

**ADVANCED  
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MATERIALS**

Supporting Information

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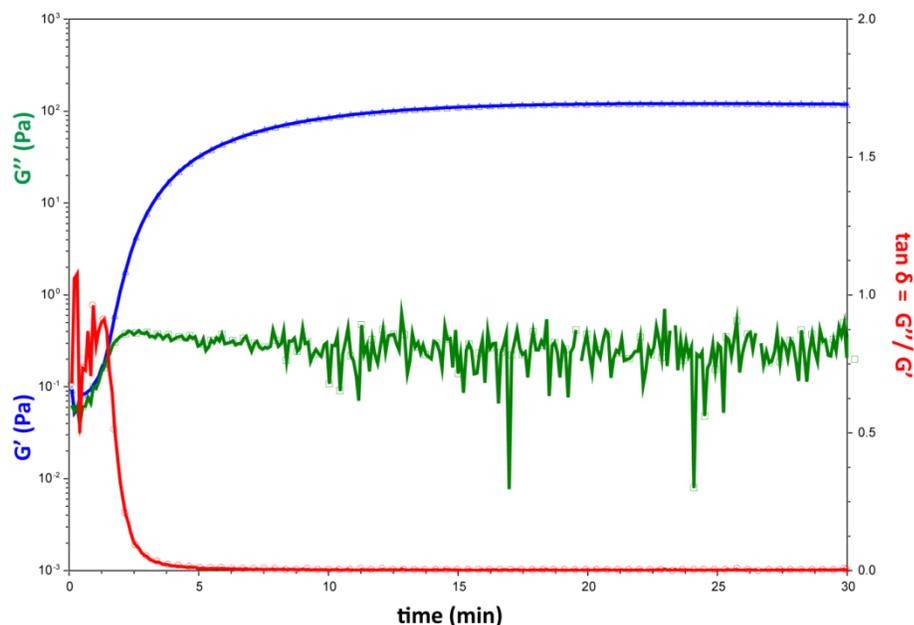
A Versatile Biosynthetic Hydrogel Platform for Engineering of Tissue Analogues

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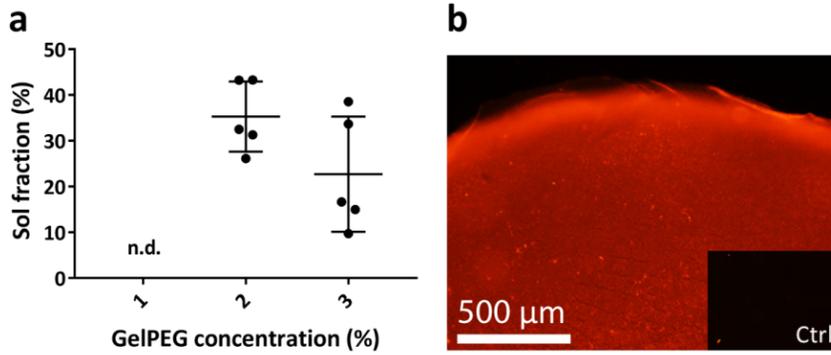
## Supporting Information

### Supplementary comment 1.

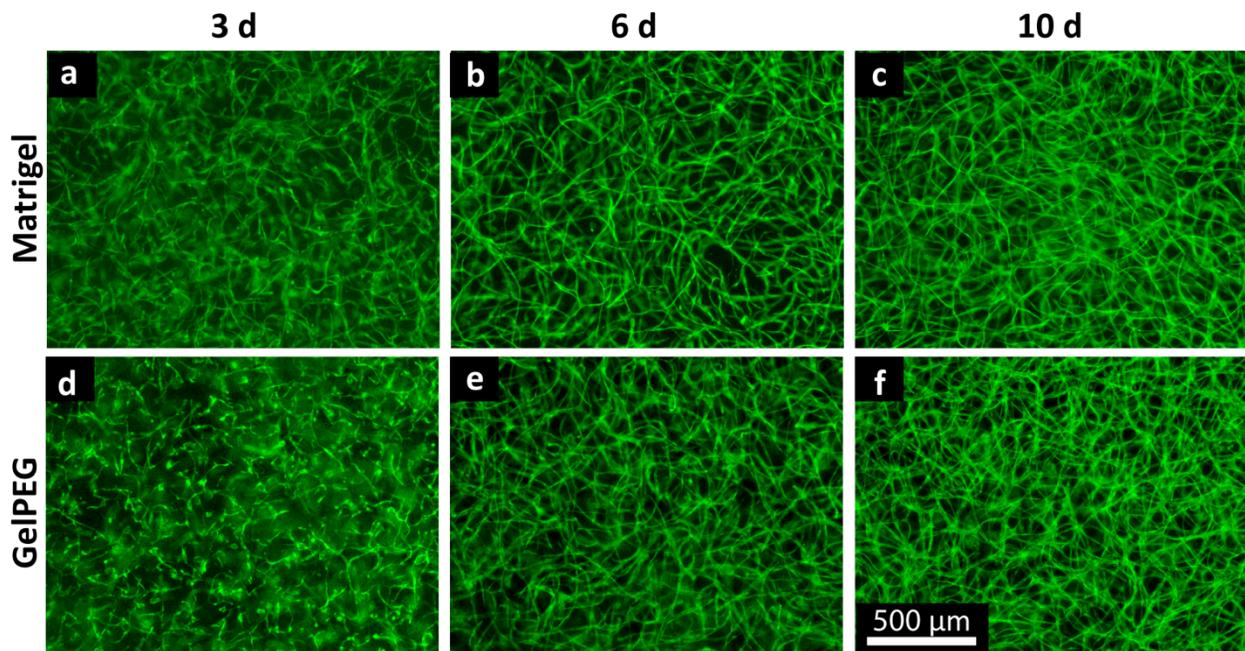
We hypothesized, that while the amino acid sequence containing a glutamine is a very specific sequence that the lysine (Lys) containing sequence is more generic. Indeed, hydrogels did not form in the absence of PEG-Gln when PEG-Lys was combined with gelatin. To exclude the formation of intramolecular crosslinks, pure gelatin was crosslinked at a concentration of 10% w/v with FXIIIa. Under present crosslinking conditions, pure gelatin did not form a hydrogel and the polymer solution remained at a low viscosity (data not shown), demonstrating the need for the substrate-specific Gln-sequence for FXIIIa mediated crosslinking.



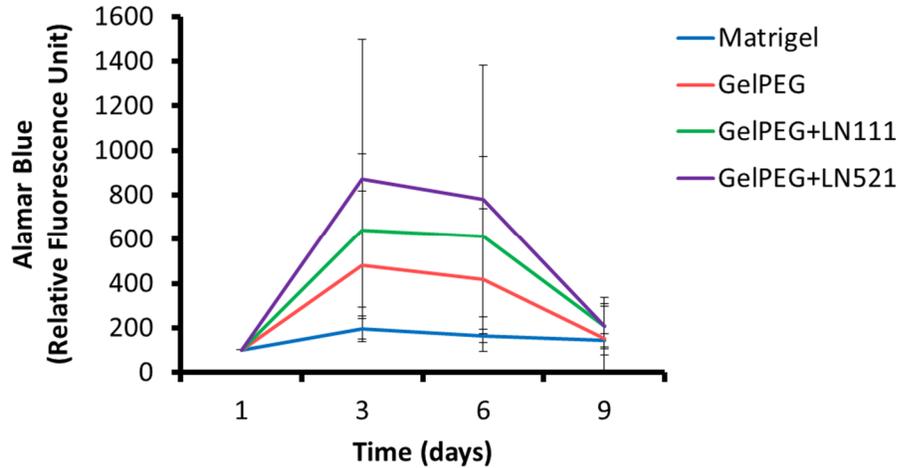
**Supplementary Figure 1.** Hydrogel formation of gelatin and PEG mediated by FXIIIa. The point of gelation occurred at about 2 minutes and the crosslinking was completed after about 15 minutes, when  $G'$  reached a plateau. Depicted data are from a representative measurement;  $n= 3$  independent experiments



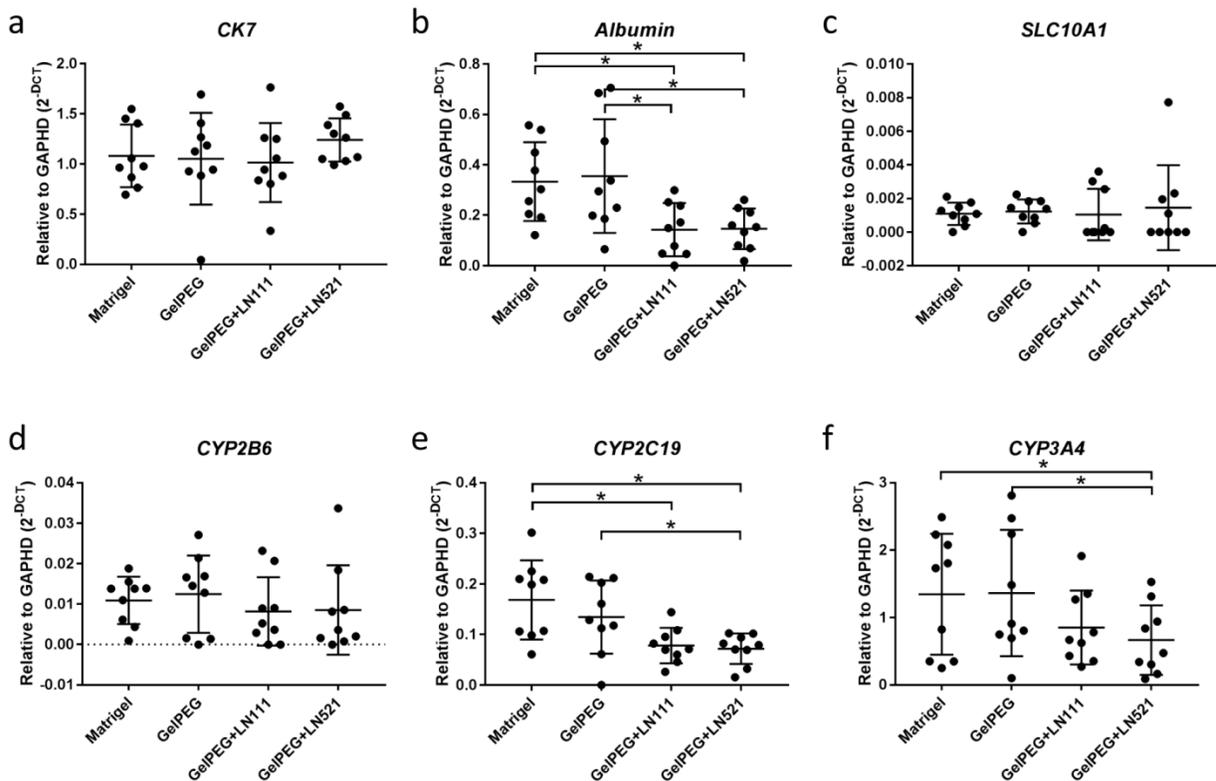
**Supplementary Figure 2.** Hydrogel sol fraction and LN binding in gelPEG hydrogel networks. **a** Sol fraction in dependence of gelPEG concentration. **b** Anti-LN-subunit  $\alpha 5$  staining of LN521-laden 3% w/v gelPEG hydrogel compared to unloaded hydrogel (insert) after 1 d swelling in TBS. Data is depicted as mean + SD,  $n=5$



**Supplementary Figure 3.** Pre-vascular network formation in GFP-ECFC and MSC co-cultures under vasculogenic culture conditions. **a-c** GFP-ECFCs in Matrigel form vascular-like networks on day 3 which are remodeled up to day 10. **d-f** GFP-ECFCs in gelPEG hydrogels form vascular-like structures with an initial temporal delay, resulting in comparable networks to Matrigel on day 6 and 10. (widefield fluorescence,  $N=3$ ,  $n=3$ )



**Supplementary Figure 4.** Metabolic activity of liver organoids over culture time in Matrigel and gelPEG-based hydrogels. Liver organoids exhibited an increase in metabolic activity from day 1 to 3, which gradually decreases over culture time, resulting in comparable activity levels as liver organoids cultured in Matrigel after 9 days culture. Data is depicted as mean  $\pm$  SD;  $N=3$ ,  $n=5$



**Supplementary Figure 5.** Gene expression levels of liver organoids that were cultured for 9 days in Matrigel and gelPEG-based hydrogels. **a** The cytokeratin *KRT7* (*CK7*) was equally expressed in all

hydrogels. **b** Albumin expression was decreased in LN-laden gelPEG hydrogels. **c** *SLC10A1*, encoding a liver-specific sodium/bile acid cotransporter, was comparably expressed in all hydrogel compositions. The cytochrome family of enzymes **d** CYP2B6 **e** CYP2C19 and **f** CYP3A4 were comparably expressed in Matrigel and gelPEG, whereas LN-laden hydrogels showed partially lower expression levels. Data is depicted as mean + SD; *N*= 3, *n*= 3

## Tables

**Supplementary table 1:** Primer sets used for qPCR analysis

Human gene	Forward primer	Reverse primer
<u>Pre-vascularized bone:</u>		
<i>PECAM1</i>	GCAGTGGTTATCATCGGAGTG	TCGTTGTTGGAGTTCAGAAGTG
<i>CDH5</i>	AAGCAGGCCAGGTATGAGAT	TGTGTA CTGGTCTGGGTGAAG
<i>CSPG4</i>	GAAGGAGGACGGACCTCAAG	GATCAGCTGCTCTCCACCATT
<i>ACTA2</i>	ATGCCATCATGCGTCTGGAT	ACGCTCAGCAGTAGTAACGA
<i>BGLAP</i>	CCTCACACTCCTCGCCCTAT	GCTTGGACACAAAGGCTGCAC
<i>SPP1</i>	GCCGAGGTGATAGTGTGGTT	GTGGGTTTCAGCACTCTGGT
<i>GAPDH</i>	CAACGGATTTGGTCGTATTGGG	TGCCATGGGTGGAATCATATTGG
<u>Liver organoids:</u>		
<i>YWHAZ</i>	ACTTTTGGTACATTGTGGCTTCAA	CCGCCAGGACAAACCAGTAT
<i>GAPDH</i>	TGCACCACCAACTGCTTAGC	GGCATGGACTGTGGTCATGAG
<i>KRT7</i>	GGACATCGAGATCGCCACCT	ACCGCCACTGCTACTGCCA
<i>CYP2C19</i>	GGGACAGAGACAACAAGCA	CCTGGACTTTAGCTGTGACC
<i>ALB</i>	GTTCGTTACACCAAGAAAGTACC	GACCACGGATAGATAGTCTTCTG
<i>SLC10A1</i>	GATATCACTGGTGGTTCTC	ATCATCCCTCCCTTGATGAC
<i>CYP3A4</i>	CACAGGCTGTTGACCATCAT	TTTTGTCCTATAAGGGCTTT
<i>CYP2B6</i>	CTACCAAGATCAAGAGTTCCTG	ATTTCAAGAAGCCAGAGAAGAG