Table S1. Primers used for generation of RNA precursor substrates, assessment of spliced product, un-spliced substrate, and total cDNA from self-excision and maturase activity assays, and qPCR standards. Annealing temperatures and expected product sizes are given.

Substrate	Forward primer	Reverse primer	Expected product size	T_A
		RNA precursor substrates		
Rps12-2	*rps12*exon3'splice For GTTGCCAGAGTACGATTA[1]	*rps12*exon3'splice Rev TGCTTTTTGACCCCATA[2]	753 bp	48 °C
Rpl2	*rpl2* splice For CAAGTGAAATCCAATCC[1]	*rpl2* splice Rev CGTTTACCTAGCCAACA[2]	1247 bp	48 °C
		Spliced product assessment from activity assays		
Rps12-2	*rps12*exon3'splice For GTTGCCAGAGTACGATTA	*rps12* intron2 span Rev CCCATATTTAGAACGCC	190 bp	50 °C
Rpl2	*rpl2*intronspan F CTACCTTTGACCGATATGC	*rpl2* splice Rev CGTTTACCTAGCCAACA	240 bp	50 °C

qPCR standards				
Rps12-2	*rps12*exon3'splice For GTTGCCAGAGTACGATTA[1]	*rps12*exon3'splice Rev TGCTTTTTGACCCCATA[2]	212 bp	48 °C
Rpl2	*rpl2* splice For CAAGTGAAATCCAATCC[1]	*rpl2* splice Rev CGTTTACCTAGCCAACA[2]	584 bp	48 °C
Detection of un-spliced product				
Rps12	*rps12*exon3'splice For GTTGCCAGAGTACGATTA	*Rps12in2*Rev GTTAGCCATACACTTCACA	277 bp	52 °C
Rpl2	*rpl2* splice For CAAGTGAAATCCAATCC	*rpl2* exon1-intron Rev CTTGCTGCCGTTACTCA	472 bp	48 °C
Detection of Total RNA				
Rps12	*rps12*exon3'splice For GTTGCCAGAGTACGATTA	*Rps12*exon2Rev TACTGCGACAGCATCTAG	168 bp	52 °C
Rpl2	*rpl2* splice For CAAGTGAAATCCAATCC	*rpl2* exon1Rev CCCATAGTGTATGAGACA	225 bp	48 °C

[1] Gateway *attB1* primer sequence GGGGACAAGTTTGTACAAAAAAGCAGGCTTC added to forward primer to facilitate BP cloning.

[2] Gateway *attB2* primer sequence GGGGACCACTTTGTACAAGAAAGCTGGGTT added to reverse primer to facilitate BP cloning.