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Supplemental Information

LINC00680 and TTN-AS1 Stabilized by EIF4A3

Promoted Malignant Biological Behaviors

of Glioblastoma Cells

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(1) binding site name	HPAM7_26664						
Target Location	chr6:58273119-58273139[-]						
RBP Name	elF4All						
Target Name	LINCODESO						
Target Transcripts	ENST00000418765 ENST00000450081 ENST00000418368 ENST00000399751 ENST00000422882						
ClipSeq ReadNum	1000						
(2) binding site name	HPAM1_44003						
Target Location	chr6:58273121-58273136[-]						
R8P Name	eF4AII						
Target Name	LINCODERD						
Target Transcripts	ENST00000418765 ENST00000450081 ENST00000418368 ENST00000399751 ENST00000422682						

nding Score 0.083

GUCGAAA

GGGUCGAAAA

C GGAG GU A G A UGG

GAG GUU

1760-1778

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 ENITODO00389134

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 Cripties Resolver
 ENITOD000197264
 ENITOD000389134



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Position on chromosom Conserved Species: Biod

	Predicted consequential pairing of target region (top) and miRNA (bottom)	Site type
Position 2320-2327 of EGR3 3' UTR	5' AUUUGUAGGGGUAUACAGGUUUA 	8mer
Position 2320-2327 of EGR3 3' UTR hsa-miR-320b	5'AUUUGUAGGCGUAUAAGCUUUA IIIIII 3' AACGGAGAGUUGGGUCGAAAA	8mer
Position 2320-2327 of EGR3 3' UTR hsa-miR-320c	5* AUUUGUAGGCGUAUACAGCUUUA 3* UGGGAGAGUUGGGUCGAAAA	8mer
Position 2320-2327 of EGR3 3' UTR hsa-miR-4429	5' AUUUGUAGGCGUAUACAGCUUUA 3' GCGGAGAGUCGGGUCGAAAA	8mer
Position 2320-2327 of EGR3 3' UTR hsa-miR-320a	5'AUUUGUAGGGUAUACAGCUUUA 3' AGCDGGAGAGUUGGGUCGAAAA	8mer

















Supplemental figure 1. Results of the in-silico experiments and expression and

survival of EIF4A3 and EGR3 from databases

A. Putative binding sites of EIF4A3 and LINC00680 from Database: Starbase V2.0

(http://starbase.sysu.edu.cn/starbase2/). B. Putative binding sites of EIF4A3 and TTN-AS1 from Database: Starbase V2.0 (http://starbase.sysu.edu.cn/starbase2/).C. Putative binding sites of LINC00680 and miR-320b from Database: LncBase (http://diana.imis.athena-innovation.gr/DianaTools/index.php?r=lncBase/index). D. Putative binding sites of TTN-AS1 and miR-320b from Database: LncBase (http://diana.imis.athena-innovation.gr/DianaTools/index.php?r=lncBase/index). E. Putative binding sites of miR-320b and EGR3 mRNA from Database: TargetScan (http://www.targetscan.org/vert 72/). F. Putative binding sites of EGR3 and PKP2 promotor from Database: Jaspar (http://jaspar.genereg.net/). G. Expression of EIF4A3 from database Database: Oncomine (https://www.oncomine.org/resource/main.html), ###p<0.001 versus normal brain tissue. H. Expression of EGR3 from database Database: Oncomine, ###p<0.001 versus normal brain tissue. I. Survival data of EIF4A3 from Database: GEPIA (http://gepia.cancer-pku.cn/detail.php). J. Survival data of EGR3 from Database: GEPIA.

Supplemental figure 2. Results of experiments using BTICs

A. GFAP staining by Immunofluorescence. Secondary antibody was labeled with FITC. Green fluorescence demonstrated GFAP in BTICs. B. Expression of EIF4A3 determined by western blot. **p < 0.01 versus normal human astrocytes (HA). C. Expression of LINC00680 and TTN-AS1 determined by PCR (n=3). ##p < 0.01 versus HA. D. CCK-8 assay was conducted to investigate the effect of EIF4A3, LINC00680 and TTN-AS1 on proliferation in BTICs. *p < 0.05 versus sh-NC group (empty vector);

**p < 0.01 versus sh-NC group; #p < 0.05 versus sh-EIF4A3 group; p < 0.05 versus sh-LINCC0680 group, &p < 0.05 versus sh-TTN-AS1 group. E. Flow cytometry analysis of EIF4A3, LINC00680 and TTN-AS1 knockdown in BTICs. *p < 0.05 versus sh-NC group (empty vector); **p < 0.01 versus sh-NC group; #p < 0.05 versus sh-EIF4A3 group; \$p < 0.05 versus sh-LINCC0680 group, &p < 0.05 versus sh-TTN-AS1 group. F. Transwell assays were used to measure the effect of EIF4A3, LINC00680 and TTN-AS1 on cell migration and invasion in BTICs. Representative images and statistical plots were presented. *p < 0.05 versus sh-NC group (empty vector); **p < 0.050.01 versus sh-NC group; #p < 0.05 versus sh-EIF4A3 group; p < 0.05 versus sh-LINCC0680 group, &p < 0.05 versus sh-TTN-AS1 group. Scale bars represent 80 µm. G. Expression of LINC00680(left) and TTN-AS1(right) were detected in BTICs with EIF4A3 knockdown by PCR. *p < 0.05 versus sh-NC group. H. CCK-8 assay was conducted to investigate the effect of miR-320b on proliferation in BTICs. *p < 0.05versus pre-NC group, #p<0.05 versus anti-NC group. I. Flow cytometry analysis effect of miR-320b on apoptosis in BTICs. *p < 0.05 versus pre-NC group, #p<0.05 versus anti-NC group. J. Transwell assays were used to measure the effect of miR-320b on cell migration and invasion in BTICs. Representative images and statistical plots were presented. *p < 0.05 versus pre-NC group, #p<0.05 versus anti-NC group. Scale bars represent 80 μ m. Data are presented as the mean \pm SD (n = 3 in each group).

Supplemental figure 3. Results of experiments using BTICs

A. Real-time PCR analysis for EIF4A3, LINC00680 and TTN-AS1 regulating miR320b expression in BTICs. *p < 0.05 versus sh-NC group; **p < 0.01 versus sh-NC

group; #p < 0.05 versus sh-EIF4A3 group; \$p < 0.05 versus sh-LINCC0680 group, &p < 0.05 versus sh-TTN-AS1 group. B. Real-time PCR analysis for miR-320b modulating LINC00680 and TTN-AS1 expression in BTICs. *p < 0.05 versus pre-NC group, #p<0.05 versus anti-NC group. C. CCK-8 assay was conducted to investigate the effect of LINC00680/TTN-AS1 and miR-320b on proliferation in BTICs. **p < 0.01 versus sh-NC+ pre-NC group. D. Flow cytometry analysis of LINC00680/TTN-AS1 and miR-320b in BTICs. **p < 0.01 versus sh-NC+ pre-NC group. E. Transwell assays were used to measure the effect of LINC00680/TTN-AS1 and miR-320b on cell migration and invasion in BTICs. Representative images and statistical plots were presented. **p < 0.01 versus sh-NC+ pre-NC group. Scale bars represent 80 µm. Data are presented as the mean \pm SD (n = 3). F. Western blot was used to detect expression of EGR3 in BTICs. G. Expression of EGR3 mRNA in BTICs was detected by Real-time PCR. H. CCK-8 assay was conducted to investigate the effect of EGR3 on proliferation in BTICs. *p < 0.05 versus sh-NC group. I. Flow cytometry analysis of EGR3 in BTICs. *p < 0.05versus sh-NC group. J. Transwell assays were used to measure the effect of EGR3 on cell migration and invasion in BTICs. Representative images and statistical plots were presented. Data are presented as the mean \pm SD (n = 3 in each group). *p < 0.05 versus sh-NC group. Scale bars represent 80 μ m. Data are presented as the mean \pm SD (n = 3). K. CCK-8 assays were performed in BTICs with the altered expression of miR-320b and EGR3. *p < 0.01 versus anti-NC group; #p < 0.05 versus anti-miR-320b group. L. Flow cytometry analysis of BTICs with the altered expression of miR-320b and EGR3. *p < 0.05 versus anti-NC group; #p < 0.05 versus anti-miR-320b group. M.

Quantification number of migration and invasion cells treated with anti-miR-320b and sh-EGR3. Representative images and statistical plots were presented. Scale bars represent 80 μ m. *p < 0.05 versus anti-NC group; #p < 0.05 versus anti-miR-320b group. Data are presented as the mean \pm SD (n = 3).

Supplemental figure 4. Results of experiments using BTICs

A. Expression of EGR3 was regulated in BTICs lines with knockdown of EIF4A3 and/or LINC00680/TTN-AS1. *p < 0.05 versus sh-NC group; **p < 0.01 versus sh-NC group; #p < 0.05 versus sh-EIF4A3 group; \$p < 0.05 versus sh-LINCC0680 group, &p < 0.05 versus sh-TTN-AS1 group. B. Expression of EGR3 was regulated in BTICs lines with knockdown or over-expression of miR-320b. **p < 0.01 versus pre-NC group, ##p<0.01 versus anti-NC group. C. Expression of EGR3 was regulated by LINC00680/TTN-AS1 and miR-320b. **p < 0.01 versus sh-NC+ pre-NC group. Data are presented as the mean \pm SD (n = 3). D. Western blot analysis of PKP2 in BTICs knockdown of LINC00680 or TTN-AS1. Data are presented as the mean \pm SD (n = 3 in each group). *p < 0.05 versus sh-NC group. E.Western blot analysis of PKP2 in BTICs regulated by LINC00680/TTN-AS1 and miR-320b. **p < 0.01 versus sh-NC+ pre-NC group. F. Western blot analysis of PKP2 in BTICs regulated by miR-320b. **p < 0.01 versus pre-NC group, ##p<0.01 versus anti-NC group. G.Western blot analysis of PKP2 in BTICs regulated by miR-320b and EGR3. **p < 0.01 versus anti-NC group; #p < 0.05 versus anti-miR-320b group. H. Western blot analysis of PKP2 in BTICs regulated by EGR3. **p < 0.01 versus sh-NC group. Data are presented as the mean \pm SD (n = 3).

EIF4A3	Forward primer 5'-GATGCCGATGAACGTTGCTG-3'				
	Reverse primer	5'-GGTGGTGGCACCTTAGAAGTAT-3'			
LINC00680	Forward primer	5'-CCATCGACTGGCTCATCACAA-3'			
	Reverse primer	5'-GGGGCAAGGCAAATCAATACC -3'			
TTN-AS1	Forward primer	5'-ACACTCATCATCCAGCCTGC-3'			
	Reverse primer	5'-TGGCCATCATTGCAAGCTAAC-3'			
miR-320b	Forward primer	5'-AATTAATCCCTCTCTTTCTAGTTCT-3'			
	Reverse primer	5'-AGTTAATTTGTTTGCCCTCTCAA-3'			
EGR3	Forward primer	5'-GGTGACCATGAGCAGTTTGC-3'			
	Reverse primer	5'-TAGGTCACGGTCTTGTTGCC-3'			

Table S1. Primers used for RT-qPCR.

Table S2. Sequences for sh-RNA, agomir and antagomir

sh-EIF4A3	5'-GGATATTCAGGTTCGTGAAAC-3'
sh-LINC00680	5'-GCACTTGAAGAGAGGTTTACC-3'
sh-TTN-AS1	5'-GCAATGATGGGCCACTAAATTG-3'
sh-EGR3	5'-CTAAACCAACTGCCTGACAAT-3'
miR-320b agomir	5'- AAAGCUGGGUUGAGAGGG CAA -3'
miR-320b antagomir	5'- UUGCCCUCUCAACCCAGCUUUU-3'

Table S3. Primers used for ChIP experiments

Binding			Produ	Annealing
site or		Sequence	ct size	temperature
Control			(bp)	(°C)
PCR1	Forward primer	5'-GTGACTCTCGAGGGCTCACC-3'	192	60.8
	Reverse primer	5'-CGGCCTTCCCTGAGGAGG-3'		
PCR2	Forward primer	5'-GCAGTGGCTCACGCCTGTAATC-3'	261	61.2
	Reverse primer	5'-GAGTGCAGTGGTGTGATCTCAGC-3'		