

Supplementary material for *Spruce sugars and poultry hydrolysate as growth medium in repeated fed-batch fermentation processes for production of yeast biomass*

Spruce sugars and poultry hydrolysate as growth medium in repeated fed-batch fermentation processes for production of yeast biomass

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Supplementary material

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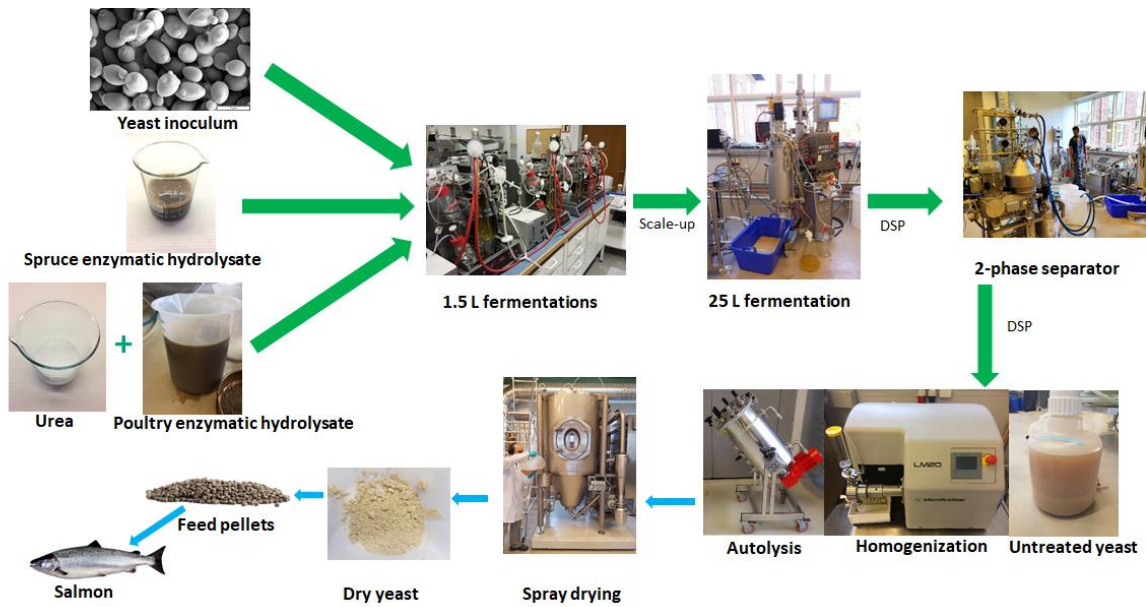


Figure S1. Process flow of study. Green arrows indicate processing steps included in this study. Blue arrows indicate planned further processing steps.

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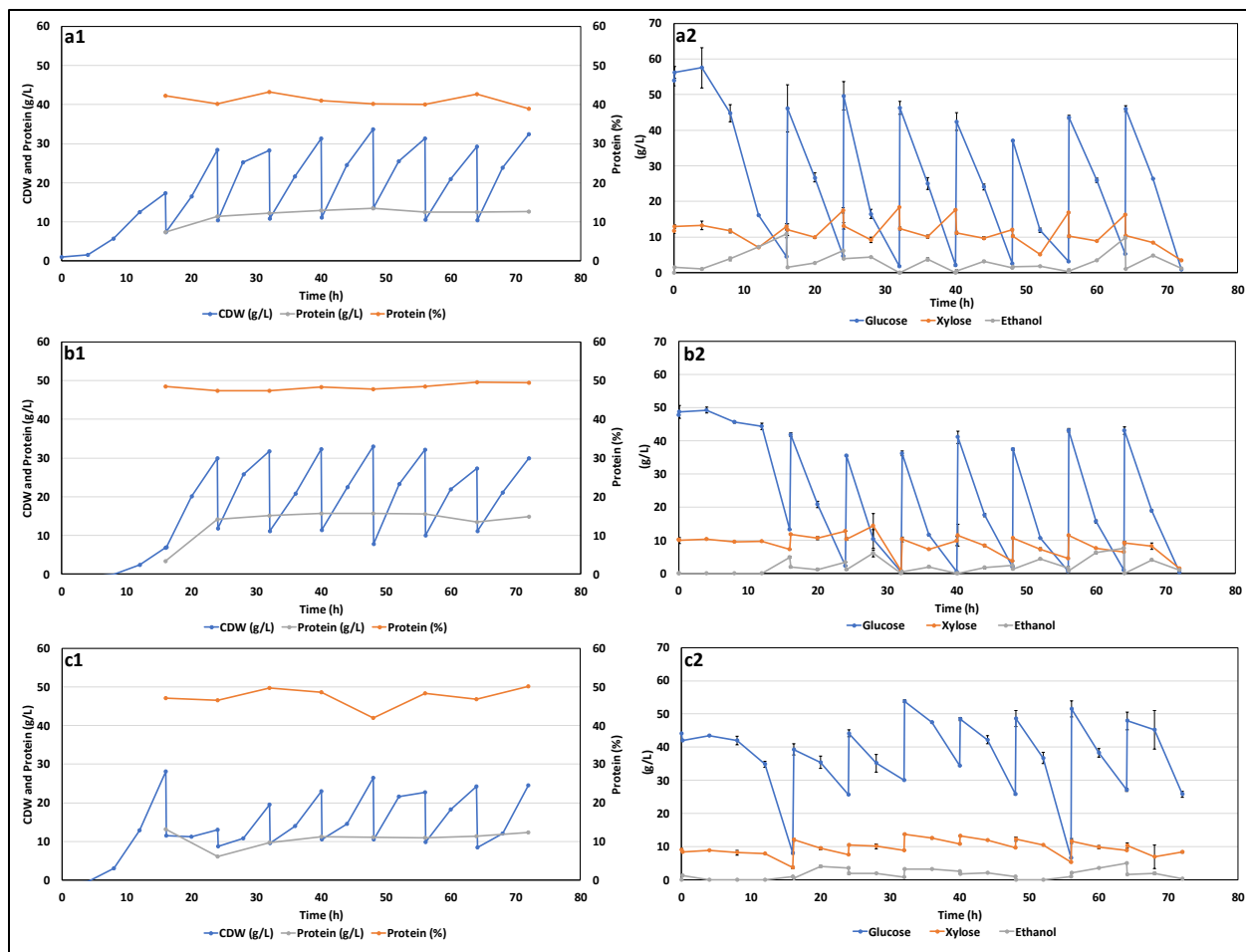


Figure S2. Repeated fed-batch cultivation of three yeast strains grown on 100% poultry hydrolysates and BALI™ hydrolysates in a 2.5 L fermenter. The starting volume was 1.5 L and the fermentation lasted 72 h. Panels labeled 1 show accumulation of cells (blue lines) and protein (grey lines), as well as the protein content of the cells (orange lines); panels labeled 2 show glucose (blue lines), xylose (orange lines) and ethanol (grey lines). a, *C. jadinii*; b, *W. anomalus*; c, *B. adenivorans*. Growth was monitored by measuring the CDW (g/L, blue lines) every 4 h. For the samples taken from 16 h and onwards, the protein content (orange lines) of the dried cells was measured using the Kjeldahl method (w/w %). The concentration of yeast protein (g/L; grey lines) was calculated by multiplying CDW (g/L; blue lines) with the protein content (w/w %). Acetic acid and lactic acid production were negligible for all yeasts (results not shown).

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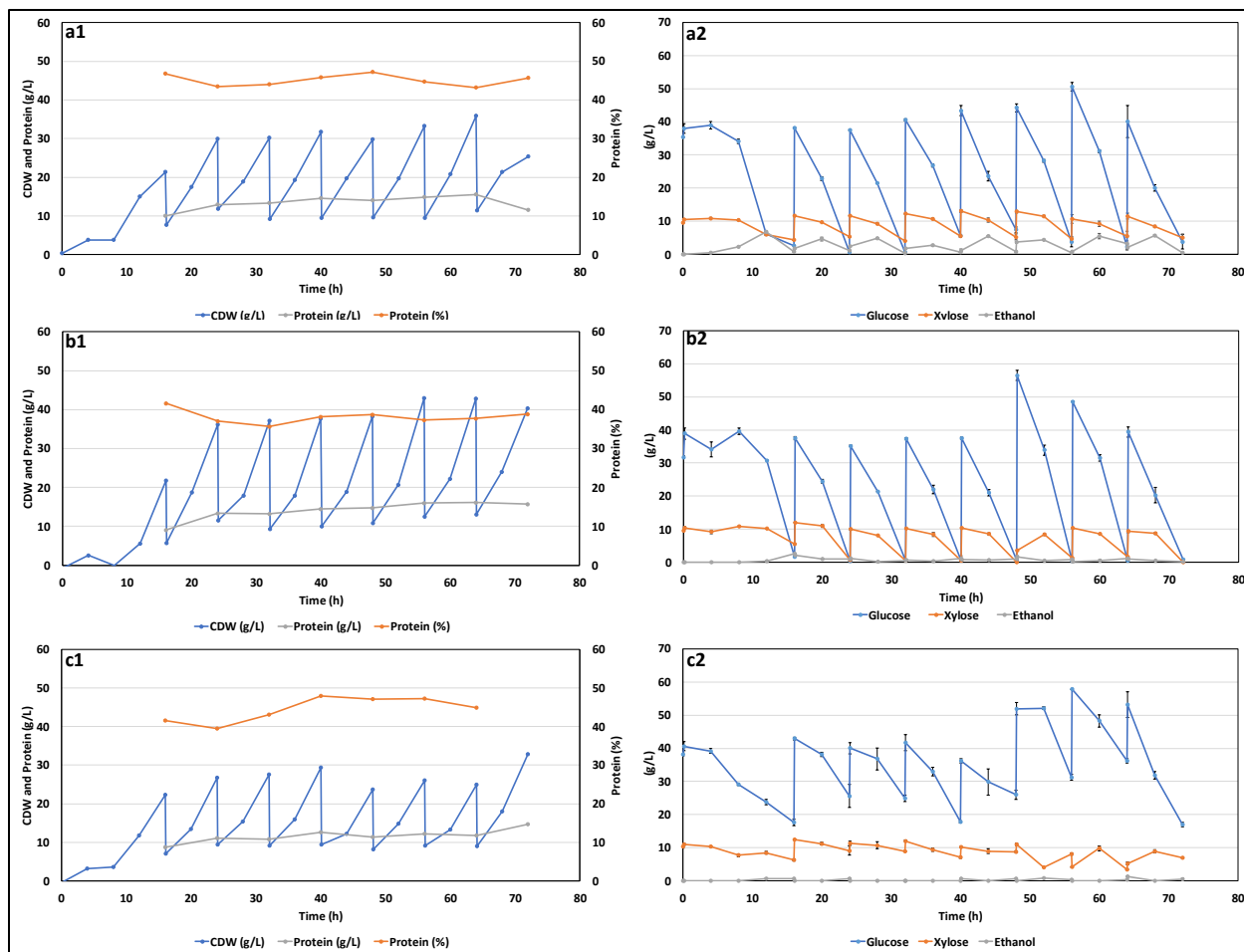


Figure S3. Repeated fed-batch cultivation of three yeast strains grown on 40% poultry hydrolysates and 60% urea with BALI™ hydrolysates in a 2.5 L fermenter. The starting volume was 1.5 L and the fermentation lasted 72 h. Panels labeled 1 show accumulation of cells (blue lines) and protein (grey lines), as well as the protein content of the cells (orange lines); panels labeled 2 show glucose (blue lines), xylose (orange lines) and ethanol (grey lines). a, *C. jadinii*; b, *W. anomalus*; c, *B. adenivorans*. Growth was monitored by measuring the CDW (g/L, blue lines) every 4 h. For the samples taken from 16 h and onwards, the protein content (orange lines) of the dried cells was measured using the Kjeldahl method (w/w %). The concentration of yeast protein (g/L; grey lines) was calculated by multiplying CDW (g/L; blue lines) with the protein content (w/w %). Acetic acid and lactic acid production were negligible for all yeasts (results not shown).

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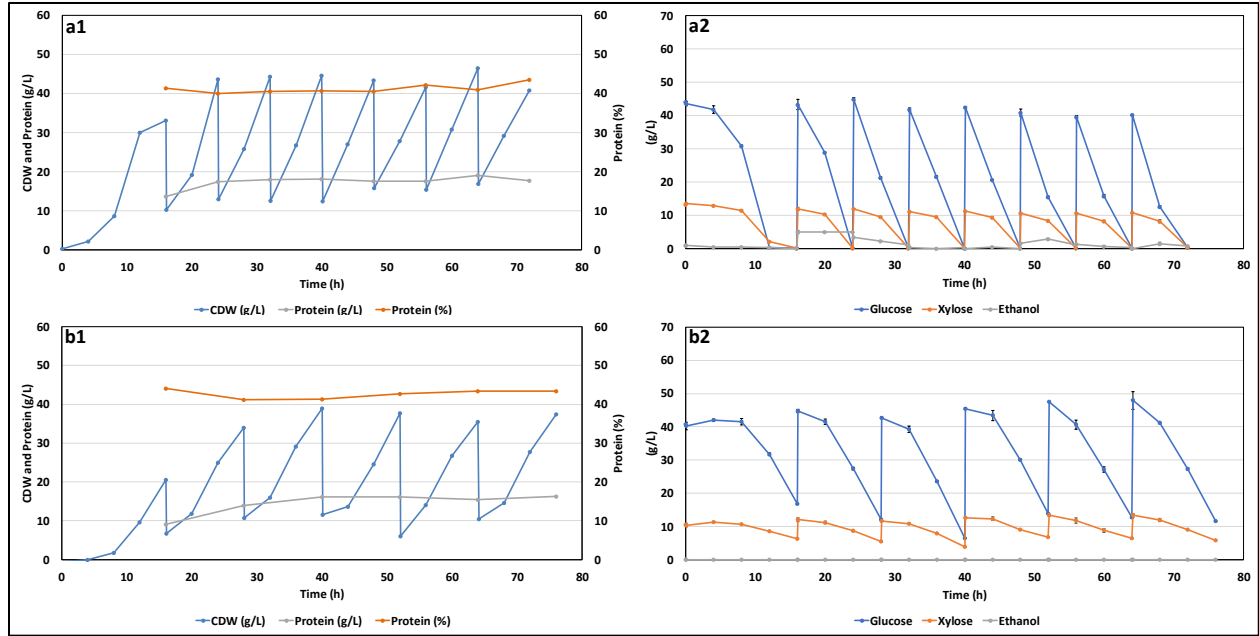


Figure S4. Repeated fed-batch cultivation of two yeast strains grown on 60% poultry hydrolysates with 40% urea and BALI™ hydrolysates in a 2.5 L fermenter. The starting volume was 1.5 L and the fermentation lasted 72 h and 76 h, respectively. Panels labeled 1 show accumulation of cells (blue lines) and protein (grey lines), as well as the protein content of the cells (orange lines); panels labeled 2 show glucose (blue lines), xylose (orange lines) and ethanol (grey lines). a, *W. anomalus*; b, *B. adenivorans*. Growth was monitored by measuring the CDW (g/L, blue lines) every 4 h. For the samples taken from 16 h and onwards, the protein content (orange lines) of the dried cells was measured using the Kjeldahl method (w/w %). The concentration of yeast protein (g/L; grey lines) was calculated by multiplying CDW (g/L; blue lines) with the protein content (w/w %). Acetic acid and lactic acid production were negligible for both yeasts (results not shown).

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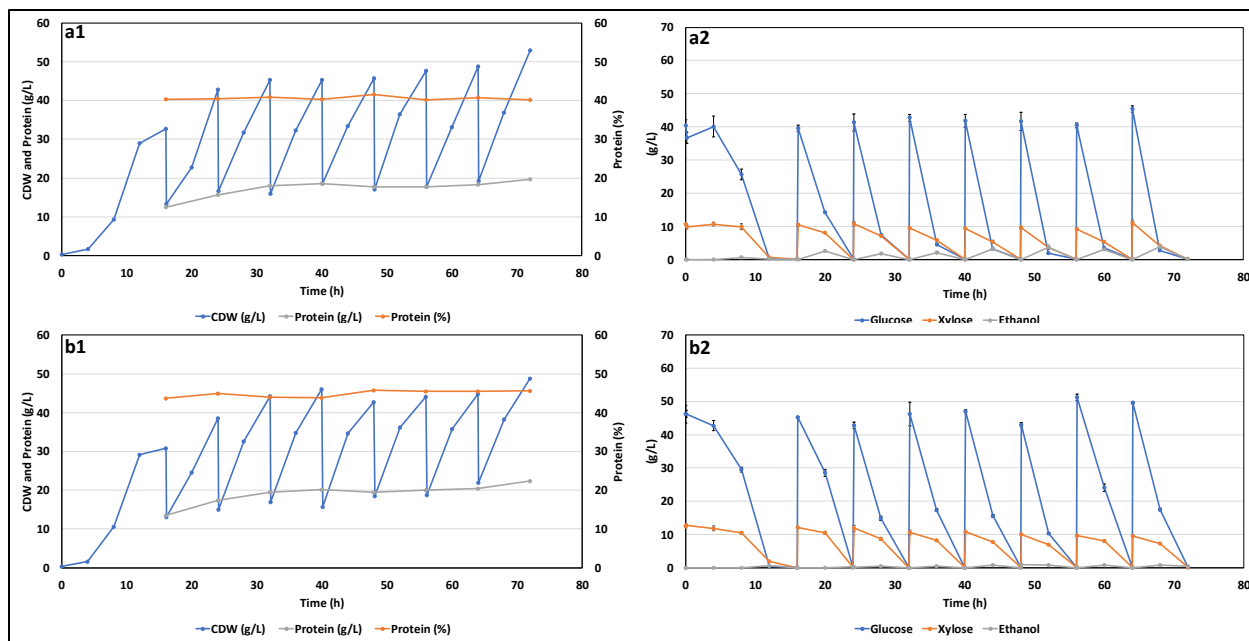


Figure S5. Repeated fed-batch cultivation of *W. anomalus* grown on 60% poultry hydrolysates, 40% urea and biotin or 80% poultry hydrolysates, 20% urea with BALI™ hydrolysates in a 2.5 L fermenter. The starting volume was 1.5 L and the fermentation lasted 72 h. a, 60% poultry hydrolysates, 40% urea and biotin b, 80% poultry hydrolysates, 20% urea. Panels labeled 1 show accumulation of cells (blue lines) and protein (grey lines), as well as the protein content of the cells (orange lines); panels labeled 2 show glucose (blue lines), xylose (orange lines) and ethanol (grey lines). Growth was monitored by measuring the CDW (g/L, blue lines) every 8 h. For the samples taken from 16 h and onwards, the protein content (orange lines) of the dried cells was measured using the Kjeldahl method (w/w %). The concentration of yeast protein (g/L; grey lines) was calculated by multiplying CDW (g/L; blue lines) with the protein content (w/w %). Acetic acid and lactic acid production were negligible on both media (results not shown).

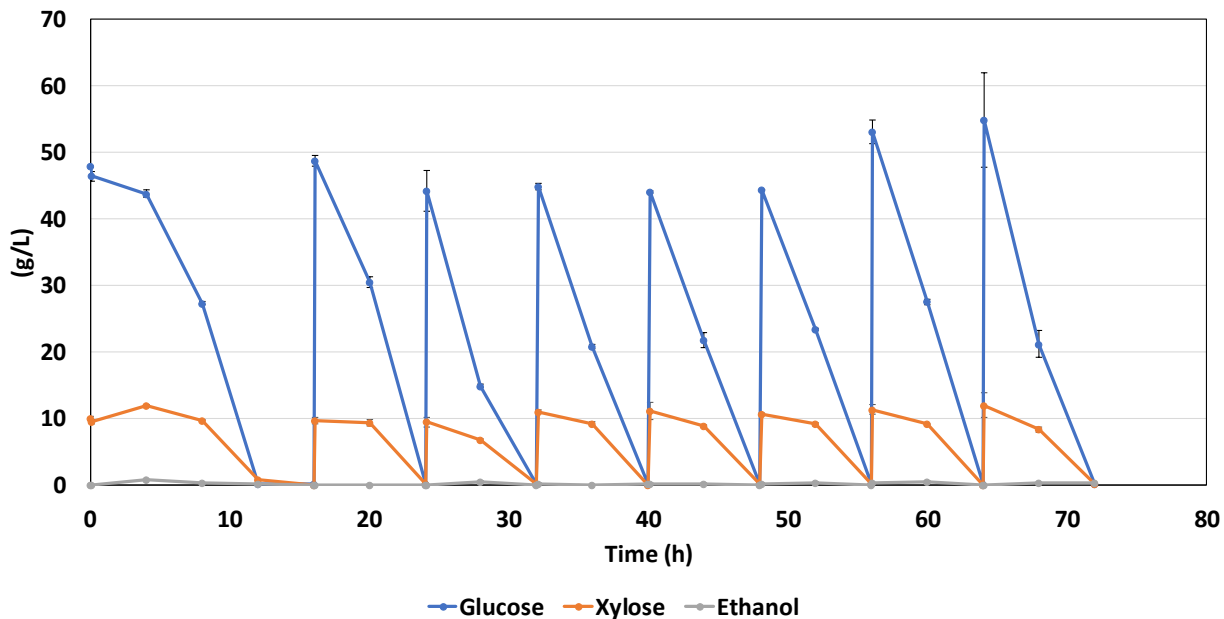


Figure S6. Repeated fed-batch cultivation of *W. anomalus* grown on 80% poultry hydrolysates, 20% urea with BALI™ hydrolysates in a 42 L fermenter. The starting volume was 25 L and the fermentation lasted 72 h. The figure shows glucose (blue lines), xylose (orange lines) and ethanol (grey lines). Acetic acid and lactic acid production were negligible (results not shown). Values are means \pm SD (n = 2).

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Table S1 Composition of BALI™ spruce hydrolysate

Content	BALI™
Dry matter (w/w %)	62.3
Density (kg/L)	1.286
Total sugars (% DM)	90.0
Glucose	68.2
Xylose	5.4
Mannose	6.5
Other sugars ^a	9.9
Acids ^b	1.2
Glycerol	0.2
Lignin	5.3

^a Sum of fructose, arabinose, galactose, gentobiose and cellobiose. ^b Sum of lactic, formic and acetic acid.

Table S2. CDW and protein content in 1.5 L batch fermentations (24h).

	Sampling time (h)	Poultry hydrolysates + BALI™	
		CDW (g/L)	Protein content (%)
<i>C. jadinii</i>	12	13.4 ± 1.2	47.1 ± 0.7
	24	17.4 ± 1.7	45.1 ± 0.9
<i>W. anomalus</i>	12	17.7 ± 0.4	50.8 ± 0.4
	24	29.7 ± 0.2	45.9 ± 0.3
<i>B. adenivorans</i>	12	16.4 ± 1.6	47.2 ± 0.4
	24	44.0 ± 0.0	41.9 ± 2.6

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Table S3. The numbers and assumptions related to the industrial evaluation of replacing 10 % of the annual Norwegian fish feed protein by yeast produced using the media described in this study (a detailed description of the calculations is presented below).

	Quantity	Unit
<i>Numbers related to replacing 10 % of fish feed by yeast: *</i>		
Produced fish feed per year ¹	1 630 000	tonnes
Replace 10 % of the feed with SCP	163 000	tonnes
Biomass yield on glucose	0.66	g/g
Amount glucose needed for industrial production	250 000	tonnes
Amount of cellulose **	220 000	tonnes
Amount of dry spruce ²	510 000	tonnes
Amount of wet spruce ³	962 000	tonnes
% of the annual spruce harvest in Norway ⁴	11.3	%
Amount BIOCO hydrolysate needed (80 %, 4.69 g N/L)	330 000	tonnes
Amount urea needed (20 %, 1.17 g N/L)	12 550	tonnes
Q_X^*	3.92	g/L/h
Q_X (converted to tonnes per liter per year) *	0.03434	tonnes/L/year
Total fermentation volume required to produce 163 000 tonnes of SCP	4747	m ³
No. of bioreactors of 300 m ³ with a working volume of 200 m ³ required to produce the yeast	24	reactors
<i>Numbers related to replacing 10 % of fish feed protein by yeast protein: *</i>		
Amount feed protein per year (50% of the feed)	815 000	tonnes
Replace 10 % of the protein with SCP	81 500	tonnes
Q_P^*	1.87	g/L/h
Q_P (converted to tonnes per liter per year) *	0.01638	tonnes/L/year
Total fermentation volume required to produce 81 500 tonnes of protein	4 975	m ³
No. of bioreactors of 300 m ³ with a working volume of 200 m ³ required to produce the SCP	25	reactors

* Based data from the pilot-scale fermentation of *W. anomalus*. Note that the protein content of the yeast was 47.8 %, whereas the protein content of the feed is 50 %.

** Conversion of cellulose to glucose increases the mass by a factor 1.11. Note that the calculation assumes that saccharification of cellulose happens with 100 % efficiency.

Calculations for Table S3:

BALI Hydrolysate (Spruce)

Production of fish feed per year = 1 630 000 tonnes

If 10 % is replaced

$$\frac{1\,630\,000\text{ tonnes}}{10} = 163\,000\text{ tonnes}$$

The biomass yield based on glucose in our experiment is 0.66 g/g

$$\frac{163\,000\text{ tonnes SCP}}{0.66\frac{\text{g}}{\text{g}}} \cong 250\,000\text{ tonnes of glucose are needed}$$

The Conversion from glucose to cellulose is 1.11 (180/162, Molecular weight ratio of free glucose to glucose units in cellulose)

$$\frac{250\,000\text{ tonnes of glucose}}{1.11} \cong 220\,000\text{ tonnes of cellulose}$$

The amount of cellulose in dried spruce is 43 %

$$\frac{220\,000\text{ tonnes of cellulose}}{0.43} \cong 510\,000\text{ tonnes of dry spruce}$$

The moisture content for Norwegian spruce is 53 %

$$510\,000\text{ tonnes of dry spruce}/0.53 \cong 962\,000\text{ tonnes of wet spruce}$$

Around $10.0 \times 10^6 \text{ m}^3$ (8.5×10^6 wet tonnes) of spruce are annually harvested in Norway

Thus,

$$\frac{962\,000\text{ tonnes of wet spruce}}{8.5 \times 10^6\text{ tonnes}} \times 100 \cong 11.3\% \text{ of the annual spruce harvest in Norway}$$

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BIOCO Hydrolysate:

Nitrogen originating from BIOCO in fermentations using 80 % BIOCO hydrolysate: 4.69 g/L nitrogen

Nitrogen conversion factor (Kjeldahl method) = 6.25

$$4.69 \left(\frac{g}{L} N \right) \times 6.25 = 29.3 \frac{g}{L} \text{ protein}$$

Assuming 250×10^3 tonnes of glucose are needed for industrial production

and

$50 \times 10^{-6} \frac{\text{tonnes}}{L}$ of glucose are used in our experiment:

The total fermentation volume would be

$$\frac{250 \times 10^3 \text{ tonnes of glucose}}{50 \times 10^{-6} \frac{\text{tonnes}}{L}} = 5000 \times 10^6 L$$

$29.3 \frac{g}{L}$ protein in fermentation from BIOCO $\times 500 \times 10^6 L = 146\,500$ tonnes of protein

*= **330 000 tonnes of BIOCO hydrolysate** (44% protein)*

Urea:

Nitrogen content in fermentations using 20 % Urea = 1.17 g/L nitrogen

$1.17 \frac{g}{L}$ urea nitrogen was used for every $50 \frac{g}{L}$ of glucose

60.06 g urea in 1 mol urea

28 g nitrogen in 1 mol urea

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$$\frac{28 \text{ g/mol}}{60.06 \text{ g/mol}} = 0.466$$

$$1.17 \frac{\text{g}}{\text{L}} \text{nitrogen} / 0.466 = 2.51 \text{ g/L urea in the fermentations}$$

$$\frac{250 \times 10^3 \text{ tonnes of glucose} \times 2.51 \frac{\text{g}}{\text{L}} \text{ urea}}{50 \frac{\text{g}}{\text{L}} \text{ glucose}} \cong \mathbf{12\ 550 \text{ tonnes of urea}}$$

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