

SUPPLEMENTARY MATERIAL

Innate T- $\alpha\beta$ lymphocytes as new immunological components of anti-tumoral “off-target” effects of the tyrosine kinase inhibitor dasatinib

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Supplementary methods

Supplementary Table 1. CML Patients characteristics.

Group	n	Age range	Age median	Age mean	Sex ratio
DasaPEG	54	20-65	48	46	1.7

All patients were confirmed Philadelphia chromosome-positive.

Supplementary Table 2.

Anti-mouse antibodies used in this study.

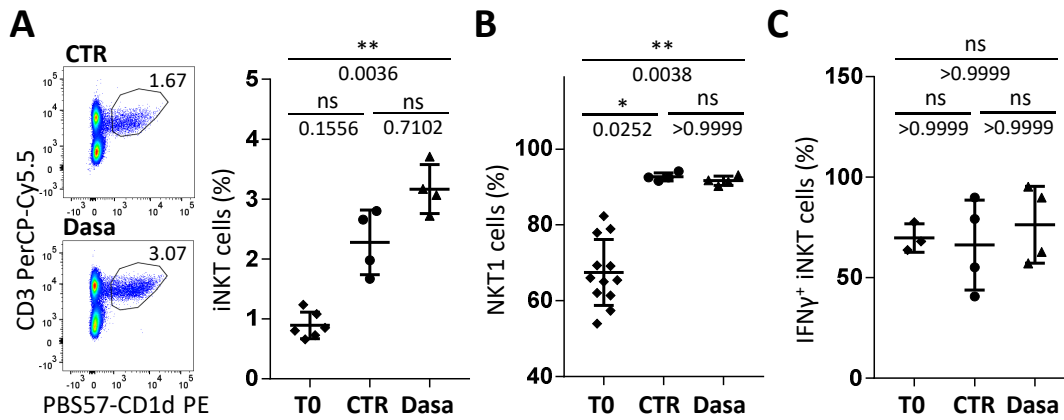
Antibody	Fluorochrome	Clone	Supplier	Reference
CD69	BV421	H1.2F3	BD Biosciences	562920
CD3 ϵ	PerCP-Cy5.5	17A2	BD Biosciences	100218
TCR- β	PerCP-Cy5.5	H57-597	BioLegend	109228
CD8	BV510	53-6.7	BD Biosciences	563068
CD44	PE-Cy7	IM7	BD Biosciences	560569
CD62L	PE	MEL-14	BD Biosciences	553151
CD49d	Vioblue	R1-2	Miltenyi	130-102-404
CD122	APC	TM-B1	BioLegend	123214
PLZF	AF488	Mags.21F7	eBioscience	53-9320-82
ROR- γ T	APC	B2D	eBioscience	17-6981-82
T-bet	PE-Cy7	4B10	BioLegend	644823
panNK CD49b	APC-Cy7	DX5	BioLegend	108920
CD4	PE	RM4-5	BD Biosciences	553049
CD24	PE	M1/69	BD Biosciences	553262
Eomes	AF488	Dan11mag	eBioscience	53-4875
IFN γ	PE/PE-Cy7	XMG1.2	BD Biosciences	554412/557649

Supplementary Table 3.

Anti-human antibodies used in this study.

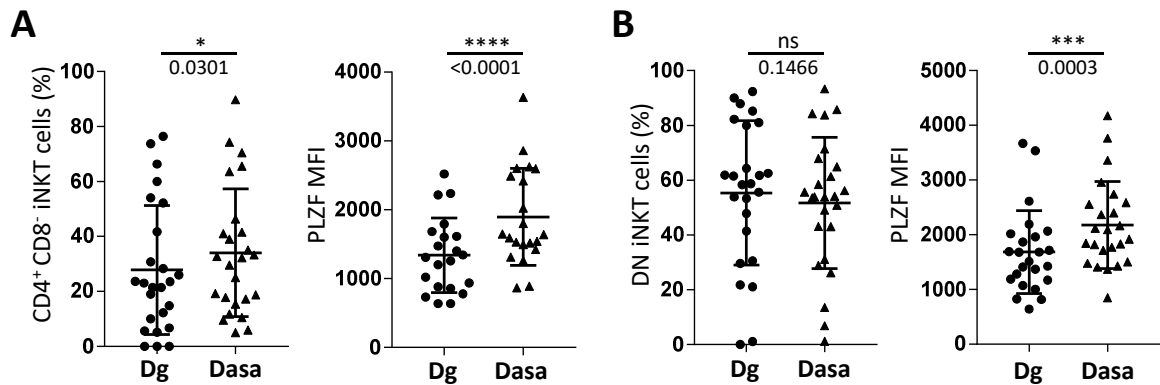
Antibody	Fluorochrome	Clone	Supplier	Reference
TCR- $\alpha\beta$	BV421	IP26	BioLegend	306722
CD3	BV421	UCHT1	BioLegend	300434
CD161	PerCP-Cy5.5	HP-3G10	BioLegend	339908
CD49d	BV510	9F10	BioLegend	304318
CD4	FITC	RPA-T4	BioLegend	300506
TCR V α 24-J α 18	APC	6B11	BioLegend	342908
CD8	PE-Cy7	RPA-T8	BD Pharmingen	557746
Eomes	eFluor®660	WD1928	eBiosciences	50-4877-42
PLZF	PE	P3.62.8.1	eBiosciences	12-4714-82
KIR2D	PE	NKVFS1	Miltenyi Biotec	130-092-688
KIR3DL1/KIR3DL2 (CD158e/k)	PE	5.133	Miltenyi Biotec	130-095-205
NKG2A (CD159a)	PE	REA110	Miltenyi Biotec	130-098-814

Supplementary Figure 1



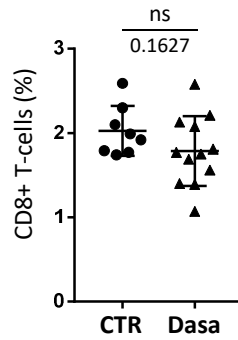
Supplementary Figure 1. IL-15 promotes Th1 differentiation of iNKT cells *in vitro*. (A, B and C) BALB/c Eomes-GFP derived splenocytes were directly analyzed (T0, n=12) or cultured 7 days in the presence of IL-15 with (Dasa, n=4) or without (CTR, n=4) dasatinib. iNKT cells percentages (A) and differentiation (B) into NKT1 subtype (T-bet⁺ PLZF^{int}) were analyzed in live lymphocytes by flow cytometry. (C) Splenocytes were further stimulated for 16h with IL-12 and IL-18 and IFN γ secretion was analyzed in iNKT cells. Statistical analysis: Kruskal-Wallis.

Supplementary Figure 2



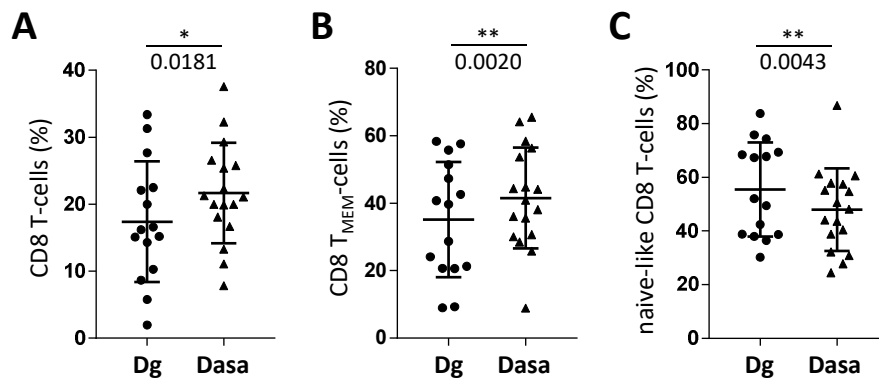
Supplementary Figure 2. In CML patients under dasatinib treatment, iNKT cells are increased. PBMCs isolated from patients (n=25) at CML diagnosis (Dg) or after 3 months of dasatinib treatment (Dasa) were analyzed by flow cytometry for iNKT cells. **(A)** Percentage of CD4⁺ CD8⁻ iNKT cells and PLZF MFI in this subset. **(B)** Percentage of DN (CD4⁻ CD8⁻) iNKT cells and PLZF MFI in this subset. Statistical analysis: paired two-tailed Wilcoxon test.

Supplementary Figure 3



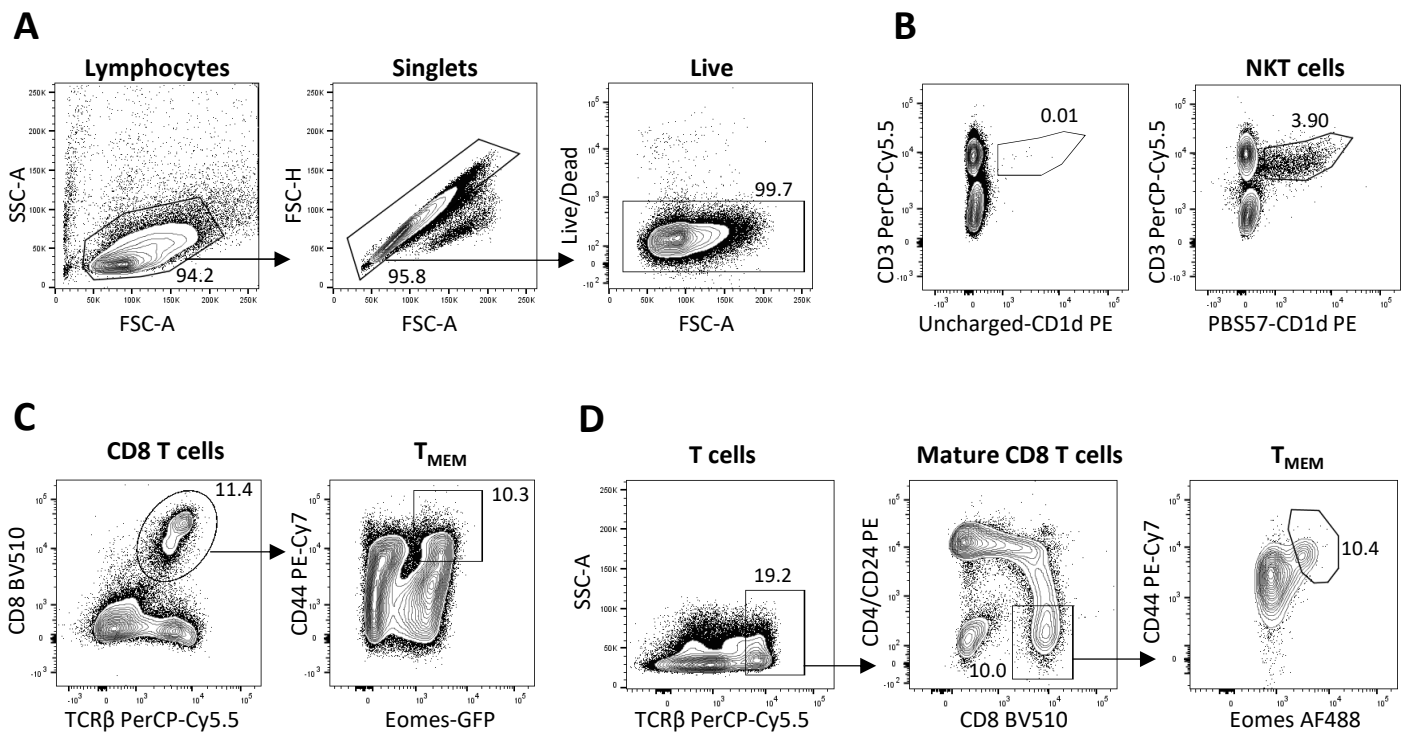
Supplementary Figure 3. Dasatinib does not affect the total CD8 T-cells compartment *in vivo*. Flow cytometry analysis of thymic cells from BALB/c WT mice orally gaved with dasatinib (Dasa, n=12) or its excipient (CTR, n=8) for 8 weeks. Analysis of TCR β ⁺ CD8⁺ T-cells among live mature lymphocytes (CD4⁻ CD24⁻). Statistical analysis: two-tailed Mann-Whitney test.

Supplementary Figure 5



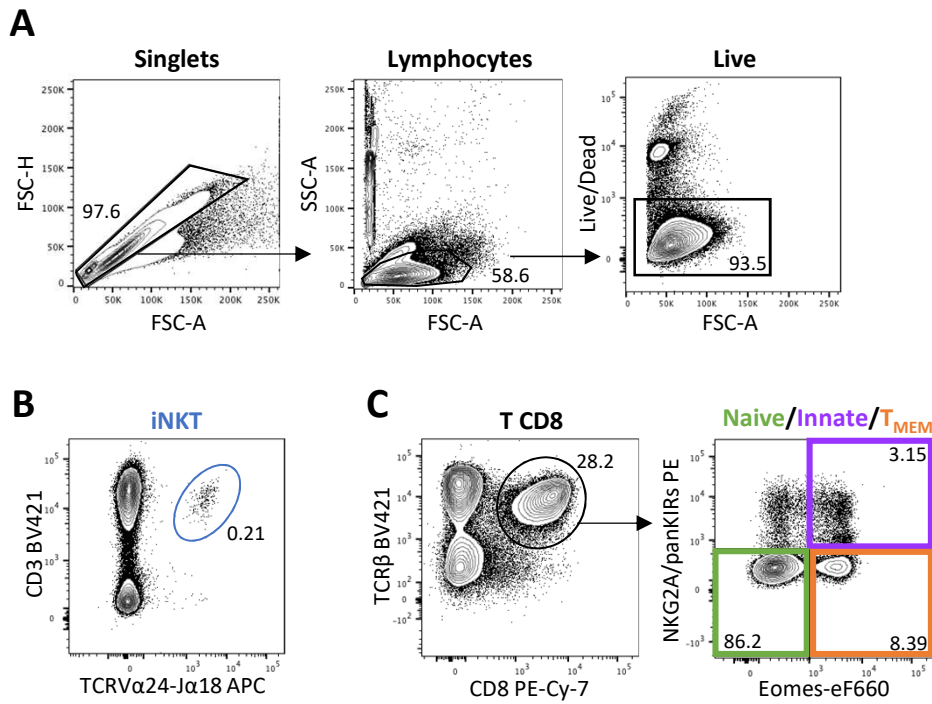
Supplementary Figure 5. CD8 T-cells subtypes repartition is modified in CML patients under dasatinib treatment. PBMCs isolated from patients (n=15) at CML diagnosis (Dg) or after 3 months of dasatinib treatment (Dasa) were analyzed by flow cytometry for CD8 T-cells (A) CD8 T_{MEM} cells (B) and naive-like CD8 T-cells. Statistical analysis: paired two-tailed Wilcoxon test.

Supplementary Figure 6



Supplementary Figure 6. Mouse gating trees for iNKT and T_{MEM} cells, related to Figures 1, 2 and 3. (A) Common gating tree to define live lymphocytes in spleen and thymus. (B) iNKT cells are defined as CD3⁺ PBS57-CD1d⁺ in spleen and thymus, uncharged CD1d tetramers were used as controls. (C and D) Gating tree for TCR β ⁺ CD8⁺ T_{MEM} cells populations in spleen (C) and thymus (D) preparations from BALB/c WT or BALB/c Eomes-GFP mice, respectively, based on CD44 and Eomes expression. Representative plots from a 10 weeks-old mice are shown.

Supplementary Figure 7



Supplementary Figure 7. Gating tree for iNKT, naive-like, T_{MEM} and innate CD8 T cells, related to Figure 4. Human gating tree to define live iNKT cells (CD3⁺ TCRVα24-Jα18⁺) and live TCRβ⁺ CD8⁺ naive-like, T_{MEM} and innate CD8 T cells populations in PBMC-derived preparations based on panKIR/NKG2A and Eomes expression. Representative plots from a 43 years-old human are shown.