Novel and potent dopamine D₂ receptor (D₂R) Go-protein biased agonists

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CHEMISTRY

PAM 2 was prepared by amide coupling between racemic 2-methylindoline and thionyl chloride activated benzo[d]thiazole-2-carboxylic acid, as reported in Scheme 1A. Compound 9 (Scheme 1B) was also prepared by reacting benzo[d]thiazole-2-carboxylic acid with 4-aminobutanol and 1,1'-carbonyldiimidazole (CDI) to afford the intermediate amide alcohol 23. 23 was then converted into the corresponding bromine analogue 24 via an Appel reaction before being inserted in the N-5 position of sumanirole¹⁻² by nucleophilic substitution, carried out in a sealed vessel, at a high temperature and in the absence of base. The absence of organic or inorganic base was necessary to minimize intra-molecular cyclization of 24 and/or the more favorable sumanirole N-1 alkylation.² Similarly, 10 was prepared starting from (\pm) -2-methylindoline to obtain the intermediate 25 by amidation with 4-bromobutanoic acid followed by sumanirole N-5 alkylation (Scheme 1C). Sumanirole N-5 alkylation with bromoalkane intermediates produced moderate to low yields, thus reductive amination was used to successfully obtain N-5 regiospecificity in almost quantitative yields. 11 (diastereometric mixture) and its (R,S)-12 and (R,R)-13 single enantiomers were prepared, as depicted in Scheme 1D, starting from the commercially available (\pm) -, (S)- or (R)-2-methylindoline, respectively. The indoline nitrogen was reacted with phosgene and 4-aminobutanol to afford the alcohol intermediates, which were then oxidized to the corresponding aldehydes with Dess-Martin periodinane (DMP) and conjugated to sumanirole in the N-5 position via sodium triacetoxyborohydride-mediated reductive amination. Diastereomeric excess and purity of the final resolved diastereoisomers were determined via chiral HPLC, in comparison with the diastereomeric mixture (HPLC chromatograms; Figure S1), using methods described in detail in the experimental section. Finally, compound 8, presenting

the benzothiazole ring substituted in position 5 as the SP, was prepared following Scheme 1E. Amidation of the activated benzo[d]thiazole-5-carboxylic acid, followed by oxidation of the terminal alcohol to the corresponding aldehyde and consequent reductive amination led to the desired product. In Schemes 2 and 3 the synthetic routes to obtain the bivalent analogues embedding the 1,2,3,4 tetrahydroquinoline PP are shown. Specifically, Cbz- protected (R)-3amino-1-methoxy-3,4-dihydroquinolin-2(1H)-one, a key intermediate isolated from the enantiospecific sumanirole synthesis,¹ was deprotected and dialkylated with two *n*-propyl substituents (35) before being reduced in the presence of borane dimethylsulfide complex and subsequently N-substituted with N-(4-bromobutyl)-1H-indole-2-carboxamide¹ to obtain 14. In order to synthesize the mono-propyl analogue 17, 34 was first reacted with propionyl chloride and then reduced to obtain the *n*-propyl secondary amine intermediate 38 which was readily alkylated with N-(4-bromobutyl)-1H-indole-2-carboxamide. The benzothiazole- and 2methylindoline-containing analogues depicted in Scheme 3 were prepared from the intermediates 23 and 26, respectively. 23 was oxidized to the corresponding aldehyde and from the subsequent reductive amination with N-methyl-1,2,3,4-tetrahydroquinolin-3-amine¹ both mono- and disubstituted products 15 and 18 were isolated. Compound 16 was obtained from alkylation of Nmethyl-1,2,3,4-tetrahydroquinolin-3-amine with 40 under basic conditions.

Compound 19, presenting a *trans*-cyclopropyl ring in the connecting linker, was prepared according to the multistep approach described in Scheme 4. The preparation of the *trans*-cyclopropyl linker was slightly modified from what was previously reported,³ allowing an overall higher yield synthesis with intermediates that were easily analyzed by chiral HPLC (methods in the experimental section) to confirm the retention of stereochemistry throughout the entire synthetic route (chiral HPLC analyses of intermediate **42** and final product **19** in Figures

S2 and S3). In detail, commercially available predominantly trans ethyl 2-formylcyclopropane-1-carboxylate was reacted first with nitromethane in the presence of lithium tert-butoxide, then with methanesulfonyl chloride and triethylamine to generate the pure *trans* nitro-olefin 42 in almost quantitative yield, over two steps. 42 was analyzed in chiral HPLC to confirm the formation of the sole trans racemic mixture (Figure S2) in 50:50 enantiomeric ratio. Simultaneous reduction of both the nitro-olefin and the ester functional groups with lithium aluminum hydride yielded intermediate 43, which was then reacted with indole-2-carboxylic acid to afford 44. Both 43 and 44 showed spectroscopic data identical to the same intermediates prepared previously, via a different synthetic approach.³ Finally, oxidation of the primary alcohol and reductive amination with sumanirole led to final product 19. 19 was analyzed in chiral HPLC to confirm the lack of racemization during the synthetic steps and the formation of the pure trans-cyclopropyl racemic mixture. Indeed, the chiral HPLC chromatogram (Figure S3) shows the existence of only the two cyclopropyl *trans* enantiomers (enantiomeric ratio 47:53), despite the potential formation of eight total stereoisomers that could have occurred in the case of an undesired racemization at the cyclopropyl or at the **sumanirole** scaffold. Thus, these data confirmed the stereospecificity of this alternative synthetic route. 20, differing from 19 only due to the shift of the cyclopropyl ring one methylene unit farther from the PP and closer to the SP, was prepared as described in Scheme 5. Trans-48 was obtained following the procedure by Kumar et al.,³ which described in detail the isolation of the *trans* cycloproyl methanamino-Opyranil protected intermediate 46. Finally, alcohol 48 was oxidized and coupled with sumanirole to obtain 20. 20 was analyzed by chiral HPLC, analogously to 19, to confirm the lack of racemization and the presence of only the *trans*-cyclopropyl enantiomeric mixture (Figure S4, enantiomeric ratio 49:51).

 $1^{1,2}$ (G-protein D₂R biased full agonist) was resynthesized, in high yield, via a slightly modified synthetic procedure, where the final coupling step between **sumanirole** in *N*-5 position and the *N*-(4-bromobutyl)-1H-indole-2-carboxamide was obtained by reductive amination in presence of the corresponding aldehyde (*N*-(4-oxobutyl)-1*H*-indole-2-carboxamide), sodium triacetoxyborohydride, and catalytic amount of acetic acid. All the obtained spectroscopic data where consistent with the previously published ones.¹

Experimental Methods

All chemicals and solvents were purchased from chemical suppliers unless otherwise stated, and used without further purification. All melting points were determined on an OptiMelt automated melting point system and are uncorrected. The ¹H and ¹³C NMR spectra were recorded on a Varian Mercury Plus 400 instrument. Proton chemical shifts are reported as parts per million (\delta ppm) relative to tetramethylsilane (0.00 ppm) as an internal standard. Coupling constants are measured in Hz. Chemical shifts for ¹³C NMR spectra are reported as parts per million (δ ppm) relative to deuterated CHCl₃ or deuterated MeOH (CDCl₃ 77.5 ppm, CD₃OD 49.3 ppm). Chemical shifts, multiplicities and coupling constants (J) have been reported and calculated using Vnmrj Agilent-NMR 400MR or MNova 9.0 software. Gas chromatography-mass spectrometry (GC/MS) data were acquired (where obtainable) using an Agilent Technologies (Santa Clara, CA) 6890N GC equipped with an HP-5MS column (cross-linked 5% PH ME siloxane, 30 m \times 0.25 mm i.d. \times 0.25 µm film thickness) and a 5973 mass-selective ion detector in electron-impact mode. Ultrapure grade helium was used as the carrier gas at a flow rate of 1.2 mL/min. The injection port and transfer line temperatures were 250 and 280 °C, respectively, and the oven temperature gradient used was as follows: the initial temperature (100 °C) was held

for 3 min and then increased to 295 °C at 15 °C/min over 13 min, and finally maintained at 295 °C for 10 min. All column chromatography was performed using a Teledyne Isco CombiFlash RF flash chromatography system and silica gel (Merck, 230-400 mesh, 60Å) or preparative thin layer chromatography (silica gel, Analtech, 1000 μ m). Microanalyses were performed by Atlantic Microlab, Inc. (Norcross, GA) and agree with $\pm 0.4\%$ of calculated values (table S1). HRMS (mass error less than 5 ppm) and MS/MS fragmentation analysis were performed on a LTQ-Orbitrap Velos (Thermo-Scientific, San Jose, CA) coupled with an ESI source in positive ion mode to confirm the assigned structures and regiochemistry. HPLC analysis was performed using an Agilent system coupled with DAD (Diode Array Detector). Separation of the analyte, purity and enantiomeric/diastereomeric excess determinations were achieved at 40 °C using one of the following different methods (the specific method used for each compound is mentioned in the detailed reaction description). Method A): Agilent Poroshell 120 EC-C18 (4.6 x 50 mm, 2.7 µm) column. The mobile phase used (0.5 mL/min flow rate) was composed of 0.1% acetic acid in H₂O and acetonitrile with gradient elution, starting with 10% (organic) linearly increasing to 60% up to 5 min, maintaining at 60% (5-15 min) and re-equilbrating to 10%. The total run time was 20 min. Method B): Chiralcel OD (Daicel Corporation CPI Company) column (4.6 x 50 mm, 3 µm). The mobile phase used (0.5 mL/min flow rate) was composed of 0.1% diethylamine in 2-propanol and hexanes with gradient elution, starting with 10% of 2-propanol linearly increasing to 20% up to 5 min, maintaining at 20% (5-20 min) and re-equilibrating to 10%. The total run time was 25 min. Method C): Chiralcel OZ-H (Daicel Corporation CPI Company) column (4.6 x 250 mm, 5 μ m). The mobile phase used (0.5 mL/min flow rate) was composed of 10% 2-propanol in hexanes with isocratic elution. The total run time was 60 min. Method D): Chiralpak AD-H (Daicel Corporation CPI Company) column (4.6 x 250 mm, 5 µm). The mobile

phase used (0.5 mL/min flow rate) was composed of 0.1% diethylamine in 2-propanol and hexanes with gradient elution, starting with 20% of 2-propanol linearly increasing to 30% up to 15 min, maintaining at 30% (15-160 min) and re-equilibrating to 20%. The total run time was 170 min. **Method E)** Chiralpak AD-H (Daicel Corporation CPI Company) column (4.6 x 250 mm, 5 μ m). The mobile phase used (1.0 mL/min flow rate) was composed of 40% 2-propanol (in presence of 0.1% diethylamine) in hexanes with isocratic elution. The total run time was 60 min. Optical rotations were determined using a Jasco DIP-370 polarimeter. Unless otherwise stated, all the compounds were evaluated to be >95% pure on the basis of combustion analysis, NMR, GC-MS, and HPLC-DAD.

Scheme 1.



a) SOCl₂, DCM, DMF, TEA; b) 1,1-CDI, 4-aminobutanol, THF; c) PPh₃, CBr₄, ACN; d) **sumanirole**, DMF, 100 °C, sealed vessel; e) SOCl₂, DCM, TEA, 4-bromobutanoic acid; f) COCl₂, 4-aminobutanol, THF, TEA; g) DMP, DCM, from 0 °C to RT; h) **sumanirole**, Na(OAc)₃BH, AcOH, DCE.

Benzo[d]thiazol-2-yl(2-methylindolin-1-yl)methanone (2). To a solution of benzo[d]thiazole-2-carboxylic acid (1.0g, 5.6 mmol) in dichloromethane (50 mL) at room temperature, was added a solution of thionyl chloride (1.0g, 1.6 mL, 8.4 mmol) in *N*,*N*-dimethylformamide (2 mL).

Reaction was heated to reflux for 3 hours, before cooling to room temperature and concentrating under reduced pressure to afford the crude acid chloride. Residue was dissolved in DCM, and to the stirring solution was added 2-methylindoline (0.74g, 5.6 mmol), followed by triethylamine (1.6 ml, 11.5 mmol), and the reaction allowed to stir overnight at room temperature. Upon completion, reaction mixture was then washed with a 10% aqueous solution of Na₂CO₃. The organic phase was dried over anhydrous Na₂SO₄. Filtration and removal of solvent under vacuum afforded the crude product which was subsequently purified via Combiflash chromatography, eluting with 10% EtOAc in hexanes to afford 2 as a white solid. After washing with a 10% solution of diethyl ether in hexanes, product was dried via vacuum filtration to a constant weight (0.246 g/ 15% yield). ¹H NMR (400 MHz, CDCl₃) δ 8.37 (d, J = 8.0 Hz, 1H), 8.18 - 8.09 (m, 1H), 8.03 - 7.95 (m, 1H), 7.53 (dtd, J = 20.6, 7.2, 1.3 Hz, 2H), 7.29 (d, J = 7.3Hz, 2H), 7.14 (t, J = 7.4 Hz, 1H), 6.09 (br s, 1H), 3.52 (dd, J = 15.6, 8.8 Hz, 1H), 2.79 (d, J =15.6 Hz, 1H), 1.40 (d, J = 6.3 Hz, 3H) ppm. ¹³C NMR (101 MHz, CDCl₃) δ 158.64, 153.66, 141.76, 136.53, 131.64, 127.50, 126.75, 126.50, 125.17, 124.85, 121.91, 118.95, 56.70, 36.60, 22.49 ppm. GC-MS (EI) 294.1 m/z (M+). Anal. (C₁₇H₁₄N₂OS ⁺0.25H₂O) C, H, N. M.p.: 133-134 °C

N-(4-Hydroxybutyl)benzo[d]thiazole-2-carboxamide (23). To a stirring solution of benzo[d]thiazole-2-carboxylic acid (1.0g, 5.6 mmol) in tetrahydrofuran (20 mL) at room temperature, was added 1,1'-carbonyldiimidazole (1.0g, 6 mmol), followed by dropwise addition of *N,N*-dimethylformamide (2 mL) to solubilize any undissolved solids. 4-aminobutan-1-ol (0.5g, 0.517 mL, 5.6 mmol) was subsequently added dropwise into solution, and the reaction allowed to stir overnight at room temperature. Reaction was concentrated under vacuum to

afford the crude product which was purified via Combiflash chromatography, eluting with 100% EtOAc to afford the desired compound as a viscous yellow oil **(0.586 g/ 42% yield).** ¹H NMR (400 MHz, CDCl₃) δ 7.94 (dd, J = 8.2, 1.2 Hz, 1H), 7.85 (dd, J = 8.1, 1.2 Hz, 1H), 7.76 (t, J = 6.2 Hz, 1H), 7.41 (dtd, J = 25.6, 7.6, 1.2 Hz, 2H), 3.66 (t, J = 6.2 Hz, 2H), 3.48 (q, J = 6.6 Hz, 2H), 3.17 (s, 1H), 1.80 – 1.57 (m, 4H) ppm.

N-(4-Bromobutyl)benzo[d]thiazole-2-carboxamide (24). To a stirring solution of **23** (0.5g, 2 mmol) in anhydrous acetonitrile (10 mL) at room temperature and under inert atmosphere, was added triphenylphosphine (0.8 g, 3 mmol), followed by carbon tetrabromide (1.0g, 3 mmol). Reaction was heated to reflux, and allowed to proceed for 4 hours. Upon completion of the reaction, solvent was stripped under vacuum and precipitated solids were resuspended in a 2N aqueous solution of NaOH. This aqueous solution was subsequently extracted 3x with EtOAc and the combined organic fractions were then dried over anhydrous Na₂SO₄. Filtration and removal of solvent under reduced pressure afforded the crude alkyl bromide, which was then purified via Combiflash chromatography, eluting with EtOAc:Hexanes (2:8) to afford the desired product as a pale red solid (0.381 g / 61% yield). ¹H NMR (400 MHz, CDCl₃) δ 8.05 (ddt, *J* = 8.2, 1.3, 0.6 Hz, 1H), 7.96 (ddd, *J* = 7.9, 1.4, 0.7 Hz, 1H), 7.54 (ddd, *J* = 8.4, 7.1, 1.3 Hz, 1H), 7.50 – 7.42 (m, 2H), 3.60 – 3.42 (m, 4H), 2.06 – 1.92 (m, 2H), 1.90 – 1.79 (m, 2H) ppm.

(R)-N-(4-(Methyl(2-oxo-1,2,5,6-tetrahydro-4H-imidazo[4,5,1-ij]quinolin-5-

yl)amino)butyl)benzo[d]thiazole-2-carboxamide (9). **24** (0.124 g, 0.4 mmol) and **sumanirole** (0.100 g, 0.4 mmol) were charged to a sealed vessel and solubilized in *N*,*N*-dimethylformamide (10 mL). Reaction was heated to approximately 100°C, and allowed to proceed with vigorous

stirring for two hours. Upon completion of the reaction, solvent was concentrated under vacuum, and the remaining residue purified by preparatory thin-layer chromatography, eluting with MeOH:chloroform (5:95) to afford the desired product (**0.051g**, **29% yield**) ¹H NMR (400 MHz, CDCl₃) δ 9.94 (s, 1H), 8.03 – 7.84 (m, 3H), 7.54 – 7.40 (m, 2H), 7.01 – 6.79 (m, 3H), 4.26 – 4.17 (m, 1H), 3.61 – 3.41 (m, 3H), 3.26 (tt, *J* = 9.9, 4.7 Hz, 1H), 3.04 – 2.79 (m, 2H), 2.71 – 2.56 (m, 2H), 2.41 (s, 3H), 1.82 – 1.56 (m, 4H) ppm. ¹³C NMR (101 MHz, CDCl₃) δ 164.23, 159.93, 154.93, 152.88, 137.03, 127.30, 126.69, 126.55, 126.07, 124.12, 122.36, 121.41, 119.73, 118.46, 107.33, 57.55, 53.44, 39.85, 38.15, 27.19, 26.92, 25.30 ppm. HRMS (ESI positive) calculated 436.1802; found 436.1814 (+H⁺). [α]²³_D: 0.83 (0.120 g/100 mL in MeOH). Anal. (C₂₃H₂₅N₅O₂S · 1.5H₂O) C, H, N. M.p.: 63 °C (dec.)

4-Bromo-1-(2-methylindolin-1-yl)butan-1-one (25). To a stirring solution of 4-bromobutanoic acid (2.5 g, 1.6 mL, 15 mmol) in dichloromethane (50 mL) at room temperature, was added thionyl chloride (2.7 g, 1.65 mL, 22.7 mmol) and the reaction heated to reflux for three hours. Reaction mixture was then stripped of solvent under vacuum, and the residue resuspended with the dropwise addition of a solution of 2-methylindoline (2.0 g, 15 mmol) in dichloromethane (50 ml). Once addition was complete, triethylamine (5 mL, 35.8 mmol) was added to the reaction mixture, and stirred overnight at room temperature. The reaction mixture was subsequently decanted into a separatory funnel and washed with a 10% Na₂CO₃ solution, followed by a 10% HCl solution. The organic phase was then dried over anhydrous Na₂SO₄, before filtering and concentrating under vacuum to afford the crude bromide. Crude material was then purified by Combiflash chromatography, eluting with EtOAc:Hexanes (2:8) to afford the desired product (**3.56 g, 84% yield).** ¹H NMR (400 MHz, CDCl₃) δ 8.15 (d, *J* = 8.1 Hz, 1H), 7.20 (t, *J* = 7.6 Hz,

2H), 7.02 (t, *J* = 7.4 Hz, 1H), 4.58 – 4.49 (m, 1H), 3.74 – 3.66 (m, 2H), 3.41 (dd, *J* = 16.0, 9.0 Hz, 1H), 2.69 (dtd, *J* = 29.6, 14.9, 14.3, 6.6 Hz, 3H), 2.31 – 2.17 (m, 2H), 1.31 (d, *J* = 6.4 Hz, 3H) ppm.

(5R)-5-(Methyl(4-(2-methylindolin-1-yl)-4-oxobutyl)amino)-5,6-dihydro-4H imidazo[4,5,1*ij]quinolin-2(1H)-one (10).* Compound 25 (0.112 g, 0.4 mmol) and sumanirole (0.100 g, 0.4 mmol) were charged to a sealed vessel and solubilized in N,N-dimethylformamide (10 mL). Reaction was heated to approximately 100°C, and allowed to proceed with vigorous stirring for two hours. Upon completion of the reaction, solvent was concentrated under vacuum, and the remaining residue purified by preparatory thin-layer chromatography, eluting with MeOH:chloroform (10:90) to afford the desired product as a mixture of diastereoisomers (0.04 g, **20% yield).** ¹H NMR (400 MHz, CDCl₃) δ 9.20 (s, 1H), 8.16 (d, J = 7.7 Hz, 1H), 7.19 (d, J =7.6 Hz, 2H), 7.08 - 6.88 (m, 2H), 6.88 - 6.78 (m, 2H), 4.52 (s, 1H), 4.16 (dd, J = 12.0, 4.3 Hz, 1H), 3.62 - 3.45 (m, 1H), 3.42 (d, J = 18.5 Hz, 1H), 3.25 - 3.19 (m, 1H), 2.98 - 2.80 (m, 3H), 2.79 – 2.61 (m, 4H), 2.41 (s, 3H), 2.00 – 1.91 (m, 2H), 1.29 (br s, 3H) ppm. An analytical sample was converted into the corresponding maleate salt. ¹³C NMR (101 MHz, CD₃OD) δ 171.33, 168.77, 133.61, 126.76, 126.70, 126.59, 124.55, 123.95, 122.16, 122.13, 122.09, 119.40, 118.03, 114.26, 107.98, 107.75, 65.46, 58.48, 56.16, 54.43, 38.48, 38.19, 37.33, 37.32, 36.93, 35.75, 32.15, 26.14, 22.77, 21.29, 20.04, 19.67, 14.00 ppm. HPLC-DAD Method A (Rt: 6.099 min; purity 93.3%). HRMS (ESI positive) calculated 405.2285; found 405.2295 $(+H^+)$.

N-(4-Hydroxybutyl)-2-methylindoline-1-carboxamide (26). To a stirring solution of 2-methylindoline (2.0 g, 15 mmol) and triethylamine (~5 eq.) in anhydrous tetrahydrofuran (50

mL) cooled to 0 °C and under inert atmosphere, was added dropwise a solution of phosgene (6 mL, 15% wt. in toluene) in anhydrous tetrahydrofuran (10 mL). After stirring for 1 hour at 0°C, a solution of 4-aminobutan-1-ol (1.3 g, 1.344 mL, 15 mmol) and triethylamine (~5 eq.) in tetrahydrofuran (10 mL) was added dropwise into the reaction, and allowed to warm to room temperature overnight with stirring. Reaction was subsequently concentrated under vacuum and resuspended in H₂O before extracting repeatedly with dichloromethane. Organic fractions were then combined together and dried over anhydrous Na₂SO₄. Filtration and solvent removal under vacuum afforded the crude alcohol, which was purified via Combiflash chromatography, eluting with 100% EtOAc to afford the desired product as a yellow oil (**0.850** g / **23% yield**). ¹H NMR (400 MHz, CDCl₃) δ 7.67 (d, *J* = 8.0 Hz, 1H), 7.14 (d, *J* = 7.8 Hz, 2H), 6.90 (td, *J* = 7.4, 1.0 Hz, 1H), 5.10 (s, 1H), 4.38 (ddt, *J* = 10.7, 6.5, 3.2 Hz, 1H), 3.69 (t, *J* = 5.8 Hz, 2H), 3.36 (ddd, *J* = 11.8, 9.4, 6.3 Hz, 3H), 2.60 (dd, *J* = 15.6, 1.9 Hz, 1H), 2.03 (s, 1H), 1.66 (pd, *J* = 6.7, 1.6 Hz, 4H), 1.32 – 1.21 (m, 3H) ppm.

(S)-N-(4-Hydroxybutyl)-2-methylindoline-1-carboxamide (27). Compound was prepared following the synthetic procedure described for 26 starting from (2S)-methylindoline. Observed spectroscopic data collected for 27 identical to that observed for 26.

(*R*)-*N*-(4-Hydroxybutyl)-2-methylindoline-1-carboxamide (28). Compound was prepared following the synthetic procedure described for 26 (2*R*)-methylindoline. Observed spectroscopic data collected for 28 identical to that observed for 26.

2-Methyl-N-(4-oxobutyl)indoline-1-carboxamide (29). To a well stirring solution of **26** (0.782 g, 3.15 mmol) in dichloromethane (32 mL), was added Dess-Martin Periodinane (1.536 g, 3.62 mmol) and the reaction stirred for 1 hour at room temperature. The reaction mixture was washed with 10% NaHCO₃ and the organic phase dried over anhydrous Na₂SO₄. Filtering and concentrating under vacuum afforded the crude aldehyde as a peach colored oil which was purified via Combiflash chromatography, eluting with 100% EtOAc to afford the desired product as a colorless oil (0.470 g, 60% yield). ¹H NMR (400 MHz, CDCl₃) δ 9.81 (s, 1H), 7.68 (d, *J* = 7.9 Hz, 1H), 7.20 – 7.07 (m, 2H), 6.91 (qd, *J* = 7.5, 1.1 Hz, 1H), 5.03 (br s, 1H), 4.43 – 4.24 (m, 2H), 3.44 – 3.30 (m, 3H), 2.66 – 2.55 (m, 2H), 1.92 (p, *J* = 6.9 Hz, 2H), 1.41 – 1.21 (m, 3H) ppm.

(S)-2-Methyl-N-(4-oxobutyl)indoline-1-carboxamide (30). Compound was prepared following the synthetic procedure described for 29, starting from 27. Observed spectroscopic data collected for 30 identical to that observed for 29.

(*R*)-2-Methyl-N-(4-oxobutyl)indoline-1-carboxamide (31). Compound was prepared following the synthetic procedure described for 29, starting from 28. Observed spectroscopic data collected for 31 identical to that observed for 29.

2-Methyl-N-(4-(methyl((R)-2-oxo-1,2,5,6-tetrahydro-4H-imidazo[4,5,1-ij]quinolin-5-

yl)amino)butyl)indoline-1-carboxamide (11). To a stirring solution of **sumanirole** (0.3 g, 1.47 mmol) in 1,2-dichloroethane (20 mL) at room temperature was added glacial acetic acid (catalytic, 5 drops), followed by the dropwise addition of **29** (0.418 g, 1.69 mmol) in 1,2-

dichloroethane (10 mL). After 15 minutes of continued stirring, NaBH(OAc)₃ (0.470 g, 2.21 mmol) was added in one portion and the reaction allowed to stir overnight at room temperature. Reaction was subsequently quenched with the addition of solid K₂CO₃ and allowed to stir for 5 minutes before filtering and concentrating under vacuum to afford the crude product as a viscous dark green oil. Crude material was purified by Combiflash chromatography, eluting with MeOH:DCM (15:85) to afford the desired product as a foamy white solid (0.46 g, 72% yield). ¹H NMR (400 MHz, CDCl₃) δ 9.24 (s, 1H), 7.66 (dq, J = 7.8, 1.1 Hz, 1H), 7.20 – 7.11 (m, 2H), 7.01 - 6.83 (m, 4H), 4.90 (t, J = 5.6 Hz, 1H), 4.47 - 4.34 (m, 1H), 4.18 (dd, J = 12.1, 4.5 Hz, 1H), 3.56 (dd, J = 12.0, 10.2 Hz, 1H), 3.49 (s, 1H), 3.44 – 3.32 (m, 3H), 3.22 (tt, J = 9.7, 4.8 Hz, 1H), 2.99 - 2.91 (m, 2H), 2.73 - 2.56 (m, 3H), 2.40 (s, 3H), 1.64 - 1.55 (m, 3H), 1.30 (dd, J =6.4, 1.6 Hz, 3H) ppm. ¹³C NMR (101 MHz, CDCl₃) δ 154.74, 154.64, 142.33, 129.93, 127.51, 127.25, 125.73, 125.19, 121.95, 121.56, 119.97, 118.55, 114.74, 107.26, 57.30, 55.19, 53.69, 40.27, 40.03, 38.06, 36.32, 27.93, 26.77, 25.04, 21.01 ppm. HPLC-DAD Method B (diastereoisomer A R_t: 10.679 min; diastereoisomer B R_f: 13.854 min. purity >99%; dr 49:51); method E (diastereoisomer A Rt: 13.992 min; diastereoisomer B Rf: 31.393 min. purity 95.9%; dr 49:51). The free base was converted into the corresponding oxalate salt. HRMS (ESI positive) calculated 434.2551; found 434.2562 (+H⁺). $[\alpha]_D^{23}$: -8.00 (0.125 g/100 mL in MeOH). Anal. (C₂₅H₃₁N₅O₂ · 1.25C₂H₂O₄ · 1.25H₂O) C, H, N M.p.: 118 °C (dec.)

(S)-2-Methyl-N-(4-(methyl((R)-2-oxo-1,2,5,6-tetrahydro-4H-imidazo[4,5,1-ij]quinolin-5-

yl)amino)butyl)indoline-1-carboxamide (12). Compound was prepared following the synthetic procedure described for **11**, starting from **30**. ¹H NMR (400 MHz, CDCl₃) δ 9.30 (s, 1H), 7.66 (d, J = 8.0 Hz, 1H), 7.14 (d, J = 7.8 Hz, 2H), 6.89 (ddd, J = 19.2, 16.5, 7.9 Hz, 4H), 4.91 (t, J = 5.7

Hz, 1H), 4.39 (q, J = 7.2 Hz, 1H), 4.17 (dd, J = 12.1, 4.2 Hz, 1H), 3.61 – 3.43 (m, 2H), 3.38 (dd, J = 15.7, 8.9 Hz, 3H), 3.20 (dq, J = 9.7, 5.1, 4.7 Hz, 1H), 2.97 – 2.90 (m, 2H), 2.63 (td, J = 15.6, 14.4, 6.2 Hz, 3H), 2.39 (s, 3H), 1.62 (d, J = 14.4 Hz, 3H), 1.30 (d, J = 6.3 Hz, 3H) ppm. ¹³C NMR (101 MHz, CDCl₃) δ 154.75, 154.64, 142.35, 129.91, 127.51, 127.36, 125.90, 125.19, 121.93, 121.42, 119.83, 118.50, 114.74, 107.21, 57.40, 55.18, 53.72, 40.32, 40.09, 38.14, 36.32, 27.94, 26.84, 25.20, 21.00 ppm. HPLC-DAD Method B (R_t: 11.410 min. purity 89.2%, de >99%). The free base was converted into the corresponding oxalate salt. [α]²³_D: 3.333 (0.120 g/100 mL). Anal. (C₂₅H₃₁N₅O₂ · 2C₂H₂O₄ · 1.5H₂O) C, H, N. M.p.: 52 °C (dec.). A sample of the oxalate salt was free based and analyzed via HPLC-DAD method E (R_t: 14.491 min. purity >99%, de >99%).

(R)-2-Methyl-N-(4-(methyl((R)-2-oxo-1,2,5,6-tetrahydro-4H-imidazo[4,5,1-ij]quinolin-5-

yl)amino)butyl)indoline-1-carboxamide (13). Compound was prepared following the synthetic procedure described for **11**, starting from **31**. ¹H NMR (400 MHz, CDCl₃) δ 8.61 (s, 1H), 7.66 (d, J = 8.0 Hz, 1H), 7.16 (t, J = 7.8 Hz, 2H), 7.01 – 6.83 (m, 4H), 4.87 (s, 1H), 4.46 – 4.34 (m, 1H), 4.17 (dd, J = 11.9, 4.3 Hz, 1H), 3.61 – 3.47 (m, 2H), 3.44 – 3.29 (m, 3H), 3.22 (dt, J = 9.9, 4.8 Hz, 1H), 2.96 (m, 2H), 2.73 – 2.57 (m, 3H), 2.40 (s, 3H), 1.63 – 1.56 (m, 3H), 1.31 (d, J = 6.3 Hz, 3H) ppm. ¹³C NMR (101 MHz, CDCl₃) δ 154.71, 154.37, 142.34, 129.92, 127.51, 127.34, 125.66, 125.19, 121.93, 121.48, 119.98, 118.54, 114.73, 107.12, 57.35, 55.19, 53.72, 40.28, 40.05, 38.08, 36.32, 27.93, 26.84, 25.11, 21.03 ppm. HPLC-DAD Method B (R_t: 14.468 min. purity 99.1%, de >99%). The free base was converted into the corresponding oxalate salt. $[\alpha]_D^{23}$: -20.0 (0.115 g/ 100 mL). Anal. (C₂₅H₃₁N₅O₂ · 1.5C₂H₂O₄ · H₂O) C, H, N. M.p.: 93 °C (dec.). A

sample of the oxalate salt was free based and analyzed via HPLC-DAD method E (R_t : 32.314 min. purity >99%, de >99%).

N-(4-Hydroxybutyl)benzo[d]thiazole-5-carboxamide (32)

Compound was prepared following the synthetic procedure described for **23** starting from benzo[d]thiazole-5-carboxylic acid (0.3 g, 1.7 mmol). The crude product was used in the next step without further purification.

N-(4-Oxobutyl)benzo[d]thiazole-5-carboxamide (33)

Compound was prepared following the synthetic procedure described for **29** starting from **32** (0.12 g, 0.5 mmol). Crude material was purified by Combiflash chromatography, eluting with 100% EtOAc to afford the desired product as yellow oil (**0.06 g, 48% yield**).¹H NMR (400 MHz, CDCl₃) δ 9.83 (s, J = 1.1 Hz, 1H), 9.07 (d, J = 2.1 Hz, 1H), 8.41 (dd, J = 78.8, 1.5 Hz, 1H), 8.15 – 7.50 (m, 2H), 6.73 (br s, 1H), 3.61 – 3.40 (m, 2H), 2.64 (td, J = 6.8, 1.1 Hz, 2H), 1.99 (p, J = 6.8 Hz, 2H).

(R)-N-(4-(Methyl(2-oxo-1,2,5,6-tetrahydro-4H-imidazo[4,5,1-ij]quinolin-5-

yl)amino)butyl)benzo[d]thiazole-5-carboxamide (8)

Compound was prepared following the synthetic procedure described for **11** starting from **sumanirole** (0.022 g, 0.1 mmol) and **33** (0.022g , 0.09 mmol). Crude material was purified by Combiflash chromatography, eluting with 10% CMA (CHCl₃:MeOH:NH₄OH 9:1:0.1) to afford the desired product as yellow oil (**69% yield**).¹H NMR (400 MHz, CDCl₃) δ 9.06 (s, 1H), 8.49 (dd, J = 1.7, 0.6 Hz, 1H), 8.15 – 7.73 (m, 3H), 7.08 – 6.64 (m, 4H), 4.14 (dd, J = 12.1, 4.2 Hz,

1H), 3.76 - 3.39 (m, 2H), 3.27 - 3.11 (m, 1H), 2.94 (d, J = 7.3 Hz, 2H), 2.65 (dt, J = 13.0, 6.6 Hz, 2H), 2.56 - 2.24 (m and s, 1H and 3H), 1.84 - 1.47 (m, 4H). ¹³C NMR (101 MHz, CDCl₃) δ 167.04, 155.27, 154.12, 153.08, 136.66, 133.45, 127.45, 125.59, 124.48, 122.13, 121.76, 121.40, 119.96, 118.41, 106.99, 57.28, 53.65, 40.04, 40.00, 38.19, 29.69, 27.25, 27.03, 25.25. The free base was converted into the corresponding oxalate salt. $[\alpha]_D^{23}$: -12.173 (0.115 g/100 mL). Anal. (C₂₃H₂₅N₅O₂S · C₂H₂O₄ · 3H₂O) C, H, N. M.p.: 113 °C (dec)

Scheme 2.



a) $Pd(OH)_2/C$, H_2 , EtOH, 50 psi, 12-24 hrs; b) propionyl chloride, DCM, TEA; c) propionaldehyde, Na(OAc)_3BH, AcOH, THF; d) $BH_3(CH_3)_2$, THF, from 0 °C to reflux; e) *N*-(4-bromobutyl)-1*H*-indole-2-carboxamide, K₂CO₃, DMF, 100 °C, sealed vessel; f) *N*-(4-bromobutyl)-1*H*-indole-2-carboxamide, DME, DIPEA, 100 °C, sealed vessel.

(R)-3-Amino-1-methoxy-3,4-dihydroquinolin-2(1H)-one (34). Benzyl (R)-(1-methoxy-2-oxo-1,2,3,4-tetrahydroquinolin-3-yl)(methyl)carbamate (6.55 g, 20 mmol) was added as a viscous amber oil into a hydrogenation vessel and solubilized with anhydrous ethanol (200 mL). Pd(OH)₂ on carbon (0.5 g, 0.712 mmol) was added carefully to the solution and gently shaken to ensure submersion in the solvent. The reaction mixture was subsequently placed in a Parr hydrogenation apparatus and pressurized under H₂ gas (50 psi). Reaction was allowed to proceed with vigorous mechanical shaking overnight at room temperature. After 24 hours, reaction was removed from hydrogenation apparatus and the reaction mixture filtered through celite and concentrated under vacuum to afford the crude material as a viscous oil. Crude product was purified by Combiflash chromatography, eluting with MeOH:DCM (15:85) to afford the *N*-methoxylated lactam as the major product (**3.6 g, 93.3% yield**). ¹H NMR (400 MHz, CDCl₃) δ 7.39 – 7.23 (m, 2H), 7.20 (m, 1H), 7.11 – 7.01 (m, 1H), 3.91 (s, 3H), 3.46 (dd, *J* = 3.1, 1.0 Hz, 1H), 3.10 (dd, *J* = 15.3, 6.0 Hz, 1H), 2.85 (t, *J* = 14.3 Hz, 1H), 2.06 (br s, 2H) ppm.

(*R*)-*N*-(1-Methoxy-2-oxo-1,2,3,4-tetrahydroquinolin-3-yl)propionamide (36). To a well stirring solution of **34** (5.5 g, 18.5 mmol) in dichloromethane (40 mL) at room temperature, was added dropwise a solution of propionyl chloride (4.0 g, 3.77 mL, 24 mmol) in dichloromethane (10 mL). Reaction was allowed to proceed over the following 12 hours at this temperature. Reaction mixture was then concentrated under vacuum and purified by Combiflash chromatography, eluting with MeOH:Chloroform (1:99) to afford the desired product (4.5 g, 97% yield). ¹H NMR (400 MHz, CDCl₃) δ 7.32 – 7.22 (m, 1H), 7.19 (dd, *J* = 8.1, 1.2 Hz, 2H), 7.09 (m. 1H), 6.61 (br s, 1H), 4.58 (ddd, *J* = 14.3, 6.1, 5.2 Hz, 1H), 3.90 (s, 3H), 3.48 (dd, *J* = 15.1, 6.1 Hz, 1H), 2.53 (m, 1H), 2.31 (qd, *J* = 7.6, 1.3 Hz, 2H), 1.27 – 1.09 (m, 3H) ppm.

(*R*)-*N*-*Propyl-1,2,3,4-tetrahydroquinolin-3-amine (38).* To a solution of **36** (4.10 g, 16.5 mmol) in anhydrous tetrahydrofuran (50 mL) under inert atmosphere and cooled to 0°C, was added dropwise a solution of BH_3 ·THF (15 mL, 10 M soln.). Reaction was slowly warmed to room temperature over 1 hour with continued stirring, and subsequently heated to reflux overnight. Upon completion, reaction was quenched with the dropwise addition of MeOH at 0°C until no further gas evolution was detected. Reaction mixture was then concentrated under vacuum to afford the desired crude product used in the next step without further purification.

(R)-N-(4-(Propyl(1,2,3,4-tetrahydroquinolin-3-yl)amino)butyl)-1H-indole-2-carboxamide (17).

To a stirring solution of **38** (0.173 g, 0.9 mmol) and K₂CO₃ (0.3 g, 2.1 mmol) in *N*,*N*-dimethylformamide (5 mL) at room temperature, was added dropwise a solution of *N*-(4-bromobutyl)-1*H*-indole-2-carboxamide² (0.21 g, 0.7 mmol) in *N*,*N*-dimethylformamide (5 mL). Reaction was stirred at this temperature for the following 12 hours, and then heated gently to 100°C for an additional hour. Upon completion, the reaction mixture was filtered and concentrated under vacuum to afford the crude material which was purified by Combiflash chromatography, eluting with MeOH:Chloroform (15:85) to afford the desired product (**0.096 g**, **34% yield**). ¹H NMR (400 MHz, CDCl₃) δ 9.20 (br s, 1H), 7.63 (dd, *J* = 8.0, 1.1 Hz, 1H), 7.42 (dd, *J* = 8.3, 1.0 Hz, 1H), 7.32 – 7.23 (m, 1H), 7.13 (ddd, *J* = 8.0, 7.0, 1.0 Hz, 1H), 7.06 – 6.91 (m, 2H), 6.80 (d, *J* = 2.3 Hz, 1H), 6.72 – 6.56 (m, 1H), 6.50 (ddd, *J* = 21.2, 8.3, 1.2 Hz, 1H), 6.32 (br s, 1H), 3.80 (m, 1H), 3.49 (q, *J* = 6.6 Hz, 2H), 3.41 – 3.31 (m, 1H), 3.24 – 3.00 (m, 1H), 2.87 – 2.68 (m, 2H), 2.67 – 2.41 (m, 4H), 2.15 (q, *J* = 7.6 Hz, 2H), 1.73 – 1.39 (m, 4H), 0.86 (t, *J* = 7.3 Hz, 3H) ppm. ¹³C NMR (101 MHz, CDCl₃) δ 161.48, 144.30, 136.08, 130.85, 130.54, 129.92, 127.66, 127.15, 126.82, 124.40, 121.86, 120.87, 120.61, 118.04, 117.26, 113.84, 111.85,

101.60, 54.11, 52.61, 50.18, 45.38, 44.14, 41.68, 39.58, 32.63, 29.76, 27.57, 26.22, 21.87, 11.80 ppm. The free base was converted into the corresponding oxalate salt. HRMS (ESI positive) calculated 405.26489; found 405.26477 (+H⁺). $[\alpha]_D^{23}$:-9.565 (0.115 g/100 mL in MeOH). Anal. (C₂₅H₃₂N₄O⁻¹.25C₂H₂O₄^{-0.5H₂O) C, H, N M.p.: 88-92 °C}

(*R*)-3-(*Dipropylamino*)-1-methoxy-3,4-dihydroquinolin-2(1H)-one (35). To a well stirred solution of **34** (3.6 g, 18.7 mmol) in tetrahydrofuran (25 mL) at room temperature, was added propionaldehyde (3.26 g, 4.03 mL, 56.2 mmol) along with acetic acid (0.28 g, 0.27 mL, 4.6 mmol), and finally NaBH(OAc)₃ (13.1 g, 84.6 mmol). An additional 25 mL of tetrahydrofuran was subsequently added to rinse, and the reaction stirred at room temperature overnight. Reaction was quenched with the dropwise addition of a saturated Na₂CO₃ solution, and subsequently extracted 3x with 50 mL portions of dichloromethane. Organic fractions were then combined together, dried over anhydrous Na₂SO₄, before filtering and concentrating under reduced pressure. Crude material was used in the next step without further purification.

(*R*)-*N*,*N*-*Dipropyl-1,2,3,4-tetrahydroquinolin-3-amine (37).* To a solution of **35** (1.3 g, 4.7 mmol) in anhydrous tetrahydrofuran (50 mL) under inert atmosphere and cooled to 0°C, was added dropwise a solution of BH₃·THF (5 mL, 10 M soln.). Reaction was slowly warmed to room temperature over 1 hour with continued stirring, and subsequently heated to reflux overnight. Upon completion, reaction was quenched with the dropwise addition of MeOH until no further gas evolution was detected. Reaction mixture was then concentrated under vacuum to afford the desired crude product as a viscous yellow oil in sufficient purity as to advance to the following synthetic (1.00 g, 86% yield). ¹H NMR (400 MHz, CDCl₃) δ 6.96 (t, *J* = 7.8 Hz, 2H),

6.62 (td, *J* = 7.4, 1.2 Hz, 1H), 6.49 (d, *J* = 7.9 Hz, 1H), 3.14 (d, *J* = 9.4 Hz, 2H), 2.83 (d, *J* = 7.2 Hz, 2H), 2.57 – 2.48 (m, 4H), 1.68 (d, *J* = 10.7 Hz, 1H), 1.68 (s, 1H), 1.48 (p, *J* = 7.3 Hz, 2H), 1.00 – 0.89 (m, 2H), 0.88 (t, *J* = 7.3 Hz, 6H) ppm.

(R)-N-(4-(3-(Dipropylamino)-3,4-dihydroquinolin-1(2H)-yl)butyl)-1H-indole-2-carboxamide

(14). Compound 37 (0.1 g, 0.4 mmol), along with N-(4-bromobutyl)-1H-indole-2-carboxamide² (0.15 g, 0.5 mmol), and diisopropylethylamine (0.742 g, 1 mL, 5.74 mmol) were charged to a sealed vessel and solubilized in 1,2-dimethoxyethane (20 mL). Reaction was then heated to approximately 80°C, and allowed to proceed with vigorous stirring overnight. Upon completion of the reaction, solvent was concentrated under vacuum and the remaining residue purified by Combiflash chromatography, eluting with MeOH:chloroform (5:95). Semi pure material isolated was further purified by two iterations of preparatory thin-layer chromatography, eluting with MeOH:chloroform: NH₄OH (5:95:1) to afford the desired product (0.0134g, 7% yield).). ¹H NMR (400 MHz, CDCl₃ + CD₃OD) δ 9.71 (br s, 1H), 7.61 (d, J = 8.0, 1H), 7.41 (d, J = 8.0 Hz, 1H), 7.26 – 7.24 (m, 1H), 7.12 – 6.95 (m, 3H), 6.83 (br s, 1H), 6.60 – 6.56 (m, 2H), 3.64 – 3.61 (m, 1H), 3.49 (m, 2H), 3.31 - 3.22 (m, 5H), 2.81 (br d, J = 7.9 Hz, 2H), 2.55 - 2.35 (m, 3H), 1.82 - 1.33 (m, 8H), 0.88 (td, J = 7.3, 4.0 Hz, 6H) ppm. ¹³C NMR (101 MHz, CDCl₃ + CD₃OD) δ 161.81, 144.83, 136.18, 129.70, 127.52, 127.35, 124.29, 121.84, 120.43, 116.36, 111.86, 110.78, 62.52, 52.66, 49.87, 49.66, 49.44, 49.23, 39.28, 34.74, 21.19, 23.72, 18.85, 13.80, 11.68 ppm. HPLC-DAD Method A (Rt: 8.214 min; purity 94.2%). HRMS (ESI positive) calculated 447.31184; found 447.31402 (+H⁺). $[\alpha]_D^{23}$:9.091 (0.055 g/100 mL in MeOH).

Scheme 3.



a) DMP, DCM, from 0 °C to RT; b) Na(OAc)₃BH, AcOH, DCE; c) PPh₃, CBr₄, ACN; d) K₂CO₃, ACN, 60 °C.

N-(4-Oxobutyl)benzo[d]thiazole-2-carboxamide (39). To a stirring solution of **23** (0.5 g, 1.9 mmol) in dichloromethane (20 mL) at room temperature was added Dess-Martin Periodinane (0.974 g, 2.3 mmol) and allowed to proceed over the following 1 hour. Upon completion, reaction mixture was diluted in dichloromethane and washed with a 10% aqueous solution of NaHCO₃. Organic fraction was dried over anhydrous Na₂SO₄, filtered, and concentrated under vacuum to afford the crude aldehyde as a viscous oil. Crude product was subsequently purified via Combiflash chromatography, eluting with EtOAc:Hexanes (1:1) to afford the desired product as a cloudy, viscous oil (0.495 g, 99%). ¹H NMR (400 MHz, CDCl₃) δ 9.80 (s, 1H), 8.13 – 8.03 (m, 1H), 7.98 (ddd, *J* = 8.0, 4.3, 1.1 Hz, 2H), 7.61 – 7.46 (m, 1H), 3.62 – 3.51 (m, 2H), 2.62 (td, *J* = 7.2, 1.1 Hz, 2H), 2.06 – 1.97 (m, 2H) ppm.

(R)-N-(4-(Methyl(1,2,3,4-tetrahydroquinolin-3-yl)amino)butyl)benzo[d]thiazole-2-

carboxamide (15) and (R)-N-(4-(3-((4-(Benzo[d]thiazole-2carboxamido)butyl)(methyl)amino)-3,4-dihydroquinolin-1(2H)-yl)butyl)benzo[d]thiazole-2-

carboxamide (18). To a well stirring solution of (*R*)-*N*-methyl-1,2,3,4-tetrahydroquinolin-3amine (0.353 g, 2.17 mmol) in 1,2-dichloroethane (45 mL) was added acetic acid (5 drops, catalytic*), followed by the dropwise addition of a solution of **39** (0.540 g, 2.17 mmol) in 1,2dichloroethane (5 mL). This was allowed to stir at room temperature for the following 15 minutes before the addition of NaBH(OAc)₃ (0.691 g, 3.26 mmol). The reaction was subsequently allowed to proceed over the following 85 hours. Upon completion, reaction was transferred to a separatory funnel and washed 3x with a 10% aqueous solution of NaHCO₃. Organic fraction was dried over anhydrous Na₂SO₄, filtered, and concentrated under vacuum to afford the crude products as a pale orange oil. Purification via Combiflash chromatography was conducted in gradient from 0 to 15% MeOH in DCM, and two distinct fractions were collected and concentrated under vacuum to afford two regioisomeric products as amber colored oils.

18 (77.6 mg, 5.69 % yield) eluted first. ¹H NMR (400 MHz, CDCl₃) δ 8.07 – 7.89 (m, 3H), 7.78 (dd, J = 7.9, 1.1 Hz, 1H), 7.66 – 7.58 (m, 1H), 7.58 – 7.41 (m, 2H), 7.23 (td, J = 7.5, 1.1 Hz, 1H), 7.16 – 7.10 (m, 1H), 7.10 – 7.01 (m, 1H), 7.01 – 6.85 (m, 2H), 6.57 (dt, J = 20.3, 7.7 Hz, 2H), 3.54 (q, J = 6.5 Hz, 5H), 3.51 – 3.40 (m, 2H), 3.40 – 3.25 (m, 2H), 3.06 – 2.94 (m, 2H), 2.94 – 2.81 (m, 2H), 2.58 (s, 3H), 1.75 (d, J = 6.4 Hz, 8H) ppm. free base was converted into the corresponding oxalate salt. ¹³C NMR (101 MHz, dmso) δ 165.11, 162.73, 159.96, 159.83, 153.13, 144.64, 136.52, 129.97, 127.54, 127.31, 124.31, 123.46, 118.81, 117.03, 111.79, 109.99, 57.53, 53.20, 51.78, 50.61, 49.04, 48.23, 37.17, 26.86, 26.48, 26.46, 22.96, 21.40 ppm. [α]²³_D:-

9.231 (0.130g/100 mL in MeOH). HRMS (ESI positive) calculated 627.26; found 627.26 (+H⁺). Anal. (C₃₄H₃₈N₆O₂S₂ · 3C₂H₂O₄) C, H, N

15 (45.3 mg, 5.28 % yield) eluted second. ¹H NMR (400 MHz, CDCl₃) δ 8.02 – 7.92 (m, 1H), 7.85 (d, J = 7.9 Hz, 1H), 7.73 – 7.66 (m, 1H), 7.56 – 7.43 (m, 1H), 7.29 (m, 2H), 6.99 (t, J = 8.0 Hz, 2H), 6.52 (d, J = 8.3 Hz, 1H), 3.90 (br s, 1H), 3.76 – 3.64 (m, 3H), 3.62 – 3.46 (m, 3H), 3.17 – 3.06 (m, 3H), 2.74 (s, 3H), 1.99 – 1.91 (m, 2H), 1.77 (dt, J = 21.6, 10.7 Hz, 2H) ppm. Free base was converted into the corresponding oxalate salt. ¹³C NMR (101 MHz, dmso) δ 165.11, 163.12, 159.95, 153.14, 144.72, 136.52, 129.84, 127.54, 127.30, 124.33, 123.46, 117.17, 117.02, 114.27, 65.35, 57.75, 53.15, 41.01, 39.02, 37.03, 27.55, 26.50, 21.32, 15.61 ppm. [α]_D²³:-11.0 (0.100 g/100 mL in MeOH). HRMS (ESI positive) calculated 395.19; found 395.19 (+H⁺). Anal. (C₂₂H₂₆N₄OS · 2.5C₂H₂O₄ · 1.5H₂O) C, H, N. M.p.: 83 °C (dec.)

N-(4-bromobutyl)-2-methylindoline-1-carboxamide (40). Compound was prepared following the synthetic procedure described for **24** starting from **26** (0.850 g, 3.42 mmol). Crude product was partially purified via Combiflash chromatography, eluting with EtOAc:Hexanes (gradient from 20% to 60% EtOAc in hexanes), and used in the next step without any further purification.

2-Methyl-N-(4-(methyl((R)-1,2,3,4-tetrahydroquinolin-3-yl)amino)butyl)indoline-1-

carboxamide (16). **40** (0.257 g. 0.83 mmol) along with (*R*)-*N*-methyl-1,2,3,4-tetrahydroquinolin-3-amine (0.154 g, 0.95 mmol) were added to a sealed vessel and suspended in acetonitrile (5 mL). Solubilization was induced with vigorous stirring and gentle heating (65 – 85 °C) in an oil bath. Reaction was allowed to proceed over the following 3 hours, before heating was removed. Reaction mixture was basified by the addition of solid K_2CO_3 (5 to 10 equivalents), filtered and concentrated under vacuum to afford the crude product as an amber colored oil. Crude material was purified via Combiflash chromatography in gradient from 5 to 10% MeOH in Chloroform to afford the desired product as an off-white solid (**25.8 mg, 7.95% yield**). ¹H NMR (400 MHz, CDCl₃) δ 7.75 (d, *J* = 8.1 Hz, 1H), 7.15 (d, *J* = 7.6 Hz, 2H), 7.05 – 6.87 (m, 3H), 6.68 (t, *J* = 7.4 Hz, 1H), 6.52 (d, *J* = 8.0 Hz, 1H), 5.31 (s, 1H), 4.56 – 4.47 (m, 1H), 3.68-3.24 (m, 5H), 3.03 (m, 4H), 2.62 (s and m, 3H and 1H), 1.98-1.56 (m, 4H), 1.30 (d, *J* = 6.3 Hz, 3H) ppm. ¹³C NMR (100 MHz, CDCl₃) δ 154.85, 143.52, 142.49, 129.83, 129.66, 127.54, 127.46, 125.10, 121.94, 118.41, 115.01, 114.50, 57.20, 55.02, 53.70, 38.99, 37.39, 36.42, 27.38, 21.04, 15.26 ppm. [α]²³_D: -10.909 (0.055 g/100 mL in MeOH). HRMS (ESI positive) calculated 393.2646; found 393.2657 (+H⁺). Anal. (C₂₄H₃₂N₄O · 0.5CHCl₃ · 1.5CH₃OH) C, H, N. M.p.: 47-48 °C

Scheme 4.



a) tBuOLi, tBuOH, CH₃NO₂, THF; b) MsCl, TEA, DCM; c) LAH, THF; d) Indole-2-carboxylic acid, HOBt, EDC, THF; e) DMP, DCM, from 0 °C to RT; f) **sumanirole**, Na(OAc)₃BH, AcOH, DCE.

Trans-ethyl 2-(1-hydroxy-2-nitroethyl)cyclopropane-1-carboxylate (41). To a solution of ethyl 2-formylcyclopropane-1-carboxylate (predominantly trans) (1.0 g, 7 mmol) in tetrahydrofuran/t-BuOH (10 mL, 1:1 ratio), cooled to 0 °C in an ice bath, was added dropwise nitromethane (0.641 g, 10.5 mmol). After 5 minutes of stirring, lithium tert-butoxide (0.112 g) was added cautiously to the reaction and allowed to stir at room temperature for the following 3 hours. Upon completion, the reaction was diluted with a saturated aqueous solution of NH₄Cl, and the mixture extracted with dichloromethane. Organic fractions were combined together and dried over anhydrous Na₂SO₄, before filtering and drying under vacuum to afford the desired product without further purification (1.32 g, 92.3% yield). ¹H NMR (400 MHz, CDCl₃) δ 4.59 – 4.36 (m, 2H), 4.23 – 4.04 (m, 2H), 3.99 (q, *J* = 7.0 Hz, 1H), 2.90 – 2.84 (br s, 1H), 1.77 – 1.67 (m, 1H), 1.55 (m, 1H), 1.33 – 1.16 (m, 4H), 1.16 – 0.99 (m, 1H) ppm.

Trans-ethyl (E)-2-(2-nitrovinyl)cyclopropane-1-carboxylate (42).

To a well stirring solution of 41 (8.291 g, 40.8 mmol) and methanesulfonyl chloride (7.01 g, 4.74 mL, 61.2 mmol) in dichloromethane (100 mL) under inert atmosphere and submerged in a 0 °C ice bath, was added dropwise a solution of triethylamine (10.322 g, 14.227 mL, 102 mmol) in dichloromethane (50 mL). Reaction was allowed to warm to room temperature with stirring over the following 18 hours. Upon completion, reaction mixture was concentrated under vacuum and subsequently diluted in H₂O. This aqueous mixture was extracted with diethyl ether, and the organic fraction combined together and dried over anhydrous Na₂SO₄, before filtering, and concentrating under vacuum to afford the crude product as a yellow oil. Crude material was purified twice via Combiflash chromatography, first eluting in gradient from 0 to 90% EtOAc in Hexanes, and semi-pure material re-purified, eluting in gradient from 0 to 24% EtOAc in Hexanes. Finally, the purified racemic mixture was quantitated by chiral HPLC. HPLC-DAD Method C (Enantiomer A Rt: 30.379 min; Enantiomer B Rt: 37.560 min; purity 99.9%; er 50:50). ¹H NMR (400 MHz, CDCl₃) δ 7.14 (d, J = 13.2 Hz, 1H), 6.74 (dd, J = 13.2, 10.6 Hz, 1H), 4.16 (qd, J = 7.1, 1.6 Hz, 2H), 2.23 - 2.10 (m, 1H), 1.97 (m, 1H), 1.66 (dtd, J = 8.6, 5.4, 3.2 Hz, 1H),1.38 – 1.14 (m, 4H) ppm.

Trans-(2-(2-aminoethyl)cyclopropyl)methanol (43). Lithium aluminum hydride (0.4 g, 10.37 mmol) was charged to a round bottom flask under inert atmosphere and cooled to 0 °C in an ice bath before being cautiously suspended with the dropwise addition of anhydrous tetrahydrofuran (30 mL). To this stirring suspension was added dropwise a solution of **42** (0.640 g, 3.46 mmol) in anhydrous tetrahydrofuran (10 mL), and the reaction allowed to warm to room temperature with vigorous stirring over following 18 hours. Upon completion of the reaction, excess hydride

was quenched via the dropwise addition of MeOH, followed by 2N aqueous solution of NaOH. Anhydrous Na_2SO_4 was added into stirring solution to remove excess water, and solids were removed via filtration over Celite. Filtrate was concentrated under vacuum to afford the crude desired amino alcohol used in the next step without further purification (**0.386 g, 97% yield**).

Trans-N-(2-(2-(hydroxymethyl)cyclopropyl)ethyl)-1H-indole-2-carboxamide (44). As previously described,³ to a stirring solution of 1H-indole-2-carboxylic acid (0.391 g, 2.4 mmol) in a 1:1 mixture of THF:DCM (20 mL) cooled to 0 °C in an ice bath, was added 1-Ethyl-3-(3dimethylaminopropyl)carbodiimide (0.548 g, 2.8 mmol) and Benzotriazol-1-ol (0.356 g, 2.6 mmol). After 30 minutes of stirring at 0 °C, a solution of 43 (0.250 g, 2.2 mmol) in dichloromethane (10 mL) was added dropwise, followed by the dropwise addition of N,Ndiisopropylethylamine (0.397 g, 0.535 mL, 3 mmol) into the reaction mixture and allowed to stir overnight warming to room temperature. Upon reaction completion, solvent was stripped under vacuum and resultant residue diluted in EtOAc. Organic phase was washed with a 20% aqueous solution of NaHCO₃, before drying over anhydrous Na₂SO₄, filtering, and concentrating under reduced pressure. Crude material was purified via Combiflash chromatography, eluting with EtOAc:Hexanes (4:6) to afford the desired product (0.275 g, 49% yield). ¹H NMR (400 MHz, CDCl₃) δ 9.56 (m, 1H), 7.63 (dd, J = 7.9, 1.0 Hz, 1H), 7.41 (dt, J = 8.3, 1.0 Hz, 1H), 7.36 (m, 1H), 7.12 (ddd, J = 8.0, 7.0, 1.0 Hz, 1H), 6.97 (m, 2H), 4.06 – 3.75 (m, 2H), 3.49 – 3.22 (m, 1H), 3.01 (dd, J = 11.0, 9.5 Hz, 1H), 2.01 - 1.88 (m, 1H), 1.17 - 1.03 (m, 1H), 0.96 (dq, J = 9.0, 4.6)Hz, 1H), 0.70 (tt, J = 9.4, 4.3 Hz, 1H), 0.40 (ddt, J = 16.7, 8.4, 5.0 Hz, 2H) ppm.

Trans-N-(2-(2-formylcyclopropyl)ethyl)-1H-indole-2-carboxamide (45). As previously described,³ to a stirring solution of 44 (0.280 g, 1.1 mmol) in dichloromethane (15 mL) at 0 °C was added Dess-Martin Periodinane (0.583 g, 1.38 mmol) and allowed to proceed over the following 1 hour at room temperature. Upon completion, reaction mixture was diluted in dichloromethane and washed with a 10% aqueous solution of NaHCO₃. Organic fraction was dried over anhydrous Na₂SO₄, filtered, and concentrated under vacuum to afford the crude aldehyde. Crude product was subsequently purified via Combiflash chromatography, eluting with EtOAc:Hexanes (1:1) to afford the desired product as a viscous oil (0.119 g, 43% yield). ¹H NMR (400 MHz, CDCl₃) δ 9.49 (s, 1H), 9.10 (d, *J* = 5.2 Hz, 1H), 7.65 (dt, *J* = 8.2, 1.0 Hz, 1H), 7.45 (dt, *J* = 8.3, 1.0 Hz, 1H), 7.34 – 7.23 (m, 1H), 7.14 (ddd, *J* = 8.0, 7.0, 1.0 Hz, 1H), 6.84 (dd, *J* = 2.2, 0.9 Hz, 1H), 6.37 (s, 1H), 3.60 (q, *J* = 6.7 Hz, 2H), 1.80 – 1.67 (m, 3H), 1.46 (m, 1H), 1.36 (dt, *J* = 9.0, 4.6 Hz, 1H), 1.00 (ddd, *J* = 8.1, 6.4, 4.7 Hz, 1H) ppm.

Trans-N-(2-(2-((methyl((R)-2-oxo-1,2,5,6-tetrahydro-4H-imidazo[4,5,1-ij]quinolin-5-

yl)amino)methyl)cyclopropyl)ethyl)-1H-indole-2-carboxamide (19). To a stirring solution of **sumanirole** (0.083 g, 0.41 mmol) in 1,2-dichloroethane (10 mL), was added Acetic Acid (5 drops, catalytic), followed by the dropwise addition of **45** (0.120 g, 0.47 mmol) in 1,2-dichloroethane (5 mL). After stirring for 30 minutes, NaBH(OAc)₃ (0.130 g, 0.61 mmol) was added slowly into the reaction and allowed to stir overnight at room temperature. Upon completion, reaction was quenched with the addition of 0.1 g of K₂CO₃, filtered and concentrated under vacuum to afford the crude product which was purified by Combiflash chromatography, eluting with MeOH:chloroform (1:9) to afford the desired product **(89 mg, 49% yield).** ¹H NMR (400 MHz, CDCl₃) δ 9.66 (d, *J* = 18.5 Hz, 1H), 8.92 (s, 1H), 7.61 (d, *J* =

8.2 Hz, 1H), 7.44 (d, J = 8.3 Hz, 1H), 7.31 – 7.21 (m, 1H), 7.11 (tdd, J = 7.0, 3.8, 1.0 Hz, 1H), 7.00 – 6.90 (m, 1H), 6.90 – 6.77 (m, 3H), 6.59 – 6.51 (m, 1H), 4.18 (dd, J = 12.0, 4.2 Hz, 1H), 3.63 – 3.47 (m, 3H), 3.25 (m, 1H), 2.92 (t, J = 7.5 Hz, 2H), 2.60 – 2.39 (s and m, 3H and 2H), 1.65 – 1.52 (m, 2H), 0.76 (m, 1H), 0.61 (m, J = 7.2 Hz, 1H), 0.46 – 0.35 (m, 2H) ppm. ¹³C NMR (101 MHz, CDCl₃) δ 161.70, 154.52, 154.53, 136.32, 136.30, 136.31, 130.85, 130.83, 127.58, 127.30, 127.29, 125.73, 124.37, 121.81, 121.51, 120.57, 119.91, 119.92, 118.42, 118.35, 111.99, 107.22, 106.56, 102.00, 101.97, 58.62, 58.41, 57.33, 57.22, 39.90, 39.73, 38.66, 38.61, 33.59, 33.54, 27.31, 27.16, 17.58, 17.11, 17.05, 15.92, 10.83, 10.73, ppm. HPLC-DAD Method D (diastereoisomer A R_i: 114.571 min; diastereoisomer B R_i: 138.375 min; purity 97.7%; dr 50:50). The free base was converted into the corresponding oxalate salt. $[\alpha]_D^{23}$: 4.545 (0.110 g/100 mL in MeOH). HRMS (ESI positive) calculated 444.2394; found 444.2384 (+H⁺). Anal. (C₂₆H₂₉N₅O₂ · 2C₂H₂O₄) C, H, N. M.p.: 113 °C (dec.)





a) Indole-2-carboxylic acid, HOBt, EDC, THF; b) 10% HCl in H₂O, THF; c) DMP, DCM, from 0 °C to RT; d) **sumanirole**, Na(OAc)₃BH, AcOH, DCE.

Trans-N-((2-(2-((tetrahydro-2H-pyran-2-yl)oxy)ethyl)cyclopropyl)methyl)-1H-indole-2-

carboxamide (47). As previously described,³ to a stirring solution of 1H-indole-2-carboxylic acid (1 g, 6.2 mmol) in tetrahydrofuran (20 mL) cooled to 0 °C in an ice bath, was added 1-Ethyl-3-(3-dimethylaminopropyl)carbodiimide (1.39 g, 7.24 mmol) and Benzotriazol-1-ol (0.977 g, 7.24 mmol). After 30 minutes of stirring at 0 °C, a solution of 46 (1.03 g, 5.17 mmol) in tetrahydrofuran (20 mL) was added dropwise, followed by the dropwise addition of *N*,*N*-diisopropylethylamine (0.397 g, 0.535 mL, 3 mmol) into the reaction mixture and allowed to stir overnight warming to room temperature. Upon reaction completion, solvent was stripped under vacuum and resultant residue diluted in EtOAc. Organic phase was washed with a 20% aqueous solution of NaHCO₃, before drying over anhydrous Na₂SO₄, filtering, and concentrating under reduced pressure. Crude material was purified via Combiflash chromatography, eluting with EtOAc:Hexanes (3:7) to afford the desired product as a mixture of diasteroisomers (0.285 g, 17% yield). ¹H NMR (400 MHz, CDCl₃) δ 9.51 (s, 1H), 7.64 (d, *J* = 8.1 Hz, 1H), 7.44 (dq, *J* =

8.4, 1.0 Hz, 1H), 7.32 - 7.23 (m, 1H), 7.13 (ddd, J = 8.0, 7.0, 1.0 Hz, 1H), 6.94 (ddd, J = 13.8, 2.2, 0.9 Hz, 1H), 6.58 - 6.51 (m, 1H), 4.61 (ddd, J = 22.1, 5.1, 2.6 Hz, 1H), 4.02 - 3.78 (m, 2H), 3.69 - 3.46 (m, 3H), 3.14 (dddd, J = 16.7, 13.3, 8.2, 4.7 Hz, 1H), 2.07 - 1.31 (m, 8H), 0.80 (m, 2H), 0.57 - 0.38 (m, 2H) ppm.

Trans-N-((2-(2-hydroxyethyl)cyclopropyl)methyl)-1H-indole-2-carboxamide (48). As previously described,³ to a stirring solution of **47** (0.3 g, 0.88 mmol) in tetrahydrofuran (6 mL) at room temperature, was added dropwise a 10% aqueous solution of HCl and allowed to stir overnight. Upon completion, reaction was diluted with water and extracted with EtOAc. Organic fractions were combined together, dried over anhydrous Na₂SO₄, filtered, and concentrated under vacuum. Crude material was purified by Combiflash chromatography, eluting in gradient from 60 to 80% EtOAc in Hexanes to afford the desired product **(0.199 g, 88% yield).** ¹H NMR (400 MHz, CDCl₃ + CD₃OD) δ 7.60 (dt, *J* = 8.1, 1.0 Hz, 1H), 7.40 (dd, *J* = 8.4, 1.0 Hz, 1H), 7.23 (ddd, *J* = 8.3, 7.0, 1.2 Hz, 1H), 7.14 (m, 1H), 6.93 (br s, 1H), 3.77 (m, 3H), 2.95 – 2.83 (m, 1H), 1.79 (dq, *J* = 14.3, 5.3 Hz, 1H), 1.53 – 1.45 (br s, 1H), 1.27 – 1.07 (m, 1H), 0.77 (dtt, *J* = 30.4, 9.0, 4.5 Hz, 2H), 0.42 (ddt, *J* = 37.7, 8.3, 5.0 Hz, 2H) ppm.

Trans-N-((2-(2-oxoethyl)cyclopropyl)methyl)-1H-indole-2-carboxamide (49). As previously described,³ to a stirring solution of **48** (0.2 g, 0.77 mmol) in dichloromethane (15 mL) at room temperature and opened to atmosphere was added Dess-Martin Periodinane (0.408 g, 0.96 mmol) and allowed to proceed over the following 1 hour. Upon completion, reaction mixture was diluted in dichloromethane and washed with a 10% aqueous solution of NaHCO₃. Organic fraction was dried over anhydrous Na₂SO₄, filtered, and concentrated under vacuum to afford the

crude aldehyde. Crude product was subsequently purified via Combiflash chromatography, eluting with EtOAc:Hexanes (1:1) to afford the desired product (0.179 g, 90% yield). ¹H NMR (400 MHz, CDCl₃) δ 9.92 (s, 1H), 9.35 (s, 1H), 7.71 (dt, J = 8.1, 1.0 Hz, 1H), 7.61 (br s, 1H), 7.44 (dq, J = 8.2, 1.0 Hz, 1H), 7.32 – 7.23 (m, 2H), 7.14 (ddd, J = 8.0, 6.9, 1.0 Hz, 1H), 4.16 – 4.05 (m, 1H), 3.10 (dd, J = 19.5, 3.8 Hz, 1H), 2.69 (ddd, J = 13.6, 9.8, 2.6 Hz, 1H), 2.14 – 2.01 (m, 1H), 0.90 – 0.75 (m, 2H), 0.68 (dt, J = 8.3, 5.3 Hz, 1H), 0.53 (dt, J = 8.3, 5.3 Hz, 1H) ppm.

Trans-N-((2-(2-(methyl((R)-2-oxo-1,2,5,6-tetrahydro-4H-imidazo[4,5,1-ij]quinolin-5-

yl)amino)ethyl)cyclopropyl)methyl)-1H-indole-2-carboxamide (20). To a stirring solution of sumanirole (0.123 g, 0.61 mmol) in 1,2-dichloroethane (10 mL), was added Acetic Acid (5 drops, catalytic), followed by the dropwise addition of 49 (0.180 g, 0.7 mmol) in 1,2dichloroethane (5 mL). After stirring for 30 minutes, NaBH(OAc)₃ (0.193 g, 0.91 mmol) was added slowly into the reaction and allowed to stir overnight at room temperature. Upon completion, reaction was quenched with the addition of solid K_2CO_3 , filtered and concentrated under vacuum to afford the crude product which was purified by Combiflash chromatography, eluting with MeOH:chloroform (1:9) to afford the desired product (96 mg, 37% yield). ¹H NMR $(400 \text{ MHz}, \text{CDCl}_3) \delta 9.62 \text{ (s, 1H)}, 8.90 \text{ (br s, 1H)}, 7.59 \text{ (t, } J = 8.9 \text{ Hz}, 1\text{H}), 7.47 - 7.38 \text{ (m, 1H)}, 7.47 - 7.38 \text{ (m, 1H)}, 7.47 - 7.38 \text{ (m, 1H)}, 7.47 - 7.38 \text{ (m, 2H)}, 7.47 - 7.48 \text{ (m, 2H)}, 7.47 + 7.48 \text{ (m, 2H)}, 7.47 + 7.48 \text{ (m, 2H)}, 7.47 + 7.48 \text{ (m, 2H)}, 7.48 \text{ (m$ 7.30 - 7.21 (m, 1H), 7.10 (dd, J = 8.1, 7.0 Hz, 1H), 6.95 (dq, J = 7.6, 4.0 Hz, 1H), 6.91 - 6.79(m, 3H), 6.56 (br s, 1H), 4.13 (ddd, J = 36.2, 12.1, 4.2 Hz, 1H), 3.70 - 3.48 (m, 2H), 3.34 - 3.15(m, 2H), 2.95 (dd, J = 11.4, 6.8 Hz, 2H), 2.75 – 2.57 (m, 1H), 2.42 – 2.32 (s and m, 3H and 1H), 1.55 (m, 1H), 1.25 (m, 1H), 0.89 (m, 1H), 0.73 (m, 1H), 0.54 - 0.42 (m, 1H), 0.38 (td, <math>J = 7.9, 4.0 Hz, 1H) ppm. HPLC-DAD Method D (diastereoisomer A Rt: 117.044 min; diastereoisomer B R_t : 129.707 min; purity 90.2%; dr 48:52). The free base was converted into the corresponding

oxalate salt. ¹³C NMR (101 MHz, d₆-DMSO) δ 163.60, 161.44, 153.75, 136.81, 132.33, 127.51, 127.22, 126.81, 123.58, 121.79, 121.35, 120.05, 119.35, 116.03, 115.65, 112.66, 107.30, 105.04, 102.83, 57.86, 57.76, 55.80, 54.58, 53.89, 42.79, 37.57, 32.46, 29.12, 26.25, 25.14, 18.79, 17.85, 17.66, 15.04, 10.54 ppm. [α]_D²³: -13.571 (0.140 g/100 mL in MeOH). HRMS (ESI positive) calculated 444.2394; found 444.2381 (+H⁺). Anal. (C₂₆H₂₉N₅O₂ · 2C₂H₂O₄ · 1.25H₂O) C, H, N. M.p.: 55 °C (dec.)

Scheme 6.



a) DMP, DCM, from 0 °C to RT; b) sumanirole, Na(OAc)₃BH, AcOH, DCE.

N-(4-oxobutyl)-1*H*-indole-2-carboxamide (51)

To a stirring solution of 50^2 (0.5 g, 2.15 mmol) in dichloromethane (25 mL) at 0 °C was added Dess-Martin Periodinane (1.05 g, 2.48 mmol) and allowed to proceed over the following 1 hour at room temperature. Upon completion, reaction mixture was diluted in dichloromethane and washed with a 10% aqueous solution of NaHCO₃. Organic fraction was dried over anhydrous Na₂SO₄, filtered, and concentrated under vacuum to afford the crude aldehyde. Crude product was partially purified via Combiflash chromatography, eluting with EtOAc:Hexanes (1:1), and used in the next step without any further purification.

(*R*)-*N*-(4-(methyl(2-oxo-1,2,5,6-tetrahydro-4*H*-imidazo[4,5,1-*ij*]quinolin-5-yl)amino)butyl)-1*H*-indole-2-carboxamide (1)

To a stirring solution of **sumanirole** (0.285 g, 1.40 mmol) in 1,2-dichloroethane (18 mL), was added Acetic Acid (5 drops, catalytic), followed by the dropwise addition of **51** (0.370 g, 1.6 mmol) in 1,2-dichloroethane (10 mL). After stirring for 30 minutes, NaBH(OAc)₃ (0.445 g, 2.1 mmol) was added slowly into the reaction and allowed to stir overnight at room temperature. Upon completion, reaction was quenched with the addition of solid K₂CO₃, filtered and concentrated under vacuum to afford the crude product which was purified by Combiflash

chromatography, eluting with MeOH:chloroform (15:85) to afford the desired product (0.359 g, 61.3% yield). ¹H and ¹³C NMR chemical shifts were consistent with previously reported data.¹ The free base was converted into the corresponding oxalate salt. HRMS (ESI positive) calculated 418.22; found 418.22 (+H⁺), error -1.5ppm. Anal. ($C_{24}H_{27}N_5O_2 \cdot C_2H_2O_4 \cdot H_2O$), calculated C 59.42, H 5.95, N 13.33; found C 59.68, 6.10, 13.14. A sample of the oxalate salt was free based and analyzed via HPLC-DAD: Chiralpak AD-H (Daicel Corporation CPI Company) column (4.6 x 250 mm, 5 µm). The mobile phase used (1 mL/min flow rate) was composed of 0.1% diethylamine in 2-propanol and hexanes with gradient elution, starting with 30% of 2-propanol linearly increasing to 40%. Maximum absorbance was observed at λ 280 nM. Purity and enatiomeric excess were determined to be >95% and >99%, respectively.

| Compounds | Calculated | | | Found | | | | |
|-----------|---|------------|----------------------|-------------------------------------|--------------------------|------------------------|--|--|
| | C% | H% | N% | C% | H% | N% | | |
| 2 | 68.32 | 4.89 | 9.37 | 68.58 | 4.81 | 9.42 | | |
| | GC-MS | (EI) 294.1 | <i>m/z</i> (M+ | -) | 1 | 1 | | |
| 8 | 51.81 | 5.74 | 12.08 | 51.62 | 5.35 | 11.96 | | |
| | 59.72 | 6.10 | 15.14 | 59.73 | 5.71 | 14.88 | | |
| 9 | HRMS = | 436 1802 | 2 (+H ⁺) | HRMS | = 436.18 | 14 (+H ⁺) | | |
| | 11002 (+11) | | | HRMS | = 458.16 | 34 (+Na ⁺) | | |
| | HRMS = | 405 228 | 5 (+H+) | HRMS | = 405.22 | 95 (+H ⁺) | | |
| 10 | | 100.2200 | (111) | HRMS = 427.2113 (+Na ⁺) | | | | |
| | HPLC-DAD = purity 93.3% | | | | | | | |
| 11 | 58.09 | 6.38 | 12.32 | 58.48 | 6.28 | 12.17 | | |
| | HRMS = $434.2551 (+H^+)$ HRMS = $434.2562 (+H^+)$ | | | | | | | |
| | HPLC-D | AD = fig | ures S1A | and S1B | | | | |
| 12 | 54.37 | 5.98 | 10.93 | 54.55 | 5.89 | 10.54 | | |
| | HPLC-DAD = figures S1A and S1B | | | | | | | |
| 13 | 57.33 | 6.19 | 11.94 | 57.66 | 6.40 | 11.83 | | |
| | HPLC-DAD | | | | AD = figures S1A and S1B | | | |
| 14 | HRMS = 447.31184 (+H ⁺) HRMS = 447.31402 (+H ⁺) | | | | | | | |
| | HPLC-DAD = purity 94.2% | | | | | | | |
| 15 | 50.15 | 5.30 | 8.66 | 50.15 | 5.17 | 8.47 | | |
| | HRMS = $395.19 (+H^+)$ | | | HRMS = $395.19 (+H^+)$ | | | | |

 Table S1: Microanalysis, Mass Spectroscopy and HPLC-DAD data

| 16 | 62.42 | 7.76 | 11.20 | 62.31 | 7.33 | 10.80 | |
|----|--------------------------|-----------|-----------------------|--------------------------|--------------------------|------------------------|--|
| | HRMS = $393.2649 (+H^+)$ | | | HRMS | HRMS = $393.2657 (+H^+)$ | | |
| 17 | 62.78 | 6.80 | 10.65 | 63.09 | 6.55 | 10.56 | |
| | HRMS = | 405.2648 | 39 (+H ⁺) | HRMS | = 405.26 | 447 (+H ⁺) | |
| 18 | 53.56 | 4.94 | 9.37 | 53.66 | 5.16 | 8.99 | |
| | HRMS = $627.26 (+H^+)$ | | | HRMS = $627.26 (+H^+)$ | | | |
| | 57.78 | 5.33 | 11.23 | 57.38 | 5.70 | 11.29 | |
| 19 | HRMS = $444.2394 (+H^+)$ | | | HRMS = $444.2384 (+H^+)$ | | | |
| | HPLC-DAD = figures S3 | | | | | | |
| | 55.77 | 5.54 | 10.84 | 55.46 | 5.54 | 11.18 | |
| 20 | HRMS = $444.2394 (+H^+)$ | | | HRMS = $444.2381 (+H^+)$ | | | |
| | HPLC-D | AD = figu | ures S4 | | | | |

| D2s-BRET Constructs | | Quinpirole | Sumanirole ^b | 1 ^b | 2 | 8 | 9 | 11 | 12 | 13 | 19 | 20 |
|------------------------|--------------|-----------------|-------------------------|-----------------|---------------|-----------------|--------------------|----------------|------------|---------------|-----------------|-----------------|
| | Emax | 100.0 + 6.0 | 122.0 + 7.5 | 93.4 ± | 16.2 ± | 86.7 ± | 108.7 ± | 110.9 ± | 65.5 ± | 67.8 ± | 49.1 ± | 93.4 ± |
| Gi1 | (%) | 100.0 ± 0.0 | 133.0 ± 7.5 | 5.5 | 6.1 | 4.3 | 8.0 | 6.8 | 6.6 | 6.0 | 2.9 | 5.5 |
| activation (10 min) | EC50 (nM) | 86.7 ± 29.7 | 543.3 ± 216.4 | 124.7 ± 60.5 | 0.5 ± 0.5 | 103.3 ± 29.8 | 1,166.8 ± 382.8 | 66.5 ± 23.7 | 812.8 ± | 222.3 ± 100.2 | 174.6 ± 58.0 | 124.7 ± 60.5 |
| | _ | | | | | | | | 555.7 | | | |
| GoA | Emax | 100.0 ± 5.3 | 94.8 ± 2.9 | 66.4 ± | 29.3 ± | 89.2 ± | 82.7 ± | 86.1 ± | 67.3 ± | 68.9 ± | 69.5 ± | 66.4 ± |
| activation | (%) | | | 4.1 | 5.8 | 6.4 | 7.4 | 6.1 | 6.7 | 6.1 | 4.0 | 4.1 |
| (10 min) | EC50 | 27.2 + 12.7 | 74 9 1 1 7 9 | 1.1 ± | 200.4 ± | 62+20 | 136.8 ± | 30.1 ± | 115.3 | 0.8 ± | 18.4 ± | 11.00 |
| (10 mm) | (nM) | 37.2 ± 12.7 | 74.0 ± 17.0 | 0.9 | 147.3 | 0.2 ± 2.9 | 61.8 | 13.1 | ± 56.3 | 0.4 | 6.8 | 1.1 ± 0.9 |
| cAMP | Emax | 100.0 ± | 114.1 + 2.2 | 95.9 ± | 24.2 ± | 95.3 ± | 80.1 ± | 72.9 ± | 96.8 ± | 88.7 ± | 44.2 ± | 95.9 ± |
| inhibition | (%) | 11.7 | 114.1 ± 2.2 | 3.3 | 14.1 | 9.4 | 13.2 | 14.1 | 11.7 | 11.3 | 3.2 | 3.3 |
| (10 min) | EC50 | 13.9 ± 8.5 | 32.2 ± 5.4 | 0.3 ± | 26.5 ± | 7.2 ± 4.1 | 96.1 ± | 67.6 ± | 78.5 ± | 76.7 ± | 17.1 ± | 0.3 ± 0.1 |

Table S2. D₂-Mediated Gi and Go activation, cAMP inhibition and β -arrestin2 recruitment^a

| (nM) | 0.1 | 26.3 | 65.1 | 50.6 | 44.8 | 45.2 | 7.6 | |
|---|--------------------|---------------|----------------------|-----------------------|-------------|-------------------------|---------------|--|
| | | | | | | | | |
| ^a Potency (expressed as EC50) and ef | fficacy values (%, | normalized t | o quinpirole Emax) | for hD ₂ R | expressed | in HEK29 | 93 cells. The | |
| values represent the arithmetic mean | ± SEM of at least | three indepen | dent experiments, ea | ach perform | ned in trip | licate. ^b Da | ta previously | |
| reported ² | | | | | | | | |

| Compounds | D ₄ R |
|----------------------------|--------------------------------------|
| | $\underline{K_i \pm \text{SEM}(nM)}$ |
| Sumanirole | 1610 ± 297 |
| 1 | 899 ± 154 |
| 11 | 30.9 ± 2.61 |
| (<i>R</i> , <i>S</i>)-12 | 25.5 ± 3.18 |
| (R,R)-13 | 44 ± 5.95 |
| 19 | 1490 ± 142 |
| 20 | 1320 ± 110 |

Table S3. $[^{3}H]$ -(R)-(+)-7-OH-DPAT radioligand competition binding data on D₄R

The values represent the arithmetic mean from at least three independent experiments, each performed in triplicate. The competition experiments were performed in the presence of 3 nM [³H]-(*R*)-(+)-7-OH-DPAT, similarly to what previously described,⁴ and membrane preparations from HEK293 cells expressing hD₄R (final concentration of proteins in the experiment ~30-60 µg/well). IC₅₀ values for each compound were determined from dose-response curves and K_i values were calculated using the Cheng-Prusoff equation;⁵ K_d values for [³H]-(*R*)-(+)-7-OH-DPAT were determined via separate homologous competitive binding experiments using cold (±)-7-OH-DPAT.

| Table | S4 . | Off-target | binding | for | 1 ^a |
|--------|-------------|------------|---------|-----|-----------------------|
| 1 4010 | ~ •• | Oll taiget | omanns | 101 | - |

| Binding Assay Results for 1 | | | | | |
|---|-----------------------|---------------------------|------------------|--|--|
| Receptor [³ H]Ligand | IC ₅₀ (nM) | K _i (nM) ± SEM | Hill Slope ± SEM | | |
| DA D ₁ [³ H]SCH23390 | >9200* | >7900* | * | | |
| 5HT _{1A} [³ H]8-OH-DPAT | 137 ± 29 | 127 ± 27 | -0.63 ± 0.07 | | |
| 5HT _{2A} [³ H]5-HT | 1,940 ± 410 | $1,440 \pm 260$ | -1.06 ± 0.18 | | |
| 5HT _{2C} [³ H]5-HT | 3,080 ± 390 | $2,440 \pm 270$ | -0.75 ± 0.04 | | |

^{*}If some experiments yielded Ki, values less than 10 μ M and other experiments yielded Ki, values greater than 10 μ M, the latter experiments were assigned a value of 10 μ M and averages calculated. The actual value is greater than that average and no standard error is reported. No Hill slope or SEM was calculated due to the low affinity of the compound ^aData were obtained through the NIDA Addiction Treatment Discovery Program contract (ADA151001) with Oregon Health & Science University.

| Binding Assay Results for 11 | | | | | |
|---|-----------------------|---------------------------|------------------|--|--|
| Receptor [³ H]Ligand | IC ₅₀ (nM) | K _i (nM) ± SEM | Hill Slope ± SEM | | |
| DA D ₁ [³ H]SCH23390 | $>10 \mu\mathrm{M}^*$ | $>10 \mu\mathrm{M}^*$ | * | | |
| 5HT _{1A} [³ H]8-OH-DPAT | 25.6 ± 3.0 | 23.7 ± 2.8 | -0.48 ± 0.01 | | |
| 5HT _{2A} [³ H]5-HT | $4,120 \pm 740$ | 3,090 ± 490 | -0.70 ± 0.10 | | |
| 5HT _{2C} [³ H]5-HT | >10 µM* | $>10 \mu\mathrm{M}^*$ | * | | |

| Fable S5. | Off-target | binding for | 11 ^a |
|-----------|------------|-------------|------------------------|
|-----------|------------|-------------|------------------------|

*No Hill slope or SEM was calculated due to the low affinity of the compound. ^aData were obtained through the NIDA Addiction Treatment Discovery Program contract (ADA151001) with Oregon Health & Science University.

| Binding Assay Results for (R,R)-13 | | | | | |
|---|-----------------------|---------------------------|------------------|--|--|
| Receptor [³ H]Ligand | IC ₅₀ (nM) | K _i (nM) ± SEM | Hill Slope ± SEM | | |
| DA D ₁ [³ H]SCH23390 | $>10 \mu\mathrm{M}^*$ | $>10\mu\mathrm{M}^*$ | * | | |
| 5HT _{1A} [³ H]8-OH-DPAT | 35.8 ± 4.4 | 33.2 ± 4.0 | -0.51 ± 0.01 | | |
| 5HT _{2A} [³ H]5-HT | $3,700 \pm 1,000$ | $2,710 \pm 670$ | -0.71 ± 0.04 | | |
| 5HT2C [³ H]5-HT | $>10 \mu\mathrm{M}^*$ | $>10 \mu\mathrm{M}^*$ | * | | |

Table S6. Off-target binding and functional data for (R,R)-13^a

*No Hill slope or SEM was calculated due to the low affinity of the compound. ^aData were obtained through the NIDA Addiction Treatment Discovery Program contract (ADA151001) with Oregon Health & Science University.



Figure S1A. 11, (*R*,*S*)-**12** and (*R*,*R*)-**13** – Chiral HPLC resolution OD (5 cm)



Figure S1B. 11, (*R*,*S*)-**12** and (*R*,*R*)-**13** – Chiral HPLC resolution AD-H (25 cm). In the top-left corner of the first panel are reported the blank chromatograms of 2-PrOH and CHCl₃ used to dissolve the samples.



Figure S2. 42 – Chiral HPLC resolution OZ-H (25 cm)



Figure S3. trans-19 - Chiral HPLC resolution AD-H (25 cm)



Figure S4. trans-20 - Chiral HPLC resolution AD-H (25 cm)



Figure S5. Sumanirole - Chiralpak AD-H (Daicel Corporation CPI Company) column (4.6 x 250 mm, 5 μ m). The mobile phase used (1 mL/min flow rate) was composed of 0.1% diethylamine in 2-propanol and hexanes with isocratic elution (10% of 2-propanol). Maximum absorbance was observed at λ 280 nM and enantiomeric excess was >99%.



Figure S6. 1 - Chiralpak AD-H (Daicel Corporation CPI Company) column (4.6 x 250 mm, 5 μ m). The mobile phase used (1 mL/min flow rate) was composed of 0.1% diethylamine in 2-propanol and hexanes with gradient elution, starting with 30% of 2-propanol linearly increasing to 40%. Maximum absorbance was observed at λ 280 nM and enantiomeric excess was >99%.



Figure S7. Dose response curves (normalized to quinpirole E_{max}) used for table 2. A. cAMP inhibition, B. β -arrestin 2 recruitment, C. Gi1 activation, and D. GoA activation (Black – quinpirole, gray – sumanirole, magenta – 2, blue – 9, red – 11, purple – 19, brown – 20, light blue – 12, orange – 13, yellow – 8, green – 1). Curves are fitted to a monophasic sigmoidal dose response. The error bars represent S.E.M.



Figure S8. Bias factor values, taken from table 3, represented as web of bias, similarly to what previously shown by Klein Herenbrink C. et al.⁶ Bias factors in logarithmic scale for D_2 -Mediated Gi vs. Go activation, cAMP inhibition vs. β -arrestin2 recruitment, and Go activation vs. β -arrestin2 recruitment.

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