

Figure S1 - Ruillier et al- Revision

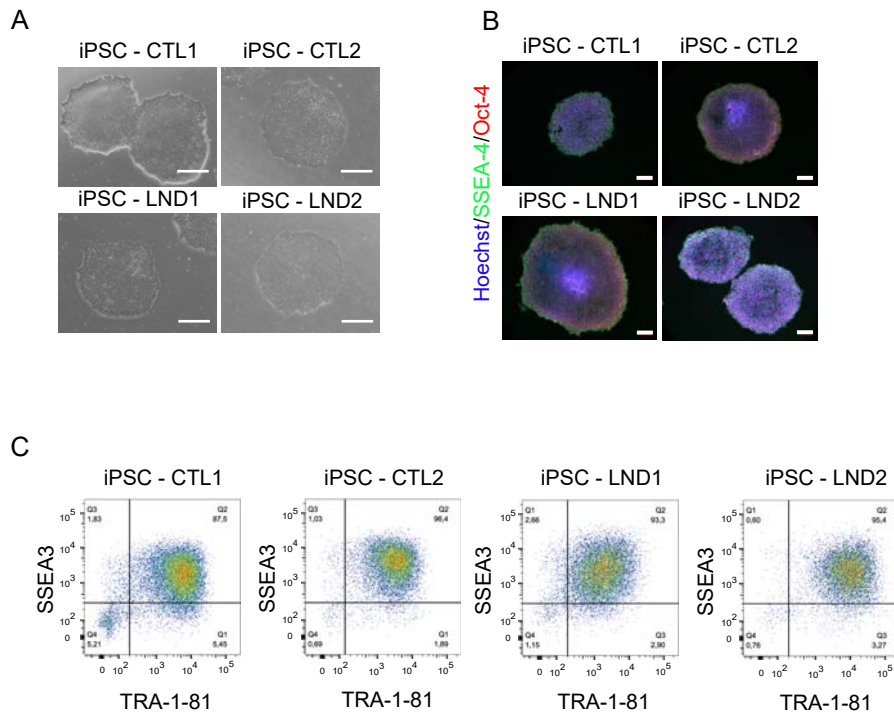


Figure S1: CTL and LND donor-derived iPSC

A - Bright field microscopy of colonies with typical iPSC morphology. Control HGPRT competent cells: CTL1 and CTL2. Lesch-Nyhan disease HGPRT- deficient cells: LND1 and LND2. Scale bar = 200 μ m. B- Representative images of self-renewing HGPRT competent (CTL1 and CTL2) and deficient (LND1 and LND2) iPSC lines expressing the 2 pluripotency markers SSEA-4 (green) and Oct-4 (red) in colonies. Scale bar = 200 μ m. C - Flow cytometry analysis of SSEA3 and TRA-1-81 pluripotency markers.

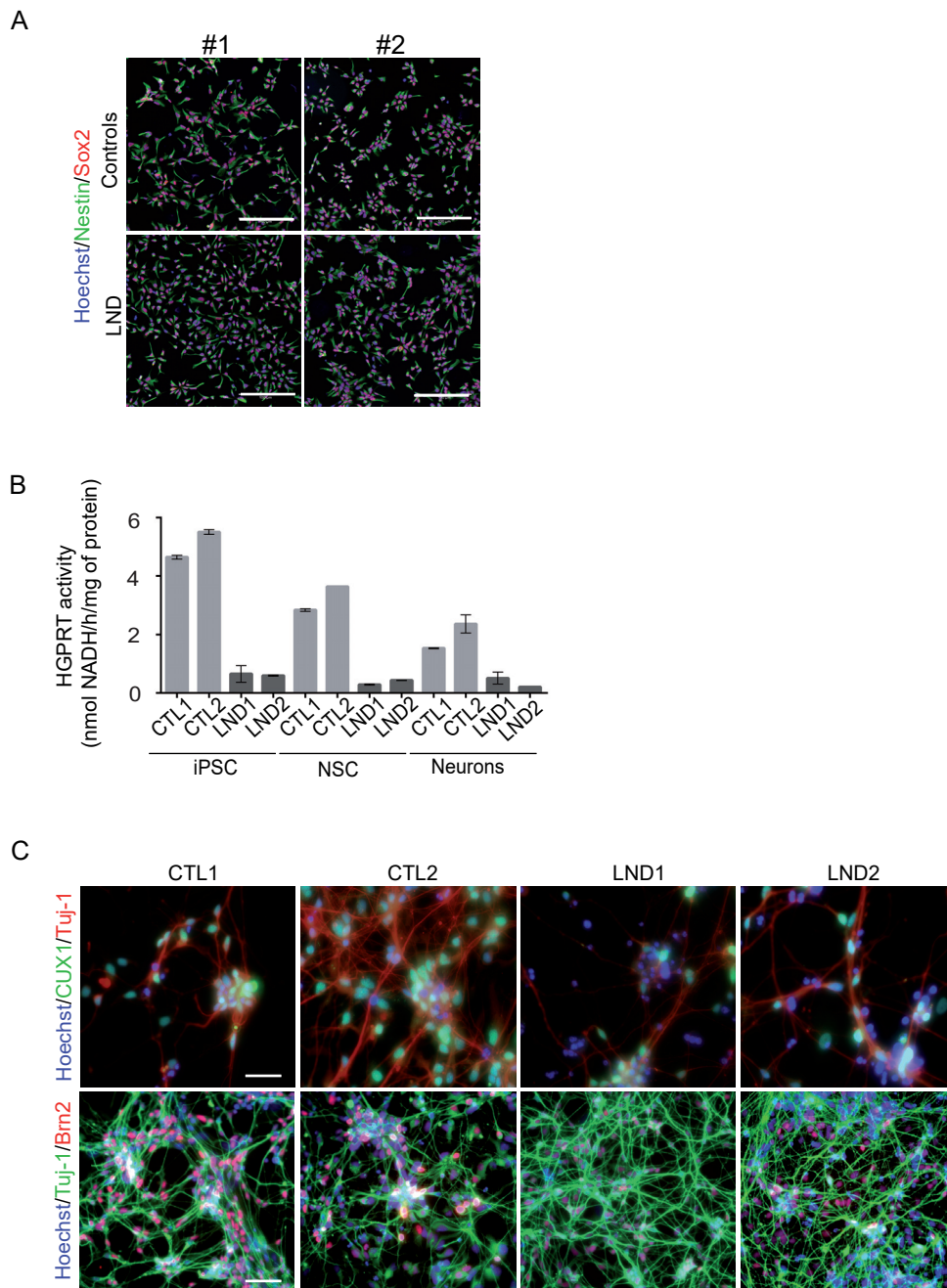


Figure S2: Phenotypical characterization of neurons differentiated from CTL and LND stem cells

A - Representative images of neural stem cells (NSC) expressing Nestin (green) and Sox2 (red). Scale bar = 200 μ m. B - HGPRT enzymatic activity in CTL and LND-derived iPSC, NSC and neurons. The results are expressed as nmol of NADH produced per hour and normalized to total protein content. C – Immunostaining of the cortical superficial layer markers CUX1 and Brn2 in Tuj-1 positive neurons after 18 days of NSC differentiation. Scale bar: CUX1 = 25 μ m, Brn2 = 50 μ m.

Figure S3 - Ruillier et al- Revision

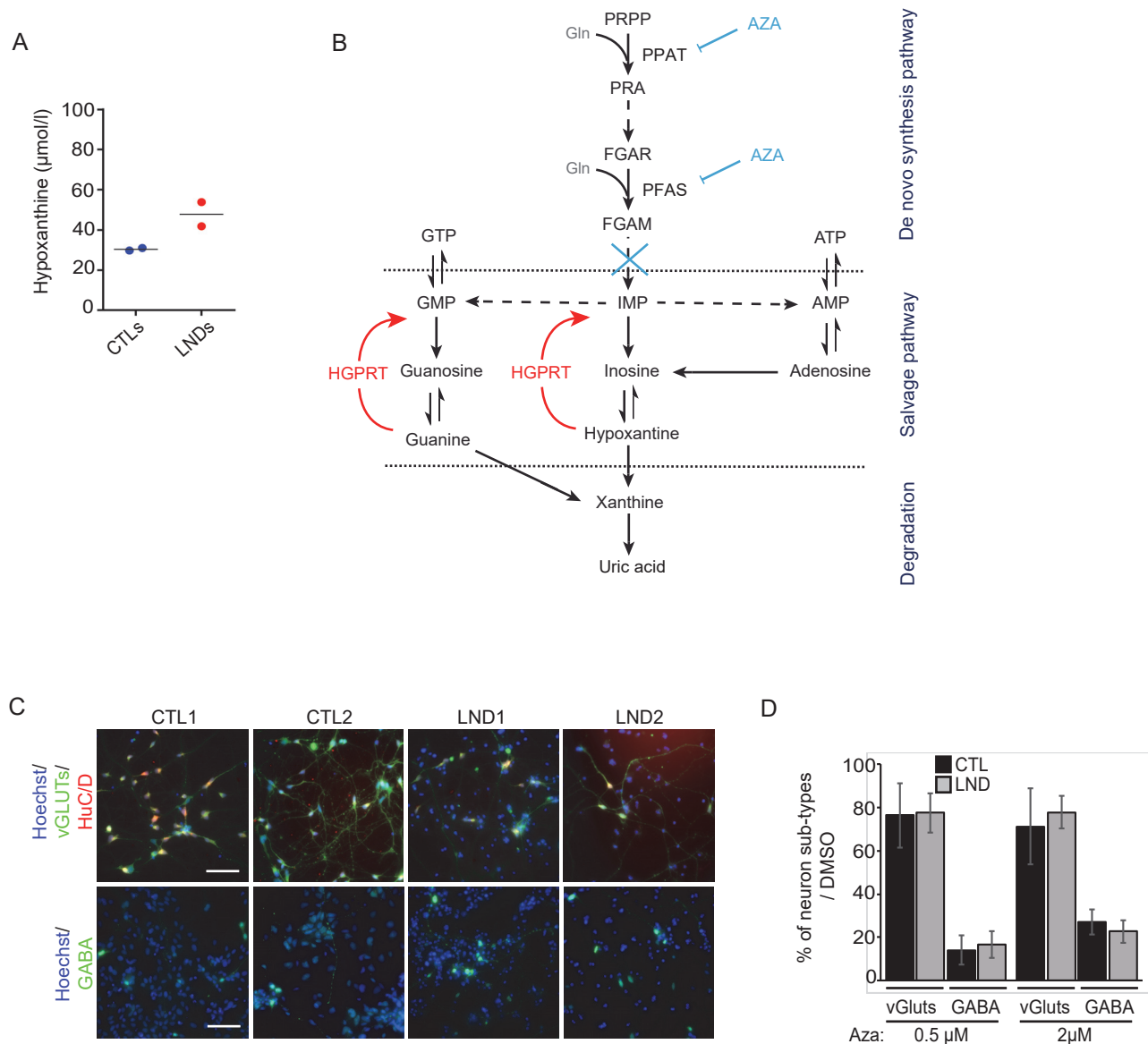


Figure S3: Inhibition of de novo synthesis using asazaserine

A - Quantification of hypoxanthine synthesis in CTL and LND cells. B-. Inhibition of purine de novo synthesis by Azaserine. C- Immunostaining of glutamatergic and GABAergic neuronal sub-types in CTL and LND cells treated with DMSO 0.1% or azaserine 2 µM from day 5 of differentiation. Scale bar = 25µm. D- Quantification of the different neuronal sub-types after DMSO or azaserine treatment. Results are expressed as mean +/- SD of 2 cell lines in 3 biological replicate. C-D: The analysis of neuronal subtypes were performed at day 18 of differentiation.

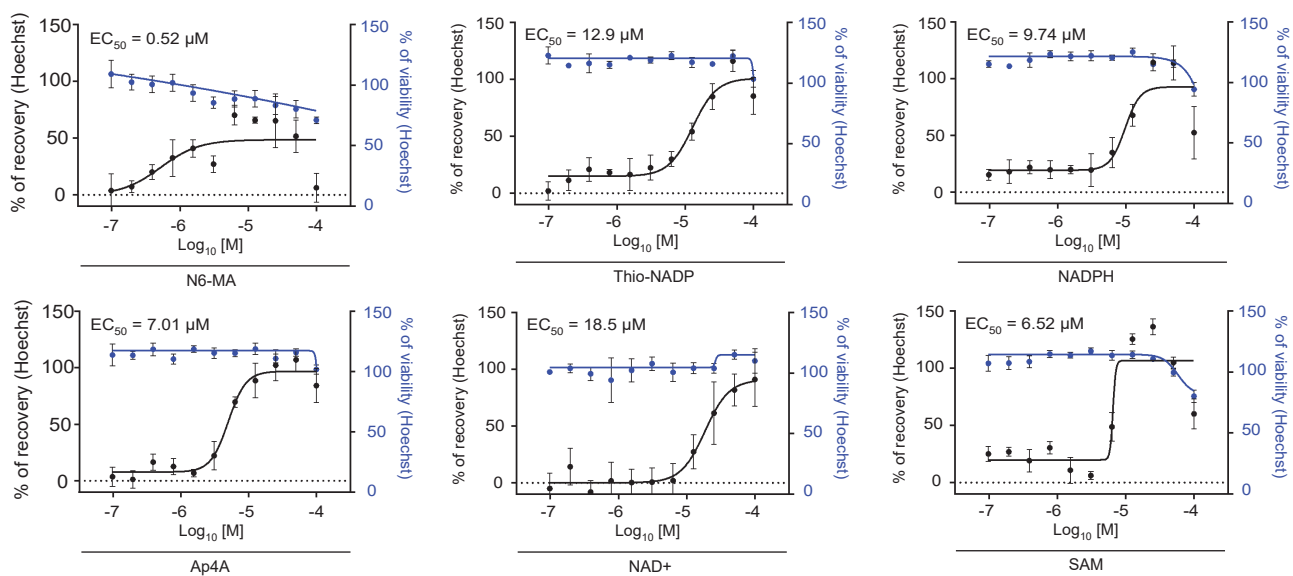


Figure S4: Dose-response analysis of the 6 hit compounds efficiency using Hoechst staining as a read-out.

The black curve corresponds to the percentage of recovery after treatment of LND1 NSC with 1.0 μM azaserine. The blue line shows the percentage of viability after treatment with the drug alone without azaserine, representing the toxicity of the compound itself. Results are expressed as mean ± SD of 4 technical replicates (2 biological replicates).

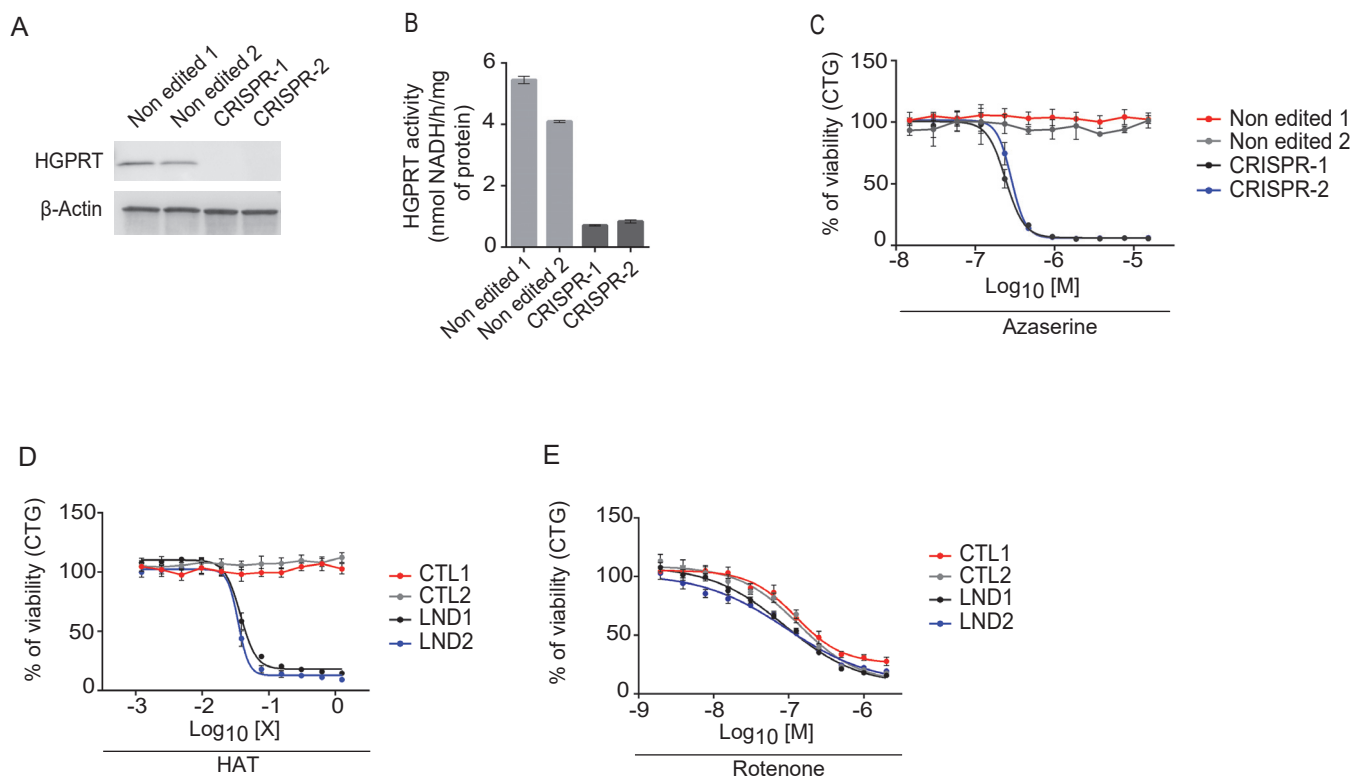


Figure S5: Primary hits compounds specificity.

A- Western-blot analysis of HGPRT protein expression in CRISPR/Cas-9 edited NSC-derived from the human embryonic stem cells line SA001 (CTL3). Two non-edited clones and 2 clones invalidated for HGPRT (CRISPR-1 and CRISPR-2) were compared. β -actin was used as a loading control. B - Enzymatic HGPRT activity measurement in the same clones. C – Azaserine selective toxic effect upon HGPRT-edited CRISPR NSC. Results are expressed as mean \pm SD of 4 technical replicates. CTG: Cell-TiterGlo. D- CTG estimation of cell viability after treatment of CTL and LND cell lines with increasing concentrations of HAT medium (Hypoxanthine, Aminopterin and Thymidine containing medium). The X axis represents HAT medium concentration express as log_{10} (X), where the X is the dilution factor of the commercial HAT complete medium. Results are expressed as mean \pm SD of 4 technical replicates and 2 biological replicates. E- CTG estimation of cell viability after treatment of CTL and LND cell lines with increasing concentrations of rotenone on CTL and LND NSC lines. X axis represents compound concentration expressed as log_{10} (Rotenone concentration in molar M). Results are expressed as mean \pm SD of 4 technical replicates and 2 biological replicates.

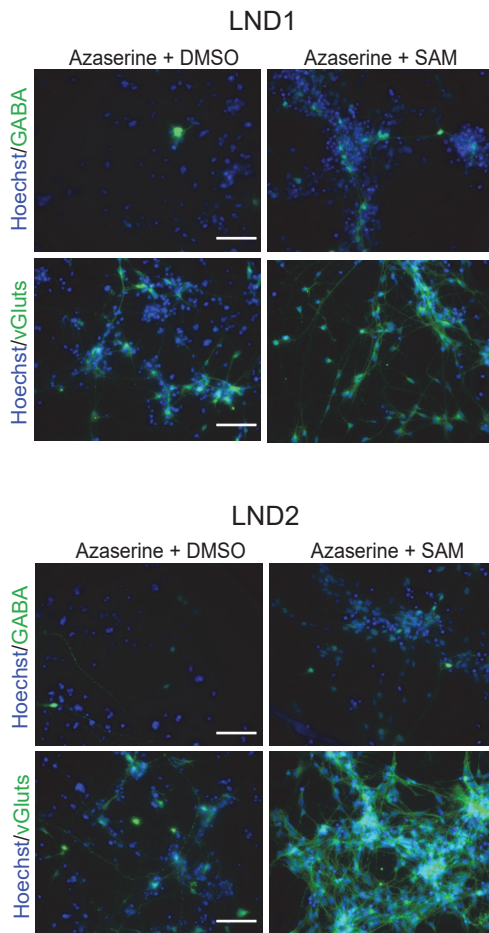


Figure S6: Analysis of neuronal subtypes rescued by SAM after azaserine treatment of LND cells.

GABA: GABAergic neurons, vGluts: Glutamatergic neurons, SAM : S-adenosylmethionine.

Scale bar = 50 μ m.

Table S1: List of CTL and LND hiPSC or hESC lines

Study code	Cell type	Donnor code	Cell of origin	Provider	Karyotype	HGPRT mutation	Clinical description
CTL1	hiPSC	GM01869	fibroblasts	Coriell, USA	46, XY	no	unaffected
CTL2	hiPSC	GM04603	fibroblasts	Coriell, USA	46, XY	no	unaffected
CTL3	hESC	SA001	Blastocyst	Cellartis, Sweden	46, XY	no	unaffected
LND1	hiPSC	GM02227	fibroblasts	Coriell, USA	46, XY	inv/del, ex6-9 Edwards et al, 1989	LND
LND2	hiPSC	GM23784	fibroblasts	Coriell, USA	46, XY	IVS7 + 5 G>A Gibbs et al, 1990	LND

Table S2: List of primary antibodies, providers and dilutions.

Antibody	Host	Reference	Provider	Dilution
HPRT1	Rabbit	15059-1-AP	Proteintech, Rosemont, IL, USA	1:500
PRTFDC1	Rabbit	11986-1-AP	Proteintech, Rosemont, IL, USA	1:250
SSEA-4	Mouse	4755S	Cell Signaling, Danvers, MA, USA	1 :500
Oct-4	Rabbit	2840S	Cell Signaling, Danvers, MA, USA	1 :500
Nestin	Mouse	MAB5326	Merck, Darmstadt, Germany	1 :500
Sox2	Rabbit	AB5603	Merck, Darmstadt, Germany	1:500
Ki-67	Mouse	MAB4190	Merck, Darmstadt, Germany	1:500
HuC/D	Mouse	A-21271	Thermo Scientific, Waltham, MA, USA	1:250
Tubulin β -3 (Tuj-1)	Rabbit	802001	Biolegend, San Diego, CA, USA	1:1000
TRA-1-81 AF647	Mouse	330706	Biolegend, San Diego, CA, USA	2 μ g/ml
SSEA-3 PE	Rat	330312	Biolegend, San Diego, CA, USA	16 μ g/ml

Table S3: Table summarizing the 32 primary hits from LOPAC, Prestwick and SelleckChem libraries.

Library	Conc.	CAS	Compound name	% Recovery	Z-Score Run	Z-Score Plate
LOPAC	10 μ M	102783-36-8	P1,P4-Di(adenosine-5') tetraphosphate triammonium	181.99	39.97	16.68
LOPAC	10 μ M	19254-05-8	Thio-NADP sodium	65.68	14.78	12.87
LOPAC	10 μ M	1867-73-8	N6-Methyladenosine	64.55	14.53	10.39
LOPAC	10 μ M	2646-71-1	NADPH tetrasodium	59.48	13.43	9.6
Prestwick	5 μ M	53-84-9	Nadide	17.11	4.26	6.06
Prestwick	5 μ M	66-81-9	Cycloheximide	13.55	3.49	4.93
LOPAC	10 μ M	85666-17-7	Furegrelate sodium	12.45	3.25	1.29
Prestwick	5 μ M	5536-17-4	Vidarabine	12.30	3.22	4.53
Prestwick	5 μ M	1867-73-8	N6-Methyladenosine	11.29	3.00	4.98
LOPAC	10 μ M	130506-22-8	6-Nitroso-1,2-benzopyrone	9.86	2.69	1.85
Prestwick	5 μ M	21679-14-1	Fludarabine	9.81	2.68	5.62
LOPAC	10 μ M	134523-03-8	Atorvastatin calcium salt trihydrate	9.71	2.66	3.42
LOPAC	10 μ M	38819-10-2	Ara-G hydrate	7.46	2.17	1.91
LOPAC	10 μ M	501-36-0	Resveratrol	7.48	2.17	1.91
Prestwick	5 μ M	73573-88-3	Mevastatin	6.78	2.02	3.29
PW Phyto	10 μ M	6027-98-1	Harmaline hydrochloride dihydrate	100.55	8.88	8.88
SelleckChem	5 μ M	827022-32-2	PD-0332991 (Palbociclib) HCl	20.00	6.57	6.08
SelleckChem	5 μ M	827022-33-3	PD0332991 (Palbociclib) Isethionate	18.66	6.17	7.21
SelleckChem	5 μ M	1144068-46-1	WYE-125132	14.73	5.01	4.83
SelleckChem	5 μ M	1009298-09-2	AZD8055	13.72	4.70	5.48
SelleckChem	5 μ M	741713-40-6	R547	10.40	3.72	3.57
SelleckChem	5 μ M	1207360-89-1	GDC-0349	10.09	3.63	3.41
SelleckChem	5 μ M	343787-29-1	CP-673451	9.9	3.57	3.36
SelleckChem	5 μ M	1009298-59-2	AZD2014	8.56	3.17	3.67
SelleckChem	5 μ M	147526-32-7	Pitavastatin calcium (Livalo)	8.35	3.11	3.59
SelleckChem	5 μ M	1056634-68-4	Momelotinib (CYT387)	5.81	2.36	2.25
SelleckChem	5 μ M	670220-88-9	Crenolanib (CP-868596)	5.56	2.29	2.17
SelleckChem	5 μ M	936890-98-1	OSI-027	5.52	2.27	2.17
SelleckChem	5 μ M	1224844-38-5	INK 128 (MLN0128)	5.46	2.25	2.58
SelleckChem	5 μ M	1223001-51-1	Torin 2	5.46	2.25	2.58
SelleckChem	5 μ M	928037-13-2	Golvatinib (E7050)	5.20	2.18	2.49

SelleckChem	5 μ M	50357-45-4	Pentamidine HCl	5.04	2.13	2.43
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