

## **Supplementary information**

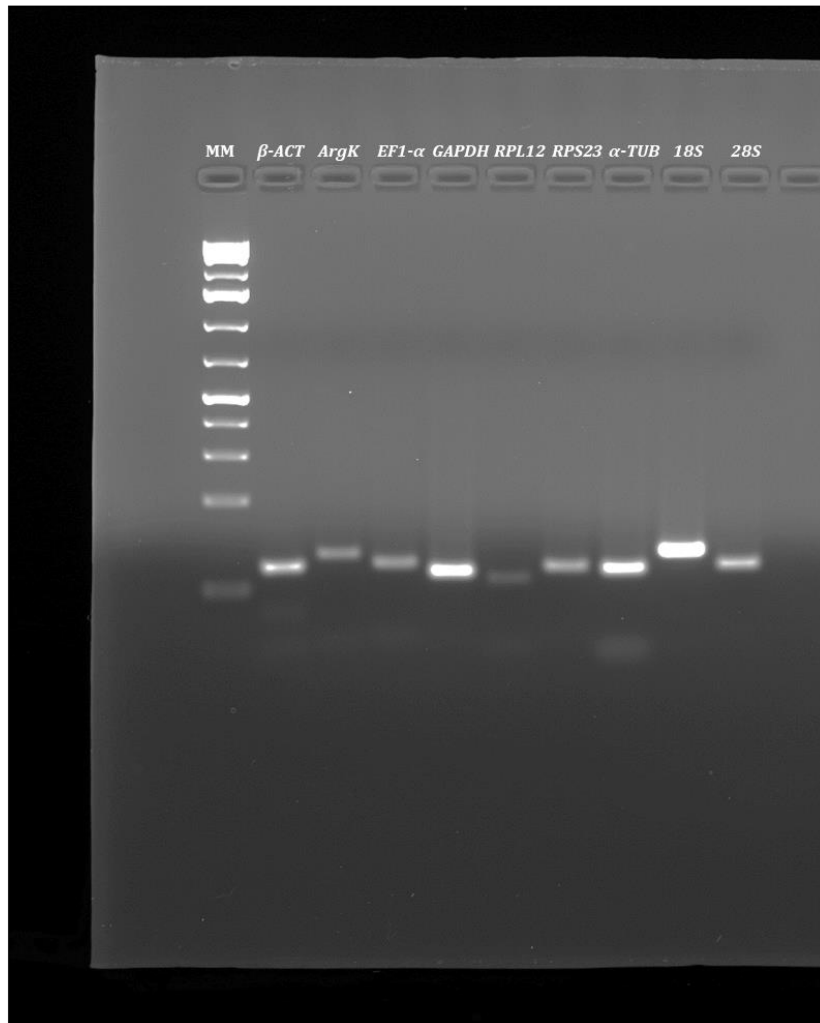
### **Selection of reference genes for normalization of RT-qPCR data in gene expression studies in *Anthonomus eugenii* Cano (Coleoptera: Curculionidae)**

Daniele H. Pinheiro <sup>1,2</sup> & Blair D. Siegfried <sup>1\*</sup>

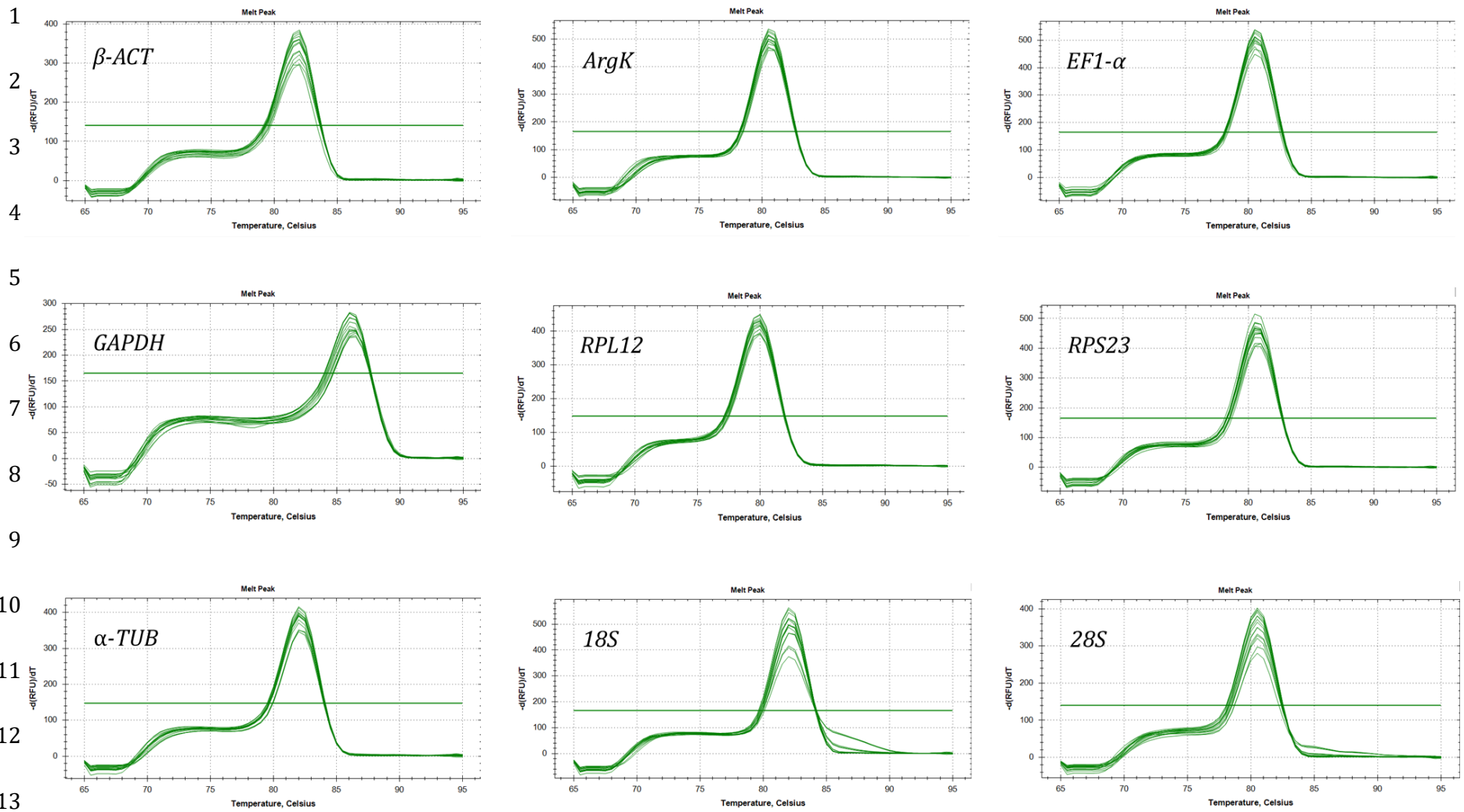
<sup>1</sup> University of Florida, Entomology and Nematology Department, Charles Steinmetz Hall, P. O. Box 110620, Gainesville, FL, 32611-0620, United States.

<sup>2</sup> Present address: Embrapa Genetic Resources and Biotechnology, Parque Estação Biológica, W5 Norte, P. O. Box 02372, Brasília, DF, 70770-917, Brazil.

\* Correspondence and requests for materials should be addressed to B.D.S. (email: bsiegfried1@ufl.edu).



**Supplementary Fig. S1.** Agarose gel (1.5%) indicates the amplification of a single fragment of candidate reference genes.



14 **Supplementary Fig. S2.** Melting curves generated by RT-qPCR.

15 **Table S1.** Comprehensive stability ranking of candidate reference genes according to BestKeeper, geNorm and NormFinder.

Experimental condition	Comprehensive ranking	Gene	BestKeeper	GeNorm	NormFinder	Mean
Developmental stages	1	<i>EF1-<math>\alpha</math></i>	1	3	1	1.66
	2	<i>18S</i>	2	5	2	3
	2	<i>RPL12</i>	4	1	4	3
	3	<i>28S</i>	3	4	5	4
	4	<i>RPS23</i>	5	2	7	4.66
	5	<i><math>\alpha</math>-TUB</i>	8	6	3	5.66
	6	<i>GAPDH</i>	6	7	6	6.33
	7	<i><math>\beta</math>-ACT</i>	7	9	9	8.33
	7	<i>ArgK</i>	9	8	8	8.33
Sex	1	<i>RPS23</i>	1	2	3	2
	2	<i>RPL12</i>	2	1	4	2.33
	3	<i>GAPDH</i>	3	6	2	3.66
	4	<i><math>\alpha</math>-TUB</i>	6	5	1	4
	5	<i>EF1-<math>\alpha</math></i>	5	3	6	4.66
	6	<i>28S</i>	4	4	7	5
	7	<i>18S</i>	7	7	5	6.33
	8	<i>ArgK</i>	9	8	8	8.33
	9	<i><math>\beta</math>-ACT</i>	8	9	9	8.66

Continued

16

17

18

Low temperature	1	<i>GAPDH</i>	1	1	1	1
	2	<i>α-TUB</i>	2	2	3	2.33
	3	<i>EF1-α</i>	3	3	2	2.66
	4	<i>18S</i>	4	4	4	4
	5	<i>28S</i>	5	6	7	6
	5	<i>ArgK</i>	7	5	6	6
	6	<i>RPS23</i>	5	9	5	6.33
	7	<i>RPL12</i>	9	7	8	8
High temperature	8	<i>β-ACT</i>	8	8	9	8.33
	1	<i>α-TUB</i>	3	1	4	2.66
	1	<i>RPS23</i>	4	2	2	2.66
	2	<i>GAPDH</i>	5	3	1	3
	3	<i>EF1-α</i>	7	4	3	4.66
	4	<i>RPL12</i>	6	5	7	6
	4	<i>28S</i>	1	9	8	6
	5	<i>18S</i>	2	8	9	6.33
All temperatures	6	<i>ArgK</i>	8	7	5	6.66
	7	<i>β-ACT</i>	9	6	6	7
	1	<i>α-TUB</i>	1	2	2	1.66
	2	<i>GAPDH</i>	5	1	1	2.33
	3	<i>RPS23</i>	4	6	3	4.33
	4	<i>ArgK</i>	7	3	4	4.66
	5	<i>EF1-α</i>	8	4	5	5.66
	5	<i>RPL12</i>	6	5	6	5.66
6	<i>18S</i>	3	8	7	6	
7	<i>28S</i>	2	9	8	6.33	
8	<i>β-ACT</i>	9	7	9	8.33	

Continued

Starvation	1	<i>RPL12</i>	1	3	1	1.66
	2	<i>α-TUB</i>	3	2	4	3
	2	<i>18S</i>	5	1	3	3
	3	<i>RPS23</i>	2	5	5	4
	4	<i>GAPDH</i>	7	4	2	4.33
	5	<i>EF1-α</i>	4	6	6	5.33
	6	<i>ArgK</i>	6	7	7	6.66
	7	<i>28S</i>	9	8	8	8.33
	8	<i>β-ACT</i>	8	9	9	8.66
dsRNA	1	<i>α-TUB</i>	1	3	1	1.66
	2	<i>RPL12</i>	3	1	2	2
	3	<i>RPS23</i>	4	2	3	3
	4	<i>18S</i>	5	4	6	5
	4	<i>28S</i>	2	5	8	5
	5	<i>EF1-α</i>	6	6	5	5.66
	6	<i>GAPDH</i>	8	8	4	6.66
	7	<i>ArgK</i>	7	7	7	7
	8	<i>β-ACT</i>	9	9	9	9

20

21

22

23

24

25 **Table S2.** Sequence and amplicon size of the primers used for cloning.

Primer	Sequence 5'-3'	Tm (°C)	Size (bp)	Reference
ArgK-F	CCTGTTTCGACCCTATCATCGARGAYTAYCA	55	733	Yang et al., 2016
ArgK-R	GTCGTAGATACCACCTTCAGCTTCNGTRTGTC			
EF1- $\alpha$ -F	GCCGGTACYGGWGAATTCGAAGC	55	290	Pinheiro et al., 2019
EF1- $\alpha$ -R	CATCCCTTGAACCBGGCATCTT			
RPL12-F	ATGCCMCAAARTTCGAYCCSA	55	365	Pinheiro et al., 2019
RPL12-R	GCCATDGAKCKDGGCCTCATTTG			
RPS23-F	TGGGCYGACAARGAKTACAARAAAGC	55	300	This study
RPS23-R	CCTTRAATCTDACWCCRGGGAATATC			
$\alpha$ -TUB-F	ATCAAGACCAARCGTACCATCC	55	257	Pinheiro et al., 2019
$\alpha$ -TUB-R	TCAGAGAAYTCWCCTTCYTCCATACC			
18S-F	GTTGCGGTTAAAAAGCTCGT	50	644	This study
18S-R	GTGAGGTTTCCCGTGTGAG			
28S-F	CGAGATTCCCCTGTCCCTA	55	532	This study
28S-R	ACTGAACATCGGGATCAAGC			
RpII140-F	TTGGAGTTYTTGGAGGAGTGGTC	55	581	This study
RpII140-R	CCCATGGCTTGYTTDCCCAT			
Pro $\alpha$ -2-F	CAAATYGARTATGCCTTRGCTGC	50	581	Pinheiro et al., 2019
Pro $\alpha$ -2-R	TCACAAATHCCDACTTCAATRTTATC			

Tm - Melting temperature used in PCR amplification reactions.

26

27

28

1 **References**

2 Yang, C. *et al.* Selection of reference genes for RT-qPCR analysis in a predatory biological  
3 control agent, *Coleomegilla maculata* (Coleoptera: Coccinellidae). *Sci. Rep.* **5**, 18201 (2016).

4 Pinheiro, D. H., Taylor, C. E., Wu, K. & Siegfried, B. D. Delivery of gene-specific dsRNA by  
5 microinjection and feeding induces RNAi response in Sri Lanka weevil, *Mylocerus*  
6 *undecimpustulatus undatus* Marshall. *Pest Manag. Sci.* (2019). doi:10.1002/ps.5601

7