

Supplemental material

On the Origin of the Anomalous Behavior of Lipid Membrane Properties in the Vicinity of the Chain-Melting Phase Transition

A. Kuklin, D. Zabelskii, I. Gordeliy, J. Teixeira, A. Brûlet, V. Chupin, V. Cherezov and V. Gordeliy

December 16, 2019

SANS data

Representative examples of the SANS diffraction data of 4% (v/v) DMPC MLVs in D_2O are shown in Figure S1A for $T = 18^\circ\text{C}$, 23°C and 24°C . The data are presented in Iq^2 vs q coordinates (Kratky-Porod plot) after normalization and background subtraction. Multiplication of the scattering intensity $I(q)$ by the Lorentz factor q^2 was done to correct for the scattering from randomly-oriented multilayers. The first order diffraction peak around $q = 0.1\text{\AA}^{-1}$ is clearly visible at all temperatures, and an additional peak around $q = 0.05\text{\AA}^{-1}$, although weak, is visible at $T = 18^\circ\text{C}$. It corresponds to a ripple phase of DMPC with the periodicity of 120\AA . The repeat distances, d , of multilayers were calculated using the Bragg equation: $2d \sin \theta = n\lambda$ [1].

For SANS experiments with 1% (v/v) DMPC ULVs in D_2O selected scattering curves are shown for $T = 18$ and 25°C (Figure S1B). The results are plotted using the Kratky-Porod plot in log-lin representation according to the asymptotic expression for the scattering from lipid bilayers [2]:

$$I(q) = \frac{I_0}{q^2} \exp(-R_T^2 q^2) \quad (\text{S1})$$

which can be applied when $1/\sqrt{S} < q < 1/R_T$ where S is the vesicle surface area and R_T is the bilayer radius of gyration which is given by:

$$R_T^2 = \frac{\int_S \rho(z) z^2 dz}{\int_S \rho(z) dz} \quad (\text{S2})$$

Calculations of the bilayer thickness using R_T were done by using $d_b = R_T \sqrt{12}$ with subsequent correction for the hydration of polar lipid heads and inhomogeneity of the scattering [3]:

$$d'_b = d_b \left(1 - 1.26 \frac{V_W}{V_L} \right) \quad (\text{S3})$$

where $V_W = n_W \nu_{WL} = 7.2 \cdot 30\text{\AA}^3 = 216\text{\AA}^3$ is the volume of water molecules inside the bilayer and V_L is the volume of a DMPC molecule ($V_L = 1101\text{\AA}^3$ [4])

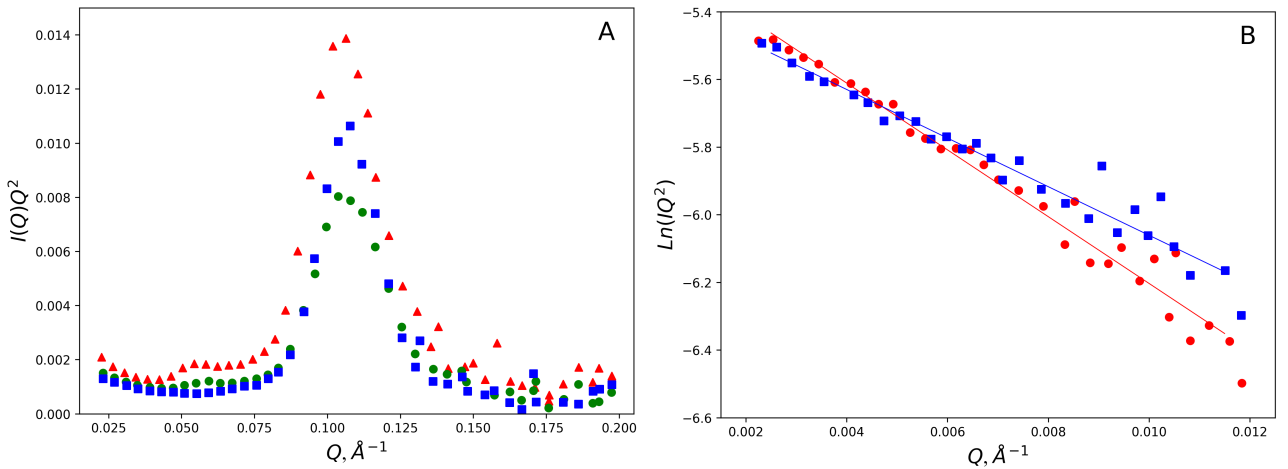


Figure S1: (A) Representative examples of diffraction data for 4% (v/v) DMPC MLVs in D_2O presented in Iq^2 vs q^2 plot at $T = 18^\circ\text{C}$, 23°C and 24°C and colored in red, green and blue, respectively. (B) Examples of SANS data for 1% (v/v) DMPC ULVs in D_2O presented in $\ln(Iq^2)$ vs q^2 plot at $T = 18^\circ\text{C}$ and 25°C and colored in red and blue, respectively

NSE spectra fitting

Spin echo signal from undulating vesicles consist of two different parts: Static form-factor of a vesicle, which corresponds to the contribution of the diffusion term and quasielastic form-factor which accounts for undulations of the vesicle:

$$I(q, t, r) = e^{-Dq^2t} V_s^2 (\Delta\rho)^2 I_1(q, t, r) \quad (\text{S4})$$

where I_1 is given by:

$$I_1(q, t, r) = \left(f_0(qr) + \sum_{l=1} \frac{2l+1}{4\pi} \langle a_l(0)a_l(t) \rangle \right) \quad (\text{S5})$$

$f_0 = (\sin(qr)/qr)^2$ is the static form factor, $f_1 = ((l+2)j_l(qr) - qrj_{l+1}(qr))^2$ is the quasielastic form-factor, r is the radius of a vesicle, V_s is its scattering volume, $\Delta\rho$ is the scattering density contrast, j_l is the l_{th} Bessel function, $D = kT/6\pi\eta r^*$ is the diffusion coefficient, where $r^* = r + 25\text{\AA}$ is the hydrodynamic radius, η is the D_2O viscosity, k_B is the Boltzmann constant and $\langle a_l(0)a_l(t) \rangle = \langle a_l^2 \rangle \exp(-\omega_l t)$ is the autocorrelation function of the undulation amplitude, where the average undulation amplitude is given by:

$$\langle |a_l^2| \rangle = \frac{k_B T l^2 (l+1)^2 (l+2)(l-1)}{K_c (2l+1)(2l^2+2l+1)} \quad (\text{S6})$$

K_c is the bending rigidity and l is the undulation mode. The undulation frequency is given by the expression:

$$\omega_l = \frac{K_c l^2 (l+1)^2 (l+2)(l-1)}{\eta r^3 (2l+1)(2l^2+2l+1)} \quad (\text{S7})$$

The polydispersity of vesicles is taken into account by a Gaussian distribution:

$$p(r) = \sqrt{\frac{1}{2\pi\sigma^2}} \exp\left(-\frac{(r-R_0)^2}{2\sigma^2}\right) \quad (\text{S8})$$

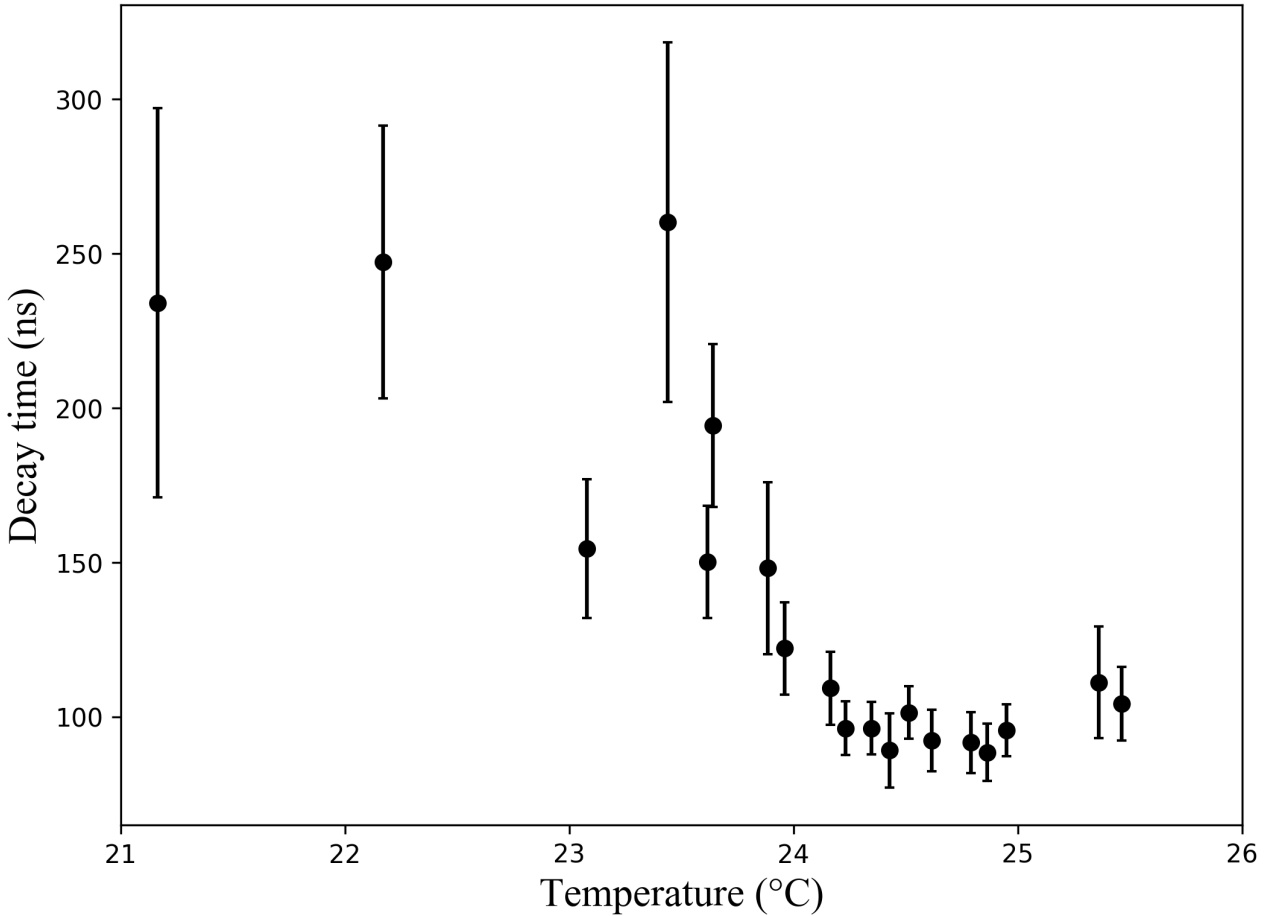


Figure S2: Temperature dependence of the decay time for 4% (v/v) DMPC ULVs in D_2O obtained with single exponential fits of the NSE data near the chain-melting phase transition region.

The scattering intensity from polydisperse vesicles can be written as:

$$I(q, t) = \int_{r_{min}}^{r_{max}} I(q, t, r)p(r)dr \quad (\text{S9})$$

For vesicles made with a small volume extrusion apparatus, the polydispersity $\sigma_R \approx 0.3R_0$, as it was shown previously in [5]. For diffusion motion of rigid vesicles, the NSE decay time is given by:

$$\tau = \frac{1}{Dq^2} = \frac{6\pi\eta r^*}{kTq^2} \quad (\text{S10})$$

The NSE data can be ambiguously treated with any number of exponents in quasielastic form-factor, although NSE signal can be easily overfitted. The decay times of the MLVs obtained from single exponential fits of the data are presented at Figure S2. The data clearly demonstrate that decay time dependence decreases by factor of 3, when it should nearly the same, that indicate the strong contribution from the undulation term which should be taken into account during data processing.

References

- [1] V. Gordeliy and M. Kiselev, "Definition of lipid membrane structural parameters from neutronographic experiments with the help of the strip function model," *Biophysical journal*, vol. 69, no. 4, pp. 1424–1428, 1995.
- [2] W. Knoll, K. Ibel, and E. Sackmann, "Small-angle neutron scattering study of lipid phase diagrams by the contrast variation method," *Biochemistry*, vol. 20, no. 22, pp. 6379–6383, 1981.
- [3] V. Gordeliy, L. Golubchikova, A. Kuklin, A. Syrykh, and A. Watts, "The study of single biological and model membranes via small-angle neutron scattering," in *Trends in Colloid and Interface Science VII*, pp. 252–256, Springer, 1993.
- [4] J. F. Nagle and S. Tristram-Nagle, "Structure of lipid bilayers," *Biochimica et Biophysica Acta (BBA)-Reviews on Biomembranes*, vol. 1469, no. 3, pp. 159–195, 2000.
- [5] R. C. MacDonald, R. I. MacDonald, B. P. M. Menco, K. Takeshita, N. K. Subbarao, and L.-r. Hu, "Small-volume extrusion apparatus for preparation of large, unilamellar vesicles," *Biochimica et Biophysica Acta (BBA)-Biomembranes*, vol. 1061, no. 2, pp. 297–303, 1991.