Supplementary Table and Figures

Insight into DNA substrate specificity of PARP1 catalyzed DNA poly(ADP-ribosyl)ation

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| | 3' CGACACGGGAGTTGGCTTAAGTGTTCGGATCT |
|-----|--|
| S13 | 5' GCTGTGCCCTCAA AGCCTAGA HEG 3' CGACACGGGAGTTGGCTTAAGTGTTCGGATCT CGATCT |
| S14 | 5 ' GCTGTGCCCTCAACCGAA TCACAAGCCTAGA HEG 3 ' CGACACGGGAGTTGGCTTAAGTGTTCGGATCT D HEG Linker |
| S15 | 5' GCTGTGCCCTCAA CGAATTCACAAGCCTAGAGTTACATTAGCAGATACG 3' CGACACGGGAGTTGGCTTAAGTGTTCGGATCTCAATGTAATCGTCTATGC D HEG 4 |
| S16 | 5 ' GCTGTGCCCTCAACCGAAT CACAAGCCTAGAGTTACATTAGCAGATACG 3 ' CGACACGGGAGTTGGCTTAAGTGTTCGGATCTCAATGTAATCGTCTATGC \bigcirc HEG 4 |
| S17 | 5' GCTGTGCCCTCAACCGAATT CACAAGCCTAGAGTTACATTAGCAGATACG 3' CGACACGGGAGTTGGCTTAA-GTGTTCGGATCTCAATGTAATCGTCTATGC \bigcirc HEG 4 |
| S18 | 5 ' GCTGTGCCCTCAACCGAATTCAC AGCCTAGAGTTACATTAGCAGATACG 3 ' CGACACGGGAGTTGGCTTAAGTGTTCGGATCTCAATGTAATCGTCTATGC D HEG 4 |
| S19 | <pre> tt CGTGTGCCCTCAACCGAATTCACAAGCCTAGA HEG S' GGTGTGCCCTCAACCGAATTCACAAGCCTAGA HEG S' CGACACGGGAGTTGGCTTAAGTGTTCGGATCT Linker tt </pre> |
| S20 | 5' GCTGTGCCCTCAA CGAATTCACAAGCCTAGA 3' CGACACGGGAGTTGGCTTAAGTGTTCGGATCT HEG 4 |
| S21 | الالتى 5 ' TCATAGGCTTAGTCGTCATTCGCTGTGCCCTCAACCGAATTCACAAGCCTAGA HEG 3 ' CGACACGGGAGTTGGCTTAAGTGTTCGGATCT Linker |
| S22 | 5' GCTGTGCCCTCAACCGAATTCACAAGCCTAGA HEG 3' CTTACTGCTGATTCGGATACTCGACACGGGAGTTGGCTTAAGTGTTCGGATCT Linker |



Figure S1. ADP-ribosylation of S1ⁿ DNA duplexes containing 5'-otherhangs by PARP1 at a non-saturating concentration of NAD⁺. [³²P]labelled S1ⁿ DNA duplexes (20 nM) were combined with 50 nM PARP1 in the presence of 50 μ M NAD⁺ for 15 min at 37°C under standard reaction conditions. The data on PARP-catalysed formation of DNA ADP-ribosylation products are presented as mean ± SD from three independent experiments.



Figure S2. PARP1 DNA PARylation activity in presence of HPF1. (**A**) The 5'-[³²P]labelled S1⁷ DNA duplex (20 nM) was incubated with PARP1 (10 nM) and varying concentrations of HPF1 under standard reaction conditions. The products of the reaction were separated using denaturing PAGE. (**B**) Comparison of DNA PARylation activities of PARP1 (10 nM) toward S1ⁿ DNA substrates (20 nM) in presence of 100 nM HPF1 under standard reaction conditions. The purified human HPF1 protein was kindly provided by Dr Ivan Ahel (University of Oxford, U.K.). The error bars represent the standard deviation (n = 3).



Figure S3. Validation of PARP1-dependent PARylation of 3'-phosphorylated DSB termini of DNA substrates S2 and S10-13 by CIP-induced dephosphorylation. After incubation with PARP1, the 5'-[³²P]labelled DNA samples were heated for 10 min at 85°C, and the resulting [³²P]labelled DNA PARylation products were further incubated with 10 U of CIP for 30 min at 37°C.



Figure S4. ADP-ribosylation of S1ⁿ DNA duplexes containing 5'-otherhangs by PARP2 and PARP3. [32 P]labelled S1ⁿ DNA duplexes (20 nM) were combined with 50 nM PARP3 or PARP2 in the presence of 1 mM NAD⁺ in ADPR buffer (20 mM HEPES-KOH pH 7.6, 50 mM KCl, 5 mM MgCl₂, 1 mM DTT and 100 µg/mL BSA). The mixture was incubated for 10 min (PARP3) or 30 min (PARP2) at 37°C. The data on PARP-catalysed formation of DNA ADP-ribosylation products are presented as mean ± SD from three independent experiments.