Science Advances

advances.sciencemag.org/cgi/content/full/6/13/eaaz2129/DC1

Supplementary Materials for

SKP1 drives the prophase I to metaphase I transition during male meiosis

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Published 25 March 2020, *Sci. Adv.* **6**, eaaz2129 (2020) DOI: 10.1126/sciadv.aaz2129

This PDF file includes:

Fig. S1. Localization of SKP1 to the SC in oocytes.

Fig. S2. Inactivation of the Rec8 gene by CRISPR-Cas9-mediated genome editing.

Fig. S3. Targeted inactivation of the *Skp1* gene.

Fig. S4. H1t expression and CREST localization in WT and Skp1^{cKO} spermatocytes.

Fig. S5. Localization of SKP1 in WT and $Trip13^{Gt/Gt}$ pachytene spermatocytes.

Fig. S6. Precocious chromosome desynapsis and persistence of RPA2 foci in E18.5 *Skp1*^{cKO} oocytes.

Fig. S7. Expression and localization of PLK1, BUB1, CENP-C, and CCNB1 in spermatocytes.

SUPPLEMENTARY MATERIALS



Fig. S1. Localization of SKP1 to the SC in oocytes. Surface spread nuclei of wild type E18.5 oocytes were immunostained with anti-SKP1 and SYCP2 antibodies. (**A**) Leptotene. (**B**) Late zygotene or early pachytene. (**C**) Pachytene. (**D**) Diplotene. Arrowheads (**B**) indicate the absence of SKP1 in unsynapsed regions on the synaptonemal complexes at the late zygotene /early pachytene stage. Arrows (**D**) designate the localization of SKP1 to the synapsed regions at the diplotene stage. Scale bar, 10 μm.



Fig. S2. Inactivation of the *Rec8* **gene by CRISPR-Cas9–mediated genome editing.** The sequences and locations of the sgRNAs are shown. In the *Rec8* knockout, exons 2- 19 are deleted.



Fig. S3. Targeted inactivation of the *Skp1* gene. (A) Schematic diagram of the *Skp1* targeting strategy. The mouse *Skp1* gene consists of 6 exons and encodes a protein of 163 amino acids. In the conditional *Skp1*^f allele, exons 3-5 are floxed. Exons 3-5 encode residues 33-153. Deletion of exons 3-5 (359 nt) not only removes the majority of the protein but also results in a frameshift in the resulting *Skp1* mutant transcript. HyTK is a double selection marker and enables hygromycin-positive selection and thymidine kinase-negative selection. (B) Histological analysis of *Skp1*^{f/+} control and *Skp1*^{f/-} *Ddx4*-Cre testes at postnatal day 6 (PND6) through 6 months. *Ddx4*-Cre is germ cell-specific but constitutively active (*20*). Note the complete loss of germ cells in *Skp1*^{fl/-} *Ddx4*-Cre testes at PND10 and beyond. Scale bar, 25 μ m. (C) Lack of SKP1 signal on the synaptonemal complexes in *Skp1*^{cKO} pachytene spermatocytes. Scale bar, 10 μ m.



Fig. S4. H1t expression and CREST localization in WT and $Skp1^{cKO}$ spermatocytes. (A) Staging of pachynema by H1t expression. In wild type, H1t is absent in early pachynema, appears in mid-pachynema, and increases its abundance in late pachynema and diplonema. In $Skp1^{cKO}$ spermatocytes, early pachynema with no unsynapsed ends are H1t-negative, some pachynema with split ends are H1t-negative (Y1 pachynema) and thus at the early pachytene stage, and the remaining pachynema with split ends are H1t-positive (Y2 pachynema) and thus are at the midpachytene stage. However, Y2 pachynema could also be considered as early diplonema. (B) Count of H1t-negative (Y1) and H1t-positive (Y2) $Skp1^{cKO}$ pachytene spermatocytes with unsynapsed split ends. (C) CREST marks centromeres. Pachytene spermatocytes were examined by surface nuclear spread analysis. Arrows indicate some of the desynapsed chromosome ends. (D) The centromeric ends are preferentially desynapsed in $Skp1^{cKO}$ pachytene spermatocytes. Scale bars, 10 µm.



Fig. S5. Localization of SKP1 in WT and *Trip13*^{Gt/Gt} **pachytene spermatocytes.** Surface nuclear spread analysis of wild type and *Trip13*^{Gt/Gt} spermatocytes was performed by immunostaining of SKP1 and SYCP3. Scale bar, 10 μm.



Fig. S6. Precocious chromosome desynapsis and persistence of RPA2 foci in E18.5 *Skp1*^{cKO} oocytes. *Skp1*^{cKO} oocytes were collected from E18.5 *Skp1*^{fl/-} *Ddx4*-Cre embryos. (A) Precocious chromosome desynapsis in *Skp1*^{cKO} oocytes. Surface spread nuclei of E18.5 oocytes were immunostained with anti-SYCP1 and anti-SYCP2 antibodies. Pachytene and diplotene oocytes are shown. Graph on the right shows increased percentage of diplotene spermatocytes in E18.5 *Skp1*^{fl/-} *Ddx4*-Cre embryos. Data were from three independent experiments. (B) Persistence of RPA2 foci in *Skp1*^{cKO} diplotene oocytes. Surface spread nuclei of E18.5 oocytes were immunostained with anti-RPA2 and anti-SYCP3 antibodies. Graph on the right shows the count of RPA2 foci. Scale bars, 10 μ m.



