

**Software:** CLC Genomics Workbench 8

Black = applied for SP1,SP2 and SP3 data

Blue = applied for SP3 data

Green = applied for SP1 and SP2 data

## 1. Demultiplex

File: JGJ0HAZ01.sff

Define tags:

Barcode length: 11

Sequence: 1-1000

Set barcode options:

Search both strands = yes

Barcodes: MID sequences Roche (1-6)

Result handling:

Create list of reads without barcode = yes

Create report = no

Save = yes

## 1. Workflow Map reads to reference beta

### a. Trim sequences

Quality trimming:

Ambiguous trim = Yes

Ambiguous limit = 2

Quality trim = Yes

Quality limit = 0,05 = phred score = 20

Adapter trimming:

Trim adapter list = NA Trim adapter library 2

Use colorspace = No

Search on both strands: Yes

Sequence filtering:

Remove 5' terminal nucleotides = No Yes

Number of 5' terminal nucleotides = NA 30

Remove 3' terminal nucleotides = No Yes

Number of 3' terminal nucleotides = NA 30

Discard short reads = Yes

Minimum number of nucleotides in reads = 15

Discard long reads = Yes

Maximum number of nucleotides in reads = 1.000

Result handling:

Save discarded sequences = Yes

Save broken pairs = No

Create report = Yes

Result handling: Save

➔ SP3 data files saved as \*.fastq and shared via DataHub

### b. Map reads to reference data

Select sequencing reads

Trimmed reads. For 454 data enter both MID's.

References

Use reference files mentioned above. Consensus sequences per sample derived from the consensus sequences of the different institutions

Masking mode = No masking

Exclude annotated = NA

Include annotated only = NA

Mapping options:

Mismatch cost = 2

Cost of insertions and deletions = Affine gap cost

Insertion open cost = 7

Insertion extend cost = 2

Deletion open cost = 7

Deletion extend cost = 2

Length fraction = 0,7

Similarity fraction = 0,9

Global alignment = No

Non-specific match handling = Map randomly

Results handling:

Output mode = Create stand-alone read mappings

Create report = Yes

Collect un-mapped reads = Yes

Save = yes

## 2. Workflow Realign and detect variants

Local realignment

Realignment settings:

Realign unaligned ends = Yes

Multi-pass realignment = 2

Guidance-variant track = Not set

Result handling:

Output mode = Create reads track

Output track of realigned regions = No

Indels and Structural variants

Select read mappings

Locally realigned file

Select settings

P-Value threshold = 0,0001

Maximum number of mismatches = 3

Filter variants = Yes

Minimum number of reads = 2

Reference masking: NA

Result handling:

Create report = No

Create breakpoints = No

Create InDel variants = Yes

Create structural variations = No

Save = yes

Local realignment

## Realignment settings:

Realign unaligned ends = Yes  
Multi-pass realignment = 2  
Guidance-variant track = Locally realigned (InDel)-file  
Force realignment to guidance-variants = No

## Result handling:

Output mode = Create reads track  
Output track of realigned regions = No

## Low frequency variant detection

## Select read mappings

Locally realigned – locally realigned files

## Low frequency variant parameters

Required significance (%) = 1,0

## General filters

Ignore positions with coverage above = 100.000  
Restrict calling to target regions = Not set  
Ignore broken pairs = Yes  
Ignore non-specific matches = Reads  
Minimum coverage = 2  
Minimum count = 2  
Minimum frequency (%) = 1,0

## Noise filters

Base quality filter = Yes  
Neighborhood radius = 5  
Minimum central quality = 0  
Minimum neighborhood quality = 0  
Read direction filter = Yes  
Direction frequency (%) = 5,0  
Relative read direction filter = Yes  
Significance (%) = 1,0  
Read position filter = Yes  
Significance (%) = 1,0  
Remove pyro-error variants = No (454 data checked with and without, no difference for mSNV identification)

## Result handling:

Create track = Yes  
Create annotated table = Yes  
Create report = No