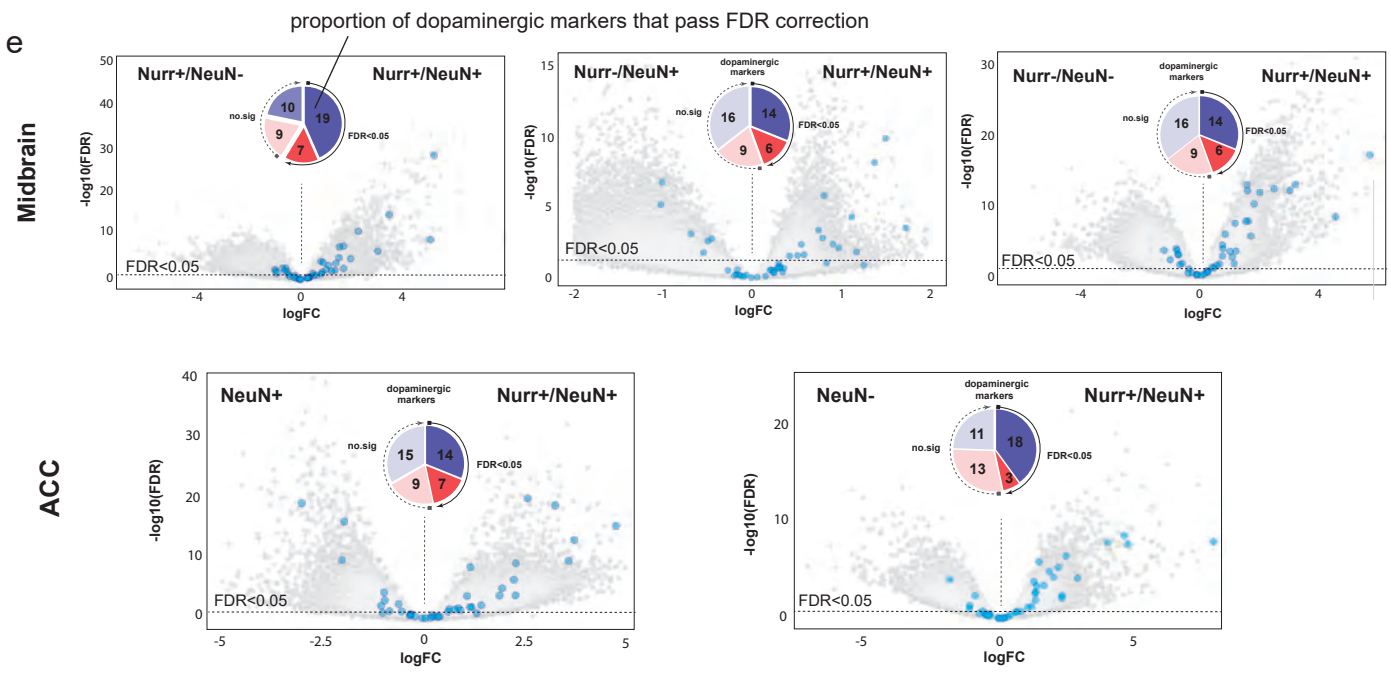
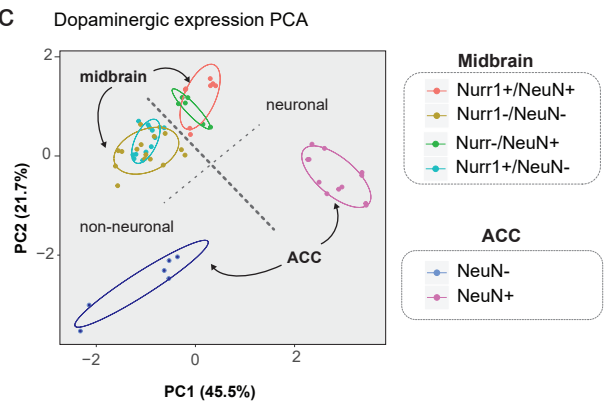
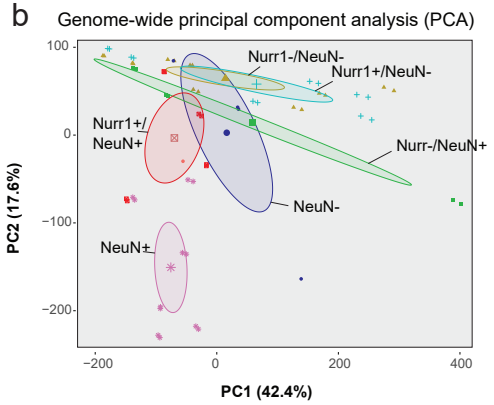
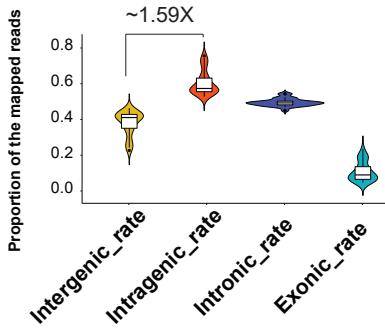
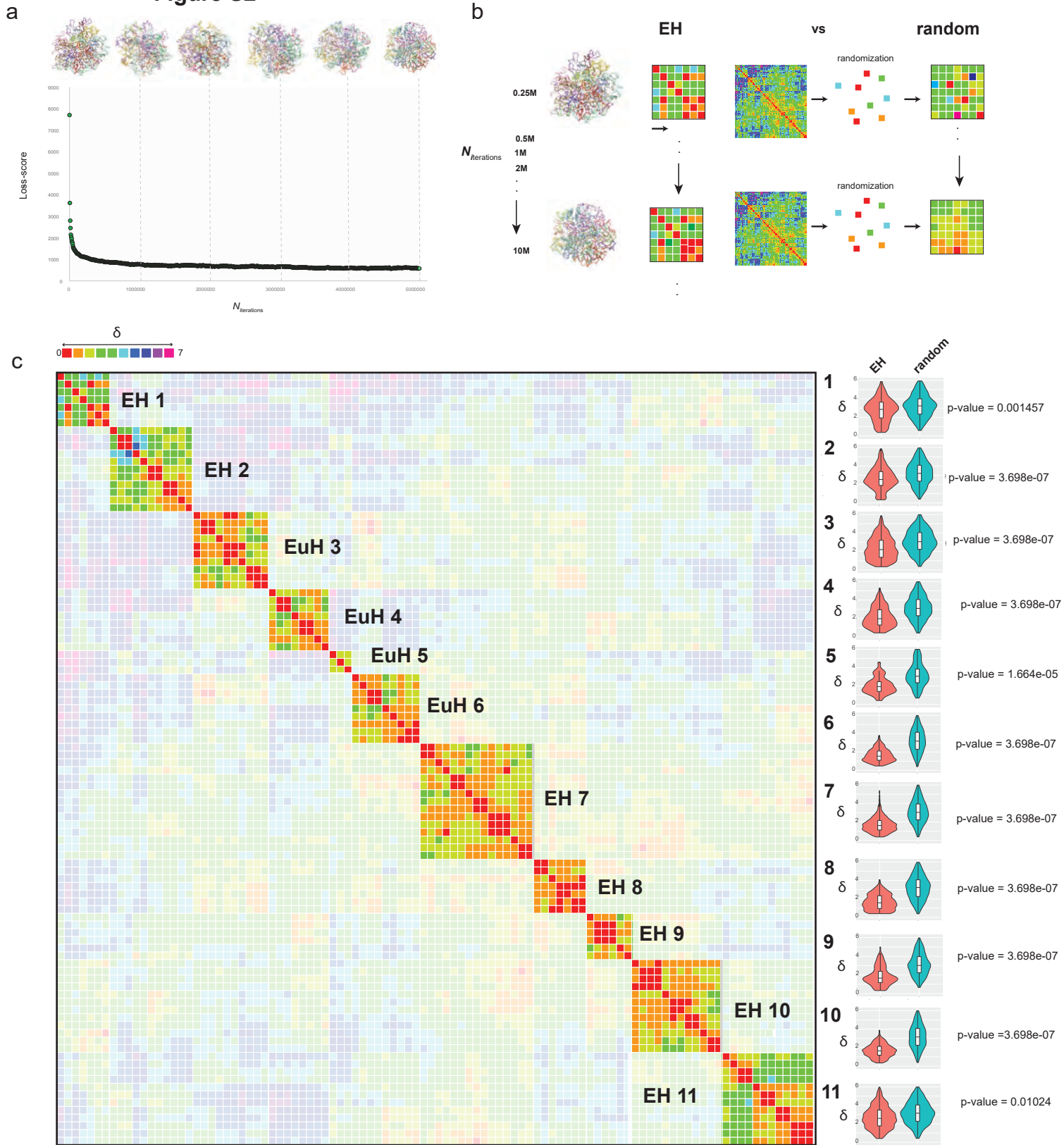


Figure S1



Supplementary Figure 1. Transcriptomic analysis of sorted nuclei from anterior cingulate cortex (ACC) and the ventral midbrain. **a)** Genomic features of mapped reads from nucRNAseq datasets (*Additional File 2: Table S2*) using RNA-SeQC³⁹. Violin plots show proportions of intergenic, intragenic, intronic and exonic fractions for the N=30 cell type specific nucRNAseq libraries (*Additional File: Table S2*). **b)** Genome-wide PCA analysis of nucRNAseq from six sorted nuclei populations (4 midbrain, 2 cortical nuclei populations) as indicated. **c)** PCA analyses of 45 transcripts selected from mouse midbrain single cell RNAseq resource⁴¹ for the six sorted nuclei populations as indicated. **d)** Differential nucRNA-seq analysis of five different nuclei populations from midbrain and cortex, each compared against midbrain dopaminergic neuron *Nurr1*⁺/*NeuN*⁺ using edgeR¹¹⁰ wrapper tool RUVseq⁴⁰. Pie charts denote the proportion of dopaminergic markers that pass the FDR cutoff and the direction of the fold change (blue denotes upregulated and red downregulated in *Nurr1*⁺/*NeuN*⁺ population).

Figure S2

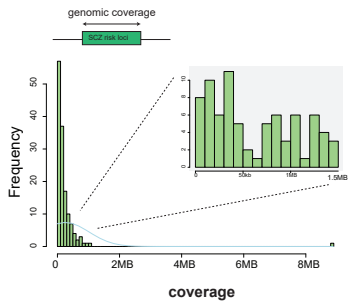


Supplementary Figure 2. *In silico* Nurr1⁺/NeuN⁺ chrom3D model stability across multiple iterations.

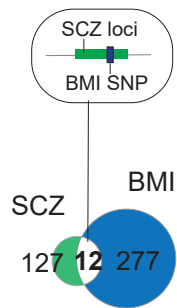
Benchmark analysis for the *in-silico* chrom3D model stability across multiple iterations **a)** genome-wide and **(b,c)** by Euclidean hotspot (b,c). **a)** Loss score plot measuring conformation sparsity of the *in-silico* model with successive increase in iterations (0.25M to 5M) across different runs of chrom3D. Note that the model starts to become stable after 1M of iterations. **b, c)** In order to assess the statistical confidence for each EH to be found with the exact same pairwise distance cluster between its domains, chrom3D was run 12 times with multiple iterations (from 250k, 500k, 1M, to 10M of iterations) and EH pair-wise distances were calculated along with random domains pair-wise distances in the model. As the distribution of the distances do not follow a Normal distribution (tested by a Shapiro and Andersen normality test), a non-paired Wilcox test was used to estimate the significance to find the same structure against randomness across all chrom3D iterations.

Figure S3

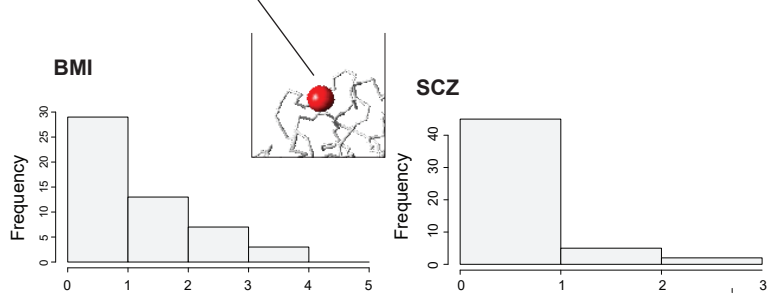
a SCZ risk loci genomic coverage span



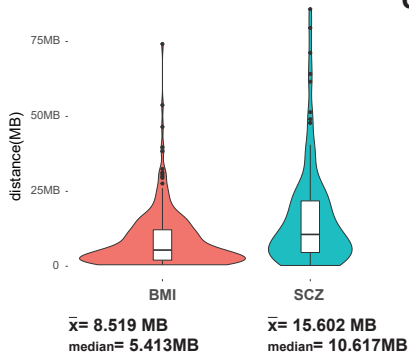
c Traits intersection



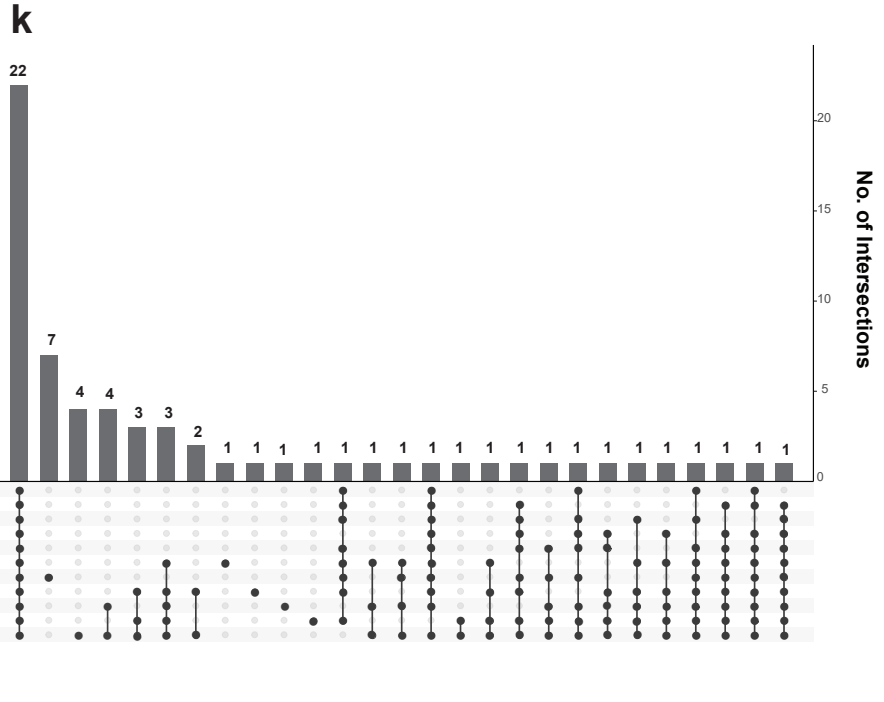
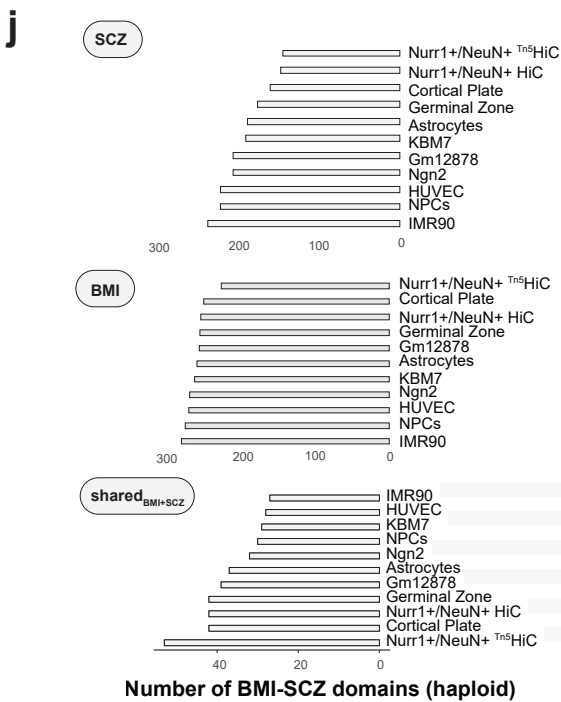
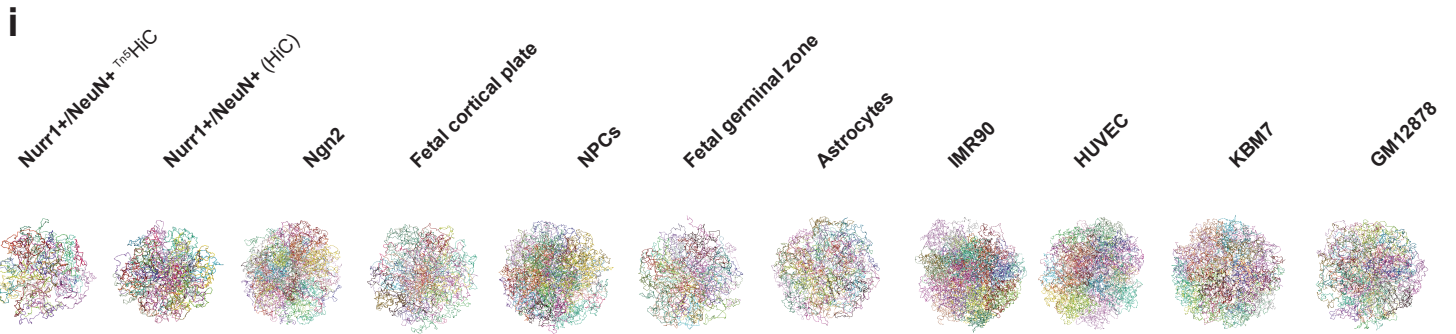
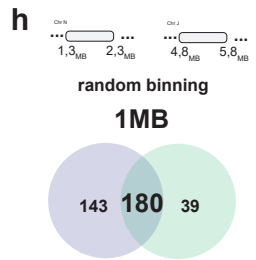
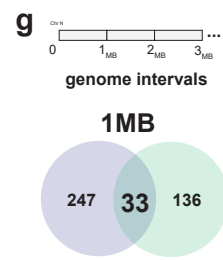
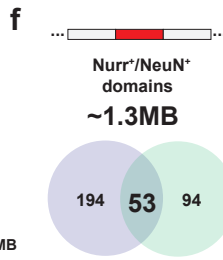
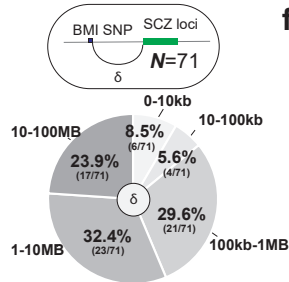
e SNPs/risk loci rate per BMI+SCZ domains



b Intergenic distance - SNPs/risk loci



d Distance among traits

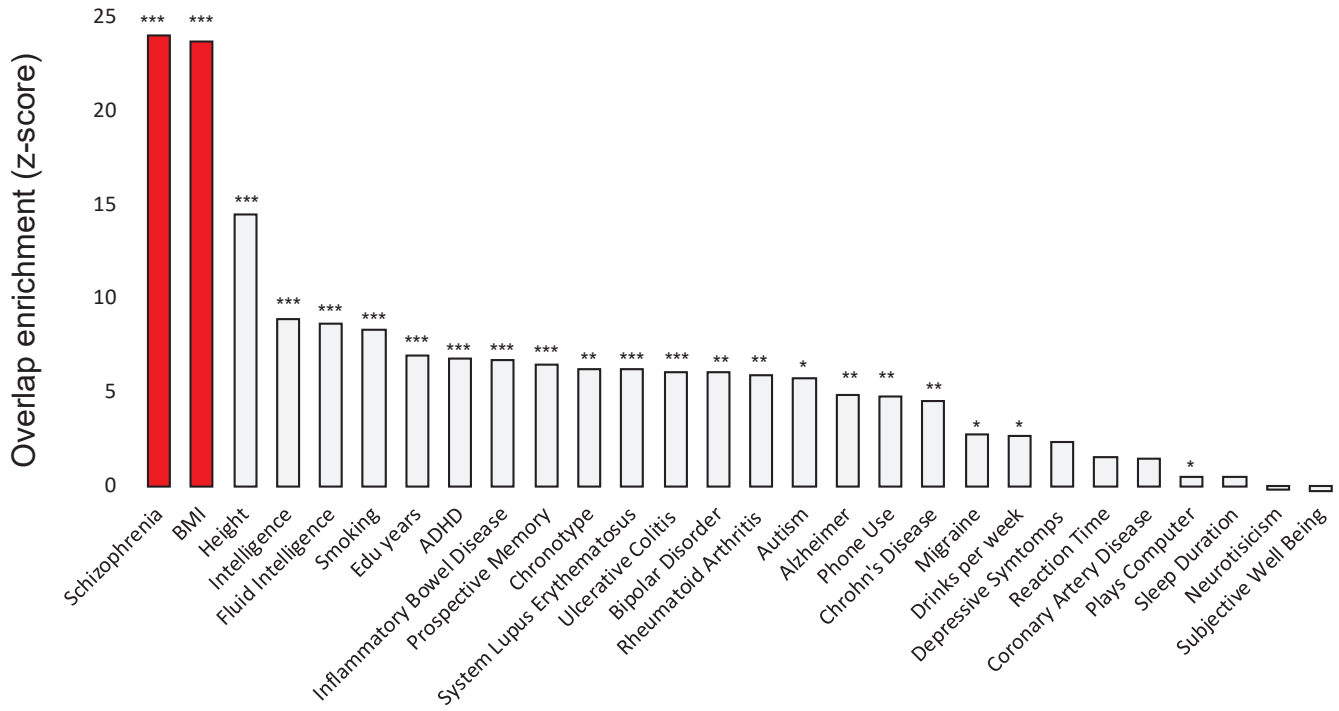


Supplementary Figure 3 Linear and 3D conformation characterization of BMI and SCZ.

a) Histogram of schizophrenia genomic coverage. SCZ loci average in length $\sim 256\text{kb} \pm 749$. **b)** Genomic distance between BMI SNPs (left) and schizophrenia risk loci (right) **c)** BMI SNPs and schizophrenia loci genomic Venn plot intersection showing that only 12 BMI SNPs fall into schizophrenia risk loci. **d)** 2D linear genome exploration of contiguous BMI and schizophrenia polymorphisms (N=71 SCZ risk loci located next to BMI risk SNP or vice versa). Pie chart showing the distance interval percentage among BMI SNPs and schizophrenia risk loci (interval hits versus total number showed in parenthesis). **e)** Histogram of BMI SNPs (left) and schizophrenia risk loci (right) falling into the same *Nurr1*⁺/*NeuN*⁺ 3D model domain. **f)** Intersection of both BMI and schizophrenia traits with *Nurr1*⁺/*NeuN*⁺ 3D model domains. **g)** genomic intervals observed at 1Mb and **h)** computed by random binning of 1Mb. **i)** *In silico* 3D cell models from Hi-C datasets generated from various cell types and tissues as indicated, at 50kb and 1MB resolution of intra-chromosomal and inter-chromosomal interactions respectively with 5M Chrom3D iterations (see Methods). **j)** Intersection of (top) schizophrenia risk) loci, (middle) BMI SNPs and (bottom) co-localization of both traits in 3D domains of all cell types present in the analysis. **k)** Number of BMI-SCZ domains shared across all 3D models.

Figure S4

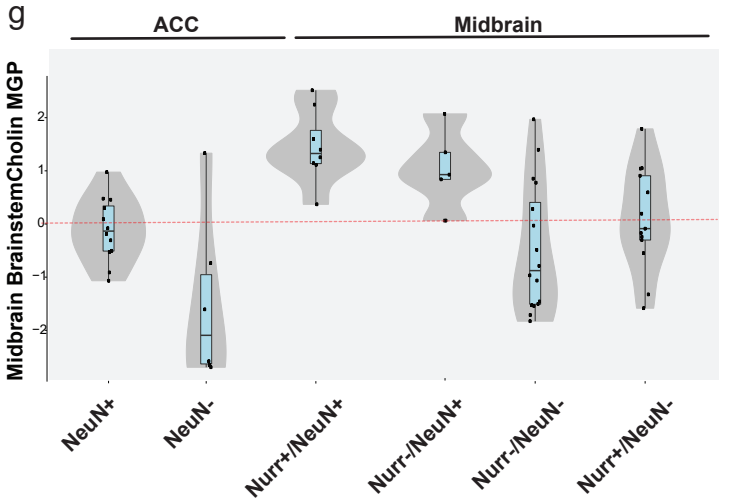
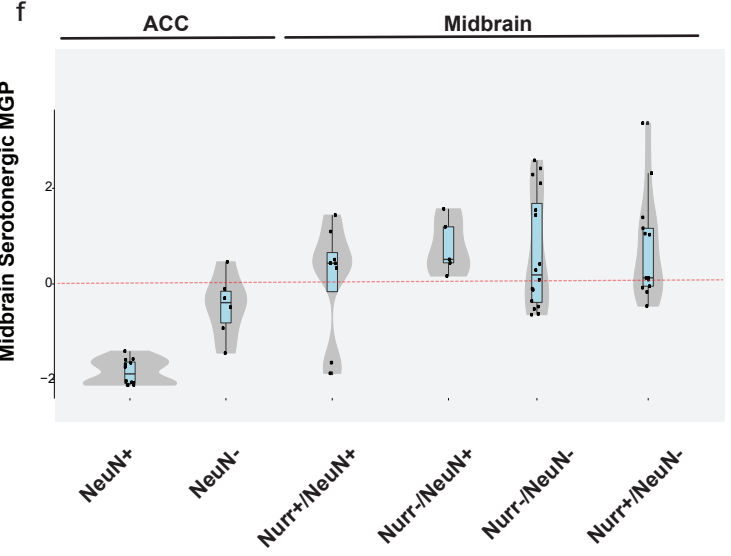
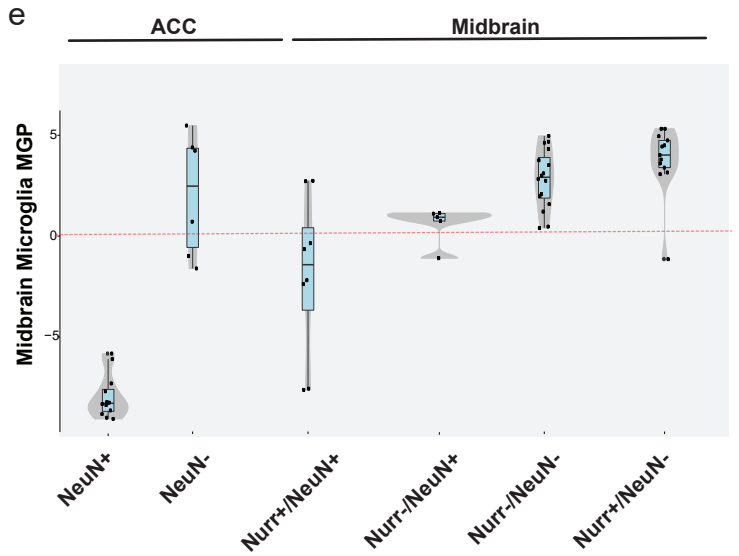
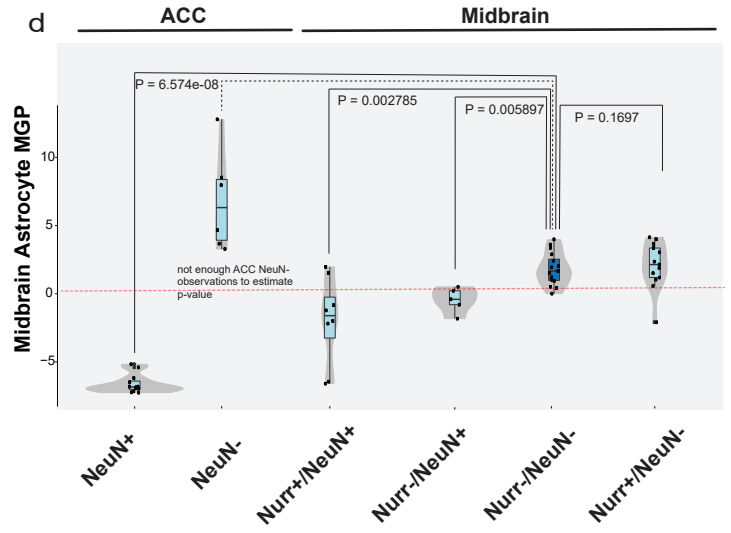
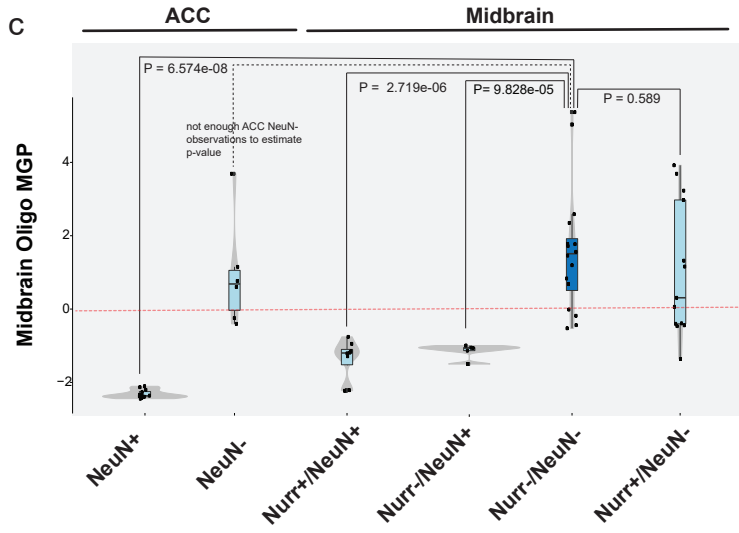
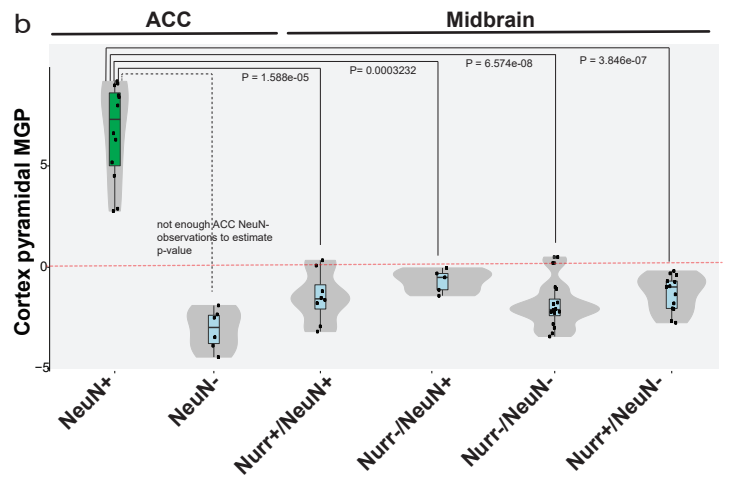
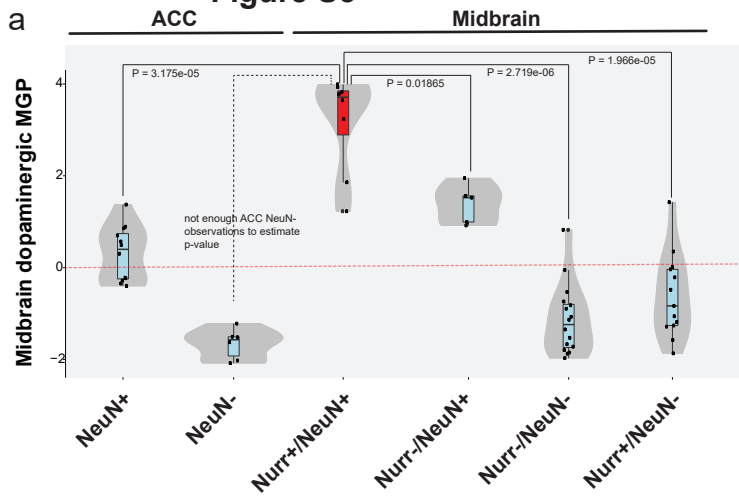
BMI-SCZ_{domains} vs other traits



Supplementary Figure 4: Permutation analysis of BMI-SCZ shared domains versus 28 GWAS

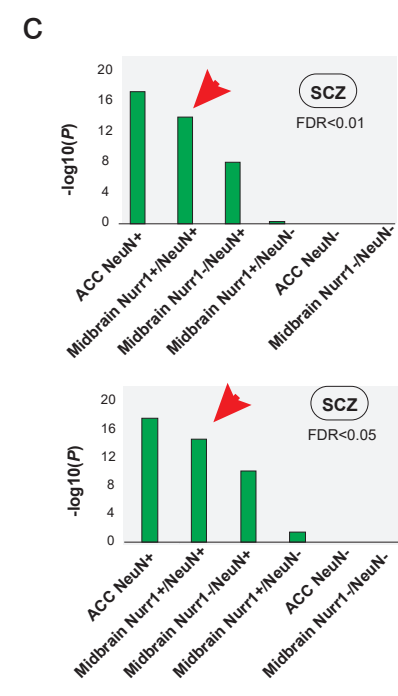
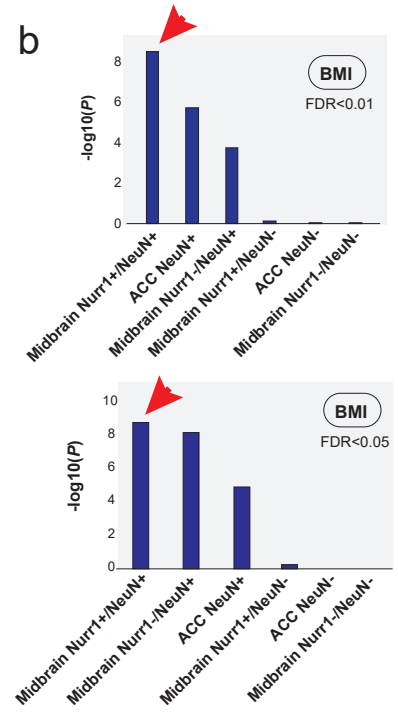
traits. Bar plot showing the strength of the association of BMI-SCZ shared domains over 28 traits (selection of significant SNPs at p value $< 5 \times 10^{-8}$). The overlap enrichment is measured by z-scores that were calculated permuting BMI-SCZ domains 10k times randomly over all domains called in Nurr1⁺/NeuN⁺ nuclei. Values represent the distance between the expected and the observed value, measured in standard deviations. Note that BMI and SCZ are the two top ranking GWAS; other traits were also significantly enriched (asterisks, *** p value < 0.001 , ** p value < 0.01 , * p value < 0.05) but the strength of the associations are considerable less when compared to BMI and SCZ each separately (red bars). The following GWAS summary statistics were used in this analyses (PMID numbers): Type 2 Diabetes (30297969); Drinks per week and Ever smoker(30643258); Prospective memory, Fluid intelligence score, Migraine, Phone use, Gaming defined as Plays computer (31427789); Reaction time (29844566); Height (25282103); Coronary artery disease (26343387); Crohn's disease, Ulcerative Colitis; Inflammatory Bowel disease (26192919); Rheumatoid Arthritis (24390342); Systemic Lupus Erythematosus (26502338); BMI (25673413); Chronotype and Sleep duration (27494321); Insomnia (28604731); Depressive Symptoms, Neuroticism, Subjective Well Being (27089181); Intelligence (28530673); Education Years (28197077); Attention Deficit/Hyperactivity Disorder=ADHD (30478444); Alzheimer's Disease (24162737); Autism (30804558); Depression (); Bipolar Disorder (31043756); SCZ (25056061).

Figure S5



Supplementary Figure 5. Cell-specific marker gene expression. Gene expression by cell type, using single-cell curated data from *markerGeneProfile* utility in *neuroexpresso*⁴². The four nuclei subpopulations from ventral midbrain (Nurr1⁺/NeuN⁺, Nurr1⁻/NeuN⁺, Nurr1⁺/NeuN⁻, Nurr1⁻/NeuN⁻) and two cortical (NeuN⁺ and NeuN⁻) subpopulations of nuclei of the present study were compared for marker gene expression representing seven cell types including brainstem dopaminergic and cholinergic and serotonergic neurons, microglia and oligodendrocytes and astrocytes, and cortical pyramidal neurons, as indicated on Y-axes. **a)** Violin plots show significantly increased expression of dopaminergic marker genes in Nurr1⁺/NeuN⁺ MDN nuclei populations compared to each of the remaining five non-dopaminergic nuclei populations (Wilcoxon p). **b)** cortical pyramidal neuron marker expression is highest in cortex ACC NeuN⁺. **c)** oligodendrocyte, **d)** astrocyte, **e)** microglia, **f)** serotonergic and **g)** cholinergic marker gene expression.

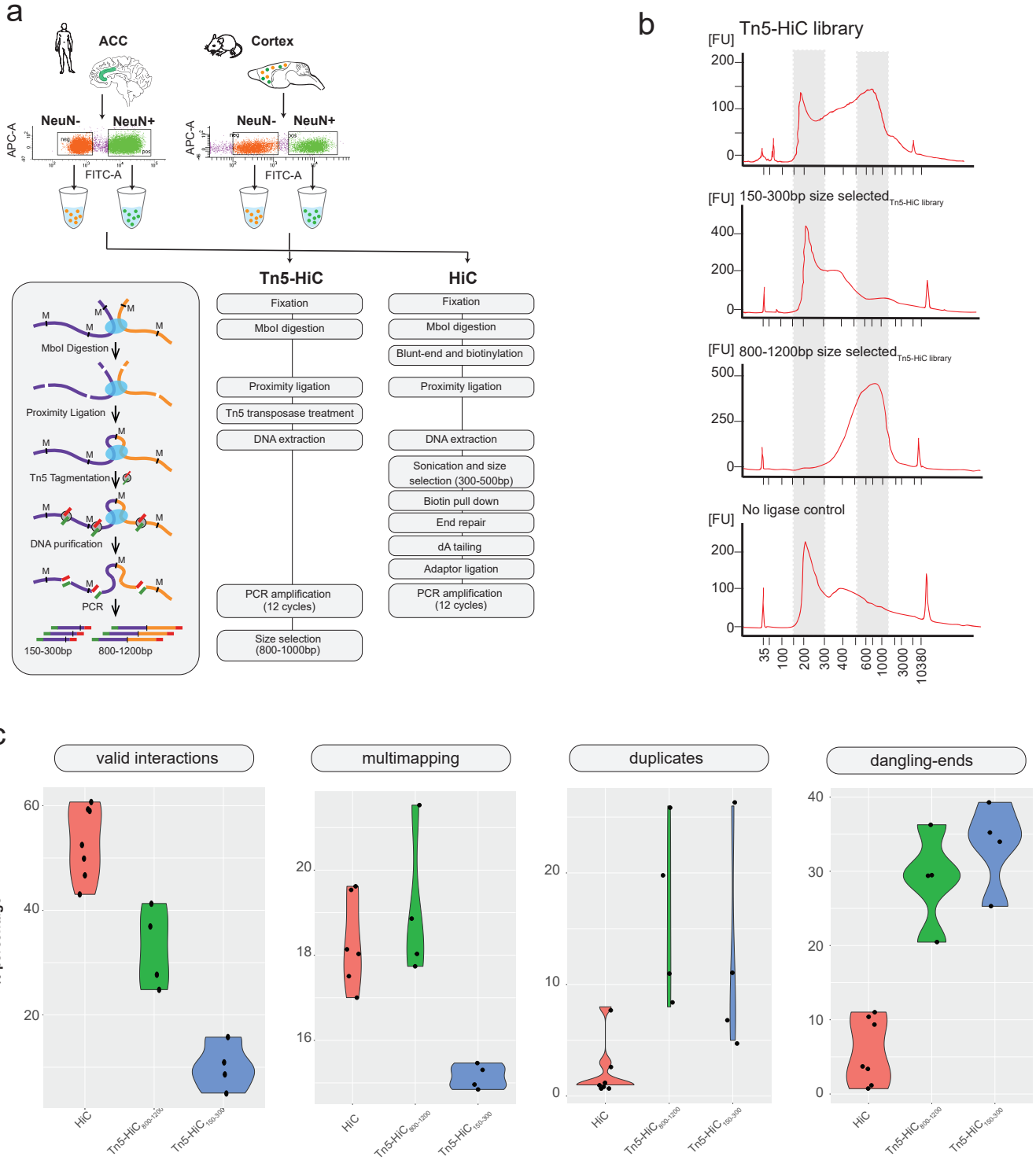
Figure S6



Supplementary Figure 6. Trait association of expressed genes in six neuronal and non-neuronal populations collected from midbrain and cerebral cortex.

MAGMA GWAS association of expressed genes in neuronal versus non-neuronal populations. **a)** At different FDR thresholds (0.01 and 0.05), genes enriched in each sorted population after differential expression analysis (using edgeR¹¹⁰ wrapper tool RUVseq⁴⁰) were piped into MAGMA association trait analysis. Color scale represents the log p-value and boxes showing “***”: represent p-values significant at 0.01 level after FDR correction of multiple testing across all traits and gene sets; or boxes with “*”: p-values significant at 0.05 level. **b)** BMI and **c)** SCZ associate on ranking for each cell population at selected genes at FDR<0.01 (upper bar plot) or FDR<0.05 (lower barplot). Notice *Nurr1*⁺/*NeuN*⁺ MDN (marked by red arrowhead) rank top among for BMI and second for SCZ, among the six populations of nuclei. The following GWAS summary statistics were used in this analyses (PMID numbers): Type 2 Diabetes (30297969); Drinks per week and Ever smoker(30643258); Prospective memory, Fluid intelligence score, Migraine, Phone use, Gaming defined as Plays computer (31427789); Reaction time (29844566); Height (25282103); Coronary artery disease (26343387); Crohn’s disease, Ulcerative Colitis; Inflammatory Bowel disease (26192919); Rheumatoid Arthritis (24390342); Systemic Lupus Erythematosus (26502338); BMI (25673413); Chronotype and Sleep duration (27494321); Insomnia (28604731); Depressive Symptoms, Neuroticism, Subjective Well Being (27089181); Intelligence (28530673); Education Years (28197077); Attention Deficit/Hyperactivity Disorder=ADHD (30478444); Alzheimer’s Disease (24162737); Autism (30804558); Depression (); Bipolar Disorder (31043756); SCZ (25056061).

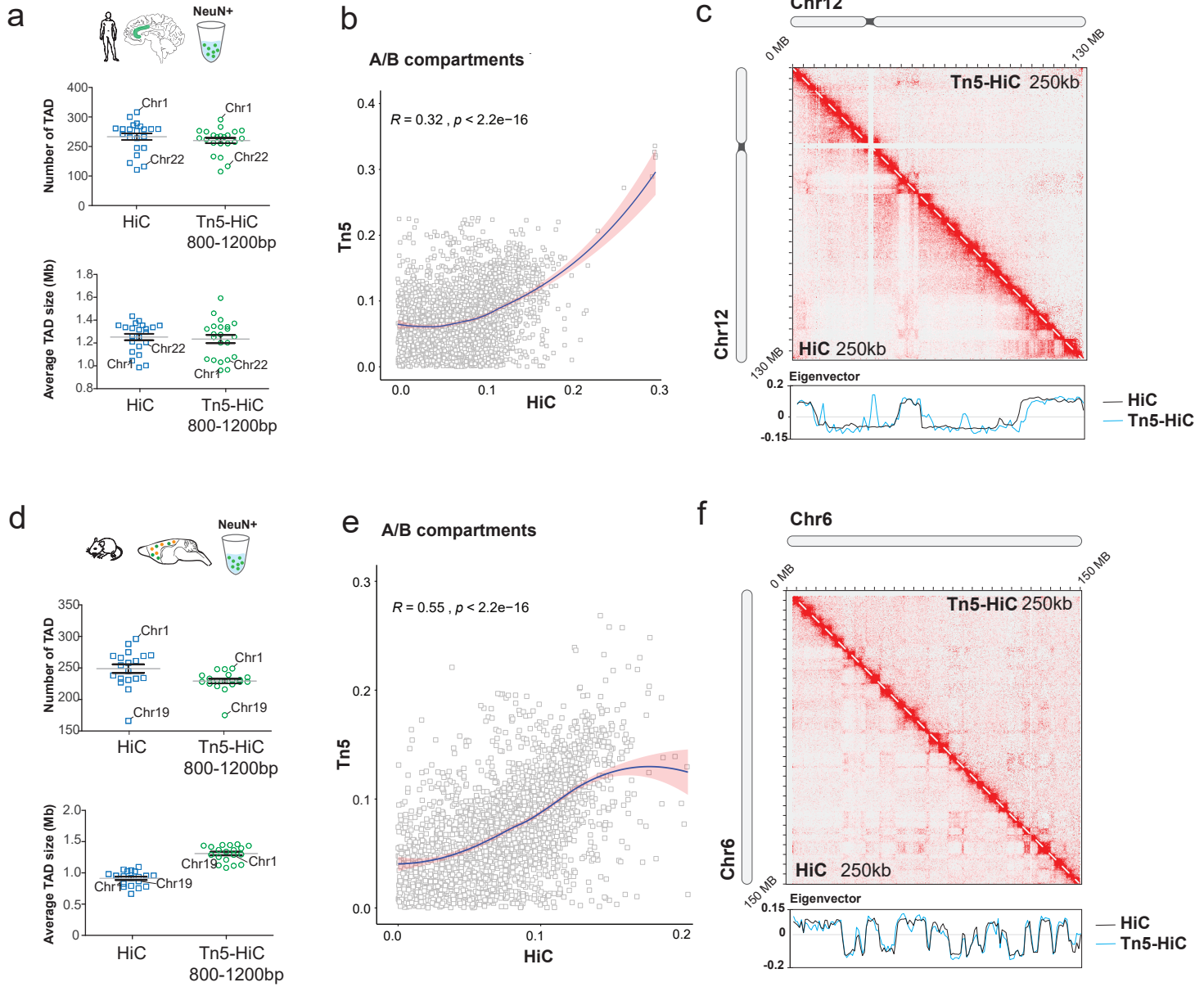
Figure S7



Supplementary Figure 7. $Tn5$ HiC versus HiC workflow and quality controls.

- a)** $Tn5$ HiC and HiC procedures for side-by-side comparison of both techniques. Two $Tn5$ HiC libraries sizes were produced for each cell-type and species with different fragment sizes: 150-300bp and 800-1200bp.
- b)** Bioanalyzer plots of the different fragment libraries ranges produced by $Tn5$ HiC protocol, from top to bottom plot: without size selection, 150-300bp, 800-1200bp and no ligase control. **c)** Valid interaction and three common artifacts (multi-mapping, dangling ends, duplicates) found in $Tn5$ HiC method compared to HiC (see also *Additional File 2: Table S1*).

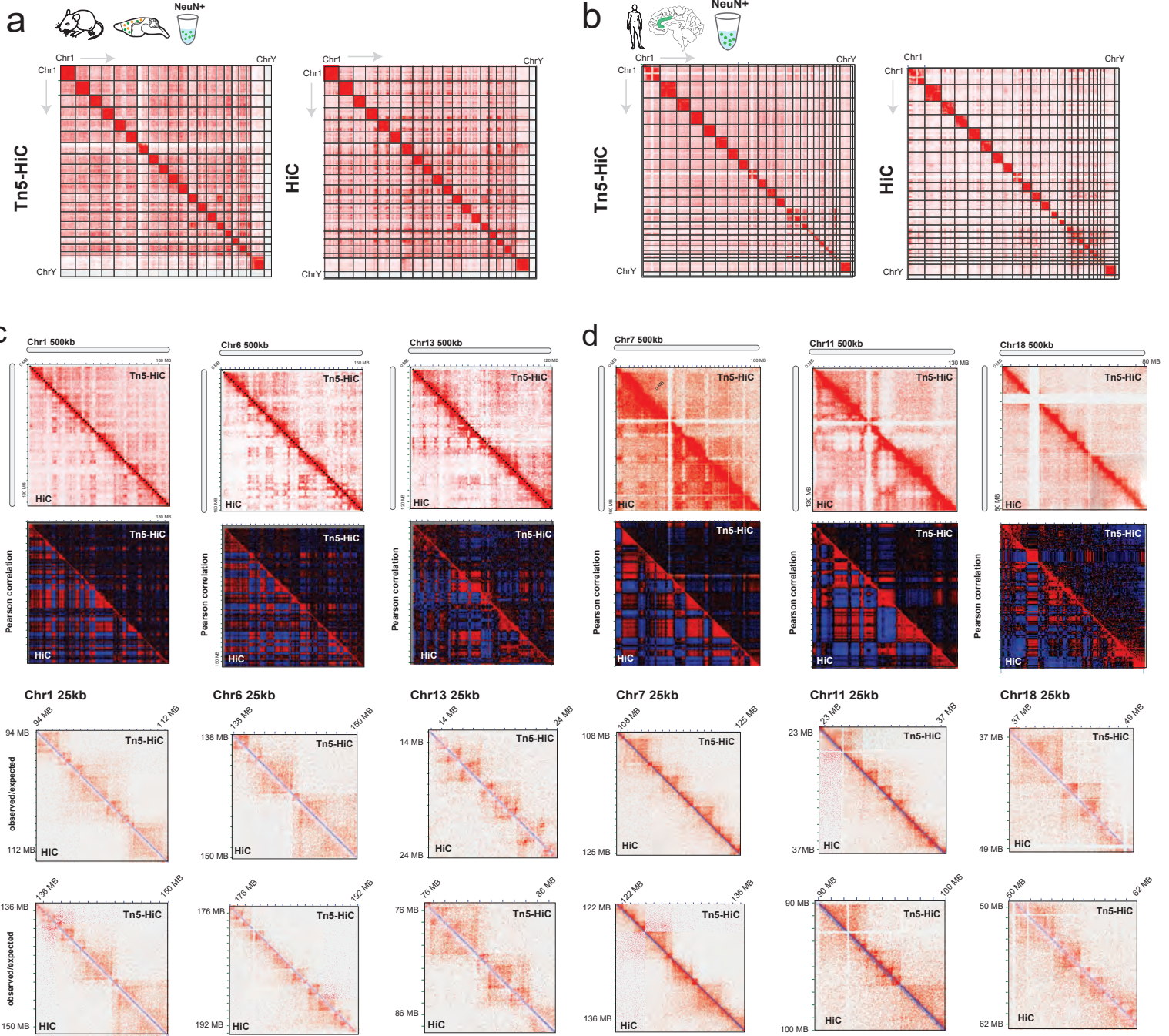
Figure S8



Supplementary Figure 8. Topological Associated Domains (TADs) and A/B compartment calling in $Tn5$ Hi-C and HiC.

Human and mouse NeuN⁺ $Tn5$ HiC_{800-1200bp} and conventional HiC libraries were subsampled and bootstrapped to allow for direct comparison. **a, d)** Number of TADs and average size of (top) human and (bottom) mouse libraries, analyzed with TADtree¹¹¹. Each datapoint = 1 autosomal chromosome. **b, e)** Loess regression curve to estimate correlation among A/B compartment calls from HiC and $Tn5$ HiC libraries, represented by the Pearson correlation values produced by chromosome-wide Eigenvector. **c, f)** Interaction matrix heatmap of human and mouse $Tn5$ HiC (upper right from diagonal) and HiC (lower left from diagonal) at 250kb resolution. Eigenvector plots showing A/B compartment calls along (top) human chromosome 12 and (bottom) mouse chromosome 6 (blue: $Tn5$ HiC, black: HiC).

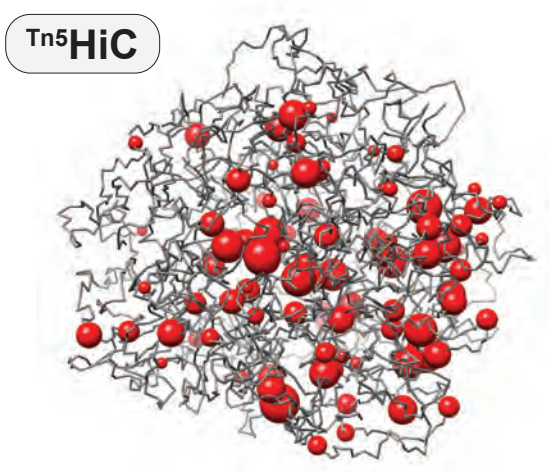
Figure S9



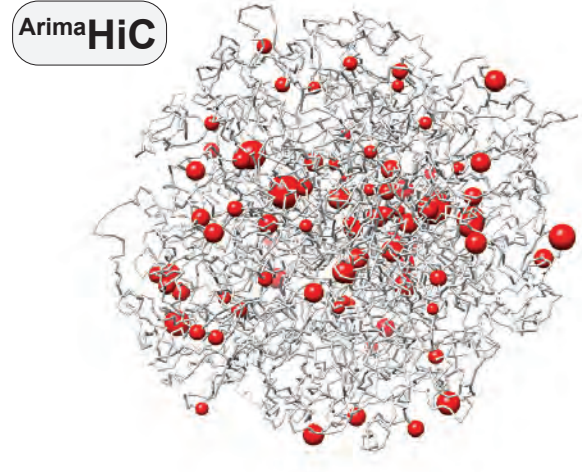
Supplementary Figure 9. Neuronal mouse and human Tn5-HiC interactions matrices. Interaction matrices examples of subsampled $Tn5$ HiC and HiC libraries NeuN⁺ samples. **a,b)** Genome-wide mouse (a) and human (b) heatmap interaction matrix at 2.5MB resolution of $Tn5$ HiC (left) and HiC (right). **c, d)** Interaction matrices of $Tn5$ HiC and HiC at (top) 500kb and (bottom) 25kb. Upper right from diagonal, $Tn5$ HiC and lower left from diagonal, HiC library. Also shown are Pearson correlation at 500kb.

Figure S10

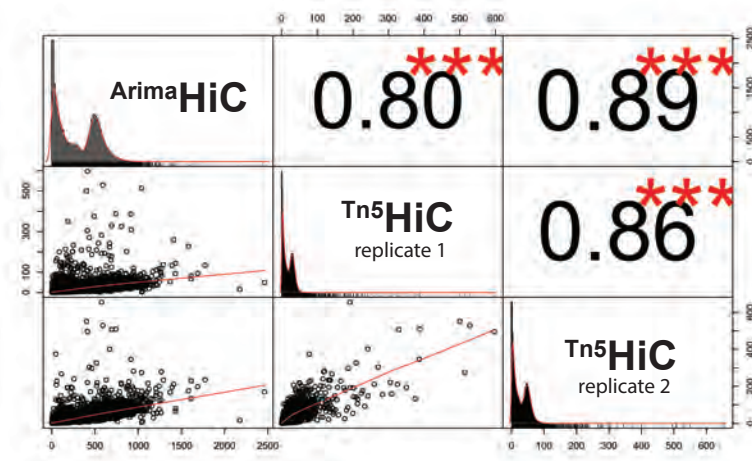
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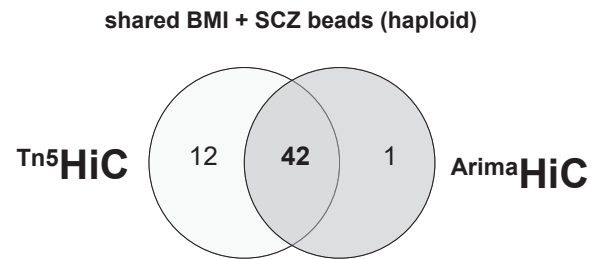
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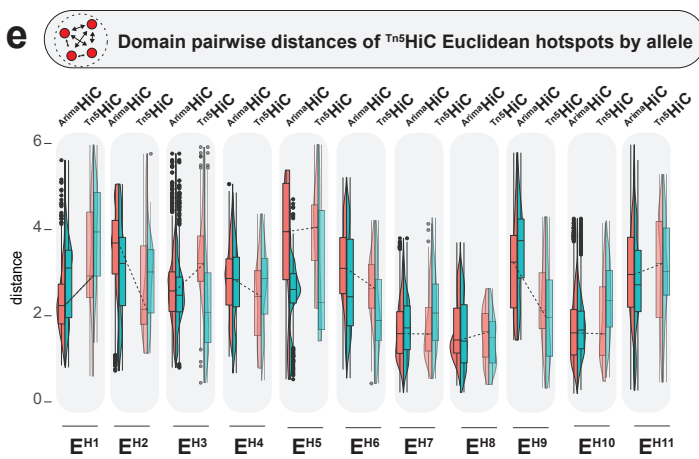
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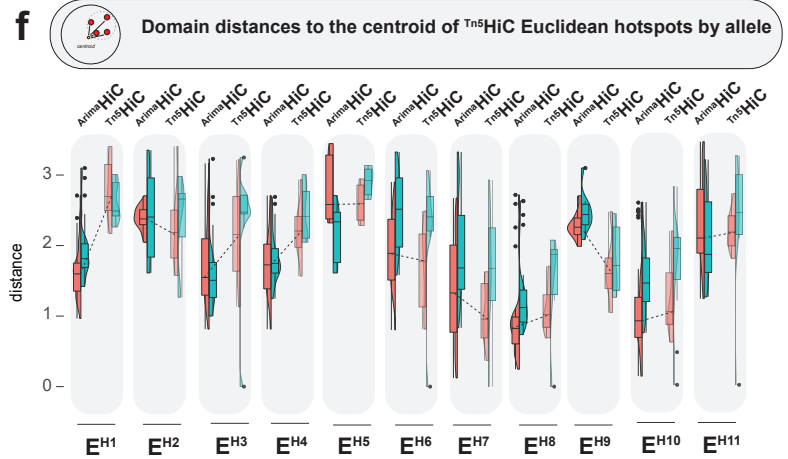
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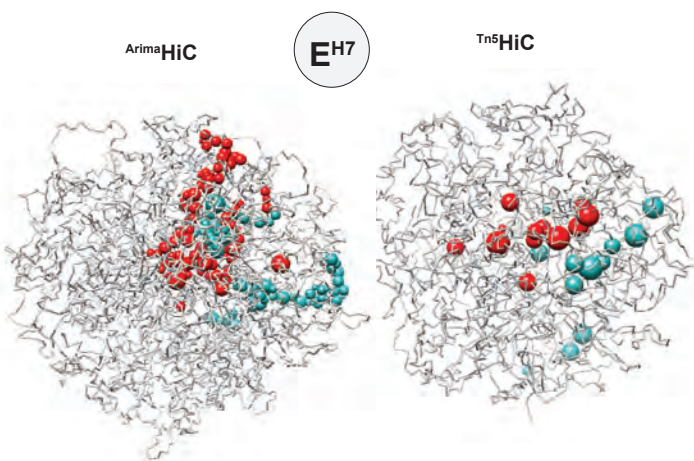
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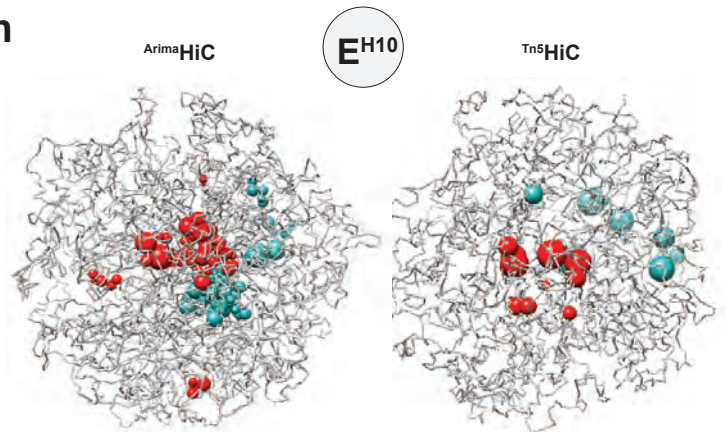
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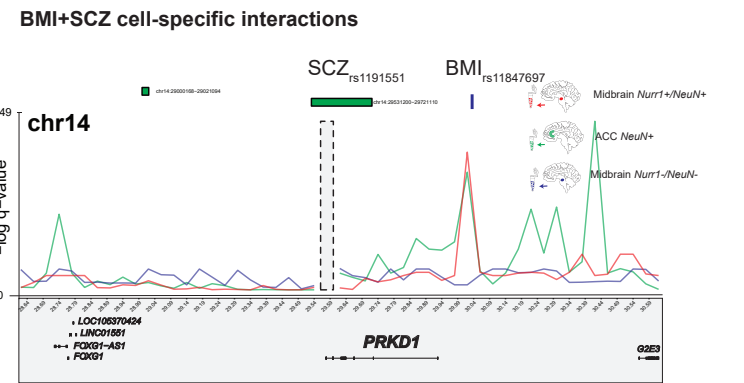
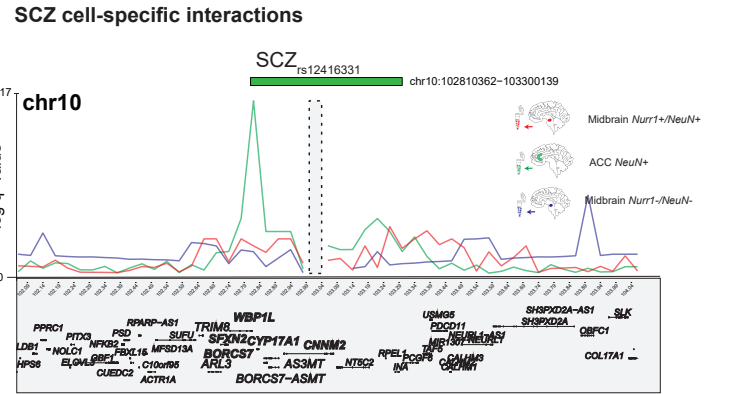
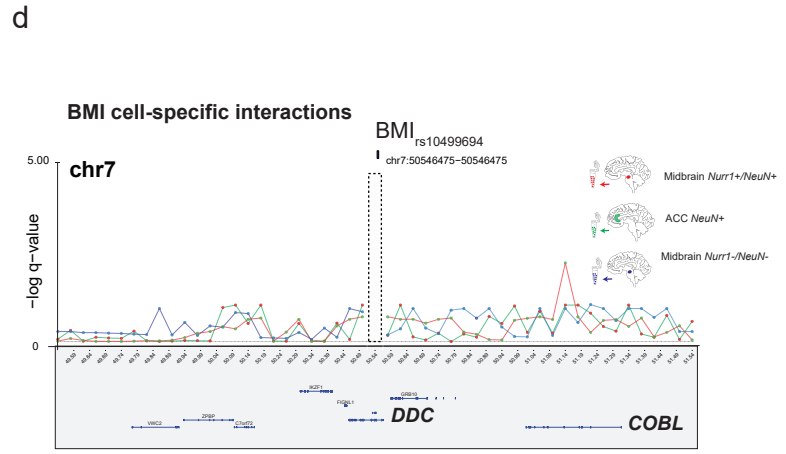
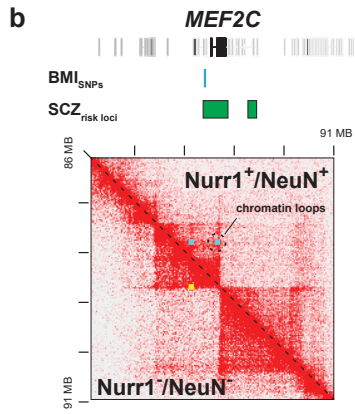
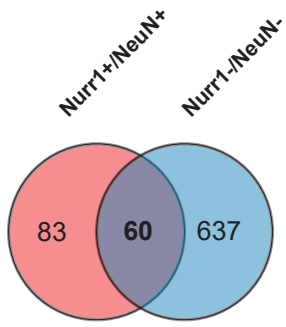
h



Supplementary Figure 10. *Nurr1*⁺/*NeuN*⁺ *Tn5*HiC and *Arima*HiC.

Comparison of *Tn5*HiC *Nurr1*⁺/*NeuN*⁺ and *Arima*HiC *Nurr1*⁺/*NeuN*⁺ domain-based chrom3D models. Note that due to the higher proportion of valid interactions pairs (~17% of the total number of pairs of *Tn5*HiC versus the ~35% of *Arima*HiC libraries, the resolution of the 3D model is considerably higher in *Arima*HiC libraries. **a, b**) *In silico* 3D models of MDN *Nurr1*⁺/*NeuN*⁺ spatial genomes. Note that domains (beads) are smaller in *Arima*HiC model (**b**, left) compared to *Tn5*HiC (**a**, right), reflecting better signal to noise. Average domain width, *Arima*Hi-C $1.16\text{Mb} \pm 1.75$ (median=0.9Mb) and *Tn5*Hi-C was $1.68\text{Mb} \pm 2.98\text{Mb}$ (median=0.95Mb) **c**) Correlation of intrachromosomal interactions at 40kb resolution genome-wide, of *Arima*HiC and two replicates of *Tn5*HiC libraries.*** $P < 0.001$. **d**) Venn plot of BMI and schizophrenia domains intersections, showing that 42/43 domains (called by the Arrowhead program in *Nurr1*⁺/*NeuN*⁺ *Arima*Hi-C) showing co-localization of BMI and SCZ risk sequences fully overlap with the 50 shared domains in *Nurr1*⁺/*NeuN*⁺ *Tn5*Hi-C. **e-g**) chrom3D diploid models of MDN *Nurr1*⁺/*NeuN*⁺ *Arima*Hi-C and *Tn5*Hi-C. Alleles represented by red and green colors, respectively. **e, f**) *Arima*HiC and *Tn5*HiC Euclidean distance comparison of **e**) pair-wise distances among distinct domains comprising Euclidean hotspots $E^{H1} - E^{H11}$ of the *Tn5*HiC *Nurr1*⁺/*NeuN*⁺ model and **f**) distances to the centroid for each hotspot ($E^{H1} - E^{H11}$) domains. **g**) Euclidean hotspot E^{H7} and **h**) Euclidean hotspot E^{H10} mapped in (left) *Arima*HiC and (right) *Tn5*Hi-C chrom3D diploid models.

a Figure S11

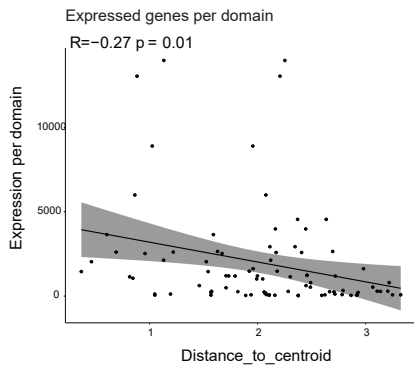


Supplementary Figure 11. Cell-specific chromosomal conformations enriched for cognitive and psychiatric traits

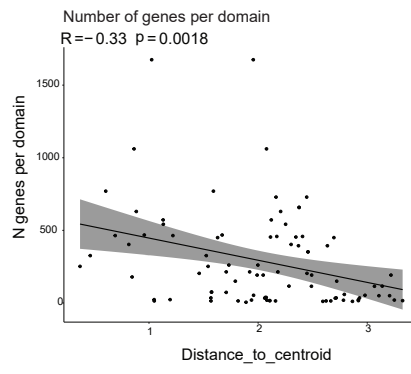
a) Venn intersection among $Nurr1^+/NeuN^+$ and $Nurr1^-/NeuN^-$ HICCUPS⁵⁴ for loops called significant both at 5 and 10kb resolution. **b)** Example of loop called for *MEF2C* genomic locus associated with BMI and schizophrenia risk. Above diagonal heatmap (25kb resolution) shows midbrain $Nurr1^+/NeuN^+$ contact map with two chromatin loops (marked in blue) and below diagonal heatmap shows $Nurr1^-/NeuN^-$ single loop. **c)** LD-score partitioned heritability⁶¹ plot using for midbrain $Nurr1^+/NeuN^+$ and $Nurr1^-/NeuN^-$ loops at 0.01 and 0.05 FDR thresholds (HICCUPS⁵⁴). **d)** Binomial chromosomal contact mapping at the site of specific BMI and SCZ risk polymorphisms. Contact enrichment over the genomic region of the highlighted bin (vertical gray bar) as indicated. Lines represent contact enrichments, (red) midbrain $Nurr1^+/NeuN^+$, (green) FC/ACC $NeuN^+$ and (blue) midbrain $Nurr1^-/NeuN^-$.

Figure S12

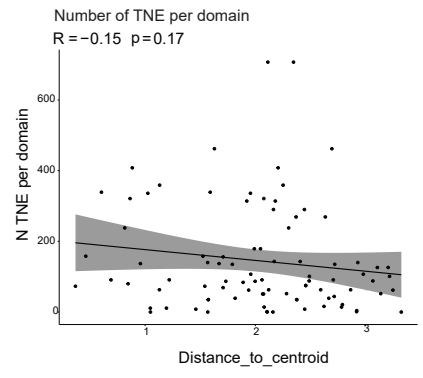
a



b



c



Supplementary Figure 12. Gene expression to nuclear centroid correlation analysis.

Correlations plots for gene expression, number of genes and TNE transcribed non-coding element per domain in relation with the proximity to the nuclear centroid. **a)** Expressed genes per domain to nuclear centroid. **b)** Number of genes per domain to nuclear centroid. **c)** Number of TNE per domain to nuclear centroid. For all plots, the 100 domains carrying both BMI and SCZ risk SNPs were used as input.

Euclidean hotspot domains to nuclear centroid

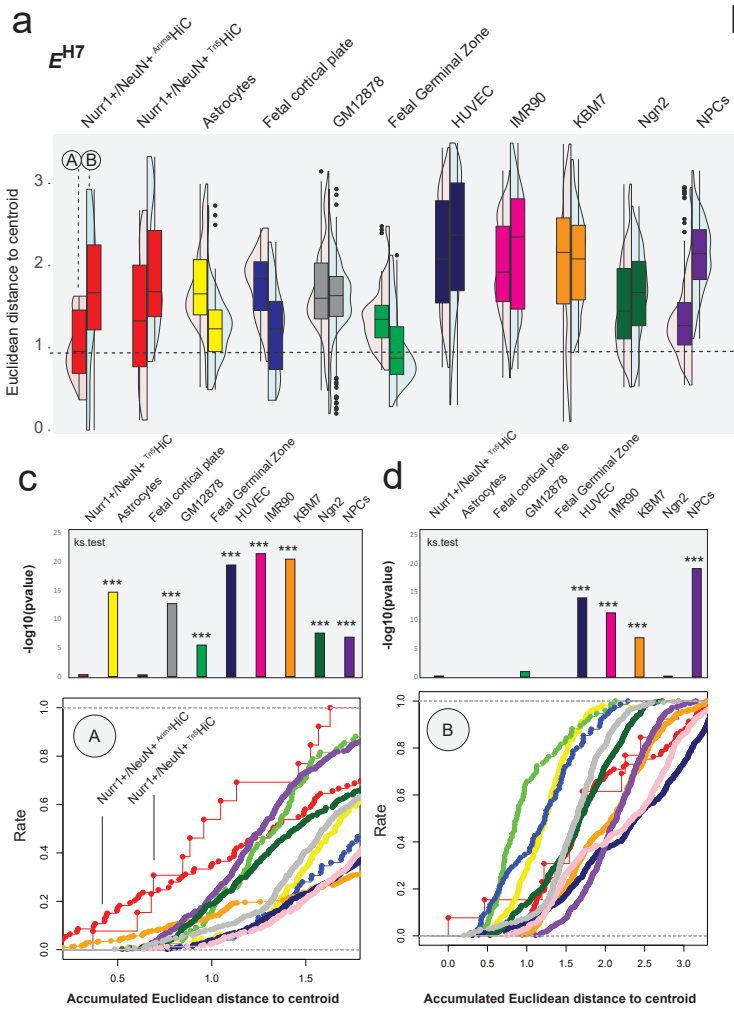
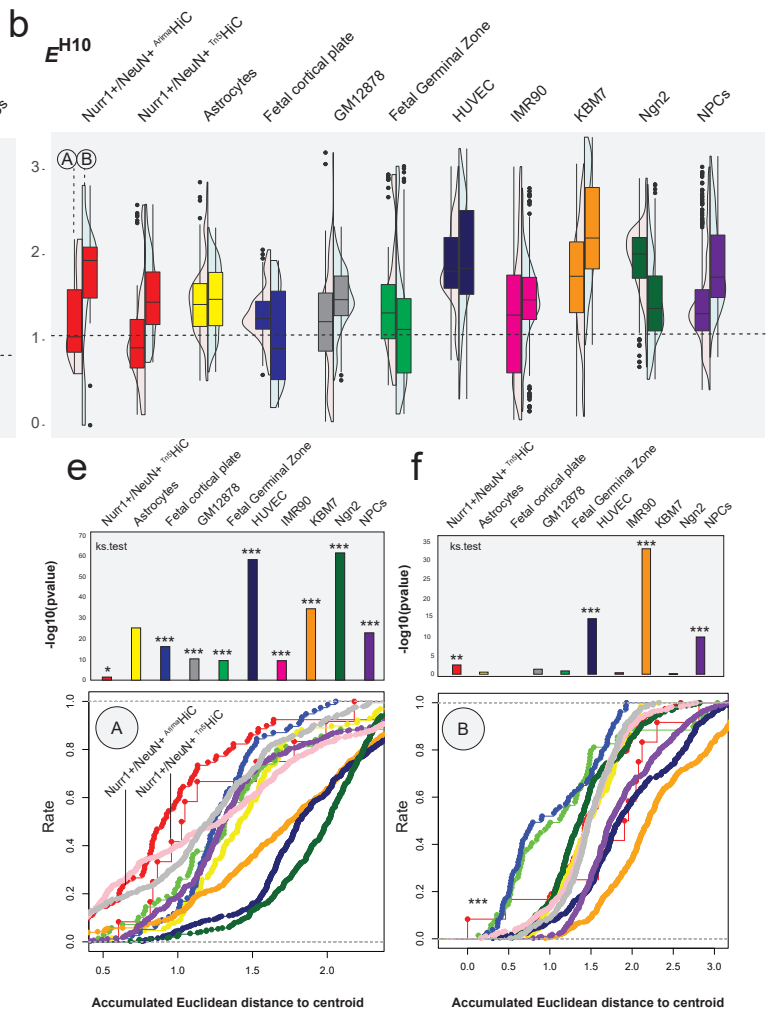
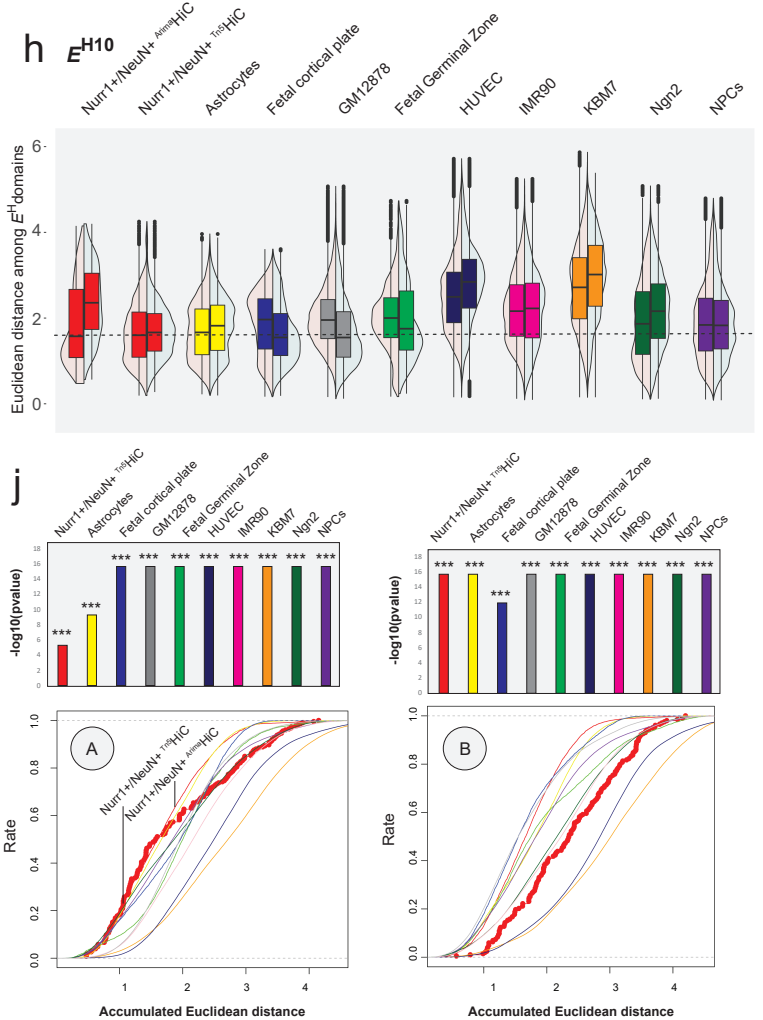
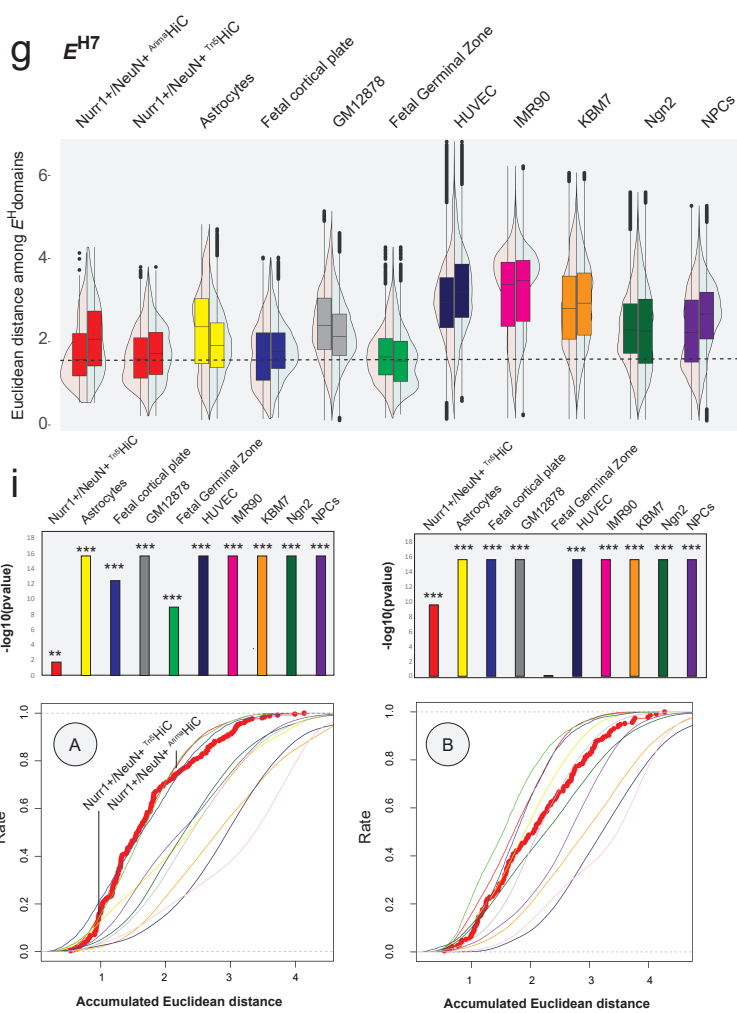


Figure S13



Pairwise distances among Euclidean hotspot domains

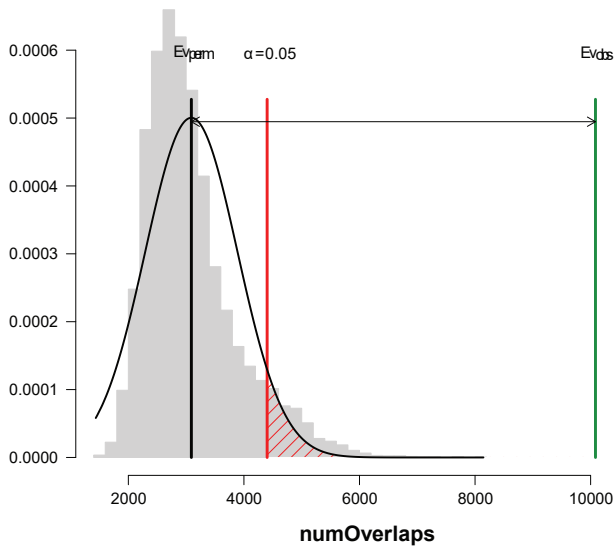


Supplementary Figure 13. Cell-type specific conformations of Euclidean hotspots E^{H7} and E^{H10} across multiple cell types.

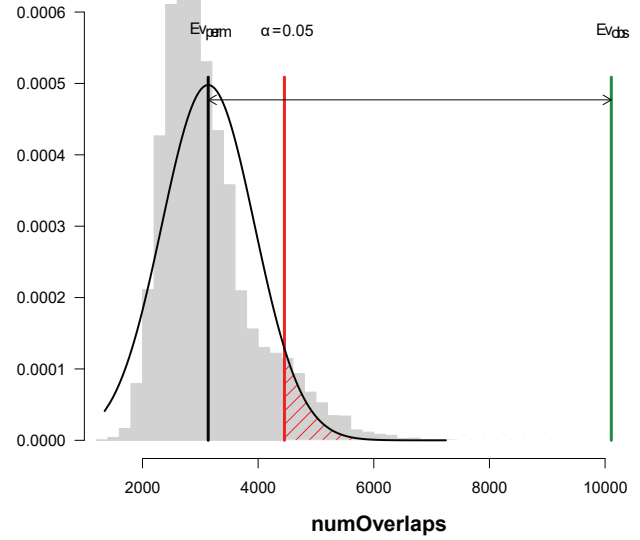
(a-f) Nuclear centroid and **(g-j)** pairwise distances of domains comprising E^{H7} and E^{H10} across 11 HiC datasets including two types of MDN Nurr1⁺/NeuN⁺ libraries (^{Arima}Hi-C and ^{Tn5}Hi-C), and 9 previously published Hi-C datasets generated from various other cell lines and tissues. **(a,g)** E^{H7} and **(b,h)** E^{H10} violin plots of **(a,b)** distance-to-nuclear centroid and **(g,h)** pairwise distances among hotspot domains in chrom3D diploid models, shown for both alleles 'A' and 'B' across multiple cell types, from left to right: (red boxplots) MDN Nurr1⁺/NeuN⁺ ^{Arima}HiC, MDN Nurr1⁺/NeuN⁺ ^{Tn5}HiC; (yellow boxplots) stem cell-derived astrocytes²⁸; (navy blue boxplots) fetal brain cortical plate²⁴; (gray boxplots) GM12878 cell line³⁰; (green boxplots) fetal brain germinal zone²⁴; (dark blue, pink and orange boxplots) HUVEC, IMR90 and KBM7 cell lines³⁰; (green and purple boxplots) Neurogenin 2 (NGN2) induced neurons and neural precursor cells (NPC)²⁸. Dashed line denotes median of 'A' domain distance-to-centroid for ^{Arima}HiC Nurr1⁺/NeuN⁺. Note that E^{H7} is biased towards A alleles while E^{H10} is 'monoallelic'. **(c, d)** E^{H7} and **(e,f)** E^{H10} ^{Arima}Hi-C library-to-other library comparison of domain distances to centroid shown separately for **(c,e)** 'A' and **(d,f)** 'B' alleles. **(i)** E^{H7} and **(j)** E^{H10} ^{Arima}Hi-C library-to-other library comparison of pairwise distances among hotspot domains shown separately (left) 'A' and (right) 'B' alleles. **(c-f)** and **(i,j)** (Kolmogorov–Smirnov test, *(**, ***) P< 0.05(0.01, 0.001).

Figure S14

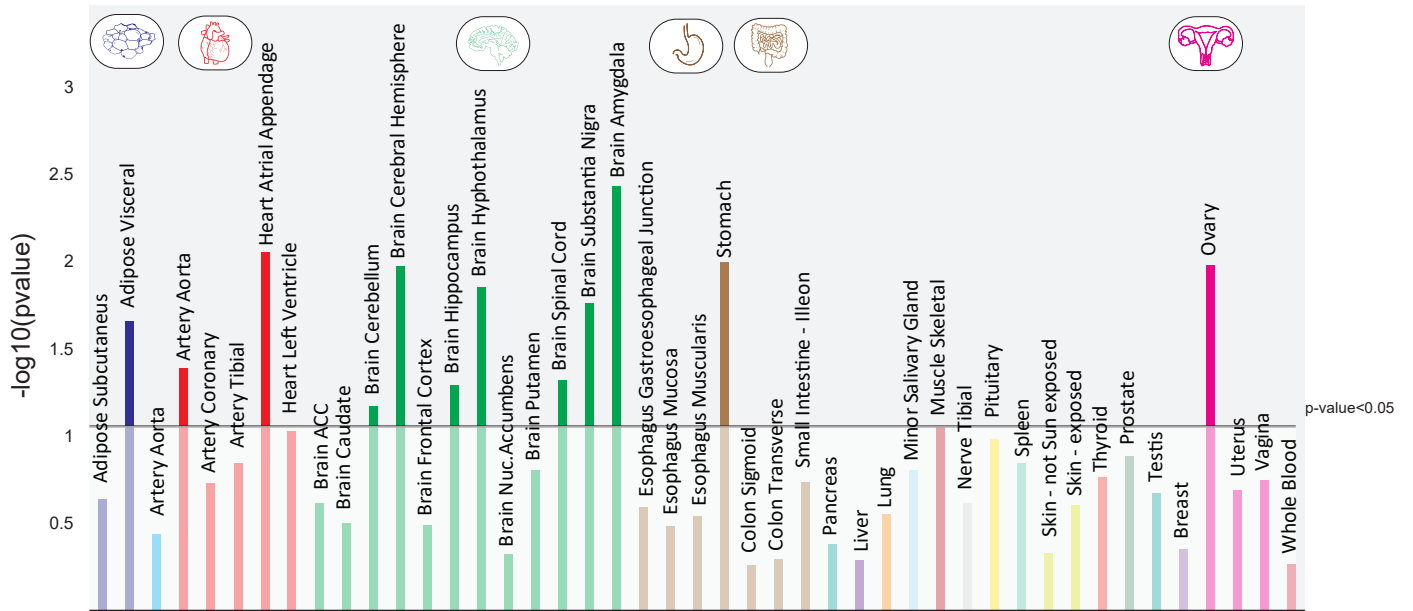
a **clumped** $r^{0.2}$ p -value: 0.0001
 Z -score: 8.771
 n perm: 10000
 randomization: resampleRegions



b **clumped** $r^{0.6}$ p -value: 0.0001
 Z -score: 8.698
 n perm: 10000
 randomization: resampleRegions



c



Supplementary Figure 14: 'Cross-disorder' chromosomal contacts interconnecting BMI with SCZ risk variants are enriched for tissue-specific eQTLs.

a,b) eQTL from EH 1-11, with clumping for each gene using the PLINK 1.90 software at a clump distance of 250kb and an r^2 of **a)** 0.2 and **b)** 0.6. Notice significant enrichment for eQTLs at the E^H 1-11. **c)** BMI-SCZ contacts associated with GTEX cis eQTLs unique to specific tissues, as indicated. Note significant enrichment for brain and adipocyte-rich tissues.