



Supplementary Materials

Structural diversity of sense and antisense RNA hexanucleotide repeats associated with ALS and FTLD

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Figure S1. Canonical and non-canonical base pairs. Schematic presentation of **a**) Watson-Crick G-C base pair, **b**) hemi-protonated C⁺-C base pair and **c**) G-G base pair in N1-carbonyl symmetric geometry.



Figure S2. Cytosine H5/H6 correlation region of 2D TOCSY NMR spectrum of $r(G_2C_4)_4$. Spectrum was recorded at pH 7.0 and 25 °C at mixing time of 80 ms and concentration of 0.9 mM per strand.



Figure S3. ¹H NMR spectra of r(G₂C₄)₄ in 10% ²H₂O recorded at 5, 25 and 37 °C. At 25 °C the pH was 7.0. All NMR spectra were acquired at concentration of 0.9 mM per strand.



Figure S4. UV melting experiment of homodimer adopted by $r(G_2C_4)_4$. The absorbance at 260 nm was measured during one cycle of heating (black) and cooling (red) the sample in the temperature range between 20 and 90 °C. Temperature of half transition for homodimer adopted by $r(G_2C_4)_4$ was determined for all four oligonucleotide concentrations from melting curve obtained during heating. All samples were prepared in 20 mM Li⁺ cacodylate buffer with pH 7.0. Concentrations of $r(G_2C_4)_4$ were **a**) 2.6, **b**) 3.0, **c**) 7.7, **d**) 9.3 and **e**) 20 μ M per strand.



Figure S5. Imino region of ¹H NMR spectra of $r(G_2C_4)_4$ in 10% ²H₂O at different pH. All NMR spectra were acquired at 25 °C and 0.3 mM oligonucleotide concentration per strand. Signal corresponding to hemi-protonated C⁺-C base pairs is marked with an arrow.

Table 1. Translational diffusion coefficients (Dt) of $r(G_2C_4)_4$ at different pH values. All 2D DOSY spectra were acquired at 25 °C and concentration of 0.3 mM per strand. Standard deviation of the data set was ± 0.05 .

рН	Dt [* 10^(-10) m ² s ⁻¹]
7.0	0.91
6.5	0.94
6.0	1.25
5.5	1.30
5.0	1.35
4.5	1.40



Figure S6. Imino region of ¹H NMR spectra of $r(G_2C_4)_4$ in 10% ²H₂O at different pH and in the presence of 10% *w*/*v* PEG. All NMR spectra were acquired at 25 °C and concentration of 0.3 mM per strand. The vertical scales of the spectra with pH ranging from 5.5 to 4.5 have been increased six-fold (6×).



Figure S7. CD spectra of $r(G_2C_4)_4$ with pH ranging from 4.5 to 7.0. Arrows represent the direction of change in pH. All CD spectra were acquired at 25 °C and concentration of 100 μ M per strand.



Figure S8. 20% native PAGE of r(G₂C₄)₄ at pH 6.0 and 5 °C. Concentration of r(G₂C₄)₄ was 0.3 mM per strand. DNA ladder was used as a standard.



Figure S9. Imino-imino spectral region of 2D NOESY spectrum of $r(G_2C_4)_4$. Spectrum was recorded at pH 4.5 and 5 °C at mixing time of 100 ms and concentration of 0.7 mM per strand. Above 2D NOESY spectrum is 1D ¹H NMR spectrum of $r(G_2C_4)_4$ recorded at pH 4.5, 5 °C and oligonucleotide concentration of 0.7 mM per strand. NOE contact (marked with red circle) confirms the presence of hemi-protonated C⁺-C base pairs within the hairpin.



Figure S10. Imino regions of ¹H NMR spectra of $r(G_2C_4)_4$ in 10% ²H₂O at 5, 25 and 40 °C. All NMR spectra were acquired at concentration of 0.3 mM per strand. The pH was 4.5 at 25 °C. Numbers under signals represent integral values. The vertical scales of the spectral regions between δ 15.6 to 16.8 ppm have been increased six-fold (6×).



Figure S11. UV melting experiment of hairpin adopted by $r(G_2C_4)_4$. The absorbance at 260 nm was measured during one cycle of heating (black) and cooling (red) the sample in the temperature range between 20 and 90 °C. All samples were prepared in 20 mM Li⁺ cacodylate buffer with pH 5.0. Concentrations of $r(G_2C_4)_4$ were **a**) 2.6, **b**) 3.0, **c**) 7.7, **d**) 9.3 and **e**) 20 μ M per strand.



Figure S12. ¹H NMR spectra of $r(G_4C_2)_4$ in 10% ²H₂O at 0, 7, 15, 20, 25 and 37 °C. All NMR spectra were acquired at concentration of 0.3 mM per strand. The pH was 6.0 at 25 °C.



Figure S13. Imino regions of ¹H NMR spectra of $r(G_4C_2)_4$ in 10% ²H₂O at 0 and 25 °C. Both NMR spectra were acquired at pH 6.0 and concentration of 0.3 mM per strand. Numbers under signals represent integral values.



Figure S14. Imino and aromatic region of ¹H NMR spectra of $r(G_4C_2)_4$ in 10% ²H₂O at different pH. Arrows represent the direction of change in pH. All NMR spectra were acquired on 800 MHz spectrometer at 25 °C and concentration of 0.3 mM per strand.



Figure S15. Imino regions of ¹H NMR spectra of r(G₄C₂)₄ in 10% ²H₂O at concentration of 0.05, 0.1, 0.2, 0.3, and 1.1 mM per strand. All NMR spectra were acquired at pH 6.0 and 25 °C. Numbers under signals represent integral values.

standard [bp]	r(G₄C₂)₄ 0.3 mM	r(G₄C₂)₄ 0.2 mM	r(G₄C₂)₄ 0.1 mM
50	* .		
35			
25			-
20			
		1	
15			

Figure S16. 20% native PAGE of $r(G_4C_2)_4$ at pH 6.0 and 5 °C. Concentration of $r(G_4C_2)_4$ was 0.3, 0.2 and 0.1 mM per strand. DNA ladder was used as a standard.



Figure S17. CD spectra of equimolar mixture of $r(G_2C_4)_4$ and $r(G_4C_2)_4$ before (black) and after (red) annealing. Both CD spectra were acquired at pH 6.0, 25 °C and concentration of 100 μ M per sense and antisense strand.



Figure S18. ¹H NMR spectra of equimolar mixture of $r(G_2C_4)_4$ and $r(G_2C_4)_4$ before and after annealing. Δ represents NMR spectrum after annealing. Both NMR spectra were acquired at 25 °C, pH 6.0 and concentration of 0.2 mM per sense and antisense strand.



Figure S19. 20% native PAGE of $r(G_4C_2)_4$, $r(G_2C_4)_4$ and $r(G_4C_2)_4 + r(G_2C_4)_4$ at pH 6.0 and 5 °C. In all samples concentration of $r(G_4C_2)_4$ and $r(G_2C_4)_4$ was 0.1 mM per strand. DNA ladder was used as a standard.