

Supporting Information

Antibody-Linked Fluorogen-Activating Proteins for Antigen Detection and Cell Ablation

**Daniel S. Ackerman^{†,§}, Burcin Altun[‡], Dmytro Kolodieznyi^{††,§}, Marcel P. Bruchez^{†,††,§},
Andrew Tsourkas[‡], and Jonathan W. Jarvik^{†,§,*}**

[†]Department of Biological Sciences, ^{††}Department of Chemistry, and [§]Molecular Biosensors and Imaging Center, Carnegie Mellon University, Pittsburgh, Pennsylvania, 15213.

[‡]Department of Bioengineering, University of Pennsylvania, Philadelphia, Pennsylvania, 19104.

*Corresponding author: Jonathan W. Jarvik, Mellon Institute Room 257, 4400 Fifth Avenue, Pittsburgh, Pennsylvania, 15213. Phone: (412) 398-0490. Fax: (412) 268-7129. E-mail:

jarvik@cmu.edu.

A

```

MTFKLIINGKTLKGEITIEAVDA*EAEKIFKQYANDY
GIDGEWTYDDATKTFTVTEGGSGGSYPYDVPDYAGAQ
PAQAVVTQEPSVTVSPGGTVILTTCGSGTGAVTSGHYA
NWFQQKPGQAPRALIFDTDKKYSWTPGRFSGSLLGAK
AALTISDAQPEDEAEYYCSLSDVDGYLFGGGTQLTVL
SGGGSGGGSGGGSGGGSGGGSQAVVTQEPSVTVSPGG
TVILTTCGSGTGAVTSGHYANWFQQKPGQAPRALIFDT
DKKYSWTPGRFSGSLLGAKAALTISDAQPEDEAEYYC
SLSDVDGYLFGGGTQLTVLS

```

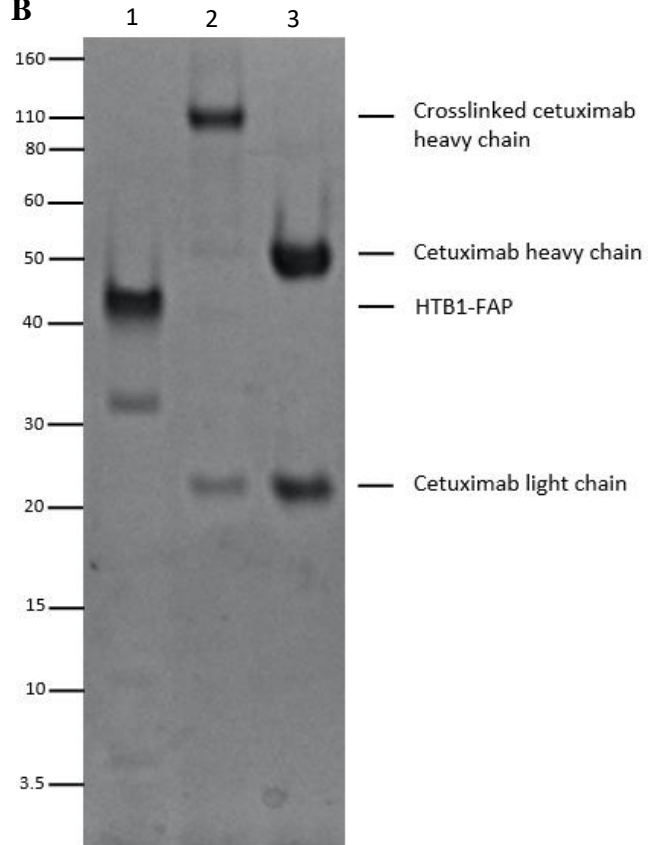
B

Figure S1. HTB1-FAP(dL5**) reagent conjugated to cetuximab. (A) Amino acid sequence of HTB1-FAP polypeptide (316 amino acids). The location of the p-benzoyl-L-phenylalanine (BPA) residue is indicated by an asterisk. (B) SDS PAGE showing crosslinked polypeptides. Lane 1: HTB1-FAP polypeptide. Lane 2: Crosslinked cetuximab. Lane 3: Cetuximab alone.

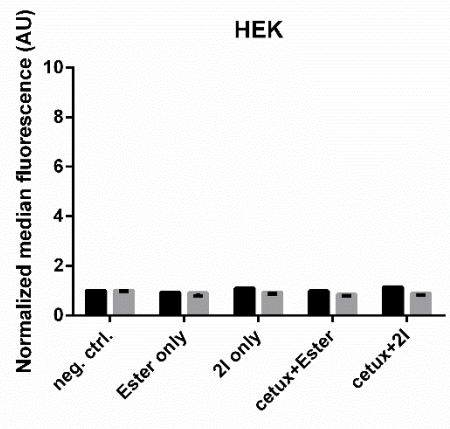
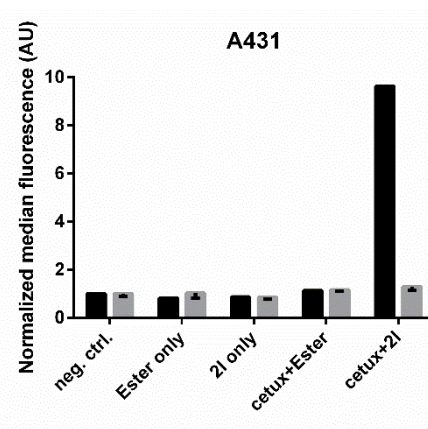
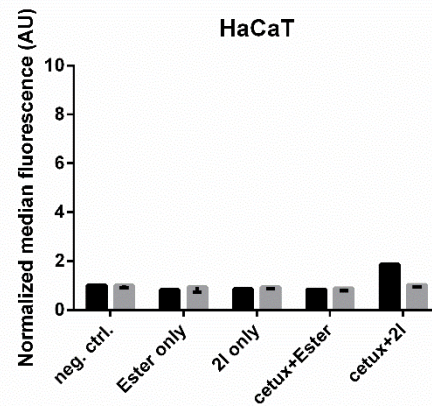
A**B****C**

Figure S2. Flow cytometric analysis of cytotoxic response via Eth-D staining in HEK, HaCaT, and A431 cells after dL5***-cetuximab* and MG-2I treatment and photoirradiation. Gray bars are from irradiated cells, and black bars are from non-irradiated cells. Neg. ctrl. = no antibody or dye. Ester/2I only = only fluorogen, no FAP/antibody. Cetux+Ester/2I = antibody and indicated fluorogen. Data from (A) HEK-293 cells, (B) HaCaT cells, and (C) A431 cells are shown. Numbers shown are the normalized average median FL2 signal (EthD-1 fluorescence) in arbitrary units normalized to the negative controls. Each bar is therefore the fold increase in EthD-1 signal over the negative control. Each bar is the average of three replicates \pm SD.

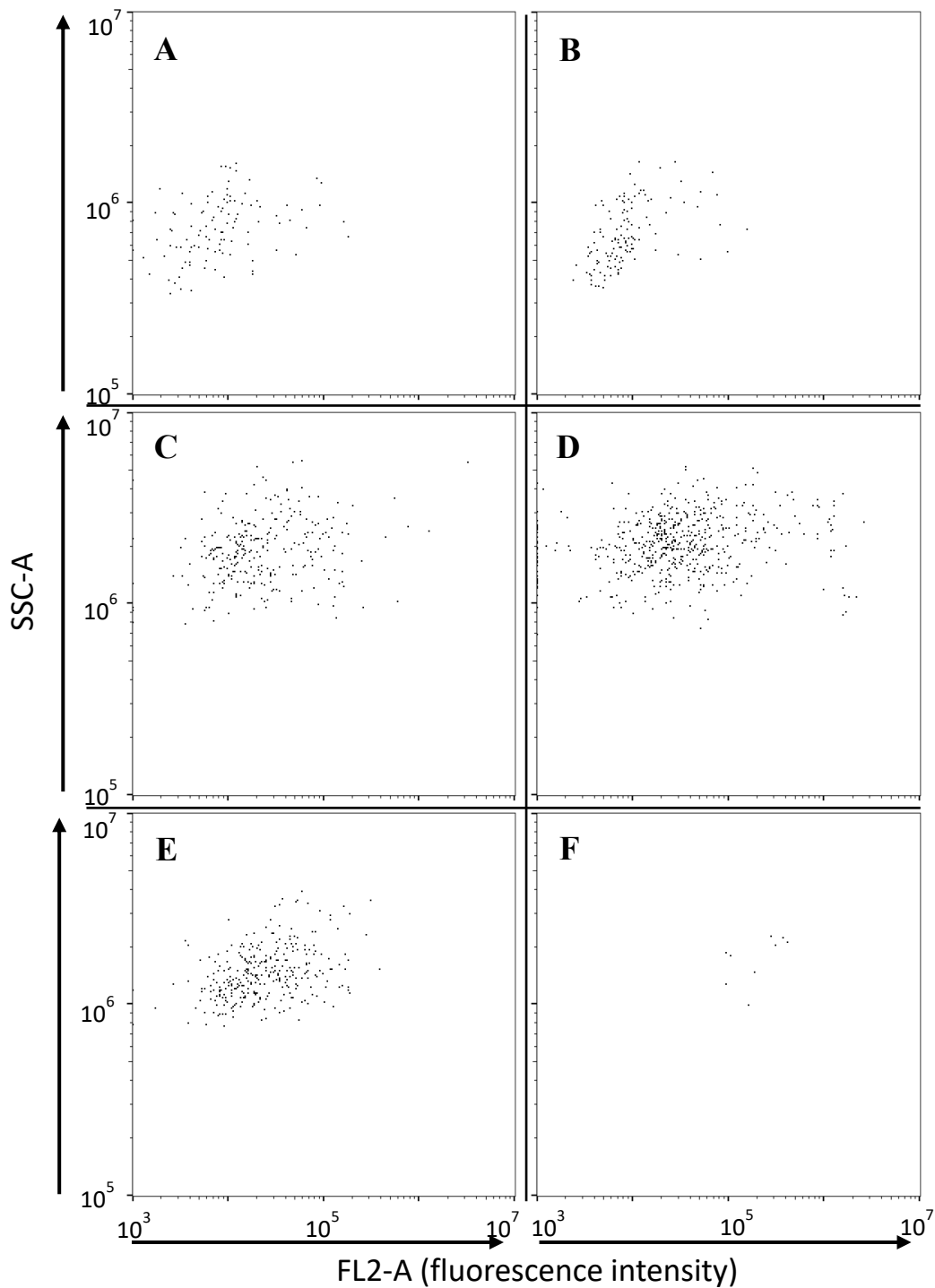


Figure S3. Representative dot plots from flow cytometry data shown in Figure S2. (A) and (B) HEK-293 cells. (C) and (D) HaCaT cells. (E) and (F) A431 cells. All cells shown were exposed to dL5**⁻-cetuximab and the photosensitizing fluorogen MG-2I. Cells in the right column were irradiated with 640nm light, while cells in the left column were not irradiated. Axes are side scatter vs. fluorescence intensity in the FL-2 channel (both in arbitrary units). The FL-2 channel shows signal from the dead cell stain EthD-1.

Table S1. Total number of singlet cells analyzed per condition to generate the data shown in Figure S2. Irr. = cells irradiated at 640nm. Non-irr. = cells not irradiated. Neg. ctrl. = no antibody or dye. Ester/2I only = only fluorogen, no FAP/antibody. Cetux+Ester/2I = antibody and indicated fluorogen.

HEK-293 cells		
	Irr.	Non-irr.
Neg. ctrl.	551	314
Ester only	433	248
2I only	601	387
Cetux+Ester	534	331
Cetux+2I	455	392
HaCaT cells		
	Irr.	Non-irr.
Neg. ctrl.	1350	901
Ester only	1546	1317
2I only	1385	1451
Cetux+Ester	1644	1047
Cetux+2I	1779	857
A431 cells		
	Irr.	Non-irr.
Neg. ctrl.	1105	907
Ester only	946	1020
2I only	709	1089
Cetux+Ester	497	802
Cetux+2I	12	725