

SUPPLEMENTARY INFORMATION

Diatoms synthesize sterols by inclusion of animal and fungal genes in the plant pathway

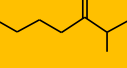
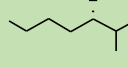
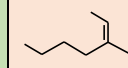
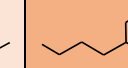

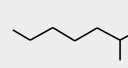
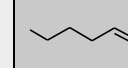
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Supplementary Table 1: Sterol composition of centric and pennate diatoms based on the 5-cholesten skeleton

Diatom species	Phytosterols					Animal sterols	
							
<i>Cyclotella cryptica</i> ^a							
<i>Skeletonema marinoi</i> ^a							
<i>Skeletonema costatum</i> ^b							
<i>Thalassiosira weissflogii</i> ^a							
<i>Pseudonitzschia arenysensis</i> ^a							
<i>Phaeodactylum tricornutum</i> ^a							
<i>Phaeodactylum tricornutum</i> ^b							
<i>Thalassiosira pseudonana</i> ^b							
<i>Cylindrotheca closterium</i> ^b							
<i>Fragilariopsis cylindrus</i> ^b							

^aCurrent study; ^bIn agreement with *Limnol. Oceanogr.*, 55(1), 2010, 91–105

Supplementary Table 2: List of main identified orthologs obtained using nodal sterol pathway translated sequences from *S. marinoi* and *C. cryptica* compared vs *T. oceanica*, *T. pseudonana*, *F. cylindrus*, *P. tricornutum*, *PN. multistriata* and *C. crispus* genomes. In addition, the sequence of the ERG2 from *Saccharomyces cerevisiae* was used as further query.

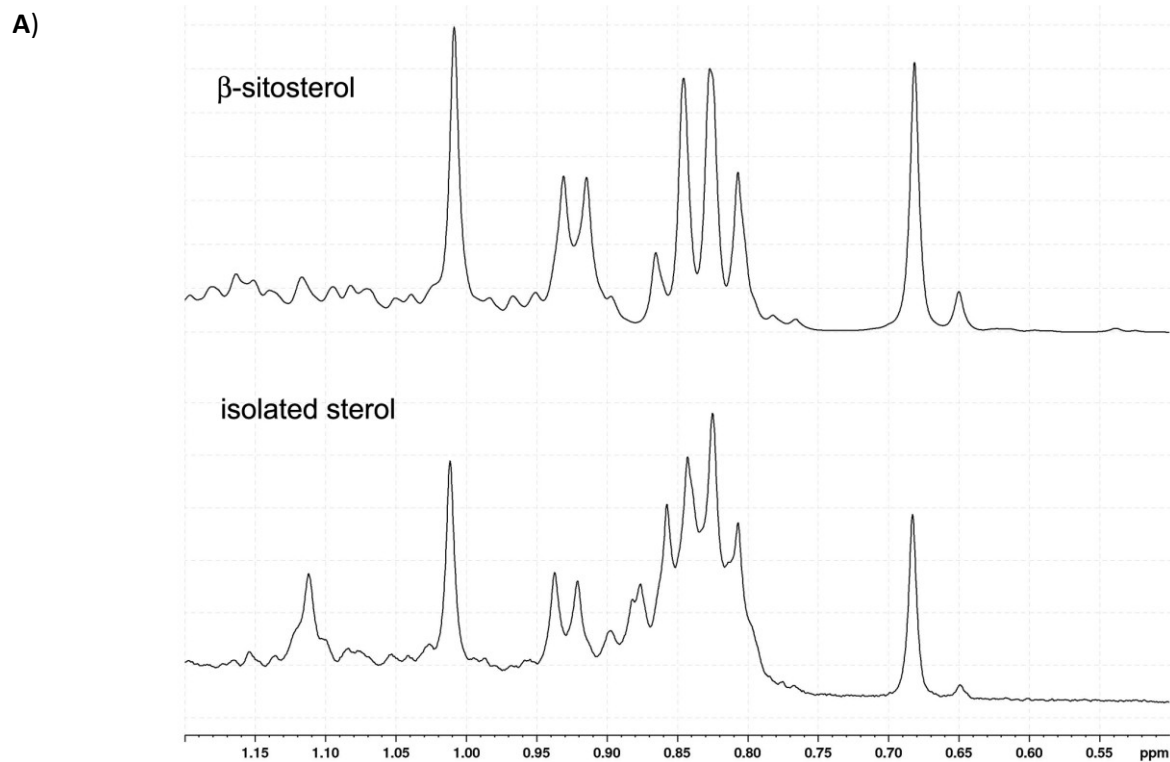
		Main identified orthologs in sequenced genomes of marine diatoms											
Sterol biosynthesis key genes		<i>T.oceanica</i>	Identity	<i>T.pseudonana</i>	Identity	<i>F.cylindrus</i>	Identity	<i>P.tricornutum</i>	Identity	<i>PN.multistriata</i>	Identity	<i>C.crispus</i>	Identity
<i>HMG-CoA synthase</i>	HMGS	EJK77540.1	84%	XP_002286567.1	87%	OEU16766.1	72%	XP_002185047.1	65%	VEU40541.1	65%	XP_005712048.1	37%
<i>HMG-CoA reductase</i>	HMGR	EJK68891.1	73%	XP_002289576.1	89%	OEU16221.1	78%	XP_002185302.1	74%	not determined		not determined	
<i>Mevalonate kinase</i>	MEVK	EJK44829.1	75%	XP_002287787.1	83%	OEU13577.1	69%	XP_002177012.1	26%	VEU38327.1	69%	XP_005711370.1	37%
<i>Squalene synthase</i>	SQS	EJK68341.1	77%	XP_002288355.1	88%	OEU14974.1	70%	XP_002180940.1	72%	VEU34553.1	65%	XP_005712480.1	36%
<i>Cycloartenol synthase</i>	CAS	EJK52279.1	64%	XP_002287432.1	85%	OEU13816.1	68%	XP_002185678.1	71%	VEU33723.1	57%	XP_005715573.1	45%
<i>Cycloartenol-C-24-methyltransferase</i>	SMT1	EJK49936.1	37%	XP_002287743.1	69%	OEU12190.1	50%	XP_002178531.1	42%	VEU39293.1	26%	XP_005710912.1	25%
<i>Cycloeucaleanol cycloisomerase</i>	CPI	EJK52525.1	68%	XP_002291862.1	72%	OEU13221.1	52%	XP_002184217.1	59%	VEU40796.1	52%	XP_005714180.1	41%
<i>Sterol 14 demethylase</i>	CYP	EJK78043.1	81%	XP_002289823.1	91%	OEU16403.1	59%	XP_002185538.1	65%	VEU38469.1	61%	XP_005711653.1	46%
<i>Delta 14 sterol reductase</i>	CYP	EJK57401.1	34%	XP_002287678.1	74%	OEU08604.1	56%	XP_002183899.1	35%	VEU34167.1	51%	XP_005715240.1	33%
<i>C8 sterol isomerase (ScERG2)</i>	ERG2											XP_005718833.1	47%
<i>C-5 sterol desaturase</i>	DWF7	EJK66009.1	38%	XP_002295550.1	80%	OEU21245.1	51%	XP_002179834.1	55%	VEU39255.1	65%	XP_005719132.1	30%
<i>7 dehydrocholesterol reductase</i>	DWF5	EJK73338.1	76%	XP_002289717.1	79%	OEU22434.1	70%	XP_002184305.1	38%	VEU39255.1	63%	XP_005719145.1	34%
<i>24 dehydrocholesterol reductase (Sm-TR10988)</i>	DHCR24					OEU21854.1	43%			VEU38364.1	42%	XP_005713924.1	35%
<i>Delta(24(24(1)))-sterol reductase</i>	24SR	EJK57400.1	30%	XP_002287183.1	31%	OEU08604.1	35%	XP_002182735.1	32%	VEU34167.1	31%	XP_005715240.1	29%

Supplementary Table 3: Comparison analysis of plants, fungal, yeast and human enzymes involved key reaction of sterol synthesis vs the *S. marinoi* and *C. cryptica* sequenced transcriptomes. Resulted hits showing alignment query coverage $\leq 39\%$ were considered not significant and dark grey highlighted.

Organisms	Descriptions	Genes	Best BLAST hit			
			<i>S. marinoi</i>	Statistics	<i>C. cryptica</i>	Statistics
<i>Arabidopsis thaliana</i>	<i>Squalene epoxidase</i>	SQE1	No hit		No hit	
<i>Homo sapiens</i>	<i>Squalene epoxidase</i>	SQE	No hit		No hit	
<i>Pheodactylum tricornutum</i>	<i>Alternative squalene epoxidase</i>	AltSQE	TR7561	QC = 58% I = 25% P= 42%	TR29442	QC = 50% I = 26% P= 43%
<i>Arabidopsis thaliana</i>	<i>Cycloarthenol synthase</i>	CAS1	TR12960	QC = 86% I = 47% P= 64%	TR3126	QC = 86% I = 48% P= 64%
<i>Neurospora crassa</i>	<i>Lanosterol synthase</i>	LSS		QC = 92% I = 38% P= 54%		QC = 86% I = 39% P= 55%
<i>Homo sapiens</i>	<i>Lanosterol synthase</i>	LSS		QC = 89% I = 44% P= 60%		QC = 90% I = 45% P= 60%
<i>Aspergillus nidulans</i>	<i>Lanosterol synthase</i>	LSS		QC = 87% I = 32% P= 49%		QC = 87% I = 32% P= 48%
<i>Arabidopsis thaliana</i>	<i>Sterol 14 demethylase</i>	CYP10A1	TR7602	QC = 76% I = 25% P= 43%	TR12199	QC = 77% I = 25% P= 41%
<i>Arabidopsis thaliana</i>	<i>Sterol 14 demethylase</i>	CYP10A2		QC = 41% I = 29% P= 48%		QC = 42% I = 30% P= 31%
<i>Arabidopsis thaliana</i>	<i>C8-sterol isomerase</i>	AT1G05440	TR11754	QC = 15% I = 23% P= 51%	No hit	
<i>Arabidopsis thaliana</i>	<i>C8-sterol isomerase</i>	HYD1	TR27203	QC = 24% I = 23% P= 55%	TR38445	QC = 23% I = 41% P= 50%
<i>Saccharomyces cerevisiae</i>	<i>C8-sterol isomerase</i>	ERG2	TR4901	QC = 22% I = 29% P= 57%	TR41732	QC = 18% I = 32% P= 51%
<i>Aspergillus nidulans</i>	<i>C8-sterol isomerase</i>	ERG2	TR2164	QC = 12% I = 42% P= 70%	TR9781	QC = 19% I = 36% P= 54%
<i>Neurospora crassa</i>	<i>C8-sterol isomerase</i>	ERG2	TR4901	QC = 22% I = 29% P= 57%	TR17006	QC = 21% I = 27% P= 45%
<i>Arabidopsis thaliana</i>	<i>Sterol methyltransferase 1</i>	SMT1	TR612	QC = 89% I = 46% P= 65%	TR44522	QC = 98% I = 43% P= 53%
<i>Chlamydomonas reinhardtii</i>	<i>Sterol methyltransferase</i>	SMT1-2	TR612	QC = 84% I = 38% P= 53%	TR45147	QC = 84% I = 40% P= 53%
<i>Arabidopsis thaliana</i>	<i>Sterol methyl-oxidase</i>	SMO1-1	TR11964	QC = 62% I = 27% P= 43%	TR32604	QC = 60% I = 31% P= 45%
<i>Arabidopsis thaliana</i>	<i>Sterol methyl-oxidase</i>	SMO1-2	TR30213	QC = 81% I = 26% P= 45%		QC = 80% I = 23% P= 42%
<i>Arabidopsis thaliana</i>	<i>Sterol methyl-oxidase</i>	SMO1-3	TR11964	QC = 70% I = 27% P= 45%		QC = 56% I = 30% P= 45%
<i>Arabidopsis thaliana</i>	<i>Sterol methyl-oxidase</i>	SMO2-1	TR319	QC = 84% I = 23% P= 40%		QC = 58% I = 25% P= 52%
<i>Arabidopsis thaliana</i>	<i>Sterol methyl-oxidase</i>	SMO2-2	TR28078	QC = 78% I = 28% P= 46%		QC = 59% I = 33% P= 52%
<i>Homo sapiens</i>	<i>C4 methyl sterol oxidase</i>	SMO	TR11964	QC = 86% I = 25% P= 43%		QC = 75% I = 24% P= 45%
<i>Saccharomyces cerevisiae</i>	<i>Sterol-4-alpha-carboxylate 3-dehydrogenase</i>	ERG26	TR28078	QC = 98% I = 25% P= 42%	TR14638	QC = 98% I = 29% P= 48%

<i>Homo sapiens</i>	<i>3-beta-hydroxysteroid-Delta(8),Delta(7)-isomerase</i>	EBP	No hit		No hit	
<i>Arabidopsis thaliana</i>	<i>C5-sterol desaturase</i>	DWF7	TR28093	QC = 58% I = 23% P= 42%	TR48724	QC = 51% I = 25% P= 43%
<i>Homo sapiens</i>	<i>C5-sterol desaturase</i>	SC5D	TR28093	QC = 79% I = 25% P= 43%	TR48724	QC = 48% I = 32% P= 43%
<i>Arabidopsis thaliana</i>	<i>7-dehydrocholesterol reductase</i>	DWF5	TR2636	QC = 86% I = 40% P= 55%	TR24068	QC = 86% I = 41% P= 56%
<i>Homo sapiens</i>	<i>24-dehydrocholesterol reductase</i>	DHCR24	TR10998	QC = 95% I = 50% P= 67%	TR28124	QC = 24% I = 25% P= 44%
<i>Arabidopsis thaliana</i>	<i>Delta 24 sterol reductase</i>	DWF1		QC = 92% I = 37% P= 54%		QC = 11% I = 33% P= 52%
<i>Solanum lycopersicum</i>	<i>Sterol side chain reductase</i>	SSR2		QC = 92% I = 39% P= 53%		QC = 25% I = 25% P= 42%
<i>Physalis alkekengi</i>	<i>Delta 24 sterol reductase</i>	24ISO		QC = 91% I = 39% P= 53%		QC = 23% I = 25% P= 45%
<i>Saccharomyces cerevisiae</i>	<i>Delta 24(24) sterol reductase</i>	ERG4	TR213	QC = 94% I = 43% P= 59%	TR29411	QC = 93% I = 43% P= 58%
<i>Saccharomyces cerevisiae</i>	<i>Delta 22 sterol desaturase</i>	ERG5	TR7602	QC = 69% I = 22% P= 49%	TR3228	QC = 40% I = 28% P= 43%

Supplementary Figure 1: NMR (600 MHz, CDCl₃) identification of clionasterol from *S. marinoi* and *C. cryptica*. (A) Comparison of ¹H NMR upfield-shifted signals of standard β-sitosterol and the natural 24 ethyl sterol isolated from the diatoms; (B) ¹H-NMR data of clionasterol and β-sitosterol, and comparison with the literature assignment (Nes *et al.* Lipids (1976) 11: 118).

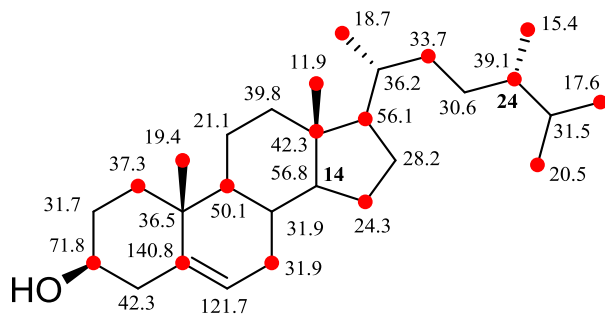


B)

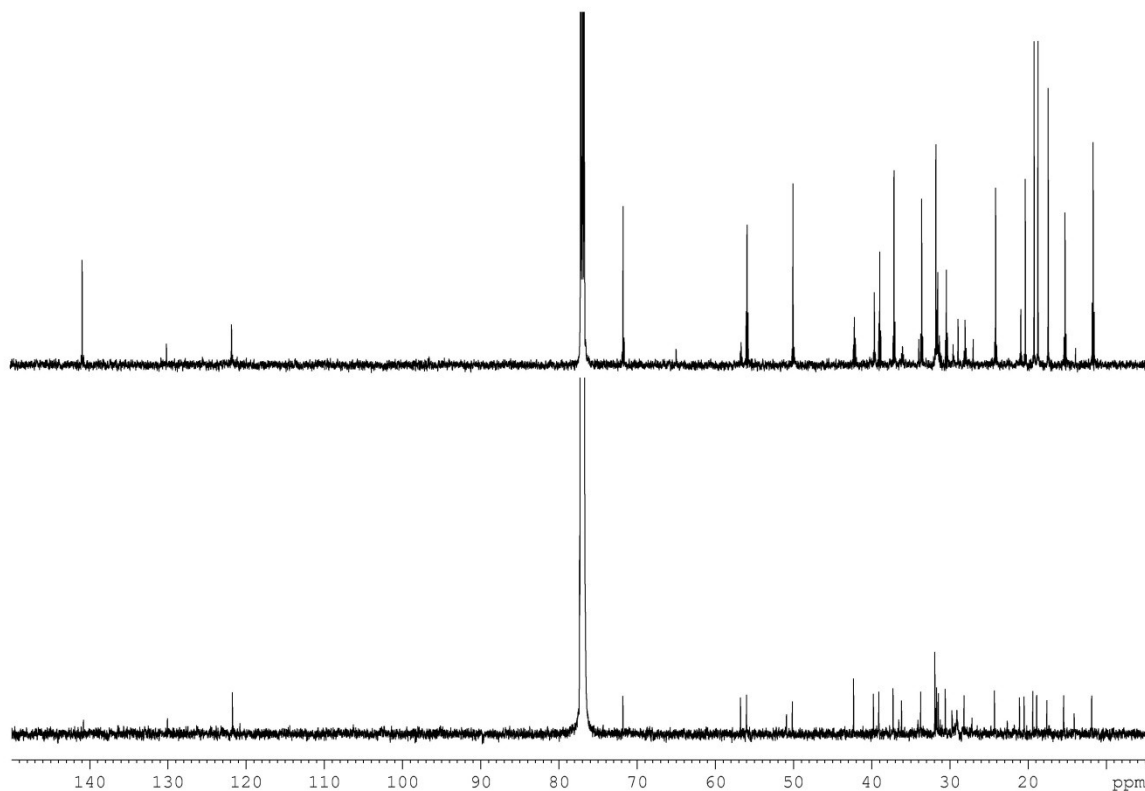
Positions	Experimental data		Literature data	
	Clionasterol ^a	β-Sitosterol ^b	Clionasterol	β-Sitosterol
18	0.6800	0.6826	0.6818	0.6818
19	1.0090	1.0090	1.0090	1.0090
21	0.9247	0.9243	0.9272	0.9227
26	0.8308	0.8371	0.8318	0.8364
27	0.8106	0.8163	0.8091	0.8136
29	0.8578	0.8462	0.8545	0.8454

^a sample isolated from *C. cryptica*; ^b standard.

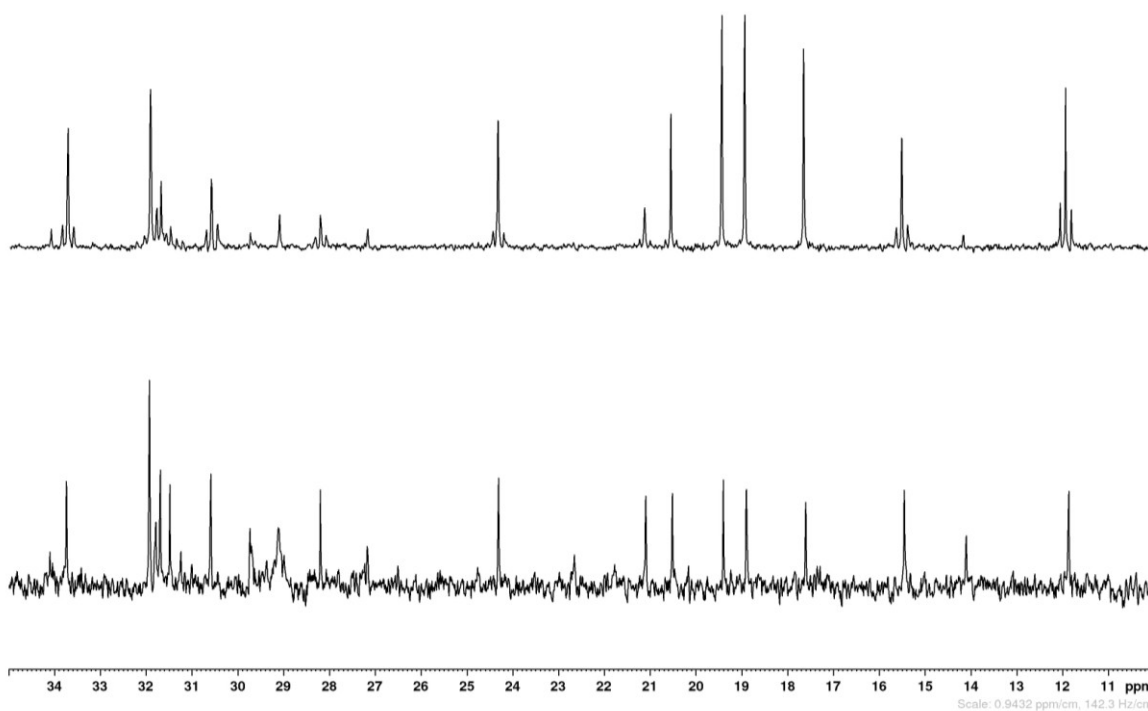
Supplementary Figure 2: Incorporation of 1-¹³C-glucose into dihydrobrassicasterol of *C. cryptica*. Enriched carbons are shown by a red spot on the structure. **(A)** Full and **(B-D)** four enlarged regions of the ¹³C-NMR spectra (CDCl₃, 125 MHz) of labeled (top trace) and unlabeled (down trace) dihydrobrassicasterol. Intensity of ¹³C signals of labeled and unlabeled spectra are normalized on C14 (56.79 ppm).



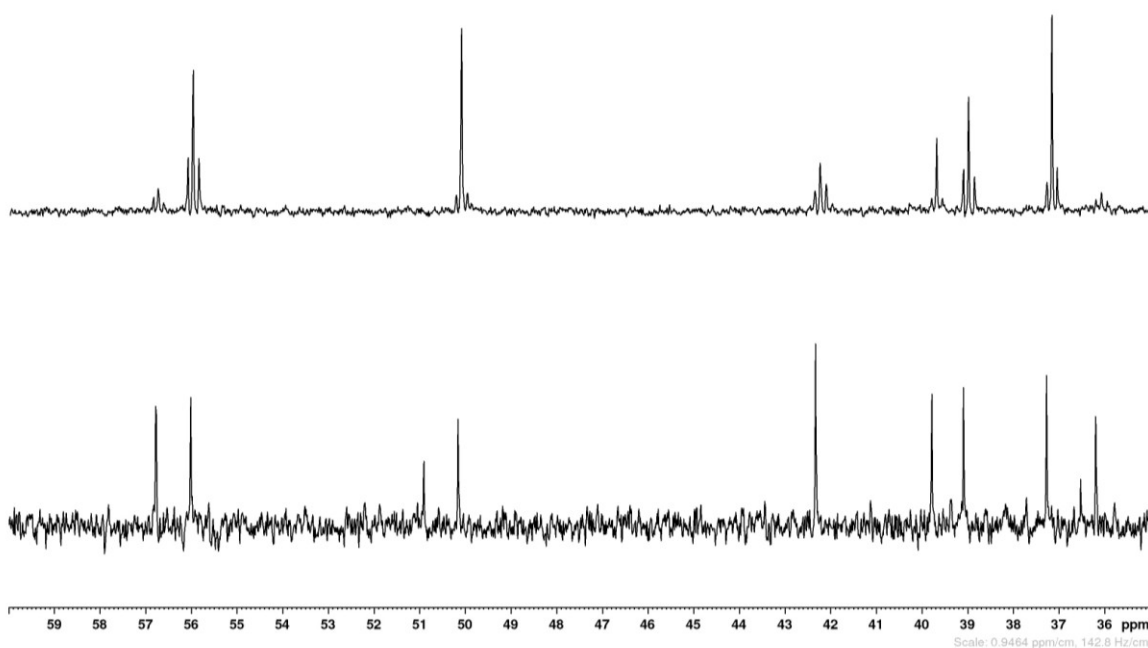
A) Full spectral width (155-0 ppm).



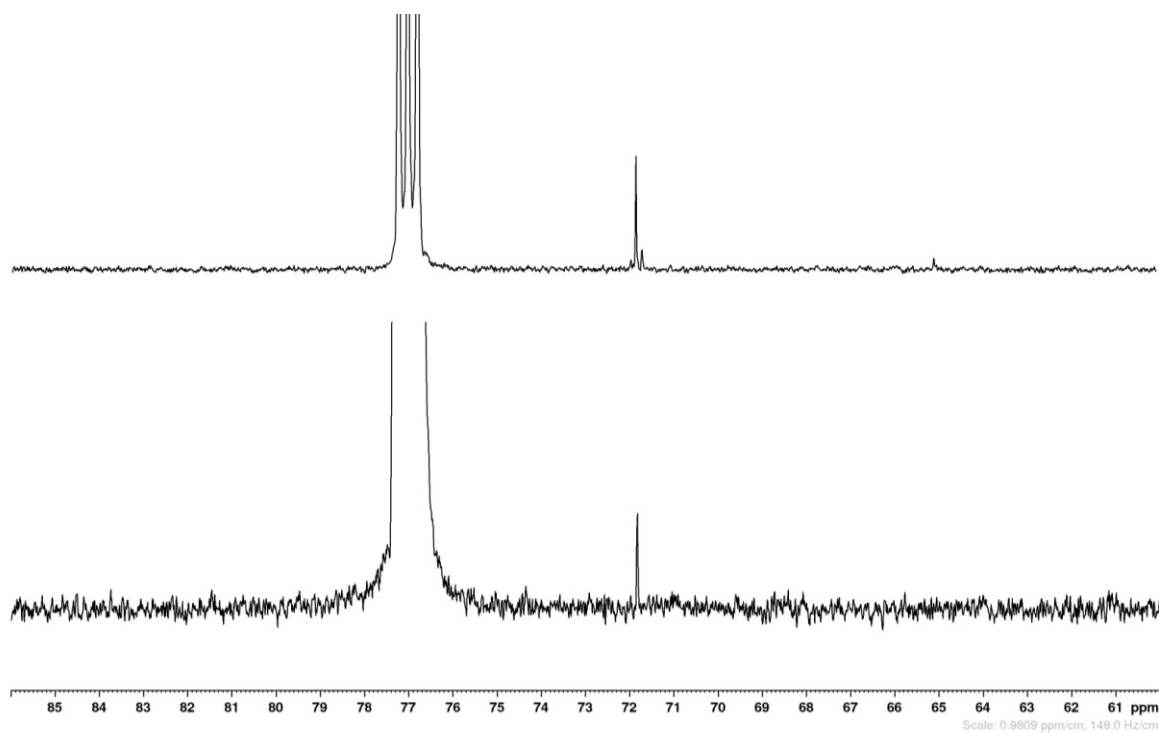
B) Region between 35 and 10 ppm.



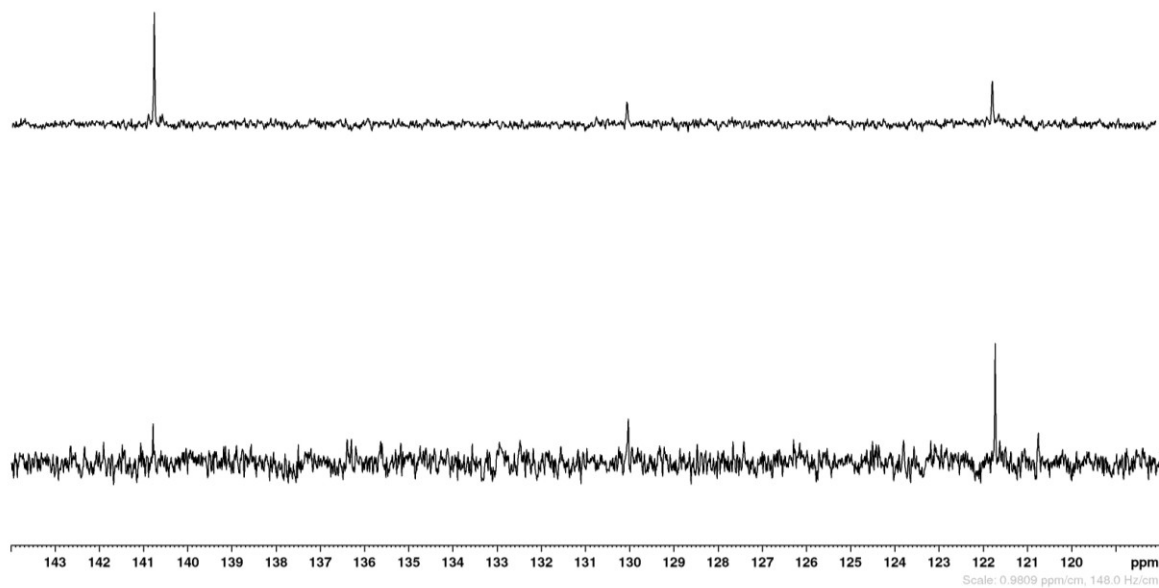
C) Region between 60 and 35 ppm.



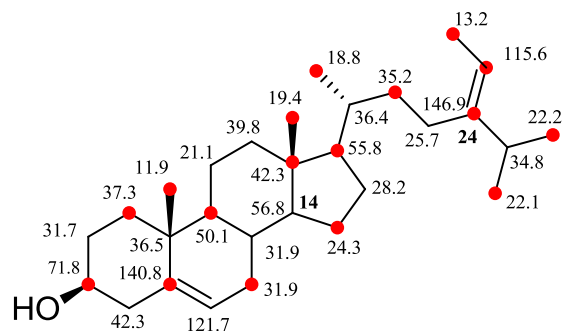
D) Region between 85 and 60 ppm.



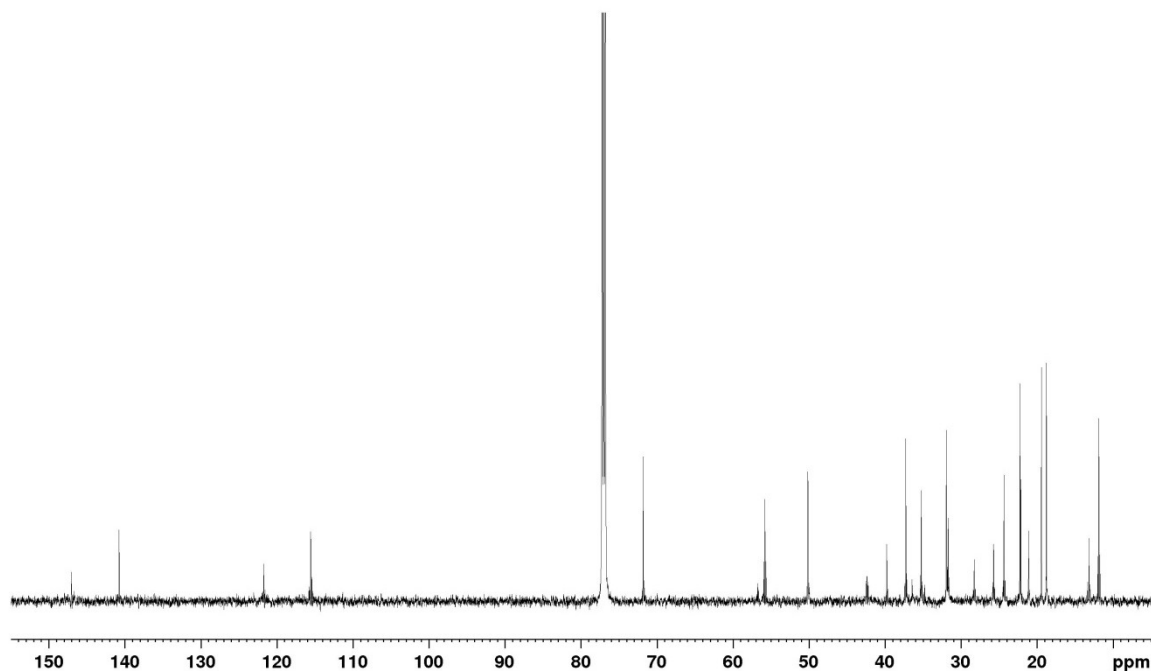
E) Region between 144 and 118 ppm



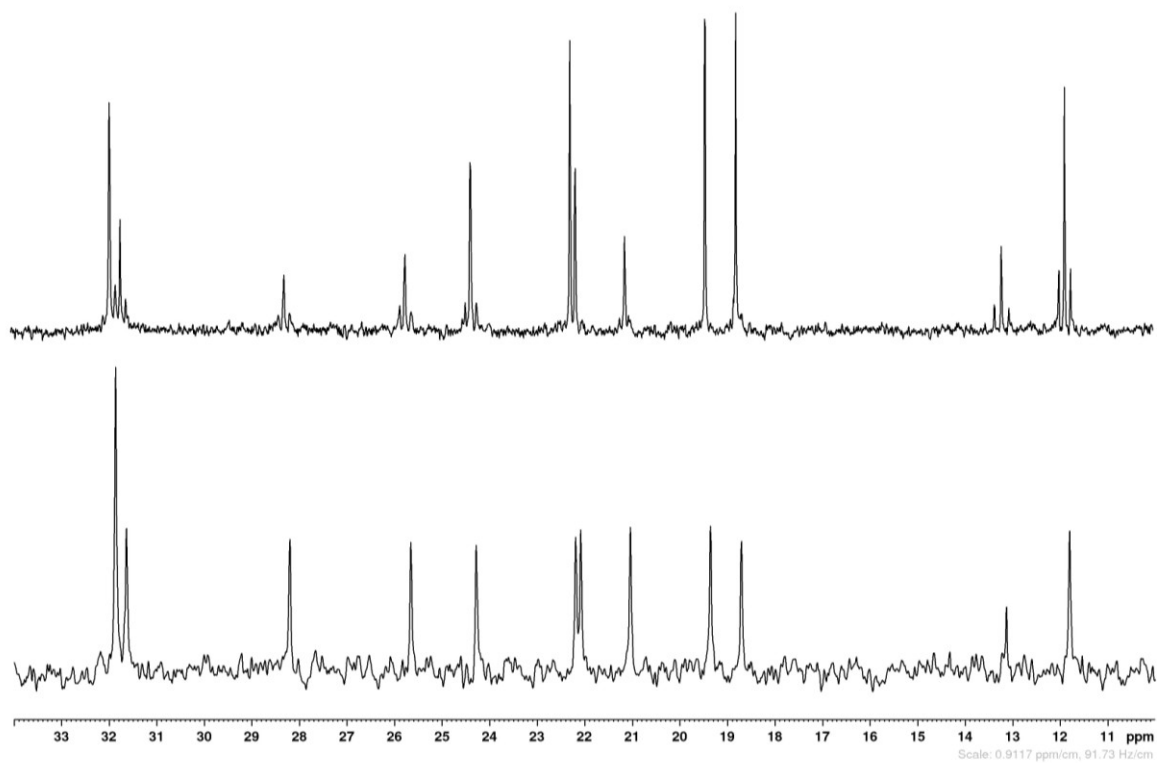
Supplementary Figure 3: Incorporation of 1-¹³C glucose into fucosterol of *C. cryptica*. Enriched carbons are shown by a red spot on the structure. **(A)** Full and **(B-D)** enlarged regions of the ¹³C-NMR spectra (CDCl₃, 125 MHz) of labeled (top trace) and unlabeled (down trace) fucosterol. Intensity of ¹³C signals of labeled and unlabeled spectra are normalized on C14 (56.79 ppm).



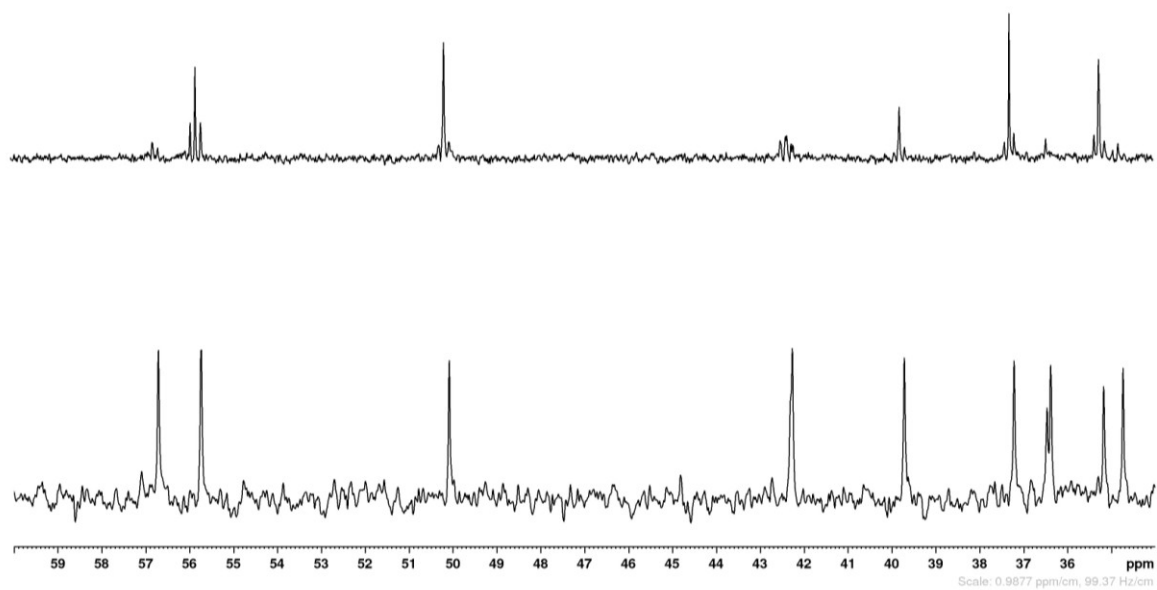
A) Full spectral width (160-0 ppm).



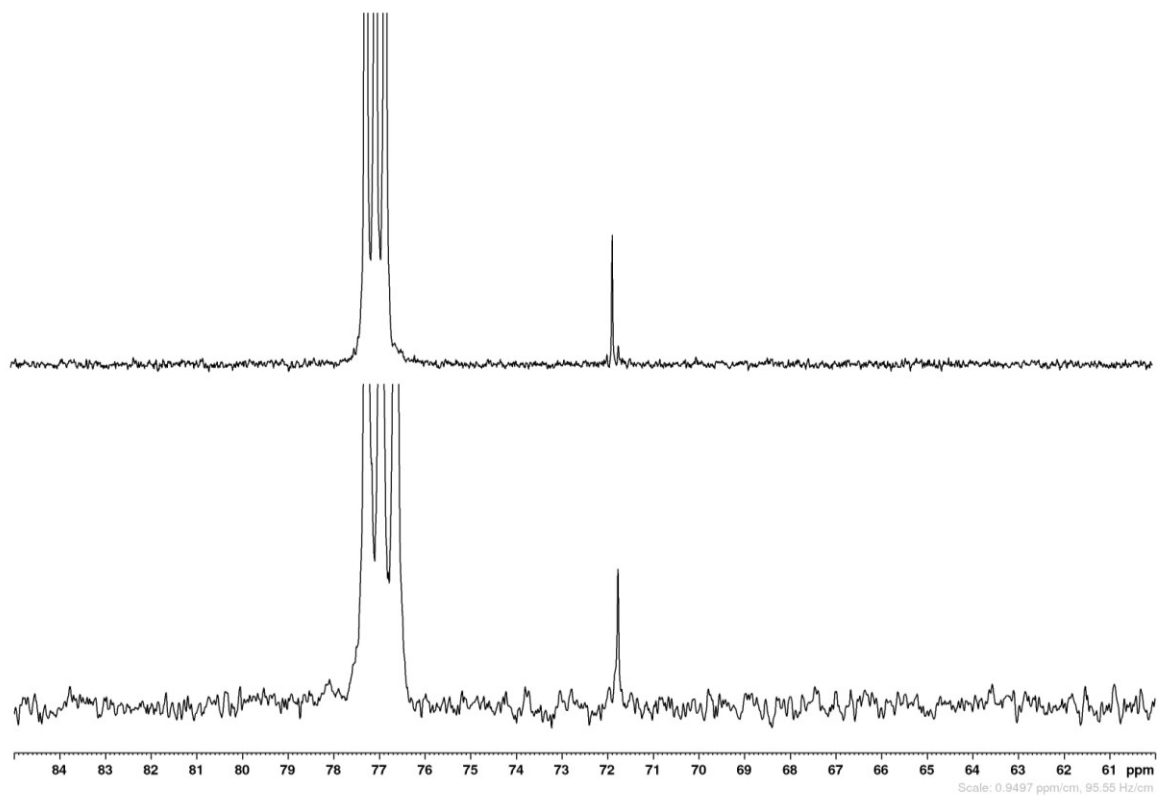
B) Region between 35 and 10 ppm.



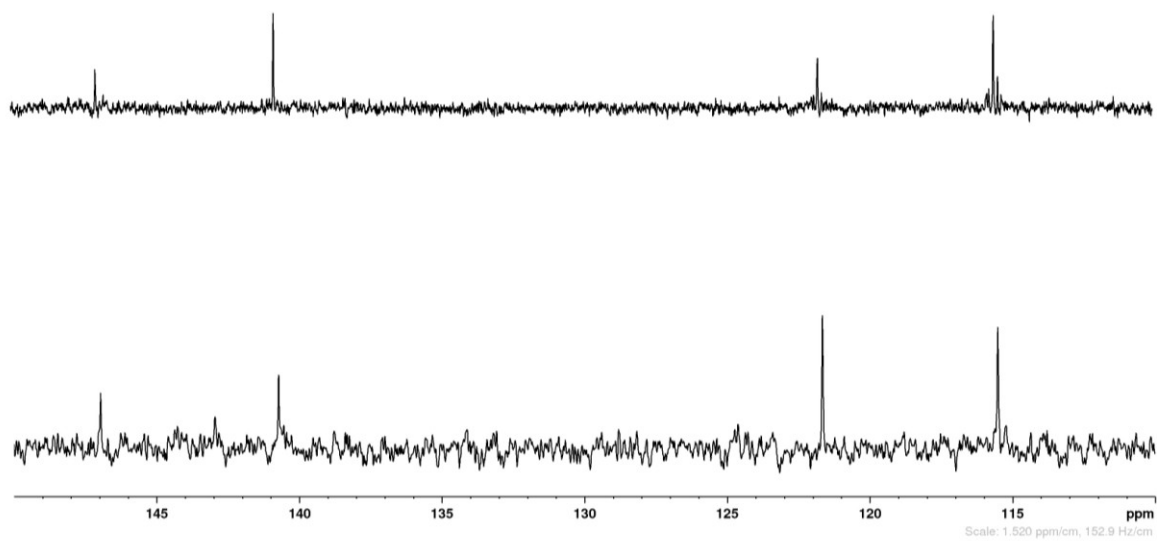
C) Region between 60 and 35 ppm.



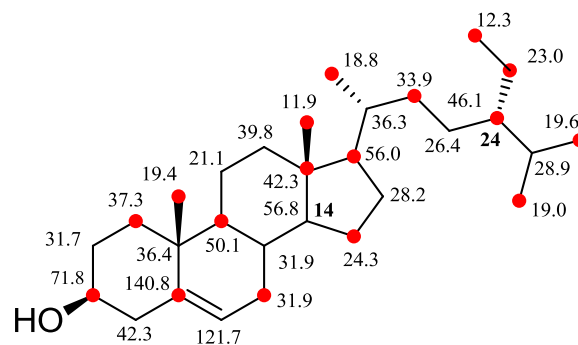
D) Region between 85 and 60 ppm.



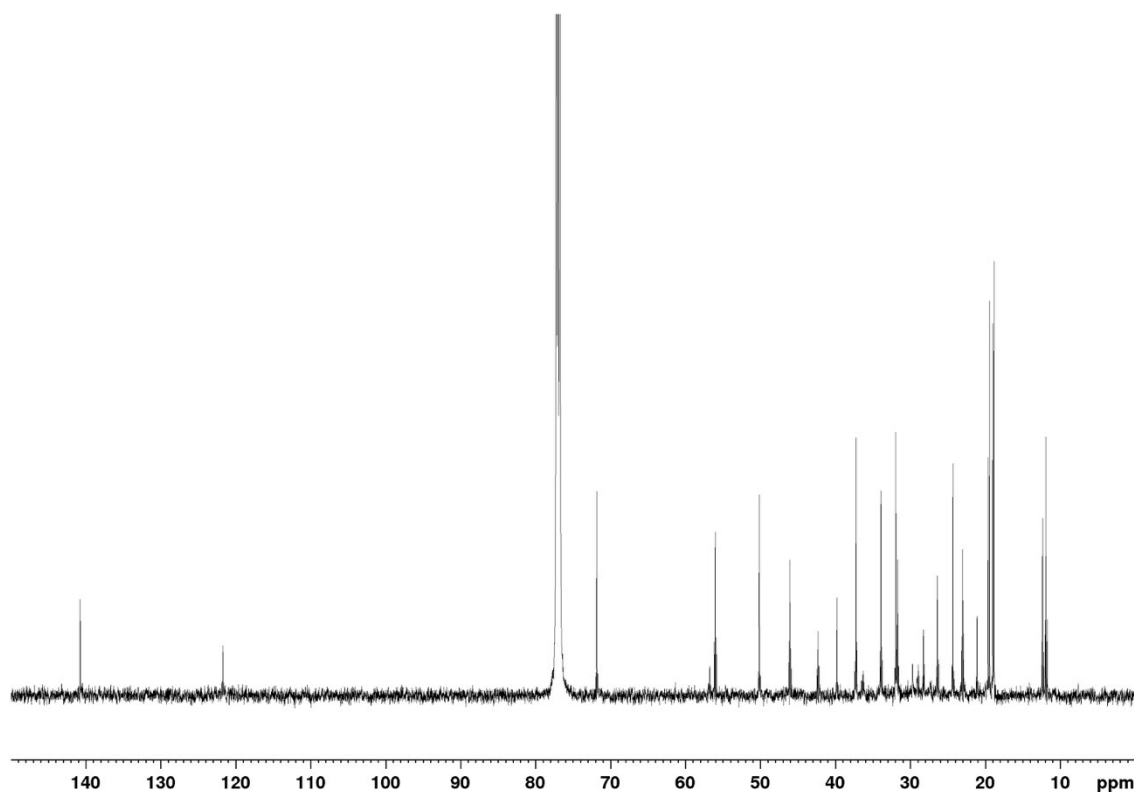
E) Region between 150 and 110 ppm.



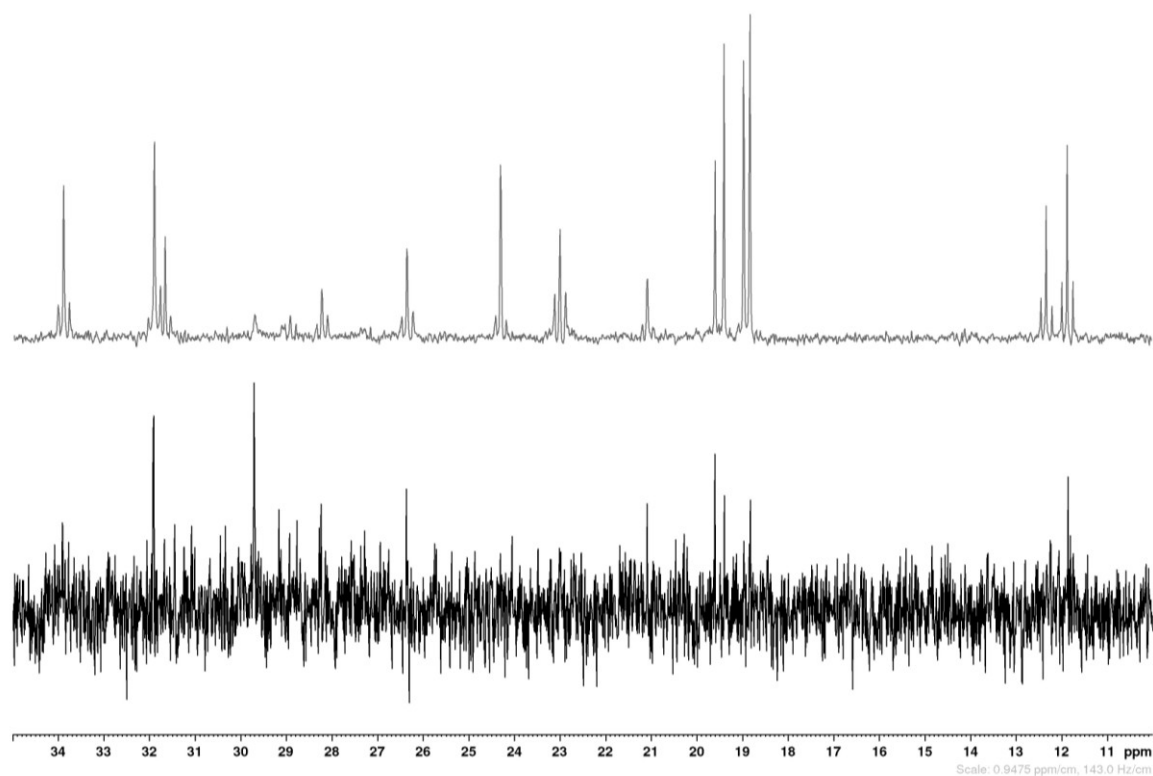
Supplementary Figure 4: Incorporation of 1-¹³C glucose into clionasterol of *C. cryptica*. Enriched carbons are shown by a red spot on the structure. **(A)** Full and **(B-D)** enlarged regions of the ¹³C-NMR spectra (CDCl₃, 125 MHz) of labeled clionasterol.



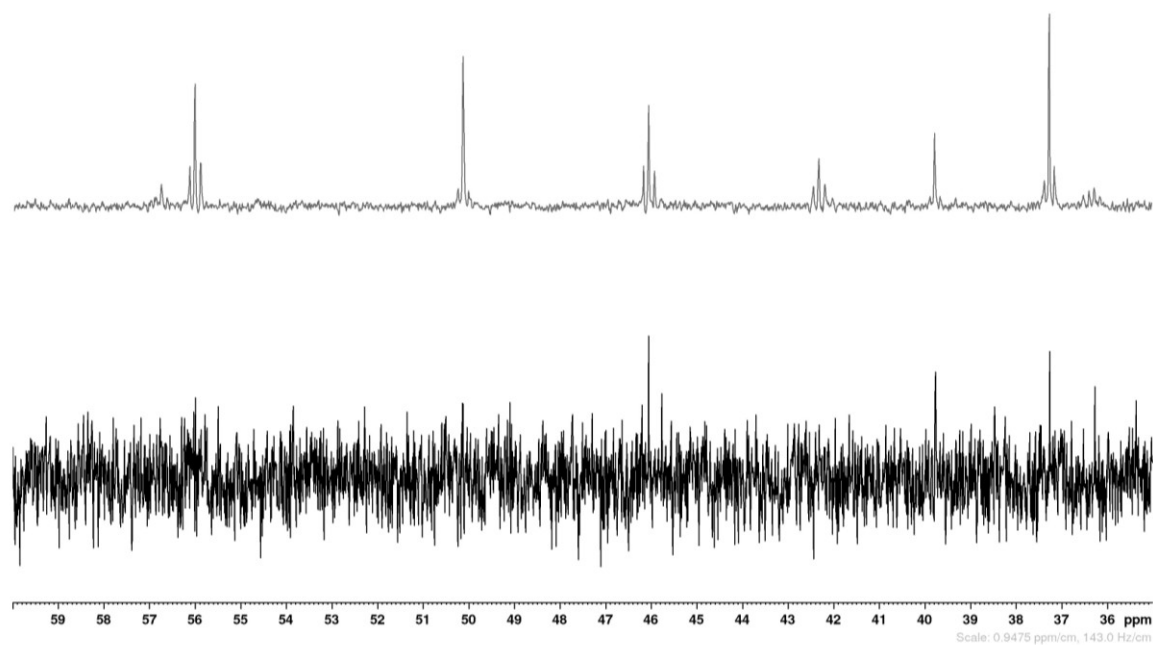
A) Full spectral width (160-0 ppm).



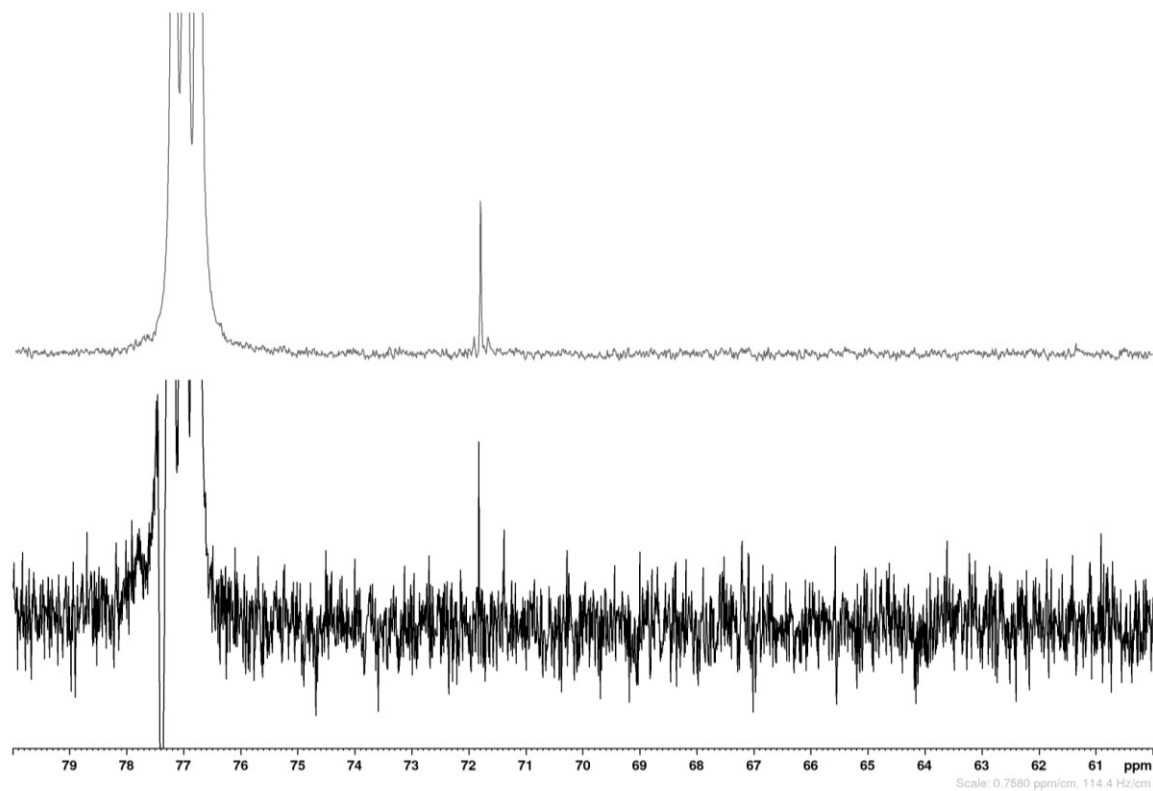
B) Region between 35 and 10 ppm.



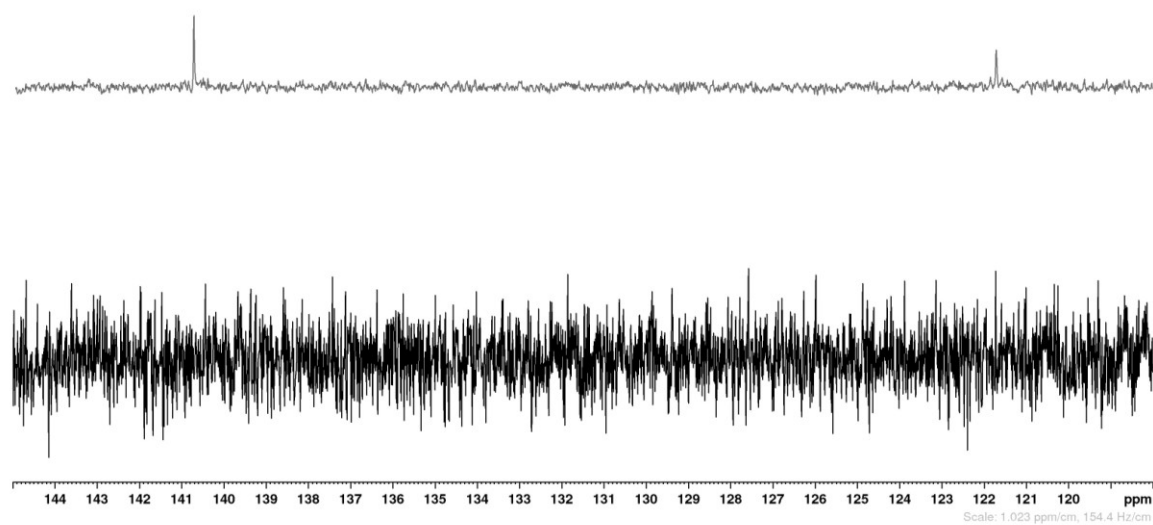
C) Region between 60 and 35 ppm.



D) Region between 85 and 60 ppm.



E) Region between 145 and 118 ppm.



Supplementary Figure 5: Alignment of amino acid sequences of the catalytic region of the putative proteins of *S. marinoi* and *C. cryptica* with homologous regions from CAS and LSS in animals, yeast and fungi.

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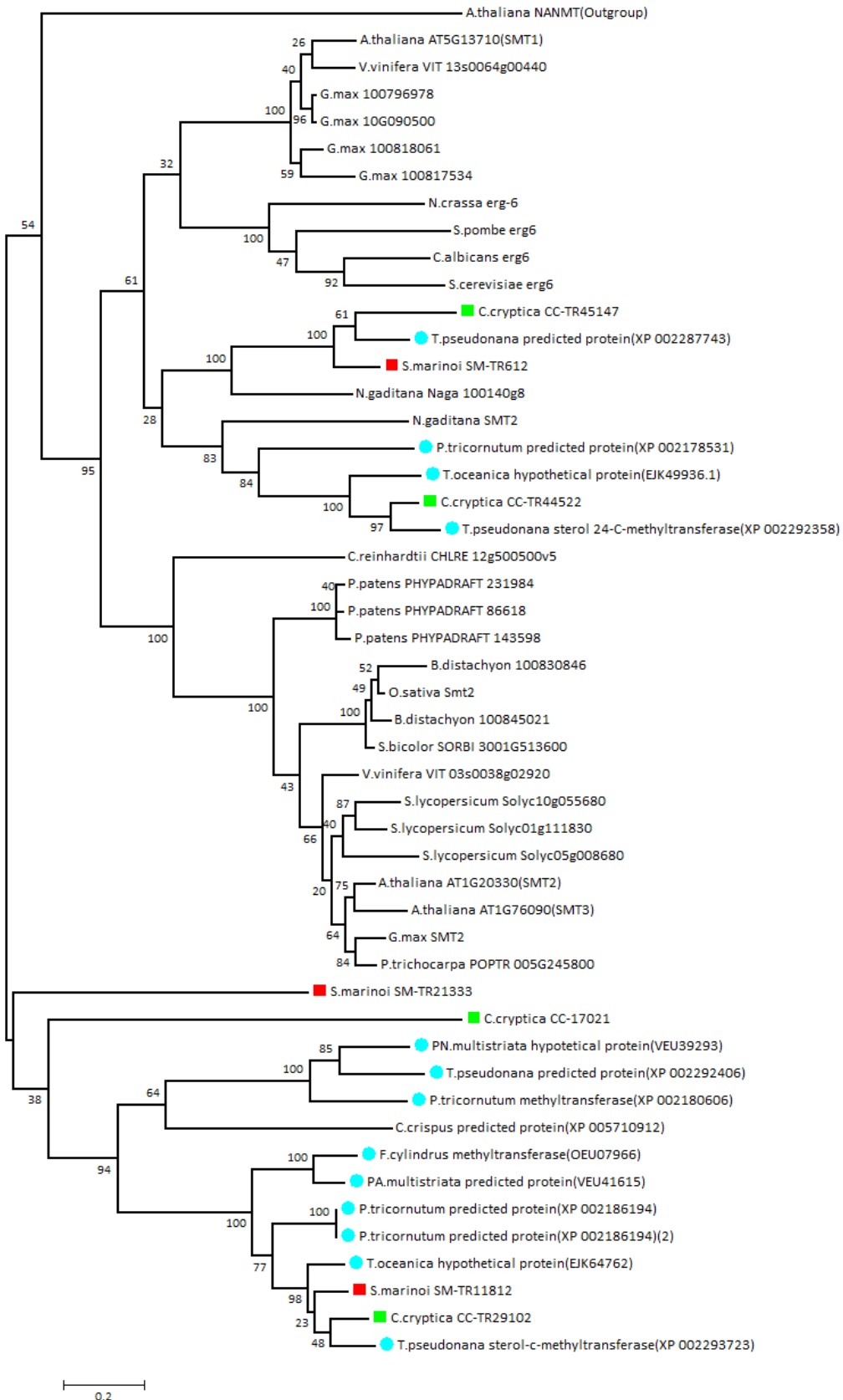
A.nidulans_LSS      342  DSNDLTNSTGVSVD AFLNMI AFYC--KEGPD SKALRRIQTASLEYLWMGPRGMQVMSIH 399
H.sapiens_LSS      325  ADDRFTKSI SIGPI SKTINMLV RWY--VDGPASTAFQEHVSRI PDYLWMLDGMKMQGTTN 382
A.thaliana_CAS1    355  YEDENTRYICIGPVNKV LNMLCCWV---EDPNSEAFKLHLPRIHDFLWLAEDGMKMQGYN 411
A.thaliana_CAS2    264  YEDENTRYICIGPVNKV LNMLCCWV---EDPNSEAFKLHLPRIHDFLWLAEDGMKMQGYN 320
S.marinoi_CAS      776  AEDLQTNFIDIGPVNKALNMVSAFHSANNDINDPAVRSHMMRPDYLWVAEDGMKMQGYN 835
C.criptica_CAS     299  AEDLQTNFYIDIGPVNKALNMVAAYHAANNNDINDPAVQSHMMRPDYLWLAEDGMKMQGYN 358
N.crassa_LSS       348  MEDENTDYSIDLAPVNAAMNTVCCYV--RDGPGAYSVRRHVERLDEVLWVNAEGMLCNGTIN 405
S.cerevisiae_LSS   328  TELQNTDSL CIAPVNQAF CALVTLI--EEGVDSEAFQRLQYRFKDALPHGPGQGMTIMGTN 385

A.nidulans_LSS     400  AGHTWETA FVLQAYAEGGLSKVPEIQAAIERAYKYLVEQQHV-----VDYPE 446
H.sapiens_LSS      383  GSQLWDTAFAIQALLEAGGHRPEFSSCLQKAHEFLRLSQVP-----DNPP 428
A.thaliana_CAS1    412  GSQLWDTGF AIQAILATNLVE--EYGPVLEKAHSFVKNSQVL-----EDCPG 456
A.thaliana_CAS2    321  GSQLWDTGF AIQAILATNLVE--EYGPVLEKAHSFVKNSQVL-----EDCPG 365
S.marinoi_CAS      836  GSQCWDTSF AIQAVWECGLLD--KFPILSAKVWAFLE RTQILSTEC SQSTPAYQYESC-D 892
C.criptica_CAS     359  GSQCWDTSF AIQAVRECGLLD--HFPLLSTKVWAYLE RTQILSTETSQSSPAFYESC-D 418
N.crassa_LSS       406  GVQCWDTAF AIQAIMDAGLTEDPRWRPMLIKSLEFLEDQQIR-----ENVK 451
S.cerevisiae_LSS   386  GVQTDWCAFAIQYFFVAGLAERPEFYNTIVSAYKFLCHAQFD-----TECVP 432

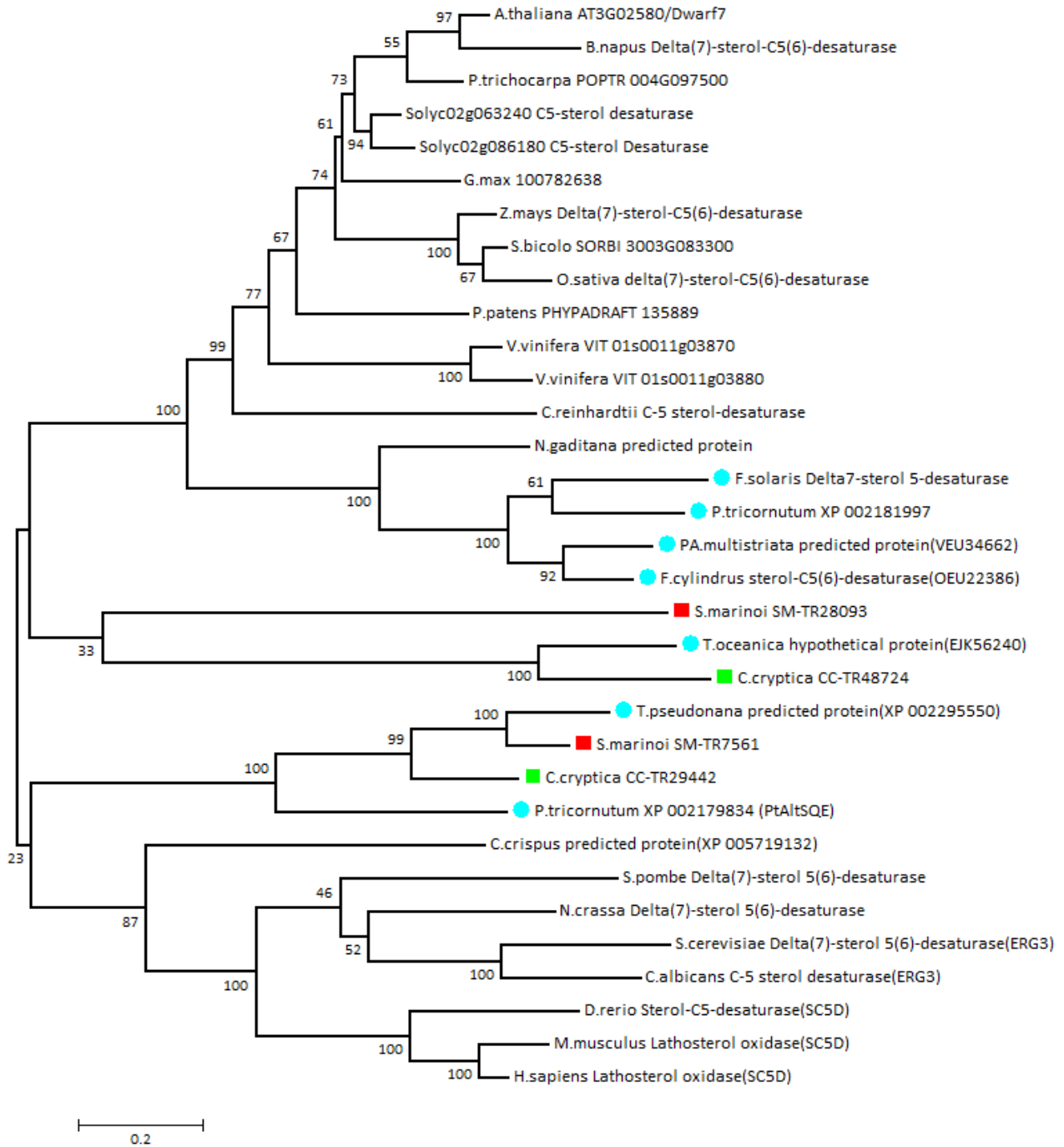
A.nidulans_LSS     447  DSQCHFFSRL--GGWPFSTRYQGNVCS DCTGEALKSILMIE-RDSRFT-----RLTT 495
H.sapiens_LSS      429  DYQ-KYYRQMRKGGFSFSTLDGWI VSDCTAEALKAVLLLQEKCPHVT-----EHIP 479
A.thaliana_CAS1    457  DLN-YWYRHISKGAWPFSTADHGWPIS DCTAEGLKAALLLS-KVPKEIVG-----EPID 508
A.thaliana_CAS2    366  DLN-YWYRHISKGAWPFSTADHGWPIS DCTAEGLKAALLLS-KVPKAI VG-----EPID 417
S.marinoi_CAS      893  SRD-KFYRHVSKGGWPFSTSAHGWPIS DCTGEGGLKGV LALM-DSPIVMDGVKRGILKDIE 950
C.criptica_CAS     419  NRD-KFYRHVSKGGWPFSTSAHGWPIS DCTGEGGLKGV LALM-NSPIVVEAVKTGV LKSID 473
N.crassa_LSS       452  DQD-KCYRHPRKGAWAFSNKDG YAVSDCVSEALKSVIILQ-KTPGFP-----TLID 501
S.cerevisiae_LSS   433  GS----YDRKRKGAWGFSTKTQGYT VADCTAEAIKAIIMVK-NSPVFSEV-----HHMIS 482

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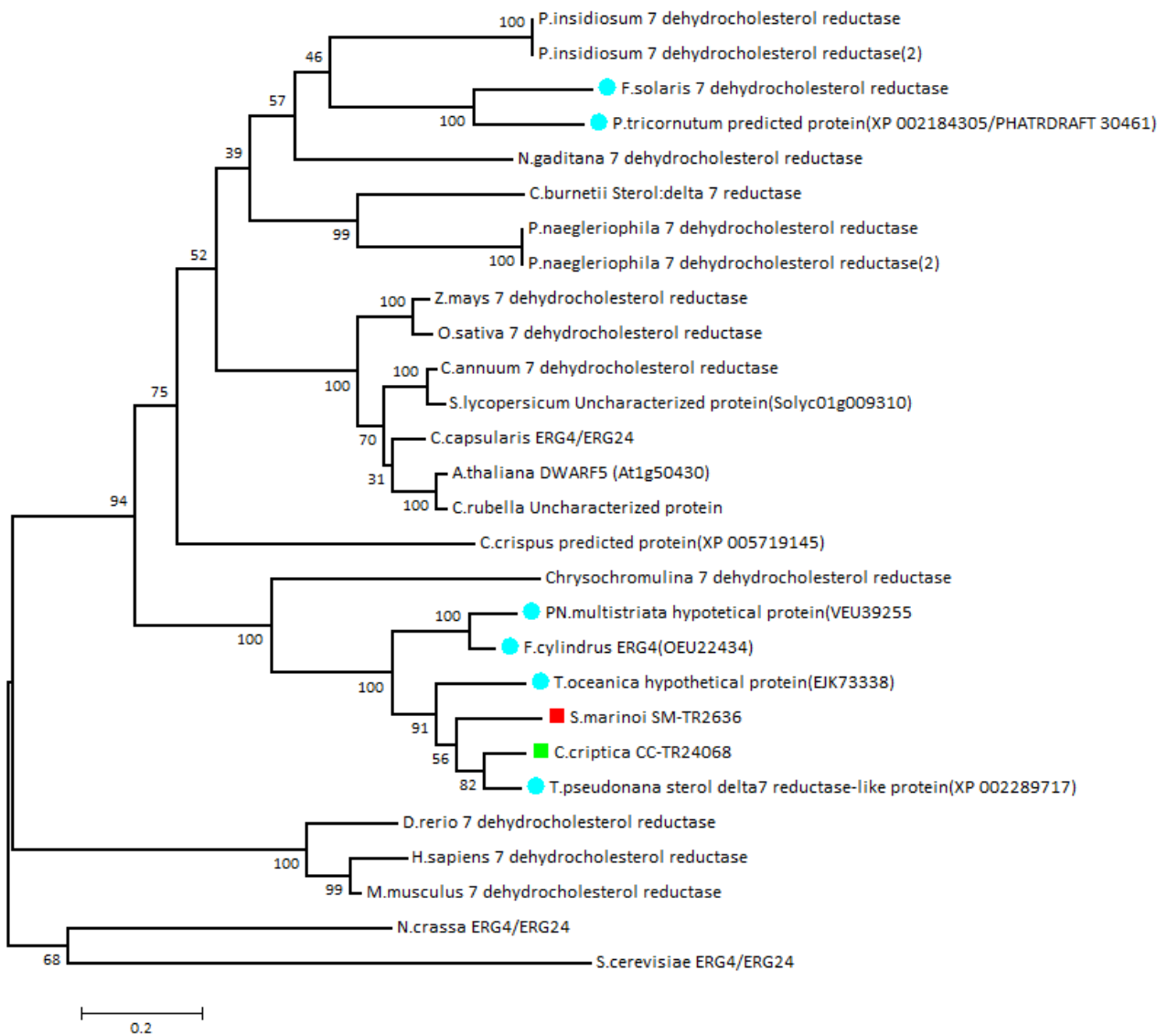

Supplementary Figure 6: Rooted phylogenetic tree of Sterol Methyl Transferases (SMTs) from various organisms. The bootstrapping test (replicate = 100) is indicated on each node in order to test the phylogeny. Nicotinate methyltransferase (NANMT) was used as outgroup. Red square = *S. marinoi*; Green square = *C. cryptica*; Light blue spot = other diatom.



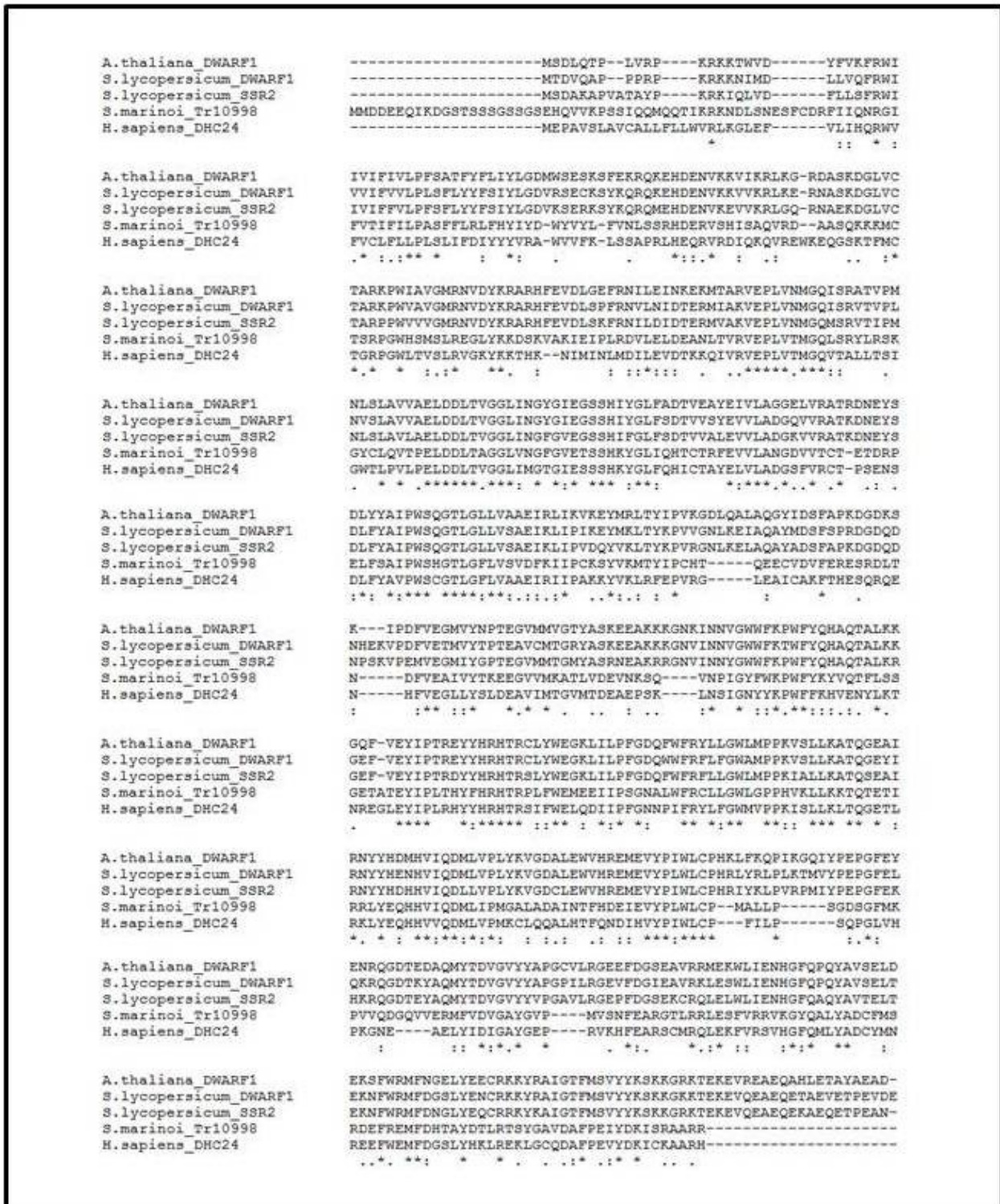
Supplementary Figure 7: Un-rooted phylogenetic trees of key enzymes of C5-sterol desaturase, including DWF7 and ERG3, from various organisms. The bootstrapping test (replicate = 100) is indicated on each node in order to test the phylogeny. Red square = *S. marinoi*; Green square = *C. cryptica*; Light blue spot = other diatom.



Supplementary Figure 8: Un-rooted phylogenetic trees of key enzymes of DWF5, ERG4 and 7 dehydrocholesterol reductase from various organisms. The bootstrapping test (replicate = 100) is indicated on each node in order to test the phylogeny. Red square = *S. marinoi*; Green square = *C. cryptica*; Light blue spot = other diatom.



Supplementary Figure 9: Alignment of aminoacid sequences of DWF1 from *A. thaliana* and *S. lycopersicum*, SSR2 from *S. lycopersicum* and 24-DHCR from *Homo sapiens* and Sm-TR10998 from *S. marinoi*.



SUPPORTING MATERIAL

Diatoms synthesize sterols by inclusion of animal and fungal enzymes in the plant pathway

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FIGURE SM1. ^1H NMR spectrum (600 MHz, C_6D_6) of acetylated 24-methylene cholesterol from *S. marinoi*.

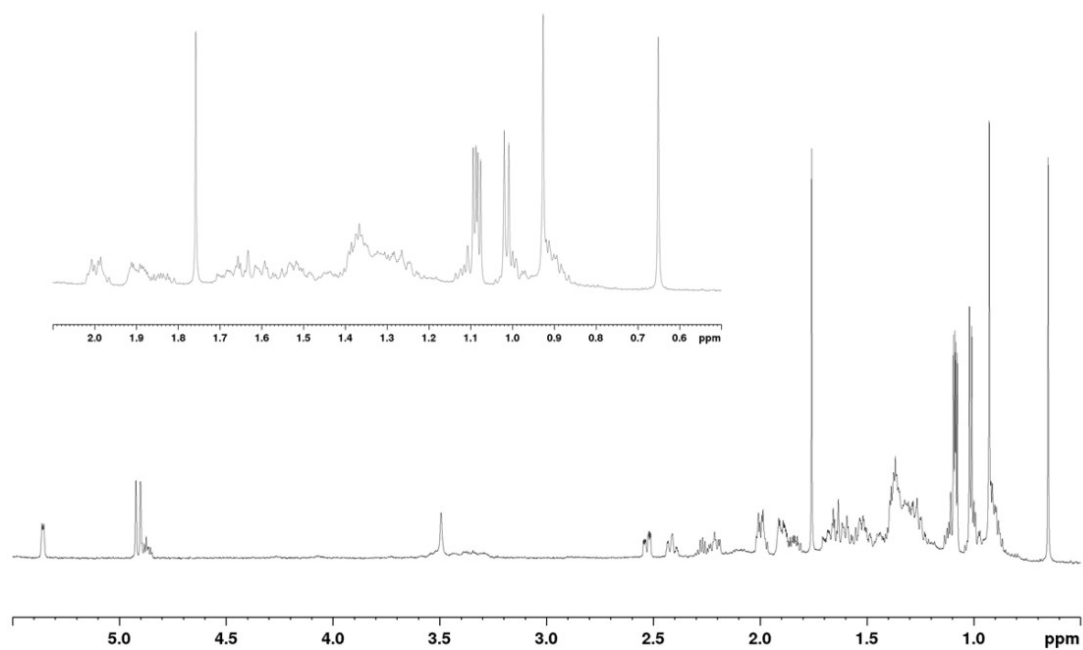


FIGURE SM2. ^1H NMR spectrum (600 MHz, CDCl_3) of natural dihydrobrassicasterol.

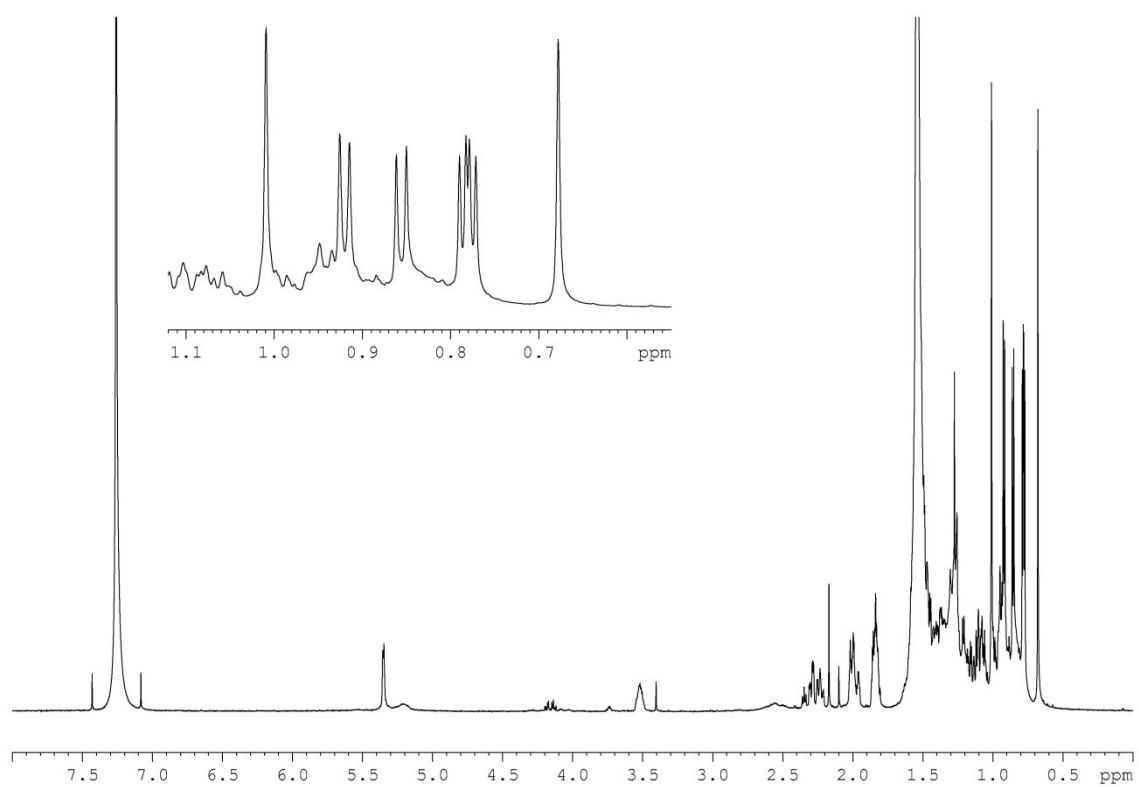


FIGURE SM3. ^{13}C NMR spectrum (125 MHz, CDCl_3) of natural dihydrobrassicasterol.

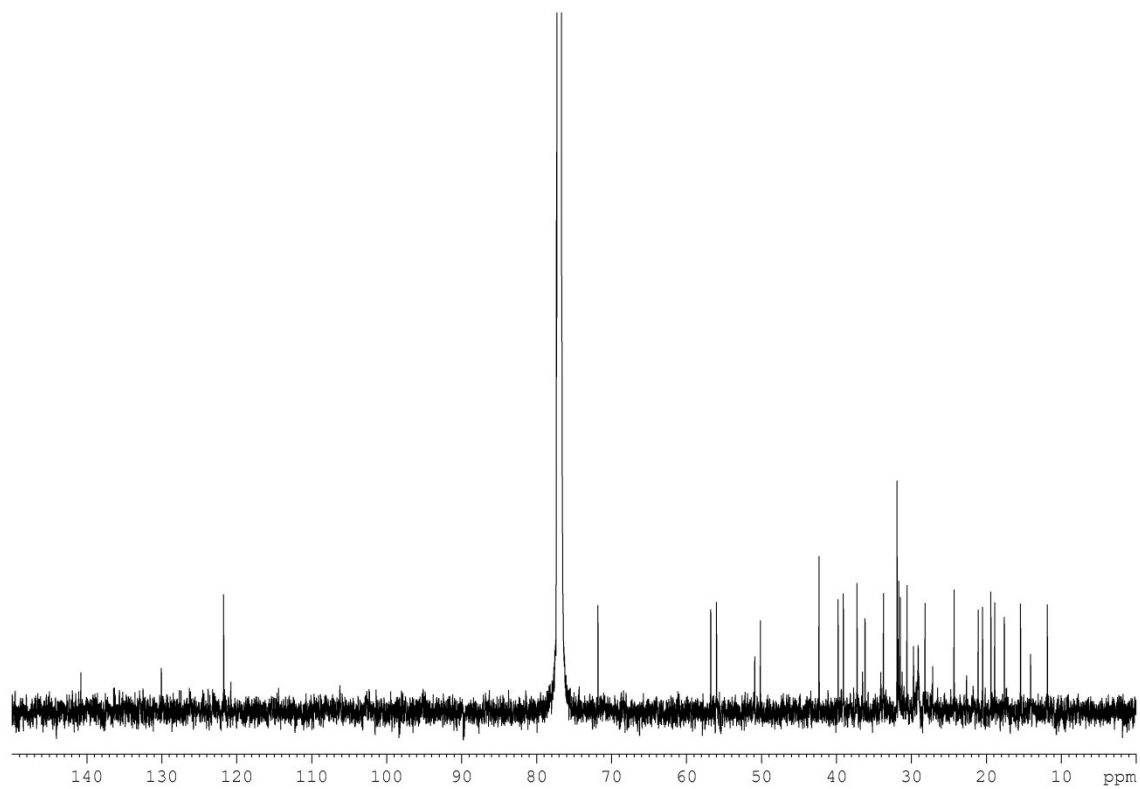


FIGURE SM4. ^1H NMR spectrum (600 MHz, CDCl_3) of clionasterol from *C. cryptica*.

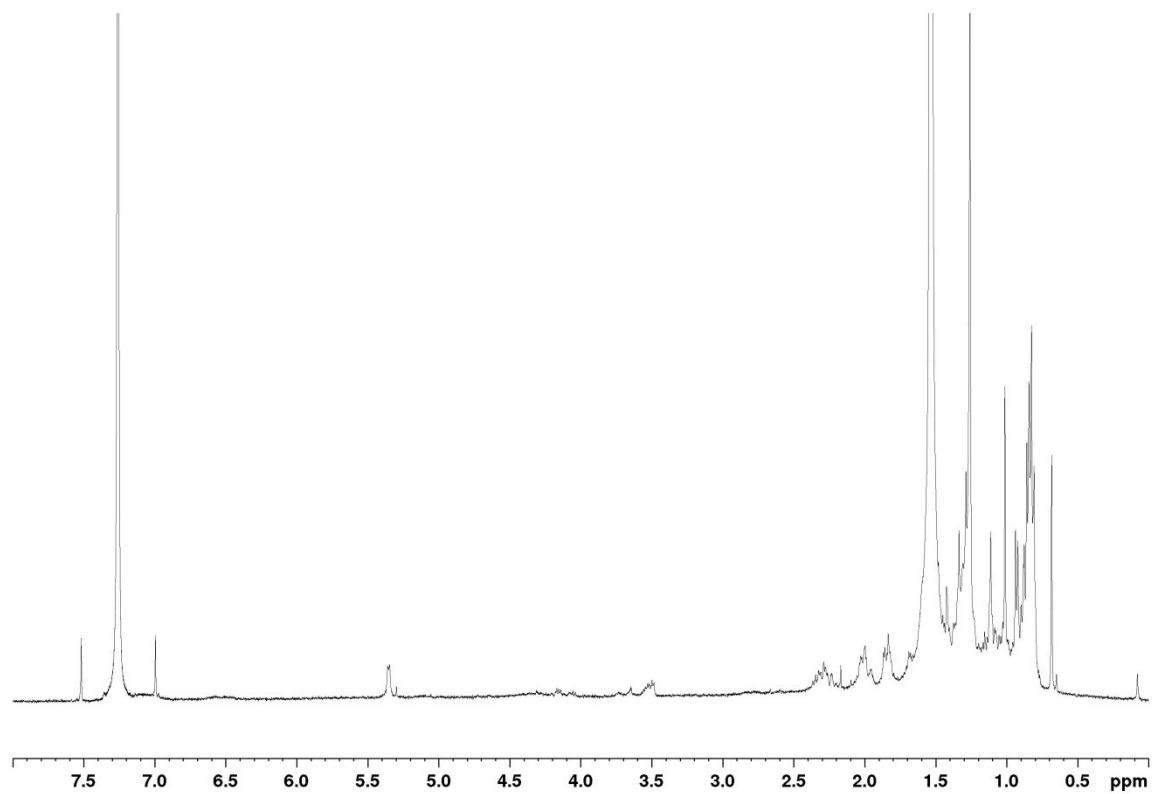
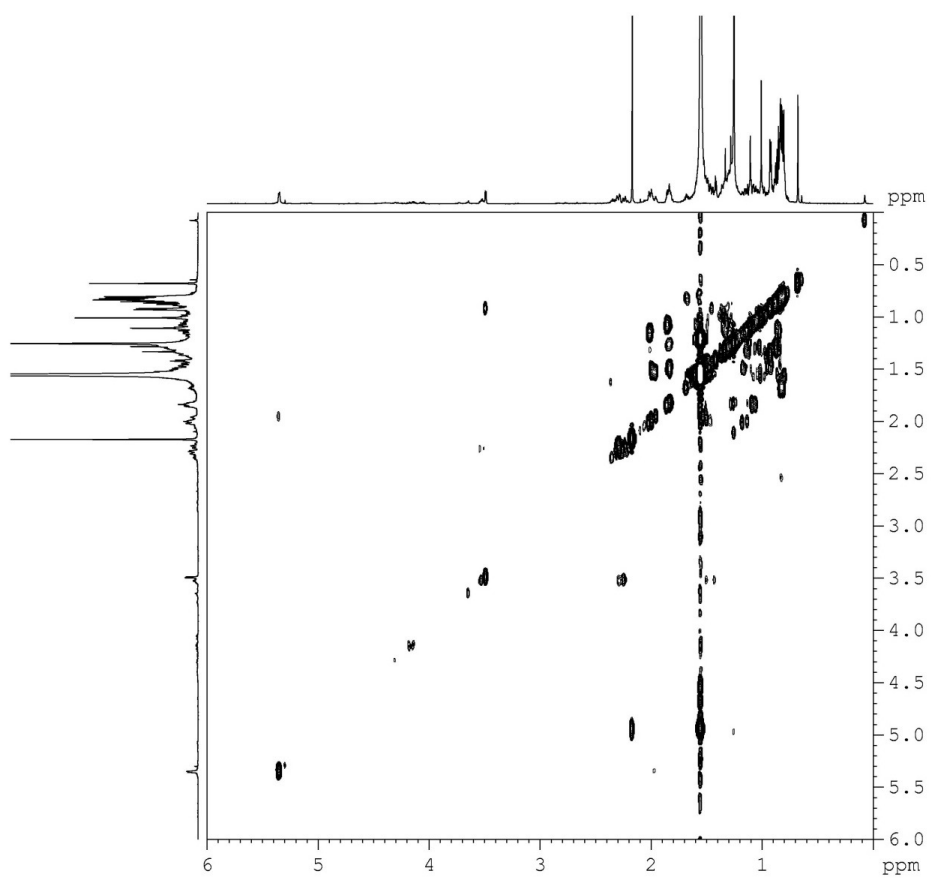


FIGURE SM5. COSY spectrum (600 MHz, CDCl₃) of clionasterol from *C. cryptica*.



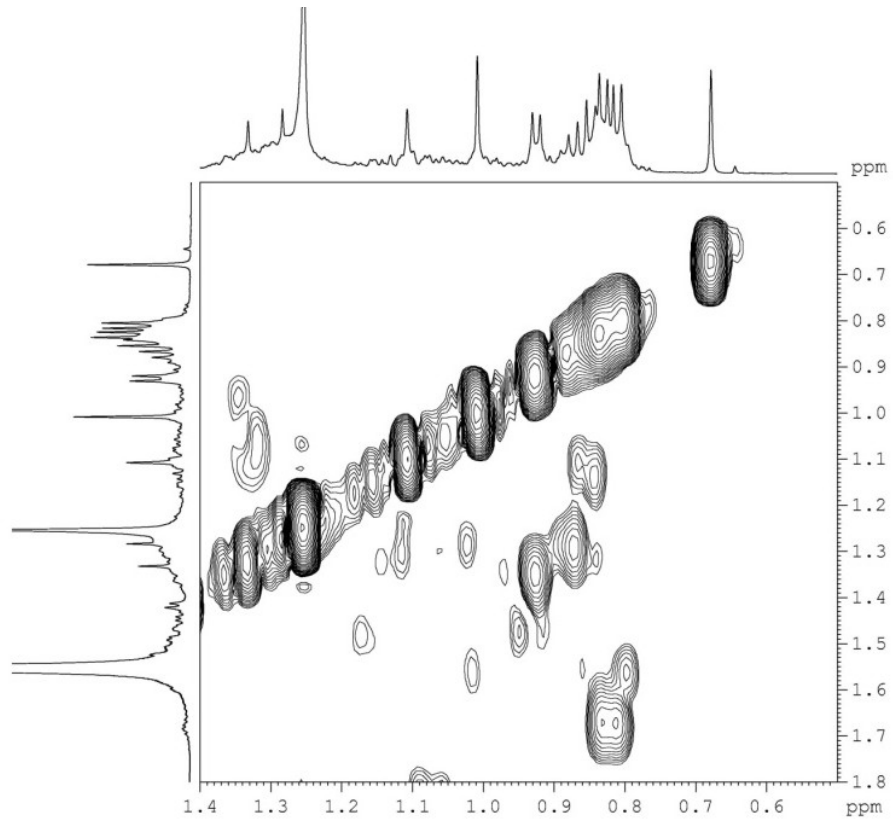


FIGURE SM6. HSQC spectrum (600 MHz, CDCl₃) of clonasterol from *C. cryptica*.

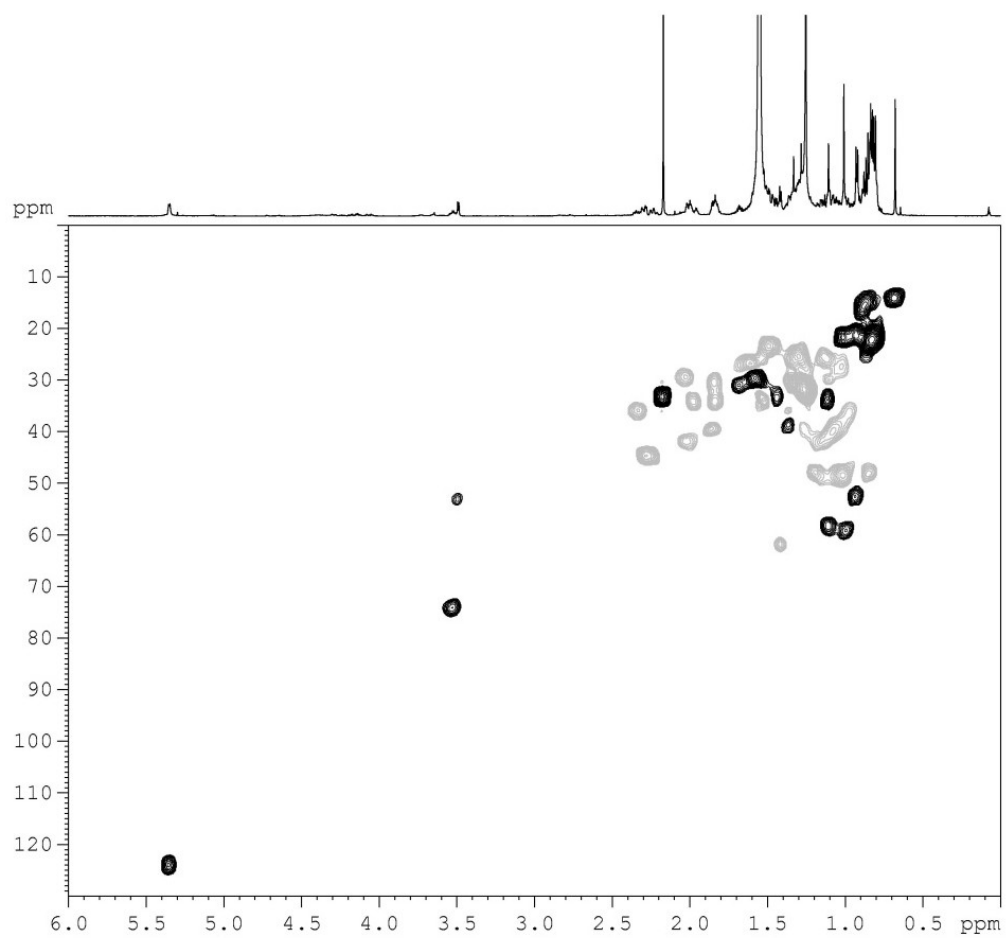


FIGURE SM7. ^1H NMR spectrum (600 MHz, C_6D_6) of natural fucosterol.

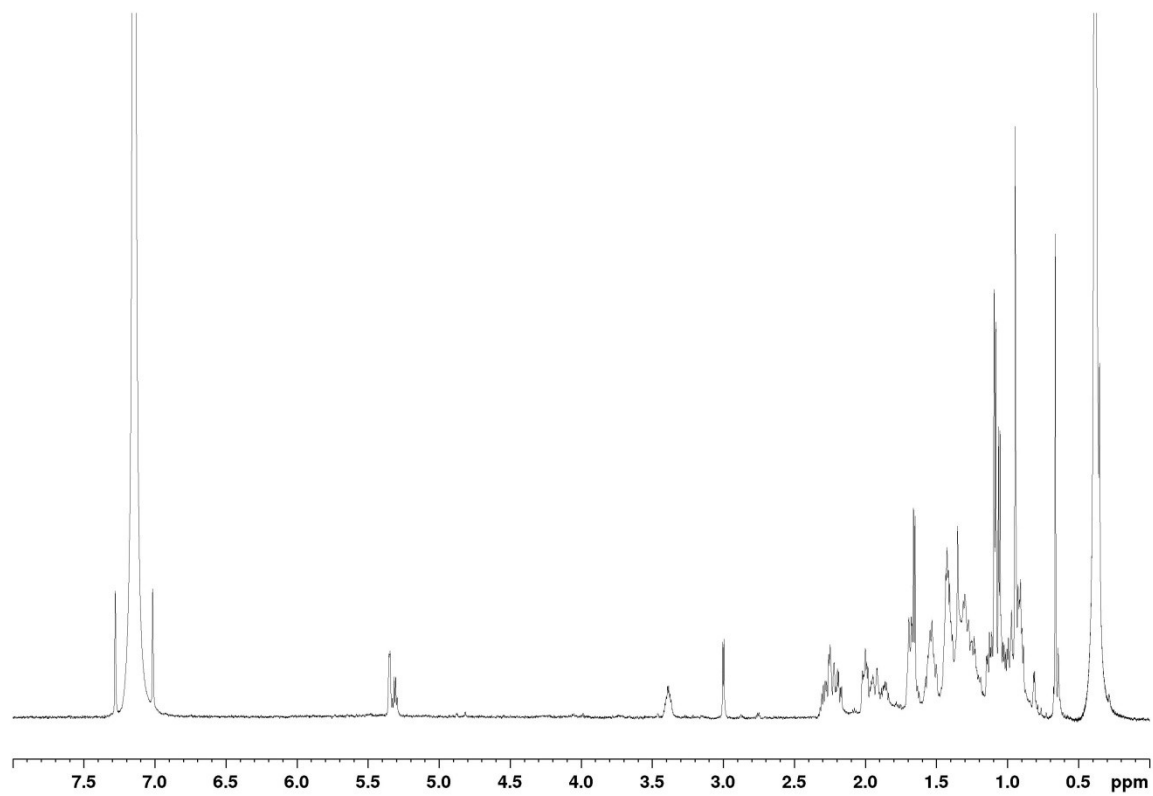


FIGURE SM8. ^1H NMR spectrum (600 MHz, CDCl_3) of acetyl brassicasterol from *P. tricornutum*.

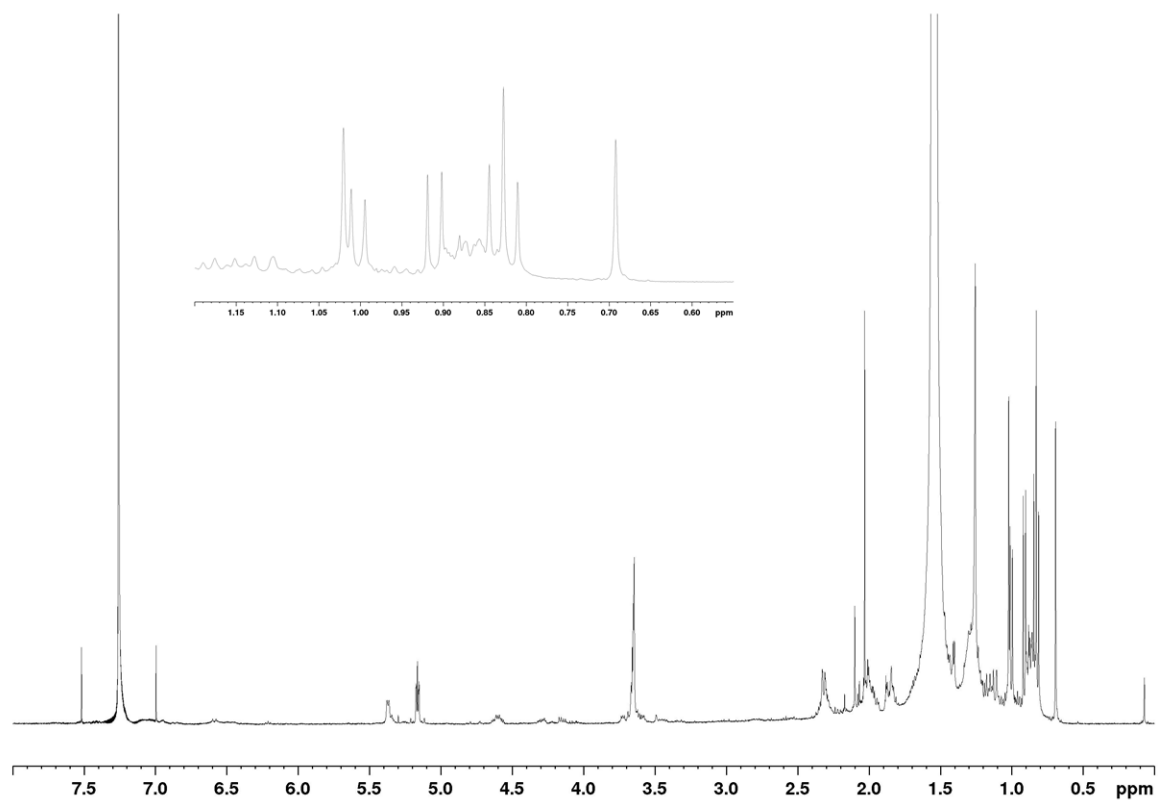


FIGURE SM9. COSY spectrum (600 MHz, CDCl_3) of acetyl brassicasterol from *P. tricornutum*.

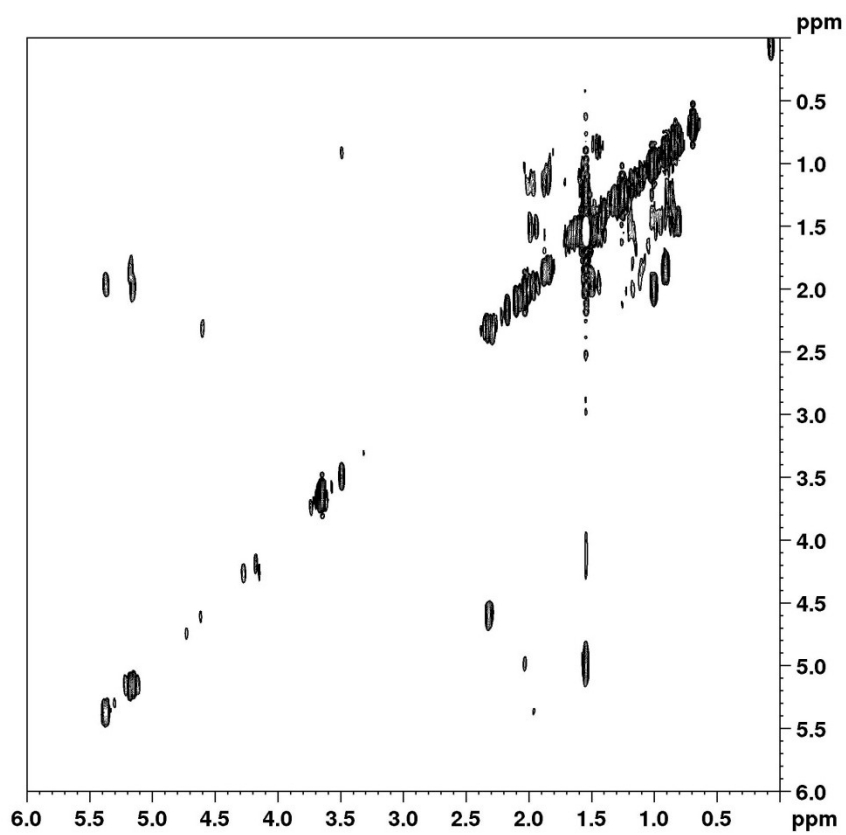


FIGURE SM10. Nodal sterol biosynthesis related translated transcripts comparison between *Skeletonema marinoi* (Queries) and *Cyclotella cryptica* transcriptome (Database).

- *Mevalonate Kinase: SM-MEVK (TR10489) obtains= CC-MEVK (TR35862)*



Query Coverage: 99.68%

Identities: 84.3%

Positives: 92.7%

- *Squalene synthase: SM-SQS (TR5837) obtains=CC-SQS (TR905)*



Query Coverage: 83.4%

Identities: 76.2%

Positives: 86.8%

- *Cycloartenol synthase: SM-CAS (TR12960) obtains=CC-CAS (TR3126)*



Query Coverage: 61.5%

Identities: 84.7%

Positives: 92.6%

Cycloartenol-C-24-methyltransferase: SM-SMT1 (TR612) obtains = CC-SMT1 (TR45147)



Query Coverage: 91.8%

Identities: 72%

Positives: 82.3%

- *Sterol methyl transferase: SM-SMT2 (TR11812) obtains= CC-SMT2(TR29102)*



Query Coverage: 84.6%

Identities: 72.7%

Positives: 78.5%

- *Sterol 14 demethylase: SM-CYP (TR7602) obtains= CC-CYP (TR23734)*

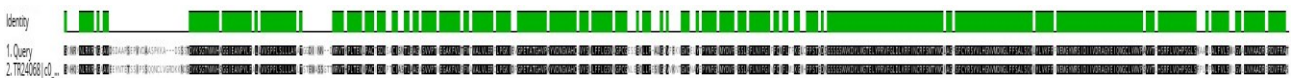


Query Coverage: 68.2%

Identities: 47%

Positives: 64.8%

- *7 dehydrocholesterol reductase: SM-DWF5 (TR2636) obtains = CC-DWF5 (TR24068)*



Query Coverage: 99.5%

Identities: 74%

Positives: 83.7%

- *Delta(24(24(1)))-sterol reductase: SM-24SR (TR213) obtains= CC-24SR (TR29411)*



Query Coverage: 100%

Identities: 75.5%

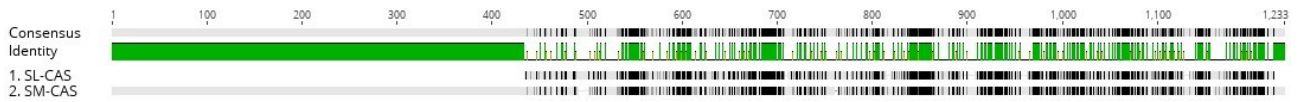
Positives: 85.2%

Alignments and outputs were performed using the Geneious software (Kearse et al., 2012). Blast investigation were performed using *S.marinoi* translated transcripts as queries; database was constructed using *C. cryptica* transcriptome. Reverse investigation were performed to confirm the results. This investigation revealed equal results (Data not showed).

Kearse M, Moir R, Wilson A, Stones-Havas S, Cheung M, Sturrock S et al. 2012. Geneious Basic: An integrated and extendable desktop software platform for the organization and analysis of sequence data. *Bioinformatics* 15;28 (12) 1647-9.

FIGURE SM11. Nodal sterol biosynthesis related translated transcripts alignments between *Skeletonema marinoi* and *Solanum lycopersicum*

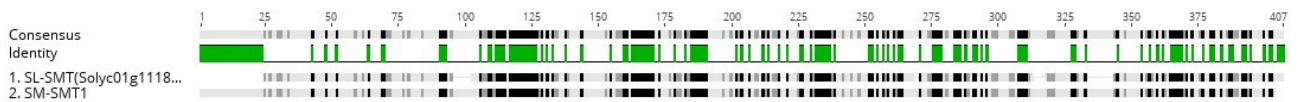
- *Cycloartenol synthase, SM-CAS (TR12960).*



Identities: 43.7%

Positives: 58.2%

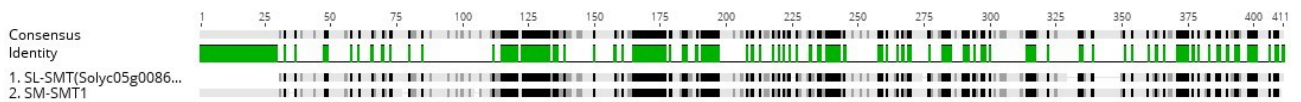
- *Cycloartenol-C-24-methyltransferase, SM-SMT1 (TR612)*



Identities: 32.4%

Positives: 50%

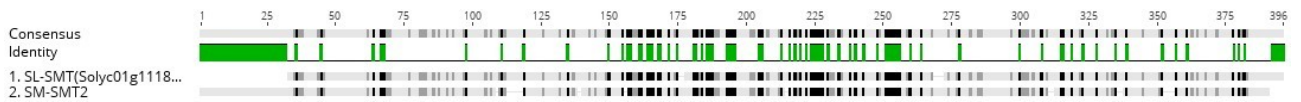
and



Identities: 34.2%

Positives: 50.4%

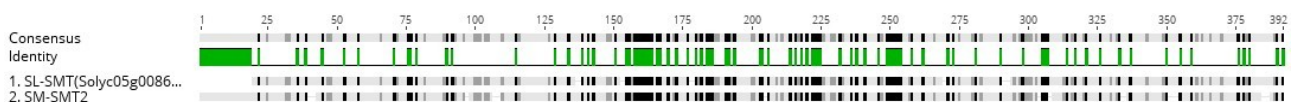
- *Sterol methyl transferase, SM-SMT2 (TR11812)*



Identities: 20.6%

Positives: 39%

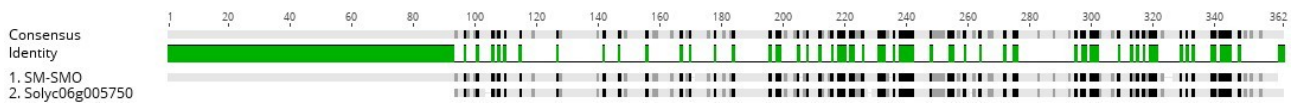
and



Identities: 23.3%

Positives: 39.7%

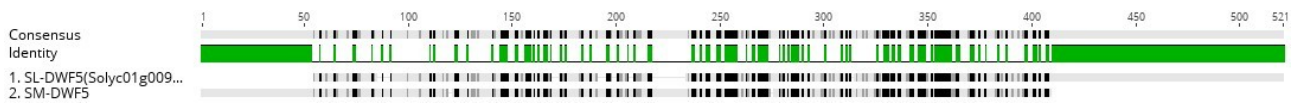
- *C-4 sterol methylsterol monooxygenase, SM-SMO(TR11964)*



Identities: 25%

Positives: 41%

- *7 dehydrocholesterol reductase, SM-DWF5 (TR2636)*

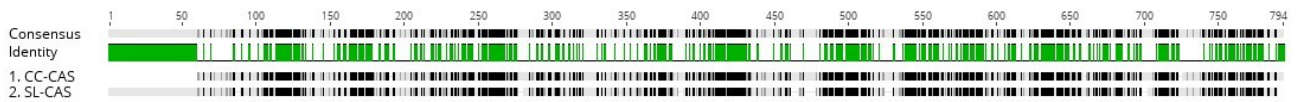


Identities: 31%

Positives: 47.3%

FIGURE SM12. Nodal sterol biosynthesis related translated transcripts alignments between *Cyclotella cryptica* and *Solanum lycopersicum*

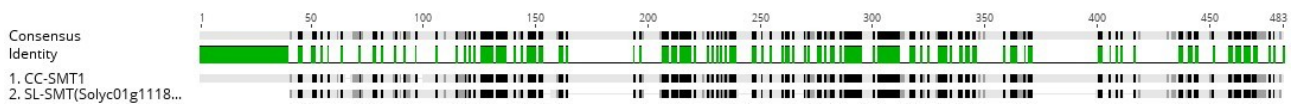
- *Cycloartenol synthase, CC-CAS (TR3126).*



Identities: 46%

Positives: 60.8%

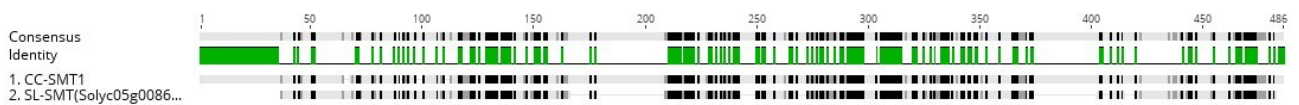
- *Cycloartenol-C-24-methyltransferase, CC-SMT1 (TR45147)*



Identities: 33%

Positives: 45.2%

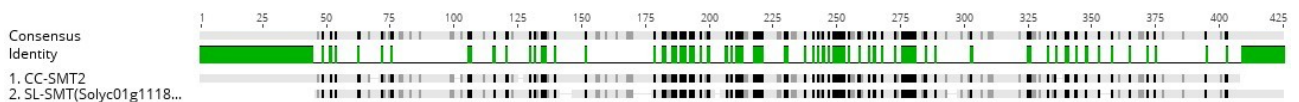
and



Identities: 33%

Positives: 45.9%

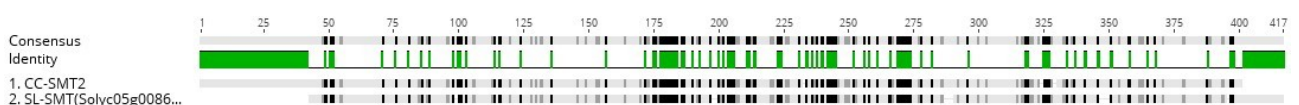
- *Sterol methyl transferase, CC-SMT2 (TR29102)*



Identities: 21.8%

Positives: 36.9%

and



Identities: 22.6%

Positives: 37.6%

- *7 dehydrocholesterol reductase, CC-DWF5 (TR24068)*



Identities: 35.5%

Positives: 52.6%

Alignments and outputs were performed using the Geneious software Alignments and outputs were performed using the Geneious software (Kearse et al., 2012)

Kearse M, Moir R, Wilson A, Stones-Havas S, Cheung M, Sturrock S et al. 2012. Geneious Basic: An integrated and extendable desktop software platform for the organization and analysis of sequence data. *Bioinformatics* 15;28 (12) 1647-9.