

Figure S1. Characterization of TLR7 and TLR8 in BLaER1 Monocytes, Related to Figure 1

(A) Volcano plot showing gene expression differences of differentiated versus un-differentiated BLaER1 cells. The negative log_{10} pvalues (y axis) are plotted against the log_{10} fold changes in gene expression (x axis). Significantly (adjusted p-value <0.05 and absolute fold change >2) upregulated genes are highlighted in red, downregulated genes are highlighted in blue. TLR4, 7 and 8 are specifically highlighted. (B) BLaER1 Ctrl. and *TLR8-/-* cells were stimulated using Lipofectamine 2000 with and without RNA40^S. (C) BLaER1 cells were stimulated with different amounts of RNA40^O. The unstimulated control shown in (B) and (C) is the same as in Figure 1D, as it is derived from the same experiment. (D) Different RNAs were tested for TLR8 activation. P20 is a self-complementary RNA forming hairpins, whereas P20-5M is only a partially self-complementary RNA. Data are depicted as mean + SEM of three (B and C) or four (D) independent experiments. Statistics indicates significance by two-way ANOVA: ***p ≤ 0.001, **p ≤ 0.01, *p ≤ 0.05, ns = not significant.



Figure S2. *RNASE1^{-/-}*, *RNASE2^{-/-}*, and *RNASE6^{-/-}* Cells Show No Reduced Response to RNA40 Stimulation, Related to Figure 2

(A–C) BLaER1 control, *RNASE1, RNASE2* and *RNASE6*-deficient cells were stimulated with RNA40^S, TL8-506, R848 and LPS. Controls are identical for *RNASE1-/-* and *RNASE6^{-/-}* cells as they derive from the same experiment. **(D)** THP-1 wild type, *RNASET2^{-/-}* and *TLR8^{-/-}* cells were stimulated as indicated. **(E)** BLaER1 control and *RNASET2^{-/-}* cells were stimulated as indicated. a they derive from the same experiment. Data are depicted as mean + SEM of three (A–D) or four (E) independent experiments. Statistics indicates significance by two-way ANOVA: ****p ≤ 0.0001; ***p ≤ 0.01, *p ≤ 0.01, *p ≤ 0.05, ns = not significant.



Figure S3. Identifying Possible RNase T2 Degradation Products by *In Vitro* Digestion of RNA40, Related to Figure 3

(A) List of all possible fragments associated with the *in vitro* digest of RNA40^s with RNase T2, analyzed by HPLC/MALDI-TOF. All found and calculated ([M-H]⁻) masses are shown. The color code refers to the probability that fragments could be assigned to the one depicted. (B) Most likely hits of RNA40^s digested with RNase T2 based on the analysis from (A). (C) Assuming RNase T2 is only capable of cleaving RNA40 between G and U residues, all possible fragments are depicted. The ticks indicate which of these fragments could be confirmed by HPLC/MALDI-TOF. (D) Urea gel of *in vitro* digested RNA P20 and P20-5M with RNase T2. P20 is a self-complementary RNA fragment and forms hairpins, whereas P20-5M is only partially selfcomplementary RNA. One representative gel of two independent experiments is shown.



Figure S4. Cells Deficient in RNase A Family Enzymes Show Different RNA40 Catabolism in Comparison with *RNASET2^{-/-}* Cells, Related to Figure 4

(A) Whole cell lysate of unstimulated BLaER1 cells with indicated genotypes were analyzed by LC-MS. Data are normalized to Ctrl. cells (note logarithmic scale). (B–D) Whole cell lysate of control, $RNASE1^{-/-}$, $RNASE2^{-/-}$ and $RNASE6^{-/-}$ BLaER1 cells stimulated with RNA40^s was analyzed by LC/MS (14 h after stimulation). RNA40^s derived metabolites are shown. Data are normalized to Ctrl. cells (note logarithmic scale). Data are depicted as mean + SEM of three independent experiments. Statistics indicates significance by a Welch's unequal variances t test. ***p ≤ 0.001, **p ≤ 0.01, *p ≤ 0.05; ns, not significant; n.d., not detected.



Figure S5. Crystal Structure of the Second Pocket of TLR8, Related to Figure 5

Crystal structure of the second pocket of TLR8 bound to UG (PDB, 4R07). Yellow dashed lines indicate hydrogen bonds whereas green dashed lines show π - π -interactions. The atoms are color coded as follows: nitrogen, blue; oxygen, red; phosphor, orange; carbon, light blue and gray.



Figure S6. RNase T2 Degradation Products Bypass the Lack of RNase T2, Related to Figure 6

(A) Urea gel of full-length and RNase T2 digested (dAdC)₇-UUGUCU. (B) BLaER1 cells of indicated genotypes were stimulated with either digested or undigested (dAdC)₇-UUGUCU (0.9 μ g/condition) and the indicated controls. (C) Urea gel of the following full-length ONs: (dAdC)₇-UUGUCU, (dAdC)₇-UUG, (dAdC)₇-UU and (dAdC)₈. (D) Control stimulation of BLaER1 cells associated with (Figure 6G). (E) Control stimulation of BLaER1cells associated with (Figure 6H). Data are depicted as mean + SEM of three independent experiments or one of two representative gels is shown. Statistics indicates significance by two-way ANOVA. ****p ≤ 0.0001; ***p ≤ 0.001, **p ≤ 0.05, ns = not significant.



Figure S7. ¹⁵N-Labeled *S. aureus* Detection in Myeloid Cells Depends on RNase T2 Upstream of TLR8, Related to Figure 7

(A) Schematic overview of experimental setup. *S. aureus* was grown in ¹⁵N-labeled medium, harvested and used to stimulate differentiated BLaER1 cells. 1 h after infection gentamicin was added and another 14 h later IL-6 release was measured by ELISA. (B) Stimulation of BLaER1 cells with different MOI of ¹⁵N-labeled *S. aureus*. Data are depicted as mean + SEM of three independent experiments. Statistics indicates significance by two-way ANOVA. ****p \leq 0.0001; ***p \leq 0.001, **p \leq 0.01, *p \leq 0.05, ns = not significant.