

Fig. S1.  $\beta$ -arrestin recruitment to ACKR3<sub>WT</sub> induced by CCX662 and CXCL11 in BRET experiments. (A) Potency of activation relative to CXCL12 was calculated from  $\Delta pEC_{50}$ =  $pEC_{50,CXCL12}$ - $pEC_{50,CXCL12}$ - $pEC_{50,CXCL12}$ - $pEC_{50,CXCL12}$ . (B) Efficacy of CCX662 and CXCL11 normalized to CXCL12. Bars show the average and standard errors from five independent experiments. Significant differences for CCX662 and CXCL11 compared to CXCL12 are noted: \*P<0.05, \*\*P<0.01, \*\*\*P<0.001 from one-way ANOVA with Dunnett's multiple comparison test.



**Fig. S2. Expression of ACKR3 variants in HEK cells. (A)** Total expression in HEK293T cells determined by luminescence. Data were normalized to ACKR3<sub>WT</sub> expression from the same experiment according to Lum<sub>mutant</sub>/Lum<sub>WT</sub> x 100. (**B**) Surface expression of ACKR3 in HEK293T cells quantified from anti-HA antibody staining. (**C**) Same samples as in (A) stained with an anti-ACKR3 antibody. Surface expression of ACKR3<sub>d17</sub> is not detectable with the anti-ACKR3 antibody since the truncation removes the epitope for antibody binding. (**D**) Constitutive association of GFP10-β-arrestin-2 in HEK293T cells to Rluc-fused receptors. Data in are means ± SEM of three or more experiments.



**Fig. S3. Kinetics of association of CXCL12 with ACKR3 on live** *Sf9* **cells.** (**A**) Representative example of association curve in the absence (total binding) and presence (non-specific binding) of the small molecule agonist CCX777 determined from the geometric mean fluorescent intensity (GMFI) of FITC-conjugated anti-HA antibody bound to CXCL12-HA. (**B**) Specific binding obtained by subtracting non-specific from total binding in (A). The fit of the association data is significantly improved with the use of a two-phase exponential function (solid line) compared to a single-phase exponential (dotted line). Data are representative of five experiments.



Fig. S4. SPR in nanodiscs and detergent micelles. (A) Analysis of CXCL12 binding for ACKR3 in nanodiscs. Plot of  $k_{obs}$  determined from fitting the association phase of the SPR curves at different concentrations of chemokine using a shared value for the dissociation rate constant (k<sub>d</sub>). Linear regression indicates that the data fits to a straight line with a correlation coefficient (R<sup>2</sup>) of 0.997. (B) Thermostability of ACKR3 in complex with biotinylated CXCL12 determined from CPM assays.  $\Delta T_m$  was determined as  $T_{m,ACKR3:CXCL12-biotin}$ - $T_{m,ACKR3:CXCL12}$ . Bars show the average and standard error of three measurements. (C) SPR curves for the binding of ACKR3 in DDM/CHS detergent micelles to immobilized CXCL12-biotin at different concentrations of receptor. (D and E) Association (k<sub>a</sub>) and dissociation (k<sub>d</sub>) constants determined from fitting SPR data in micelles and nanodiscs from three separate experiments.



Fig. S5. N-terminal cytochrome b562-RIL fusion protein did not affect CXCL12 dissociation rate or folding of ACKR3. (A) Dissociative half-life for CXCL12 with ACKR3<sub>WT</sub> and bril-ACKR3<sub>WT</sub> in live *Sf9* cells. Bars correspond to averages and standard errors from n=4 (ACKR3<sub>WT</sub>) or n=8 (bril-ACKR3<sub>WT</sub>). (B) SDS-PAGE of ACKR3<sub>WT</sub> and bril-ACKR3<sub>WT</sub> purified from *Sf9* cells. (C) Thermostability of purified ACKR3<sub>WT</sub>:CXCL12<sub>WT</sub> and bril-ACKR3<sub>WT</sub>:CXCL12<sub>WT</sub> in detergent micelles measured by CPM fluorescence. Peaks at 61°C correspond to the midpoints of unfolding (Tm) values. (D) Mean and standard errors of three T<sub>m</sub> measurements for ACKR3<sub>WT</sub> and bril-ACKR3<sub>WT</sub>.



**Fig. S6. BRET signal increase after addition of 2µM chemokine for ACKR3**<sub>wT</sub> and **ACKR3**<sub>d29</sub>. % ligand-induced BRET was calculated as a percentage of CXCL12-induced BRET with ACKR3<sub>wT according to (BRET<sub>2uM ligand</sub>-BRET<sub>no ligand</sub>)/ (BRET<sub>ACKR3-WT,2uM CXCL12</sub>-BRET<sub>ACKR3-WTno ligand</sub>) x 100. Data are means ± SEM of three experiments.</sub>

	Ref	Author	CXCR4	ACKR3
1	(33)	Hanes	2.1	0.0014
2	(47)	Hoffmann	23.8	2.6
3	(35)	Szpakowska	12.9	1.8
4	(38)	Montpas		5.9
5	(34)	Benredjem		3.0
6	(48)	Wijtmans		0.63
7	(49)	Canals		0.2
8	(50)	Zabel		0.2
9	(50)	Zabel		0.07
10	(50)	Zabel		0.1
11	(4)	Burns		0.2
12	(5)	Rajagopal		0.2
13	(51)	Balabanian		0.8
	Average 1-3:		12.9	1.47
Average all:				1.21

Table S1. IC<sub>50</sub> values (nM) for equilibrium binding studies reported in literature. Ref, reference; first author listed.