Supplementary Information

Bacterial chemotaxis towards polysaccharide pectin by pectin-binding protein

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An excel file for Table S1 was separately uploaded.

Table S2.	Primers	used in	this	study.	
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Primer	Sequence (5'-3')
nKS F1	AAAAATGGCAGCGCTGGCAGTCCTT
pKS R2	GGCGGTGCTACAGAGTTCTTGAAGT
$r-3kbn \rightarrow 6kbn f$	TGATCGACCTGCAAGAGCGCATCCA
$r-3kbn \rightarrow 6kbn r$	TGTTCAAGCGCTCGACGCGAGTGCT
$f \rightarrow 3$ 2khn	GGGTCGTCGTCAAAGCGTCCTGTCA
$r_{-6kbn} \rightarrow 9kbn f$	
$r_{-6kbn} \rightarrow 9kbn r$	GCTCAGGTCAACATGAGCAACCTGC
$f_{-3} 2khn \rightarrow 6 2khn f$	TGCTCAAGCCACGTTTGGCCGCGTT
$f_{-3} 2kbp \rightarrow 6 2kbp r$	CACGACGCGCCCAATGTTCACCTCA
$r_{0}hh_{1$	GGTCGTAGCTGTCGTCTCAGTCATT
$r_{-9kbn} \rightarrow 12kbn r$	TTTCACGCAAGCGCCGATGCTGGAA
$f \in 2kbn \rightarrow 0.2kbn f$	
$f \in 2kbp \rightarrow 0.2kbp r$	
$1-0.2 \text{KOp} \rightarrow 9.2 \text{KOp} 1$	
$1 - 12 K 0 \mu \rightarrow 1$ f 0 21 km $\rightarrow 121$ km f	
$1-9.2 \text{KOp} \rightarrow 12 \text{KOp}$	
$1-9.2 \text{KOp} \rightarrow 12 \text{KOp}$	
$1-12K0p \rightarrow 1$	
F-1289838~	
1-3135268~	
sph2942 BamHI infusion f	
sph2942 BamHI infusion r	AACGIIGCCCGGAICCGACAAGGGCAGAIACACGI
sph1119 BamHI infusion f	
sph1119 BamHI infusion r	AACGTTGCCCGGATCGTCGATTACGCAACCAGCCG
sph1117 BamHI infusion f	TCGCGCGACCGGATCCCGTCGTACCACCGTTAACC
sph1117 BamHI infusion r	AACGTTGCCCGGATCGTATCGGAGAAGTTCGCCCG
sph1118 BamHI infusion f	TCGCGCGACCGGATCACGAACTTGCCGATCGACCA
sph1118 BamHI infusion r	AACGTTGCCCGGATCATTGGCGCCCATACCTCCAG
sph2733 BamHI infusion f	TCGCGCGACCGGATCTTCAAGCTGGGCTACCTCGC
sph2733 BamHI infusion r	AACGTTGCCCGGATCGGACGTACGGATGATGTCGA
sph1118 BamHI infusion f new	TCGCGCGACCGGATCTGATGATCACGGGGATCACC
sph1118 BamHI infusion r new	AACGTTGCCCGGATCTTAAAACGGCCGCGCAACCA
sph1117 f	AAGCGATACCACATCAGCGTCCACT
sph1117 r	AGAAGCGTATGGCGCTGTACGACCA
sph1118 f	TACGCTTCTTCATGCTTTCGTTCGG
sph1118 r	TCGTAGTTCATCCCTAGGAGGACAA
sph1119 f	ATGAGTCGACCCCTTCATTTTCTGC
sph1119 r	TCAGGCGTTTGAACTACCTGATGTC
sph1117 f inverse	AAGTCATCGGCACGGTTCGTCCGGC
sph1117 r inverse	TGAGTCTTGGCGGCGACGAACTGGT
sph1118 f inverse	GACGGTGTAATCGTCAACCTTCGAA
sph1118 r inverse	TGGCTATGGACCGTGACGAGATCAA
sph1119 f inverse	GGGCGACCTCCAGTGCCTCGGTATT
sph1119 r inverse	AATGACCGCGCACGCCATGCACGGA
SPH1118 NdeI(S)	GGCATATGGCTGCTTTCACGCAAGCGCCGA
SPH1118 NdeI(L)	GGCATATGATGAGGTTTTACTCTCGCAAGTTTGCG
Km-F	GGGGGCGCTGAGGTCTGCCTCGTGAAG
Km-R	GGGGGAAAGCCACGTTGTGTCTCAAAA
sph1118-NdeI in fusion	AAGGAGATATACATATGGCTGCTTTCACGCAAGCGCCGA
sph1118-XhoI in fusion	GGTGGTGGTGCTCGAGCTTGGAGAAGTACCACTGCT



Figure S1. Import and degradation of alginate polysaccharide by strain A1 cells. Extracellular alginate is incorporated to the periplasm through the cell-surface pit and bound to the periplasmic binding protein (AlgQ1 or AlgQ2). The binding protein delivers the polysaccharide to the ABC transporter (AlgM1-AlgM2/AlgS-AlgS), which imports alginate to the cytoplasm with an energy from ATP hydrolysis by AlgS-AlgS. Alginate is degraded to constituent monosaccharides by three endotype (A1-I, II, and III) and a exotype (A1-IV) alginate lyases.



Figure S2. Assimilation of pectin by strain A1 cells. Bacterial cells were cultured in test tubes (n = 3). Values indicate the average optical density at 600 nm (OD₆₀₀) as follows: triangle, nonmotile wild-type strain A1 cells; circle, strain A1-MP cells; rhombus, strain A1-M5 cells ; square, *sph1118*-complemented strain Δ *sph1118* cells derived from strain A1-MP; cross, strain Δ *sph1118* cells derived from strain A1-MP; plus, *sph1117-sph1118*-complemented A1-M5 cells. Error bars indicate standard deviations.



Figure S3. Purification of SPH1118. (**A**) Elution profile of the recombinant SPH1118 from gel filtration column chromatography. (**B**) SDS-PAGE profile of the recombinant SPH1118. Lane S, proteins before subjected to the gel filtration column chromatography; lanes 72-94, fractions cotaining the eluted protein from the chromatography. This electrophoretic profile is not an image cropped from different parts of the same gel or from different gels.