Analogous and diverse functions of APSES-type transcription factors in the morphogenesis of the entomopathogenic fungus, *Metarhizium rileyi*

Caiyan Xin^{a#}, Jinping Zhang^{a#}, Siji Nian^a, Guangxi Wang^a, Zhongkang Wang^b, Zhangyong Song^a*, Guangwei Ren^c

a School of Basic Medical Sciences, Southwest Medical University, Luzhou 646000, People's Republic of China.

b Chongqing Engineering Research Center for Fungal Insecticide, School of LifeScience, Chongqing University, Chongqing 400030, People's Republic of China.c Institute of Qingdao Economic Crop Chinese Academy of Agricultural Sciences,

Qingdao 266101, People's Republic of China.

These authors contributed equally to this work.

Author 1: Given name: Caiyan	Family name: Xin
Email: xincy0211@126.com	
Author 2: Given name: Jinping	Family name: Zhang
Email: zhangjpwcx@163.com	
Author 3: Given name: Siji	Family name: Nian
Email: sijinian@swmu.edu.cn	
Author 4: Given name: Guangxi	Family name: Wang
Email: <u>2371966379@qq.com</u>	
Author 5: Given name: Zhongkang	Family name: Wang
Email: w-zk@163.com	
Author 6: Given name: Zhangyong	Family name: Song

Email: <u>szy83529@163.com</u>

Tel/Fax: +86-0830-3161506

Author 7: Given name: Guangwei

Family name: Ren

Email: renguangwei@caas.cn

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Annotation from NCBI Nr	Gene								Up/Down	FDR
protein database	Name	GW1_FPKM	GW2_FPKM	GW3_FPKM	GS1_FPKM	GS2_FPKM	GS3_FPKM	log2FC		
Cellulase	Cel	7.641	5.207	5.092	1.429	0.991	0.967	-2.213	Down	6.32E-05
Cytochrome P450	Cp450	0.609	0.346	0.162	2.113	1.627	1.288	2.357	Up	1.80E-6
Heat shock protein 101	H101	90.944	107.584	84.997	882.414	713.167	717.310	3.214	Up	9.46E-45
GATA transcription factor	NsdD	4.169	4.167	3.989	9.684	9.626	8.946	1.450	Up	1.94E-08
Heat shock protein 90	H90	554.026	352.991	567.606	1395.486	1391.291	1242.497	1.634	Up	2.08E-24
Cell wall beta-glucan synthesis	Cwb	1340.449	1271.608	1278.913	1121.157	1099.003	1045.579	-0.0677	Normal	0.8755
Protein-tyrosine phosphatase	Ptp	65.210	66.009	70.078	43.187	44.949	41.099	-0.454	Normal	0.120
bZIP transcription factor HapX	HapX	106.516	100.291	97.252	43.285	37.952	41.278	-1.125	Down	5.01E-10
MAP kinase kinase EMK1	Emk1	114.815	107.860	100.243	129.258	112.718	119.672	0.3439	Normal	0.181
Cytochrome c peroxidase	Сср	94.366	99.565	99.157	58.498	88.807	61.365	0.6509	Normal	0.00093
ATPase assembly factor ATP10	Atp10	89.720	83.323	80.491	60.525	63.557	62.765	-0.255	Normal	0.391
Mob1/phocein	Mob	23.610	17.309	22.918	25.562	16.770	18.760	0.6037	Normal	0.2491

Table S1 Gene expression in $\Delta MrStuA$ mutants relative to wild-type (WT) during dimorphic transition

DNA-repair protein rad2	Rad2	6.245	10.622	9.246	8.141	9.635	9.040	0.596 Normal	0.221
ChaC-like protein	Clp	56.621	45.068	56.537	52.606	51.923	22.517	0.8753 Normal	0.131

Abbreviation used: RPKM: fragments per kilobase million; FDR: false discovery rate; FC: fold change; Up/Down: up-/down-regulated. GW1, GW2, and GW3 were three samples of the WT strain during dimorphic transition. GS1, GS2, and GS3 were three samples of the $\Delta MrStuA$ mutants during dimorphic transition.

Annotation from NCBI Nr	Gene								Up/Down	FDR
protein database	Name	GW1_FPKM	GW2_FPKM	GW3_FPKM	GX1_FPKM	GX2_FPKM	GX3_FPKM	log2FC		
Cytochrome P450	Cp450	0.609	0.346	0.162	0.169	0.280	0.161	-0.824	Normal	0.371
GATA transcription factor	NsdD	4.169	4.167	3.989	10.440	12.405	10.568	1.561	Up	7.58E-11
Cellulase	Cel	7.641	5.207	5.092	0	0	0		Down	7.12E-31
Heat shock protein 101	H101	90.944	107.584	84.997	219.451	335.662	256.553	1.554	Up	3.02E-06
Heat shock protein 90	H90	554.026	352.991	567.606	1395.564	962.506	1596.451	1.452	Up	2.36E-05
Cell wall beta-glucan syntheses	Cwb	1340.449	1271.608	1278.913	3061.632	2841.773	2976.183	1.222	Up	2.99E-20
Protein-tyrosine phosphatase	Ptp	65.210	66.009	70.078	13.032	13.125	11.998	-2.366	Down	1.32E-20
MAP kinase kinase EMK1	Emk1	114.815	107.860	100.243	3061.631	2841.772	2976.182	1.222	Up	2.98E-20
Cytochrome c peroxidase	Сср	94.366	99.565	99.157	297.820	292.440	299.596	2.050	Up	1.31E-34
ATPase assembly factor ATP10	Atp10	89.720	83.323	80.491	85.463	83.348	78.939	-0.000419	Normal	0.997
Mob1/phocein	Mob	23.610	17.309	22.918	14.267	15.234	14.845	-0.372	Normal	0.1642
DNA-repair protein rad2	Rad2	6.245	10.622	9.246	15.466	16.311	15.420	0.00005	Normal	0.833
ChaC-like protein	Clp	56.621	45.068	56.537	48.013	24.020	46.171	-0.3970	Normal	0.238

Table S2 Gene expression in $\triangle MrXbp$ mutants relative to WT during dimorphic transition

bZIP transcription factor	HapX 106 516	100 201	07 252	28 235	25.508	23.601	-1.942 Down	1.64E-28
НарХ	100.510	100.291	91.232	20.235	25.508	23.001		
Abbreviation used: RPKM:	fragments per kilobase n	nillion; FDR: fa	alse discovery	y rate; FC: fold	change; Up/De	own: up-/dov	vn-regulated. GW1,	GW2, and

GW3 were three samples of the WT strain during dimorphic transition. GX1, GX2, and GX3 were three samples of the $\Delta MrXbp$ mutants during dimorphic transition.

Annotation from NCBI Nr	Gene Name	W1 FDKM	W? FDKM	W3_FPK	N1 FDKM		N2 FDKM		Up/Down	FDR
protein database		VV 1_F F K.IVI	VV 2_F F KIVI	Μ	N1_FFKW	N2_FPKM	IN 3_FF KIVI	log2FC		
Heat shock protein 101	H101	298.837	123.399	390.334	3331.313	29995.36	1220.111	2.737	Up	0.00562
Ubiquitin-conjugating	Uce	16 510	10 925	15 171	0.741	0.200	0.552	5 009	D	2 00E 10
enzyme		10.518	12.835	15.171	0.741	0.399	0.553	-5.098	Down	2.00E-10
Heat shock protein 90	H90	841.821	1072.921	982.204	3700.62	4038.17	3571.412	1.342	Up	0.0307
Mob1/phocein	Mob	268.787	282.770	252.793	88.544	75.437	92.271	-2.099	Down	0.000104
ChaC-like protein	Clp	23.874	44.283	46.679	14.739	13.585	15.763	-1.709	Down	0.000405
MAP kinase kinase EMK1	Emk1	39.946	50.166	55.233	29.705	21.825	30.360	-1.227	Down	0.002
Cell surface protein (Mas1)	Mas1	1.649	1.525	1.871	20.657	25.723	39.557	3.626	Up	1.13E-06
Sodium/calcium exchanger	Sem	0 (04	10.075	11 427	7 001	10 700	11 500	0.261	NT 1	0.600
membrane region		9.694	10.275	11.437	/.801	10.792	11.529	-0.361	Normal	0.629
DNA-repair protein rad2	Rad2	8.583	3.697	2.232	29.018	24.406	37.987	2.30	Up	2.31E-10
Cytochrome c peroxidase	Сср	193.788	133.213	165.189	145.535	93.391	89.990	-1.513	Down	0.00187
Membrane protein Tapt1	Tapt l	12.885	15.915	11.571	12.765	14.495	12.518	0.619	Normal	0.3194
GATA transcription factor	NsdD	13.459	7.622	9.299	16.071	11.646	9.490	0.312	Normal	0.650

Table S3 Gene expression in $\triangle MrStuA$ mutants relative to WT during microsclerotium development

Carbohydrate-binding Wsc	Wsc	25.114	23.851	20.4269	23.877	74.704	52.111	1.243	Up	0.00227
ATPase assembly factor	Atp10	<u>91 900</u>	125 420	<u>81 020</u>	54 520	40 112	45 221	1 500	D	0.000406
ATP10		01.090	155.420	01.929	54.550	40.112	45.551	-1.309	Dowii	0.000400

Abbreviation used: RPKM: fragments per kilobase million; FDR: false discovery rate; FC: fold change; Up/Down: up-/down-regulated. W1, W2, and W3 were three samples of the WT strain during microsclerotium development. S1, S2, and S3 were three samples of the $\Delta MrStuA$ mutants during microsclerotium development.

Annotation from NCBI Nr protein	Gene Name	W1 FDKM	W? FDKM	W3 FDKM	N1 FDKM		N3 FDVM		Up/Down	FDR
database		VV 1_FF KIVI	VV 2_FF KIVI	VV 3_FF KIVI		N2_FPKM	N 3_F F K IVI	log2FC		
Heat shock protein 101	H101	274.482	113.266	355.170	537.836	575.774	318.860	0.752	Normal	0.314
Ubiquitin-conjugating enzyme	Uce	16.518	12.835	15.171	0.741	0.399	0.553	-3.288	Down	0.00616
Heat shock protein 90	H90	841.821	1072.921	982.204	975.138	915.535	3571.412	-0.209	Normal	0.766
Cytochrome c peroxidase	Сср	158.170	109.121	132.960	302.973	255.471	282.015	0.324	Normal	0.548
Sodium/calcium exchanger	Sem	0 604	10 275	11 427	44.079	55 074	51 010	2.069	L	2 ACE 11
membrane region		9.094	10.275	11.437	44.978	55.074	51.212	2.008	Up	3.40E-11
DNA-repair protein rad2	Rad2	8.583	3.697	2.232	30.8019	30.950	30.974	2.486	Up	2.07E-14
Carbohydrate-binding Wsc	Wsc	25.114	18.687	23.168	61.716	67.843	61.721	1.541	Up	2.12E-06
Mob1/phocein	Mob	268.787	282.770	252.793	99.832	89.361	93.163	-1.693	Down	0.000143
ATPase assembly factor ATP10	Atp10	81.779	135.227	81.833	50.840	50.797	51.808	-1.1207	Down	0.00164
ChaC-like protein	Clp	23.791	44.159	46.555	12.551	9.961	10.557	-1.855	Down	0.00005
MAP kinase kinase EMK1	Emk1	39.892	50.094	55.162	56.886	49.716	44.039	-0.1085	Normal	0.7758
Cell surface protein (Mas1)	Mas1	1.649	1.525	3.355	4.483	3.519	39.557	0.9753	Normal	0.3424
GATA transcription factor	NsdD	13.459	7.622	9.299	16.172	8.558	10.645	0.5103	Normal	0.3257

Table S4 Gene expression in $\triangle MrXbp$ mutants relative to WT during microsclerotium development

Membrane protein Tapt1	Tapt1	12.067	15.719	11.571	5.147	6.397	5.439	-1.249 Down	0.00303
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Abbreviation used: RPKM: fragments per kilobase million; FDR: false discovery rate; FC: fold change; Up/Down: up-/down-regulated. W1, W2, and W3 were three samples of the WT strain during microsclerotium development. X1, X2, and X3 were three samples of the $\Delta MrXbp$ mutant during microsclerotium development.



Fig. S1 Structural and phylogenetic analysis of MrStuA and MrXbp proteins

(http://www.ebi.ac.uk/Tools/pfa/iprscan). (B) Phylogenetic analysis of MrStuA protein. The numbers at the nodes represent the results of bootstrap analyses (1000 replicates) carried out using the neighbor-joining method. The aligned sequences of StuA protein are from Metarhizium anisopliae (KFG88133.1); Metarhizium guizhouense ARSEF 977 (KID87765.1); Metarhizium majus ARSEF 297 (KID98874.1); Metarhizium brunneum ARSEF (XP 014545496.1); 3297 Metarhizium album ARSEF 1941 (KHN97002.1); Metarhizium robertsii ARSEF 23 (XP_007819177.1); Pochonia chlamydosporia 170 (XP_018139603.1); Metarhizium acridum CQMa 102 (XP_007810169.1); Ustilaginoidea virens (KDB12946.1); Hypocrella siamensis (ANH22586.1); Fusarium sp. AF-12 (RMJ13252.1); Cordyceps sp. RAO-2017 (PHH78192.1); Purpureocillium lilacinum (PWI75332.1); Hirsutella minnesotensis 3608 (KJZ71075.1); Stachybotrys chlorohalonata IBT 40285 (KFA66336.1); Neonectria ditissima (KPM35181.1). (C) Phylogenetic analysis of MrXbp protein. The numbers at the nodes represent the results of bootstrap analyses (1000 replicates) carried out using the neighbor-joining method. The aligned sequences of MrXbp protein are from *Metarhizium majus* ARSEF 297 (KID96819.1); Metarhizium robertsii ARSEF 23 (XP_007821239.2); Metarhizium guizhouense ARSEF 977 (KID92324.1); Metarhizium robertsii (EXV05007.1); Metarhizium acridum CQMa 102 (XP_007812764.1); Tolypocladium paradoxum (POR39629.1); Cordyceps fumosorosea ARSEF 2679 (XP_018703649.1); Beauveria bassiana ARSEF 2860 (XP_008593781.1); Ustilaginoidea virens (KDB17052.1); Trichoderma (OTA01006.1); Trichoderma citrinoviride (XP_024754116.1); parareesei

Trichoderma longibrachiatum ATCC 18648 (PTB80558.1); Beauveria brongniartii RCEF 3172 (OAA45796.1); Trichoderma reesei RUT C-30 (ETS05362.1); Ophiocordyceps sinensis CO18 (EQL01683.1); Claviceps purpurea 20.1 (CCE33631.1).





(A) Construction of knockout and complementation (CP) vectors. (B) PCR characterization of (B-1) $\Delta MrStuA$ mutants and (B-2) $\Delta MrXbp$ mutants, wild-type (WT), and CP strains. Open reading frame was PCR (Lanes 1-2 in B-1: 1-*MrStuA* of WT, 2- $\Delta MrStuA$ -1; Lanes 1-3 in B-2: 1-*MrXbp* of WT, 2- $\Delta MrXbp$ -1, 3- $\Delta MrXbp$ -2); The *hph* was screened (Lanes 3-4 in B-1: 3-*hph* of WT, 4- $\Delta MrStuA$ -1; Lanes 4-6 in B-2: 4-*hph* of WT, 5- $\Delta MrXbp$ -1, 6- $\Delta MrXbp$ -2). The *hph* and genomic sequence outside the flank regions was PCR (Lanes 5-8 in B-1: 5-Left Frame (LF)-WT, 6-Right Frame (RF)-WT, 7-LF- $\Delta MrStuA$ -1, 8-RF- $\Delta MrStuA$ -1; Lanes 7-12 in B-2: 7-LF-WT, 8-RF-WT, 9-LF- $\Delta MrXbp$ -1, 10-RF- $\Delta MrXbp$ -1, 11-LF- $\Delta MrXbp$ -2, 12-RF- $\Delta MrXbp$ -2). M, DNA molecular size markers (DL 5000, Takara, Beijing). The $\Delta MrStuA$ -1 mutants and $\Delta MrXbp$ -1 mutants were subjected to further

experiments. (C) qPCR analysis of *MrMid2* in the different strains. Error bars represent standard error. * P < 0.05, ** P < 0.01, denote significant differences compared with WT in AM cultures.

Fig. S3 Gene ontology (GO) annotation of differentially expressed genes (DEGs) and all genes in the (A) $\Delta MrStuA$ and (B) $\Delta MrXbp$ mutants during dimorphic transition



Fig. S4 Clusters of orthologous groups (COG) classifications in the (A) $\Delta MrStuA$



and (B) $\Delta MrXbp$ mutants during dimorphic transition





- B: Chromatin structure and dynamics C: Energy production and conversion
- D: Cell cycle control, cell division, chromosome partitioning
- E: Amino acid transport and metabolism F: Nucleotide transport and metabolism
- G: Carbohydrate transport and metabolism
- H: Coenzyme transport and metabolism
- I: Lipid transport and metabolism
- J: Translation, ribosomal structure and biogenesis
- K: Transcription
- L: Replication, recombination and repair
- M: Cell wall/membrane/envelope biogenesis
- N: Cell motility
- O: Posttranslational modification, protein turnover, chaperones
- P: Inorganic ion transport and metabolism
- Q: Secondary metabolites biosynthesis, transport and catabolism R: General function prediction only
- S: Function unknown
- T: Signal transduction mechanisms
- U: Intracellular trafficking, secretion, and vesicular transport
- V: Defense mechanisms
- W: Extracellular structures
- X: Mobilome: prophages, transposons
- Y: Nuclear structure
- Z: Cytoskeleton





Fig. S6 GO annotation of DEGs and all genes in the (A) $\triangle MrStuA$ and (B) $\triangle MrXbp$ mutants during microsclerotium development



Fig. S7 COG classifications in the (A) $\triangle MrStuA$ and (B) $\triangle MrXbp$ mutants during









- X: Mobilome: prophages, transposons
- Y: Nuclear structure
- Z: Cytoskeleton



Fig. S8 KEGG enrichment pathways of DEGs in the (A) $\Delta MrStuA$ and (B) $\Delta MrXbp$

mutants during microsclerotium development



Fig. S9 RT-qPCR of WT, $\triangle MrStuA$ and $\triangle MrXbp$ mutants during (A) dimorphic

transition and (B) microsclerotium development





Fig. S10 GO annotation of DGEs and all genes in the two morphogenesis in the (A) $\Delta MrStuA$ and (B) $\Delta MrXbp$ mutants



Fig. S11 COG classifications of consensus sequence in the two morphogenesis in the

(A) $\triangle MrStuA$ and (B) $\triangle MrXbp$ mutants



Fig. S12 KEGG enrichment pathways of DGEs in the two morphogenesis in the (A) $\triangle MrStuA$ and (B) $\triangle MrXbp$ mutants



Fig. S13 Venn diagram showing the number of shared differentially expressed genes



in the three fungal transcription factors

The distribution of shared upregulated and downregulated genes in wild-type (WT), $\Delta MrXbp$, $\Delta MrStuA$, and $\Delta MrAp1$ mutants during microsclerotium development. CK1, CK2, and CK3 were three replicates of the WT strain. *MrXbp 1*, *MrXbp2*, and *MrXbp3* were three replicates of the $\Delta MrXbp$ mutants. *MrStuA*, *MrStuA2*, and *MrStuA3* were three replicates of the $\Delta MrStuA$ mutants. *MrAp1-1*, *MrAp1-2*, and *MrAp1-3* were three replicates of the $\Delta MrAp1$ mutants.