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Last updated by author(s):	Feb 11, 2020

Reporting Summary

Nature Research wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Research policies, seeAuthors & Referees and theEditorial Policy Checklist.

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FOI	an statistical analyses, confirm that the following items are present in the figure regend, table regend, main text, or interhous section.
n/a	Confirmed
	The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement
	🗴 A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
	The statistical test(s) used AND whether they are one- or two-sided Only common tests should be described solely by name; describe more complex techniques in the Methods section.
	🗴 A description of all covariates tested
	🗴 A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
	A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
	For null hypothesis testing, the test statistic (e.g. <i>F</i> , <i>t</i> , <i>r</i>) with confidence intervals, effect sizes, degrees of freedom and <i>P</i> value noted <i>Give P values as exact values whenever suitable.</i>
×	For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
×	For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
×	\square Estimates of effect sizes (e.g. Cohen's d , Pearson's r), indicating how they were calculated
	Our web collection on <u>statistics for biologists</u> contains articles on many of the points above.

Software and code

Policy information about availability of computer code

Data collection	Zeiss Zen 2011
	NimOS 08032018
	Deltavision softWoRx 6.1.1
	GeneSys
	GE/Omega
	Topspin

FIJI 1.52n Data analysis Excel 2013

> GraphPad ver.8.1.2 MATLAB ThunderSTORM TrackMate

Python (custom scripts)

DANGLE PROCHECK-NMR Origin 7.0 SigmaPlot 12.0 Modeller 9.19

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors/reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Research guidelines for submitting code & software for further information.

Data

Policy information about <u>availability of data</u>

All manuscripts must include a <u>data availability statement</u>. This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A list of figures that have associated raw data
- A description of any restrictions on data availability

The data supporting corresponding author	-	f the study are available in the article and its Supporting Information or archived in the PDB or available upon request from the		
Field-spe	ecific r	reporting		
Please select the o	ne below tha	at is the best fit for your research. If you are not sure, read the appropriate sections before making your selection.		
🗶 Life sciences		Behavioural & social sciences		
For a reference copy of	the document w	rith all sections, see <u>nature.com/documents/nr-reporting-summary-flat.pdf</u>		
Life scier	nces s	tudy design		
All studies must dis	sclose on the	ise points even when the disclosure is negative.		
Sample size	For TIRF, SIN	RF, SIM and confocal microscopy 30 cells were analysed per strain/condition.		
Data exclusions		or single particle analysis (figure 1 b and c) trajectories were eliminated if they occurred outside cell's coordinates. When determining ffusion coefficients from MSD curves, if the determined diffusion coefficients were negative they were discarded.		
Replication	All experime all the assay	riments were performed in at least triplicate independent experiments in accordance with well-established reporting procedures for ssays		
Randomization		ribe how samples/organisms/participants were allocated into experimental groups. If allocation was not random, describe how covariates controlled OR if this is not relevant to your study, explain why.		
Blinding	The investig	he investigators were not blinded to the samples identity.		
Reportin	g for s	specific materials, systems and methods		
		ors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.		
Materials & ex				
n/a Involved in th	·	n/a Involved in the study		
Antibodies	•	ChIP-seq		
x Eukaryotic	cell lines	Flow cytometry		
Palaeontology MRI-based neuroimaging				
X Animals ar	nd other organ	isms		
=1=	search particip	ants		
X Clinical dat	ta			
Antibodies				
Antibodies used		Anti-rabbit (Sigma #A6154), Anti-Pal (in house, Bonsor et al., 2009), Anti-TolB (in house, Bonsor et al., 2009)		
Validation data is available at the manufacturer's website		Validation data is available at the manufacturer's website		