

b

Protein	Number of peptides (Unique)	Number of peptides (Total)	% of aa sequence coverage
Kv4.2	27	126	30.00%
Kv4.1	7	11	14.92%
Kv4.3	15	17	25.19%
DPP6	30	40	33.99%
DPP10	26	26	29.27%
KChIP1	9	9	22.47%
KChIP2	5	5	16.30%
KChIP3	6	7	35.94%
KChIP4	12	16	43.60%

e

Pin1 sequence:

MADEEKLPPG WEKRMSRSSG RVYYFNHITN ASQWERPSGN SSSGGKNGQG EPARVRCSHL LVKHSQSRRP SSWRQEKITR <u>TKEEALELIN</u> <u>GYIQK</u>IKSGE EDFESLASQF SDCSSAKARG <u>DL</u> GAFSRGQM QKPFEDASFA LRTGEMSGPV FT <u>DSGIHIIL RTE</u>

f

Peptide	∆corr	Xcorr
TKEEALELINGYIQK	0.362	4.21
GDLGAFSR	0.185	2.20
GQMQKPFEDASFALR	0.482	3.63
TGEMSGPVFTDSGIHIILRTE	0.509	4.54
TGEMSGPVFTDSGIHIILR	0.595	5.07
	Peptide TKEEALELINGYIQK GDLGAFSR GQMQKPFEDASFALR TGEMSGPVFTDSGIHIILRTE TGEMSGPVFTDSGIHIILR	Peptide∆corrTKEEALELINGYIQK0.362GDLGAFSR0.185GQMQKPFEDASFALR0.482TGEMSGPVFTDSGIHIILRTE0.509TGEMSGPVFTDSGIHIILR0.595

	# of	# of
Gene	peptides	peptides
symbol	(Unique)	(Total)
EIF3A	53	56
	52	54
	<u>JZ</u>	40
CANDT	41	42
SPIAN1	39	39
EIF4G1	35	39
EIF3B	33	33
LISP9X	32	33
MYOG	20	20
	30	30
EIF4G2	29	31
PSMD1	29	29
MYH9	27	27
CYFIP1	25	27
	25	26
	25	20
KIF5B	25	25
EIF3C	24	25
SPTBN1	22	22
AP3B1	21	21
MVO1C	10	10
	10	10
ATP2B1	18	18
UBE3C	18	18
USP7	16	16
PSMD2	15	21
MVO1D	15	15
	CI CI	15
PPP6R3	13	13
ARHGEF2	13	13
CLASP2	11	19
USP24	11	11
	11	11
	10	11
AP2A1	10	11
USP48	10	10
PIK3R4	10	10
USP10	10	10
ARHGAP17	9	10
	0	0
	9	9
MYOID	9	9
AP1B1	8	8
KIF11	8	8
UBA52	7	52
KIE5A	7	7
PI4KA	/	/
CAND2	7	7
USP19	6	6
ARHGEF1	6	6
CYFIP2	6	6
	6	6
LAROZ	0	0
USP5	5	11
USP15	5	6
PPP6R1	5	6
LIBE20	5	5
	5	5
UDE4D	5	5
KIF23	5	5
MYO1E	5	5
CLASP1	5	5
FIF4G3	5	5
URB1	5	5
	3	3
	4	4
XPO6	4	4
RAB3GAP1	4	4
PTPN23	4	4
PPP4R1	3	3
AD242	2	2
	<u> </u>	<u> </u>
ARFGEF1	3	3
EIF2AK4	3	3

С

h		# .4	# .6
u		# OT	# OT
	Gene	peptides	peptides
	symbol	(Unique)	(Total)
	RPS18	25	39
	RPS9	17	37
	RPS19	16	30
	RPS16	16	30
	RPS7	15	20
	RPS25	13	16
	RPS13	12	14
	PSMB2	12	12
	RPS14	11	14
	PSMB1	11	14
	PSMB4	11	13
		11	13
		11	12
		11	10
		10	10
			10
		9	13
	KAB/A	9	12
	RPL10	9	
	RAB1A	9	9
	RPS11	9	9
	RPL9	8	12
	RHOA	8	9
	EIF5A	8	9
	RAB13	8	8
	RPS24	7	12
	UBE2M	7	7
	RAC1	7	7
	DTYMK	7	7
	UBA52	6	10
	PIN1	6	6
	RPS8	6	6
	PARK7	6	6
	RPS5	6	6
	MYL6	6	6
	ATP5F1	5	6
	RAN	5	6
	PTGES3	5	5
	AP1S2	5	5
	RHER	5	5
			6
		4	5
		4	<u> </u>
		4	4
	BUOC	4	4
	KHUG	4	4
		4	4
	ARLO	4	4
	RAB6A	4	4
	RAB21	4	4
	RPL35	4	4
	CMPK1	4	4
	RPL18	3	4
	SURF4	3	4
	EIF3K	3	3
	AP1S1	3	3
	ARF6	3	3
	ARF4	3	3
	VAMP7	3	3
	ARF5	3	3

Supplementary Figure 1. Pin1 is a binding partner of Kv4.2.

a, Silver stain showing the Kv4.2 complex isolated with a TAP-Kv4.2 pulldown assay in cultured hippocampal neurons. M: marker; Ctl: control. **b**, The numbers of unique and total peptides, and the percent (%) amino acid sequence coverage for each protein that are presented in hippocampal neurons. **c**, High molecular weight proteins that are identified by TAP-Kv4.2 pulldown assay in HEK-293T cells. **d**, Low molecular weight proteins that are identified by TAP-Kv4.2 pulldown assay in HEK-293T cells. **e**, Amino acid sequence coverage obtained for the (human) Pin1 protein from HEK-293T cells. **f**, Pin1 protein sequence report. Xcorr: Sequest cross-correlation score; Δ corr: Xcorr difference between the top ranked and next best sequence.



b

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- Kv4.1 RDFVAAIISIPTPPANTPDESQPSSPGGGGRAGSTLRNSSLGTPCLFPETVKISSL
- Kv4.2 PYVTTAIISIPTPPVTTPEGDDRPESPEYSGGNIVRVSAL
- Kv4.3 SQITTAIISIPTPPALTPEGESRPPPASPGPNTNIPSIASNVVKVSVL



Supplementary Figure 2. Pin1 binding to Kv4.2 is phosphorylation-dependent and Pin1 binds to A-type voltage-gated potassium channels.

a, Lambda protein phosphatase treatment significantly reduced Kv4.2 pull-downed by GST-Pin1, suggesting Pin1 binding to Kv4.2 is phosphorylation-dependent. n = 3 each group. Paired t-test, ***p < 0.001. **b**, Alignment of C-terminal sequences of human A-type voltage-gated potassium channels. Note that the putative Pin1 binding site is conserved among Kv4 family members. Bold residues showed a preferred Pin1 binding context. **c**, Pin1 co-immunoprecipitated (co-IP) with Kv4 members but not Kv3.4 which doesn't contain the putative Pin1 binding site when expressed in HEK-293T cells. Data was repeated in three independent experiments. Data are presented as mean \pm SEM.

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				H M	T602A	T607A	T602,607A	
	602	607					_	– 98kD
Kv4.2 WT	PTPPVT	TPEGDDRPESPEYS		GFP-Kv4.2		-	-	
Kv4.2 T60	2A PAPPVT	TPEGDDRPESPEYS	Kv	v4.2-pT602		-	13	– 98kD
Kv4.2 T60	7A PTPPVT	APEGDDRPESPEYS						
				GFP-Kv4.2		-	_	- 98KD
Kv4.2 T60	2,7A PAPPVT	APEGDDRPESPEYS	Kv	/4.2-pT607				– 98kD



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Supplementary Figure 3. Characterization of Kv4.2 phosphorylation-specific antibodies with Kv4.2 mutants.

a, Sequences of mutated Kv4.2 regions. **b**, Kv4.2 and its point mutants were transfected into HEK-293T cells. Detergent lysates were analyzed by Western blotting with phospho-specific antibodies. Phospho-T602 and phospho-T607 antibodies of Kv4.2 specifically recognize pT602 and pT607 respectively. Data was repeated in two independent experiments. **c**, Kv4.2 were treated with or without Lambda protein phosphatase (PP) before western blot. Phospho-T602 and phospho-T607 antibodies of Kv4.2. Data was repeated in three independent experiments.



Cortex

Cortex

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Supplementary Figure 4. Enriched novel environment and seizure induce Kv4.2 phosphorylation in mouse cortex.

a, Enriched novel environment (EE, 1hr) induces phosphorylation of Kv4.2 Thr607 but not Thr602 in mouse cortex. n = 5 in each group. **p < 0.01. **b**, PTZ-induced seizure (50 mg / kg, i.p., 15 min) induces phosphorylation of Kv4.2 Thr607 but not Thr602 in mouse cortex. n = 6 forctl and 5 for PTZ. T-test, **p < 0.01. Data are presented as mean \pm SEM.



b



Supplementary Figure 5. ERK phosphorylates Kv4.2 in HEK-293T cells and enriched novel environment activates p38 in mouse cortex.

a, Kv4.2 and ERK1 or MEKDD (a constitutively active MEK mutant) constructs were co-transfected into HEK-293T cells. Lysates were analyzed by western blotting with anti-pT602 and anti-pT607 specific antibodies. n = 16 for ctl, 7 for ERK1 and MEKDD. *p < 0.05, **p < 0.01, ***p < 0.001. ERK1 and MEKDD increase Kv4.2 phosphorylation at T602 and T607, but to a small degree. **b**, EE induces the phospho-activation of p38 in mouse cortex. n = 6 in each group. T-test, ***p < 0.001. Data are presented as mean \pm SEM.



Supplementary Figure 6. Kv4.2TA mouse generation.

a, Strategy of Kv4.2 T607A mutant mouse (Kv4.2TA) generation. The mutation was introduced with the Crispr-Cas9 technique. **b**, Genomic DNA was extracted and PCR products were purified for sequencing. Sequencing data verified the mutations were produced as intended. **c**, Brain lysates from WT and Kv4.2TA mice were incubated with Kv4.2 antibody. IP samples were blotted with pT602 and pT607 antibodies. Phosphorylation of Kv4.2 T607 was disrupted in Kv4.2TA mice. The pT602 level is reduced in Kv4.2TA mice, probably because of the decrease of pT602 antibody recognition or pT607's impact on pT602. Data was repeated in three independent experiments. **d**, Immunostaining of Kv4.2 in WT and Kv4.2TA brain showing the Kv4.2 expression pattern was not altered in Kv4.2TA hippocampus. Scale bars: 100 μm. Data was repeated in four independent experiments.



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Supplementary Figure 7. Pin1 inhibition with PiB has no effect on A-current in Kv4.2TA mice.

a, Trace of transient I_A in response to PiB (4 μ M) treatment in Kv4.2TA. Inactivating I_A was isolated by subtracting total K⁺ measured from a step to +40 mV from a -120 mV pre-pulse from a subsequent step to +40 mV from -30 mV. Scale: 10 pA / 100 ms. **b**, I_A density in outside-out patches is not affected by PiB treatment in Kv4.2TA mice (n = 11 for vehicle, 12 for PiB), unpaired two-tailedT-test, p > 0.05. **c**, No significant difference in decay kinetics was observed in response to the treatment and vehicle control, unpaired two-tailedT-test, p > 0.05. Data are presented as mean \pm SEM.



Supplementary Figure 8. Kv4.2TA mice showed normal motor activity in open field test.

a, Total distance traveled in various regions of the open field apparatus by WT and Kv4.2TA mice. The mice were tested for 30 min sessions on two consecutive days. **b**, Time spent in the center or perimeter of the open field apparatus. n = 12 for WT, 16 for Kv4.2TA. Data are presented as mean \pm SEM.

Parameter	WT (mean ± SEM; n)	Kv4.2TA (mean ±SEM; n)
Rise time (ms)	11.69 ±.74; n=14	11.27 ±1.06; n=15
Half-activation (V _H) (mV)	24.76 ±3.47; n=12	34.74 ±4.2; n=11
Half-activation (k) (mV)	18.74 ±74; n=12	19.94 ±1.14; n=11
Half-inactivation (V _H)(mv)	-61.77 ±1.57; n=13	-64.55 ±2.42; n=9
Half-inactivation (k) (mV)	4.60 ±1.00; n=13	5.97 ±0.83; n=9

Supplementary Table 1. Outside-out somatic patch recordings of A-current. Inactivation kinetics, rise time and voltage dependence of activation and inactivation.









Fig. 1e







Fig. 1j



Fig. 1k



Fig. 2a



Kv4.2-pT602





Kv4.2 Kv4.2-pT607 Kv4.2-pT602

Fig. 2c



Kv4.2-pT602

Fig. 2d



Fig. 2f



Kv4.2

GST



Kv4.2



Kv4.2-pT602 Kv4.2-pT607

Fig. 2g



Kv4.2-pT607 Kv4.2-pT602

Fig. 3a

Fig. 3b





Fig. 3c



Fig. 3d

p-P38

Actin

Fig. 3e



Actin

Fig. 3f



Kv4.2-pT607

Fig. 3g



Kv4.2

Kv4.2

Kv4.2-pT607

Fig. 4a



Fig. 4b







Kv4.2

Fig. 4c





Fig. 4d







DPP6







Kv4.2

DPP6

Kv4.2

Fig. 5g





Kv4.2

Kv4.2-pT602

Kv4.2-pT607

Supplementary Fig. 2a



Supplementary Fig. 2b



Supplementary Fig. 3b



Supplementary Fig. 3c



Supplementary Fig. 4a

Supplementary Fig. 4b



Kv4.2 Kv4.2-pT602 Kv4.2-pT607



Supplementary Fig. 5b



p-P38 Actin

Supplementary Fig. 6c

