ escape to the cytoplasm upon GPCR activation, based on the structure of the M₂ receptor (PDB: 4MQT). A. Putative exit pathway for the Na⁺ ion via the center of the heptahelical bundle. B. An alternative exit pathway via the pocket between helices 1, 2, and 7. Helices 2, 6, and 7 are colored blue, green and purple, respectively; conserved residues listed in Table S2 are shown as sticks. The ionic lock residues are shown in cyan. CHARMM-GUI software [21] was applied to construct the lipid molecules of the membrane surrounding the receptors (shown as grey sticks).



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Fig. S6. Multiple sequence alignment of transmembrane segments in GPCRs and MRs. Sequences used in the alignment (listed under their UniProt IDs): PAR1 HUMAN - Human proteinase-activated receptor 1; PAR2 HUMAN - Human proteinase-activated receptor 2; OPSD HUMAN - Human visual rhodopsin; OPSD TODPA - Squid visual rhodopsin; CCR2 HUMAN - Human C-C chemokine receptor type 2; ACM3 RAT - Rat muscarinic acetylcholine receptor M3; ADRB1 MELGA - Turkey β-1 adrenergic receptor; DRD2 HUMAN - Human D₂ dopamine receptor; HRH1 HUMAN - Human histamine H1 receptor; AA2AR HUMAN - Human adenosine receptor A2a; OPRM MOUSE - Mouse μtype opioid receptor; OPRD HUMAN - Human δ-type opioid receptor. MRs: NODKS8 9FLAO - Sodium pumping rhodopsin from Dokdonia eikasta; Q8RUT8 CHLRE -Channelrhodopsin 2 from Chlamydomonas reinhardtii; W8VZW3 9FLAO - Chloride pumping rhodopsin from Nonlabens marinus; BACH HALS3 - Halorhodopsin from Halobacterium salinarum; Q2S2F8 SALRD - Xanthorhodopsin from Salinibacter ruber; S5DM51 9ACTN - Bacteriorhodopsin from Candidatus Actinomarina minuta; BACS2 NATPH - Sensory rhodopsin-2 from Natronomonas pharaonis; BACR1 HALMA -Bacteriorhodopsin-I from Haloarcula marismortui; BACR HALSA - Bacteriorhodopsin from Halobacterium salinarum; BACR2 HALS2 - Archaerhodopsin-2 from Halobacterium sp.; Q93WP2 CHLRE - Archaeal-type opsin 1 from Chlamydomonas reinhardtii; BACS1 HALSA - Sensory rhodopsin-1 from Halobacterium salinarum. Alignment of GPCRs was taken from the GPCRdb web service [18], alignment of MRs was constructed with T-Coffee [19], two alignments were merged using the structure superposition of human δ-opioid receptor (PDB 4N6H) and channelrhodopsin 2 from Chlamydomonas reinhardtii (PDB 6EIG), as described in [20].

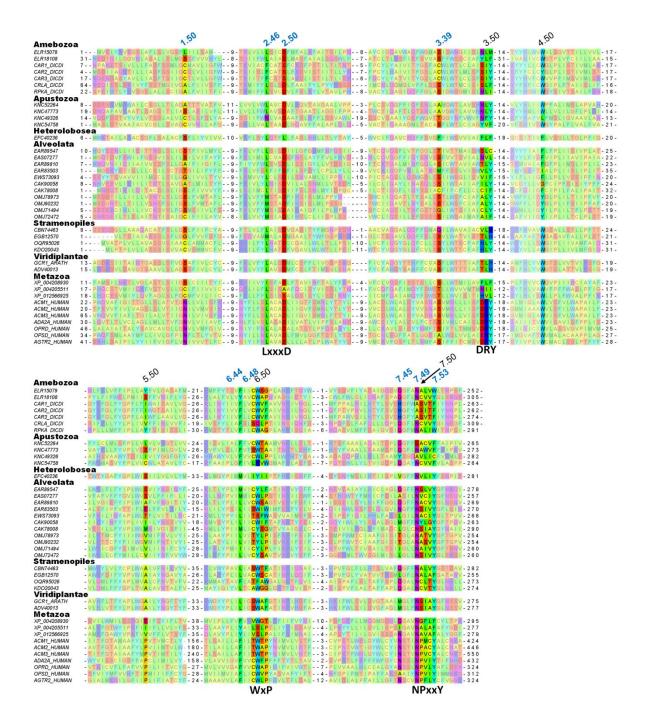


Figure S7. Multiple sequence alignment of class A GPCRs from primitive organisms (the full version of the Fig. 5 of the main text). Sequences are listed under their GenBank, UniProt or RefSeq accessions and are as follows: Amoebozoa: phosphatidylinositol 4-phosphate 5-kinase protein from *Acanthamoeba castellanii* (GenBank: ELR15079); cAMP receptor protein from *Acanthamoeba castellanii* (GenBank: ELR18108); cAMP receptor 1 from *Dictyostelium discoideum* (UniProt: CAR1_DICDI); cAMP receptor 2 from *Dictyostelium discoideum* (UniProt: CAR2_DICDI); cAMP receptor 3 from *Dictyostelium discoideum* (UniProt: CAR3_DICDI); cAMP receptor-like protein from *Dictyostelium discoideum* (UniProt: CRLA_DICDI), and G-protein-coupled receptor family protein from *Dictyostelium discoideum* (UniProt: RPKA_DICDI). Apustozoa: hypothetical protein AMSG_01092 from *Thecamonas trahens* (GenBank: KNC52264); hypothetical protein AMSG_04000 from *Thecamonas trahens* (GenBank: KNC47773); hypothetical protein AMSG_02398 from *Thecamonas trahens* (GenBank: KNC56428); PPK-1 protein from *Thecamonas trahens* (GenBank: KNC54758). Heterolobosea: predicted protein NAEGRDRAFT_72027 from *Naegleria gruberi* (GenBank: ENC40236). Alveolata: G protein coupled glucose receptor from *Tetrahymena thermophila* (GenBank: EAR89547); 7TM secretin family protein from *Tetrahymena thermophila* (GenBank: EAS07277); 7TM secretin family protein

from Tetrahymena thermophila (GenBank: EAR89810); G protein coupled glucose receptor from Tetrahymena thermophila (GenBank: EAR83503); cAMP receptor from Tetrahymena thermophila (GenBank: EWS73093); unnamed protein product from Paramecium tetraurelia (GenBank: CAK90058); hypothetical protein from Paramecium tetraurelia (Genbank: CAK78008); hypothetical protein SteCoe 21100 from Stentor coeruleus (GenBank: OMJ78973); hypothetical protein SteCoe 30276 from Stentor coeruleus (GenBank: OMJ71494); hypothetical protein SteCoe_7430 from Stentor coeruleus (GenBank: OMJ90232); hypothetical protein SteCoe_29065 from Stentor coeruleus (GenBank: OMJ72472). Stramenopiles: G-protein coupled receptor from Ectocarpus siliculosus (GenBank: CBN74463); hypothetical protein AURANDRAFT 4432 from Aureococcus anophagefferens, partial (GenBank: EGB12570); hypothetical protein ACHHYP 00504 from Achlya hypogyna (GanBank: OQR95026); hypothetical protein SPRG 14191 from Saprolegnia parasitica (GenBank: KDO20043). Viridiplantae: G-protein coupled receptor 1 from Arabidopsis thaliana (UniProt: GCR1 ARATH); G protein coupled receptor from Oryza sativa (GenBank: ADV40013). Metazoa: predicted probable G-protein coupled receptor 157 from Hydra vulgaris, partial (NCBI RefSeq: XP 004208930); predicted cAMP receptorlike protein A from Hydra vulgaris (NCBI RefSeq: XP 004205511); predicted G-protein coupled receptor 1like protein from Hydra vulgaris (NCBI RefSeq: XP 012566925); human muscarinic acetylcholine receptor M₁ (UniProt: ACM1 HUMAN); human muscarinic acetylcholine receptor M₃ (UniProt: ACM3 HUMAN); human α_{2A} adrenergic receptor (UniProt: ADA2A HUMAN); human δ-type opioid receptor (UniProt: OPRD HUMAN); human visual rhodopsin (UniProt: OPSD HUMAN).

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