

**Immunity, Volume 52**

## **Supplemental Information**

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### **Regions of the V $\gamma$ 9V $\delta$ 2 TCR and Is**

### **Essential for Phosphoantigen Sensing**

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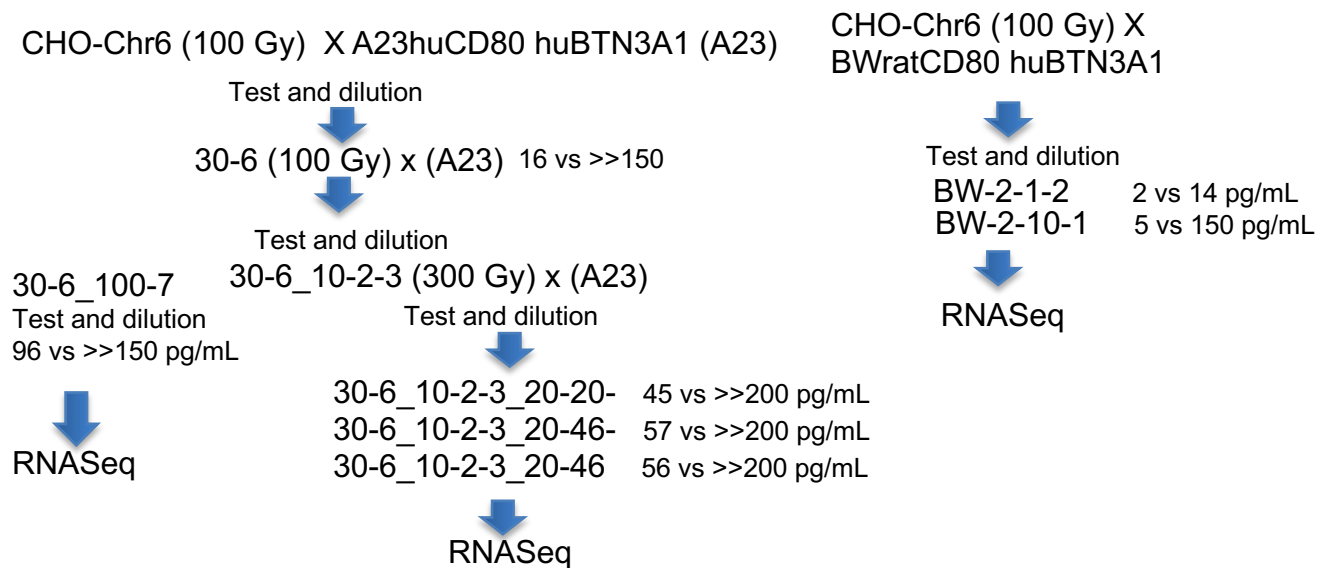
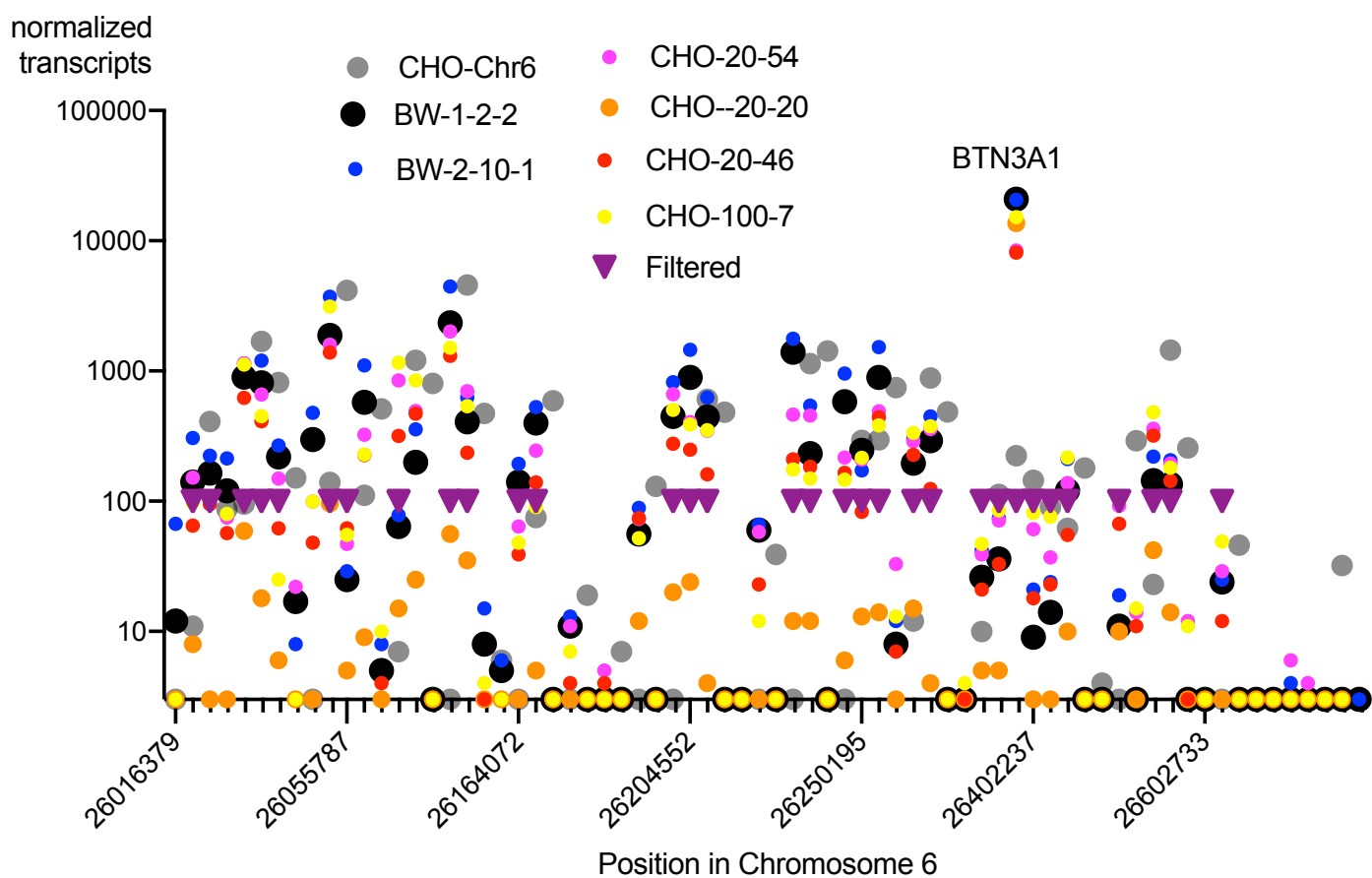
## **Supplemental Information**

### **Butyrophilin-2A1 Directly Binds Germline-Encoded**

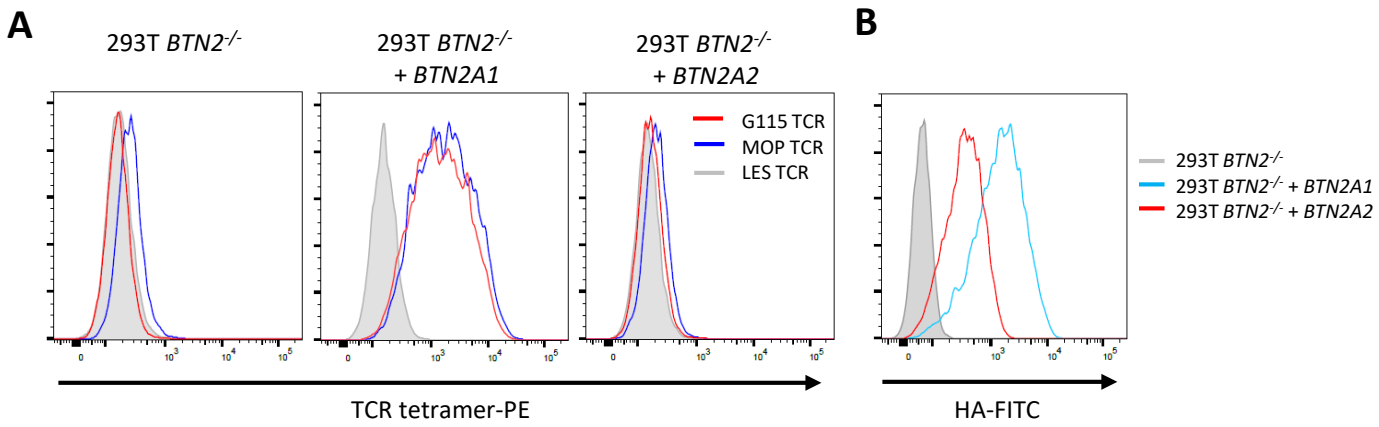
### **Regions of the V $\gamma$ 9V $\delta$ 2 TCR and Is**

### **Essential for Phosphoantigen Sensing**

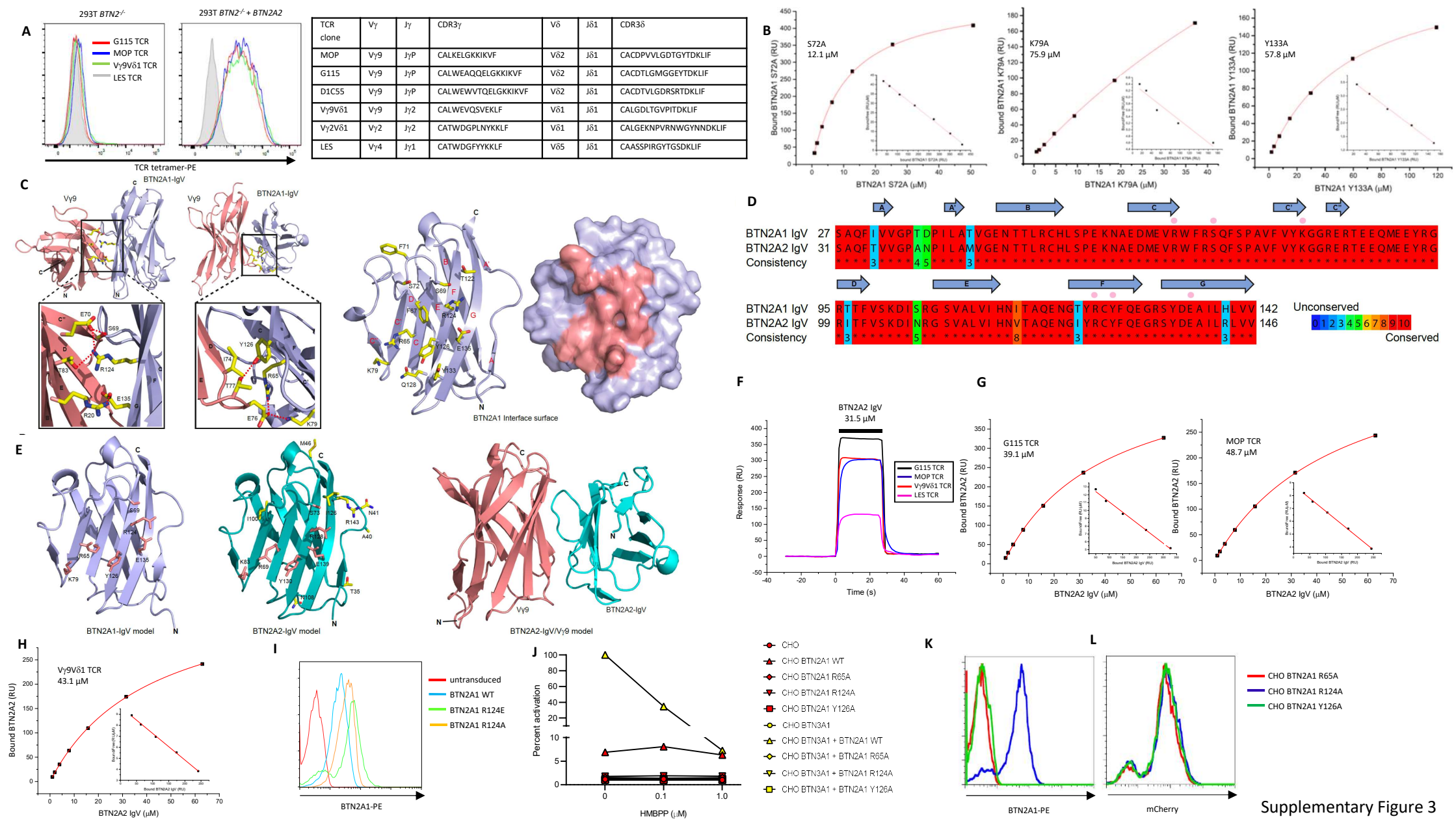
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**A****B**

**Supplementary Figure 1, related to Figure 1. Strategy used for identification of *BTN2A1* as Factor X.** (A) Origin of radiation hybrid clones selected for transcriptional analysis. IL-2 production (pg/ml) in the stimulation assay in the presence of medium or with 1  $\mu$ M HMBPP. (B) Region of human Chr 6 implicated in potentiation of P-Ag sensitisation by rodent-human radiation hybrids. Each dot represents a gene detected as  $\geq 3$  transcripts, with those highlighted in pink detected in every P-Ag-sensitizeable clone selected for analysis.



**Supplementary Figure 2, related to Figure 2. Vy9V62 TCR tetramer stains *BTN2A1* transductants.** (A) 293T *BTN2*<sup>-/-</sup> cells transduced to express HA-tagged *BTN2A1* and *BTN2A2* were stained with the indicated TCR tetramers. (B) Flow cytometry analysis of HA-tagged *BTN2A1* and *BTN2A2* in 293T *BTN2*<sup>-/-</sup> cells, as detected by anti-HA mAb.

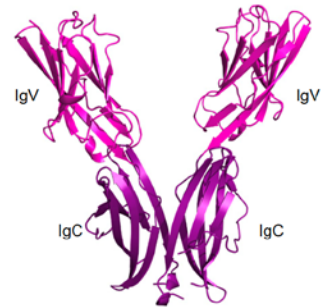
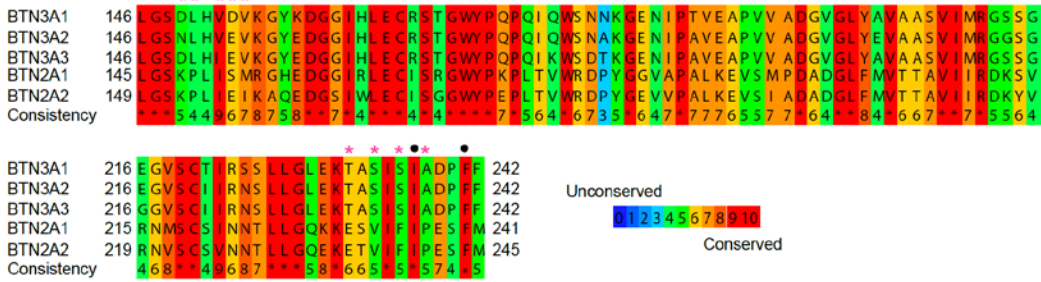


Supplementary Figure 3

**Supplementary Figure 3, related to Figure 3. BTN2A1 and BTN2A2 bind germline encoded regions of V $\gamma$ 9.** (A) (Left panel) V $\gamma$ 9V $\delta$ 1-TCR tetramer staining of BTN2A1-transduced 293T *BTN2*<sup>-/-</sup> cells. (Right panel) Comparison of CDR3 $\gamma$  sequences in G115, MOP, D1C55, and V $\gamma$ 9V $\delta$ 1 TCR clonotypes. (B) Equilibrium binding of BTN2A1 mutants S72A (left; 12.1  $\mu$ M), K79A (middle, 75.9  $\mu$ M) and Y133A (right, 57.8  $\mu$ M) to G115 TCR. Results are representative of two independent experiments. (C) (Left two panels) V $\gamma$ 9/BTN2A1 interface from the mutagenesis-informed HADDOCK model of the interaction. (Right two panels) BTN2A1 interface residues (left) forming a surface (right) proposed to interact with V $\gamma$ 9 IgV. (D) Comparison of BTN2A2 and BTN2A1 IgV sequence. Pink dots indicate conservation of BTN2A1 IgV amino acids predicted to be involved in binding V $\gamma$ 9 IgV, in BTN2A2. (E) Homology model of BTN2A2 IgV, alone and in complex with V $\gamma$ 9 TCR. (F) Specific binding of BTN2A2 IgV (31.5  $\mu$ M) to V $\gamma$ 9V $\delta$ 2 TCR as detected by SPR. (G) Equilibrium affinity and Scatchard (inset) analysis of BTN2A2 IgV binding to G115 (39.1  $\mu$ M) and MOP (48.7  $\mu$ M) V $\gamma$ 9V $\delta$ 2 TCRs. (H) Equilibrium affinity analysis of BTN2A2 IgV binding to V $\gamma$ 9V $\delta$ 1 TCR (43.1  $\mu$ M). (I) Flow cytometry staining of BTN2A1 R124A and BTN2A1 R124E mutants at surface of BTN3A1-expressing CD80<sup>+</sup> CHO cells. (J) Effects of BTN2A1 mutations on IL-2 production by TCR-MOP in response to HMBPP-treated BTN3A1 and BTN2A1 expressing CD80<sup>+</sup> CHO cells. (K) Flow cytometry staining of BTN2A1 R65A, R124A and Y126A mutants at surface of BTN3A1-expressing CD80<sup>+</sup> CHO cells. (L) mCherry reporter detection in BTN2A1 R65A, R124A and Y126A mutants-expressing CD80<sup>+</sup> CHO cells.

**A**

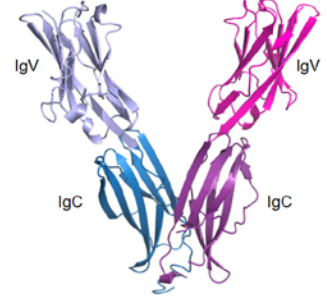
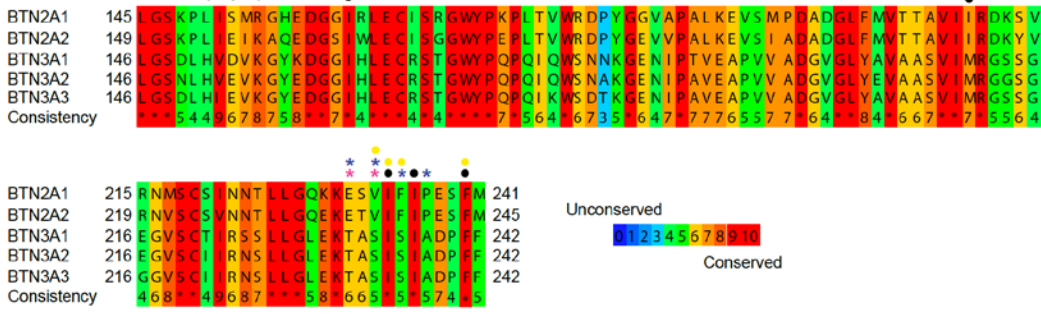
## BTN3A1 homodimer IgC-IgC dimer interface contacts



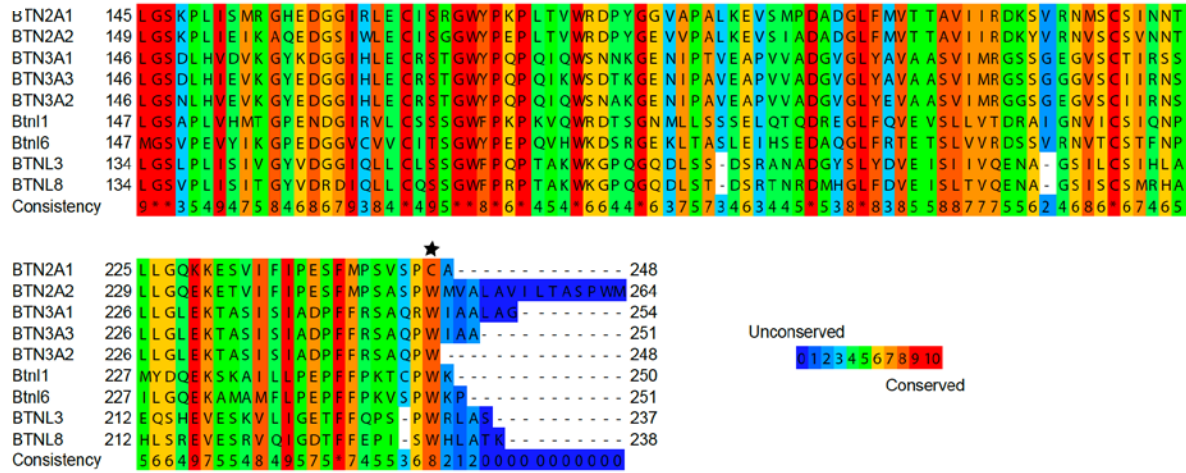
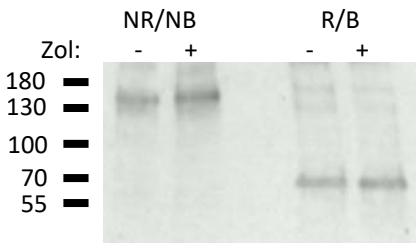
BTN3A1-BTN3A1 homodimer

**B**

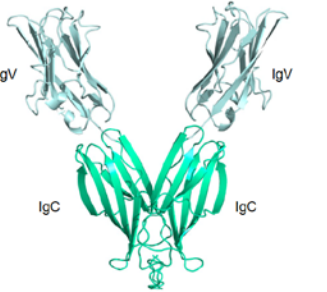
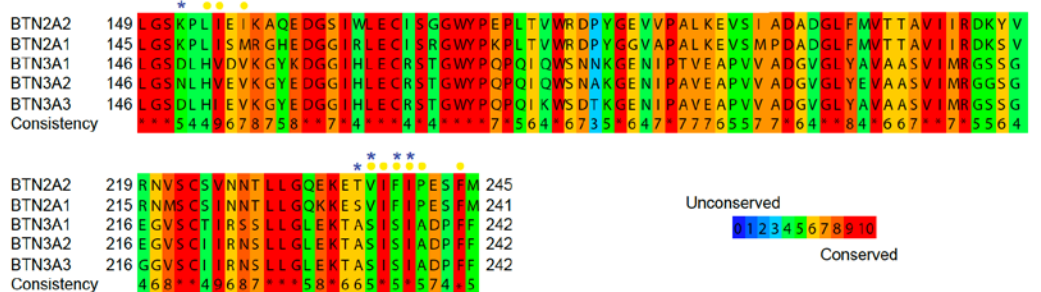
## BTN2A1-BTN3A1 heterodimer IgC-IgC dimer interface contacts



BTN2A1-BTN3A1 heterodimer

**C****D****E**

## BTN2A2 homodimer IgC-IgC dimer interface contacts



BTN2A2 Homodimer

**Supplementary Figure 4, related to Figure 4. BTN2A1 can form a homodimer involving hydrophobic IgC-IgC contacts and a disulphide linkage.** (A) Sequence alignment showing conservation of BTN3A1 IgC-IgC interface residues in BTN2A1. Pink stars indicate BTN3A1 hydrogen bonding interactions mediated by main chain atoms (D149, L150, V152, D153, V154, T232, S234, S236 and A238); black spheres indicate BTN3A1 hydrophobic interactions (V154, Y157, I237, and F241). (B) Sequence alignment showing potential for heterodimer interactions involving BTN2A1. Pink stars indicate BTN3A1 hydrogen bonding interactions mediated by main chain atoms (D149, L150, V152, V154, T232 and S234); black spheres indicate BTN3A1 hydrophobic interactions (H151, V152, D153, V154, K155, I162, M210, I235, I237 and F241); blue stars indicate BTN2A1 hydrogen bonding interactions mediated by main chain atoms (K148, M153, E231, V233, F235 and P237); yellow spheres indicate BTN2A1 hydrophobic interactions (L150, I151, M153, V233, I234, F235 and F240). (C) Sequence alignment showing unique membrane proximal Cys residue (highlighted by a black star) in BTN2A1 relative to other BTN/BTNL molecules. (D) BTN2A1 homodimer formation is not affected by Zol treatment. CD80<sup>+</sup> CHO cells expressing BTN2A1-HA and FLAG-BTN3A1 were treated +/- Zol (20  $\mu$ M) overnight, surface biotinylated, then lysed and immunoprecipitated using anti-HA antibody, and blotted with streptavidin-horseradish peroxidase. (E) BTN2A2 dimer formation. Homology model of BTN2A2 indicating potential for IgC-IgC homodimers. Blue stars indicate BTN2A2 hydrogen bonding interactions mediated by main chain atoms (K152, I155, I157, T236, V237, F239 and I240); yellow spheres indicate BTN2A2 hydrophobic interactions (L154, I155, I157, V237, I238, F239, I240, P241 and F244).



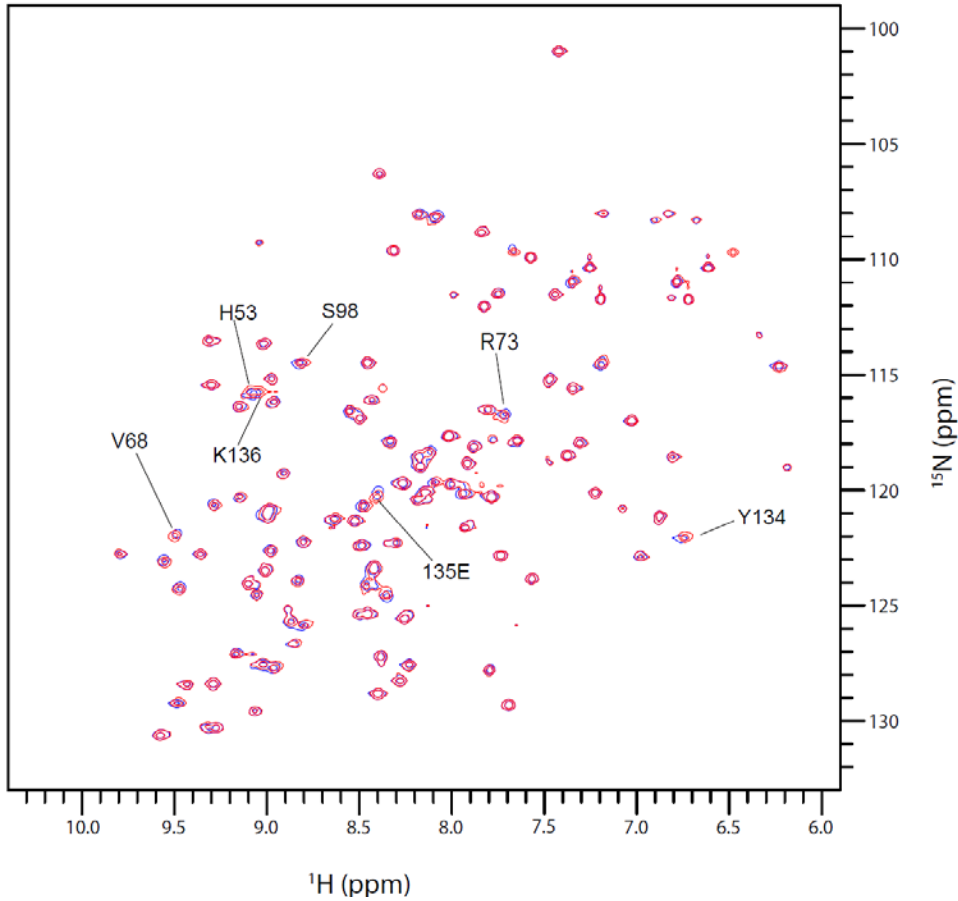
20.1 IP

A

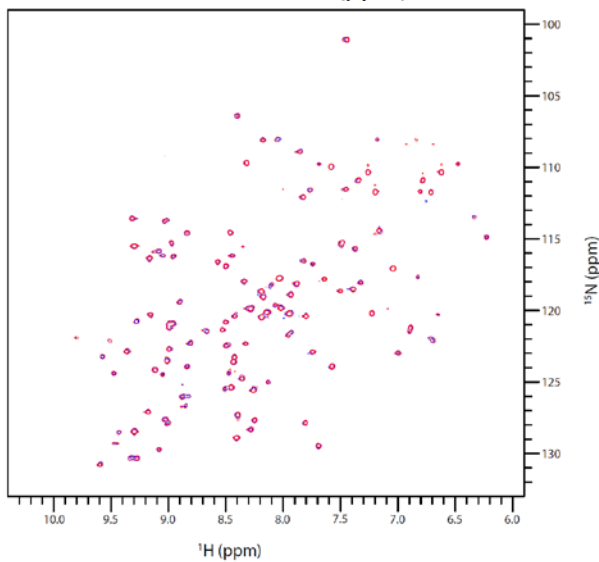
CHO cells:	2A1 HA		3A1 FLAG		2A1 HA+ 3A1 FLAG				Lysate
	+	-	+	-	+	-	+	-	
Zol	+	+	+	+	+	+	-	-	
sEGS XL	-	+	-	+	-	+	-	+	



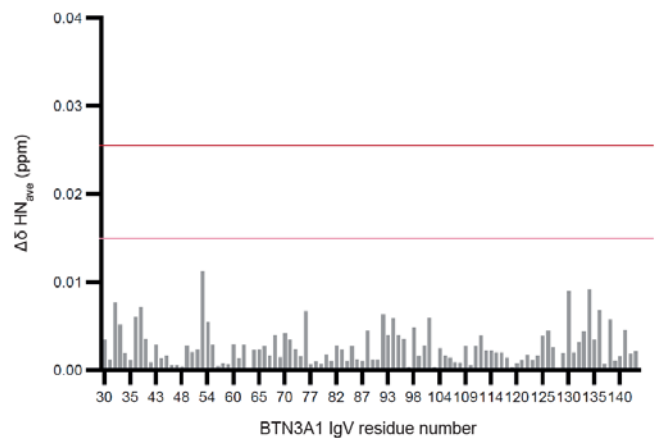
B



C



D



**Supplementary Figure 5, related to Figure 5. BTN2A1 associates with BTN3A1 via IgV-IgV contacts.** (A) SDS-PAGE following surface crosslinking and lysis of CHO CD80<sup>+</sup> cells transduced with BTN2A1-HA and FLAG-BTN3A1, and subsequent immunoprecipitation using 20.1 anti-BTN3A1 mAb, followed by western blot using anti-BTN2A1 polyclonal antibody. (B) NMR chemical shift perturbation analysis of BTN3A1 IgV domain (100  $\mu$ M) with BTN2A1 IgV. Overlay of <sup>1</sup>H-<sup>15</sup>N HSQC spectra of <sup>1</sup>H-<sup>15</sup>N-labelled BTN3A1 IgV domain in the absence (red) and presence (100  $\mu$ M; blue) of BTN2A1 IgV domain. Labelled residues are those undergoing CSPs above the threshold set in Figure 5D. (C) NMR chemical shift perturbation analysis of BTN3A1 IgV domain (120  $\mu$ M) with BTNL3 IgV. Overlay of <sup>1</sup>H-<sup>15</sup>N HSQC spectra of <sup>1</sup>H-<sup>15</sup>N-labelled BTN3A1 IgV domain in the absence (red) and presence (320  $\mu$ M; blue) of BTNL3 IgV domain. (D) Graph of chemical shift versus residue number in BTN3A1 IgV following addition of BTNL3 IgV domain as in (C). Threshold levels for significant CSPs are indicated by horizontal lines as in Figure 5D.

**Supplementary Table 1, related to main text Figure 1 and Key Resource Table. Primers and genomic markers used for characterization of radiation hybrids.**

Marker	Fwd primer	Rev primer	Source	Identifier
Ly86	AAATCTTCTTCCCCTGGGAA	GAGGATGGCTGTTAGATCATCC	This Paper	N/A
CD83	TAAGGGAAAGCCAACAATGG	CACGGTCTGTTCTTGAAGCA	This Paper	N/A
RN7SL	GAATGTGGATGGGCTGTCTC	GCCTCAAACAGTGCATTTATACTT	This Paper	N/A
MYLIP	CAGACATGCAGAAAGTTGCA	ATGCACACAGAAGATAAAGCA	This Paper	N/A
NUP153	GGACTCTCAGGCAGGAAT	TGGAACAGCCCAATAGGT	This Paper	N/A
RPL21	CCGCGGAATTCTTAGTTATCCTCT	TTTTTGCAAAGCTCACACCTTCT	This Paper	N/A
RPL36	TGTGTAGCGGAAGACATCTAGCA	TCCATGAGCACTGGTAAGAATGA	This Paper	N/A
Prolactin	GGTTTCTGATACTGGCCCGATA	GGAACGGATCATTAAGGACCTTCTC	Cathy Abbo	N/A
LOC1053	TCAGACCAAGAACAACGAGGAAC	ACTCAACACGTGTCACACACAGA	This Paper	N/A
ACOT13	GTATTGTAACAGGGTTCATGCA	CCCCAAGCTGAGACATGATT	This Paper	N/A
C6orf229	GATCCTGCTGCTCCAGAGTG	AAGCTCCTTGACGCAAGACA	This Paper	N/A
MTCO2P33	CAATTTGGCCTTCAAACGCT	AACACGTAAGGATAGGAGGGC	This Paper	N/A
RP3-425P12	AGATCATTTCTGAAGGCCGA	CCTTGCATTCTTTGGCGGGT	This Paper	N/A
SLC17A4	AGGCTGAGGCATCCCTA	TGTACCCCGCTAATAAGTT	This Paper	N/A
BTN3A2	AGCACTTTACTGATATTCATTC	AAAGTCAATACACTTTTCTACAC	This Paper	N/A
OR2B2	AATTTAAAATCACAGTGAGTAG	AGATGGAGGTTTCAGATAG	This Paper	N/A
MOG	AACTGAAAGGGTTCGGTGTG	ATTTCTGACCATTTGCCCTG	This Paper	N/A
TNFA	CAGGGTCTACACACAAATCAGTC	AAGAGAACCTGCCTGGCAGCTTGT	This Paper	N/A
HLA-DQ	GGTTATCACTCTTCTGTGATGCC	TCTGGGTCAGTGCAGGAAG	This Paper	N/A
STK38	TTGCTACTCAGCCTATTACTTTT	AAGGTTATACAGGACGACATAAC	This Paper	N/A
NFKBIE	GCCTTTTACTTCAACTGCCCTCT	ACAACAGGTGCCAGTTACAGGTT	This Paper	N/A
ICK	ACAAAAAAGCCTACTGCATTCA	CTGTCTGCCTTGGTGATGAATC	This Paper	N/A
EEF1A1	CCAACCTGAGATGTCCCTGT	TGAGTCACCCACACAAAGGA	This Paper	N/A
MAP3K7	TTGCTTAACCTTTTCCATTTCC	TCTGAATGTCACAGGCTGTACC	This Paper	N/A
CD164	GAGGCACCCATAACACTTG	TGAATTTTGGAGATGGAATGC	This Paper	N/A
PTPRK	CCACCATTATTCAATTCCAAGCAT	TGGTCTCTTAATGCCTGATTGGT	This Paper	N/A
MYB	ACAAGCATGCGTTGCCACTTCTT	CAAACACAGGATCCATGCAACA	This Paper	N/A
ULBP2	CATCAAAATGCGCATTCATC	CCAGATCAGAAATGGGTGCTT	This Paper	N/A
ESR	CTCTCTCTCTGCGCATTCAG	GAAGCCCAGAGATGCCTCAC	Cathy and S	N/A
SCAF8	TGGCACTAAATGAGGAAAAA	TGCCTTCATGGAGTTTACAT	This Paper	N/A
MAS1	CGGTCACAGTTGAGACTGTCGTCT	TTAGTATCTCATGCATATGGGATGAG	Theune et a	N/A
SLC44A4	GTCCTCTCCCACAAAAAAGTCC	TCAGCCTTCCCAGTTCC	This Paper	N/A
TRAM2	CTTATTTGGTGCCATGCAATG	AGGGTGAGAAGGACCTTTAAGG	This Paper	N/A
GPCR6	CGTGTCTCTCACCAACACC	AGCTCCACATCCTGAACACC	This Paper	N/A
ARG1	GTACTATGTGTCCATGTCATTC	TTAGTAGTTTAAAGCAGGACGTT	This Paper	N/A

**Supplementary Table 2, related to Key Resource Table and STAR methods. Cloning and CRISPR primers.**

Reagent or Resource	Source	Identifier
BTN2A-IgVgeno CRISPR Fwd (GCAGTGTTTGTGTATAAAGG)	This Paper	N/A
BTN2A-IgVgeno CRISPR Rev (CCTTTATACACAAACTGC)	This Paper	N/A
BTN2A1geno 49FCRISPR Fwd (AATGAGGGGCCATGAAGACG)	This Paper	N/A
BTN2A1geno 49FCRISPR Rev (CGTCTTCATGGCCCTCATT)	This Paper	N/A
BTN2A2geno 343CRISPR Fwd (TGAGGCCATCTACGCTCG)	This Paper	N/A
BTN2A2geno 343CRISPR Rev (CGAGGCGTAGGATGGCCTCA)	This Paper	N/A
pMIM-BTN2A1Fwd (CTTCTCTAGGCGCCGGAATTCGCCGCCACCATGGAATCAGCTGCTGCC)	This Paper	N/A
pMIM-BTN2A1Rev (TCCGCTAGCTACGTAAGATCTCTATAGGCTCTGGTGACC)	This Paper	N/A
pMIG-BTN2A2Fwd (AAATGAATTCGCCGCCACCATGGAACCAGCTGCTGCTC)	This Paper	N/A
pMIG-BTN2A2Rev (AAATAGATCTCTATAGGCTCTGGTGGTCC)	This Paper	N/A
BTN2A1IgV Geno Fwd (CCCTTTGTGAAACAGCCAG)	This Paper	N/A
BTN2A1IgV Geno Rev (CCAAAGTCACACTGTGCCAA)	This Paper	N/A
BTN2A2IgV Geno Fwd (CTGTAGCCTTGGAAATATCTGCTTC)	This Paper	N/A
BTN2A2IgV Geno Rev (CTTTGTGCAAGAGGAAGCCACTTT)	This Paper	N/A
BTN2A1 49F Geno Fwd (ATGGGGCCTATAGGTGGACCG)	This Paper	N/A
BTN2A1 49F Geno Rev (AGGGTGTGTTGATAGAGCAGGACATG)	This Paper	N/A
BTN2A2 343 Geno Fwd (AATATCTGCTTCTTTTCATCCCTG)	This Paper	N/A
BTN2A2 343 Geno Rev (GTGAAACCTATAGCTTAGCAGGGC)	This Paper	N/A
BTN2A1 R65A Fwd (AGGACATGGAGGTGGCGTGGTCCCGTCTC)	This Paper	N/A
BTN2A1 R65A Rev (GAGACCGGACACCACGCCACTCCATGTCTCT)	This Paper	N/A
BTN2A1 R124A Fwd (AAAACGGCCTACGCCCTGTACTTCCAAG)	This Paper	N/A
BTN2A1 R124A Rev (CTTGGAAAGTAACAGGCGTAGGTGCCGTTTT)	This Paper	N/A
BTN2A1 Y126A Fwd (GCACCTACCCTGTGCCTTCCAAGAAGGCA)	This Paper	N/A
BTN2A1 Y126A Rev (TGCCTTCTTGGAAAGCACAGCGTAGGTGC)	This Paper	N/A
BTN2A1 R124E Fwd (CACCTACGAATGTTACTTCCAAGAAGGCAGG)	This Paper	N/A
BTN2A1 R124E Rev (GGAAGTAACATTCTGTAGGTGCCGTTTTCC)	This Paper	N/A
BTN2A1 C247W Fwd (TGTGTCTCCCTGGGCAGTGGCCCTGCCTATC)	This Paper	N/A
BTN2A1 C247W Rev (GGGCCACTGCCAGGGAGACACACTGGGC)	This Paper	N/A
pIZ-BTN2A1 Fwd (GACCATCTCTAGAGAATTCGCCGCCACCATGGAATCAGCT)	This Paper	N/A
pIZ-BTN2A1-HA Rev (GGGAGGGAGAGGGGATCCCTAAGCGTAATCTGGAACATCGTATGGGTATGCTGATCCTAG GCTCTGGTGGGTCCCACT)	This Paper	N/A
pIZ-BTN2A2 Fwd (GACCATCTCTAGAGAATTCGCCGCCACCATGGAACCAG)	This Paper	N/A
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pIZ-BTN2A1Ldr-HA Rev (TGTGTGCACTGGTCTCAGGATCAGCATACCCATACGATGTTCCAGATTACGCTGCTGGATC CACCATGGTAATTGAATTCAATATTTAAATAATAGATct)	This Paper	N/A
pIZ-HA-BTN2A1 Fwd (CCAGATTACGCTGCTGGATCCGCCAGTTTATTGTCTGG)	This Paper	N/A
pIZ-BTN2A1 Rev (TATTATTTAAATATTGAATTCCTATAGGCTCTGGTGGTCC)	This Paper	N/A
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pIZ-BTN2A2-HA Rev (TATTATTTAAATATTGAATTCCTATAGGCTCTGGTGGTCCC)	This Paper	N/A
piH-BTN3A1Ldr-FLAG Fwd (GGATCAGCACAATTGGCCACCATGAAAATGGCAAGTTTCTCTGGCCTTCTTCTGCTCAACT TTCGTGTCTGCCTCCTTTTGTCTCAGCTGCTCATGCCTCACTCAGCTGGATCAGCAGACTAC AA)	This Paper	N/A
piH-BTN3A1Ldr-FLAG-Rev (TCCTGATGCAGATCTATTATTTAAATATTGAATTCATTTACCATGGTGGATCCAGCCTTGT CATCGTCTCCTTGTAGTCTGCTGATCC)	This Paper	N/A
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MOP Vy9-E70R Fwd (CAAATTTAGGGTGGATAGGATACCTGAAACGTC)	This Paper	N/A
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MOP Vy9-E70K Fwd (CAAATTTAAGGTGGATAGGATACCTGAAACGTC)	This Paper	N/A
MOP Vy9-E70K Rev (TCCTATCCACTTAAATTTGCCTGACGGAATG)	This Paper	N/A