

Corresponding author(s):	Kandice Levental, Ilya Levental
Last updated by author(s):	Jan 17, 2020

# **Reporting Summary**

Nature Research wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Research policies, seeAuthors & Referees and theEditorial Policy Checklist.

Statistics					
For all statistical analys	es, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.				
n/a Confirmed					
The exact sam	ple size $(n)$ for each experimental group/condition, given as a discrete number and unit of measurement				
A statement o	n whether measurements were taken from distinct samples or whether the same sample was measured repeatedly				
	test(s) used AND whether they are one- or two-sided ests should be described solely by name; describe more complex techniques in the Methods section.				
A description	🔲 🗷 A description of all covariates tested				
A description	of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons				
	A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)				
For null hypothesis testing, the test statistic (e.g. <i>F</i> , <i>t</i> , <i>r</i> ) with confidence intervals, effect sizes, degrees of freedom and <i>P</i> value noted Give <i>P</i> values as exact values whenever suitable.					
For Bayesian a	analysis, information on the choice of priors and Markov chain Monte Carlo settings				
For hierarchic	For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes				
Estimates of e	ffect sizes (e.g. Cohen's $d$ , Pearson's $r$ ), indicating how they were calculated				
'	Our web collection on <u>statistics for biologists</u> contains articles on many of the points above.				
Software and c	ode				
Policy information abou	ut <u>availability of computer code</u>				
Data collection	NIS Elements AR/BR/HC, Biorad Image Lab 6.0, PTI Felix Horiba, LSR Fortessa Cell Analyzer, SymphoTime				
Data analysis	Matlab, GraphPad Prism, Biorad Image Lab, ImageJ, Excel, Weasel, SymphoTime				
For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors/reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Research guidelines for submitting code & software for further information.					
Data					
Policy information about availability of data  All manuscripts must include a data availability statement. This statement should provide the following information, where applicable:  - Accession codes, unique identifiers, or web links for publicly available datasets  - A list of figures that have associated raw data  - A description of any restrictions on data availability					
The data sets generated during and/or analyzed during the current study are available from the corresponding author on request.					
Field-specific reporting					
Please select the one b	elow that is the best fit for your research. If you are not sure, read the appropriate sections before making your selection.				
Life sciences Behavioural & social sciences Ecological, evolutionary & environmental sciences					
For a reference copy of the do	ocument with all sections, see nature.com/documents/nr-reporting-summary-flat.pdf				

## Life sciences study design

Sample size	Numerous technical replicates were conducted for any given experiment. The maximum feasible sample sizes were used
Data exclusions	none
Replication	A minimum of 3 independent experiments were conducted for all reported results
Randomization	Describe how samples/organisms/participants were allocated into experimental groups. If allocation was not random, describe how covariates were controlled OR if this is not relevant to your study, explain why.
Blinding	all lipidomics analyses were done by an outside lab blinded to the identity of the samples

## Reporting for specific materials, systems and methods

We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, system or method listed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.

Materials & experimental systems		Me	Methods	
n/a	Involved in the study	n/a	Involved in the study	
	<b>x</b> Antibodies	x	ChIP-seq	
×	Eukaryotic cell lines		Flow cytometry	
x	Palaeontology	x	MRI-based neuroimaging	
	X Animals and other organisms		•	
x	Human research participants			
x	Clinical data			

#### **Antibodies**

Antibodies used

actin (monoclonal clone AC-15, Abcam), SREBP2 (polyclonal, Abcam), SREBP1 (monoclonal, Abcam)

Validation

Describe the validation of each primary antibody for the species and application, noting any validation statements on the manufacturer's website, relevant citations, antibody profiles in online databases, or data provided in the manuscript.

### Animals and other organisms

Policy information about studies involving animals; ARRIVE guidelines recommended for reporting animal research

Laboratory animals

female C57BL/6 mice

Wild animals

Provide details on animals observed in or captured in the field; report species, sex and age where possible. Describe how animals were caught and transported and what happened to captive animals after the study (if killed, explain why and describe method; if released, say where and when) OR state that the study did not involve wild animals.

Field-collected samples

For laboratory work with field-collected samples, describe all relevant parameters such as housing, maintenance, temperature, photoperiod and end-of-experiment protocol OR state that the study did not involve samples collected from the field.

Ethics oversight

Public Health Service and the Institutional Animal Care and Use Committee at Texas A&M University

Note that full information on the approval of the study protocol must also be provided in the manuscript.

## Flow Cytometry

### Plots

Confirm that:				
The axis labels state the marker and fluorochrome used (e.g. CD4-FITC).				
The axis scales are clearly visible. Include numbers along axes only for bottom left plot of group (a 'group' is an analysis of identical markers).				
■ All plots are contour plots with outliers or pseudocolor plots.				
A numerical value for number of cells or percentage (with statistics) is provided.				
Methodology				
Sample preparation	Describe the sample preparation, detailing the biological source of the cells and any tissue processing steps used.			
Instrument	Identify the instrument used for data collection, specifying make and model number.			
Software	Describe the software used to collect and analyze the flow cytometry data. For custom code that has been deposited into a community repository, provide accession details.			
Cell population abundance	Describe the abundance of the relevant cell populations within post-sort fractions, providing details on the purity of the samples and how it was determined.			
Gating strategy	gating was set using standard positive and negative controls for apoptosis			

🕱 Tick this box to confirm that a figure exemplifying the gating strategy is provided in the Supplementary Information.