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**Predictors of response and disease course in patients with inflammatory bowel disease treated with biological therapy
– The Danish IBD Biobank Project: Protocol for a multicenter prospective cohort study**

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Title

Predictors of response and disease course in patients with inflammatory bowel disease treated with biological therapy – The Danish IBD Biobank Project: Protocol for a multicenter prospective cohort study

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Abstract

Introduction

Inflammatory bowel diseases (IBD) are chronic diseases of unknown cause characterized by a progressive and unpredictable disease course. In the last decade, biological treatment has become a cornerstone in the treatment of IBD. However, one-in-three-to-four patients do not respond to first-line biologic agents and another third of patients see their response diminish over time. This highlights an unmet need for optimizing the use of biologicals and the prediction of treatment response. Considering the multifaceted nature of IBD, we hypothesize that multi-omics profiling of sequential samples from single patients could facilitate the discovery of predictive biomarkers of response to biological therapy and disease course.

Methods

This is a multicenter prospective cohort study which will enroll 840 biological-naïve IBD patients who initiate biological therapy in a three-year period. Primary outcomes are the occurrence of primary non-response (evaluated at week 14-16) and loss of response (evaluated during entire follow-up in patients who obtain partial or full response after induction period). Each patient will be followed-up for their clinical data for at least one year or till the end of study period (up to four years). Blood and stool samples will be collected sequentially during the first year of biological treatment. Intestinal tissue will be sampled after one year of treatment and whenever an endoscopy is performed. Samples will undergo transcriptomic, proteomic and microbial DNA analyses. Omics-data will be integrated with clinical data to identify a panel of predictive biomarkers of response to biological therapy and disease behaviour in IBD patients.

Ethics and dissemination

Ethical approval has been obtained from the Danish Ethics Committee (H-18064178). Inclusion is ongoing at three study centers and will be initiated in two additional centers. Both positive and negative study results will be disseminated through peer-reviewed journals according to STROBE guidelines, as well as presented at international conferences.

Strengths and limitations of this study

- The longitudinal design of the study and collection of sequential samples from single patients allow us to capture biomarker changes associated with critical events during biological treatment
- Integration of clinical data with data obtained from multiple types of biological material enables multi-omics analysis for addressing the multifaceted nature of inflammatory bowel diseases
- The long duration of follow-up also increases the likelihood that biomarker changes associated with degenerative changes in the intestines will be detected, which in turn could contribute to the discovery of novel molecular pathways and allow for therapeutic manipulation to halt disease progression
- Missing data are expected in some patients as all samples are collected in relation to routine visits and routine sampling, especially intestinal tissue samples at baseline, are expected to be missing in some patients
- Duration of follow-up will be limited (to one year only) in patients recruited during the last year of inclusion period

Introduction

The number of people affected by inflammatory bowel diseases (IBD) continues to increase globally, affecting up to 0.5 % of the population worldwide.^{1,2} IBD, comprising ulcerative colitis (UC), Crohn's disease (CD) and inflammatory bowel disease unclassified (IBDU) are complex, immune-mediated diseases characterized by chronic recurring inflammation in the gastrointestinal tract. Patients are often affected in their early adolescence and present with diarrhea, abdominal pain and cramps, perianal complications, as well as systemic symptoms such as fever, fatigue, joint pain and weight loss. The unpredictable and progressive disease course of IBD not only impairs the patients' quality of life, but also constitutes a socioeconomic burden.³ The annual cost of treatment is estimated to be 5.6 billion euros in Europe alone, which does not account for indirect costs related to sick leave and work disability.^{4,5}

During the last two decades, biologic agents have become a cornerstone in the treatment of severe or refractory cases of IBD to induce and maintain remission. Biologic agents are molecules targeting inflammatory mediators which have been shown to play a key role in the gut inflammation in IBD and include anti-tumor-necrosis-factor-alpha antibodies, anti-integrin-alpha4-beta7 antibodies, anti-alpha4-integrin antibodies, anti-interleukin12/23 antibodies and Janus-kinase inhibitors.⁶ Treatment with biologic agents has been shown to effectively decrease the risk of surgery and rate of hospitalization in IBD patients.⁷ However, 30-40% patients do not respond to biologics and an additional 30% of patients experience a diminished response over time.^{8,9} These patients often undergo several shifts in treatment and are exposed to excessive risk of adverse effects. Since IBD progresses over time, insufficient disease control might lead to irreversible degenerative changes in the intestine and require salvage surgery.¹⁰ We are currently unable to identify patients who will experience poor treatment response, hence we are unable to tailor biologic treatment to a given patient. Novel, but expensive, IBD treatment options are soon to be introduced and, as such, the need for measures to predict and optimize treatment outcome will only increase.^{11,12}

Recent studies of some of the more than 200 genes associated with IBD have shown how these genes lead in different ways to disruption of intestinal homeostasis and immunological tolerance with subsequent inflammation that is characteristic of IBD.^{13,14} Previous studies have primarily focused on linking genetic polymorphisms associated with IBD to the response to biological treatment, however, associations are vague and suggest that other omics profiles are also implicated.^{15,16} Recent findings indicate that mucosal inflammatory patterns and serum cytokine profiles differ between

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4 responders and non-responders to biological treatment.^{17–20} Furthermore, interactions between host
5 and gut microbiota play a pivotal role in IBD pathogenesis and should be taken into consideration
6 when predicting treatment response.^{21,22} Considering the complex nature of IBD, prediction of
7 treatment response is therefore likely to require the integration of multiple factors including genetic,
8 environmental, microbial and immunological factors into a multi-omics model, which on the other
9 hand may explain the pathobiology behind severe IBD phenotypes and intestinal damage.
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15 Here we present the study protocol for a prospective multicenter cohort study in patients with IBD
16 who are initiating biological treatment for the first time (biological-naïve patients). The Danish IBD
17 Biobank Project aims to identify a panel of predictive biomarkers associated with treatment response
18 and long-term outcomes to biological therapy in biological-naïve IBD patients.
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25 **Aims of the study**

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27 *Primary objectives* of this study are to

- 28 1) identify microbial, proteomic and transcriptomic predictors of treatment outcomes to
29 biological therapy in biological-naïve patients with IBD
- 30 2) identify microbial, proteomic and transcriptomic biomarkers of disease progression
31 and degenerative features of IBD.
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38 *Secondary objectives* are to

- 39 1) investigate treatment outcomes for biological treatment in biological-naïve IBD patients in a
40 real-life setting,
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- 43 2) evaluate adherence to national and international guidelines regarding initiation, follow-up and
44 optimization of biological therapy.
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50 **Methods**

51 Study Design

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53 This study is a multicenter, prospective cohort study which will investigate microbial, proteomic and
54 transcriptomic predictors of treatment outcomes to biological therapy in biological-naïve patients
55 with IBD. Patient enrolment was initiated in May 2019 and is currently ongoing at three study centers
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4 and will continue until May 2022. The duration of follow-up of each patient will be at least one year
5 from initiation of biological therapy or until May 2023. Clinical data and biological samples will be
6 collected at each study visit during the first year. Study visits are scheduled prior to initiation of
7 biological therapy and subsequently at routine visits for administration of biological therapy at the
8 outpatient clinic after 0, 2, and 6 weeks of treatment, and subsequently every second or third month.
9 After the first year, clinical data will be updated at least every six months until the end of follow-up
10 (Figure 1).
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16 17 18 19 Patient and public involvement

20 Patients were not involved in the process of refining the research question or the design of this study.
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26 Setting

27 The Danish IBD Biobank Project is a collaboration between the departments of gastroenterology at
28 four university hospitals located in three out of five geographic regions in Denmark. These include
29 the departments of gastroenterology at Hvidovre University Hospital, Herlev University Hospital,
30 Aarhus University Hospital and Aalborg University Hospital. Patients will be recruited from the
31 outpatient clinic or when they are admitted to hospital prior to initiation of biological therapy.
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41 Study population

42 Patients are eligible for inclusion if they are 1) diagnosed with IBD (UC, CD, IBDU) according to
43 the Copenhagen diagnostic criteria,²³ 2) aged eighteen or above, 3) starting treatment with biological
44 therapy due to IBD and have never received treatment with biological agents previously.
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49 Initiation of biological treatment is a clinical decision made by the patient's physician. Patients will
50 receive biological therapy according to the guidelines and recommendations from the Danish
51 Medicines Council, which include dosing and treatment intervals according to the drug labels.
52 According to these guidelines, patients with CD may receive: infliximab, adalimumab, vedolizumab,
53 certulizumab and ustekinumab. Patients with UC may receive: infliximab, adalimumab, vedolizumab,
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golimumab and tofacitinib. Furthermore, the study will include patients who initiate treatment with biological agents which might be approved in the future. There are no exclusion criteria in this study.

Outcome measures

Clinical response and primary non-response (PNR) to biological therapy will be evaluated after the end of the induction period, at week 14 for infliximab and vedolizumab, and week 12 for adalimumab, golimumab, certolizumab, ustekinumab and tofacitinib. Endoscopic remission will be evaluated 12 months after the start of treatment. In patients who continue biological treatment in a maintenance regime after initially achieving partial or full response, the proportion of patients with loss-of-response (LOR) will be registered after six and 12 months of biological treatment.

Clinical activity will be assessed using the Simple Clinical Colitis Activity Index (SCCAI)²⁴ for UC and IBDU patients and the Harvey-Bradshaw Index (HBI)²⁵ for CD patients. Endoscopic activity will be assessed using the Ulcerative Colitis Endoscopic Index of Severity (UCEIS)²⁶ in UC and the Simple Endoscopic Score for Crohn's Disease (SESCD)²⁷ in CD patients. Radiological activity will be assessed in CD patients who undergo imaging with MRI or abdominal CT using the Lemann Index, which indicates intestinal damage.²⁸ The Lemann Index will be evaluated at the end of follow-up by a gastroenterologist, in collaboration with a radiologist, at each study center.

Primary outcomes in this study are as follows:

1. Clinical response: defined as a decrease in SCCAI of ≥ 2 points from baseline in UC patients;²⁹ or a decrease in HBI of >3 points from baseline in CD patients.³⁰
2. Clinical remission to treatment: defined as a SCCAI of ≤ 2 in UC patients;²⁹ or a HBI of ≤ 4 in CD patients.³⁰
3. Endoscopic remission: defined as an UCEIS of ≤ 1 in UC patients;²⁶ or a SES-CD of <4 in CD patients.
4. PNR to treatment: defined as lack of improvement of symptoms and clinical signs with induction therapy, as well as patients who undergo intestinal resection or colectomy due to IBD during the period of induction therapy.
5. LOR to treatment: defined as patients achieving clinical response (as measured by clinical activity indices) during the period of induction therapy, but who later suffer from clinical

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4 relapse during maintenance therapy, including the need for rescue therapy with corticosteroids
5 or an alternative biological therapy, or surgery for IBD.
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8 6. Surgery: defined as intestinal resection or colectomy due to disease activity of IBD not
9 responding to medical therapy; or as fistula revision or drainage of abscesses after initiation
10 of biological therapy in patients with perianal CD.
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14 15 Clinical data

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17 At the time of inclusion, data on patient demographics (age, gender, ethnicity, education, height, body
18 weight), disease characteristics (disease duration, disease phenotype including disease subtype,
19 disease location, disease behavior and extra-intestinal disease manifestations), medical history,
20 history of surgery, family history of IBD, past and current medications, as well as dietary preferences,
21 will be gathered from the patient's medical record and by the use of a food frequency questionnaire.
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24 During follow-up, information on clinical disease activity, disease phenotype, current medication,
25 surgery, hospital admission, and development of comorbidity will be updated at each study visit
26 during the first year and every six months thereafter. Furthermore, results from endoscopic procedures,
27 imaging procedures, as well as results of routine blood samples (C-reactive protein, leukocyte count,
28 albumin, hemoglobin and TDM measurements) and fecal-calprotectin will be registered to evaluate
29 mucosal healing and inflammatory burden at each of the aforementioned timepoints.
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40 Biological samples

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42 Biological samples of blood, stool and intestinal tissue will be collected prospectively during the first
43 year of follow-up. Blood and stool samples will be collected immediately prior to initiation of
44 biological therapy and subsequently at each visit for drug administration of biological therapy. Upon
45 each blood sampling, a 9mL EDTA tube and a 9mL serum tube will be collected to yield plasma and
46 buffy coat and serum, respectively. Furthermore, a 2.5mL PAX-gene Blood Tube will be collected
47 for later transcriptomics analysis. Stool samples will be collected using a fecal sample collection kit
48 preserved with 96% ethanol or using a rectal foam dry swab to be immediately stored at minus 80 °C
49 after sample collection.
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57 Intestinal tissue samples will be collected at each endoscopic procedure that the patient undergoes
58 (unrelated to study participation) during follow-up and at an extra endoscopy after one year of follow-
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4 up. According to national guidelines, we expect an endoscopy to be performed at baseline,
5 immediately before initiation of biological therapy, upon change of treatment, and at least once per
6 year while the patient receives biological treatment. Intestinal tissue samples will be collected from
7 predefined locations: in CD patients, biopsies will be taken from terminal ileum, ascending and
8 sigmoid colon; in UC patients, biopsies will be taken from the ascending and sigmoid colon, as well
9 as the rectum. Two samples will be collected from each location, one sample will be treated with
10 RNA-later and handled according to instructions from the product company, while the other sample
11 will be snap-frozen in liquid nitrogen. In addition, two samples will be collected from any additional
12 area of inflammation.
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20 All biological samples will be stored at minus 80 °C until they are analyzed (Figure 2).
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25 **Data management**

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27 Clinical data will be collected using an electronic Case Report Form (e-CRF) based on the electronic
28 data capture system, REDCap (Research Electronic Data Capture), a tool designed to support data
29 capture for research studies.³¹ Biological samples will be stored at minus 80 °C until analysis of the
30 samples in batch runs. All data will be stored confidentially. The study has been approved by the
31 Danish Data Protection Agency (VD-2019-230).
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39 **Analysis plan**

40 Sample size calculation

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42 Each center is expected to start biological therapy in 5-6 biological-naïve IBD patients per month,
43 resulting in a total of 840 patients across three years. At least 30% of these are expected to be either
44 PNRs or LORs, corresponding to 252 patients. Biological samples from 200 PNR and LOR patients
45 and 70 responders will undergo primary analysis. The sample size has been calculated to be able to
46 detect a 1.3-fold upregulation of relevant genes with a common standard deviation (sigma) of one
47 and a desired power of 80% to determine a statistically significant difference ($\alpha=0.05$, two-sided test).
48 The estimated sample size is 197 subjects in the PNR/LOR group and 66 subjects in the responder
49 group.
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Analysis of biological samples

Blood and tissue samples will undergo transcriptomic analysis. RNA quality will be determined by a Bioanalyzer and RNA integrity numbers (RIN) will be calculated. RNA and miRNA expression profiles will be determined using microarray and high-throughput parallel sequencing (Illumina) to provide a global gene expression pattern using bioinformatics-based computational methods. Key pathways will be extracted by in silico annotation analysis of the transcriptome data. PCR analysis, Western blotting, and immunohistochemistry will subsequently be used to confirm expression patterns of interest. Serum and plasma will undergo characterization of preselected serum and plasma proteins using inflammation assays.

Stool samples will undergo analysis for microbiota and purification of microbiome DNA will be performed as described by Yan et al.³² Samples will undergo 16S and 18S PCR (examining bacteria, fungi and parasites) and Illumina sequencing and annotation of DNA sequences to species level. Data will thereafter be run in in-house R-scripts, which will identify both quantitative and qualitative differences in microbiota between cohorts.

Statistical analysis

Statistical programming will be carried out using the software R or SPSS. Details of the statistical analyses will be provided in the statistical analysis plan (SAP), which will be finished before data collection is completed. Comparison of demographics between patient groups will be performed using Chi-square test for nominal variables and t-test or Mann-Whitney U for ordinal variables, according to data distribution. Logistic and Cox regression models will be performed to find potential correlations between baseline characteristics and treatment outcome. Univariate and multivariate hierarchical clustering and principal component analysis will be applied to identify a panel of biomarkers which differentiate between patient groups according to their treatment response. Receiver operator characteristic (ROC) analyses will be performed to evaluate the sensitivity and specificity of single biomarkers and the composite biomarker panel. Biomarkers which are correlated to treatment response will later be assessed in a validation cohort using logistic regression and adjusted for covariates such as age, gender, disease phenotype and concomitant treatment. Statisticians and bio-informaticians will be consulted for their statistical expertise.

Discussion and expected limitations

A major strength of this project is its longitudinal design. The building of a biobank with sequential samples from single patients at different timepoints during biological treatment allows us to observe biomarker changes associated with critical events, including loss of response to specific biologic agents and disease progression. The extensive collection of data, including clinical data, laboratory data, as well as multiple types of biological material, allows us to perform multi-omics analyses that will take into account the complex interplay between host genome and host immune responses on the one hand, and gut microbes and environmental exposures on the other. In this way we will seek to identify a panel of serological, fecal and mucosal biomarkers which might assist physicians in tailoring biologic treatment to the individual IBD patient. The long duration of follow-up also allows us to study biological patterns which are associated with disease progression in IBD and the development of degenerative changes in the intestines. We thereby hope to facilitate the discovery of molecular pathways which might allow for therapeutic manipulation to halt disease progression in IBD patients.

The study does have some limitations to address. All tissue samples will be collected as part of routine endoscopies, apart from one supplementary endoscopy scheduled at one year after initiation of biological treatment. Despite national guidelines recommending that an endoscopy take place prior to initiation of biological treatment, tissue samples at baseline will inevitably be missing in some patients, since an endoscopy prior to treatment initiation will not be performed in all patients in clinical practice. Similarly, blood and stool samples are collected as part of routine sampling at regular visits, and this might challenge data comprehensiveness in some patients, for instance those lost during follow-up. All study centers also engage in randomized trials with experimental biologic agents in patients with IBD, and so we expect some patients to be excluded due to participation in alternative trials; however, this number is expected to be small.

Ethical considerations

The protocol has been approved by the Danish Ethics Committee (H-18064178) and the Danish Data Protection Agency (VD-2019-230). Patients will participate on a voluntary basis and can withdraw from the study at any time. The treatment of patients participating in the study does not differ from that of non-participating patients. All blood samples are to be collected in relation to routine sampling

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4 and tissue samples collected in relation to routine endoscopies, other than the single extra endoscopy
5 performed after one year of biological treatment that is recommended by national guidelines. Thus,
6 the current study design ensures that study participation is associated with minimal exposure to risk
7 and discomfort and any remaining potential risks are outweighed by the benefits for future patients.
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14 **Dissemination of results**

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16 Study results will be published according to the STROBE guidelines. Both negative and positive
17 results from the study will be published. Results will be submitted to publication in international peer-
18 reviewed scientific journals and presented at scientific conferences. The national patient organization
19 for patients with inflammatory bowel diseases (the Danish Colitis and Crohn's Organization) will be
20 involved to help develop the dissemination strategy and to share study results with patients. Patients
21 who participate in the project will be informed by letter of the study results if they express interest in
22 this upon study entry. Furthermore, study results and publications will be made public on the project
23 website, www.ibdbiobank.com.
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33 **Funding statement**

34
35 This study has received support from Takeda A/S through an unrestricted grant and public funds
36 hosted by the Hvidovre University Hospital. We will use private and public funds to cover the cost
37 of sample analyses and materials for sample collection and storage.
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44 **Authors' contributions**

45
46 All authors participated in manuscript conception. M.Z. is responsible for drafting the manuscript.
47
48 J.B., F.B., J.S., L.L., A.D., C.H. and A.M.P. are responsible for critical revision of the manuscript.
49
50 All authors have approved the final version of the manuscript for publication.
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57 **Conflicts of interest**

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4 MZ has no conflicts to declare.
5

6 J.B. reports personal fees from AbbVie, Janssen-Cilag, Celgene, MSD, Pfizer and grants and personal
7 fees from Takeda, all of which were unrelated to this study.
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10 F.B. reports personal fee and grants from Ferring A/S, all of which were unrelated to this study.
11

12 A.M.P reports travel grants and fees from MSD and Pfizer, all of which were unrelated to this study.
13

14 L.L. has no conflicts to declare.
15

16 A.D. has no conflicts to declare.
17

18 C.H. reports speaker fee from MSD.
19

20 J.S. reports research grant from Takeda unrelated to this study, as well as assigned unpaid national
21 coordinator of clinical trials from AbbVie, Eli Lilly and Roche and unpaid investigator at studies
22 from Arena and Zealand Pharma.
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References

1. Kaplan GG, Ng SC. Understanding and Preventing the Global Increase of Inflammatory Bowel Disease. *Gastroenterology*. 2017;152(2):313-321.e2.
2. Molodecky NA, Soon IS, Rabi DM, et al. Increasing incidence and prevalence of the inflammatory bowel diseases with time, based on systematic review. *Gastroenterology*. 2012;142(1):46-54.e42.
3. Ananthakrishnan AN, Weber LR, Knox JF, et al. Permanent Work Disability in Crohn's Disease. *Am J Gastroenterol*. 2008;103(1):154-161.
4. Burisch J, Jess T, Martinato M, Lakatos PL. The burden of inflammatory bowel disease in Europe. *J Crohn's Colitis*. 2013;7:322-337.
5. Hoivik ML, Moum B, Solberg IC, Henriksen M, Cvancarova M, Bernklev T. Work disability in inflammatory bowel disease patients 10 years after disease onset: results from the IBSEN Study. *Gut*. 2013;62(3):368-375.
6. National Board for the Use of Expensive Hospital Medication (Rådet for Dyr Sygehusmedicin). Treatment guidelines for treatment of inflammatory bowel diseases with expensive hospital medicines.
7. Peyrin-Biroulet L, Oussalah A, Williet N, Pillot C, Bresler L, Bigard M-A. Impact of azathioprine and tumour necrosis factor antagonists on the need for surgery in newly diagnosed Crohn's disease. *Gut*. 2011;60(7):930-936.
8. Loftus E V., Colombel J-F, Feagan BG, et al. Long-term Efficacy of Vedolizumab for Ulcerative Colitis. *J Crohn's Colitis*. 2016;11(4):jjw177.
9. Hanauer SB, Feagan BG, Lichtenstein GR, et al. Maintenance infliximab for Crohn's disease: the ACCENT I randomised trial. *Lancet*. 2002;359(9317):1541-1549.
10. Cosnes J, Cattan S, Blain A, et al. Long-Term Evolution of Disease Behavior of Crohn's Disease. *Inflamm Bowel Dis*. 2002;8(4):244-250.
11. White JR, Phillips F, Monaghan T, et al. Review article: novel oral-targeted therapies in inflammatory bowel disease. *Aliment Pharmacol Ther*. 2018;47(12):1610-1622.
12. Nielsen OH, Seidelin JB, Ainsworth M, Coskun M. Will novel oral formulations change the

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2
3
4 management of inflammatory bowel disease? *Expert Opin Investig Drugs*. 2016;25(6):709-
5 718.
6
7
8
9 13. Bernard K, Agnès G, Xavier Ramnik J., Khor B, Gardet A, Xavier RJ. Genetics and
10 pathogenesis of inflammatory bowel disease. *Nature*. 2011;474(7351):307-317.
11
12 14. Zhao M, Burisch J. Impact of Genes and the Environment on the Pathogenesis and Disease
13 Course of Inflammatory Bowel Disease. *Dig Dis Sci*. 2019;64(7):1759-1769.
14
15 15. Bek S, Nielsen J V., Bojesen AB, et al. Systematic review: genetic biomarkers associated
16 with anti-TNF treatment response in inflammatory bowel diseases. *Aliment Pharmacol Ther*.
17 2016;44(6):554-567.
18
19 16. de Souza HSP, Fiocchi C. Immunopathogenesis of IBD: current state of the art. *Nat Rev*
20 *Gastroenterol Hepatol*. 2016;13(1):13-27.
21
22 17. Arijs I, Li K, Toedter G, et al. Mucosal gene signatures to predict response to infliximab in
23 patients with ulcerative colitis. *Gut*. 2009;58(12):1612-1619.
24
25 18. West NR, Hegazy AN, Owens BMJ, et al. Oncostatin M drives intestinal inflammation and
26 predicts response to tumor necrosis factor-neutralizing therapy in patients with inflammatory
27 bowel disease. *Nat Med*. 2017;23(5):579-589.
28
29 19. Soendergaard C, Seidelin JB, Steenholdt C, Nielsen OH. Putative biomarkers of vedolizumab
30 resistance and underlying inflammatory pathways involved in IBD. *BMJ Open*
31 *Gastroenterol*. 2018;5(1):e000208.
32
33 20. Li Y, Soendergaard C, Bergenheim FH, et al. COX-2-PGE2 Signaling Impairs Intestinal
34 Epithelial Regeneration and Associates with TNF Inhibitor Responsiveness in Ulcerative
35 Colitis. *EBioMedicine*. 2018;36:497-507.
36
37 21. Ananthkrishnan AN, Luo C, Yajnik V, et al. Gut Microbiome Function Predicts Response
38 to Anti-integrin Biologic Therapy in Inflammatory Bowel Diseases. *Cell Host Microbe*.
39 2017;21(5):603-610.e3.
40
41 22. McIlroy J, Ianiro G, Mukhopadhyaya I, Hansen R, Hold GL. Review article: the gut
42 microbiome in inflammatory bowel disease-avenues for microbial management. *Aliment*
43 *Pharmacol Ther*. 2018;47(1):26-42.
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- 3
- 4 23. Burisch J. Crohn's disease and ulcerative colitis. Occurrence, course and prognosis during
- 5 the first year of disease in a European population-based inception cohort. *Dan Med J*.
- 6 2014;61(1):B4778.
- 7
- 8
- 9
- 10 24. Walmsley RS, Ayres RCS, Pounder RE, Allan RN. A simple clinical colitis activity index.
- 11 *Gut*. 1998;43(1):29-32.
- 12
- 13
- 14 25. Harvey RF, Bradshaw JM. A simple index of Crohn's-disease activity. *Lancet (London,*
- 15 *England)*. 1980;1(8167):514.
- 16
- 17
- 18 26. Travis SPL, Schnell D, Krzeski P, et al. Reliability and Initial Validation of the Ulcerative
- 19 Colitis Endoscopic Index of Severity. *Gastroenterology*. 2013;145(5):987-995.
- 20
- 21
- 22 27. Daperno M, D'Haens G, Van Assche G, et al. Development and validation of a new,
- 23 simplified endoscopic activity score for Crohn's disease: the SES-CD. *Gastrointest Endosc*.
- 24 2004;60(4):505-512.
- 25
- 26
- 27
- 28 28. Pariente B, Mary J-Y, Danese S, et al. Development of the Lemann index to assess digestive
- 29 tract damage in patients with Crohn's disease. *Gastroenterology*. 2015;148(1):52-63.e3.
- 30
- 31
- 32 29. Higgins PDR. Patient defined dichotomous end points for remission and clinical
- 33 improvement in ulcerative colitis. *Gut*. 2005;54(6):782-788.
- 34
- 35
- 36 30. Vermeire S, Schreiber S, Sandborn WJ, Dubois C, Rutgeerts P. Correlation between the
- 37 Crohn's disease activity and Harvey-Bradshaw indices in assessing Crohn's disease severity.
- 38 *Clin Gastroenterol Hepatol*. 2010;8(4):357-363.
- 39
- 40
- 41
- 42 31. Harris PA, Taylor R, Thielke R, Payne J, Gonzalez N, Conde JG. Research electronic data
- 43 capture (REDCap)—A metadata-driven methodology and workflow process for providing
- 44 translational research informatics support. *J Biomed Inform*. 2009;42(2):377-381.
- 45
- 46
- 47
- 48 32. Yan M, Pamp SJ, Fukuyama J, et al. Nasal microenvironments and interspecific interactions
- 49 influence nasal microbiota complexity and *S. aureus* carriage. *Cell Host Microbe*.
- 50 2013;14(6):631-640.
- 51
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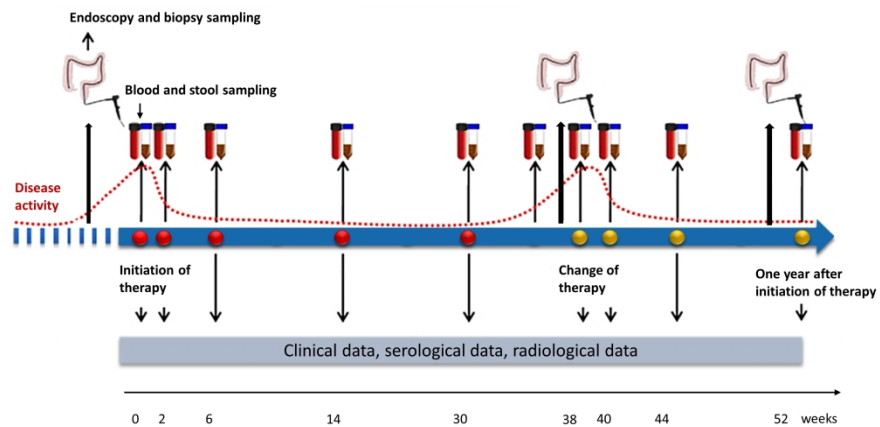


Figure 1. Study design of The Danish IBD Biobank Project.

338x190mm (300 x 300 DPI)

Sample	Medium/vial		Aliquoting	Storage
Blood	9mL EDTA tube	3500rpm at room temperature for 5 min.	3x EDTA plasma for protein analysis 1x EDTA buffy coat for DNA analysis	- 80°C
	9mL serum tube	Storage at room temperature for 30 min. 3500rpm at room temperature for 5 min.	3x serum for protein analysis	- 80°C
	2.5mL PAX-gene RNA Blood tube	Storage at room temperature for 2 hrs.	1x blood for RNA-analysis	- 20°C
	4mL EDTA tube	Direct storage.	4x EDTA whole-blood for back-up	- 80°C
Feces	Fecal tube with 96% ethanol	Direct storage.	1x fecal sample for DNA analysis	- 80°C
	Fecal Swab	Direct storage.	1x fecal sample for DNA analysis	- 80°C
Intestinal biopsy	RNA-later stabilization solution	Storage at room temperature for 24 hrs.	1x biopsy for RNA analysis	- 80°C
	Liquid nitrogen	Direct storage.	1x biopsy for back-up	- 80°C

Figure 2. Sample collection and storage in the Danishe IBD Biobank Project.

338x190mm (300 x 300 DPI)

BMJ Open

**Predictors of response and disease course in patients with inflammatory bowel disease treated with biological therapy
– The Danish IBD Biobank Project: Protocol for a multicenter prospective cohort study**

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4 **1 Title**

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7 2 Predictors of response and disease course in patients with inflammatory bowel disease treated with
8 biological therapy – The Danish IBD Biobank Project: Protocol for a multicenter prospective cohort
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24 **Abstract**

25 Introduction

26 Inflammatory bowel diseases (IBD) are chronic diseases of unknown cause characterized by a
27 progressive and unpredictable disease course. In the last decade, biological treatment has become a
28 cornerstone in the treatment of IBD. However, one-in-three-to-four patients do not respond to first-
29 line biologic agents and another third of patients see their response diminish over time. This highlights
30 an unmet need for optimizing the use of biologicals and the prediction of treatment response.
31 Considering the multifaceted nature of IBD, we hypothesize that multi-omics profiling of sequential
32 samples from single patients could facilitate the discovery of predictive biomarkers of response to
33 biological therapy and disease course.

35 Methods

36 This is a multicenter prospective cohort study which will enroll 840 biological-naïve IBD patients
37 who initiate biological therapy in a three-year period. Primary outcomes are the occurrence of primary
38 non-response (evaluated at week 14-16) and loss of response (evaluated during entire follow-up in
39 patients who obtain partial or full response after induction period). Each patient will be followed-up
40 for their clinical data for at least one year or till the end of study period (up to four years). Blood and
41 stool samples will be collected sequentially during the first year of biological treatment. Intestinal
42 tissue will be sampled after one year of treatment and whenever an endoscopy is performed. Samples
43 will undergo transcriptomic, proteomic and microbial DNA analyses. Omics-data will be integrated
44 with clinical data to identify a panel of predictive biomarkers of response to biological therapy and
45 disease behaviour in IBD patients.

47 Ethics and dissemination

48 Ethical approval has been obtained from the Danish Ethics Committee (H-18064178). Inclusion is
49 ongoing at three study centers and will be initiated in two additional centers. Both positive and
50 negative study results will be disseminated through peer-reviewed journals according to STROBE
51 guidelines, as well as presented at international conferences.

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4 53 **Strengths and limitations of this study**
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7 54 ▪ The longitudinal design of the study and collection of sequential samples from single patients
8
9 55 allow us to capture biomarker changes associated with critical events during biological
10 56 treatment
11
12 57 ▪ Integration of clinical data with data obtained from multiple types of biological material
13
14 58 enables multi-omics analysis for addressing the multifaceted nature of inflammatory bowel
15 59 diseases
16
17 60 ▪ The long duration of follow-up also increases the likelihood that biomarker changes
18
19 61 associated with degenerative changes in the intestines will be detected, which in turn could
20
21 62 contribute to the discovery of novel molecular pathways and allow for therapeutic
22 63 manipulation to halt disease progression
23
24 64 ▪ Missing data are expected in some patients as all samples are collected in relation to routine
25
26 65 visits and routine sampling, especially intestinal tissue samples at baseline, are expected to be
27 66 missing in some patients
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29 67 ▪ Patient recruitment is limited by the actual rate of initiation of biological therapy in the clinical
30
31 68 setting due to the observational design and the duration of follow-up will be limited (to one
32
33 69 year only) in patients recruited during the last year of inclusion period
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71 Introduction

72 The number of people affected by inflammatory bowel diseases (IBD) continues to increase globally,
73 affecting up to 0.5 % of the population worldwide.^{1,2} In Denmark alone, the prevalence of IBD in
74 Denmark is estimated to be 52.730 and the incidence of IBD has approached 25.9 per 100.000 person
75 years and is steadily increasing.³ IBD, comprising ulcerative colitis (UC), Crohn's disease (CD) and
76 inflammatory bowel disease unclassified (IBDU) are complex, immune-mediated diseases
77 characterized by chronic recurring inflammation in the gastrointestinal tract. Patients are often
78 affected in their early adolescence and present with diarrhea, abdominal pain and cramps, perianal
79 complications, as well as systemic symptoms such as fever, fatigue, joint pain and weight loss. The
80 unpredictable and progressive disease course of IBD not only impairs the patients' quality of life, but
81 also constitutes a socioeconomic burden.⁴ The annual cost of treatment is estimated to be 5.6 billion
82 euros in Europe alone, which does not account for indirect costs related to sick leave and work
83 disability.^{5,6}

84 During the last two decades, biologic agents have become a cornerstone in the treatment of severe or
85 refractory cases of IBD to induce and maintain remission. Biologic agents are molecules targeting
86 inflammatory mediators which have been shown to play a key role in the gut inflammation in IBD
87 and include anti-tumor-necrosis-factor-alpha antibodies, anti-integrin-alpha4-beta7 antibodies, anti-
88 alpha4-integrin antibodies, anti-interleukin12/23 antibodies and Janus-kinase inhibitors.⁷ Treatment
89 with biologic agents has been shown to effectively decrease the risk of surgery and rate of
90 hospitalization in IBD patients.⁸ In a nationwide cohort study in Denmark, the proportion of CD and
91 UC patients exposed to biological therapy was 28% and 9%, respectively, and the annual cost of
92 biologic therapy was estimated to constitute 1.9 million euros.⁹ However, 30-40% patients do not
93 respond to biologics and an additional 30% of patients experience a diminished response over
94 time.^{10,11} These patients often undergo several shifts in treatment and are exposed to excessive risk
95 of adverse effects. Since IBD progresses over time, insufficient disease control might lead to
96 irreversible degenerative changes in the intestine and require salvage surgery.¹² We are currently
97 unable to identify patients who will experience poor treatment response, hence we are unable to tailor
98 biologic treatment to a given patient. Novel, but expensive, IBD treatment options are soon to be
99 introduced and, as such, the need for measures to predict and optimize treatment outcome will only
100 increase.^{13,14}

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4 101 Recent studies of some of the more than 200 genes associated with IBD have shown how these genes
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6 102 lead in different ways to disruption of intestinal homeostasis and immunological tolerance with
7
8 103 subsequent inflammation that is characteristic of IBD.^{15,16} Previous studies have primarily focused
9
10 104 on linking genetic polymorphisms associated with IBD to the response to biological treatment,
11 105 however, associations are vague and suggest that other omics profiles are also implicated.^{17,18} Recent
12
13 106 findings indicate that mucosal inflammatory patterns and serum cytokine profiles differ between
14
15 107 responders and non-responders to biological treatment.^{19–22} Furthermore, interactions between host
16
17 108 and gut microbiota play a pivotal role in IBD pathogenesis and should be taken into consideration
18 109 when predicting treatment response.^{23,24} Considering the complex nature of IBD, prediction of
19
20 110 treatment response is therefore likely to require the integration of multiple factors including genetic,
21
22 111 environmental, microbial and immunological factors into a multi-omics model, which on the other
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24 112 hand may explain the pathobiology behind severe IBD phenotypes and intestinal damage.

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26 113 Here we present the study protocol for a prospective multicenter cohort study in patients with IBD
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28 114 who are initiating biological treatment for the first time (biological-naïve patients). The Danish IBD
29 115 Biobank Project aims to identify a panel of predictive biomarkers associated with treatment response
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31 116 and long-term outcomes to biological therapy in biological-naïve IBD patients.

32 33 117 34 35 36 118 **Aims of the study**

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38 119 *Primary objectives* of this study are to

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40 120 1) identify microbial, proteomic and transcriptomic predictors of treatment outcomes to
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42 121 biological therapy in biological-naïve patients with IBD
- 43
44 122 2) identify microbial, proteomic and transcriptomic biomarkers of disease progression
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46 123 and degenerative features of IBD.

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49 124 *Secondary objectives* are to

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51 125 1) investigate treatment outcomes for biological treatment in biological-naïve IBD patients in a
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53 126 real-life setting,
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55 127 2) evaluate adherence to national and international guidelines regarding initiation, follow-up and
56 128 optimization of biological therapy.

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130 **Methods**

131 Study Design

132 This study is a multicenter, prospective cohort study which will investigate microbial, proteomic and
133 transcriptomic predictors of treatment outcomes to biological therapy in biological-naive patients
134 with IBD. Patient enrolment was initiated in May 2019 and is currently ongoing at four study centers,
135 two additional study centers will initiate enrolment in medio 2020. Enrollment will continue until
136 May 2022. The duration of follow-up of each patient will be at least one year from initiation of
137 biological therapy or until May 2023. Clinical data and biological samples will be collected at each
138 study visit during the first year. Study visits are scheduled prior to initiation of biological therapy and
139 subsequently at routine visits for administration of biological therapy at the outpatient clinic after 0,
140 2, and 6 weeks of treatment, and subsequently every second or third month. After the first year,
141 clinical data will be updated at least every six months until the end of follow-up (Figure 1).

143 Patient and public involvement

144 Patients were not involved in the process of refining the research question or the design of this study.
145 In the future, the study aims at involving the Danish patient organization for IBD patients (Colitis and
146 Crohn's Association) in design of future studies which may arise from the current study.

148 Setting

149 The Danish IBD Biobank Project is a collaboration between the departments of gastroenterology at
150 six hospitals including five university hospitals located in four out of five geographic regions in
151 Denmark. These include the departments of gastroenterology at Hvidovre University Hospital, Herlev
152 University Hospital, Aarhus University Hospital, Aalborg University Hospital, Odense University
153 Hospital and the Hospital of Soenderjylland. Patients will be recruited from the outpatient clinic or
154 when they are admitted to hospital prior to initiation of biological therapy.

155 Apart from the included departments, the study aims to expand the collaboration with other Danish
156 hospitals in the future.

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4 158 Study population
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6 159 Patients are eligible for inclusion if they are 1) diagnosed with IBD (UC, CD, IBDU) according to
7
8 160 the Copenhagen diagnostic criteria,²⁵ 2) aged eighteen or above, 3) starting treatment with biological
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10 161 therapy due to IBD and have never received treatment with biological agents previously.

11
12 162 Initiation of biological treatment is a clinical decision made by the patient's physician. Patients will
13
14 163 receive biological therapy according to the guidelines and recommendations from the Danish
15
16 164 Medicines Council, which include dosing and treatment intervals according to the drug labels. All
17 165 participating hospitals are advised to follow the National Treatment Guidelines for biological
18
19 166 treatment of IBD patients issued by the Medicine Council,⁷ According to these guidelines, 80% of
20
21 167 CD patients initiating biological treatment due to luminal activity are expected to receive either 1)
22 168 infliximab 2) adalimumab 3) vedolizumab as 1st or 2nd line treatment, all three drugs and
23
24 169 ustekinumab may also be used as 3rd or 4th line treatment. These recommendations also apply to
25
26 170 fistulizing CD patients except for the use of vedolizumab which is only approved as 3rd or 4th line
27
28 171 treatment. In acutely severe UC, patients in need of 'rescue'-treatment with biologicals will receive
29
30 172 infliximab. In chronic active UC who will initiate biological therapy, 80% are expected to receive 1)
31 173 infliximab, 2) vedolizumab or 3) golimumab as 1st or 2nd line treatment, furthermore, tofacitinib
32
33 174 may be used as 2nd line treatment, all above-mentioned drugs and adalimumab may also be used as
34
35 175 3rd line treatment.⁷ Furthermore, the study will include patients who initiate treatment with biological
36 176 agents which might be approved in the future. There are no exclusion criteria in this study.
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41 178 Outcome measures
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43 179 Clinical response and primary non-response (PNR) to biological therapy will be evaluated after the
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45 180 end of the induction period, at week 14 for infliximab and vedolizumab, and week 12 for adalimumab,
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47 181 golimumab, certolizumab, ustekinumab and tofacitinib. Endoscopic remission will be evaluated 12
48
49 182 months after the start of treatment. In patients who continue biological treatment in a maintenance
50
51 183 regime after initially achieving partial or full response, the proportion of patients with loss-of-
52 184 response (LOR) will be registered after six and 12 months of biological treatment.
53

54 185 Clinical activity will be assessed using the Simple Clinical Colitis Activity Index (SCCAI)²⁶ for UC
55
56 186 and IBDU patients and the Harvey-Bradshaw Index (HBI)²⁷ for CD patients. Endoscopic activity will
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58 187 be assessed using the Ulcerative Colitis Endoscopic Index of Severity (UCEIS)²⁸ in UC and the
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Simple Endoscopic Score for Crohn's Disease (SESCD)²⁹ in CD patients. Radiological activity will be assessed in CD patients who undergo imaging with MRI or abdominal CT using the Lemann Index, which indicates intestinal damage.³⁰ The Lemann Index will be evaluated at the end of follow-up by a gastroenterologist, in collaboration with a radiologist, at each study center.

Primary outcomes in this study are as follows:

1. PNR to treatment: defined as lack of clinical response with induction therapy defined as a decrease in SCCAI of ≥ 2 points from baseline in UC patients;³¹ or a decrease in HBI of > 3 points from baseline in CD patients,³² as well as patients who undergo intestinal resection or colectomy due to IBD, or as fistula revision in patients with perianal CD during the period of induction therapy.
2. LOR to treatment: defined as patients achieving clinical response (as measured by clinical activity indices) during the period of induction therapy, but who later suffer from clinical relapse during maintenance therapy, including the need for rescue therapy with corticosteroids or an alternative biological therapy, or surgery for IBD.

Secondary outcomes in this study are as follows:

1. Clinical remission to treatment: defined as a SCCAI of ≤ 2 in UC patients;³¹ or a HBI of ≤ 4 in CD patients.³²
2. Endoscopic remission: defined as an UCEIS of ≤ 1 in UC patients;²⁸ or a SES-CD of < 4 in CD patients.
3. Surgery: defined as intestinal resection or colectomy due to disease activity of IBD not responding to medical therapy; or as fistula revision or drainage of abscesses after initiation of biological therapy in patients with perianal CD.

Clinical data

At the time of inclusion, data on patient demographics (age, gender, ethnicity, education, height, body weight), disease characteristics (disease duration, disease phenotype including disease subtype, disease location, disease behavior and extra-intestinal disease manifestations), medical history, history of surgery, family history of IBD, past and current medications, smoking status (current/former/never user; duration; amount), as well as dietary preferences, will be gathered from the patient's medical record and by the use of a food frequency questionnaire.

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4 218 During follow-up, information on clinical disease activity, disease phenotype, current medication,
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6 219 surgery, hospital admission, and development of comorbidity will be updated at each study visit
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8 220 during the first year and every six months thereafter. Furthermore, results from endoscopic procedures,
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10 221 imaging procedures, as well as results of routine blood samples (C-reactive protein, leukocyte count,
11 222 albumin, hemoglobin and TDM measurements) and fecal-calprotectin will be registered to evaluate
12
13 223 mucosal healing and inflammatory burden at each of the aforementioned timepoints.
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15 224 16 17 Biological samples

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20 226 Biological samples of blood, stool and intestinal tissue will be collected prospectively during the first
21
22 227 year of follow-up. Blood and stool samples will be collected immediately prior to initiation of
23
24 228 biological therapy and subsequently at each visit for drug administration of biological therapy. Upon
25
26 229 each blood sampling, a 9mL EDTA tube and a 9mL serum tube will be collected to yield plasma and
27 230 buffy coat and serum, respectively. Furthermore, a 2.5mL PAX-gene Blood Tube will be collected
28
29 231 for later transcriptomics analysis. Stool samples will be collected using a fecal sample collection kit
30
31 232 preserved with 96% ethanol or using a rectal foam dry swab to be immediately stored at minus 80 °C
32 233 after sample collection.
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34
35 234 Intestinal tissue samples will be collected at each endoscopic procedure that the patient undergoes
36 235 (unrelated to study participation) during follow-up and at an extra endoscopy after one year of follow-
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38 236 up. According to national guidelines, we expect an endoscopy to be performed at baseline,
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40 237 immediately before initiation of biological therapy, upon change of treatment, and at least once per
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42 238 year while the patient receives biological treatment. Intestinal tissue samples will be collected from
43 239 predefined locations: in CD patients, biopsies will be taken from terminal ileum, ascending and
44
45 240 sigmoid colon; in UC patients, biopsies will be taken from the ascending and sigmoid colon, as well
46
47 241 as the rectum. Two samples will be collected from each location, one sample will be treated with
48 242 RNA-later and handled according to instructions from the product company, while the other sample
49
50 243 will be snap-frozen in liquid nitrogen. In addition, two samples will be collected from any additional
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52 244 area of inflammation.
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54 245 All biological samples will be stored at minus 80 °C until they are analyzed (Figure 2).
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59 247 **Data management** 60

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Clinical data will be collected using an electronic Case Report Form (e-CRF) based on the electronic data capture system, REDCap (Research Electronic Data Capture), a tool designed to support data capture for research studies.³³ REDCap provides validated data entry and audit trails as well as anonymized data import and export. Appointed staff members at each study center have authorized access to study data, roles in the system are given according to functions and all access to the server and other server maintenances will be logged. Study setup and hosting is performed by a PhD student at Hvidovre University Hospital. Authorized staff members (one PhD student, one student research assistant, two physicians) can add data to the electronic database and will keep the database current to reflect subject status during the study period. Once the eCRF for a subject is completed, the project personnel at each local center will approve the data using an electronic signature and thereby confirm the accuracy of the data recorded. Biological samples will be stored at minus 80 °C until analysis of the samples in batch runs. All data will be stored confidentially in locked freezers located in locked rooms which are only accessible to authorized staff. The study has been approved by the Danish Data Protection Agency (VD-2019-230).

Upon study termination, electronic data will be stored for an additional ten years before deletion. Biological data will be transferred to a biobank for future research hosted at a centrally regulated biobank facility driven by the Capital Region of Denmark, this has been approved by the Data Regulatory Agency and consent from the participants will be sought upon recruitment. In order to maintain responsible data sharing and to keep patient data confidentially, data collected in the study will not be shared as an open access resource, however, researchers are welcome to apply for access to the biobank material for future projects by contacting the steering group of the project. In Denmark, collaboration with external research partners requires separate approval from the Data Regulation Agency and the establishment of a specific data processing agreement, therefore, data sharing with external partners will be decided on a case-by-case basis in the steering group.

Analysis plan

Sample size calculation

Each center is expected to start biological therapy in 5-6 biological-naïve IBD patients per month, resulting in a total of 840 patients across three years. At least 30% of these are expected to be either PNRs or LORs, corresponding to 252 patients. Biological samples from 200 PNR and LOR patients

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4 278 and 70 responders will undergo primary analysis. The sample size has been calculated to be able to
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6 279 detect a 1.3-fold upregulation of relevant genes with a common standard deviation (σ) of one
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8 280 and a desired power of 80% to determine a statistically significant difference ($\alpha=0.05$, two-sided test).
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10 281 The estimated sample size is 197 subjects in the PNR/LOR group and 66 subjects in the responder
11 282 group.
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14 283 15 16 284 Analysis of biological samples

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18 285 Blood and tissue samples will undergo transcriptomic analysis. RNA quality will be determined by a
19 286 Bioanalyzer and RNA integrity numbers (RIN) will be calculated. RNA and miRNA expression
20 287 profiles will be determined using microarray and high-throughput parallel sequencing (Illumina) to
21 288 provide a global gene expression pattern using bioinformatics-based computational methods. Key
22 289 pathways will be extracted by in silico annotation analysis of the transcriptome data. PCR analysis,
23 290 Western blotting, and immunohistochemistry will subsequently be used to confirm expression
24 291 patterns of interest. Serum and plasma will undergo characterization of preselected serum and plasma
25 292 proteins using inflammation assays.
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29 293 Stool samples will undergo analysis for microbiota and purification of microbiome DNA will be
30 294 performed as described by Yan et al.³⁴ Samples will undergo 16S and 18S PCR (examining bacteria,
31 295 fungi and parasites) and Illumina sequencing and annotation of DNA sequences to species level. Data
32 296 will thereafter be run in in-house R-scripts, which will identify both quantitative and qualitative
33 297 differences in microbiota between cohorts.
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42 298 43 44 299 Statistical analysis

45 300 Statistical programming will be carried out using the software R or SPSS. Details of the statistical
46 301 analyses will be provided in the statistical analysis plan (SAP), which will be finished before data
47 302 collection is completed. Comparison of demographics between patient groups will be performed
48 303 using Chi-square test for nominal variables and t-test or Mann-Whitney U for ordinal variables,
49 304 according to data distribution. Logistic and Cox regression models will be performed to find potential
50 305 correlations between baseline characteristics and treatment outcome. Univariate and multivariate
51 306 hierarchical clustering and principal component analysis will be applied to identify a panel of
52 307 biomarkers which differentiate between patient groups according to their treatment response.
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Receiver operator characteristic (ROC) analyses will be performed to evaluate the sensitivity and specificity of single biomarkers and the composite biomarker panel. Biomarkers which are correlated to treatment response will later be assessed in a validation cohort using logistic regression and adjusted for covariates such as age, gender, disease phenotype and concomitant treatment. Statisticians and bio-informaticians will be consulted for their statistical expertise.

Current status

The project has been initiated in May 2019 and is currently ongoing at three study centers. A total of 160 patients have been recruited over a period of seven months, the current recruitment rate is higher than estimated (105-126 patients). One study center has initiated recruitment in January 2020 and two other study centers are expected to initiate recruitment in medio 2020.

Discussion and expected limitations

A major strength of this project is its longitudinal design. The building of a biobank with sequential samples from single patients at different timepoints during biological treatment allows us to observe biomarker changes associated with critical events, including loss of response to specific biologic agents and disease progression. The extensive collection of data, including clinical data, laboratory data, as well as multiple types of biological material, allows us to perform multi-omics analyses that will take into account the complex interplay between host genome and host immune responses on the one hand, and gut microbes and environmental exposures on the other. In this way we will seek to identify a panel of serological, fecal and mucosal biomarkers which might assist physicians in tailoring biologic treatment to the individual IBD patient. The long duration of follow-up also allows us to study biological patterns which are associated with disease progression in IBD and the development of degenerative changes in the intestines. We thereby hope to facilitate the discovery of molecular pathways which might allow for therapeutic manipulation to halt disease progression in IBD patients.

The study does have some limitations to address. All tissue samples will be collected as part of routine endoscopies, apart from one supplementary endoscopy scheduled at one year after initiation of biological treatment. Despite national guidelines recommending that an endoscopy take place prior to initiation of biological treatment, tissue samples at baseline will inevitably be missing in some

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4 338 patients, since an endoscopy prior to treatment initiation will not be performed in all patients in
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6 339 clinical practice. Similarly, blood and stool samples are collected as part of routine sampling at regular
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8 340 visits, and this might challenge data comprehensiveness in some patients, for instance those lost
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10 341 during follow-up. All study centers also engage in randomized trials with experimental biologic
11 342 agents in patients with IBD, and so we expect some patients to be excluded due to participation in
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13 343 alternative trials; however, this number is expected to be small. At last, the enrolment rate will depend
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15 344 on the number of incident biological users initiating biological therapy in the clinical setting due to
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17 345 the observational study design. However, to date, the rate of enrolment has exceeded the anticipated
18 346 rate and the enrolment target is evaluated as feasible.
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22 23 348 **Ethical considerations**

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25 349 The protocol has been approved by the Danish Ethics Committee (H-18064178) and the Danish Data
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27 350 Protection Agency (VD-2019-230). Patients will participate on a voluntary basis and can withdraw
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29 351 from the study at any time. The treatment of patients participating in the study does not differ from
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31 352 that of non-participating patients. All blood samples are to be collected in relation to routine sampling
32 353 and tissue samples collected in relation to routine endoscopies, other than the single extra endoscopy
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34 354 performed after one year of biological treatment that is recommended by national guidelines. Thus,
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36 355 the current study design ensures that study participation is associated with minimal exposure to risk
37 356 and discomfort and any remaining potential risks are outweighed by the benefits for future patients.
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41 42 358 **Dissemination of results**

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44 359 Study results will be published according to the STROBE guidelines. Both negative and positive
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46 360 results from the study will be published. Results will be submitted to publication in international peer-
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48 361 reviewed scientific journals and presented at scientific conferences. The national patient organization
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50 362 for patients with inflammatory bowel diseases (the Danish Colitis and Crohn's Organization) will be
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52 363 involved to help develop the dissemination strategy and to share study results with patients. Patients
53 364 who participate in the project will be informed by letter of the study results if they express interest in
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55 365 this upon study entry. Furthermore, study results and publications will be made public on the project
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57 366 website, www.ibdbiobank.com.
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4 368 **Funding statement**

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7 369 This study has received support from Takeda Pharma A/S through an unrestricted grant and public
8
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11
12 372 A/S and Hvidovre University Hospital were not involved in the study design, collection, analysis,
13
14 373 interpretation of the data, writing of the report or the decision to submit the paper for publication.
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18 375 **Authors' contributions**

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21 376 All authors participated in manuscript conception. M.Z. is responsible for drafting the manuscript.
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24 377 J.B., F.B., J.S., L.L., A.D., C.H. and A.M.P. are responsible for critical revision of the manuscript.
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26 378 All authors have approved the final version of the manuscript for publication.
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31 380 **Conflicts of interest**

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34 381 MZ has no conflicts to declare.
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36 382 J.B. reports personal fees from AbbVie, Janssen-Cilag, Celgene, MSD, Pfizer and grants and personal
37 383 fees from Takeda, all of which were unrelated to this study.
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40 384 F.B. reports personal fee and grants from Ferring A/S, all of which were unrelated to this study.
41
42 385 A.M.P reports travel grants and fees from MSD and Pfizer, all of which were unrelated to this study.
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44 386 L.L. has no conflicts to declare.
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46 387 A.D. has no conflicts to declare.
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48 388 C.H. reports speaker fee from MSD.
49
50 389 J.S. reports research grant from Takeda unrelated to this study, as well as assigned unpaid national
51 390 coordinator of clinical trials from AbbVie, Eli Lilly and Roche and unpaid investigator at studies
52 391 from Arena and Zealand Pharma.
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References

1. Kaplan GG, Ng SC. Understanding and Preventing the Global Increase of Inflammatory Bowel Disease. *Gastroenterology*. 2017;152(2):313-321.e2. doi:10.1053/j.gastro.2016.10.020
2. Molodecky NA, Soon IS, Rabi DM, et al. Increasing incidence and prevalence of the inflammatory bowel diseases with time, based on systematic review. *Gastroenterology*. 2012;142(1):46-54.e42. doi:10.1053/j.gastro.2011.10.001
3. Lophaven SN, Lynge E, Burisch J. The incidence of inflammatory bowel disease in Denmark 1980-2013: a nationwide cohort study. *Aliment Pharmacol Ther*. 2017;45(7):961-972. doi:10.1111/apt.13971
4. Ananthakrishnan AN, Weber LR, Knox JF, et al. Permanent Work Disability in Crohn's Disease. *Am J Gastroenterol*. 2008;103(1):154-161. doi:10.1111/j.1572-0241.2007.01561.x
5. Burisch J, Jess T, Martinato M, Lakatos PL. The burden of inflammatory bowel disease in Europe. *J Crohn's Colitis*. 2013;7:322-337. doi:10.1016/j.crohns.2013.01.010
6. Hoivik ML, Moum B, Solberg IC, Henriksen M, Cvancarova M, Bernklev T. Work disability in inflammatory bowel disease patients 10 years after disease onset: results from the IBSEN Study. *Gut*. 2013;62(3):368-375. doi:10.1136/gutjnl-2012-302311
7. National Board for the Use of Expensive Hospital Medication (Rådet for Dyr Sygehusmedicin). Treatment guidelines for treatment of inflammatory bowel diseases with expensive hospital medicines. <https://rads.dk/media/4366/beh-gastro-31.pdf>. Published 2017. Accessed October 3, 2019.
8. Peyrin-Biroulet L, Oussalah A, Williet N, Pillot C, Bresler L, Bigard M-A. Impact of azathioprine and tumour necrosis factor antagonists on the need for surgery in newly diagnosed Crohn's disease. *Gut*. 2011;60(7):930-936. doi:10.1136/gut.2010.227884
9. Lo B, Vind I, Vester-Andersen MK, Bendtsen F, Burisch J. Direct and Indirect Costs of Inflammatory Bowel Disease: Ten Years of Follow Up in a Danish Population-Based Inception Cohort. *J Crohns Colitis*. May 2019. doi:10.1093/ecco-jcc/jjz096
10. Loftus E V., Colombel J-F, Feagan BG, et al. Long-term Efficacy of Vedolizumab for Ulcerative Colitis. *J Crohn's Colitis*. 2016;11(4):jjw177. doi:10.1093/ecco-jcc/jjw177
11. Hanauer SB, Feagan BG, Lichtenstein GR, et al. Maintenance infliximab for Crohn's disease: the ACCENT I randomised trial. *Lancet*. 2002;359(9317):1541-1549. doi:10.1016/S0140-6736(02)08512-4
12. Cosnes J, Cattan S, Blain A, et al. Long-Term Evolution of Disease Behavior of Crohn's Disease. *Inflamm Bowel Dis*. 2002;8(4):244-250. doi:10.1097/00054725-200207000-00002
13. White JR, Phillips F, Monaghan T, et al. Review article: novel oral-targeted therapies in inflammatory bowel disease. *Aliment Pharmacol Ther*. 2018;47(12):1610-1622. doi:10.1111/apt.14669
14. Nielsen OH, Seidelin JB, Ainsworth M, Coskun M. Will novel oral formulations change the management of inflammatory bowel disease? *Expert Opin Investig Drugs*. 2016;25(6):709-718. doi:10.1517/13543784.2016.1165204

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15. Bernard K, Agnès G, Xavier Ramnik J., Khor B, Gardet A, Xavier RJ. Genetics and pathogenesis of inflammatory bowel disease. *Nature*. 2011;474(7351):307-317. doi:10.1038/nature10209.Genetics
16. Zhao M, Burisch J. Impact of Genes and the Environment on the Pathogenesis and Disease Course of Inflammatory Bowel Disease. *Dig Dis Sci*. 2019;64(7):1759-1769. doi:10.1007/s10620-019-05648-w
17. Bek S, Nielsen J V., Bojesen AB, et al. Systematic review: genetic biomarkers associated with anti-TNF treatment response in inflammatory bowel diseases. *Aliment Pharmacol Ther*. 2016;44(6):554-567. doi:10.1111/apt.13736
18. de Souza HSP, Fiocchi C. Immunopathogenesis of IBD: current state of the art. *Nat Rev Gastroenterol Hepatol*. 2016;13(1):13-27. doi:10.1038/nrgastro.2015.186
19. Arijis I, Li K, Toedter G, et al. Mucosal gene signatures to predict response to infliximab in patients with ulcerative colitis. *Gut*. 2009;58(12):1612-1619. doi:10.1136/gut.2009.178665
20. West NR, Hegazy AN, Owens BMJ, et al. Oncostatin M drives intestinal inflammation and predicts response to tumor necrosis factor-neutralizing therapy in patients with inflammatory bowel disease. *Nat Med*. 2017;23(5):579-589. doi:10.1038/nm.4307
21. Soendergaard C, Seidelin JB, Steenholdt C, Nielsen OH. Putative biomarkers of vedolizumab resistance and underlying inflammatory pathways involved in IBD. *BMJ Open Gastroenterol*. 2018;5(1):e000208. doi:10.1136/bmjgast-2018-000208
22. Li Y, Soendergaard C, Bergenheim FH, et al. COX-2-PGE2 Signaling Impairs Intestinal Epithelial Regeneration and Associates with TNF Inhibitor Responsiveness in Ulcerative Colitis. *EBioMedicine*. 2018;36:497-507. doi:10.1016/j.ebiom.2018.08.040
23. Ananthakrishnan AN, Luo C, Yajnik V, et al. Gut Microbiome Function Predicts Response to Anti-integrin Biologic Therapy in Inflammatory Bowel Diseases. *Cell Host Microbe*. 2017;21(5):603-610.e3. doi:10.1016/j.chom.2017.04.010
24. McIlroy J, Ianiro G, Mukhopadhyaya I, Hansen R, Hold GL. Review article: the gut microbiome in inflammatory bowel disease-avenues for microbial management. *Aliment Pharmacol Ther*. 2018;47(1):26-42. doi:10.1111/apt.14384
25. Burisch J. Crohn's disease and ulcerative colitis. Occurrence, course and prognosis during the first year of disease in a European population-based inception cohort. *Dan Med J*. 2014;61(1):B4778.
26. Walmsley RS, Ayres RCS, Pounder RE, Allan RN. A simple clinical colitis activity index. *Gut*. 1998;43(1):29-32. doi:10.1136/gut.43.1.29
27. Harvey RF, Bradshaw JM. A simple index of Crohn's-disease activity. *Lancet (London, England)*. 1980;1(8167):514. doi:10.1016/s0140-6736(80)92767-1
28. Travis SPL, Schnell D, Krzeski P, et al. Reliability and Initial Validation of the Ulcerative Colitis Endoscopic Index of Severity. *Gastroenterology*. 2013;145(5):987-995. doi:10.1053/j.gastro.2013.07.024
29. Daperno M, D'Haens G, Van Assche G, et al. Development and validation of a new, simplified endoscopic activity score for Crohn's disease: the SES-CD. *Gastrointest Endosc*.

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474 2004;60(4):505-512. doi:10.1016/s0016-5107(04)01878-4

- 475 30. Pariente B, Mary J-Y, Danese S, et al. Development of the Lemann index to assess digestive
476 tract damage in patients with Crohn's disease. *Gastroenterology*. 2015;148(1):52-63.e3.
477 doi:10.1053/j.gastro.2014.09.015
- 478 31. Higgins PDR. Patient defined dichotomous end points for remission and clinical
479 improvement in ulcerative colitis. *Gut*. 2005;54(6):782-788. doi:10.1136/gut.2004.056358
- 480 32. Vermeire S, Schreiber S, Sandborn WJ, Dubois C, Rutgeerts P. Correlation between the
481 Crohn's disease activity and Harvey-Bradshaw indices in assessing Crohn's disease severity.
482 *Clin Gastroenterol Hepatol*. 2010;8(4):357-363. doi:10.1016/j.cgh.2010.01.001
- 483 33. Harris PA, Taylor R, Thielke R, Payne J, Gonzalez N, Conde JG. Research electronic data
484 capture (REDCap)—A metadata-driven methodology and workflow process for providing
485 translational research informatics support. *J Biomed Inform*. 2009;42(2):377-381.
486 doi:10.1016/j.jbi.2008.08.010
- 487 34. Yan M, Pamp SJ, Fukuyama J, et al. Nasal microenvironments and interspecific interactions
488 influence nasal microbiota complexity and *S. aureus* carriage. *Cell Host Microbe*.
489 2013;14(6):631-640. doi:10.1016/j.chom.2013.11.005

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492 **Figure legend**

493 Figure 1. Study design of the Danish IBD Biobank Project

494 Figure 2. Sample collection and storage in the Danish IBD Biobank Project

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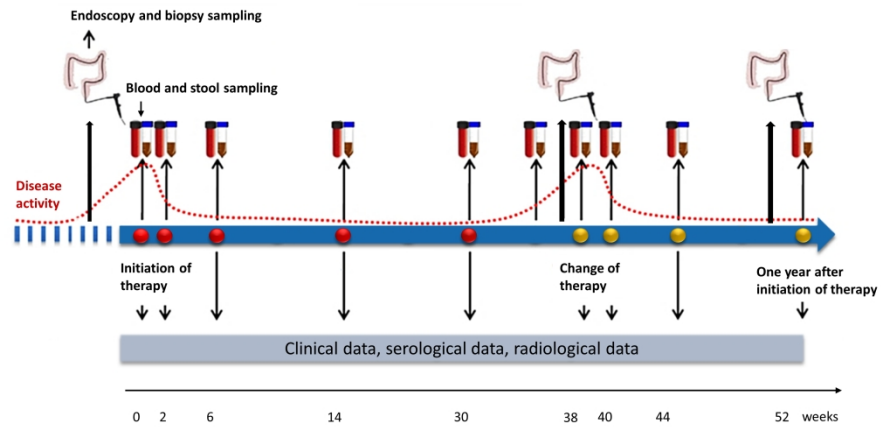


Figure 1. Study design of The Danish IBD Biobank Project.

338x190mm (300 x 300 DPI)

Sample	Medium/vial		Aliquoting	Storage
Blood	9mL EDTA tube	3500rpm at room temperature for 5 min.	3x EDTA plasma for protein analysis 1x EDTA buffy coat for DNA analysis	- 80°C
	9mL serum tube	Storage at room temperature for 30 min. 3500rpm at room temperature for 5 min.	3x serum for protein analysis	- 80°C
	2.5mL PAX-gene RNA Blood tube	Storage at room temperature for 2 hrs.	1x blood for RNA-analysis	- 20°C
	4mL EDTA tube	Direct storage.	4x EDTA whole-blood for back-up	- 80°C
Feces	Fecal tube with 96% ethanol	Direct storage.	1x fecal sample for DNA analysis	- 80°C
	Fecal Swab	Direct storage.	1x fecal sample for DNA analysis	- 80°C
Intestinal biopsy	RNA-later stabilization solution	Storage at room temperature for 24 hrs.	1x biopsy for RNA analysis	- 80°C
	Liquid nitrogen	Direct storage.	1x biopsy for back-up	- 80°C

Figure 2. Sample collection and storage in the Danishe IBD Biobank Project.

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