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Reporting Summary

Statistics

Nature Research wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Research policies, see <u>Authors & Referees</u> and the <u>Editorial Policy Checklist</u>.

For all statistical analys	es, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.				
n/a Confirmed					
☐ ☐ The exact sam	pple size (n) for each experimental group/condition, given as a discrete number and unit of measurement				
A statement of	A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly				
The statistical Only common to	The statistical test(s) used AND whether they are one- or two-sided Only common tests should be described solely by name; describe more complex techniques in the Methods section.				
A description	A description of all covariates tested				
A description	A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons				
A full descript AND variation	A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)				
For null hypothesis testing, the test statistic (e.g. <i>F</i> , <i>t</i> , <i>r</i>) with confidence intervals, effect sizes, degrees of freedom and <i>P</i> value noted Give <i>P</i> values as exact values whenever suitable.					
For Bayesian a	For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings				
For hierarchic	For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes				
\square Estimates of effect sizes (e.g. Cohen's d , Pearson's r), indicating how they were calculated					
'	Our web collection on <u>statistics for biologists</u> contains articles on many of the points above.				
Software and o	code				
Policy information abo	ut <u>availability of computer code</u>				
Data collection	Data is available on our data portal http://bio-bigdata.hrbmu.edu.cn/lmmLnc; Data sets were collected from public databases.				
Data analysis	R 3.6 was used for this study. R packages including ROCR, SummarizedExperiment, ConsensusClusterPlus, survival, pheatmap, ggplot2 were used in this manuscript.				
	om algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors/reviewers. deposition in a community repository (e.g. GitHub). See the Nature Research guidelines for submitting code & software for further information.				
Data					
Policy information abo					
- Accession codes, un - A list of figures that	include a <u>data availability statement</u> . This statement should provide the following information, where applicable: ique identifiers, or web links for publicly available datasets have associated raw data restrictions on data availability				
All data is available on ou	r database http://bio-bigdata.hrbmu.edu.cn/ImmLnc				
Field-speci	fic reporting				
<u>-</u>	elow that is the best fit for your research. If you are not sure, read the appropriate sections before making your selection.				
✓ Life sciences	Behavioural & social sciences				

Life sciences study design

All studies must disclose on these points even when the disclosure is negative.

Sample size All the cancer types were listed in supplementary tables. All analyses were based on >3 biological independent samples.

Data exclusions LncRNAs with expression values =0 in >30% samples were excluded in our analyses. This is based on previous published literature.

Replication All data analyses and findings are based on independent samples. The results were validated based on independent data sets. The

independent data sets were described in manuscript.

Randomization We randomly selected the same number of lncRNAs and performed the random test.

Blinding All data was obtained from public data resources, so blinding was not relevant.

Behavioural & social sciences study design

All studies must disclose on these points even when the disclosure is negative.

rotation must disclose on these points even when the disclosure is negative.

Study description

Briefly describe the study type including whether data are quantitative, qualitative, or mixed-methods (e.g. qualitative cross-sectional, quantitative experimental, mixed-methods case study).

Research sample

State the research sample (e.g. Harvard university undergraduates, villagers in rural India) and provide relevant demographic information (e.g. age, sex) and indicate whether the sample is representative. Provide a rationale for the study sample chosen. For studies involving existing datasets, please describe the dataset and source.

Sampling strategy

Describe the sampling procedure (e.g. random, snowball, stratified, convenience). Describe the statistical methods that were used to predetermine sample size OR if no sample-size calculation was performed, describe how sample sizes were chosen and provide a rationale for why these sample sizes are sufficient. For qualitative data, please indicate whether data saturation was considered, and what criteria

were used to decide that no further sampling was needed.

Data collection

Provide details about the data collection procedure, including the instruments or devices used to record the data (e.g. pen and paper, computer, eye tracker, video or audio equipment) whether anyone was present besides the participant(s) and the researcher, and whether the researcher was blind to experimental condition and/or the study hypothesis during data collection.

Timing Indicate the start and stop dates of data collection. If there is a gap between collection periods, state the dates for each sample cohort.

Data exclusions If no data were excluded from the analyses, state so OR if data were excluded, provide the exact number of exclusions and the rationale behind them, indicating whether exclusion criteria were pre-established.

Non-participation State how many participants dropped out/declined participation and the reason(s) given OR provide response rate OR state that no participants dropped out/declined participation.

Randomization | If participants were not allocated into experimental groups, state so OR describe how participants were allocated to groups, and if allocation was not random, describe how covariates were controlled.

Ecological, evolutionary & environmental sciences study design

All studies must disclose on these points even when the disclosure is negative.

Sampling strategy

Study description

Briefly describe the study. For quantitative data include treatment factors and interactions, design structure (e.g. factorial, nested, hierarchical), nature and number of experimental units and replicates.

Research sample

Describe the research sample (e.g. a group of tagged Passer domesticus, all Stenocereus thurberi within Organ Pipe Cactus National
Monument), and provide a rationale for the sample choice. When relevant, describe the organism taxa, source, sex, age range and
any manipulations. State what population the sample is meant to represent when applicable. For studies involving existing datasets,

describe the data and its source.

Note the sampling procedure. Describe the statistical methods that were used to predetermine sample size OR if no sample-size

calculation was performed, describe how sample sizes were chosen and provide a rationale for why these sample sizes are sufficient.

Data collection

Describe the data collection procedure, including who recorded the data and how.

Timing and spatial scale Indicate the start and stop dates of data collection, noting the frequency and periodicity of sampling and providing a rationale for

riiiiii 6 arra opatiar ocaro	these choices. If there is a gap between collection periods, state the dates for each sample cohort. Specify the spatial scale from which the data are taken				
	If no data were excluded from the analyses, state so OR if data were excluded, describe the exclusions and the rationale behind them, indicating whether exclusion criteria were pre-established.				
1 /	Describe the measures taken to verify the reproducibility of experimental findings. For each experiment, note whether any attempts to repeat the experiment failed OR state that all attempts to repeat the experiment were successful.				
	Describe how samples/organisms/participants were allocated into groups. If allocation was not random, describe how covariates were controlled. If this is not relevant to your study, explain why.				
0	Describe the extent of blinding used during data acquisition and analysis. If blinding was not possible, describe why OR explain why blinding was not relevant to your study.				
Did the study involve field	work? Yes No				
ield work, collect	ion and transport				
Field conditions	Describe the study conditions for field work, providing relevant parameters (e.g. temperature, rainfall).				
Location	State the location of the sampling or experiment, providing relevant parameters (e.g. latitude and longitude, elevation, water depth).				
Access and import/export	Describe the efforts you have made to access habitats and to collect and import/export your samples in a responsible manner and in compliance with local, national and international laws, noting any permits that were obtained (give the name of the issuing authority, the date of issue, and any identifying information).				
Disturbance	Describe any disturbance caused by the study and how it was minimized.				
Materials & experimer /a Involved in the study	Methods n/a Involved in the study ChIP-seq Flow cytometry MRI-based neuroimaging				
Animals and other or Human research part Clinical data					
Antibodies used	Describe all antibodies used in the study; as applicable, provide supplier name, catalog number, clone name, and lot number.				
Validation	Describe all antibodies used in the study; as applicable, provide supplier name, catalog number, clone name, and lot number.				
	Describe all antibodies used in the study; as applicable, provide supplier name, catalog number, clone name, and lot number. Describe the validation of each primary antibody for the species and application, noting any validation statements on the manufacturer's website, relevant citations, antibody profiles in online databases, or data provided in the manuscript.				
ukaryotic cell line	Describe the validation of each primary antibody for the species and application, noting any validation statements on the manufacturer's website, relevant citations, antibody profiles in online databases, or data provided in the manuscript.				
-	Describe the validation of each primary antibody for the species and application, noting any validation statements on the manufacturer's website, relevant citations, antibody profiles in online databases, or data provided in the manuscript.				
olicy information about <u>cel</u>	Describe the validation of each primary antibody for the species and application, noting any validation statements on the manufacturer's website, relevant citations, antibody profiles in online databases, or data provided in the manuscript.				
cukaryotic cell line olicy information about cell line source(s) Authentication	Describe the validation of each primary antibody for the species and application, noting any validation statements on the manufacturer's website, relevant citations, antibody profiles in online databases, or data provided in the manuscript. 2S Il lines				

Name any commonly misidentified cell lines used in the study and provide a rationale for their use.

Commonly misidentified lines (See <u>ICLAC</u> register)

Palaeontology

Specimen provenance Provide provenance information for specimens and describe permits that were obtained for the work (including the name of the issuing authority, the date of issue, and any identifying information).

Specimen deposition Indicate where the specimens have been deposited to permit free access by other researchers.

Dating methods

If new dates are provided, describe how they were obtained (e.g. collection, storage, sample pretreatment and measurement), where they were obtained (i.e. lab name), the calibration program and the protocol for quality assurance OR state that no new dates are provided.

Tick this box to confirm that the raw and calibrated dates are available in the paper or in Supplementary Information.

Animals and other organisms

Policy information about studies involving animals; ARRIVE guidelines recommended for reporting animal research

Laboratory animals For laboratory animals, report species, strain, sex and age OR state that the study did not involve laboratory animals.

Wild animals

Provide details on animals observed in or captured in the field; report species, sex and age where possible. Describe how animals were caught and transported and what happened to captive animals after the study (if killed, explain why and describe method; if

released, say where and when) OR state that the study did not involve wild animals.

Field-collected samples

For laboratory work with field-collected samples, describe all relevant parameters such as housing, maintenance, temperature, photoperiod and end-of-experiment protocol OR state that the study did not involve samples collected from the field.

Ethics oversight Identify the organization(s) that approved or provided guidance on the study protocol, OR state that no ethical approval or guidance was required and explain why not.

Note that full information on the approval of the study protocol must also be provided in the manuscript.

Human research participants

Policy information about studies involving human research participants

Population characteristics Describe the covariate-relevant population characteristics of the human research participants (e.g. age, gender, genotypic

information, past and current diagnosis and treatment categories). If you filled out the behavioural & social sciences study design

questions and have nothing to add here, write "See above."

RecruitmentDescribe how participants were recruited. Outline any potential self-selection bias or other biases that may be present and how

these are likely to impact results.

Ethics oversight | Identify the organization(s) that approved the study protocol.

Note that full information on the approval of the study protocol must also be provided in the manuscript.

Clinical data

Policy information about <u>clinical studies</u>

All manuscripts should comply with the ICMJE guidelines for publication of clinical research and a completed CONSORT checklist must be included with all submissions.

Clinical trial registration Provide the trial registration number from ClinicalTrials.gov or an equivalent agency.

Study protocol Note where the full trial protocol can be accessed OR if not available, explain why.

Data collection Describe the settings and locales of data collection, noting the time periods of recruitment and data collection.

Outcomes Describe how you pre-defined primary and secondary outcome measures and how you assessed these measures.

ChIP-seq

Data deposition

Confirm that both raw and final processed data have been deposited in a public database such as GEO.

Confirm that you have deposited or provided access to graph files (e.g. BED files) for the called peaks.

Data access links

May remain private before publication.

For "Initial submission" or "Revised version" documents, provide reviewer access links. For your "Final submission" document, provide a link to the deposited data.

Files in database submission	Provide a list of all files available in the database submission.		
Genome browser session (e.g. <u>UCSC</u>)	Provide a link to an anonymized genome browser session for "Initial submission" and "Revised version" documents only, to enable peer review. Write "no longer applicable" for "Final submission" documents.		
Methodology			
Replicates	Describe the experimental replicates, specifying number, type and replicate agreement.		
Sequencing depth	Describe the sequencing depth for each experiment, providing the total number of reads, uniquely mapped reads, length of reads and whether they were paired- or single-end.		
Antibodies	Describe the antibodies used for the ChIP-seq experiments; as applicable, provide supplier name, catalog number, clone name, and lot number.		
Peak calling parameters	Specify the command line program and parameters used for read mapping and peak calling, including the ChIP, control and index files used.		
Data quality	Describe the methods used to ensure data quality in full detail, including how many peaks are at FDR 5% and above 5-fold enrichment.		
Software	Describe the software used to collect and analyze the ChIP-seq data. For custom code that has been deposited into a community repository, provide accession details.		
Flow Cytometry			
Plots			
Confirm that:			
	narker and fluorochrome used (e.g. CD4-FITC).		
	visible. Include numbers along axes only for bottom left plot of group (a 'group' is an analysis of identical markers		
All plots are contour plots	s with outliers or pseudocolor plots.		
A numerical value for nur	nber of cells or percentage (with statistics) is provided.		
/lethodology			
Sample preparation	Describe the sample preparation, detailing the biological source of the cells and any tissue processing steps used.		
Instrument	Identify the instrument used for data collection, specifying make and model number.		
Software	Describe the software used to collect and analyze the flow cytometry data. For custom code that has been deposited into a community repository, provide accession details.		
Cell population abundance	Describe the abundance of the relevant cell populations within post-sort fractions, providing details on the purity of the samples and how it was determined.		
Gating strategy	Describe the gating strategy used for all relevant experiments, specifying the preliminary FSC/SSC gates of the starting cell		

Magnetic resonance imaging

Experimental design

Design type

Indicate task or resting state; event-related or block design.

Tick this box to confirm that a figure exemplifying the gating strategy is provided in the Supplementary Information.

Design specifications

Specify the number of blocks, trials or experimental units per session and/or subject, and specify the length of each trial or block (if trials are blocked) and interval between trials.

population, indicating where boundaries between "positive" and "negative" staining cell populations are defined.

Behavioral performance measures

State number and/or type of variables recorded (e.g. correct button press, response time) and what statistics were used to establish that the subjects were performing the task as expected (e.g. mean, range, and/or standard deviation across subjects).

Acquisition				
Imaging type(s)	Specify: functional, structural, diffusion, perfusion.			
Field strength	Specify in Tesla			
Sequence & imaging parameters	Specify the pulse sequence type (gradient echo, spin echo, etc.), imaging type (EPI, spiral, etc.), field of view, matrix size, slice thickness, orientation and TE/TR/flip angle.			
Area of acquisition	State whether a whole brain scan was used OR define the area of acquisition, describing how the region was determined.			
Diffusion MRI Used	Not use	ed		
Preprocessing				
Preprocessing software	Provide detail on software version and revision number and on specific parameters (model/functions, brain extraction, segmentation, smoothing kernel size, etc.).			
Normalization	If data were normalized/standardized, describe the approach(es): specify linear or non-linear and define image types used for transformation OR indicate that data were not normalized and explain rationale for lack of normalization.			
Normalization template	Describe the template used for normalization/transformation, specifying subject space or group standardized space (e.g. original Talairach, MNI305, ICBM152) OR indicate that the data were not normalized.			
Noise and artifact removal	Describe your procedure(s) for artifact and structured noise removal, specifying motion parameters, tissue signals and physiological signals (heart rate, respiration).			
Volume censoring	Define your software and/or method and criteria for volume censoring, and state the extent of such censoring.			
Statistical modeling & inferenc	e			
Model type and settings	Specify type (mass univariate, multivariate, RSA, predictive, etc.) and describe essential details of the model at the first and second levels (e.g. fixed, random or mixed effects; drift or auto-correlation).			
Effect(s) tested	Define precise effect in terms of the task or stimulus conditions instead of psychological concepts and indicate whether ANOVA or factorial designs were used.			
Specify type of analysis: Whole brain ROI-based Both				
Statistic type for inference (See Eklund et al. 2016)	Specify voxel-wise or cluster-wise and report all relevant parameters for cluster-wise methods.			
Correction	Describe the type of correction and how it is obtained for multiple comparisons (e.g. FWE, FDR, permutation or Monte Carlo).			
Models & analysis				
n/a Involved in the study Functional and/or effective co Graph analysis Multivariate modeling or pred				
Functional and/or effective connectivity		Report the measures of dependence used and the model details (e.g. Pearson correlation, partial correlation, mutual information).		
Graph analysis		Report the dependent variable and connectivity measure, specifying weighted graph or binarized graph, subject- or group-level, and the global and/or node summaries used (e.g. clustering coefficient, efficiency, etc.).		
Multivariate modeling and predictive analysis		Specify independent variables, features extraction and dimension reduction, model, training and evaluation metrics.		