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Reporting Summary

Ctatiation

Nature Research wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Research policies, seeAuthors & Referees and theEditorial Policy Checklist.

Statistics		
For all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.		
n/a Confirmed		
The exact samp	le size (n) for each experimental group/condition, given as a discrete number and unit of measurement	
A statement on	whether measurements were taken from distinct samples or whether the same sample was measured repeatedly	
	est(s) used AND whether they are one- or two-sided ts should be described solely by name; describe more complex techniques in the Methods section.	
A description of	A description of all covariates tested	
A description of	f any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons	
A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)		
For null hypothesis testing, the test statistic (e.g. <i>F</i> , <i>t</i> , <i>r</i>) with confidence intervals, effect sizes, degrees of freedom and <i>P</i> value noted Give <i>P</i> values as exact values whenever suitable.		
For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings		
For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes		
Estimates of effect sizes (e.g. Cohen's <i>d</i> , Pearson's <i>r</i>), indicating how they were calculated		
1	Our web collection on <u>statistics for biologists</u> contains articles on many of the points above.	
Software and co	ode	
	availability of computer code	
	Metamorph Software (Universal Imaging) version 7.7.9.0 was used to collect data.	
	Matlab version 2016a, Metamorph version 7.7.9.0, ImageJ and Fiji (NIH) version 2.0.0-rc-69/1.52, Microsoft Excel version 16.16.19, GraphPad Prism 8 and R Version 3.3.2 were used to analyze the data.	
For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors/reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Research guidelines for submitting code & software for further information.		
Data		
Policy information about availability of data All manuscripts must include a data availability statement. This statement should provide the following information, where applicable: - Accession codes, unique identifiers, or web links for publicly available datasets - A list of figures that have associated raw data - A description of any restrictions on data availability		
All relevant data supporting the finding of this study are available from the corresponding author upon request.		
Field-specif	ic reporting	
Please select the one be	low that is the best fit for your research. If you are not sure, read the appropriate sections before making your selection.	
Life sciences Behavioural & social sciences Ecological, evolutionary & environmental sciences		

Life sciences study design

All studies must dis	close on these points even when the disclosure is negative.
Sample size	Experiments were repeated at least 3 times, n numbers were 10 or more, with very few variability (see s.d.) and the adequate statistic tests were performed. No predetermination of sample size were performed. The sample sizes (oocytes number for each experiment) were chosen based on the number of mature follicles of mice.
Data exclusions	No data were excluded from the analysis.
Replication	Experiments were repeated at least 3 times, n numbers were 10 or more, with very few variability (see s.d). All attempts at replication were successful.
Randomization	When oocytes were collected from different mice from similar strains, they were allocated into experimental groups randomly before microinjection.
Blinding	Investigators were blinded for data analysis.
We require informatis system or method list Materials & ex n/a Involved in th	cell lines cell lines pogy d other organisms earch participants ChIP-seq Flow cytometry MRI-based neuroimaging
Antibodies	d
Antibodies used	rabbit anti-Phospho-MLC2 (Ser19) (Cell Signaling # 3671; 1:200), Alexa-594-labeled anti-rabbit (Invitrogen # A-21207; 1:400).
Validation	The rabbit anti-Phospho-MLC2 (Ser19) antibody was already described and used in mouse oocytes (Dumont et al., 2007; Schuh et al., 2008). In addition, the staining disappears after the inhibition of the activation of the protein (the antibody recognizes the active form of the protein) as shown in the manuscript.
Animals and	other organisms
Policy information	about studies involving animals; ARRIVE guidelines recommended for reporting animal research
Laboratory anima	11-week-old OFI, fmn2-/- or fmn2+/- (Leader et al., 2002) female mice were used, housed in the animal facility of the CIRB.
Wild animals	No wild animais were used for this study.
Field-collected sa	Mo field-collected samples were used for this study.
Ethics oversight	All experimental procedures used for the project have been approved by the ministry of agriculture to be conducted in our animal facility (authorization N°75-1170). The use of all the genetically modified organisms described in this project has been granted by the DGRI (Direction Générale de la Recherche et de l'Innovation: Agrément OGM; DUO-1783).

Note that full information on the approval of the study protocol must also be provided in the manuscript.