

Reporting Summary

Nature Research wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Research policies, see [Authors & Referees](#) and the [Editorial Policy Checklist](#).

Statistics

For all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.

n/a Confirmed

- The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement
- A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
- The statistical test(s) used AND whether they are one- or two-sided
Only common tests should be described solely by name; describe more complex techniques in the Methods section.
- A description of all covariates tested
- A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
- A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
- For null hypothesis testing, the test statistic (e.g. F , t , r) with confidence intervals, effect sizes, degrees of freedom and P value noted
Give P values as exact values whenever suitable.
- For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
- For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
- Estimates of effect sizes (e.g. Cohen's d , Pearson's r), indicating how they were calculated

Our web collection on [statistics for biologists](#) contains articles on many of the points above.

Software and code

Policy information about [availability of computer code](#)

Data collection

no software was used for collection of data

Data analysis

FlowJo version 10.6
FlowSOM version 1.14.1
pheatmap version 1.0.12
U-MAP version 0.2.0.0
FlowStats version 3.40.1
ggplot2 version 3.1.0
R version 3.5.2

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors/reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Research [guidelines for submitting code & software](#) for further information.

Data

Policy information about [availability of data](#)

All manuscripts must include a [data availability statement](#). This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A list of figures that have associated raw data
- A description of any restrictions on data availability

The source data underlying all figures and supplemental data are provided as a Source Data file

Field-specific reporting

Please select the one below that is the best fit for your research. If you are not sure, read the appropriate sections before making your selection.

Life sciences Behavioural & social sciences Ecological, evolutionary & environmental sciences

For a reference copy of the document with all sections, see [nature.com/documents/nr-reporting-summary-flat.pdf](https://www.nature.com/documents/nr-reporting-summary-flat.pdf)

Life sciences study design

All studies must disclose on these points even when the disclosure is negative.

Sample size	There were n=12 participants in each group (peanut allergic, peanut sensitized tolerant, healthy controls). Each sample underwent 3 in vitro stimulations. A total of 108 samples were analysed via mass cytometry.
Data exclusions	CD49b and LAG3 were excluded from manual and computational analysis due to limited expression across all samples. IL-10, IL-4 and IL-17A were excluded from manual and computational analysis due to limited expression across all samples.
Replication	findings cannot be replicated in these individuals as all cells were used for data collection
Randomization	samples were randomized prior to mass cytometry analysis and data collection
Blinding	investigators were blinded to food allergy outcome during data collection

Reporting for specific materials, systems and methods

We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, system or method listed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.

Materials & experimental systems

Methods

n/a	Involvement	Material/System
<input type="checkbox"/>	<input checked="" type="checkbox"/>	Antibodies
<input checked="" type="checkbox"/>	<input type="checkbox"/>	Eukaryotic cell lines
<input checked="" type="checkbox"/>	<input type="checkbox"/>	Palaeontology
<input checked="" type="checkbox"/>	<input type="checkbox"/>	Animals and other organisms
<input type="checkbox"/>	<input checked="" type="checkbox"/>	Human research participants
<input checked="" type="checkbox"/>	<input type="checkbox"/>	Clinical data

n/a	Involvement	Method
<input checked="" type="checkbox"/>	<input type="checkbox"/>	ChIP-seq
<input checked="" type="checkbox"/>	<input type="checkbox"/>	Flow cytometry
<input checked="" type="checkbox"/>	<input type="checkbox"/>	MRI-based neuroimaging

Antibodies

Antibodies used

Metal label Specificity Clone / Catalogue
 113In CD86 IT2.2 555662 BD
 142Nd CD19 HIB19, 3142001B, Fluidigm
 143Nd CD49b P1E6-C5, 359301 Biolegend
 144Nd IL-4 MP4-25D2, 3144010B Fluidigm
 145Nd CD4 RPA-T4, 3145001B Fluidigm
 146Nd CD8 RPA-T8, 3162015B Fluidigm
 147Sm CD20 2H7, 3147001B Fluidigm
 148Nd CD38 HB-7, BD
 149Sm CCR4 205410, 3149003A Fluidigm
 150Nd LAG3 874501, 3150016B Fluidigm
 151Eu CD123 6H6, 3151001B Fluidigm
 152Sm TNF Mab11, 3152002B Fluidigm
 153Eu CD45RA HI100, 3153001B Fluidigm
 154Sm CD3 UCHT1, 3154003B Fluidigm
 155Gd CD28 L283, 340975 BD,
 157Gd HLA-DR G46-6, 555810 BD
 158Gd CD33 WM53, 3158001B Fluidigm
 159Tb CD11c Bu15, 3159001B Fluidigm
 160Gd CD14 M5E2, 3160001B Fluidigm
 161Dy IFNg 4S.B4, 147319-85 eBioscience
 162Dy CD69 FN50, 3162001B Fluidigm
 163Dy CXCR3 G025H7, 3163004B Fluidigm
 164Dy IL-17 N49-853, 3164002B Fluidigm

165Ho CD127 A019D5, 3165008B Fluidigm
 166Er IL-2 MQ1-17h12, 3166002B Fluidigm
 167Er CD27 L128, 3167006B Fluidigm
 168Er CD40L 24-31, 3168006B Fluidigm
 169Tm CCR7 15053, MAB197 R&D Systems,
 171Yb IL-10 JES3-9D7, Biolegend
 173Yb CD25 M-A251, 555430 BD,
 176Yb CD56 NCAM16.2, 3176008B Fluidigm
 209Bi CD16 3G8, 3209002B Fluidigm

Validation

Each lot of conjugated antibody is quality control tested by CyTOF analysis of stained cells using the appropriate positive and negative cell staining and/or activation controls.

Human research participants

Policy information about [studies involving human research participants](#)

Population characteristics

Blood samples from one year old infants with peanut allergy, peanut sensitization and healthy controls were analyzed. 42% of healthy controls were male, 58% of peanut sensitization group were male and 67% of the peanut allergic group were male. Information on ancestry, family history of allergy, and other allergic conditions are specified in Table 1 of the manuscript.

Recruitment

Patients were recruited as per the HealthNuts Population Cohort study profile (Osborne et al., J. Allergy Clin Immunol 2011.)

Ethics oversight

Approval to conduct the HealthNuts study was obtained from the Victorian State Government Office for Children (reference no. CDF/07/492), the Victorian State Government Department of Human Services (reference no. 10/07), and the Royal Children's Hospital Human Research Ethics Committee (reference no. 27047).

Note that full information on the approval of the study protocol must also be provided in the manuscript.