

Supporting Information

Deficiency in the anti-apoptotic protein DJ-1 promotes intestinal epithelial cell apoptosis and aggravates inflammatory bowel disease via p53

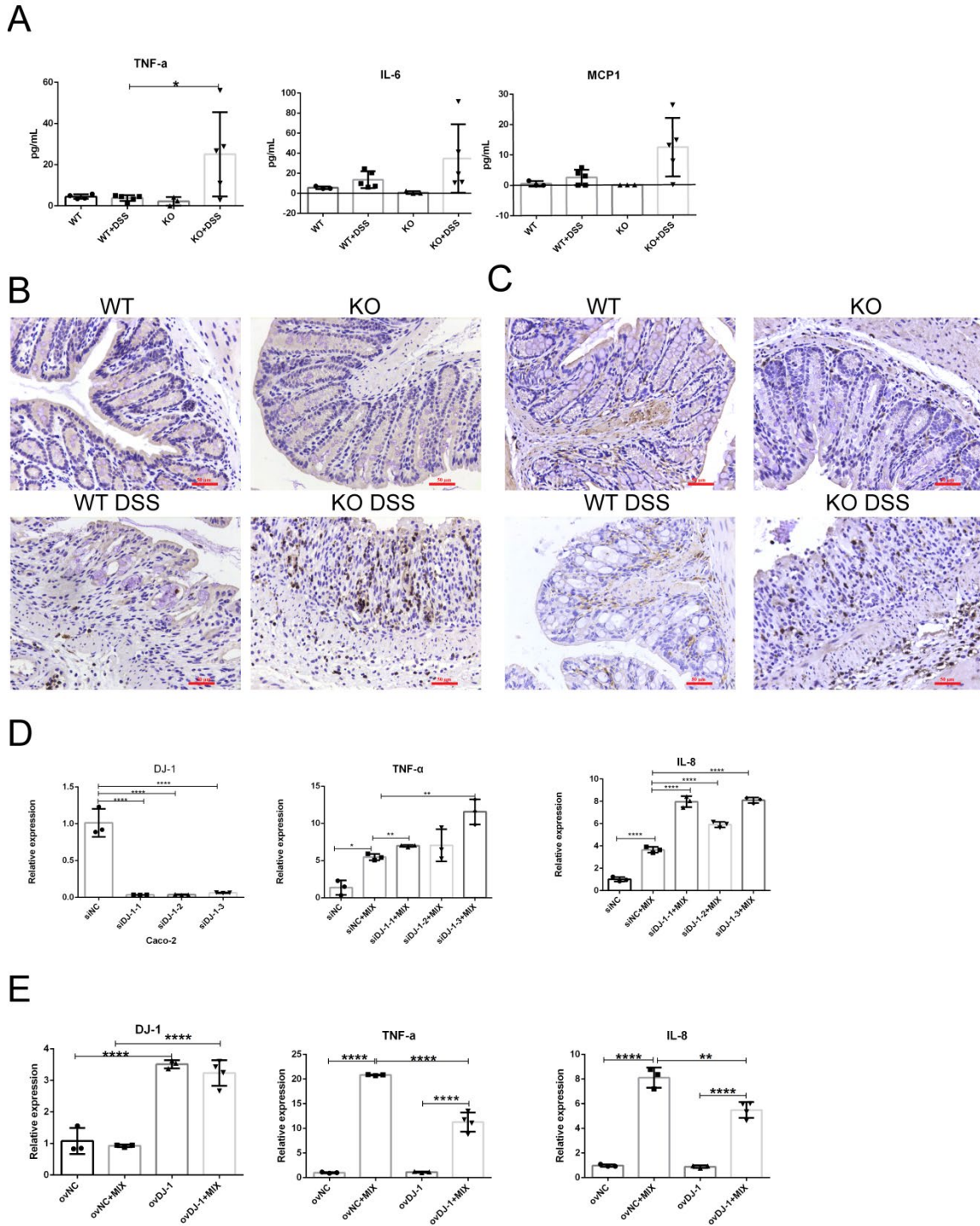
Jie Zhang^{a,*}, Min Xu^{b,*}, Weihua Zhou^a, Dejian Li^a, Hong Zhang^a, Yi Chen^a, Longgui Ning^a, Yuwei Zhang^a, Sha Li^a, Mengli Yu^a, Yishu Chen^a, Hang Zeng^a, Li Cen^a, Tianyu Zhou^a, Xinxin Zhou^a, Chao Lu^a, Chaohui Yu^a, Youming Li^{a,§}, Jing Sun^{c,§}, Xiaoni Kong^{d,§}, Zhe Shen^{a,§}

^aDepartment of Gastroenterology, the First Affiliated Hospital, School of Medicine, Zhejiang University, Hangzhou, 310003, China

^bDepartment of Liver Surgery, Renji Hospital, School of Medicine, Shanghai Jiao Tong University, 160 Pujian Road, Shanghai, 200127, China.

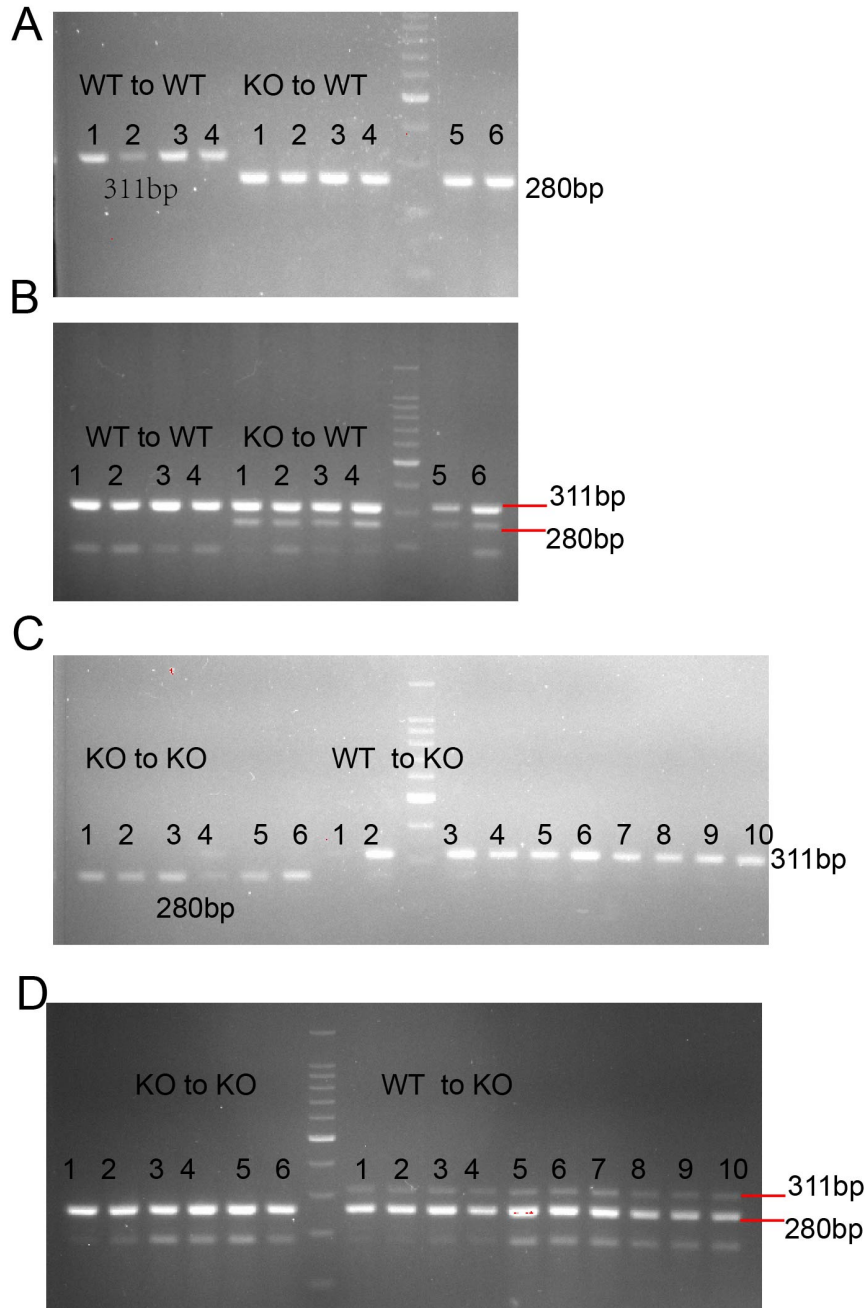
^cDepartment of Gastroenterology, Rui Jin Hospital, Affiliated to Shanghai Jiao Tong University School of Medicine Shanghai, 200031, China

^dInstitute of Clinical Immunology, Department of Liver Diseases, Shu Guang Hospital Affiliated to Shanghai University of Chinese Traditional Medicine, Shanghai, China.



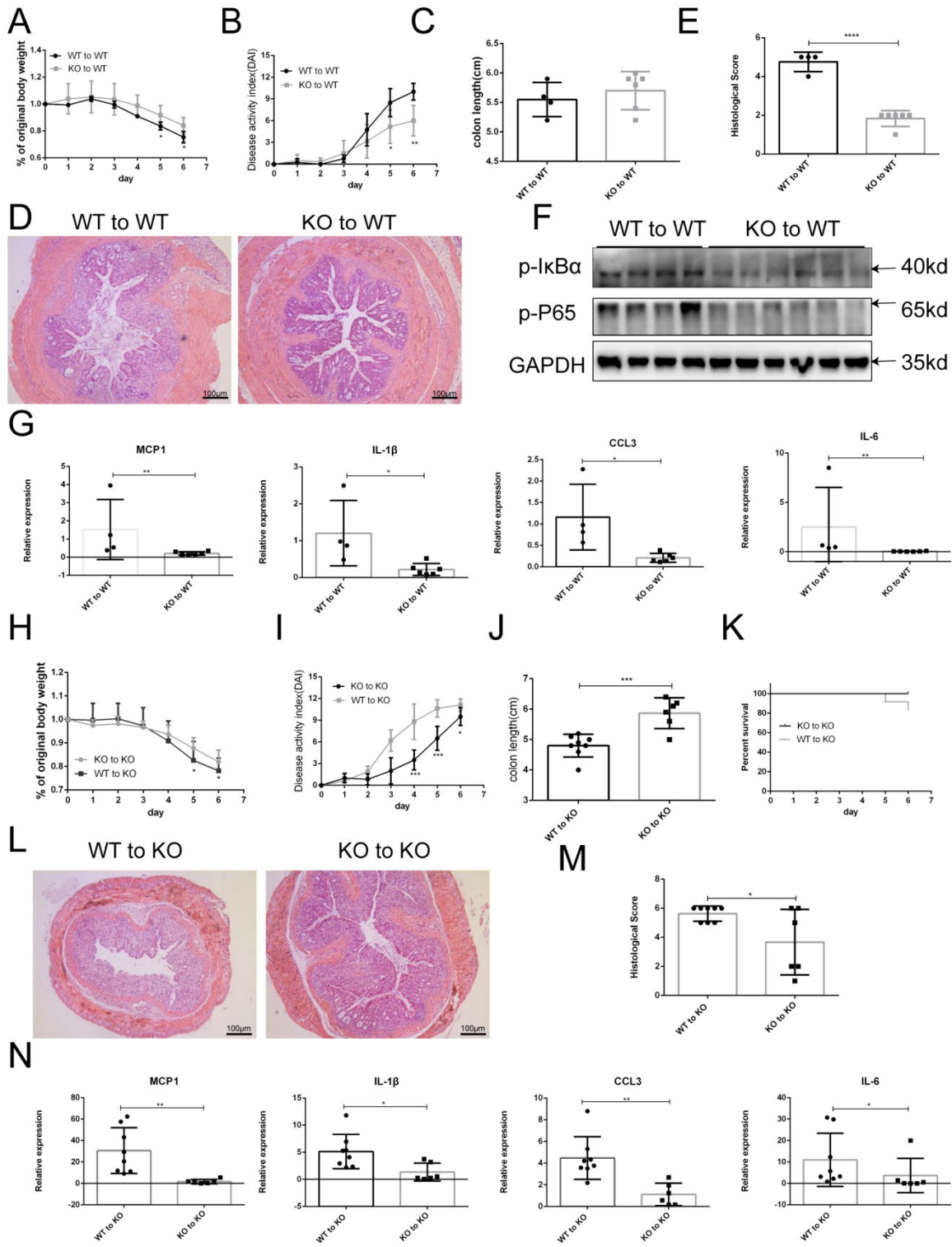
Supporting Figure 1: DJ-1 deficiency contribute to more inflammatory responses in colonic tissues. (A) Relative cytokine levels in mice serum were measured by flow cytometry. Values are expressed as the mean \pm SD, * $P < 0.05$. (B) Representative IHC of the neutrophil infiltration marker myeloperoxidase (MPO) staining in colonic sections from the mice without or after DSS administration (original magnification, 200 \times). (C) Representative IHC of F4/80 in colonic

sections from the mice treated as in A (original magnification, 200×). (D and E) mRNA expression of cytokine and chemokine in Caco-2 cell after knock down (D) or overexpression (E) of DJ-1 for 24 h, followed by MIX treatment for another 24 h. Values are expressed as the mean \pm SD. ** $P < 0.01$, *** $P < 0.001$.



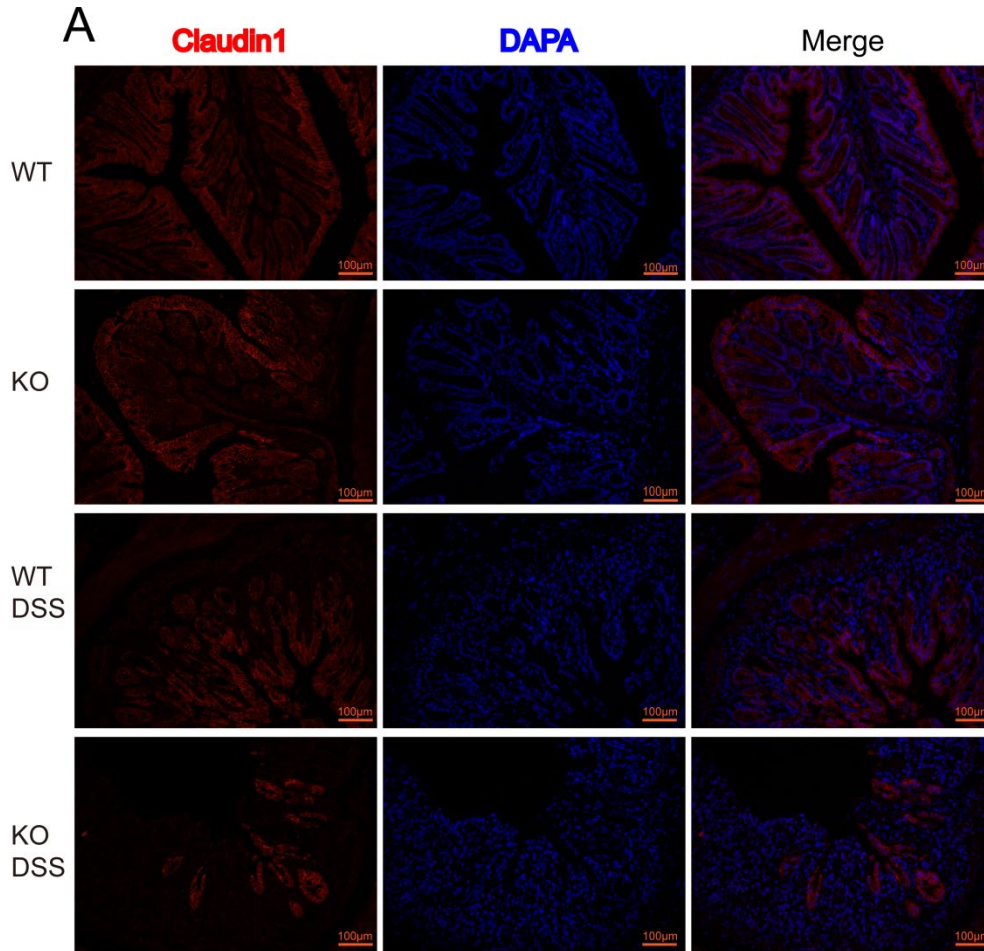
Supporting Figure 2: A: PCR analysis of whole blood cells DNA from WT recipient mice 6 week after radiation demonstrated the bone marrow were successful rebuild in WT recipient. B: PCR analysis of liver tissue from WT recipient mice 6 week after radiation. C: PCR analysis of whole blood cells DNA from KO recipient mice 6 week after radiation demonstrated the bone marrow were successful rebuild in KO recipient. D: PCR analysis of liver tissue from KO

recipient mice 6 week after radiation.

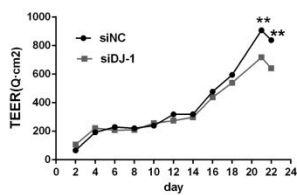


Supporting Figure 3: Absence of DJ-1 in hematopoietic cells is essential for colitis in the

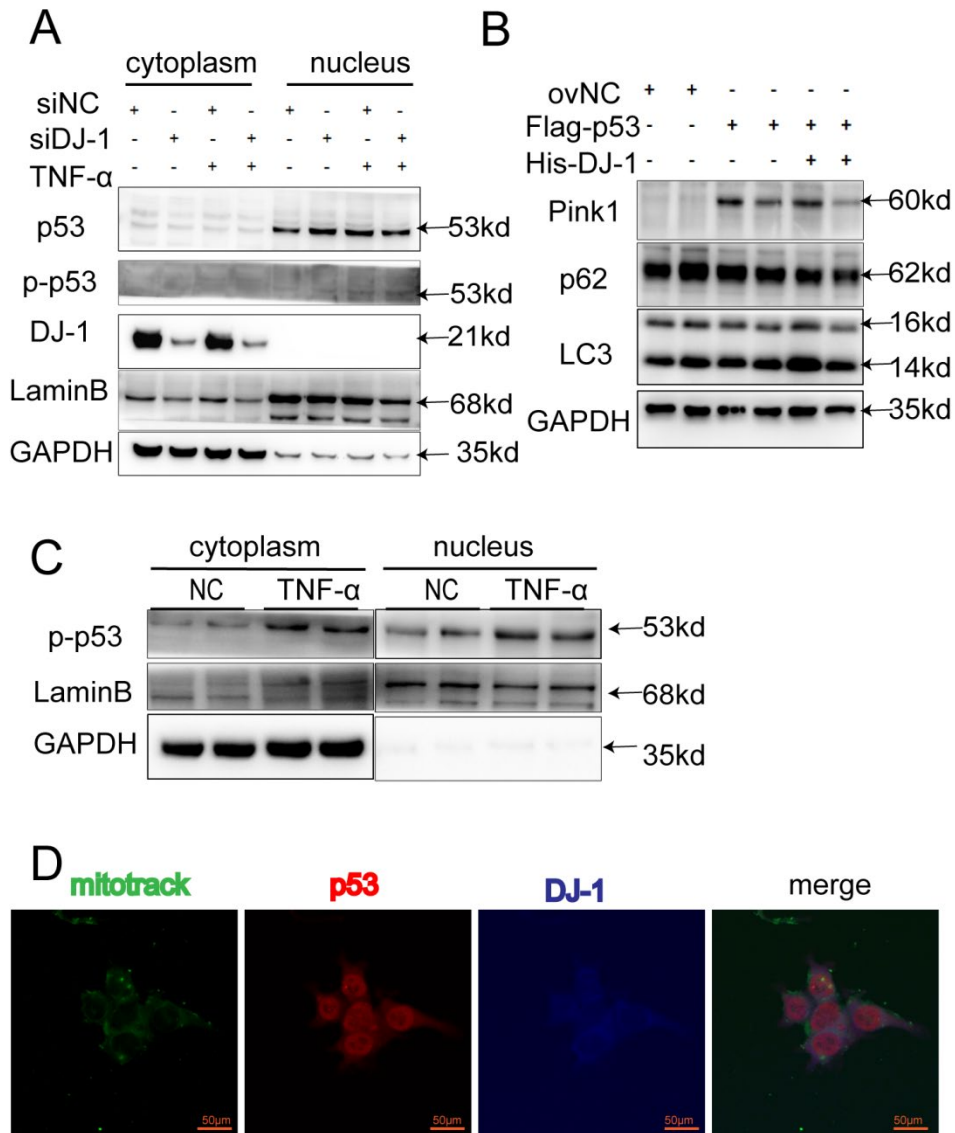
DSS-induced colitis. (A-G) The WT recipient mice received Wild-type bone marrow (WT to WT, $n=4$) and DJ-1 KO bone marrow (KO to WT, $n=6$) mice were treated with 3.5% DSS for 6 days. Body weight (A) and DAI (B) were scored daily. Mice were killed on day 6 and colon lengths (C) were measured. Values are expressed as the mean \pm SD. $**P < 0.01$, $***P < 0.001$. Histological changes (D) in colon tissues from those mice were examined by hematoxylin and eosin staining. Semiquantitative scoring of histopathology (E) (original magnification, 50 \times) was then performed as described in the Experimental Procedures. (G) Relative mRNA levels of cytokine and chemokine in colon. (F) Western blotting of I κ b α -NF- κ b protein expression in colon from mice after DSS administration. (H-N) The KO recipient mice received Wild-type bone marrow (WT to KO, $n=10$) and DJ-1 KO bone marrow (KO to KO, $n=6$) mice were treated with 3.5% DSS for 6 days. Body weight (H), DAI scores (I), colon lengths (J) and survival rate (K) were measured. Histological changes (L) (original magnification, 50 \times) by HE and Semiquantitative scoring of histopathology (M) were then performed. (N) Relative mRNA levels of cytokine and chemokine in colon. Values are expressed as the mean \pm SD. $**P < 0.01$, $***P < 0.001$.



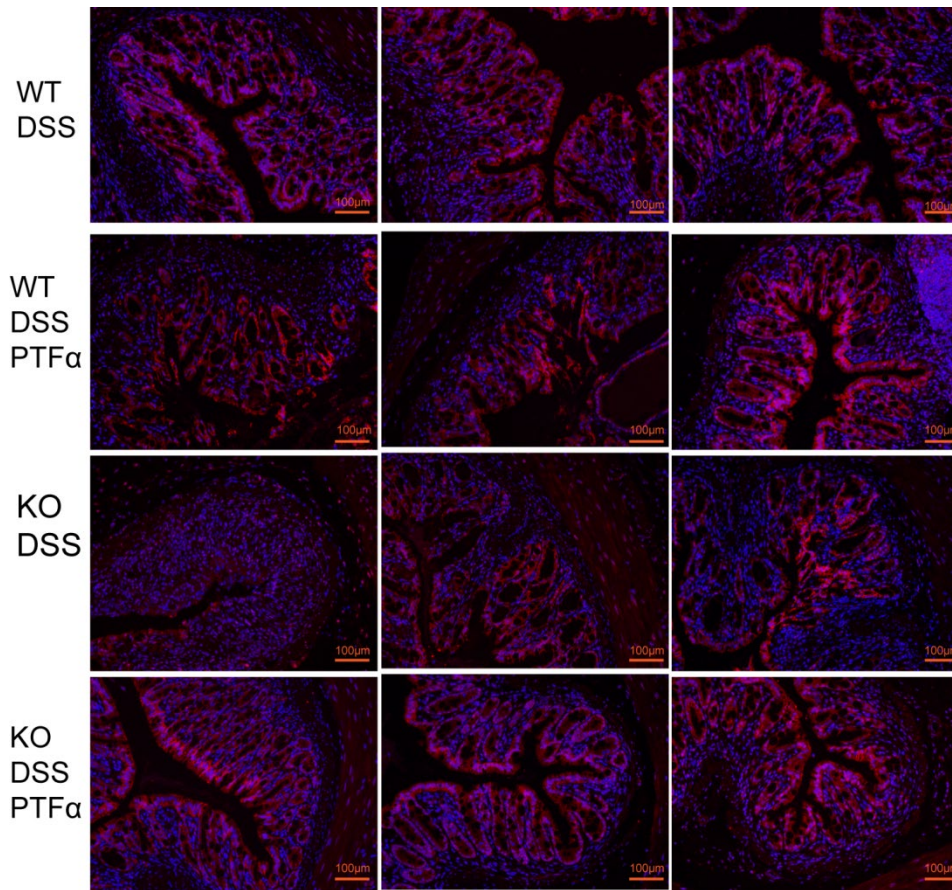
B



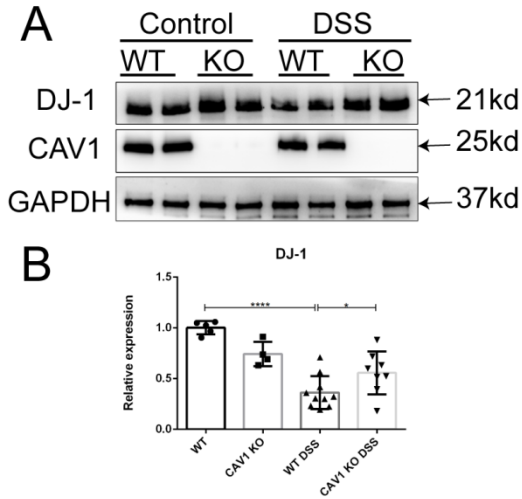
Supporting Figure 4: (A) The immunostaining of cell tight junction protein Claudin1 (Red) staining in colonic sections from the mice treated as in Figure 2 (original magnification, $\times 200$). (B) Caco-2 cells were cultured on the transwell inserts and DJ-1 were knock down by siRNA every week, when the TEER reach $800\Omega \cdot \text{cm}^2$, transwell inserts were inserted into multiple plate wells containing RAW264.7 cells. To imitate the colon inflammation, LPS ($1\mu\text{g/ml}$) was added to the basolateral compartment of this co-culture system for 24 h.



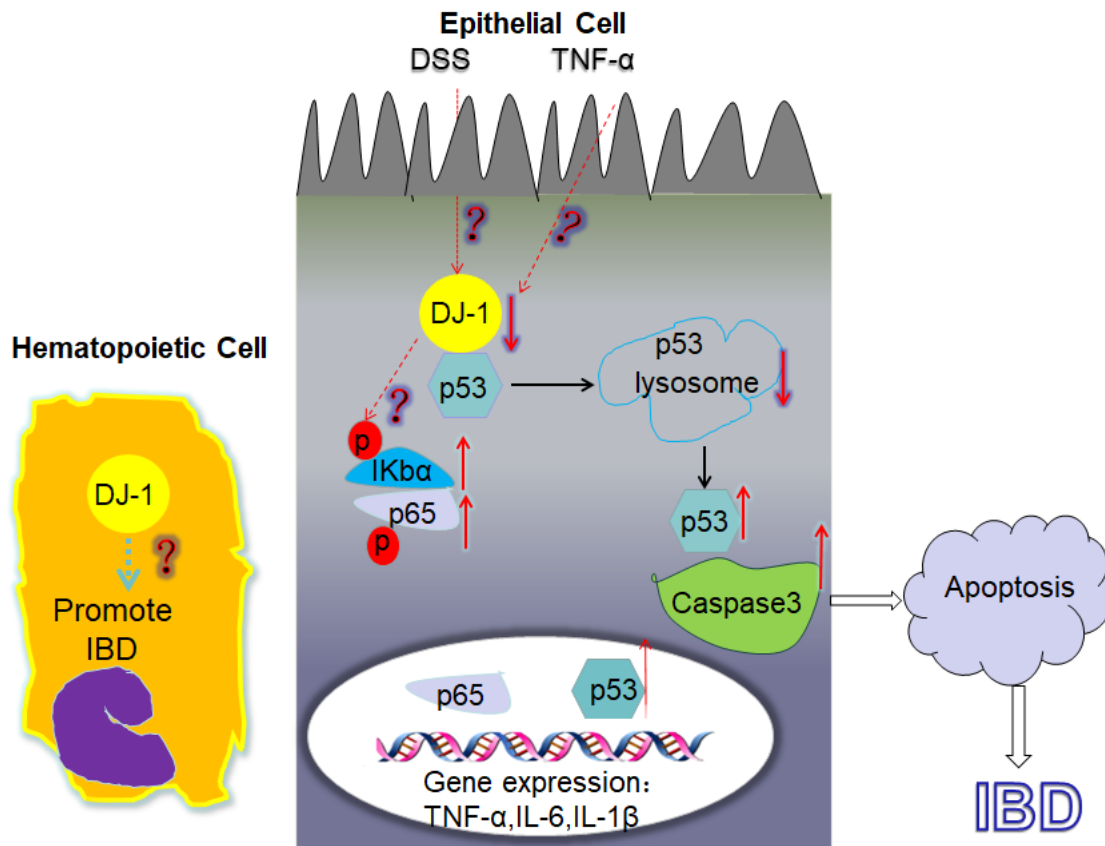
Supporting Figure 5: (A) DJ-1 was knocked down in HCT116 cells by siRNA for 24 h and then the cells were stimulated with 100 ng/mL TNF- α for 24 h. Protein were isolated from cytoplasm and nucleus. (B) 293T cell infected with Flag-tagged p53 and his-tagged DJ-1, and stimulated with 100 ng/mL TNF- α for 24 h.(C) HCT116 cell were stimulated with 100 ng/mL TNF- α for 24 h, Protein were isolated from cytoplasm and nucleus. (D) Representative immunofluorescent staining of mitotrack (green), DJ-1-specific antibody (anti-DJ-1, blue) and p53 antibody (anti-p53, red) (original magnification, 400 \times) in HCT116 cell treated with TNF- α for 24 h.



Supporting Figure 6: PFT- α was administered by intraperitoneal injection into WT mice ($n=8$) and DJ-1^{-/-} mice ($n=6$), and the control group (WT $n=5$, KO $n=7$) was intraperitoneally injected with DMSO. Claudin1 (red) staining of colon sections of mice treated as before. Nuclei were stained with DAPI (blue) (original magnification, 200 \times).



Supporting Figure 7: (A-B) Wild-type ($n=10$) and CAV1 KO ($n=8$) mice were treated with 3.5% DSS for 6 days. The DJ-1 protein level (A) and mRNA (B) level were measured in the colon tissue.



Supporting Figure 8: Mechanism schematic diagram of DJ-1 regulating IBD.

Table S1: Antibodies for Western blot

Primary antibody	Company	Cat#
DJ-1	abcam	#18257 (WB) # 76008 (IP)
p53	abcam	#26 (WB for mouse) #3133(for human)
p-P65	CST	#3039
p-JNK	CST	#4668
Actin	CST	#3077
GAPDH	CST	#5174
p-I κ b α	CST	#9246
p-IKK α/β	CST	#2697
Cleaved Caspase-3	CST	#9661
Tubulin	CST	#2148
Cleaved Caspase-7	CST	#8438
BAX	CST	#2772
PUMA	CST	#4976

Table S2: Primers used for qPCR

Gene	Primer sequence
M DJ-1 F	AAAACGCAGGGACCATACGAT
M DJ-1 R	AGGCGACTCAGATAAATTCTGTG
M MCP1 F	TTAAAAACCTGGATCGGAACCAA
M MCP1 R	GCATTAGCTTCAGATTTACGGGT
M CCL3 F	TTCTCTGTACCATGACACTCTGC
M CCL3 R	CGTGGAATCTTCCGGCTGTAG
M IL-8 F	TGTTGAGCATGAAAAGCCTCTAT
M IL-8 R	AGGTCTCCCGAATTGGAAAGG
M TNF- α F	CAGGCGGTGCCTATGTCTC
M TNF- α R	CGATCACCCCGAAGTTCAGTAG
M GAPDH F	AGGTCGGTGTGAACGGATTTG
M GAPDH R	GGGGTCGTTGATGGCAACA
M IL-6 F	TAGTCCTTCCTACCCCAATTTCC
M IL-6 R	TTGGTCCTTAGCCACTCCTTC
M IL-1 β F	CTCACAAGCAGAGCACAAGC
M IL-1 β R	TCCAGCCATACTTTAGGAAGA
H GAPDH F	TCAACGACCACTTTGTCAAGCTCA
H GAPDH R	GCTGGTGGTCCAGGGGTCTTACT
H TNF- α F	GAGGCCAAGCCCTGGTATG
H TNF- α R	CGGGCCGATTGATCTCAGC
H DJ-1 F	AACCGGAAGGGCCTGATAG
H DJ-1 R	GCAAGAGGGTGTGTTGTA ACT
H IL-8 F	ACTGAGAGTGATTGAGAGTGGAC
H IL-8 R	AACCCTCTGCACCCAGTTTTC