

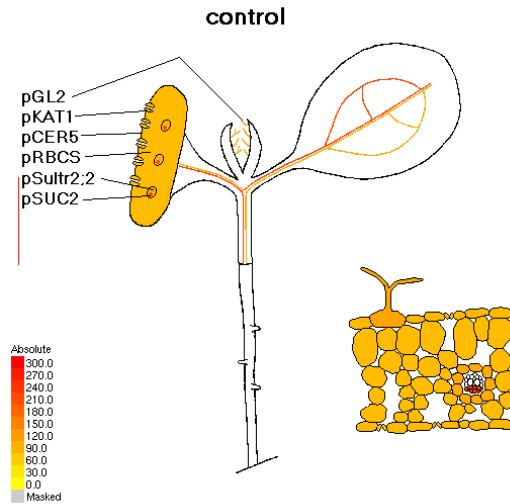
## Supplementary Information

The epidermis coordinates thermoresponsive growth through the phyB-PIF4-auxin pathway

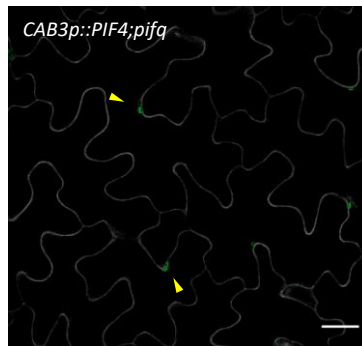
Kim et al.

**a**

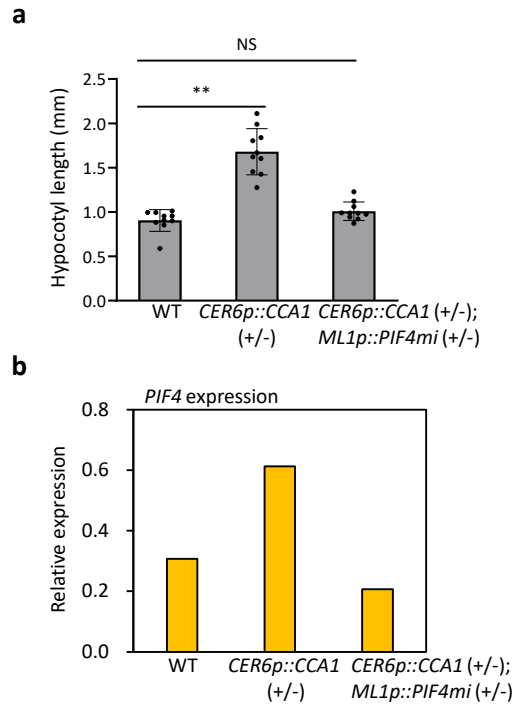
At2g43010 265248\_at *PIF4*



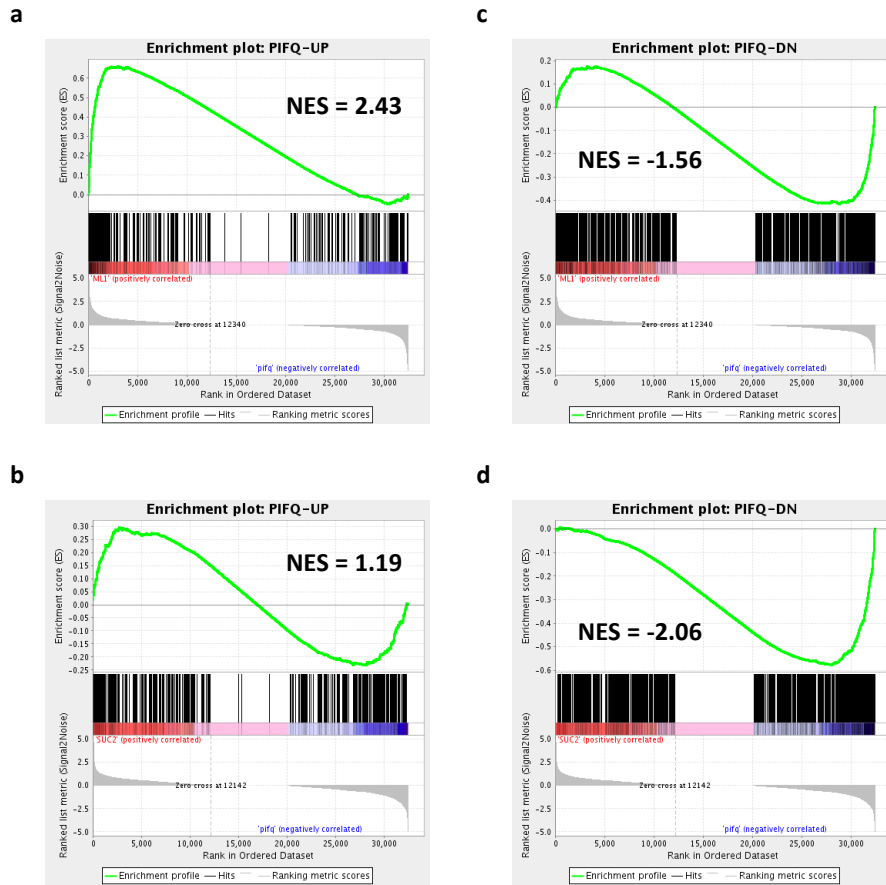
**b**



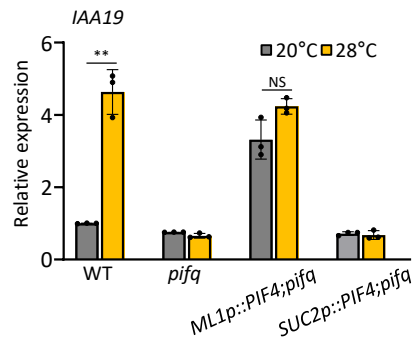
**Supplementary Fig. 1 Tissue-specific expression of *PIF4*.** **a**, *PIF4* expression in *Arabidopsis* according to the Cell-Type Specific Arabidopsis eFP browser (<http://efp.ucr.edu/>). **b**, A confocal microscopic image showing *PIF4*-YFP expression in the epidermis in *CAB3p::PIF4-YFP;pifq* transgenic plants. PI (gray) was used to counterstain the cell walls. Scale bars = 20  $\mu$ m.



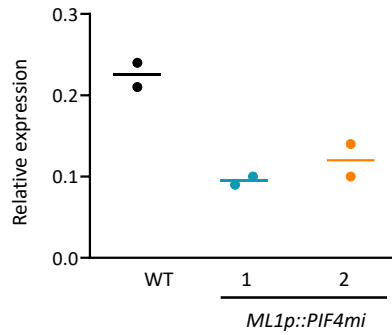
**Supplementary Fig. 2 | The long hypocotyls of *CER6p::CCA1* plants were suppressed by down-regulation of *PIF4*.** **a**, Hypocotyl lengths of seedlings grown under continuous white light at 20 °C for 7 days. Error bars indicate s.d. ( $n = 10$  plants). \*\*  $P < 0.01$  (two-tailed Student's  $t$ -test); NS, not significant (Student's  $t$ -test  $P \geq 0.05$ ). **b**, *PIF4* expression levels in the seedlings grown in the same growth condition as (a). The expression levels of *PIF4* were normalized to those of *PP2A*.



**Supplementary Fig. 3 | Gene set enrichment analysis (GSEA) of global PIF-regulated genes by tissue-specific PIF4.** The global PIF quartet-activated genes (PIFQ-UP) and -repressed genes (PIFQ-DN) were determined as differentially expressed genes in RNA-seq comparison of the wild type to *pifq* (1.5-fold, adjusted *P*-value < 0.05) (Fig. 2e, f). The enrichment of PIFQ-UP and PIFQ-DN gene sets were examined by GSEA in RNA-seq comparison of *MLI1p::PIF4;pifq* to *pifq* (a, c) or *SUC2p::PIF4;pifq* to *pifq* (b, d), respectively. NES indicates a normalized enrichment score.



**Supplementary Fig. 4 | qRT-PCR analyses of *IAA19*.** WT and transgenic seedlings were grown in 12 h light/12 h dark cycles (12L:12D) at 20 °C for 4 days and transferred under the continuous white light on the 5th day. The growth temperature was increased to 28 °C or kept at 20 °C for 4 h at ZT20-24 before harvesting for total RNA extraction. Gene expression levels were normalized to *APX3* and presented as values relative to that of wild-type (WT) kept at 20 °C. Error bars indicate s.d. ( $n=3$ ). \*\* $P < 0.01$  (two-tailed Student's  $t$ -test). NS, not significant (two-tailed Student's  $t$ -test  $P \geq 0.05$ )

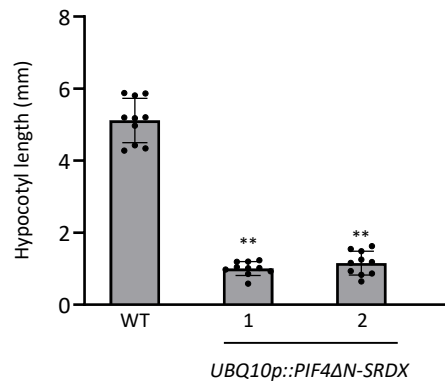


**Supplementary Fig. 5 | *PIF4* mRNA expression levels in the wild type and *ML1p::PIF4mi* seedlings.** Seedlings were grown under continuous white light at 20 °C for 5 days and harvested for RNA extraction. The gene expression levels were normalized to *PP2A*. Lines indicate median values ( $n = 2$ ).

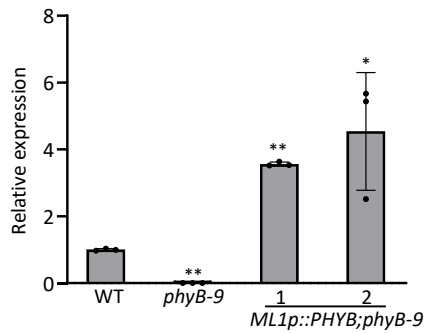
**a**



**b**

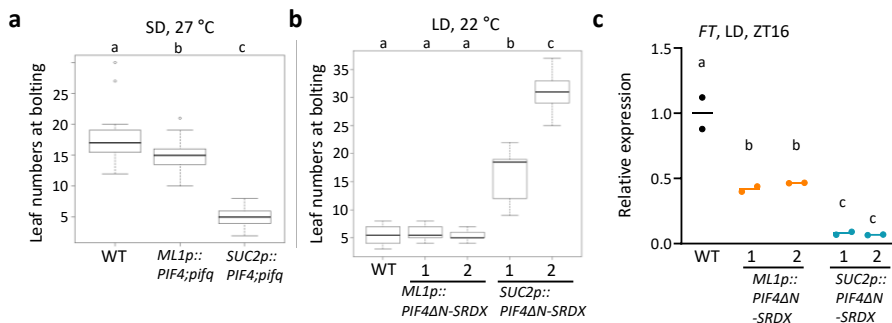


**Supplementary Fig. 6 | PIF4ΔN-SRDX inhibits hypocotyl growth at high temperatures.** Hypocotyl lengths of seedlings grown under continuous white light at 20 °C for 4 days followed by 28 °C for 3 days. Representative seedlings are shown in (a). Error bars in (b) indicate s.d. ( $n = 10$  plants).  $**P < 0.01$  (two-tailed Student's  $t$ -test).

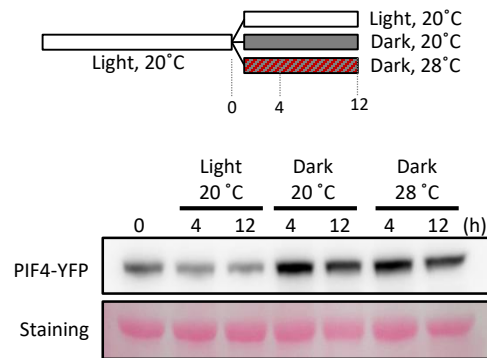


**Supplementary Fig. 7 | The expression levels of *PHYB* in WT, *phyB-9*, and *ML1p::PHYB;phyB-9* seedlings.** Seedlings were grown under continuous white light at 20 °C for 5 days and harvested for RNA extraction. The levels of *PHYB* were determined by qRT-PCR using the specific primers that amplify the wild-type *PHYB* cDNA, but not mutant *PHYB* cDNA in *phyB-9* (Supplementary Table 1). The gene expression levels were normalized to *APX3*. Error bars indicate s.d. ( $n = 3$ ). \*\* $P < 0.01$ , \* $P < 0.05$  (two-tailed Student's  $t$ -test).

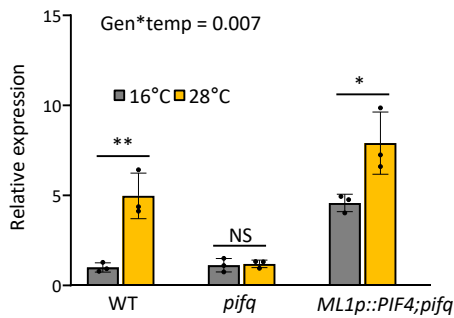




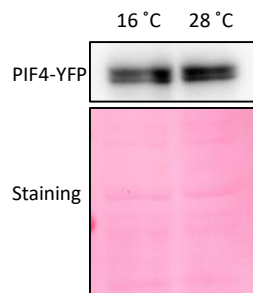
**Supplementary Fig. 8 | Vascular PIF4 plays a dominant role in the regulation of flowering. a,** Flowering time of WT, *ML1p::PIF4;pifq*, and *SUC2p::PIF4;pifq* plants grown under short days at 27 °C. Flowering time was scored as the number of rosette leaves at bolting. Letters above each box indicate significant differences based on a one-way ANOVA and Tukey's test ( $P < 0.05$ ). In the box plots (**a**, **b**), the thick lines indicate median values, the lower and upper ends of the boxes represent the 25<sup>th</sup> and 75<sup>th</sup> percentiles, and the ends of the whiskers are set at 1.5 times the interquartile range. **b,** Flowering time of WT, *ML1p::PIF4ΔN-SRDX*, and *SUC2p::PIF4ΔN-SRDX* plants grown under long days at 22 °C. Flowering time was scored as the number of rosette leaves at bolting. Letters indicate significant differences based on a one-way ANOVA and Tukey's test ( $P < 0.05$ ). **c,** *FT* mRNA expression of seedlings grown under long days at 22 °C. Seedlings were harvested at ZT16 for total RNA extraction. The *FT* expression levels were normalized to *APX3* and presented as values relative to that of WT. Lines indicate median values ( $n = 2$ ). Letters above each bar indicate significant differences based on a one-way ANOVA and Tukey's test ( $P < 0.05$ ).



**Supplementary Fig. 9 | Darkness increases the levels of epidermal PIF4 in *ML1p::PIF4;pifq*.** The *ML1p::PIF4;pifq* seedlings grown under white light at 20 °C were kept under white (20 °C) or transferred to darkness (20 or 28 °C). The seedlings were harvested after 4 or 12 h for total protein extraction. Immunoblotting was probed using an anti-GFP antibody. Ponceau S staining is shown for equal loading.

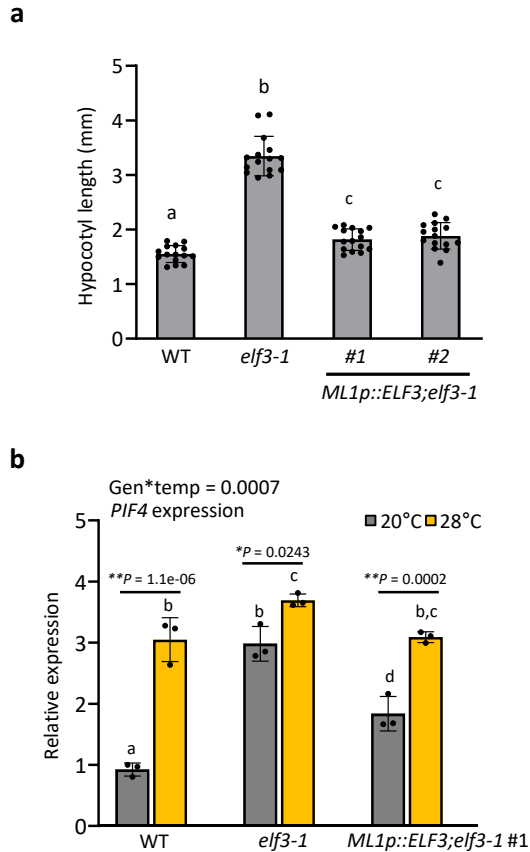


**Supplementary Fig. 10 | qRT-PCR analyses of *YUC8*.** WT, *pifq* and *ML1p::PIF4;pifq* seedlings were grown in 12 h light/12 h dark cycles (12L:12D) at 16 °C for 6 days and transferred under the continuous white light on the 7th day. The growth temperature was increased to 28 °C or kept at 16 °C for 4 h at ZT20-24 before harvesting for total RNA extraction. Gene expression levels were normalized to *APX3* and presented as values relative to that of wild-type (WT) kept at 20 °C. Error bars indicate s.d. ( $n = 3$ ). The  $P$ -value for the interaction term (genotype x temperature) calculated by two-way ANOVA is shown at the top.  $**P < 0.01$ ,  $*P < 0.05$  (two-tailed Student's  $t$ -test). NS, not significant (two-tailed Student's  $t$ -test  $P \geq 0.05$ ).



**Supplementary Fig. 11 | High-temperature effects on the stability of epidermal PIF4 protein.**

*ML1p::PIF4-YFP;pifq* seedlings were grown under white light constitutively at 16 °C or grown at 16 °C and shifted to 28 °C for 24 h. Immunoblotting was probed using an anti-GFP antibody. Ponceau S staining is shown for equal loading.



**Supplementary Fig. 12 | Epidermal ELF3 represses both hypocotyl growth and the expression of *PIF4*.** **a**, Hypocotyl lengths of seedlings grown under the continuous white light at 20 °C for 7 days. Error bars indicate s.d. ( $n = 15$  plants). Different letters above the bars indicate significant differences based on one-way ANOVA and Tukey's test ( $P < 0.05$ ). **b**, qRT-PCR analyses of *PIF4* mRNA expression. Seedlings were grown in 12 h light/12 h dark cycles (12L:12D) at 20 °C for 4 days and transferred under the continuous white light on the 5th day. The growth temperature was increased to 28 °C or kept at 20 °C for 4 h at ZT20-24 before harvesting for total RNA extraction. Gene expression levels were normalized to *APX3*. Error bars indicate s.d. ( $n = 3$ ). The  $P$ -value for the interaction term (genotype x temperature) calculated by two-way ANOVA is shown at the top. Letters above the bars indicate significant differences based on Tukey's test ( $P < 0.05$ ). Black asterisks indicate significant differences (\*\* $P < 0.01$  and \* $P < 0.05$ , two-tailed Student's  $t$ -test).

<b>qRT-PCR</b>		
<b>Gene</b>	<b>Forward</b>	<b>Reverse</b>
<i>PP2A</i>	TATCGGATGACGATTCTTCGTGCAG	GCTTGGTCGACTATCGGAATGAGAG
<i>APX3</i>	AGAGCACACTCATGGTGCCAAC	TGCTTAGCTTTCACGCCCTCAC
<i>PIF4</i>	GCCAAAACCCGGTACAAAACCA	CGCCGGTGAACATAATCTCAACATC
<i>YUC8</i>	AAACGCTCAAGGGTTCTCTTCG	CACGCACAACACCCTTTGATTTCG
<i>IAA19</i>	GGTGACAACGCGAATACGTTACCA	CCCAGTAGCATCCGATCTTTTCA
<i>IAA29</i>	AAACAGCGTTTGTTCCTTGAATG	TGGCCATCCAACAACCTTCGTAT
<i>PHYB</i>	GAAGAAGCTCGATGAGGCTTAGG	AACTGTAACCGAAAGCCTGCATC
<i>ELF3</i>	TTCATCTGGACCATCTAGTCAGC	GTTGCTTGGTTTGC GGCTGAAG
<b><i>ML1</i> and <i>SUC2</i> promoters</b>		
	<b>Forward</b>	<b>Reverse</b>
<i>ML1</i> promoter (CACC-BamHI)	CACCCAAGAACAAAACGATGCATAG	GGATCCTAACCGGTGATTACAGGAGTT
<i>SUC2</i> promoter (CACC-BamHI)	CACCAAAATCTGGTTTCATATTAATTTCA	GGATCCATTTGACAAAACCAAGAAAGTAAG
<b>ChIP-qPCR</b>		
<b>Gene</b>	<b>Forward</b>	<b>Reverse</b>
<i>PP2A</i>	CGGCTTTCATGATTCCCTCT	GCCTTAAGCTCCGTTTCCTACTT
<i>UBC30</i>	CAAATCCAAAACCTAGAAACCGAA	AACGACGAAGATCAAGAACTGGGAA
<i>rDNA</i>	CCCAAGTCAGACGAACGATT	TCTGACATGTGTGCGAGTCA
<i>YUC8</i>	TGGTCCACACAATTTTCACAG	GCAACGATGGTGATTGTTGAAG
<i>IAA19</i>	TAACCACCTTGTAAATGCCGGTC	GCAGAGACAGGTCAACTGAGGA
<i>ATHB2</i>	ACAACAAAAACAGAAGGGGTAGT	CCTTAAGCTAATCGACAAGTCAA

**Supplementary Table. 1 | Primer list for qRT-PCR, ChIP-qPCR, and vector construction**