

Synthesis, biological evaluation and molecular modeling of a novel series of fused 1,2,3-triazoles as potential anti-coronavirus agents

SUPPORTING INFORMATION

1. General experimental methods

NMR spectra were acquired at room temperature on commercial instruments (Bruker Avance 300 MHz, Bruker AMX 400 MHz or Bruker Avance II+ 600 MHz) and chemical shifts (δ) are reported in parts per million (ppm) referenced to tetramethylsilane (0.00 ppm for ^1H), or the internal (NMR) solvent signal (77.16 ppm for ^{13}C). High resolution mass spectra were acquired on a quadrupole orthogonal acceleration time-of-flight mass spectrometer (Synapt G2 HDMS, Waters, Milford, MA). For column chromatography 70-230 mesh silica 60 (E. M. Merck) was used as the stationary phase. Chemicals received from commercial sources were used without further purification. Reaction solvents (toluene) were used as received from commercial sources.

1.1. General procedure for the preparation of substituted 1,2,3-triazoles

To an oven-dried screw-capped reaction tube equipped with a magnetic stirring bar the ketone (1 eq), amine (2.8 eq), 4-nitrophenyl azide (2 eq), acetic acid (10 mol%) and 4 Å molecular sieves (50 mg) were added. The reaction mixture was dissolved in toluene (0.3M) and stirred at 100 °C for 18 hours. The crude reaction mixture was then directly purified by column chromatography (silica gel) at first with dichloromethane (DCM) as eluent to remove all 4-nitroaniline formed during the reaction followed by using a mixture of heptane and ethyl acetate as eluent to afford the corresponding 1,2,3-triazoles as yellow oil.

Ethyl-1,7-dibenzyl-5-phenyl-4,5,6,7-tetrahydro-1H-[1,2,3]triazolo[4,5-c]pyridine-7-carboxylate (14a): Yellow oil (80%). ^1H NMR (300MHz, CDCl_3): δ 7.35-7.25 (m, 8H), 7.09-7.00 (m, 6H), 6.93 (t, $J = 7.3$ Hz, 1H), 5.14 (d, $J = 16.1$ Hz, 1H), 4.73 (d, $J = 16.1$ Hz, 1H), 4.52 (d, $J = 14.2$ Hz, 1H), 4.40 (d, $J = 14.2$ Hz, 1H), 4.05 (m, 1H), 3.78 (d, $J = 12.9$ Hz, 1H), 3.65 (m, 2H), 3.37 (q, $J = 13.4$ Hz, 2H), 1.04 (t, $J = 7.1$ Hz, 3H). ^{13}C NMR (75MHz, CDCl_3): δ 171.30, 149.86, 142.41, 135.86, 135.50, 132.04, 130.38, 129.34, 128.63, 128.59, 127.81, 127.51, 127.02, 120.21, 116.00, 61.92, 58.37, 52.08, 50.64, 45.61, 41.39, 13.87. HRMS (ES+): m/z calcd for $\text{C}_{28}\text{H}_{28}\text{N}_4\text{O}_2$ $[\text{M}+\text{H}]^+$: 453.2284, found 453.2284.

Ethyl-7-benzyl-5-phenyl-1-(3,4,5-trimethoxybenzyl)-4,5,6,7-tetrahydro-1H-[1,2,3]triazolo[4,5-c]pyridine-7-carboxylate (14b): Yellow oil (85%). ^1H NMR (400 MHz, CDCl_3) δ 7.30 – 7.25(m, 6H), 7.02 – 6.99 (m, 3H), 6.91 (t, $J = 7.3$ Hz, 1H), 6.37 (s, 2H), 5.03 (d, $J = 15.7$ Hz, 1H), 4.70 (d, $J = 15.7$ Hz, 1H), 4.46 (d, $J = 14.2$ Hz, 1H), 4.37

(d, $J = 14.1$ Hz, 1H), 4.10 (dq, $J = 10.7, 7.1$ Hz, 1H), 3.81 – 3.75 (m, 11H), 3.58 (d, $J = 12.9$ Hz, 1H), 3.43 (d, $J = 13.5$ Hz, 1H), 3.33 (d, $J = 13.5$ Hz, 1H), 1.07 (t, $J = 7.2$ Hz, 3H). ^{13}C NMR (75MHz, CDCl_3): δ 171.43, 153.35, 149.80, 142.47, 137.68, 135.79, 132.01, 130.84, 130.31, 129.33, 128.64, 127.54, 120.25, 116.01, 104.74, 62.03, 60.80, 58.30, 56.24, 56.19, 52.40, 50.67, 45.61, 41.33, 13.94. HRMS (ES+) m/z calcd for $\text{C}_{31}\text{H}_{34}\text{N}_4\text{O}_5$ $[\text{M}+\text{H}]^+$: 543.2601, found 543.2600.

Ethyl-7-benzyl-1-(4-methoxybenzyl)-5-phenyl-4,5,6,7-tetrahydro-1H-

[1,2,3]triazolo[4,5-c]pyridine-7-carboxylate (14c): Yellow oil (65%). ^1H NMR (300MHz, CDCl_3): δ 7.33-7.23 (m, 5H), 7.05-6.97 (m, 6H), 6.90 (t, $J = 7.2$ Hz, 1H), 6.84-6.78 (m, 2H), 5.03 (d, $J = 15.6$ Hz, 1H), 4.75 (d, $J = 15.6$ Hz, 1H), 4.46 (d, $J = 14.2$ Hz, 1H), 4.37 (d, $J = 14.2$ Hz, 1H), 4.09 (dq, $J = 10.8$ Hz, 7.1 Hz, 1H), 3.84-3.73 (m, 5H), 3.56 (d, $J = 12.9$ Hz, 1H), 3.41 (d, $J = 13.4$ Hz, 1H), 3.27 (d, $J = 13.4$ Hz, 1H), 1.08 (t, $J = 7.1$ Hz, 3H). ^{13}C NMR (75MHz, CDCl_3): δ 171.35, 159.18, 149.82, 142.36, 135.77, 131.84, 130.32, 129.30, 128.61, 128.56, 127.47, 127.42, 120.15, 115.95, 113.94, 62.00, 58.32, 55.27, 51.76, 50.58, 45.56, 41.36, 13.94. HRMS (ES+) m/z calcd for $\text{C}_{29}\text{H}_{30}\text{N}_4\text{O}_3$ $[\text{M}+\text{H}]^+$: 483.2390, found 483.2386.

Ethyl-7-benzyl-1-(4-fluorobenzyl)-5-phenyl-4,5,6,7-tetrahydro-1H-

[1,2,3]triazolo[4,5-c]pyridine-7-carboxylate (14d): Yellow oil (80%). ^1H NMR (300 MHz, CDCl_3): δ 7.33-7.25 (m, 5H), 7.08-6.91 (m, 10H), 4.95 (d, $J = 15.8$ Hz, 1H), 4.63 (d, $J = 15.8$ Hz, 1H), 4.50 (d, $J = 14.2$ Hz, 1H), 4.36 (d, $J = 14.2$ Hz, 1H), 4.09 (dq, $J = 10.8$ Hz, 7.1Hz, 1H), 3.81-3.69 (m, 2H), 3.63 (d, $J = 12.9$ Hz, 1H), 3.39 (q, $J = 13.2$ Hz, 2H), 1.07 (t, $J = 7.1$ Hz, 3H). ^{13}C NMR (75 MHz, CDCl_3): δ 171.29, 163.93, 160.66, 149.79, 142.39, 135.85, 131.99, 131.22, 131.17, 130.33, 129.34, 129.08, 128.97, 128.69, 127.56, 120.27, 116.00, 115.60, 115.32, 62.02, 58.41, 51.38, 50.83, 45.57, 41.49, 13.93. HRMS (ES+) m/z calcd for $\text{C}_{28}\text{H}_{27}\text{N}_4\text{O}_2\text{F}_1$ $[\text{M}+\text{H}]^+$: 471.2190, found 471.2190.

Ethyl-7-benzyl-1-(3-fluorobenzyl)-5-phenyl-4,5,6,7-tetrahydro-1H-

[1,2,3]triazolo[4,5-c]pyridine-7-carboxylate (14e): Yellow oil (58%) ^1H NMR (600MHz, CDCl_3): δ 7.34 - 7.21 (m, 6H), 7.04-6.96 (m, 4H), 6.95 - 6.88 (m, 2H), 6.82 (d, $J = 7.9$ Hz, 1H), 6.77 - 6.72 (m, 1H), 4.99 (d, $J = 16.2$ Hz, 1H), 4.59 (d, $J = 16.2$ Hz, 1H), 4.52 (d, $J = 14.2$ Hz, 1H), 4.37 (d, $J = 14.2$ Hz, 1H), 4.07 (dq, $J = 10.8, 7.2$ Hz, 1H), 3.75 - 3.63 (m, 3H), 3.37 (s, 2H), 1.05 (t, $J = 7.1$ Hz, 3H). ^{13}C NMR (151 MHz, CDCl_3) δ 171.23, 163.68, 162.04, 149.81, 142.42, 138.06, 138.01, 135.89, 132.09, 130.35, 130.12, 130.07, 129.36, 128.71, 127.59, 122.67, 122.65, 120.30, 116.03, 114.83, 114.69, 114.29, 114.14, 62.01, 58.45, 51.40, 51.39, 50.88, 45.60, 41.53, 13.88. HRMS (ES+) m/z calcd for $\text{C}_{28}\text{H}_{27}\text{N}_4\text{O}_2\text{F}_1$ $[\text{M}+\text{H}]^+$: 471.2190, found 471.2187.

Ethyl-7-benzyl-1-(3-chlorobenzyl)-5-phenyl-4,5,6,7-tetrahydro-1H-

[1,2,3]triazolo[4,5-c]pyridine-7-carboxylate (14f): Yellow oil (80%) ^1H NMR (600 MHz, CDCl_3): δ 7.34 - 7.26 (m, 5zH), 7.22 - 7.18 (m, 2H), 7.05 - 6.98 (m, 5H), 6.95 - 6.90 (m, 2H), 4.91 (d, $J = 16.2$ Hz, 1H), 4.57 - 4.47 (m, 2H), 4.37 (d, $J = 14.2$ Hz, 1H), 4.08 (dq, $J = 10.8, 7.1$ Hz, 1H), 3.74 - 3.64 (m, 3H), 3.39 (q, $J = 13.2$ Hz, 2H), 1.05 (t, $J = 7.1$ Hz,

1H). ¹³C NMR (151 MHz, CDCl₃) δ 171.24, 149.81, 142.40, 137.48, 135.97, 134.54, 132.09, 130.39, 129.80, 129.38, 128.74, 128.01, 127.61, 127.26, 125.31, 120.32, 116.03, 62.06, 58.51, 51.27, 50.95, 45.61, 41.53, 13.90. HRMS (ES+) m/z calcd for C₂₈H₂₇N₄O₂Cl₁ [M+H]⁺: 487.1895, found 487.1893.

Ethyl-7-benzyl-1-(4-chlorobenzyl)-5-phenyl-4,5,6,7-tetrahydro-1H-

[1,2,3]triazolo[4,5-c]pyridine-7-carboxylate (14g): Yellow oil (67%). ¹H NMR (400 MHz, CDCl₃): δ 7.36 - 7.22 (m, 8H), 7.05 - 6.97 (m, 6H), 6.92 (t, *J* = 7.2 Hz, 1H), 4.94 (d, *J* = 16.1 Hz, 1H), 4.60 (d, *J* = 16.1 Hz, 1H), 4.51 (d, *J* = 14.2 Hz, 1H), 4.36 (d, *J* = 14.2 Hz, 1H), 4.08 (dq, *J* = 10.8, 7.1 Hz, 1H), 3.78 - 3.68 (m, 2H), 3.64 (d, *J* = 12.9 Hz, 1H), 3.40 - 3.33 (m, 2H), 1.07 (t, *J* = 7.1 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 171.27, 149.81, 142.43, 135.87, 133.99, 133.75, 132.03, 130.35, 129.36, 128.71, 128.63, 128.60, 127.60, 120.31, 116.03, 62.05, 58.44, 51.38, 50.89, 45.61, 41.56, 13.93. HRMS (ES+) m/z calcd for C₂₈H₂₇N₄O₂Cl₁ [M+H]⁺: 487.1895, found 487.1888.

Ethyl-7-benzyl-1-(2-chlorobenzyl)-5-phenyl-4,5,6,7-tetrahydro-1H-

[1,2,3]triazolo[4,5-c]pyridine-7-carboxylate (14h): Yellow oil (48%). ¹H NMR (300 MHz, CDCl₃): δ 7.35 - 7.25 (m, 8H), 7.20 - 7.15 (m, 2H), 7.07 - 7.03 (m, 2H), 7.00 - 6.97 (m, 2H), 6.92 (d, *J* = 7.3 Hz, 1H), 6.74 - 6.71 (m, 1H), 5.30 (d, *J* = 17.2 Hz, 1H), 4.80 (d, *J* = 17.2 Hz, 1H), 4.52 (d, *J* = 14.2 Hz, 1H), 4.41 (d, *J* = 14.2 Hz, 1H), 3.98 (dq, *J* = 10.8, 7.1 Hz, 1H), 3.84 - 3.63 (m, 2H), 3.61 (d, *J* = 12.9 Hz, 1H), 3.36 (d, *J* = 13.3 Hz, 1H), 3.27 (d, *J* = 13.3 Hz, 1H), 1.03 (t, *J* = 7.1 Hz, 3H). ¹³C NMR (151 MHz, CDCl₃) δ 170.92, 149.83, 142.60, 135.53, 133.68, 130.20, 129.35, 129.22, 128.84, 128.76, 128.10, 127.67, 127.03, 120.29, 116.04, 61.99, 58.48, 50.83, 49.74, 45.53, 41.48, 13.84. HRMS (ES+) m/z calcd for C₂₈H₂₇N₄O₂Cl₁ [M+H]⁺: 487.1895, found 487.1889.

Ethyl-7-benzyl-1-(3,4-dichlorobenzyl)-5-phenyl-4,5,6,7-tetrahydro-1H-

[1,2,3]triazolo[4,5-c]pyridine-7-carboxylate (14i): Yellow oil (30%). ¹H NMR (300 MHz, CDCl₃) δ 7.37 - 7.24 (m, 7H), 7.13 (d, *J* = 2.1 Hz, 1H), 7.06 - 6.88 (m, 6H), 4.76 (d, *J* = 16.1 Hz, 1H), 4.55 (d, *J* = 14.2 Hz, 1H), 4.44 (d, *J* = 16.1 Hz, 1H), 4.36 (d, *J* = 14.2 Hz, 1H), 4.14 (dq, *J* = 10.8, 7.1 Hz, 1H), 3.80 (m, 2H), 3.67 (d, *J* = 12.9 Hz, 1H), 3.46 (d, *J* = 13.3 Hz, 1H), 3.37 (d, *J* = 13.3 Hz, 1H), 1.11 (t, *J* = 7.1 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 171.24, 149.77, 142.42, 135.98, 135.63, 132.70, 132.08, 132.01, 130.43, 130.34, 129.40, 129.33, 128.82, 127.68, 126.75, 120.39, 116.06, 62.17, 58.54, 51.18, 50.79, 45.61, 41.68, 13.96. HRMS (ES+) m/z calcd for C₂₈H₂₆N₄O₂Cl₂ [M+H]⁺: 521.1505, found 521.1483.

Ethyl-7-benzyl-1-(2-chloro-6-fluorobenzyl)-5-phenyl-4,5,6,7-tetrahydro-1H-

[1,2,3]triazolo[4,5-c]pyridine-7-carboxylate (14j): Yellow oil (73%). ¹H NMR (300 MHz, CDCl₃): δ 7.35 - 7.21 (m, 8H), 7.11 - 6.98 (m, 5H), 6.94 - 6.85 (m, 1H), 5.46 (dd, *J* = 14.0, 1.6 Hz, 1H), 4.79 (dd, *J* = 14.0, 1.6 Hz, 1H), 4.37 (s, 2H), 4.29 (dq, *J* = 11.5, 7.2 Hz, 2H), 3.89 (d, *J* = 12.9 Hz, 1H), 3.63 (d, *J* = 13.5 Hz, 1H), 3.55 - 3.46 (m, 2H), 1.30 (t, *J* = 7.1 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 171.49, 163.12, 160.62, 149.89, 142.94,

136.46, 136.41, 135.38, 131.59, 130.71, 130.62, 130.16, 129.27, 128.81, 127.60, 125.52, 125.49, 120.14, 116.04, 114.40, 114.18, 62.32, 58.25, 50.63, 45.61, 45.49, 45.46, 41.01, 14.15. HRMS (ES+) m/z calcd for $C_{28}H_{26}N_4O_2Cl_1F_1$ $[M+H]^+$: 505.1800, found 505.1784.

Ethyl-7-benzyl-1-(furan-2-ylmethyl)-5-phenyl-4,5,6,7-tetrahydro-1H-

[1,2,3]triazolo[4,5-c]pyridine-7-carboxylate (14k): Yellow oil (55%). 1H NMR (300 MHz, $CDCl_3$): δ 7.33-7.25 (m, 6H), 7.0j-6.97 (m, 4H), 6.90 (t, $J = 7.3$ Hz, 1H), 6.33-6.30 (m, 2H), 5.00 (d, $J = 16$ Hz, 1H), 4.86 (d, $J = 16$ Hz, 1H), 4.42 (d, $J = 14.2$ Hz, 1H), 4.34 (d, $J = 14.2$ Hz, 1H), 4.23 (dq, $J = 10.8, 7.1$ Hz, 1H), 4.09 (m, dq, $J = 10.8, 7.1$ Hz, 1H), 3.80 (d, $J = 13$ Hz, 1H), 3.58 (d, $J = 13$ Hz, 1H), 3.50 (d, $J = 13.5$ Hz, 1H), 3.41 (d, $J = 13.5$ Hz, 1H), 1.20 (t, $J = 7.1$ Hz, 3H). ^{13}C NMR (75 MHz, $CDCl_3$): δ 171.40, 149.81, 148.18, 142.60, 142.34, 135.65, 131.64, 130.21, 129.29, 128.69, 127.52, 120.21, 116.03, 110.73, 109.66, 62.23, 58.27, 50.67, 45.77, 45.52, 41.41, 13.96. HRMS (ES+): m/z calcd for $C_{26}H_{26}N_4O_3$ $[M+H]^+$: 443.2077, found 443.2080.

Ethyl-7-benzyl-1-(2-methoxyphenethyl)-5-phenyl-4,5,6,7-tetrahydro-1H-

[1,2,3]triazolo[4,5-c]pyridine-7-carboxylate (14l): Yellow oil (56%). 1H NMR (300MHz, $CDCl_3$): δ 7.31-7.23 (m, 5H), 7.22-7.19 (m, 3H), 7.09 (dd, $J = 7.4, 1.7$ Hz, 1H), 7.01-6.96 (m, 3H), 6.90-6.87 (m, 2H), 6.83 (d, $J = 8.1$ Hz, 1H), 4.39 (d, $J = 14.2$ Hz, 1H), 4.34 (d, $J = 14.2$ Hz, 1H), 4.27 -4.11 (m, 3H), 4.09-4.01 (m, 1H), 3.87-3.83 (m, 1H), 3.78 (s, 3H), 3.53 (d, $J = 13.5$ Hz, 1H), 3.45 (d, $J = 13.5$ Hz, 1H), 3.32-3.20 (m, 3H), 1.20 (t, $J = 7.1$ Hz, 3H). ^{13}C NMR (75MHz, $CDCl_3$): 171.54, 157.61, 149.90, 142.02, 135.53, 131.66, 130.63, 130.15, 129.26, 128.50, 128.24, 127.39, 125.77, 120.68, 120.09, 116.01, 110.21, 62.03, 58.40, 55.15, 50.30, 48.91, 45.53, 40.74, 31.56, 14.05. HRMS (ES+) m/z calcd for $C_{30}H_{32}N_4O_3$ $[M+H]^+$: 497.2547, found 497.2540.

Ethyl-7-benzyl-1-(4-methoxyphenethyl)-5-phenyl-4,5,6,7-tetrahydro-1H-

[1,2,3]triazolo[4,5-c]pyridine-7-carboxylate (14m): Yellow oil (85%). 1H NMR (300 MHz, $CDCl_3$) δ 7.32 - 7.29 (m, 2H), 7.23 - 7.20 (m, 3H), 7.05 - 6.99 (m, 4H), 6.96 - 6.94 (m, 2H), 6.92 - 6.89 (m, 1H), 6.83 - 6.79 (m, 2H), 4.47 (d, $J = 14.2$ Hz, 1H), 4.35 (d, $J = 14.2$ Hz, 1H), 4.27 (dq, $J = 10.8, 7.2$ Hz, 1H), 4.17 (dq, $J = 10.8, 7.2$ Hz, 1H), 3.78 (s, 3H), 3.77 - 3.63 (m, 4H), 3.40 (s, 2H), 3.18 (ddd, $J = 13.7, 11.2, 6.2$ Hz, 1H), 3.09 (ddd, $J = 13.7, 11.2, 6.2$ Hz, 1H), 1.25 (t, $J = 7.1$ Hz, 3H). ^{13}C NMR (75 MHz, $CDCl_3$) δ 171.48, 158.37, 149.86, 141.95, 135.83, 131.50, 130.19, 129.72, 129.57, 129.34, 128.65, 127.50, 120.19, 115.99, 113.99, 62.18, 58.51, 55.25, 50.90, 50.63, 45.55, 41.23, 34.59, 14.17. HRMS (ES+) m/z calcd for $C_{30}H_{32}N_4O_3$ $[M+H]^+$: 497.2547, found 497.2538.

Ethyl-1-(2-(1H-indol-2-yl)ethyl)-7-benzyl-5-phenyl-4,5,6,7-tetrahydro-1H-

[1,2,3]triazolo[4,5-c]pyridine-7-carboxylate (14n): Yellow oil (80%). 1H NMR (300 MHz, $CDCl_3$): δ 8.02 (s, 1H), 7.54 (d, $J = 8$ Hz, 1H), 7.38-7.26 (m, 3H), 7.22-7.1 (m, 4H), 7.14-7.09 (m, 1H), 7.03 (d, $J = 8$ Hz, 2H), 6.95-6.88 (m, 4H), 4.48 (d, $J = 14.1$ Hz, 1H), 4.36 (d, $J = 14.1$ Hz, 1H), 4.22 (dq, $J = 10.8$ Hz, 7.1 Hz, 1H), 4.10 (dq, $J = 10.8$ Hz, 7.1 Hz, 1H), 3.98-3.80 (m, 2H), 3.73 (d, $J = 12.8$ Hz, 1H), 3.62 (d, $J = 12.8$ Hz, 1H), 3.41-

3.34 (m, 4H), 1.20 (t, $J = 7.1\text{Hz}$, 3H). ^{13}C NMR (75 MHz, CDCl_3): δ 171.52, 149.90, 141.99, 136.15, 135.84, 131.60, 130.19, 129.33, 128.60, 127.44, 127.18, 122.24, 121.77, 120.19, 119.54, 118.74, 116.01, 112.28, 111.13, 62.18, 58.54, 50.87, 49.70, 45.58, 41.14, 25.53, 14.12. HRMS (ES+) m/z calcd for $\text{C}_{31}\text{H}_{31}\text{N}_5\text{O}_2$ $[\text{M}+\text{H}]^+$: 506.2550, found 506.2543.

Ethyl-7-benzyl-1-(5-hydroxypentyl)-5-phenyl-4,5,6,7-tetrahydro-1H-

[1,2,3]triazolo[4,5-c]pyridine-7-carboxylate (14o): Yellow oil (57%) ^1H NMR (400MHz, CDCl_3): δ 7.33-7.28 (m, 2H), 7.27-7.22 (m, 3H), 7.02 (d, $J = 8.1\text{ Hz}$, 2H), 6.99-6.95 (m, 2H), 6.91 (t, $J = 7.2\text{ Hz}$, 1H), 4.49 (d, $J = 14.1\text{ Hz}$, 1H), 4.35-4.16 (m, 3H), 3.73-3.66 (m, 2H), 3.62 (t, $J = 6.4\text{ Hz}$, 2H), 3.55 (d, $J = 13.4\text{ Hz}$, 1H), 3.51-3.30 (m, 3H), 1.98-1.86 (m, 1H), 1.84-1.75 (m, 1H), 1.57-1.48 (m, 2H), 1.32-1.23 (m, 5H). ^{13}C NMR (100MHz, CDCl_3): δ 171.49, 149.85, 141.82, 136.04, 131.34, 130.28, 129.35, 128.65, 127.42, 120.17, 115.94, 62.46, 62.15, 58.70, 51.04, 49.33, 45.52, 41.28, 32.07, 28.88, 23.15, 14.17. HRMS (ES+) m/z calcd for $\text{C}_{26}\text{H}_{32}\text{N}_4\text{O}_3$ $[\text{M}+\text{H}]^+$: 449.2547, found 449.2546.

Ethyl-7-benzyl-1-octyl-5-phenyl-4,5,6,7-tetrahydro-1H-[1,2,3]triazolo[4,5-

c]pyridine-7-carboxylate (14p): Yellow oil (51%). ^1H NMR (300 MHz, CDCl_3): δ 7.34-7.22 (m, 5H), 7.06-6.94 (m, 4H), 6.91 (t, $J = 7.3\text{Hz}$, 1H), 4.49 (d, $J = 12\text{Hz}$, 1H), 4.36-4.15 (m, 3H), 3.68 (br s, 2H), 3.59 (d, $J = 9\text{Hz}$, 1H), 3.50-3.28 (m, 3H), 1.95-1.63 (m, 2H), 1.30-1.17 (m, 13H), 0.88 (t, $J = 6.7\text{Hz}$, 3H). ^{13}C NMR (75 MHz, CDCl_3): δ 171.51, 149.87, 141.77, 136.03, 131.28, 130.30, 129.35, 128.63, 127.41, 120.13, 115.92, 62.12, 58.71, 50.94, 49.47, 45.52, 41.23, 31.75, 29.23, 29.10, 29.05, 26.93, 22.62, 14.16, 14.10. HRMS (ES+): m/z calcd for $\text{C}_{29}\text{H}_{38}\text{N}_4\text{O}_2$ $[\text{M}+\text{H}]^+$: 475.3067, found 475.3068.

Ethyl-1-(3,4-dichlorobenzyl)-7-(4-fluorobenzyl)-5-phenyl-4,5,6,7-tetrahydro-1H-

[1,2,3]triazolo[4,5-c]pyridine-7-carboxylate (14q): Yellow oil (50%). ^1H NMR (300 MHz, CDCl_3): δ 7.39 - 7.30 (m, 3H), 7.15 (d, $J = 2.1\text{ Hz}$, 1H), 7.04 - 6.91 (m, 8H), 4.94 (d, $J = 16.0\text{ Hz}$, 1H), 4.59 - 4.52 (m, 2H), 4.36 (d, $J = 14.2\text{ Hz}$, 1H), 4.13 (dq, $J = 10.8, 7.1\text{ Hz}$, 1H), 3.78 (dq, $J = 10.8, 7.1\text{ Hz}$, 1H), 3.69 (s, 2H), 3.36 (s, 2H), 1.10 (t, $J = 7.1\text{ Hz}$, 3H). ^{13}C NMR (151 MHz, CDCl_3) δ 171.18, 163.12, 161.48, 149.72, 142.53, 135.43, 132.84, 132.21, 131.97, 131.91, 131.63, 131.61, 130.53, 129.44, 129.27, 126.68, 120.52, 116.11, 115.73, 115.59, 62.26, 58.24, 50.95, 50.88, 45.67, 40.77, 13.95. HRMS (ES+) m/z calcd for $\text{C}_{28}\text{H}_{25}\text{N}_4\text{O}_2\text{Cl}_2\text{F}_1$ $[\text{M}+\text{H}]^+$: 539.1411, found 539.1389.

Ethyl-1-(3-fluorobenzyl)-7-(4-fluorobenzyl)-5-phenyl-4,5,6,7-tetrahydro-1H-

[1,2,3]triazolo[4,5-c]pyridine-7-carboxylate (14r): Yellow oil (60%). ^1H NMR (300MHz, CDCl_3): δ 7.33-7.20 (m, 3H), 7.04-6.88 (m, 9H), 6.83 (d, $J = 7.7\text{ Hz}$, 1H), 6.76 (dt, $J = 9.6\text{ Hz}, 2.1\text{ Hz}$, 1H), 5.11 (d, $J = 16.2\text{ Hz}$, 1H), 4.68 (d, $J = 16.2\text{ Hz}$, 1H), 4.51 (d, $J = 14.2\text{ Hz}$, 1H), 4.36 (d, $J = 14.2\text{ Hz}$, 1H), 4.06 (dq, $J = 10.9\text{ Hz}, 7.1\text{ Hz}$, 1H), 3.74-3.58 (m, 3H), 3.35 (d, $J = 13.5\text{ Hz}$, 1H), 3.28 (d, $J = 13.5\text{ Hz}$, 1H), 1.04 (t, $J = 7.1\text{ Hz}$, 3H). ^{13}C NMR (151 MHz, CDCl_3): δ 171.16, 163.72, 163.08, 162.08, 161.44, 149.75, 142.52, 137.87,

137.82, 131.97, 131.91, 131.57, 131.55, 130.23, 130.17, 129.39, 122.64, 122.62, 120.43, 116.08, 115.61, 115.47, 114.98, 114.84, 114.26, 114.11, 62.10, 58.16, 51.55, 50.63, 45.68, 40.62, 13.89. HRMS (ES+) m/z calcd for $C_{28}H_{26}N_4O_2F_2$ $[M+H]^+$: 489.2096, found 489.2092.

1-(4-methoxyphenethyl)-5-phenyl-4,5,6,7-tetrahydro-1H-[1,2,3]triazolo[4,5-c]pyridine (18a): Yellow oil (85%) 1H NMR (400 MHz, $CDCl_3$): δ 7.26 (t, $J = 7.9$ Hz, 2H), 6.93 – 6.84 (m, 5H), 6.73 (d, $J = 8.5$ Hz, 2H), 4.40-4.37 (m, 4H), 3.74 (s, 3H), 3.44 (t, $J = 5.6$ Hz, 2H), 3.10 (t, $J = 6.8$ Hz, 2H), 2.23 (t, $J = 5.6$ Hz, 2H). ^{13}C NMR (100 MHz, $CDCl_3$): δ 158.68, 149.95, 141.30, 131.31, 129.80, 129.39, 129.28, 119.80, 116.40, 114.14, 55.29, 49.79, 47.19, 45.81, 35.97, 20.13. HRMS (ES+) m/z calcd for $C_{20}H_{22}N_4O_1$ $[M+H]^+$: 335.1866, found 335.1868.

1-(2-methoxyphenethyl)-5-phenyl-4,5,6,7-tetrahydro-1H-[1,2,3]triazolo[4,5-c]pyridine (18b): Yellow oil (74%). 1H NMR (400 MHz, $CDCl_3$): δ 7.26 (t, $J = 7.9$ Hz, 2H), 7.18 (t, $J = 7.8$ Hz, 1H), 6.92 (d, $J = 8.0$ Hz, 2H), 6.89 – 6.79 (m, 3H), 6.74 (t, $J = 7.4$ Hz, 1H), 4.43 (t, $J = 6.9$ Hz, 2H), 4.38 (s, 2H), 3.80 (s, 3H), 3.45 (t, $J = 5.6$ Hz, 2H), 3.15 (t, $J = 6.8$ Hz, 2H), 2.28 (t, $J = 5.6$ Hz, 2H). ^{13}C NMR (100 MHz, $CDCl_3$): δ 157.49, 149.97, 141.20, 131.12, 130.71, 129.26, 128.48, 125.58, 120.69, 119.70, 116.32, 110.19, 55.24, 47.83, 47.11, 45.85, 31.98, 20.02. HRMS (ES+) m/z calcd for $C_{20}H_{22}N_4O_1$ $[M+H]^+$: 335.1866, found 335.1869.

1-(cyclopropylmethyl)-5-phenyl-4,5,6,7-tetrahydro-1H-[1,2,3]triazolo[4,5-c]pyridine (18c): Yellow oil (90%). 1H NMR (300 MHz, $CDCl_3$): δ 7.32 - 7.23 (m, 2H), 7.03 - 6.95 (m, 2H), 6.92 - 6.82 (m, 1H), 4.43 (s, 2H), 4.12 (d, $J = 7.1$ Hz, 2H), 3.65 (t, $J = 5.7$ Hz, 2H), 2.86 (tt, $J = 5.6, 1.3$ Hz, 2H), 1.29 – 1.22 (m, 1H), 0.68 - 0.59 (m, 2H), 0.43 - 0.38 (m, 2H). ^{13}C NMR (75 MHz, $CDCl_3$): δ 150.12, 141.91, 130.29, 129.32, 119.95, 116.48, 52.82, 47.36, 46.06, 21.06, 11.05, 4.30. HRMS (ES+) m/z calcd for $C_{15}H_{18}N_4$ $[M+H]^+$: 255.1604, found 225.1606.

1-hexyl-5-phenyl-4,5,6,7-tetrahydro-1H-[1,2,3]triazolo[4,5-c]pyridine (18d): Yellow oil (48%) 1H NMR (300MHz, $CDCl_3$): δ 7.31 - 7.26 (m, 2H), 7.01-6.98 (m, 2H), 6.91 – 6.86 (m, 1H), 4.43 (s, 2H), 4.23 (t, $J = 7.3$ Hz, 2H), 3.65 (t, $J = 5.6$ Hz, 2H), 2.81 (t, $J = 5.6$ Hz, 2H), 1.91-1.82 (m, 2H), 1.32-1.26 (m, 6H), 0.87 (t, $J = 6.9$ Hz, 3H). ^{13}C NMR (75 MHz, $CDCl_3$) δ 150.10, 141.79, 130.43, 129.33, 119.99, 116.53, 48.06, 47.35, 46.05, 31.19, 29.86, 26.23, 22.42, 20.80, 13.95. HRMS (ES+) m/z calcd for $C_{17}H_{24}N_4$ $[M+H]^+$: 285.2073, found 285.2068.

1-(3-methoxyphenethyl)-5-(4-(trifluoromethyl)phenyl)-4,5,6,7-tetrahydro-1H-[1,2,3]triazolo[4,5-c]pyridine (18e): Yellow oil (90%). 1H NMR (300 MHz, $CDCl_3$): δ 7.48 (d, $J = 8.6$ Hz, 2H), 7.08 (t, $J = 7.9$ Hz, 1H), 6.90 (d, $J = 8.8$ Hz, 2H), 6.72 (ddd, $J = 8.3, 2.6, 0.9$ Hz, 1H), 6.54 (d, $J = 7.5$, 1H), 6.45 (t, $J = 2.1$ Hz, 1H), 4.48 (s, 2H), 4.43 (t, $J = 6.7$ Hz, 2H), 3.68 (s, 3H), 3.50 (t, $J = 5.6$ Hz, 2H), 3.15 (t, $J = 6.7$ Hz, 2H), 2.17 (t, J

= 5.6 Hz, 2H). ¹³C NMR (101 MHz, CDCl₃) δ 159.81, 151.93, 140.59, 138.87, 131.44, 129.76, 126.65, 126.62, 126.58, 126.54, 121.01, 114.60, 114.29, 112.56, 55.12, 49.61, 45.89, 45.19, 36.95, 19.79. HRMS (ES+) m/z calcd for C₂₁H₂₁N₄O₁F₃ [M+H]⁺: 403.1740, found 403.1736.

1-(4-methoxyphenethyl)-5-(4-(trifluoromethyl)phenyl)-4,5,6,7-tetrahydro-1H-

[1,2,3]triazolo[4,5-c]pyridine (18f): Yellow oil (85%). ¹H NMR (300 MHz, CDCl₃) δ 7.50 – 7.45 (m, 1H), 6.91 (d, *J* = 8.6 Hz, 2H), 6.87 – 6.82 (m, 2H), 6.74 – 6.69 (m, 2H), 4.48 (s, 2H), 4.40 (t, *J* = 6.7 Hz, 2H), 3.72 (s, 3H), 3.52 (t, *J* = 5.6 Hz, 2H), 3.11 (t, *J* = 6.7 Hz, 2H), 2.18 (t, *J* = 5.6 Hz, 2H). ¹³C NMR (151 MHz, CDCl₃) δ 158.72, 151.99, 140.62, 131.29, 129.81, 129.30, 126.63, 126.61, 126.58, 126.56, 114.64, 114.12, 55.26, 49.90, 45.97, 45.17, 36.01, 19.86. HRMS (ES+) m/z calcd for C₂₁H₂₁N₄O₁F₃ [M+H]⁺: 403.1740, found 403.1737.

1-(2-methoxyphenethyl)-5-(4-(trifluoromethyl)phenyl)-4,5,6,7-tetrahydro-1H-

[1,2,3]triazolo[4,5-c]pyridine (18g): Yellow oil (85%). ¹H NMR (300 MHz, CDCl₃) δ 7.49 – 7.45 (m, 2H), 7.16 (ddd, *J* = 8.2, 7.3, 1.9 Hz, 1H), 6.90 (d, *J* = 8.6 Hz, 2H), 6.81 – 6.76 (m, 2H), 6.69 (td, *J* = 7.3, 1.1 Hz, 1H), 4.47 – 4.43 (m, 4H), 3.80 (s, 3H), 3.52 (t, *J* = 5.6 Hz, 2H), 3.15 (t, *J* = 6.7 Hz, 2H), 2.22 (t, *J* = 5.6 Hz, 2H). ¹³C NMR (101 MHz, CDCl₃) δ 157.48, 151.98, 140.51, 131.09, 130.72, 128.54, 126.63, 126.59, 126.55, 126.52, 125.46, 120.66, 114.62, 110.15, 55.25, 47.88, 45.96, 45.20, 32.02, 19.65. HRMS (ES+) m/z calcd for C₂₁H₂₁N₄O₁F₃ [M+H]⁺: 403.1740, found 403.1738.

1-(4-fluorobenzyl)-5-(4-(trifluoromethyl)phenyl)-4,5,6,7-tetrahydro-1H-

[1,2,3]triazolo[4,5-c]pyridine (18h): Yellow oil (89%). ¹H NMR (300 MHz, CDCl₃) δ 7.50 – 7.47 (m, 2H), 7.22 – 7.18 (m, 2H), 7.07 – 7.01 (m, 2H), 6.95 (d, *J* = 8.7 Hz, 2H), 5.45 (s, 2H), 4.50 (br s, 2H), 3.66 (t, *J* = 5.7 Hz, 2H), 2.64 (t, *J* = 5.7 Hz, 2H). ¹³C NMR (151 MHz, CDCl₃) δ 163.54, 161.90, 152.05, 141.97, 130.62, 130.20, 130.18, 129.47, 129.41, 126.67, 126.65, 126.63, 126.60, 116.20, 116.05, 114.79, 51.47, 46.06, 45.29, 20.65. HRMS (ES+) m/z calcd for C₁₉H₁₆N₄F₄ [M+H]⁺: 377.1383, found 377.1382.

1-(3-fluorobenzyl)-5-(4-(trifluoromethyl)phenyl)-4,5,6,7-tetrahydro-1H-

[1,2,3]triazolo[4,5-c]pyridine (18i): Yellow oil (85%). ¹H NMR (300 MHz, CDCl₃) δ 7.49 (d, *J* = 8.6 Hz, 2H), 7.36 – 7.29 (m, 1H), 7.06 – 6.87 (m, 5H), 5.47 (s, 2H), 4.51 (s, 2H), 3.67 (t, *J* = 5.7 Hz, 2H), 2.65 (t, *J* = 5.7 Hz, 2H). ¹³C NMR (151 MHz, CDCl₃) δ 163.85, 162.21, 152.06, 141.98, 136.82, 136.77, 130.80, 130.75, 126.67, 126.65, 126.62, 126.60, 123.07, 123.05, 115.72, 115.58, 114.81, 114.64, 114.49, 51.47, 46.04, 45.27, 20.57. HRMS (ES+) m/z calcd for C₁₉H₁₆N₄F₄ [M+H]⁺: 377.1383, found 377.1371.

1-hexyl-5-(4-(trifluoromethyl)phenyl)-4,5,6,7-tetrahydro-1H-[1,2,3]triazolo[4,5-

c]pyridine (18j): Yellow oil (89%). ¹H NMR (300 MHz, CDCl₃) δ 7.50 (d, *J* = 8.7 Hz, 2H), 6.99 (d, *J* = 8.7 Hz, 2H), 4.51 (s, 2H), 4.23 (t, *J* = 7.3 Hz, 2H), 3.75 (t, *J* = 5.6 Hz, 2H), 2.83 (t, *J* = 5.6 Hz, 2H), 1.90 – 1.83 (m, 2H), 1.33 – 1.26 (m, 6H), 0.89–0.85 (m, 3H). ¹³C

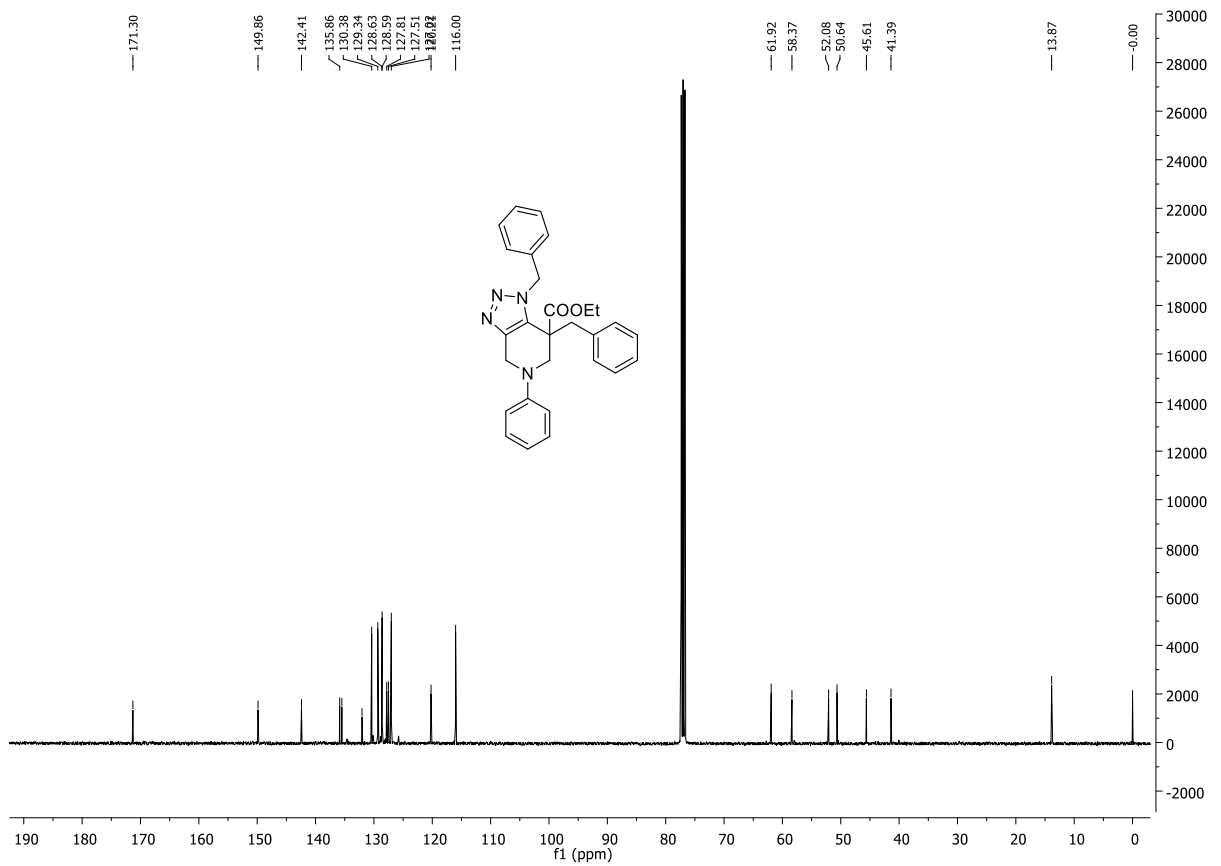
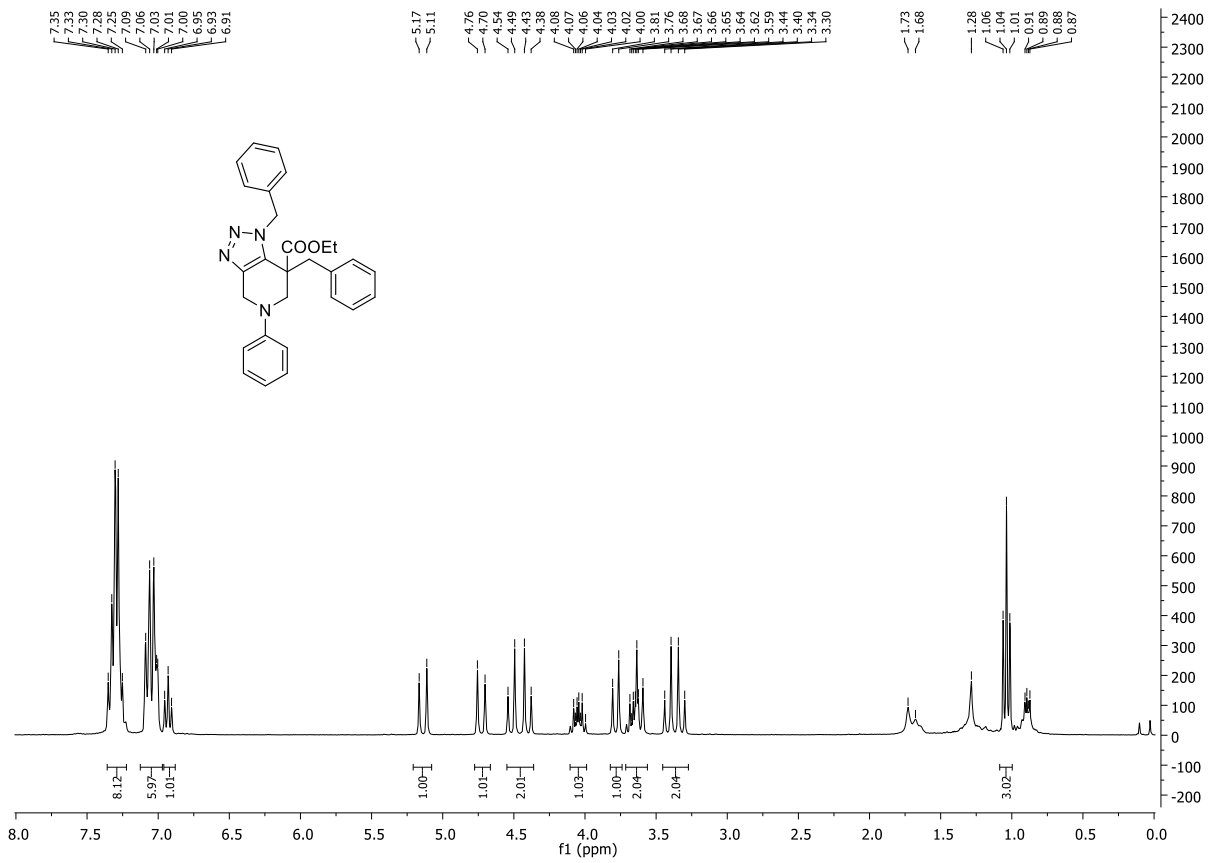
NMR (151 MHz, CDCl₃): δ 152.14, 141.15, 130.30, 126.64 (q, 4 Hz), 125.52, 123.73, 120.87, 120.65, 114.78, 48.13, 46.14, 45.33, 31.17, 29.86, 26.22, 22.42, 20.55, 13.93. HRMS (ES+) m/z calcd for C₁₈H₂₃N₄F₃ [M+H]⁺: 353.1947, found 353.1941.

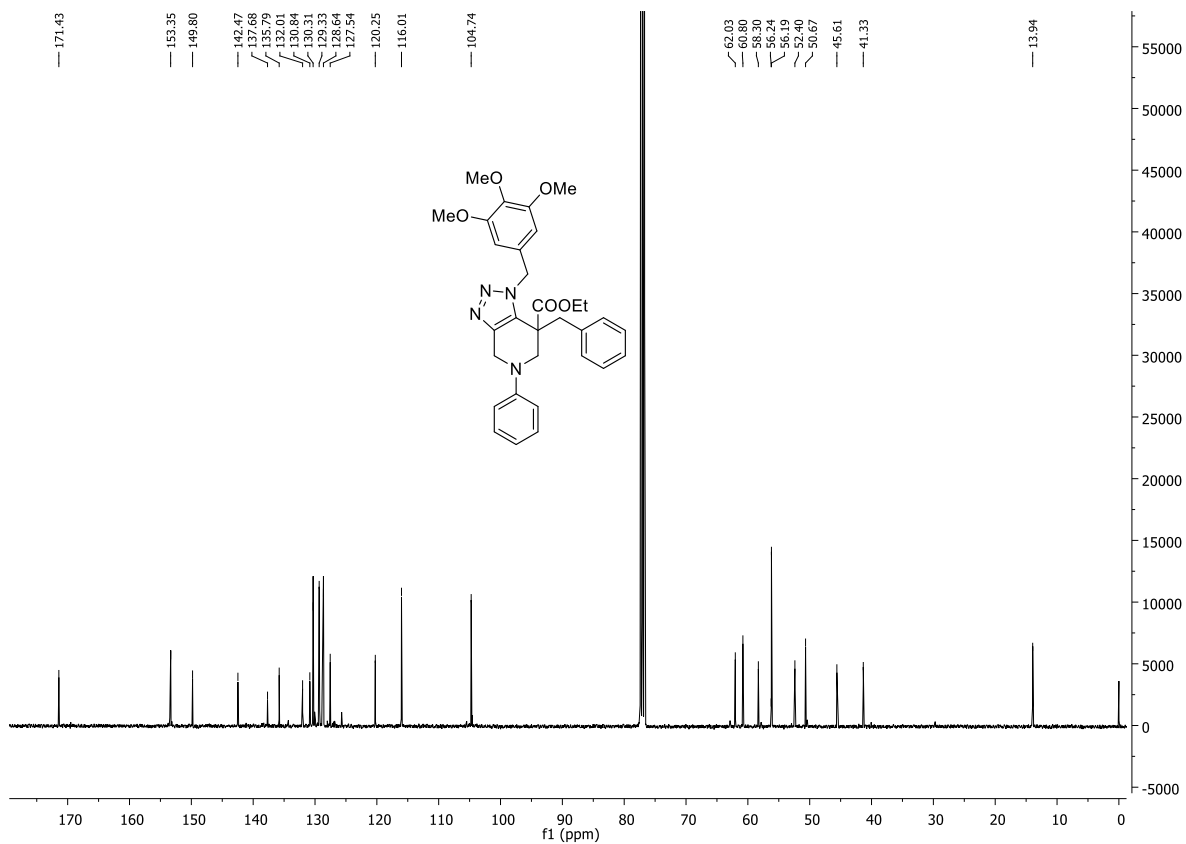
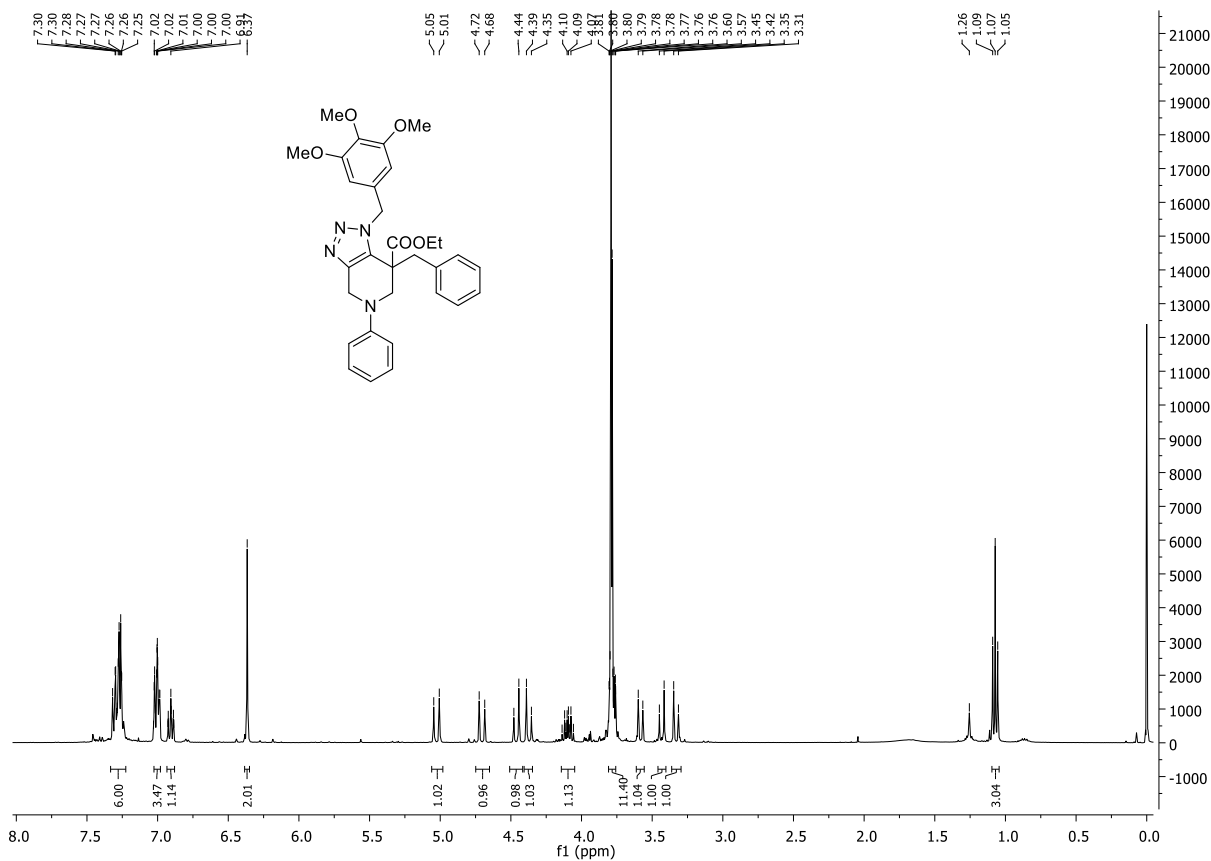
5-(4-fluorophenyl)-1-hexyl-4,5,6,7-tetrahydro-1H-[1,2,3]triazolo[4,5-c]pyridine

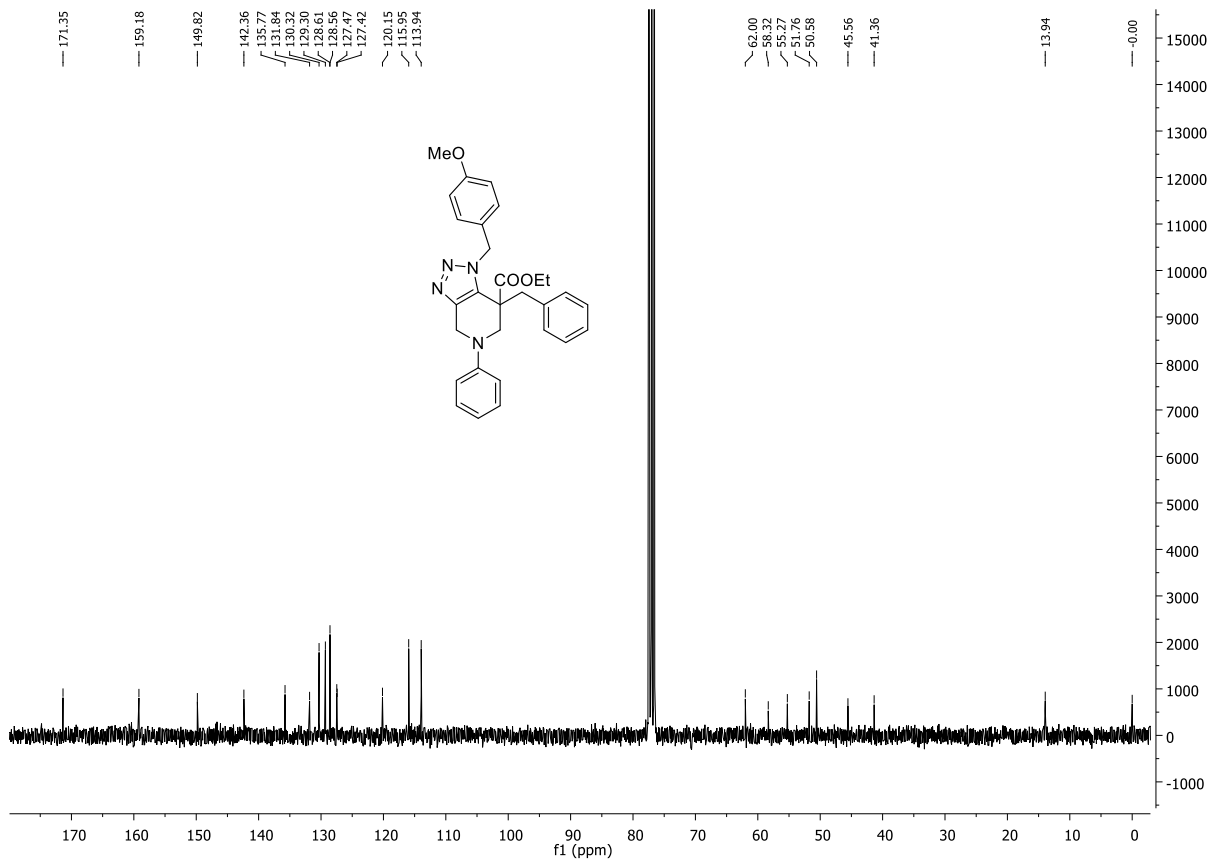
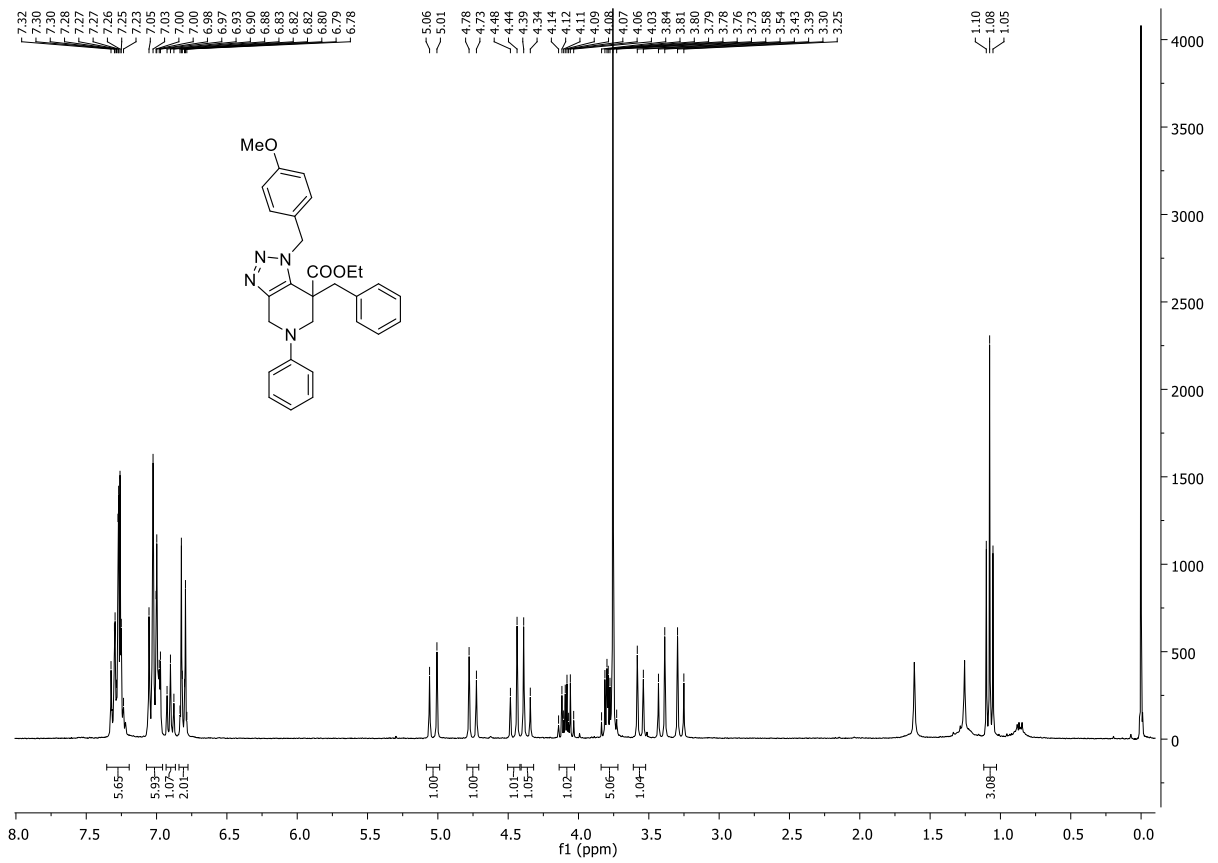
(18k): Yellow oil (91%) ¹H NMR (300 MHz, CDCl₃): δ 7.01-6.91 (m, 4H), 4.35 (br s, 2H), 4.23 (t, *J* = 7.3 Hz, 2H), 3.57 (t, *J* = 5.7 Hz, 2H), 2.80 (t, *J* = 5.7 Hz, 2H), 1.89-1.84 (m, 2H), 1.44 – 1.13 (m, 6H), 0.87 (t, *J* = 5.7 Hz, 3H). ¹³C NMR (75 MHz, CDCl₃) δ 158.90, 155.73, 146.72, 146.69, 141.63, 130.27, 118.59, 118.49, 115.90, 115.60, 48.37, 48.08, 46.94, 31.18, 29.85, 26.21, 22.42, 20.77, 13.95. HRMS (ES+) m/z calcd for C₁₇H₂₃N₄F₁ [M+H]⁺: 303.1979, found 303.1983.

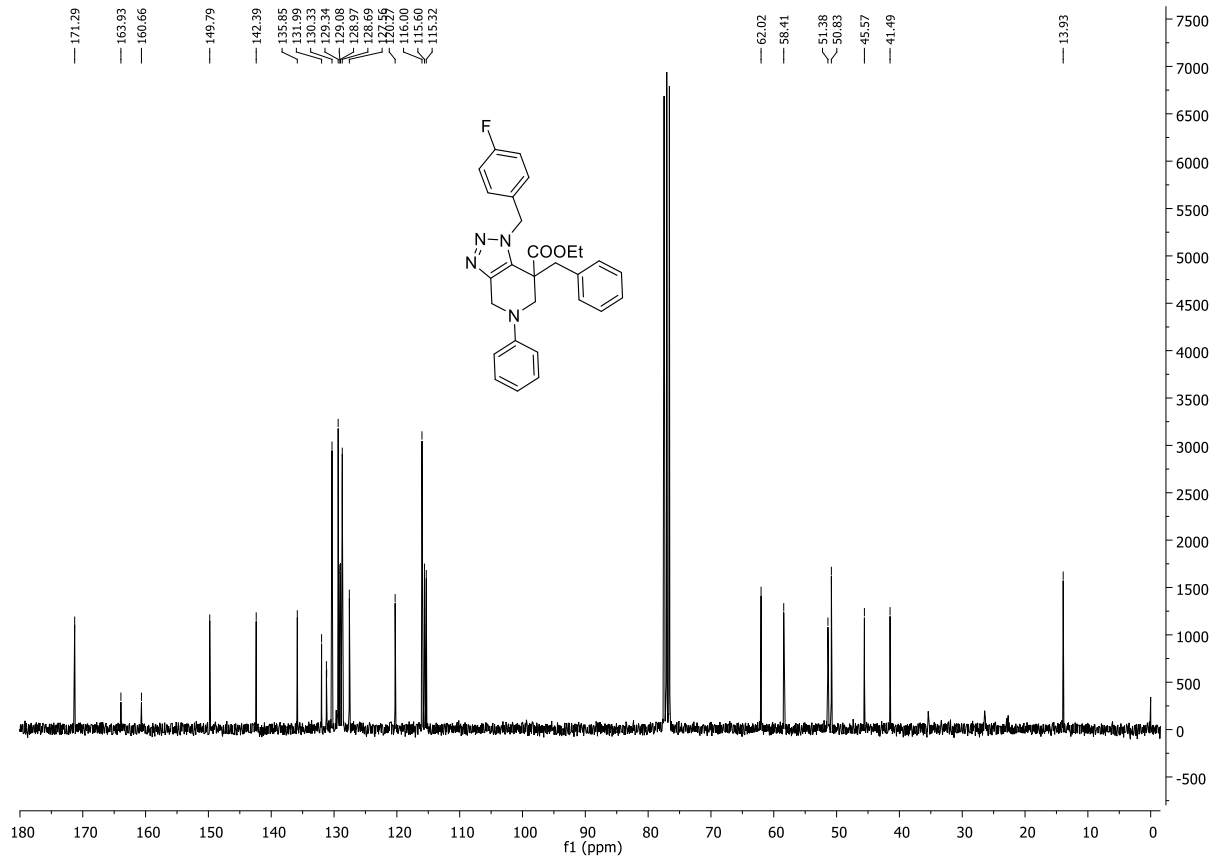
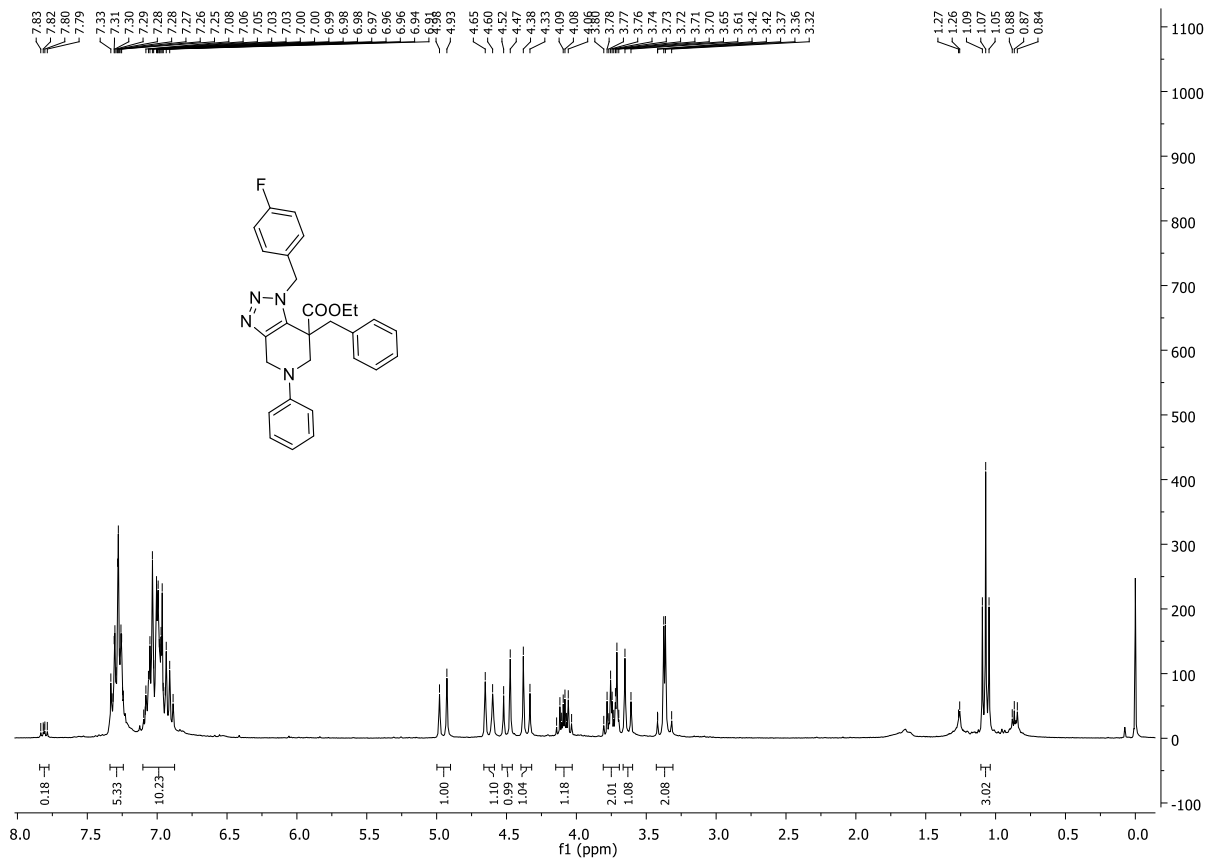
1-(2,2-diphenylethyl)-5-(4-fluorophenyl)-4,5,6,7-tetrahydro-1H-[1,2,3]triazolo[4,5-

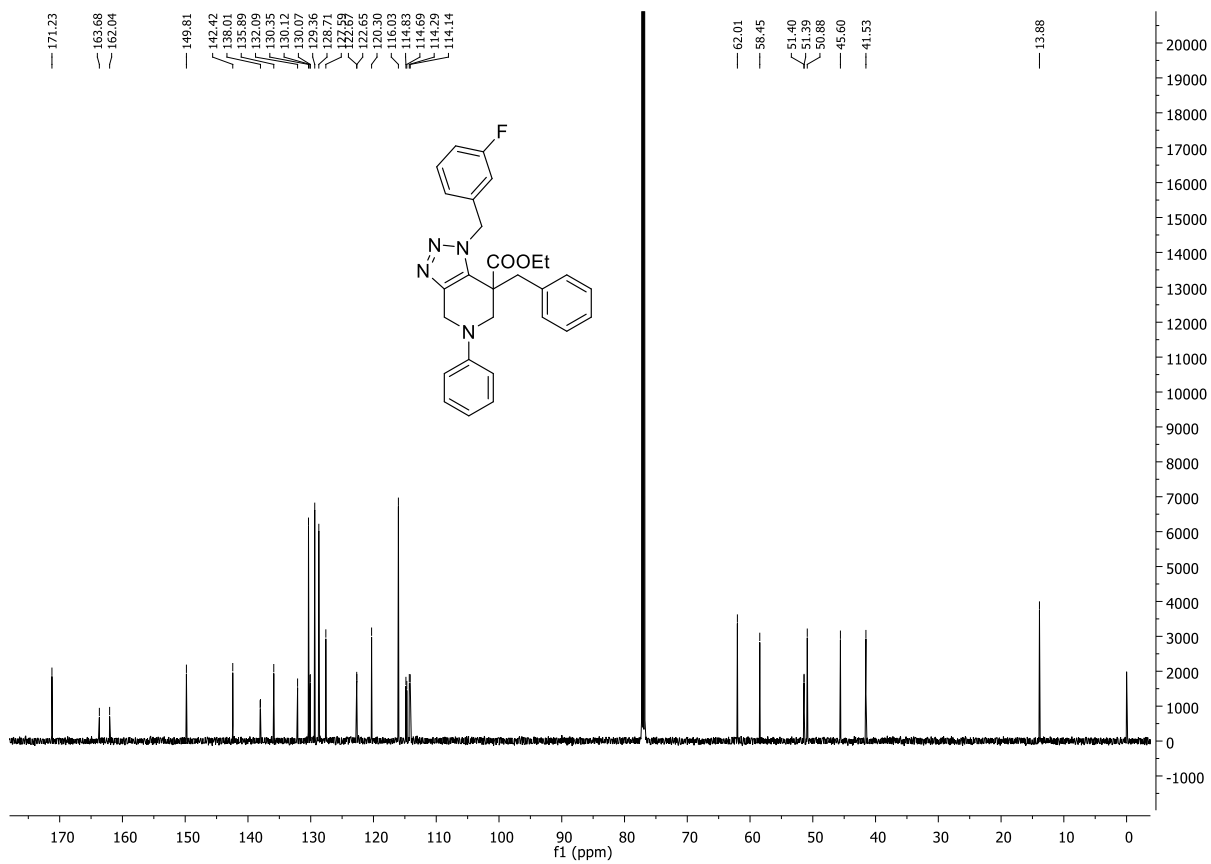
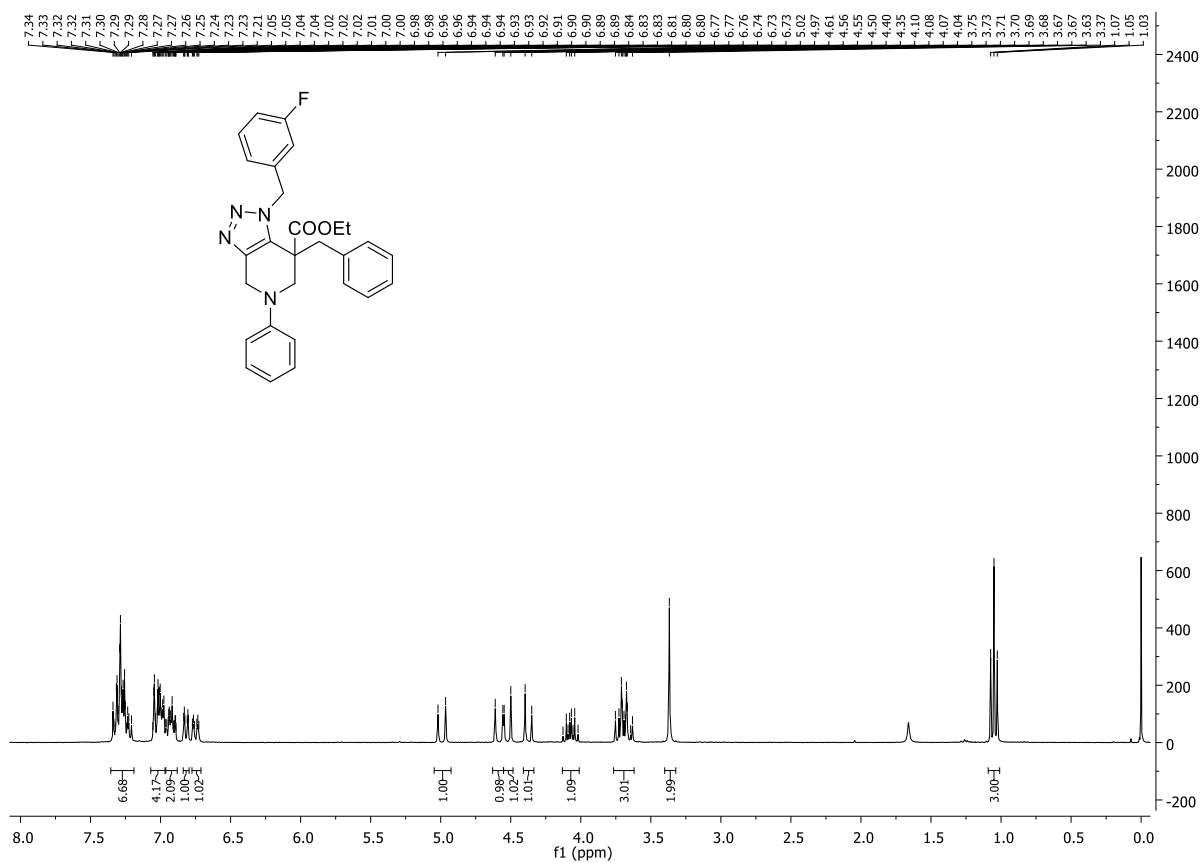
c]pyridine (18l): Yellow oil (57%). ¹H NMR (300 MHz, CDCl₃): δ 7.28-7.19 (m, 7H), 7.13 – 7.10 (m, 4H), 6.97-6.91 (m, 2H), 6.83-6.79 (m, 2H), 4.81 – 4.77 (m, 2H), 4.67 – 4.62 (m, 1H), 4.28 (s, 2H), 3.25 (t, *J* = 5.6 Hz, 2H), 1.97 (t, *J* = 5.6, 2H). ¹³C NMR (101 MHz, CDCl₃): δ 158.34, 146.37, 140.81, 140.70, 131.55, 128.74, 127.95, 127.25, 118.27, 118.20, 115.77, 115.55, 52.96, 51.71, 48.02, 46.54, 19.83. HRMS (ES+) m/z calcd for C₂₅H₂₃N₄F₁ [M+H]⁺: 399.1979, found 399.1976.

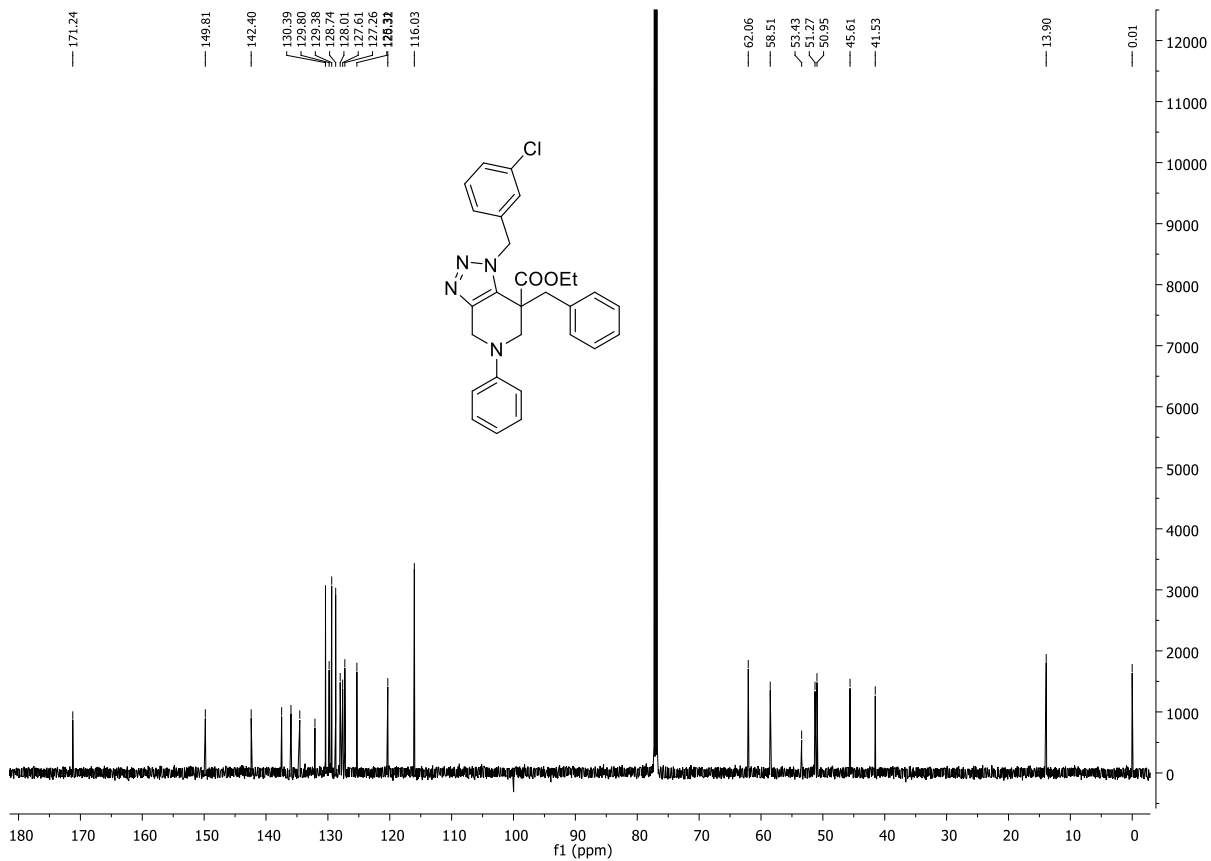
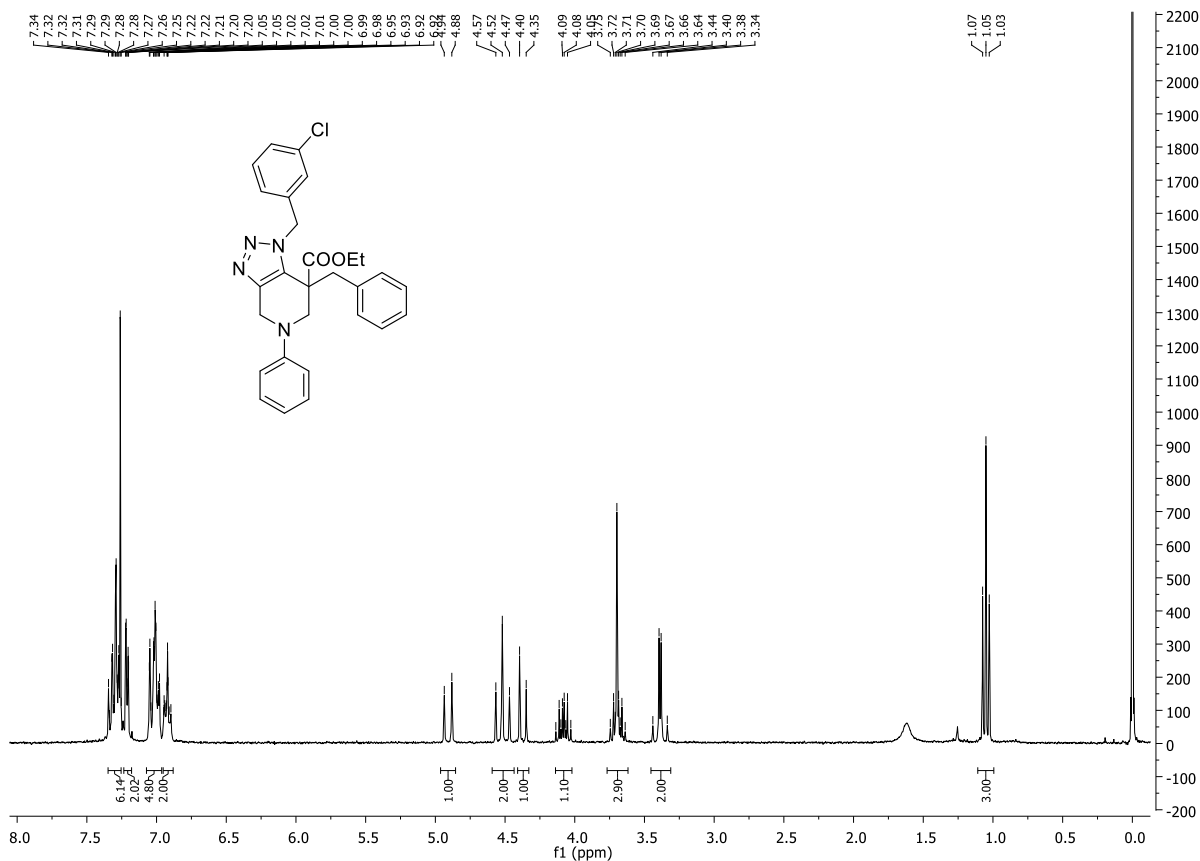


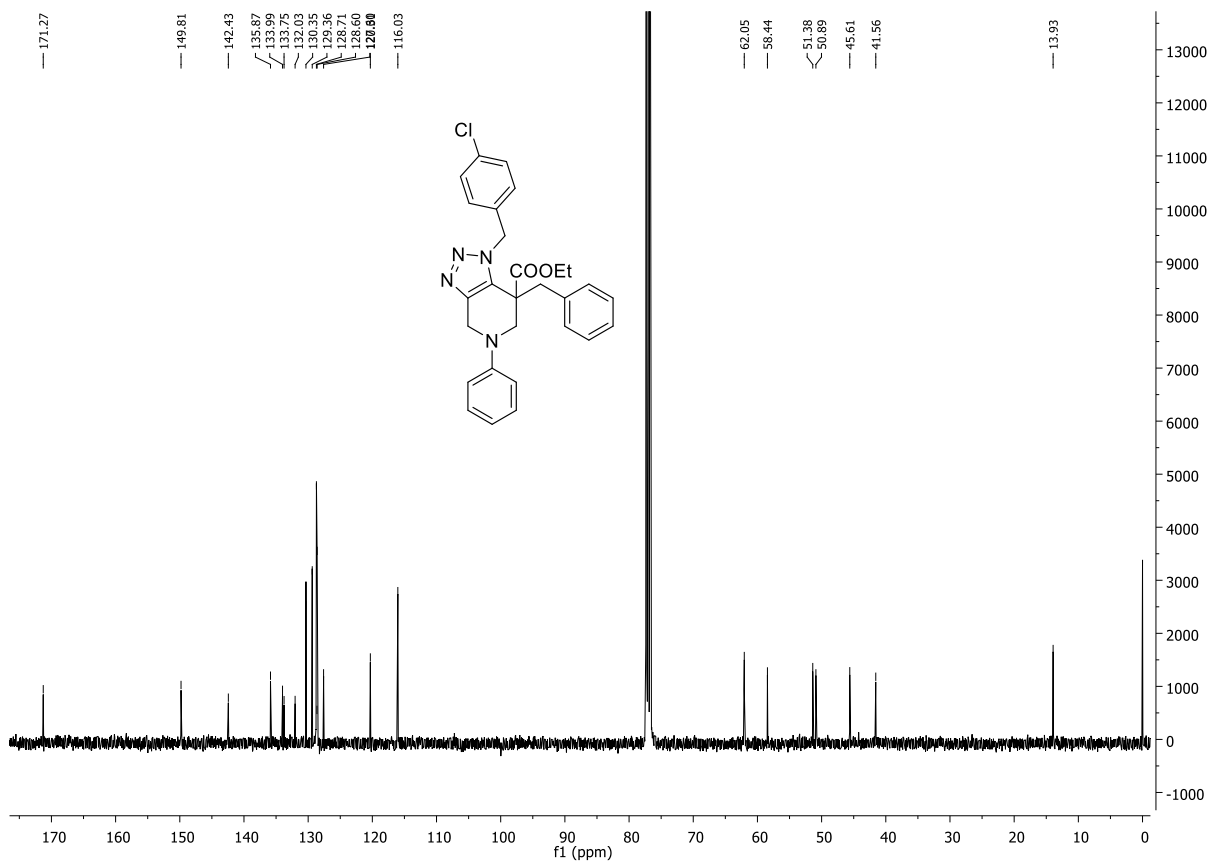
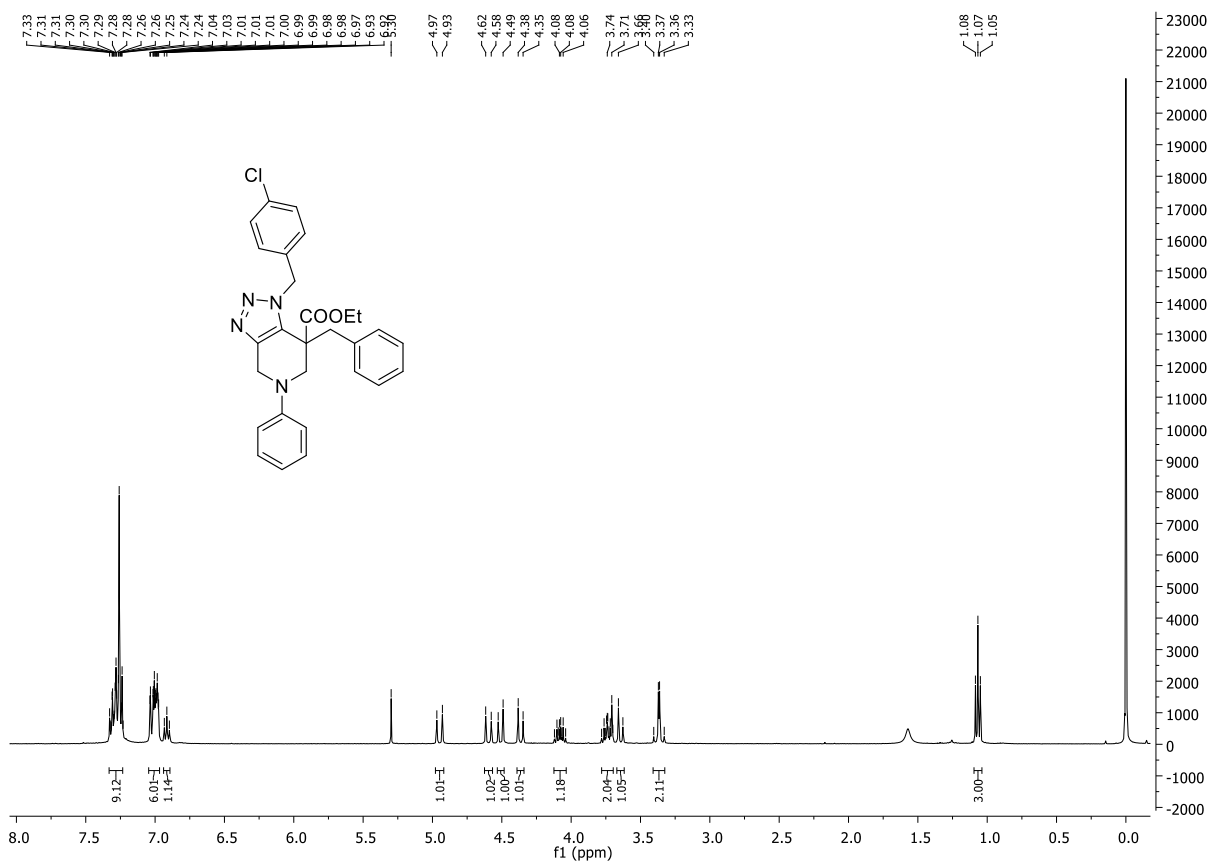


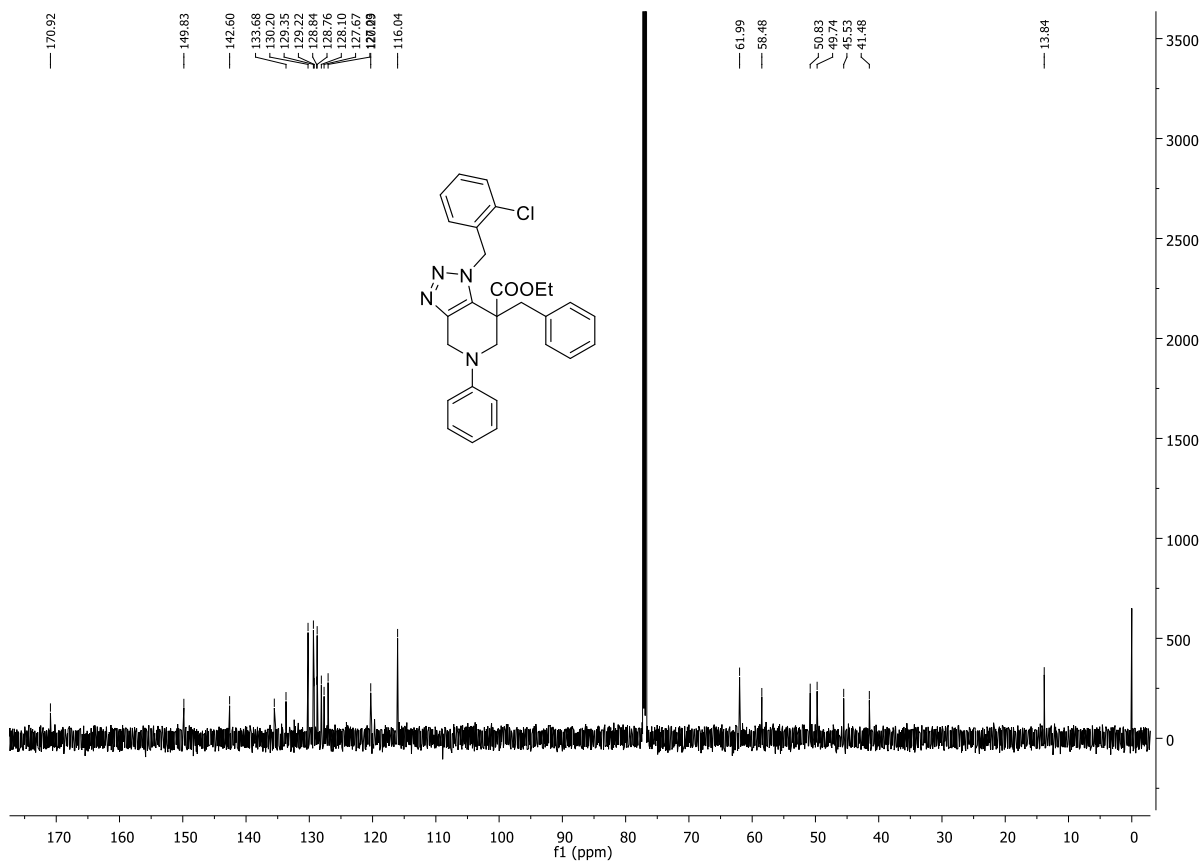
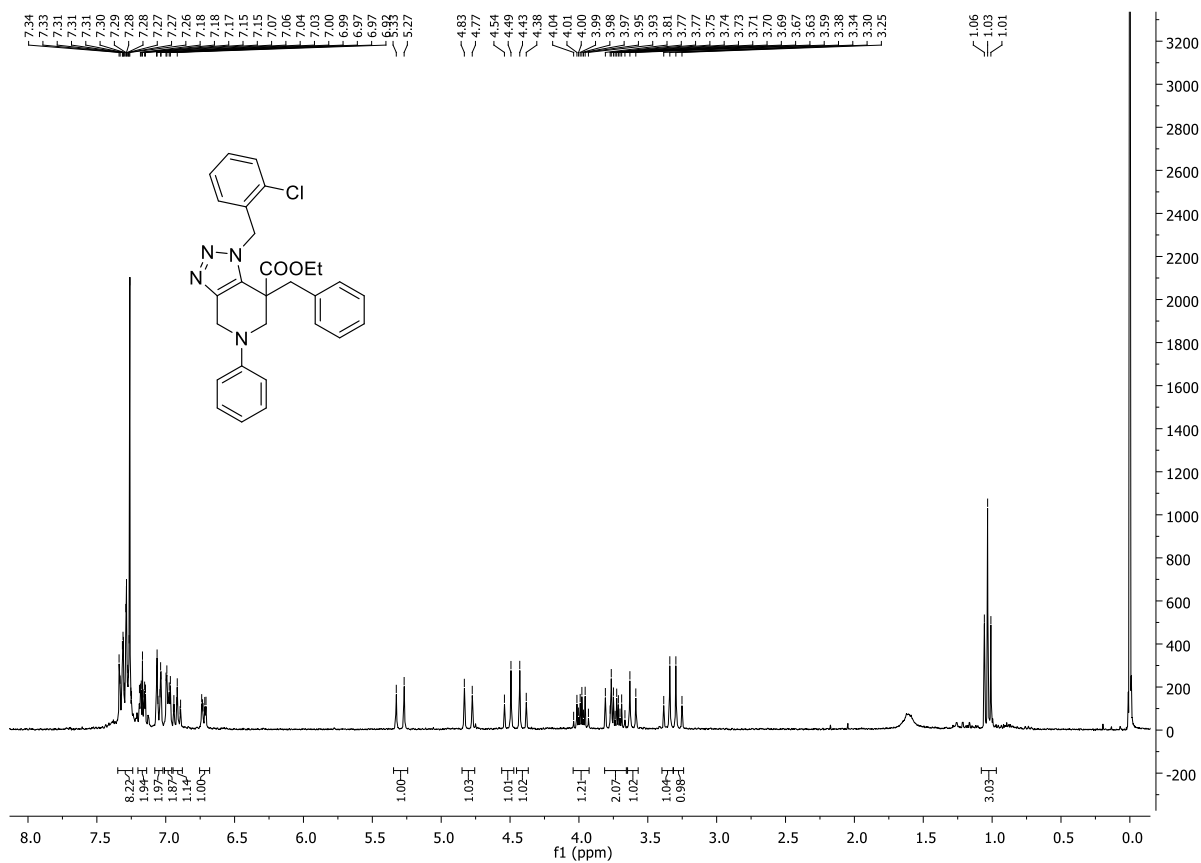


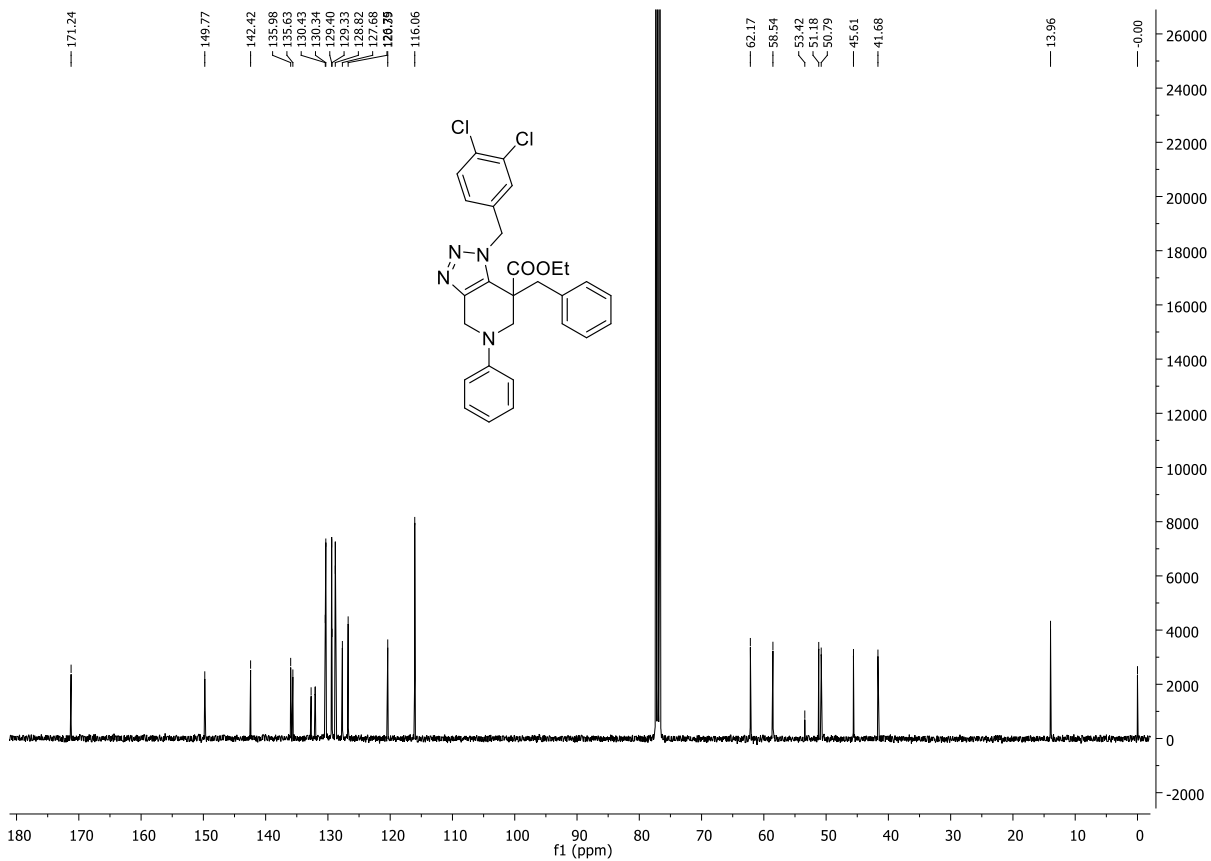
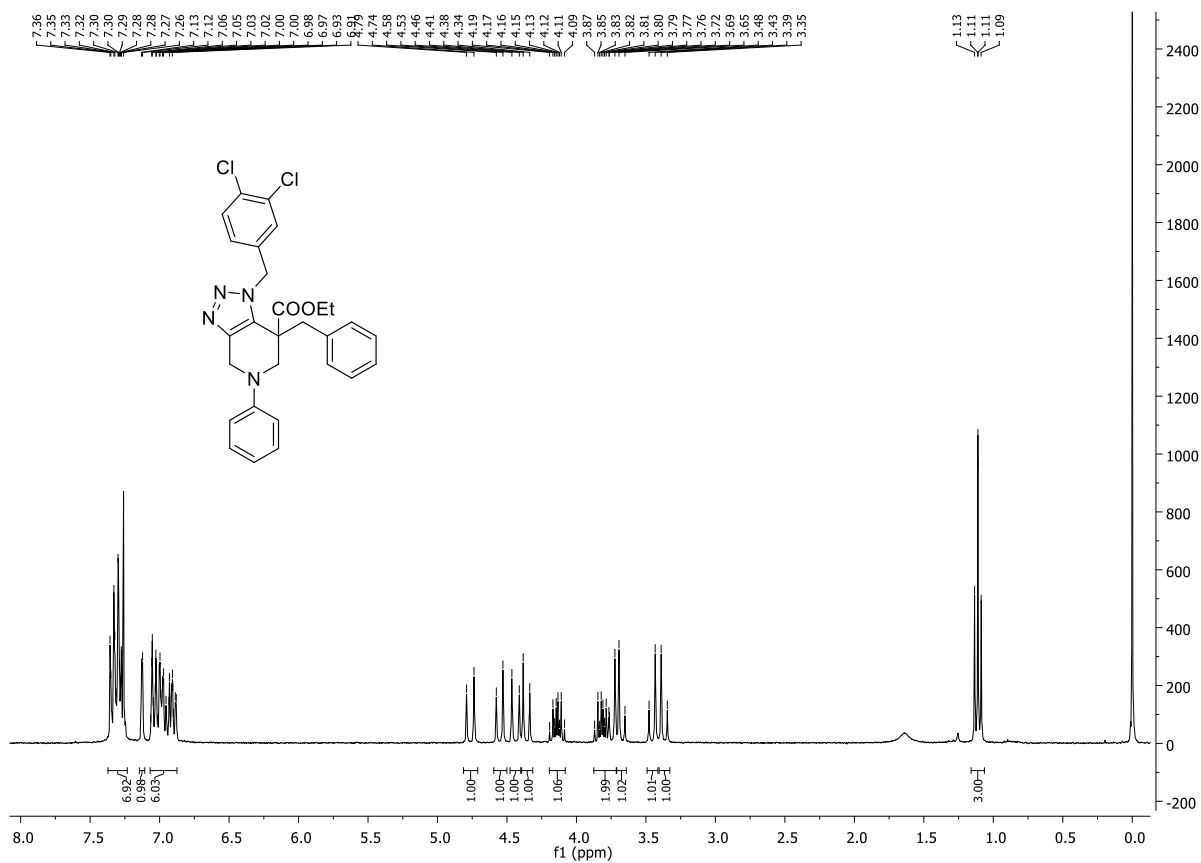


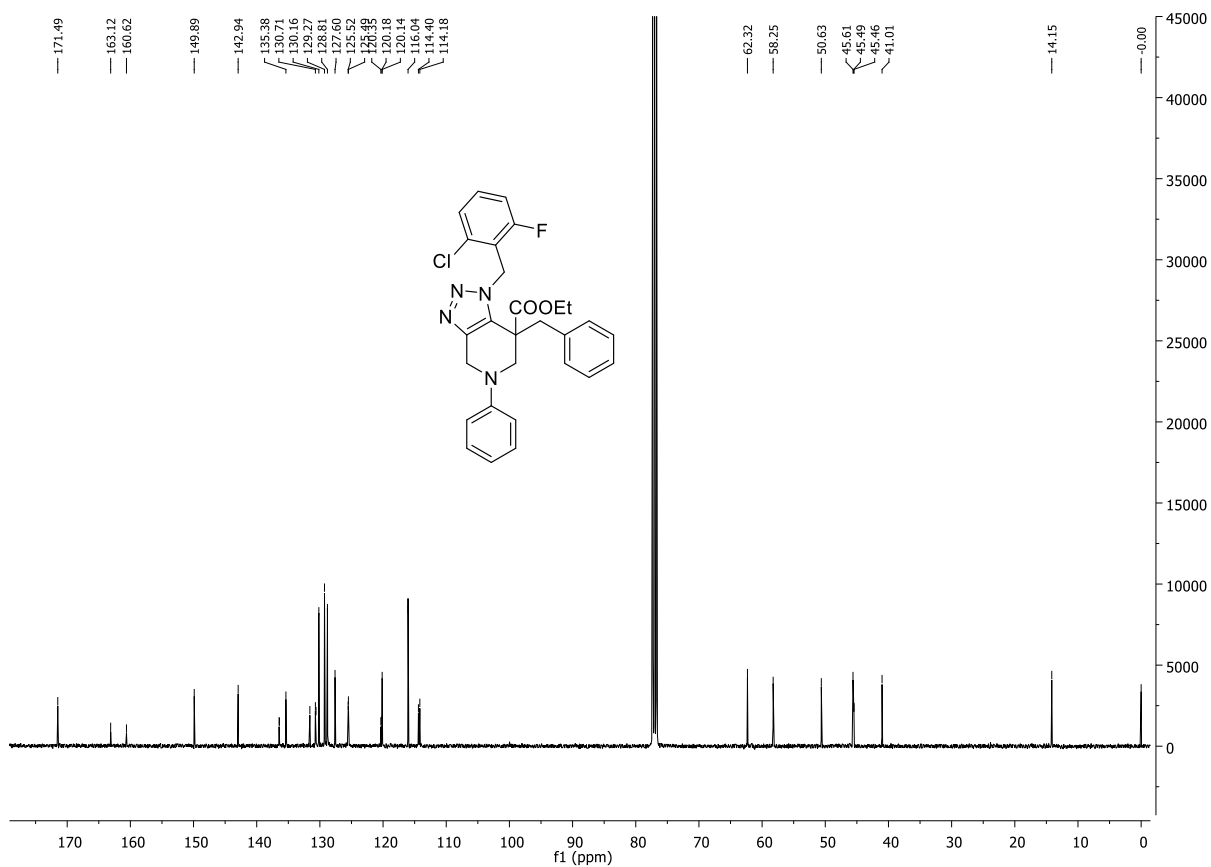
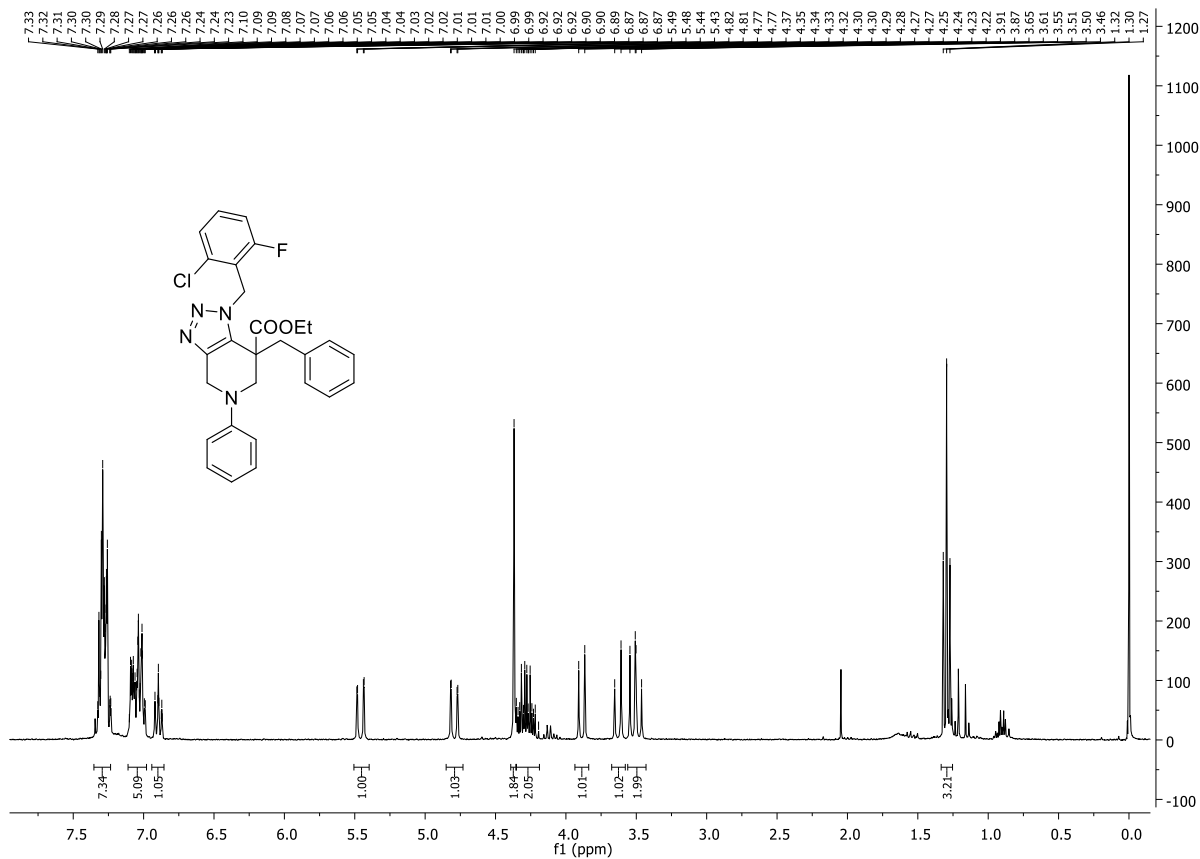


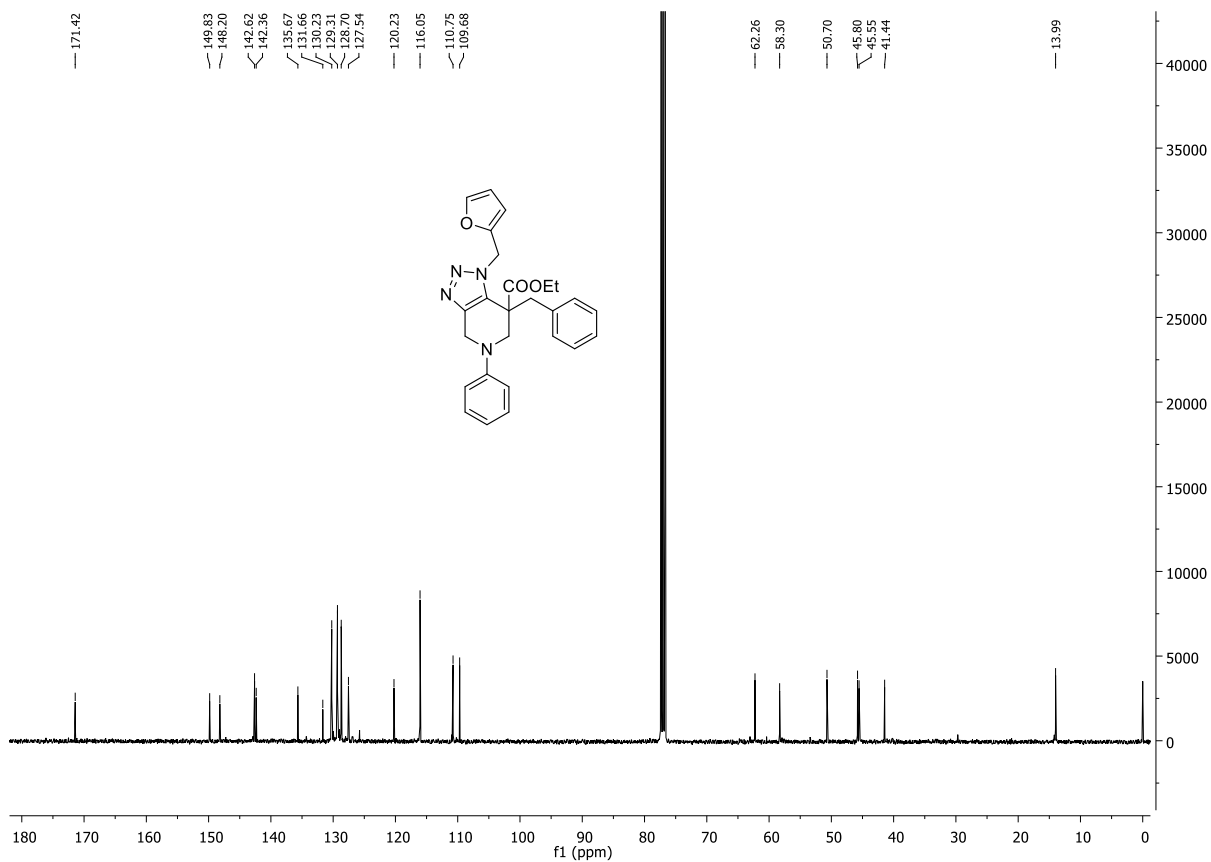
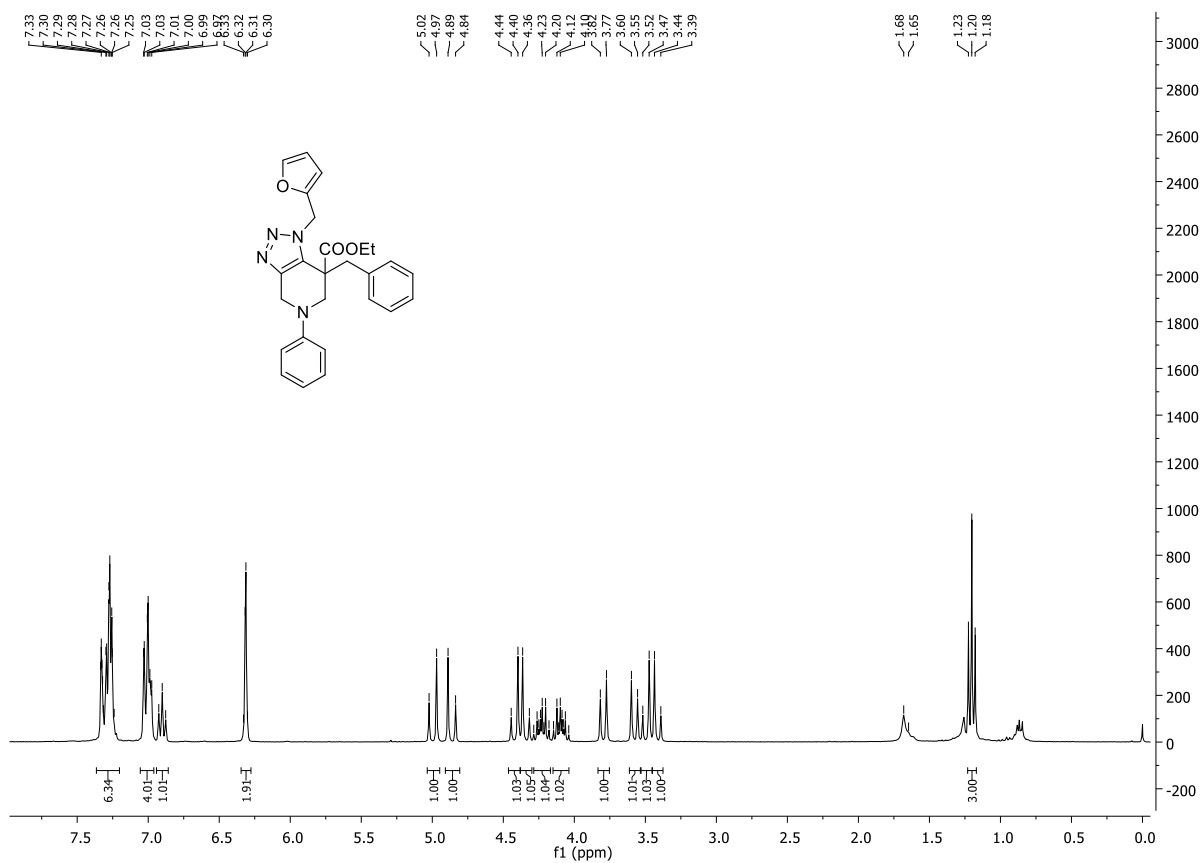


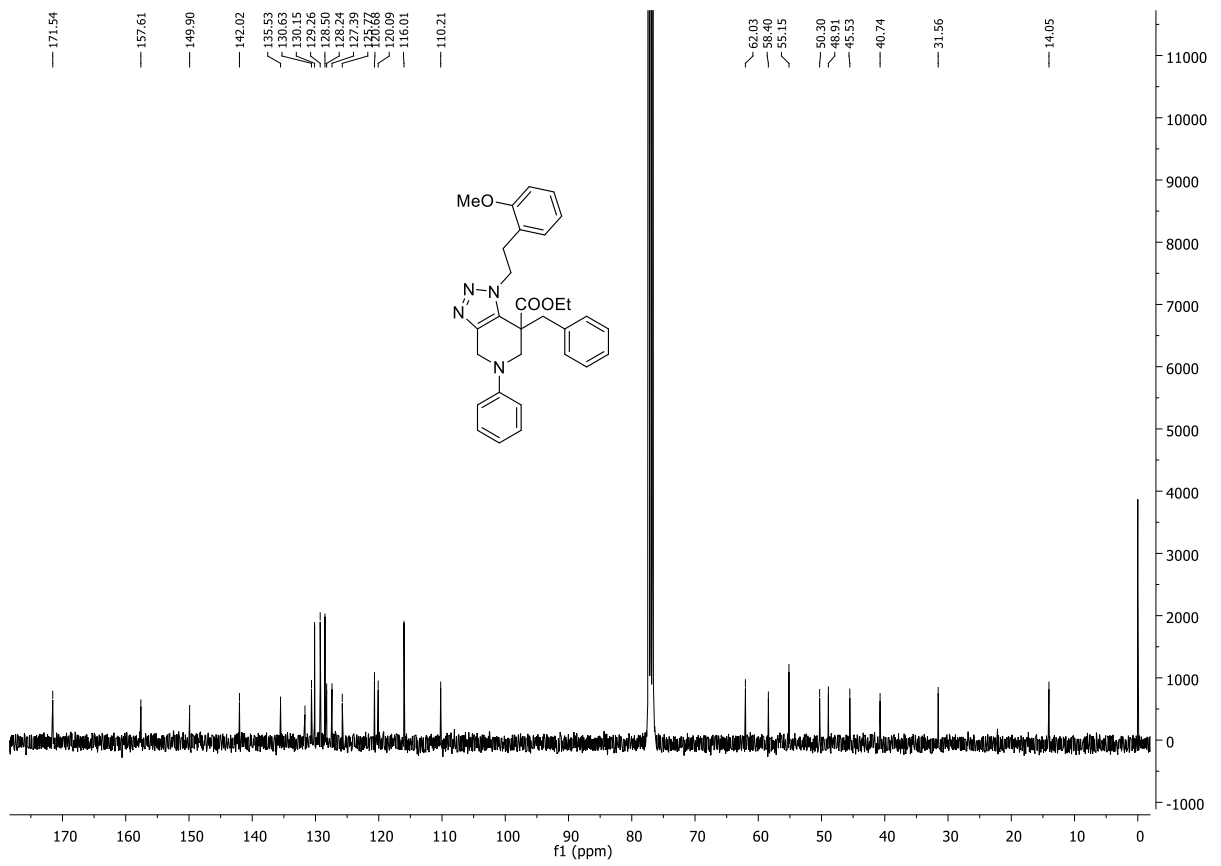
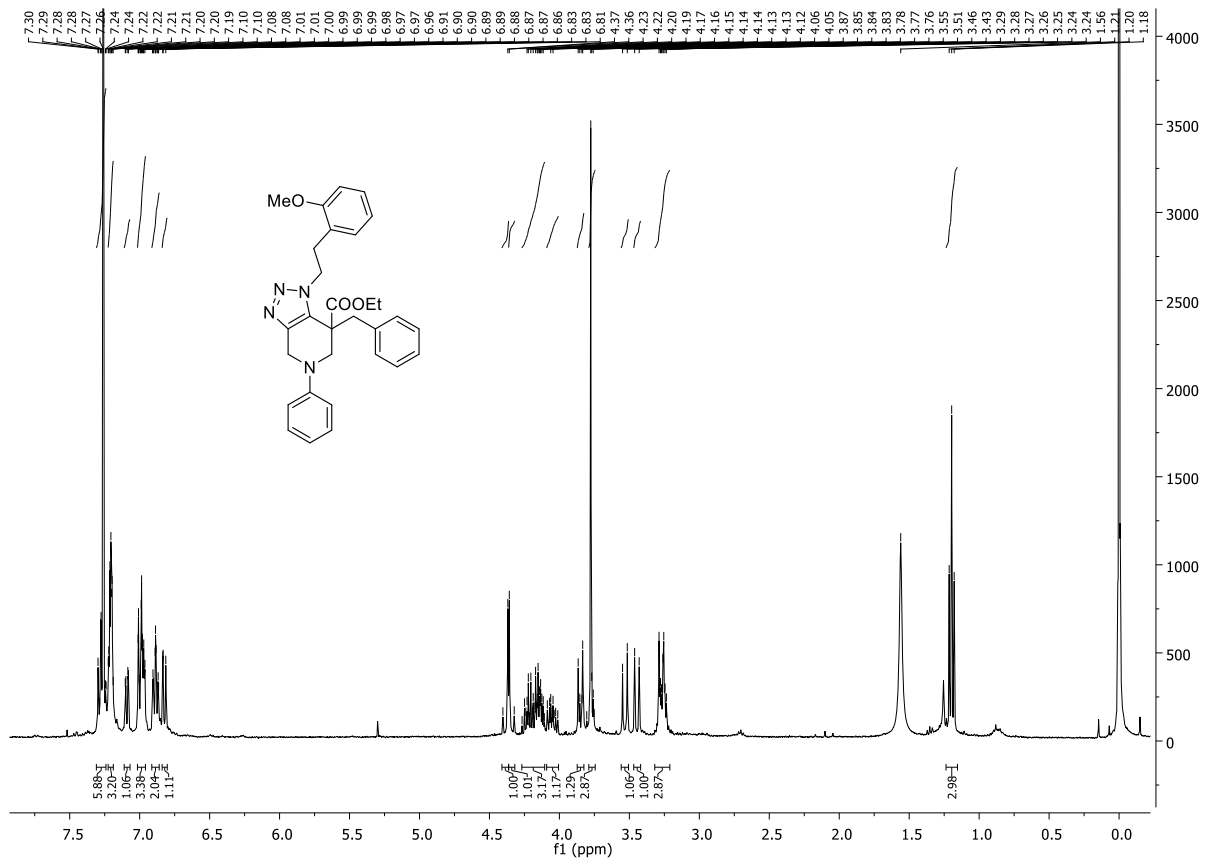


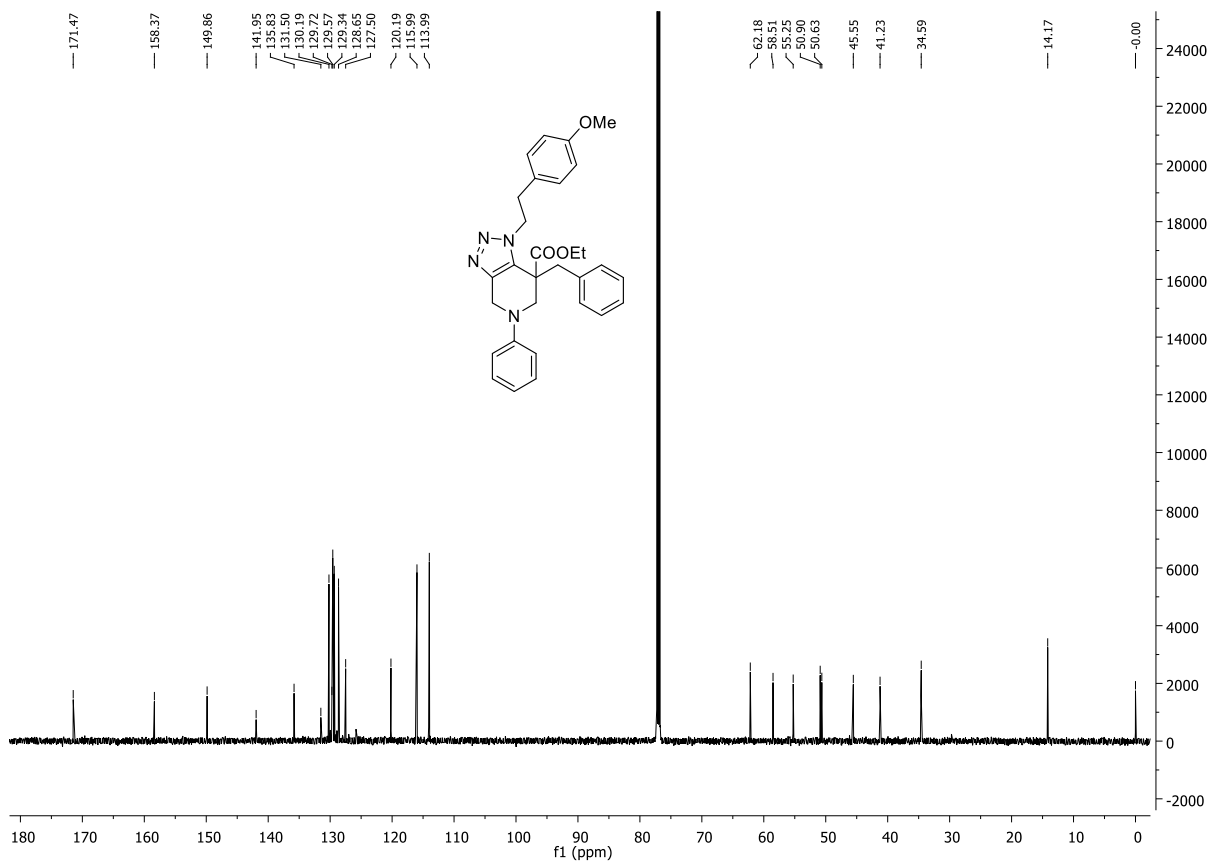
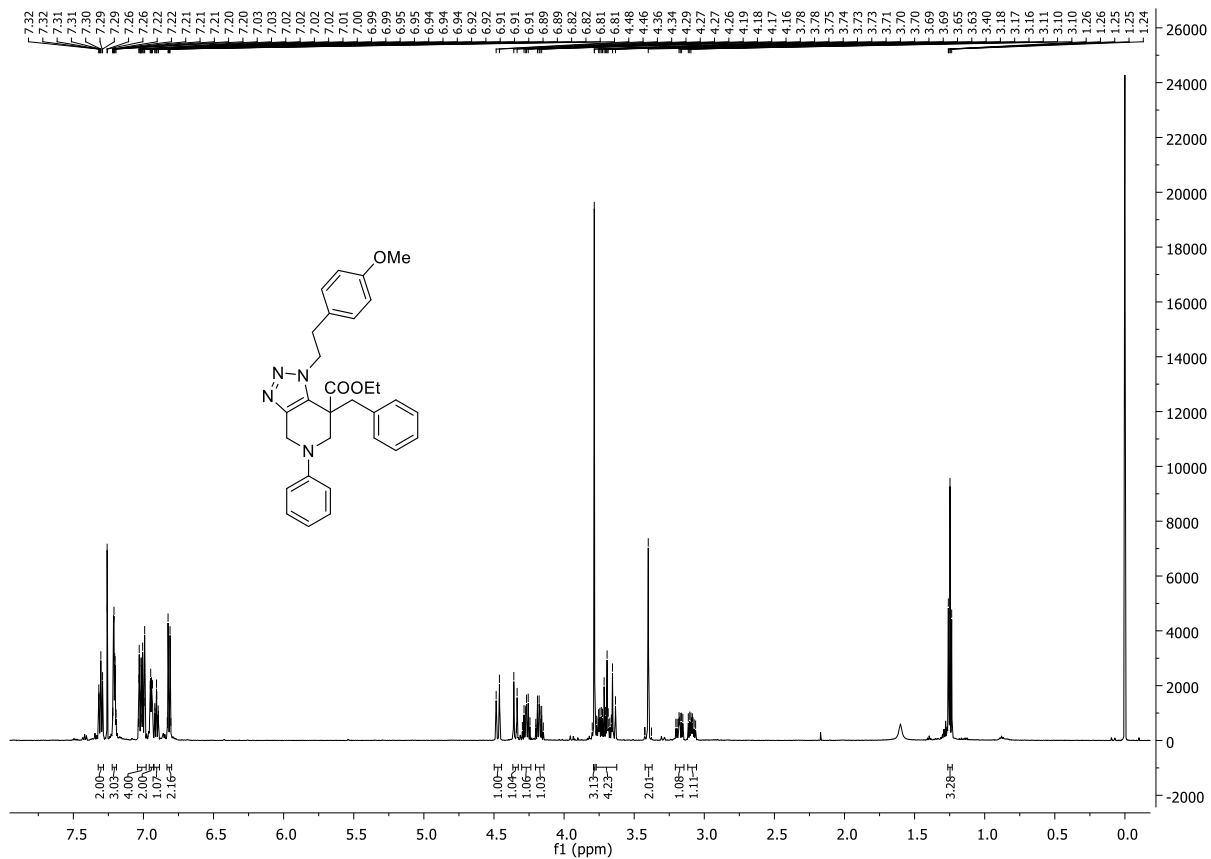


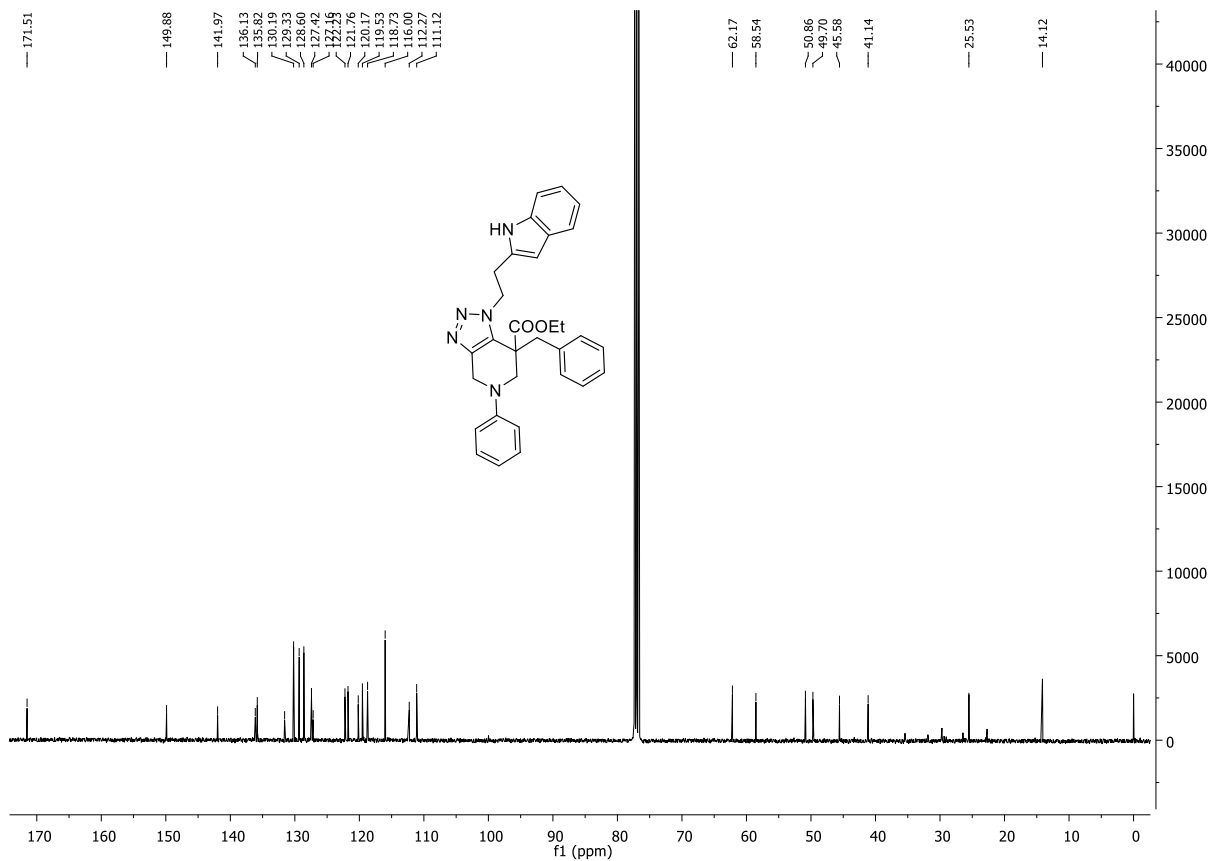
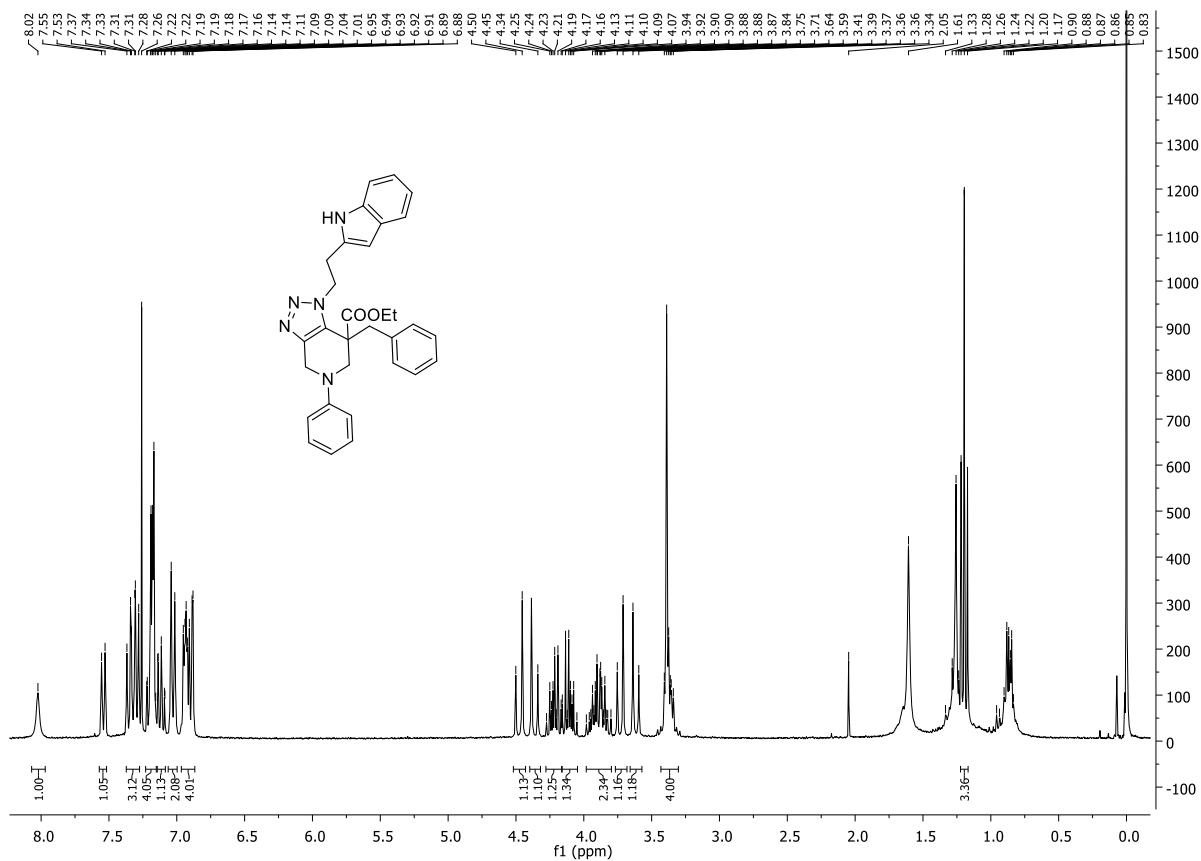


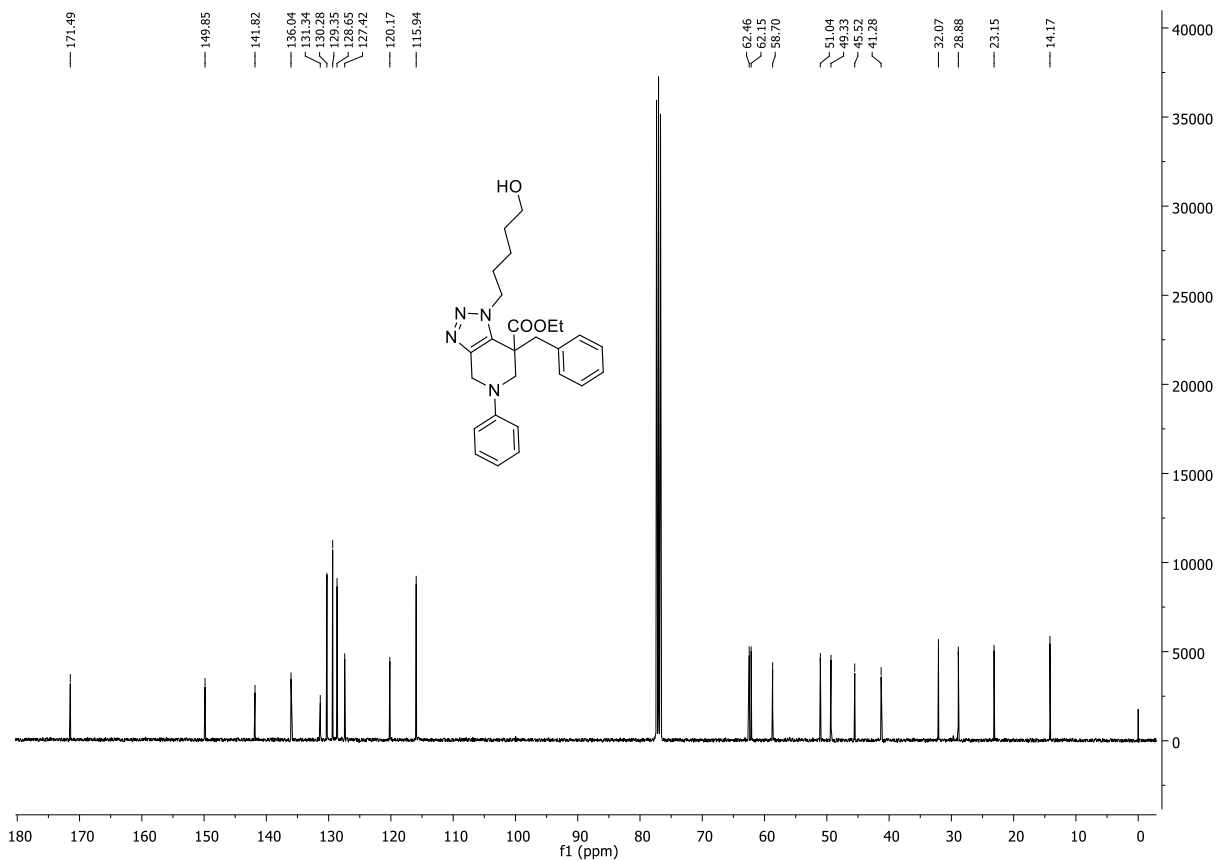
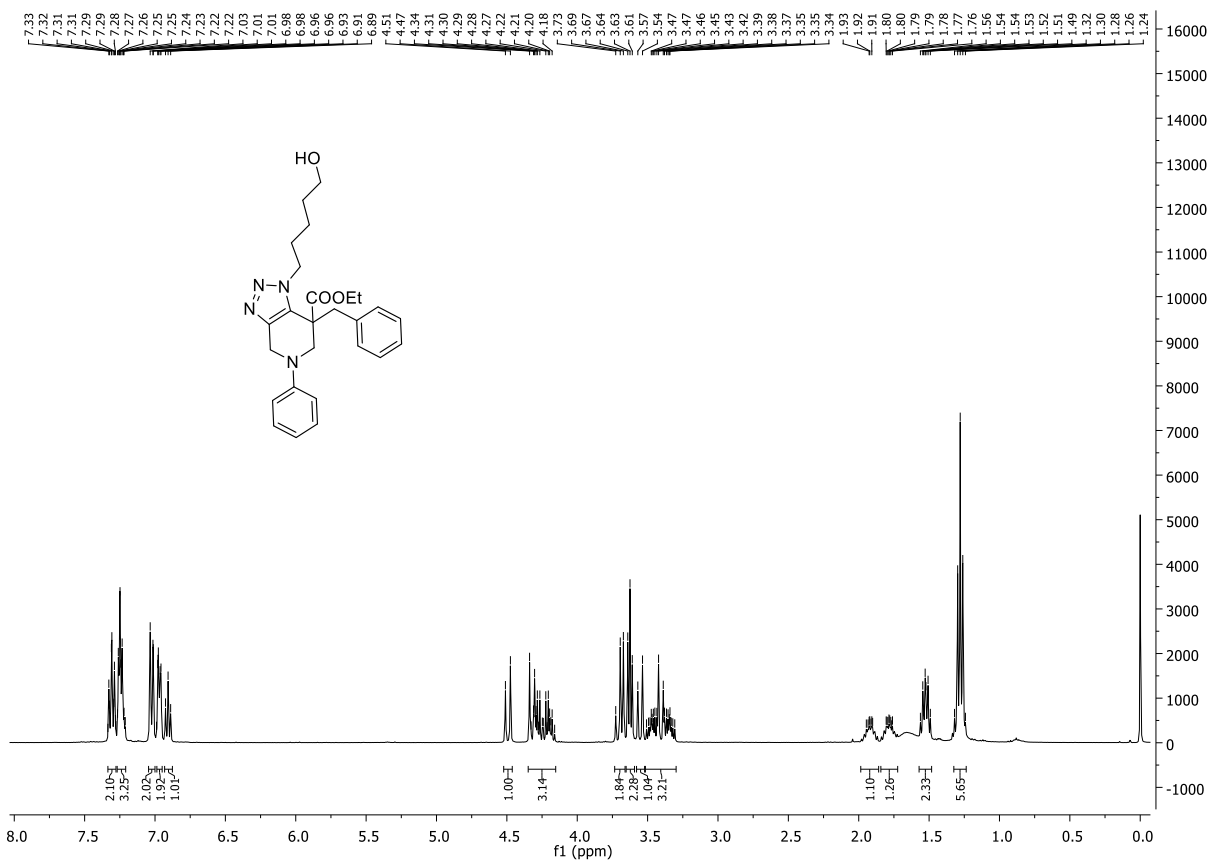


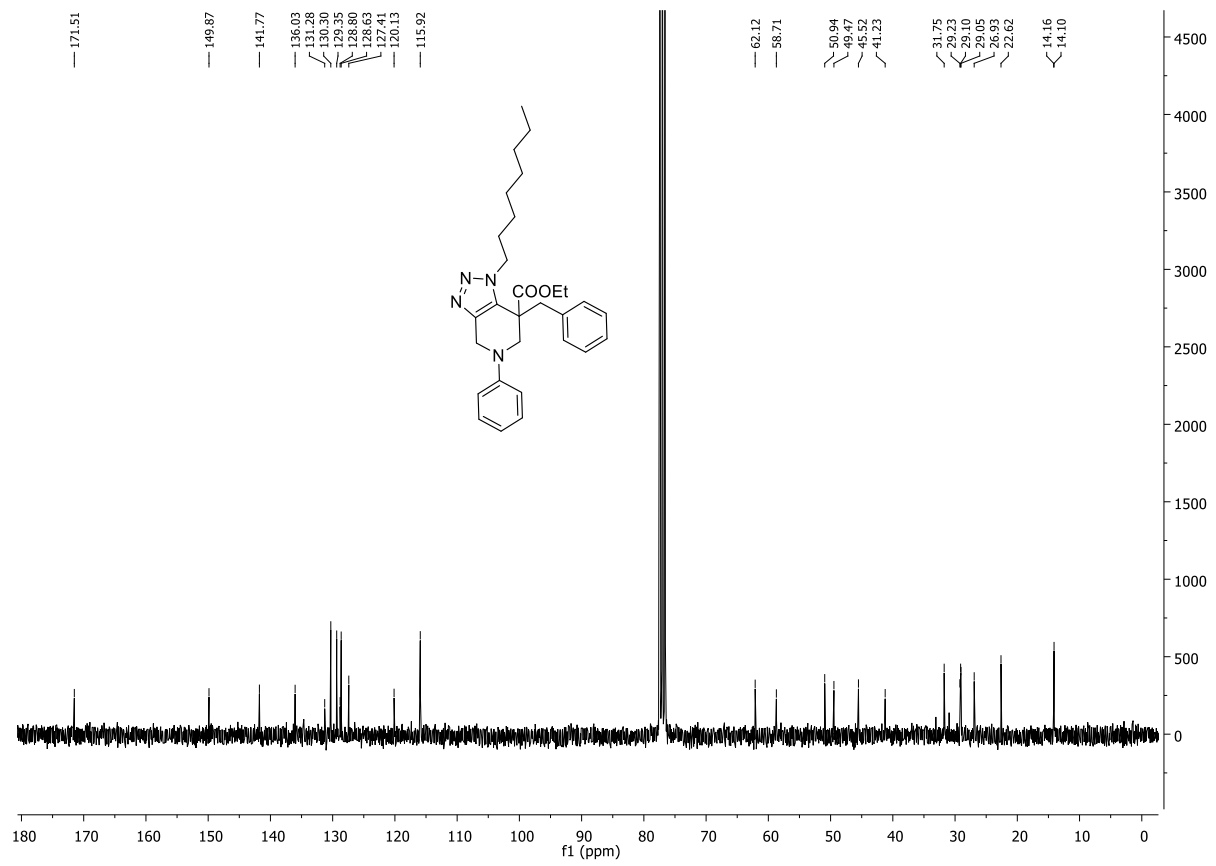
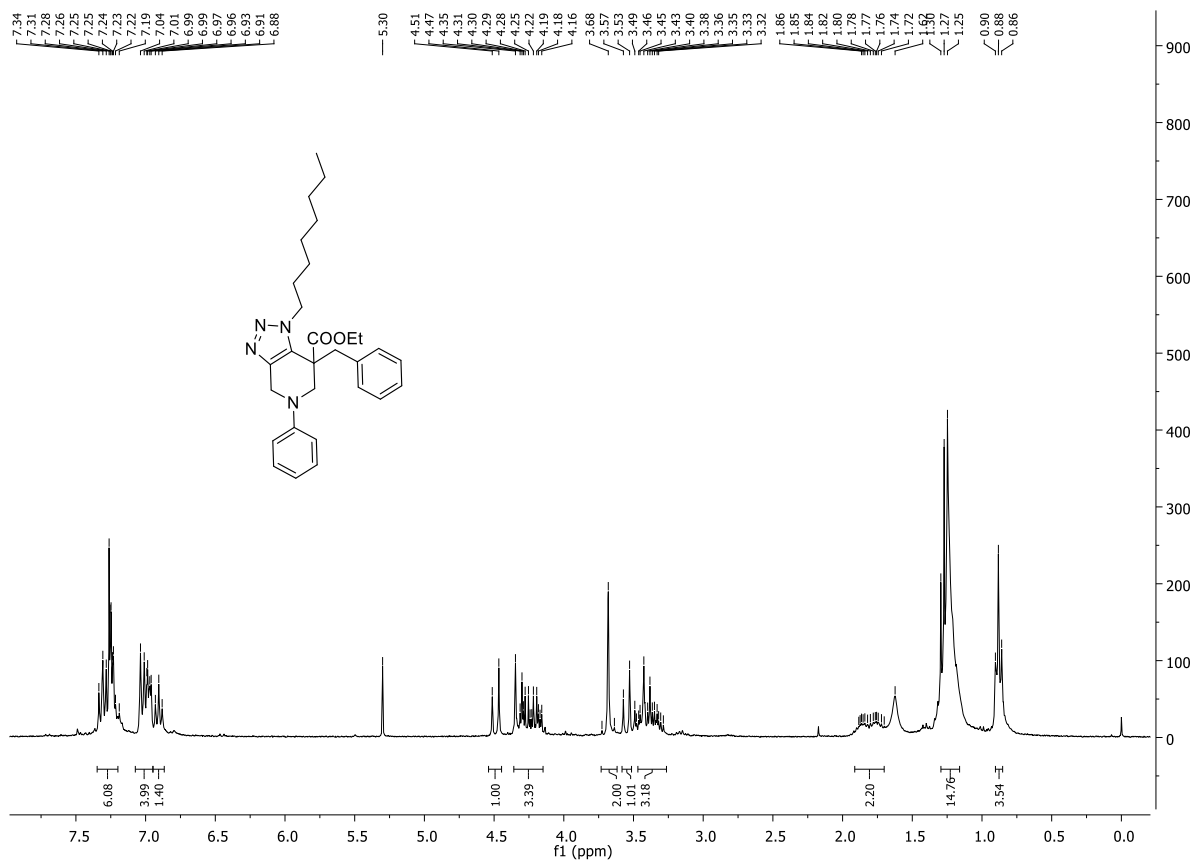


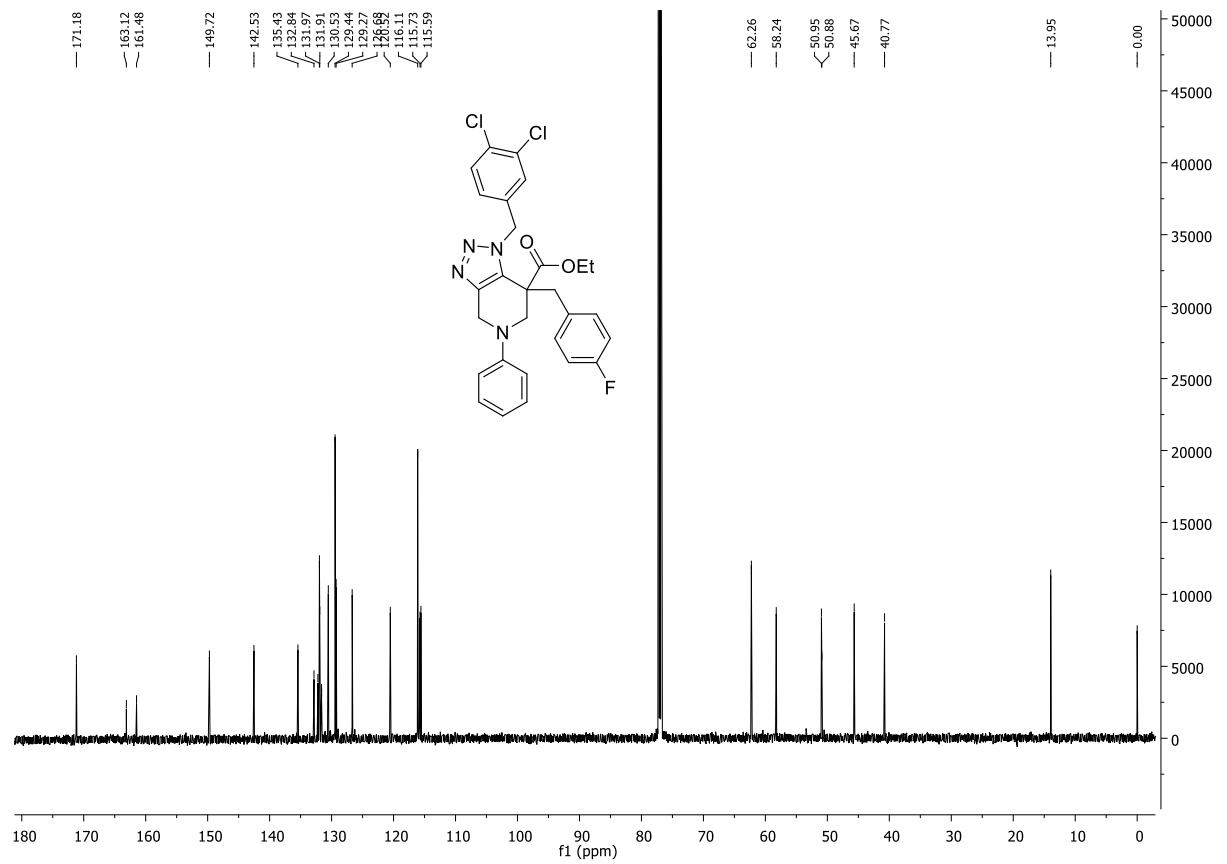
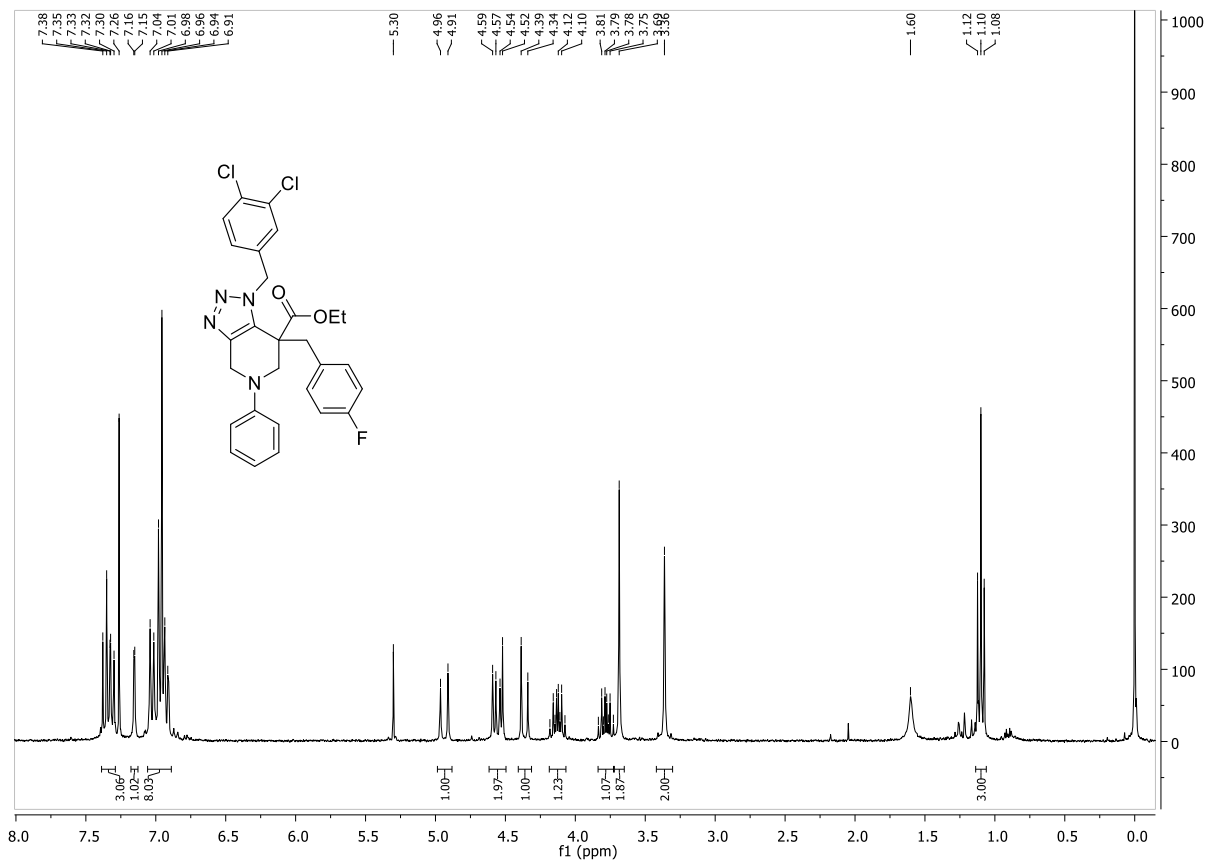


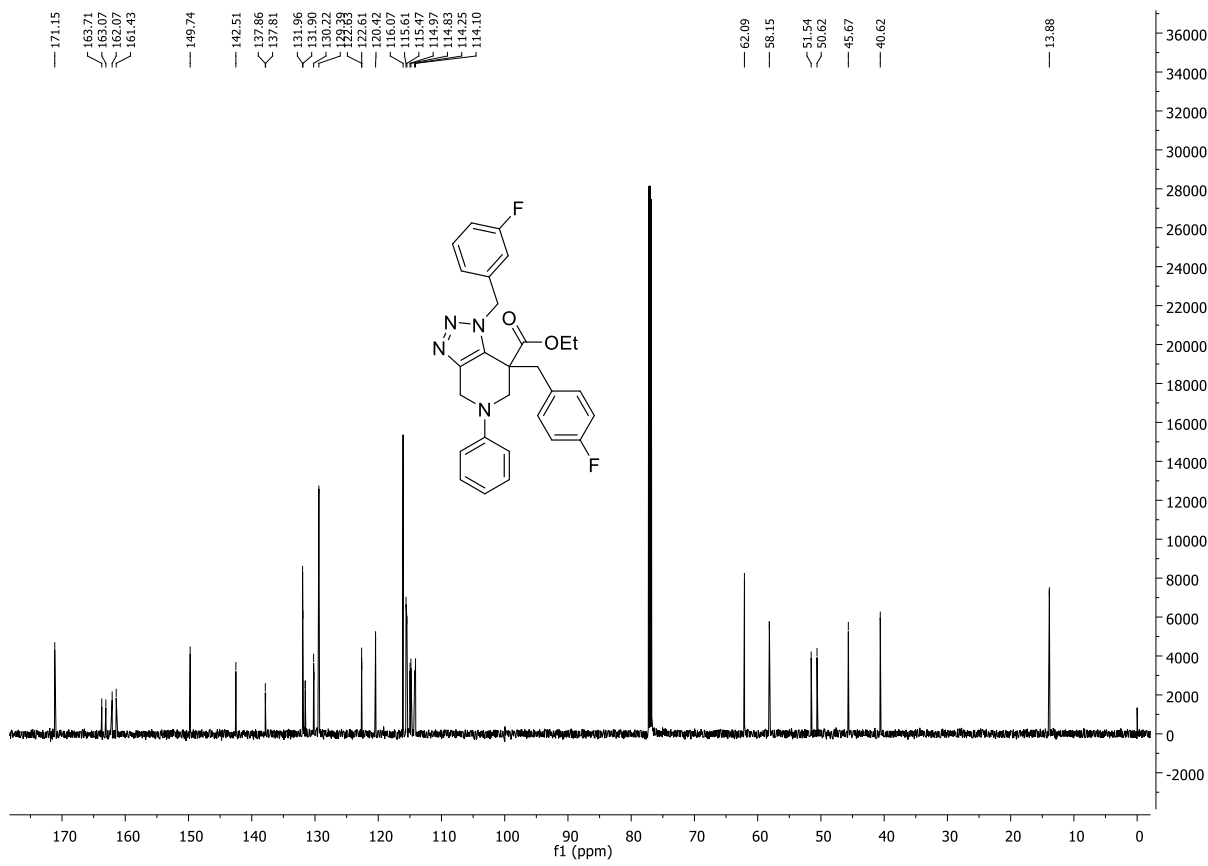
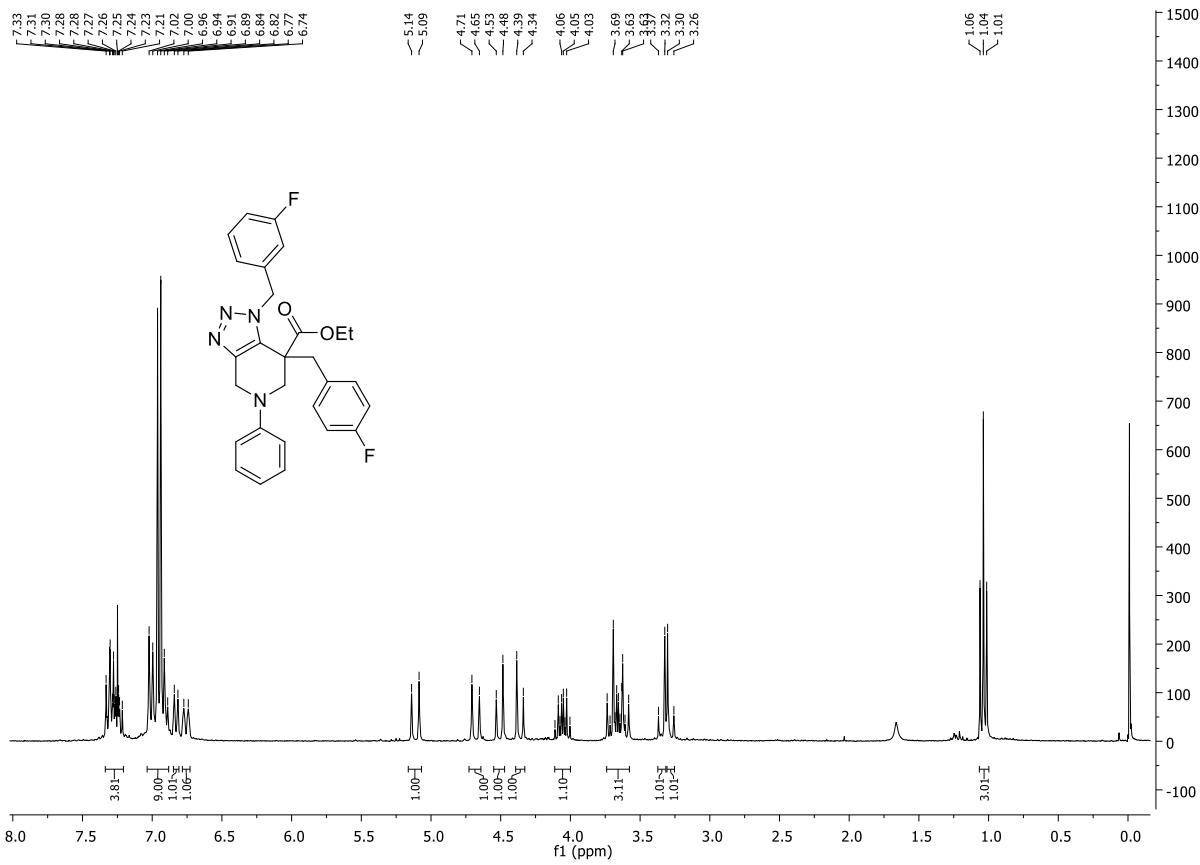


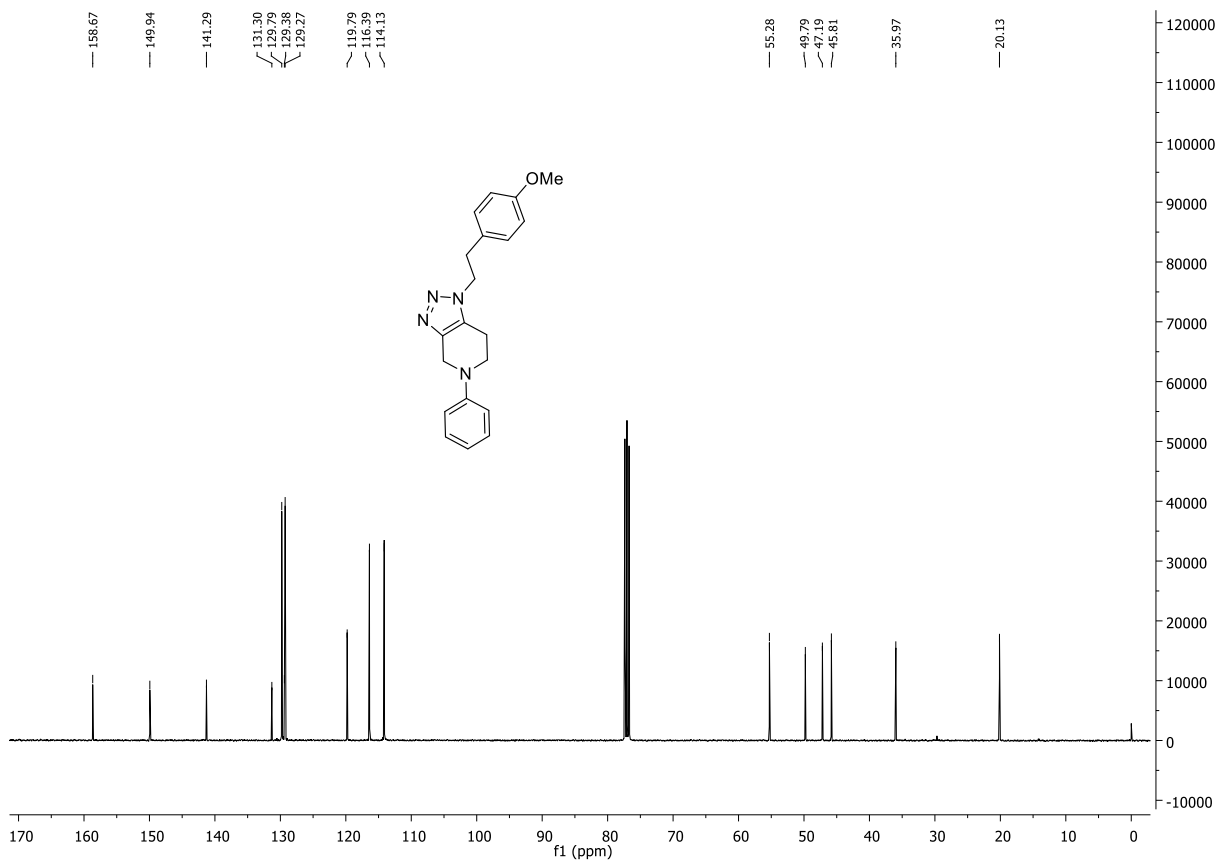
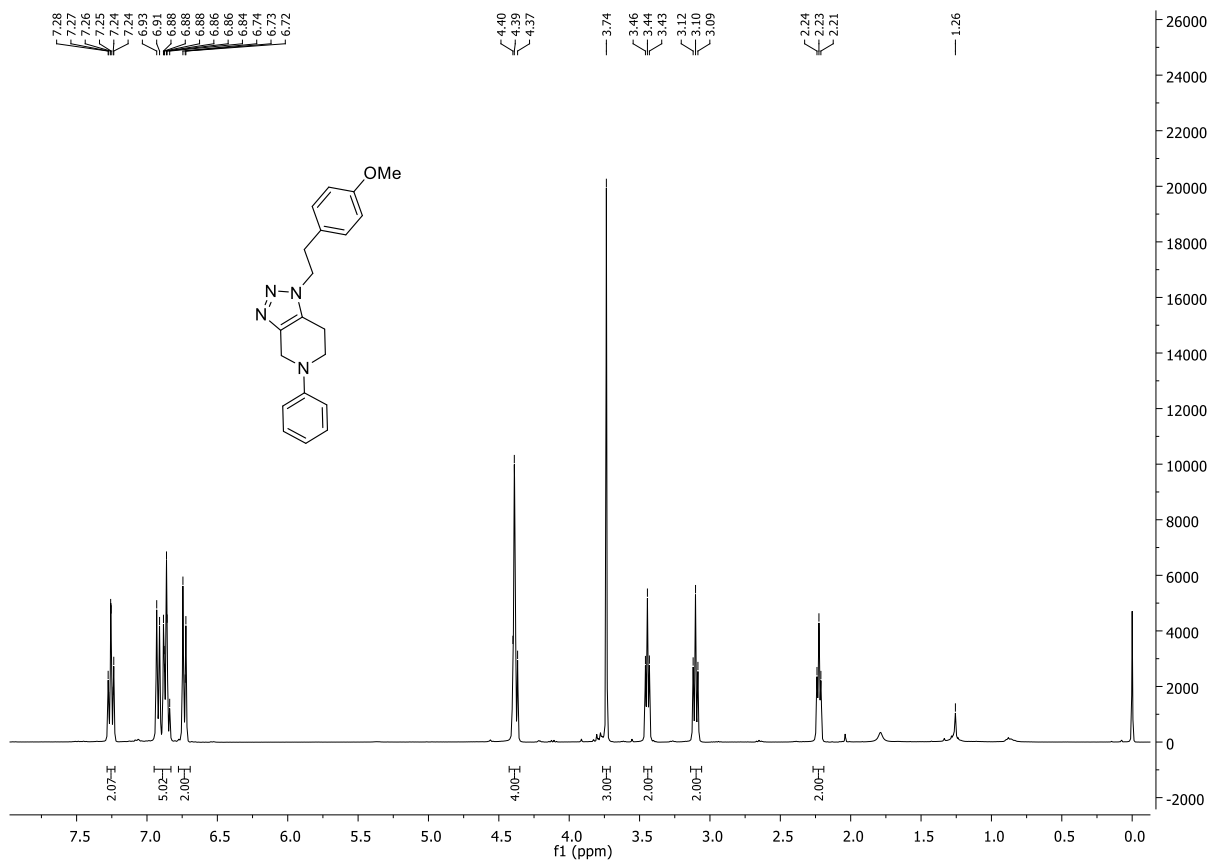


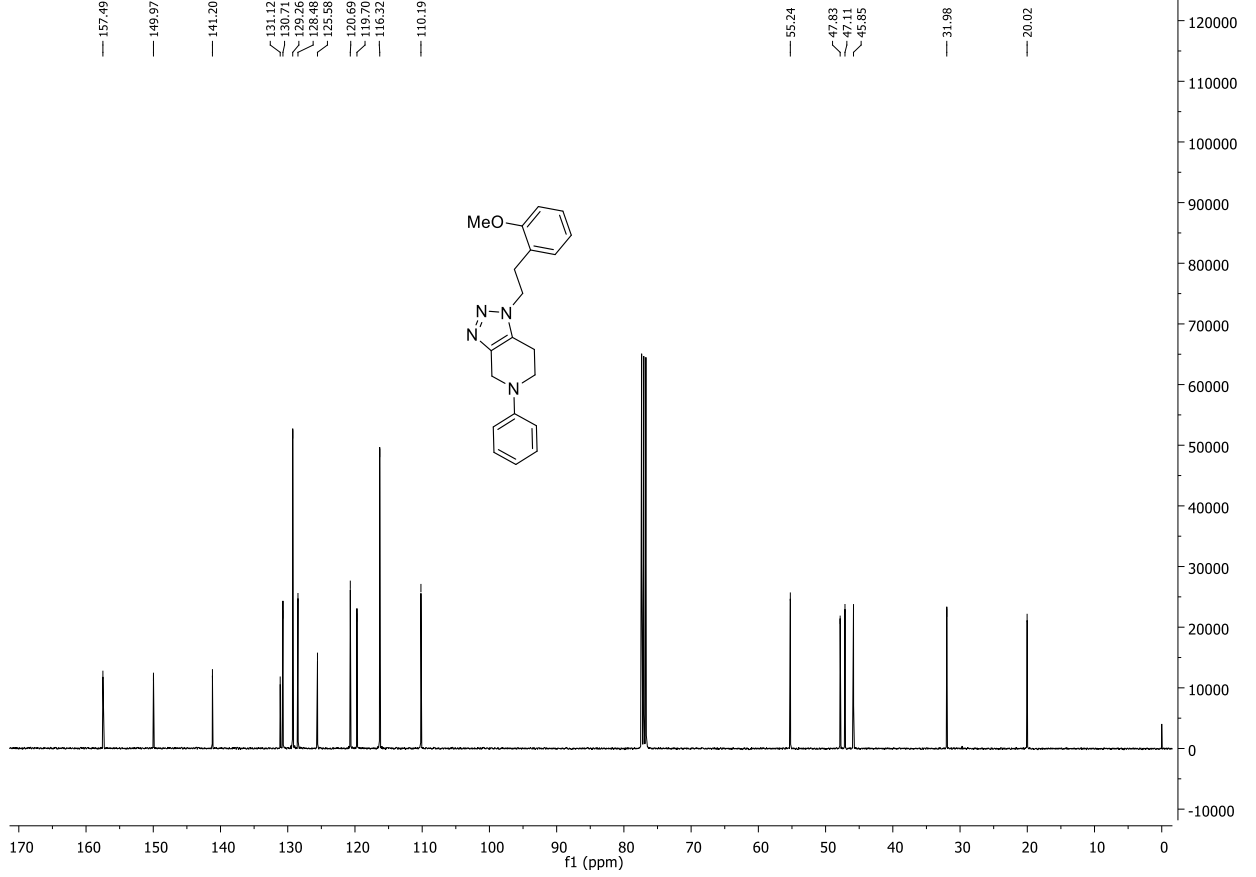
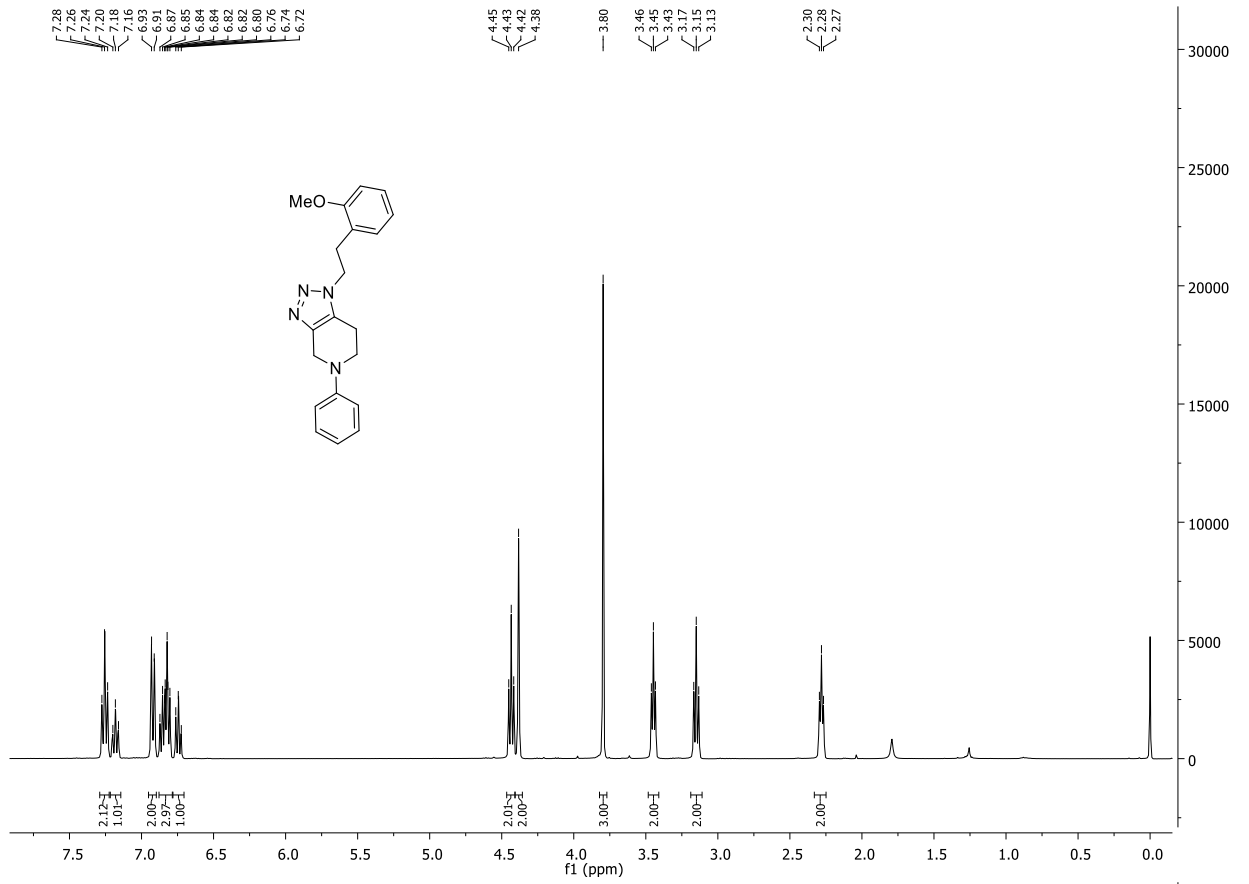


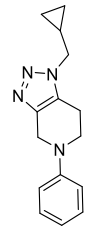
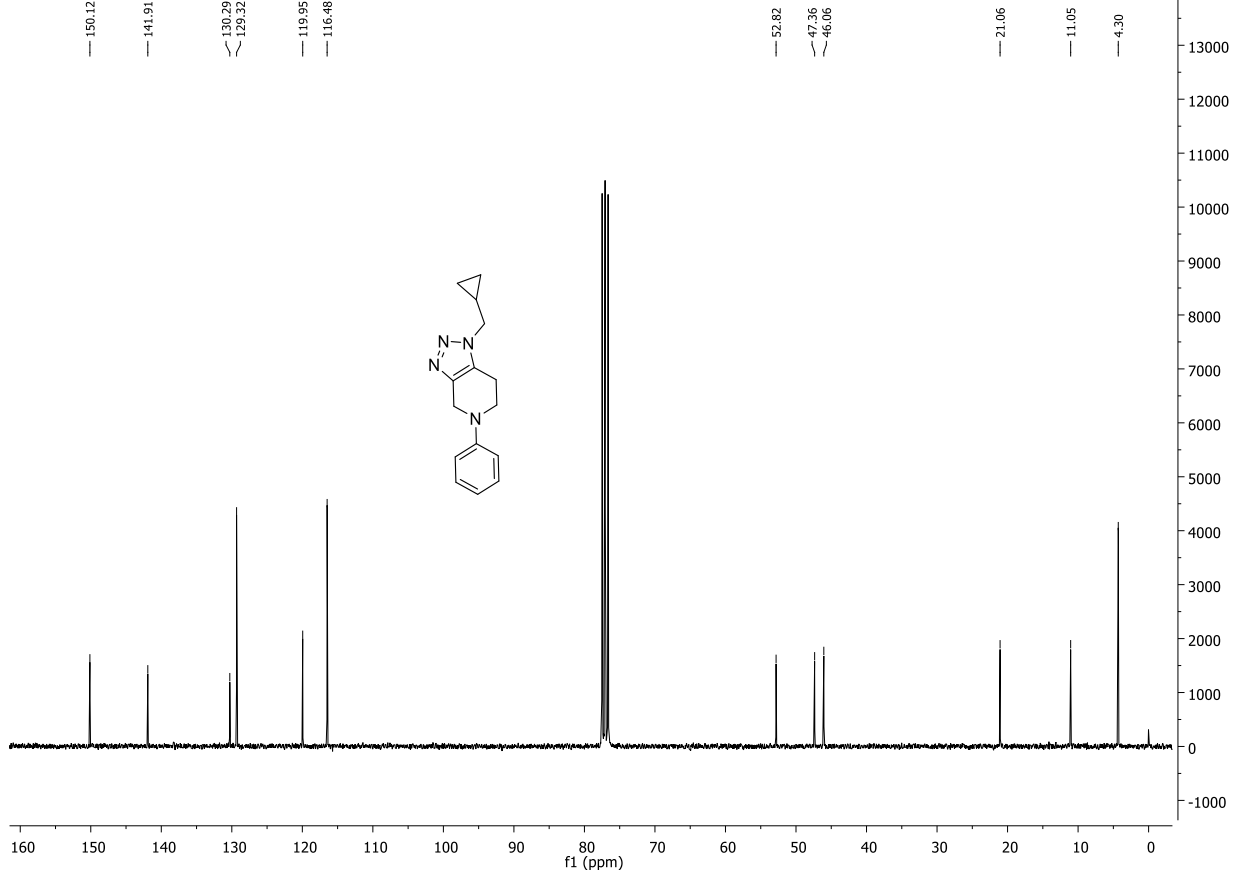
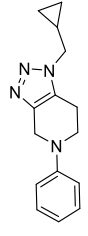
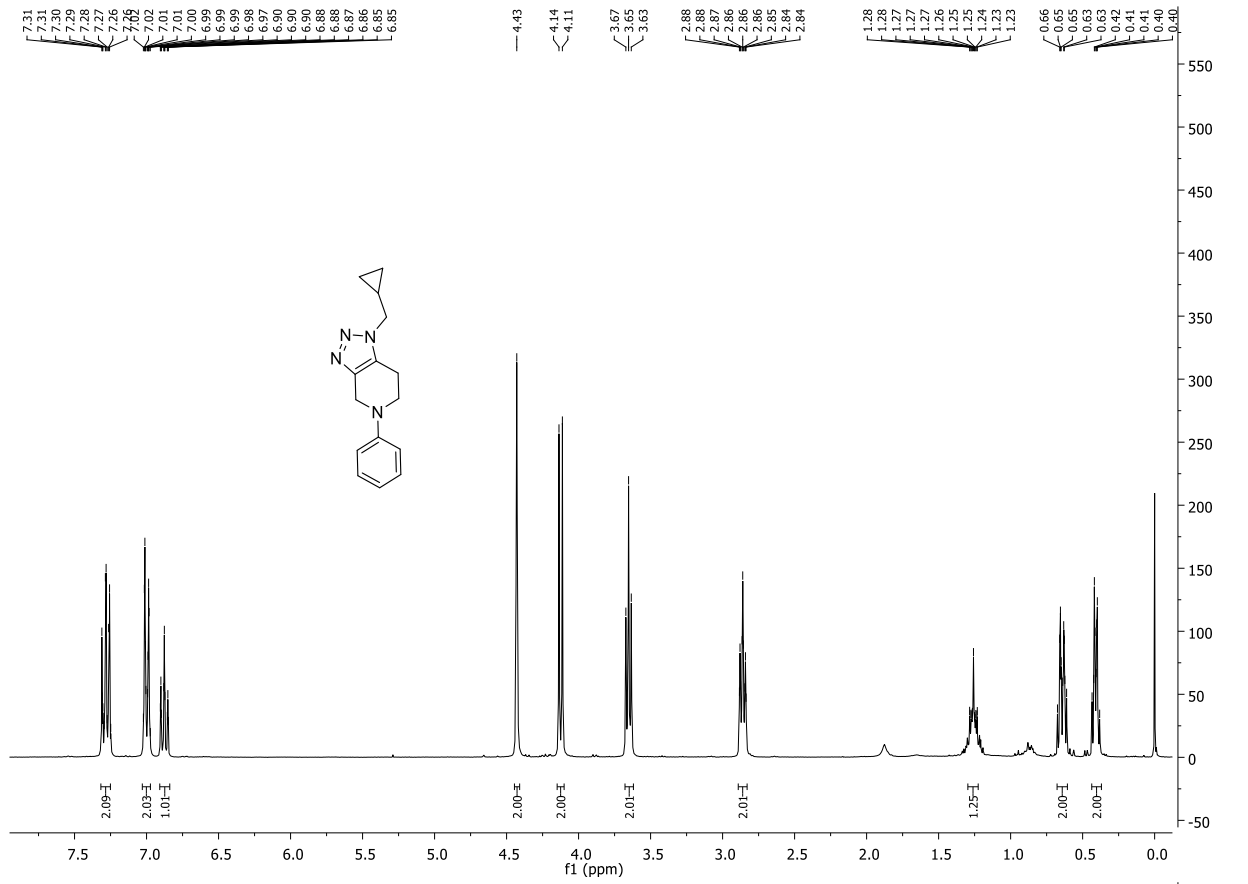


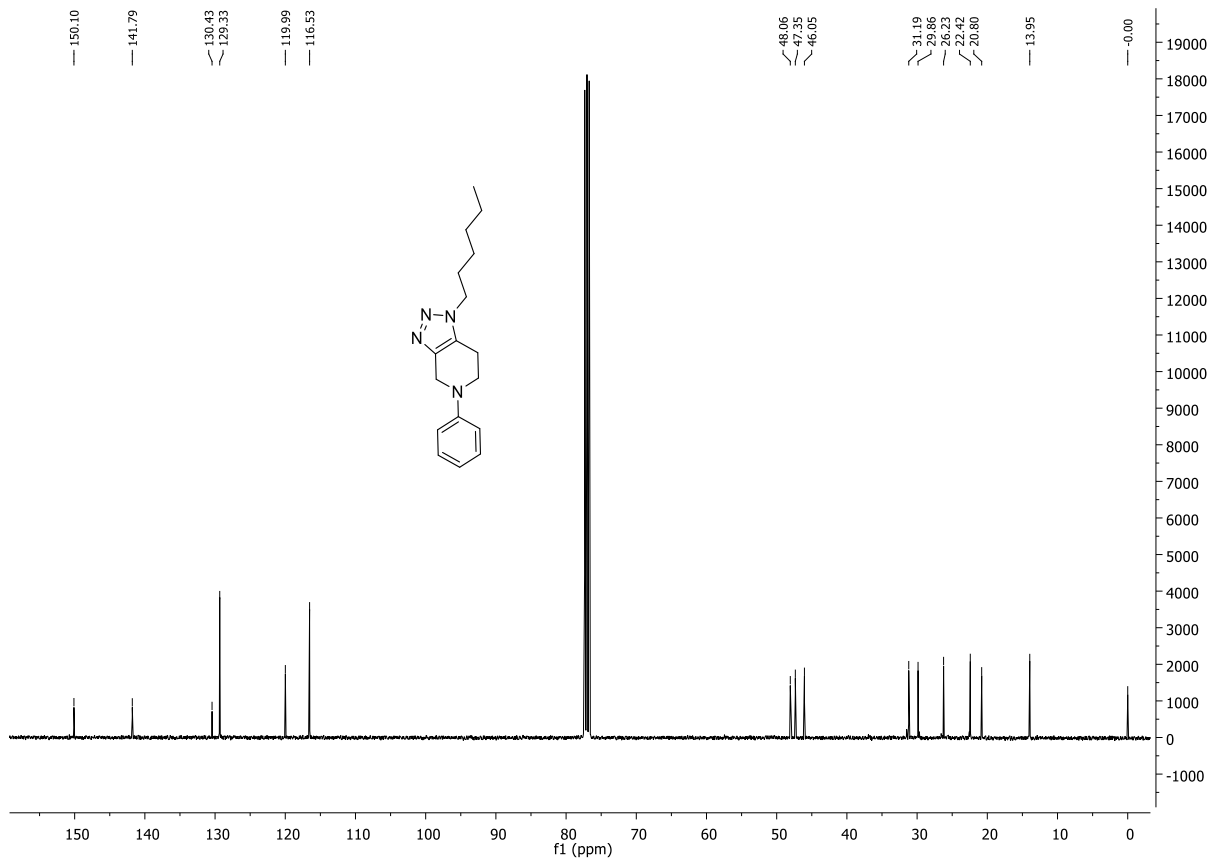
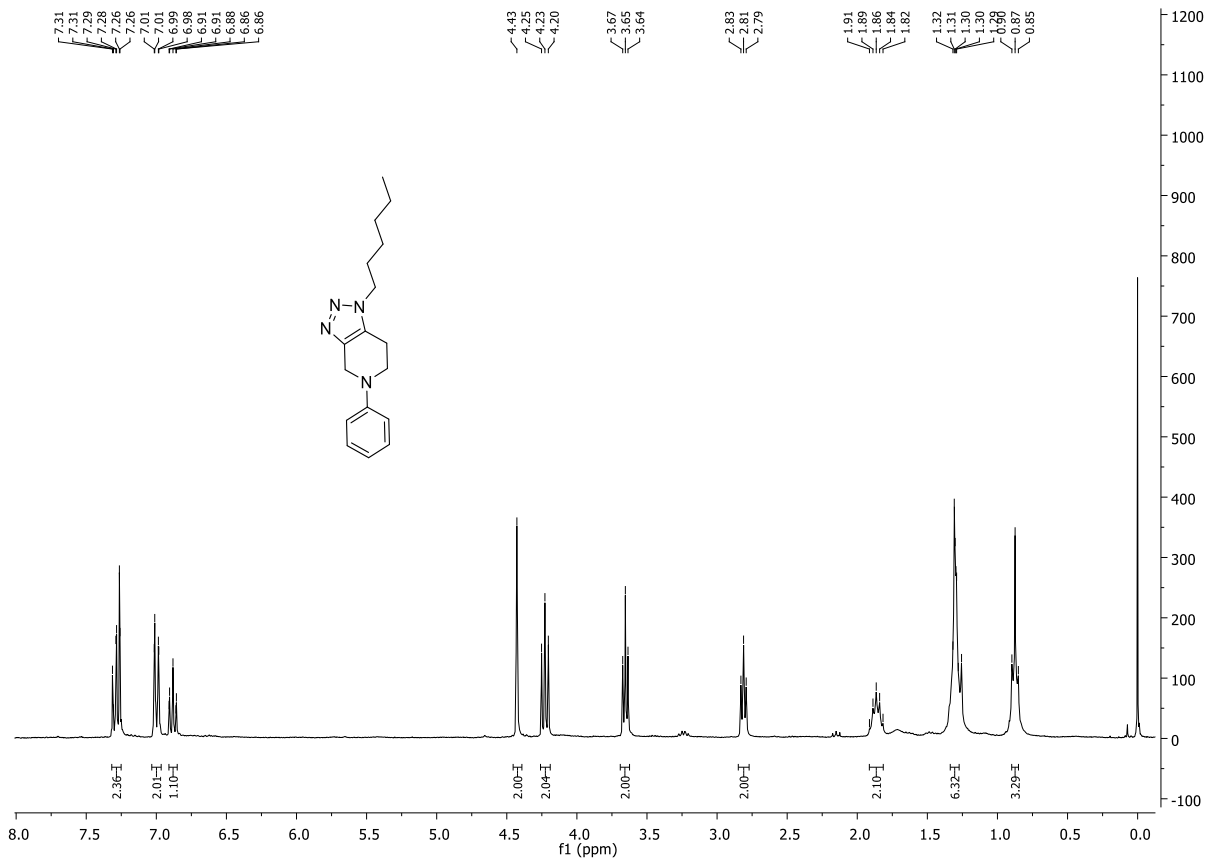


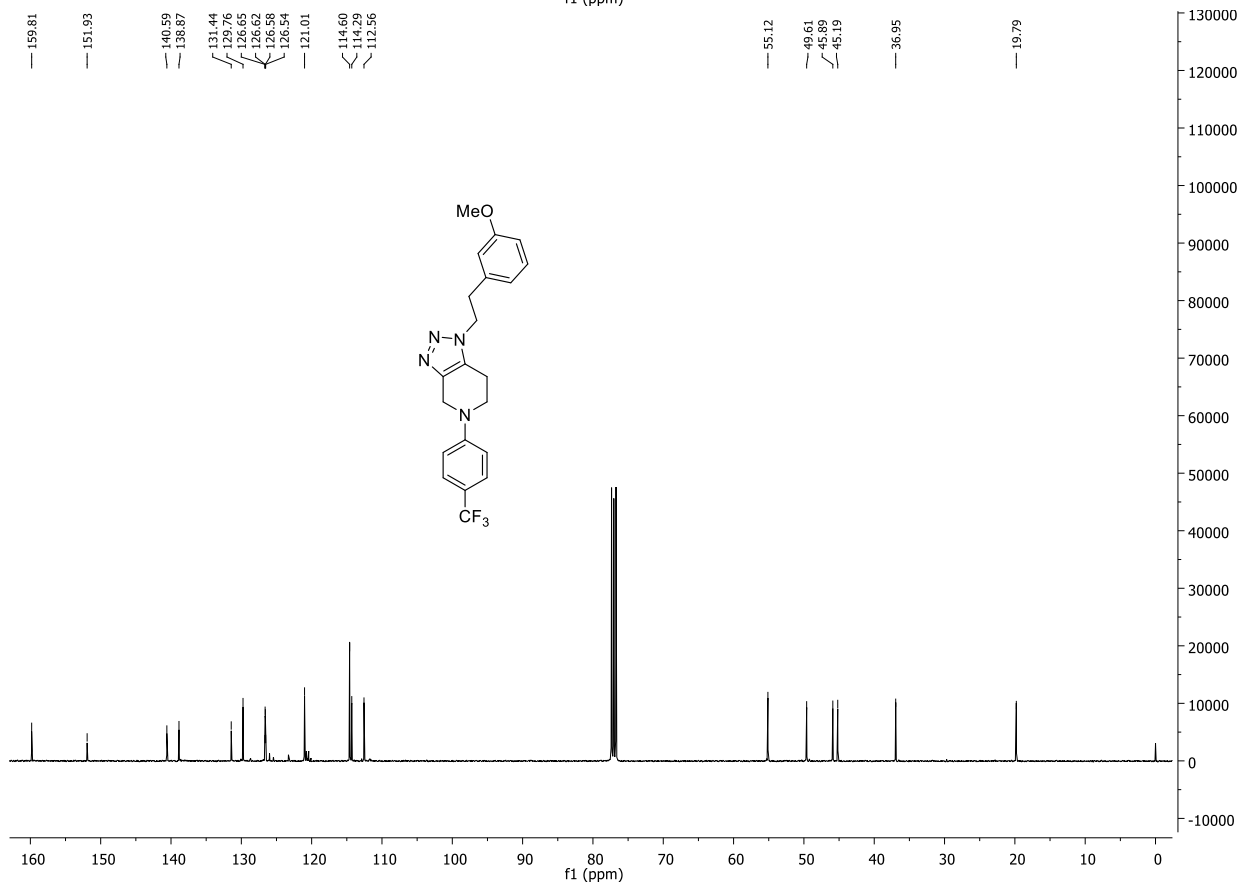
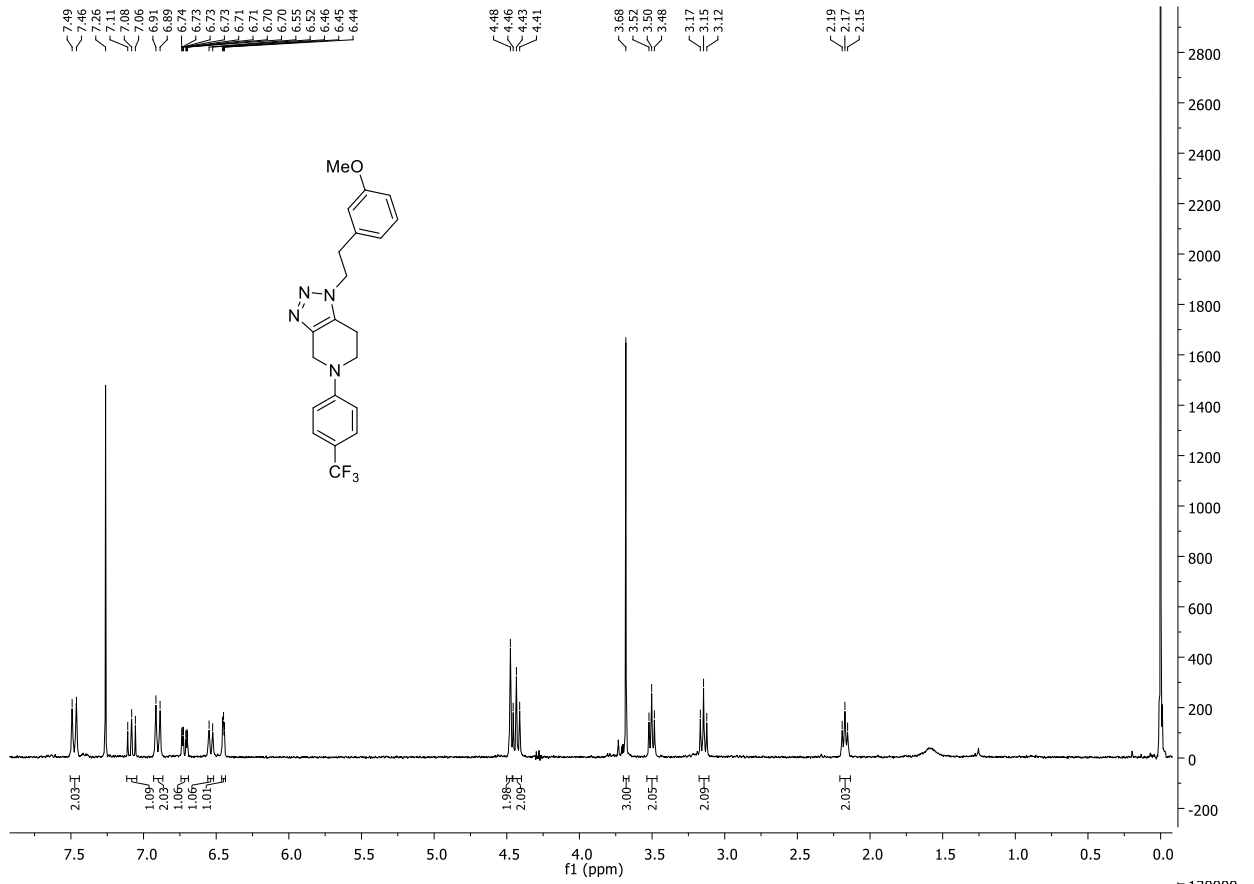


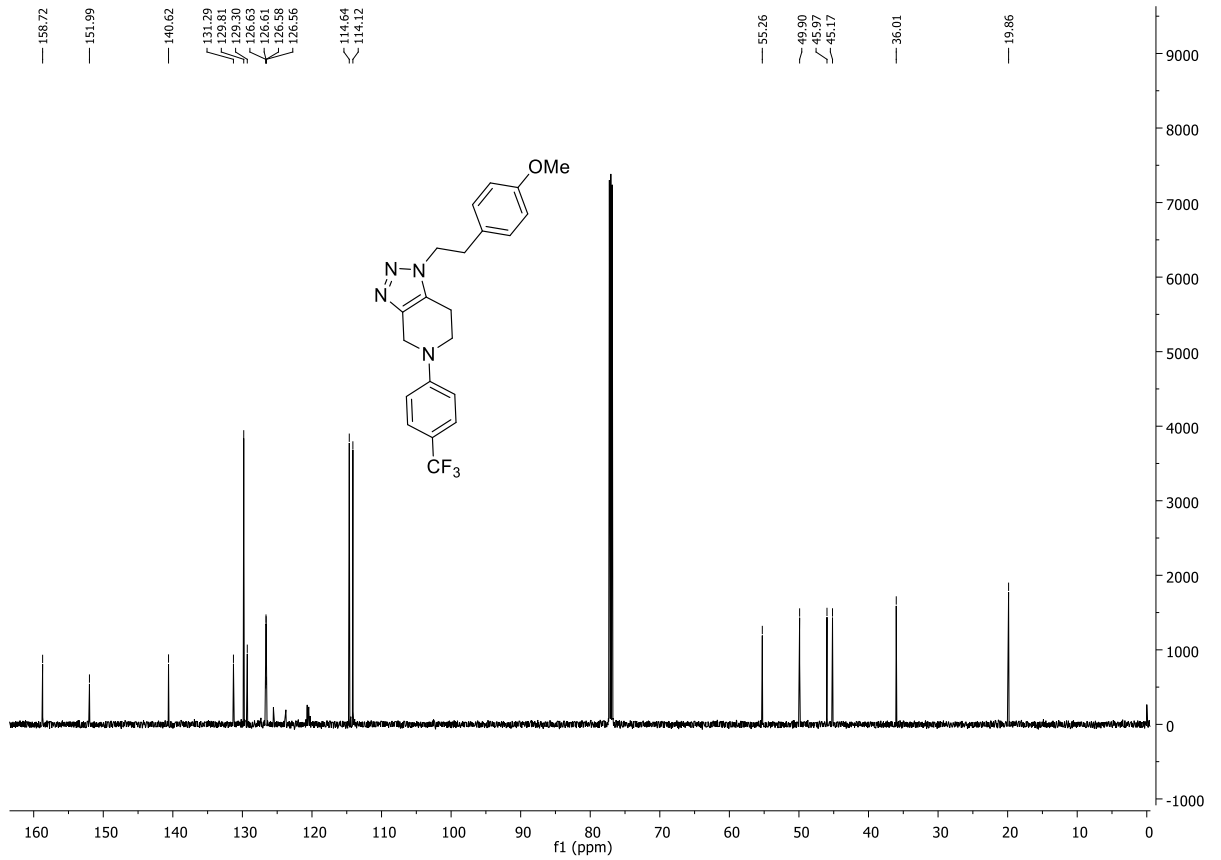
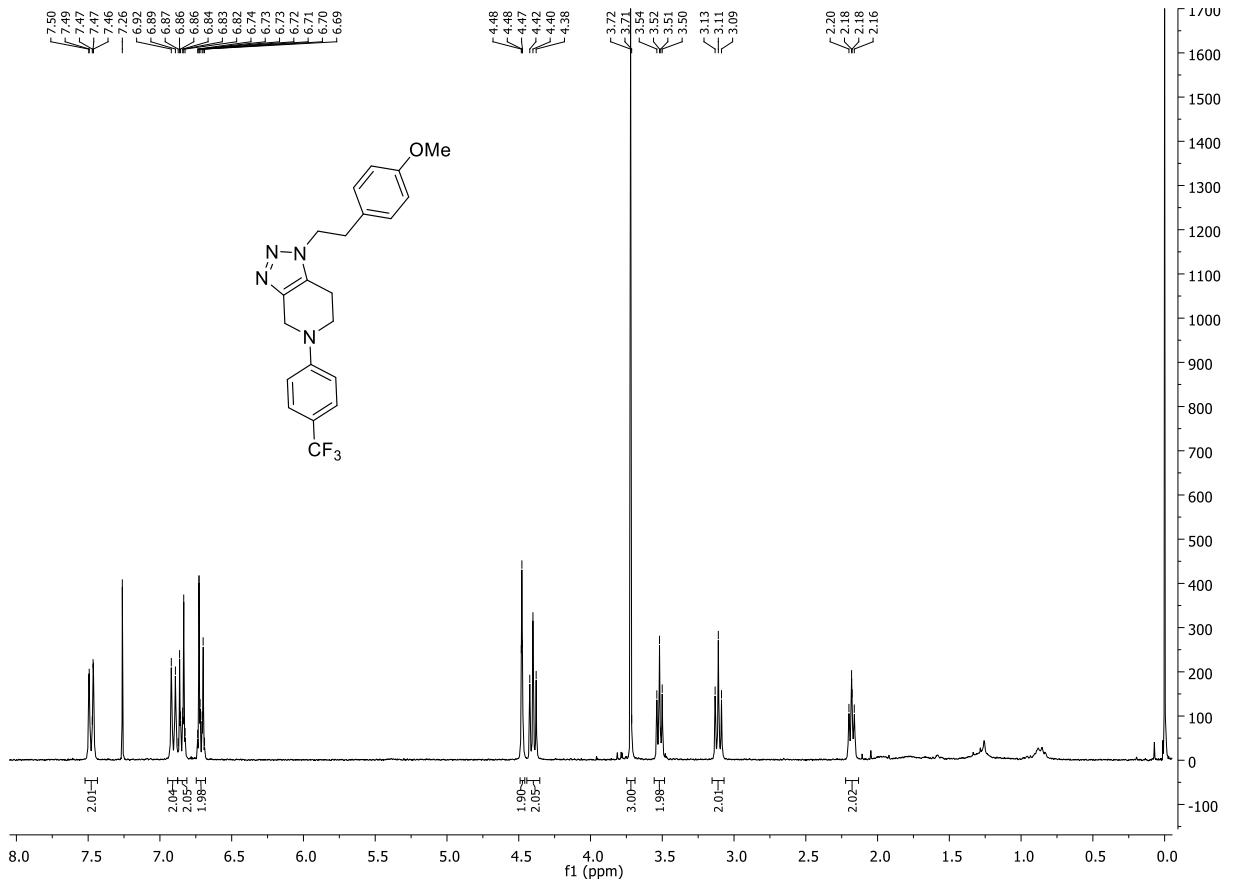


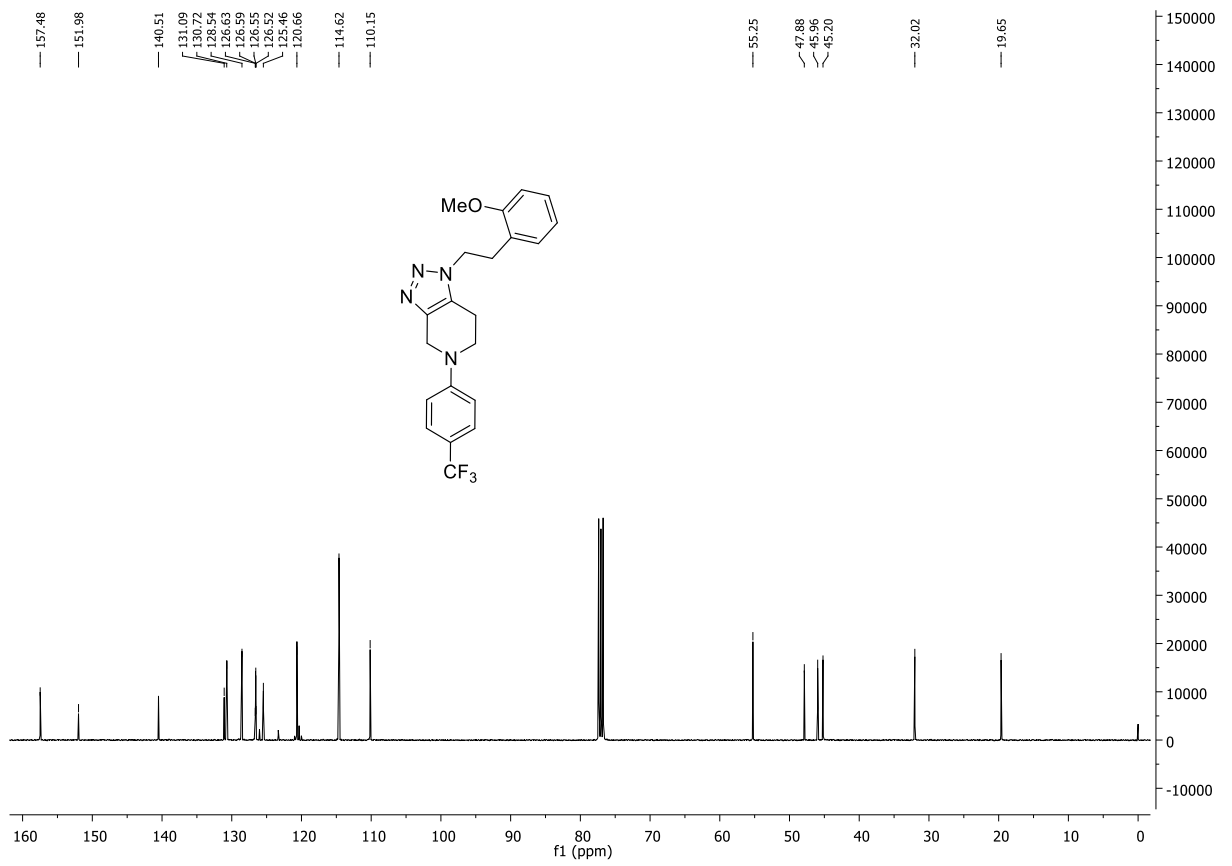
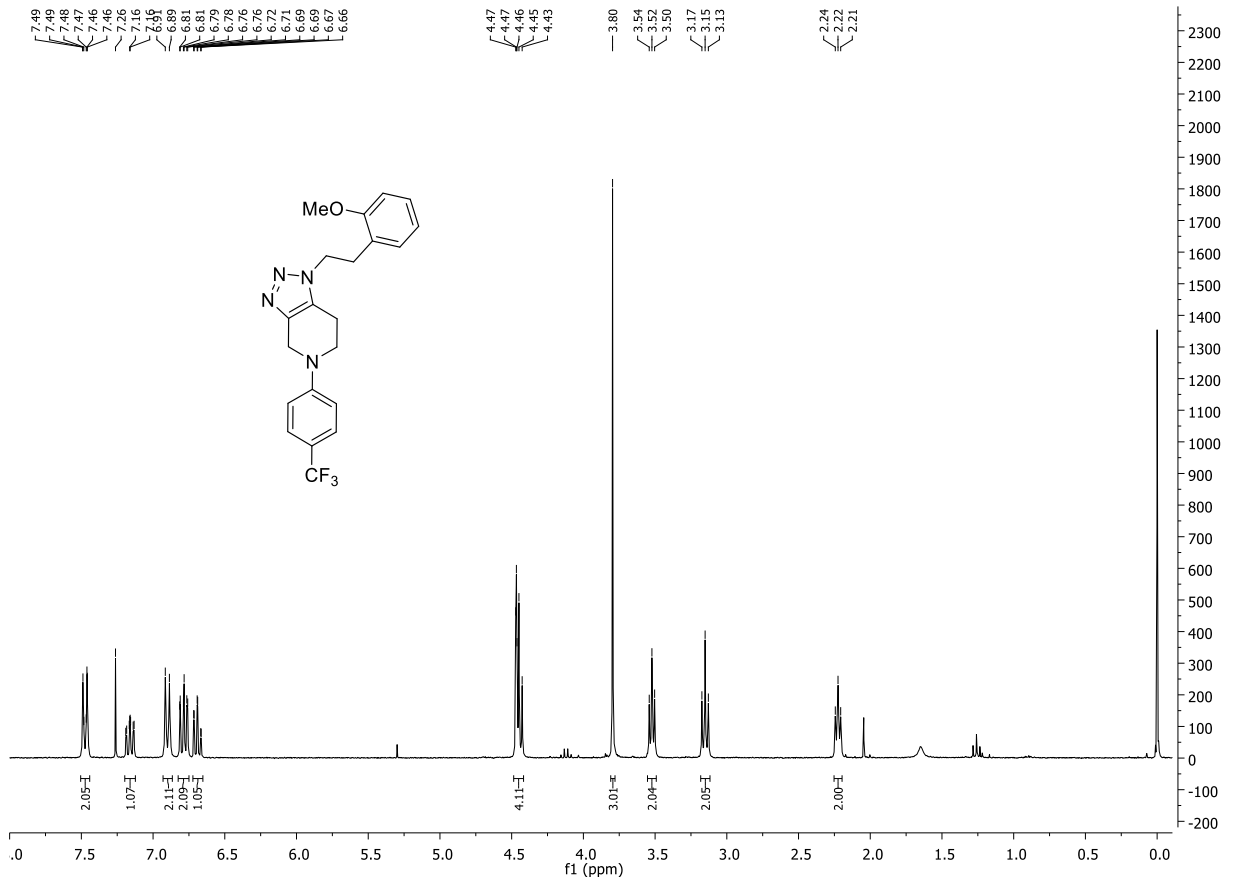


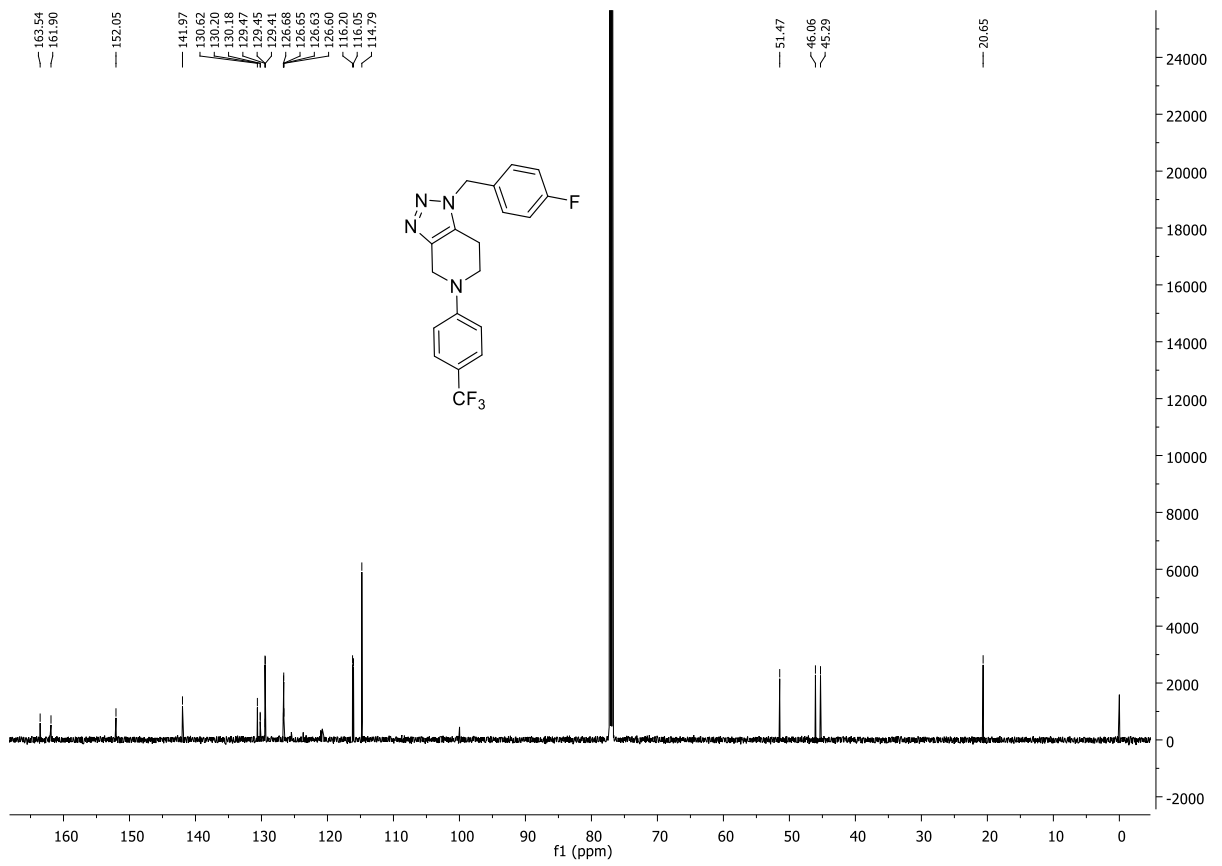
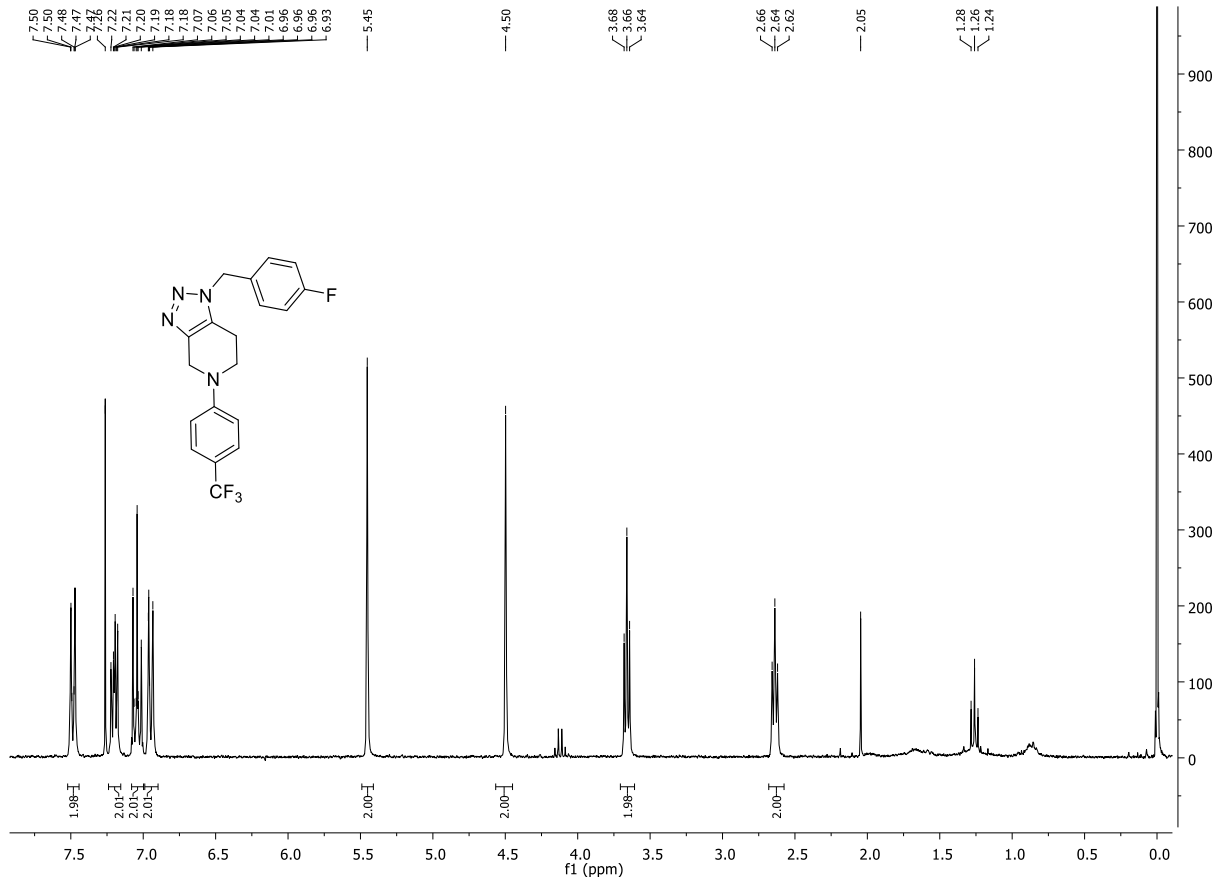


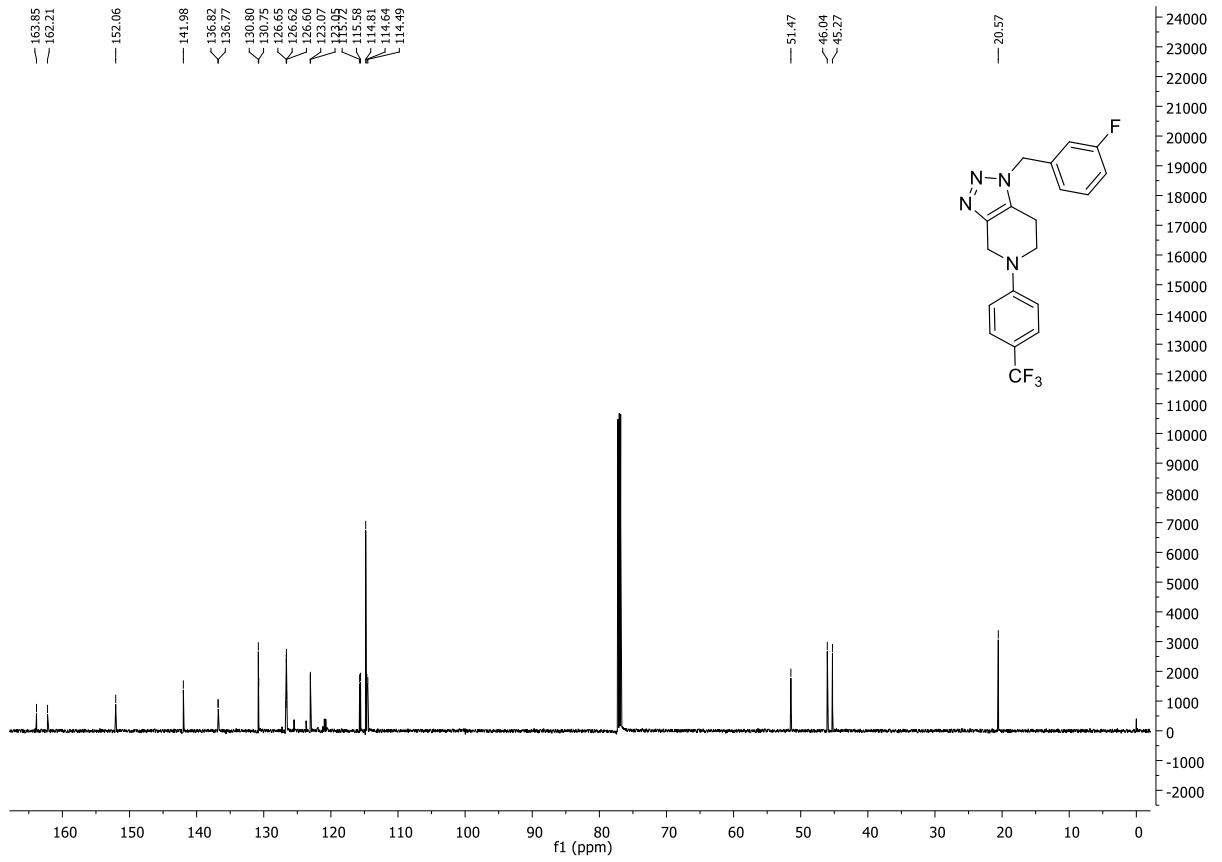
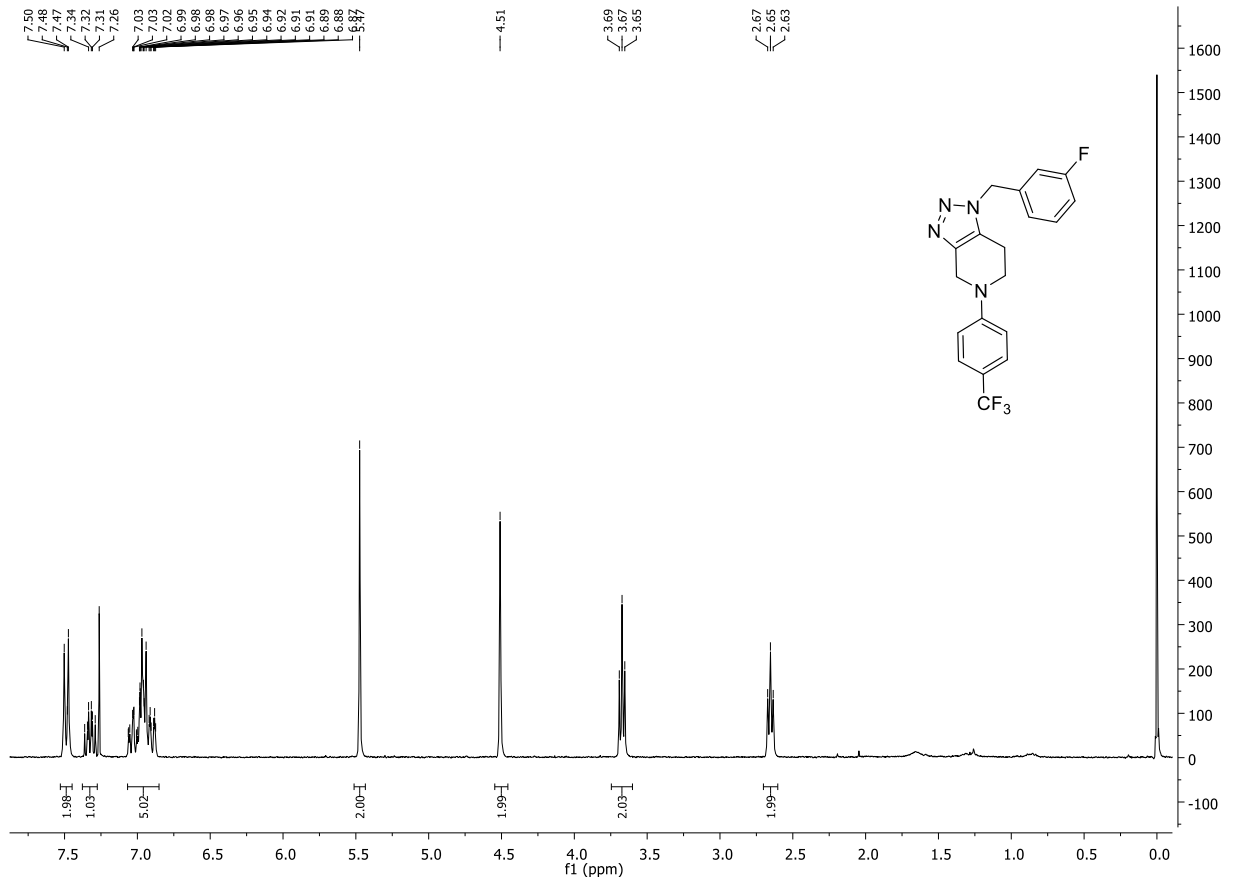


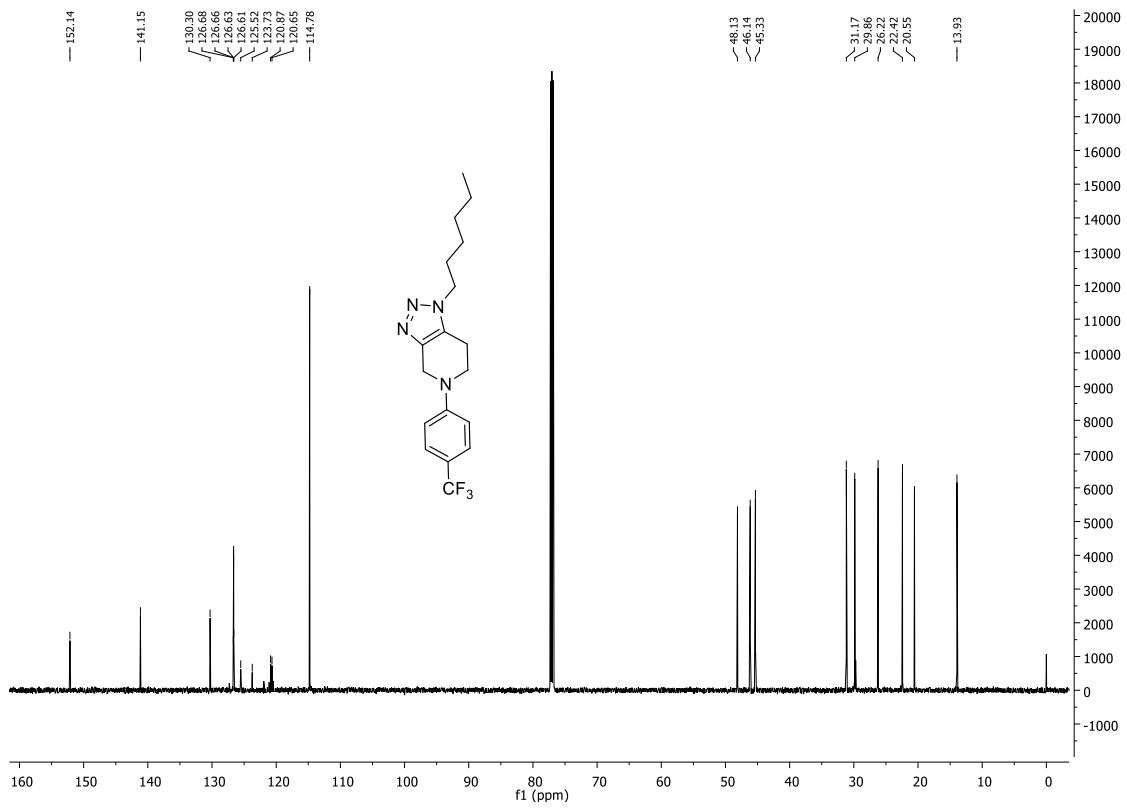
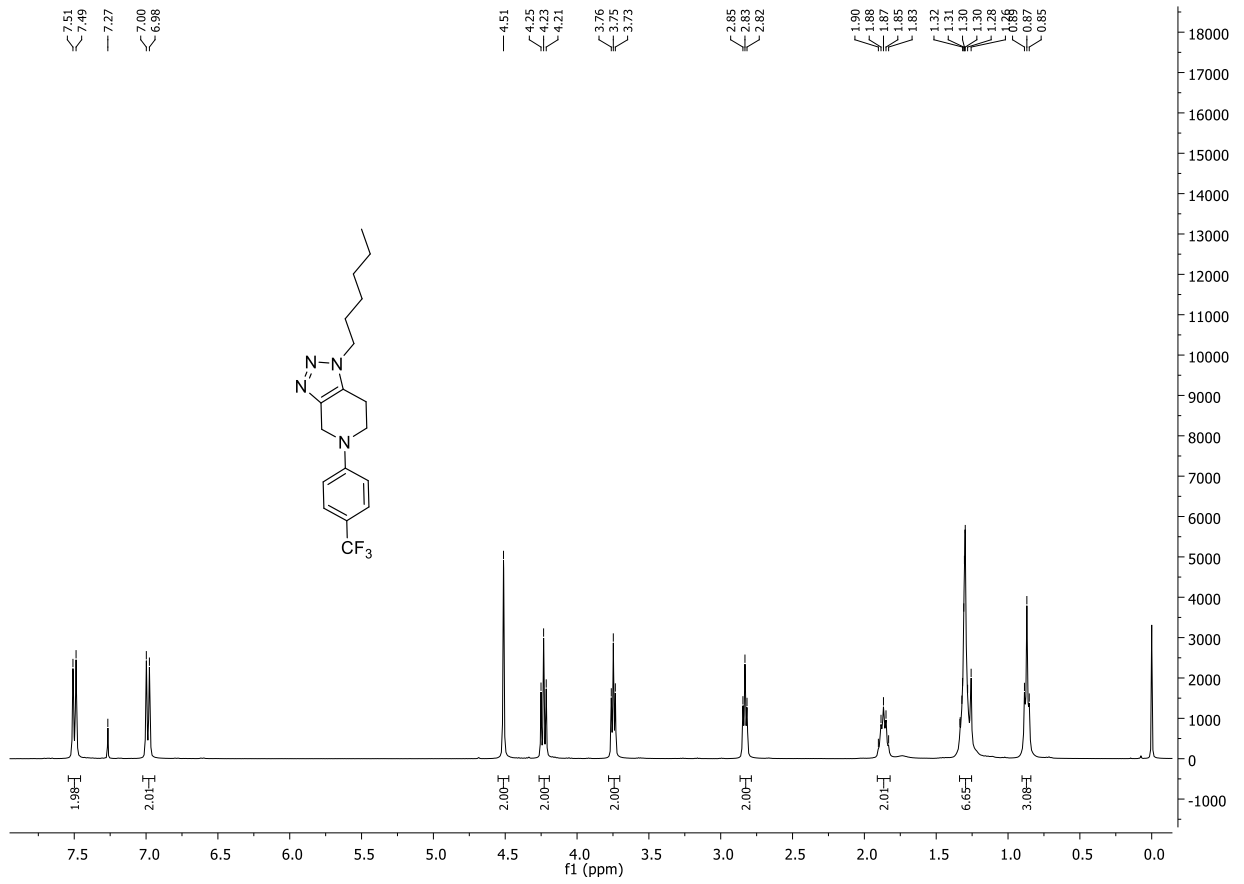


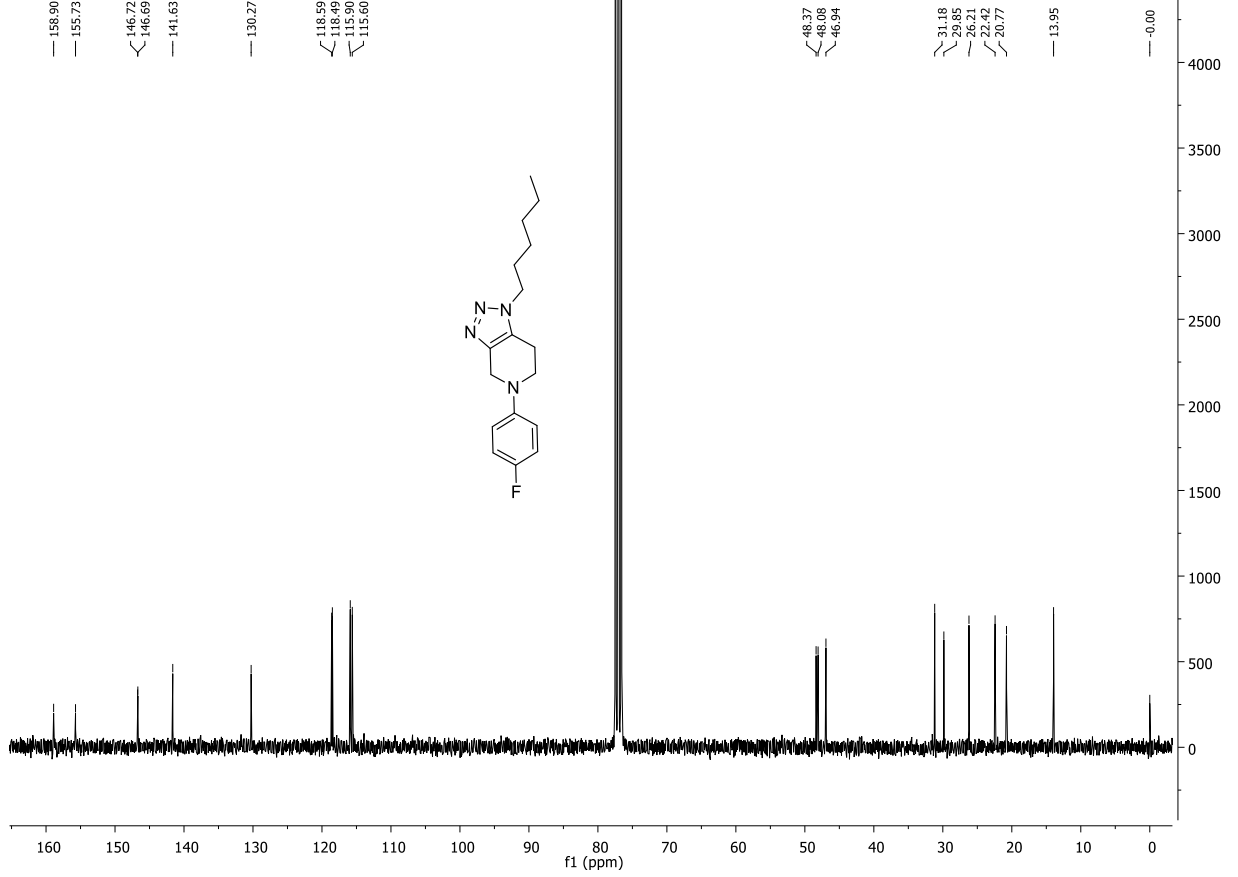
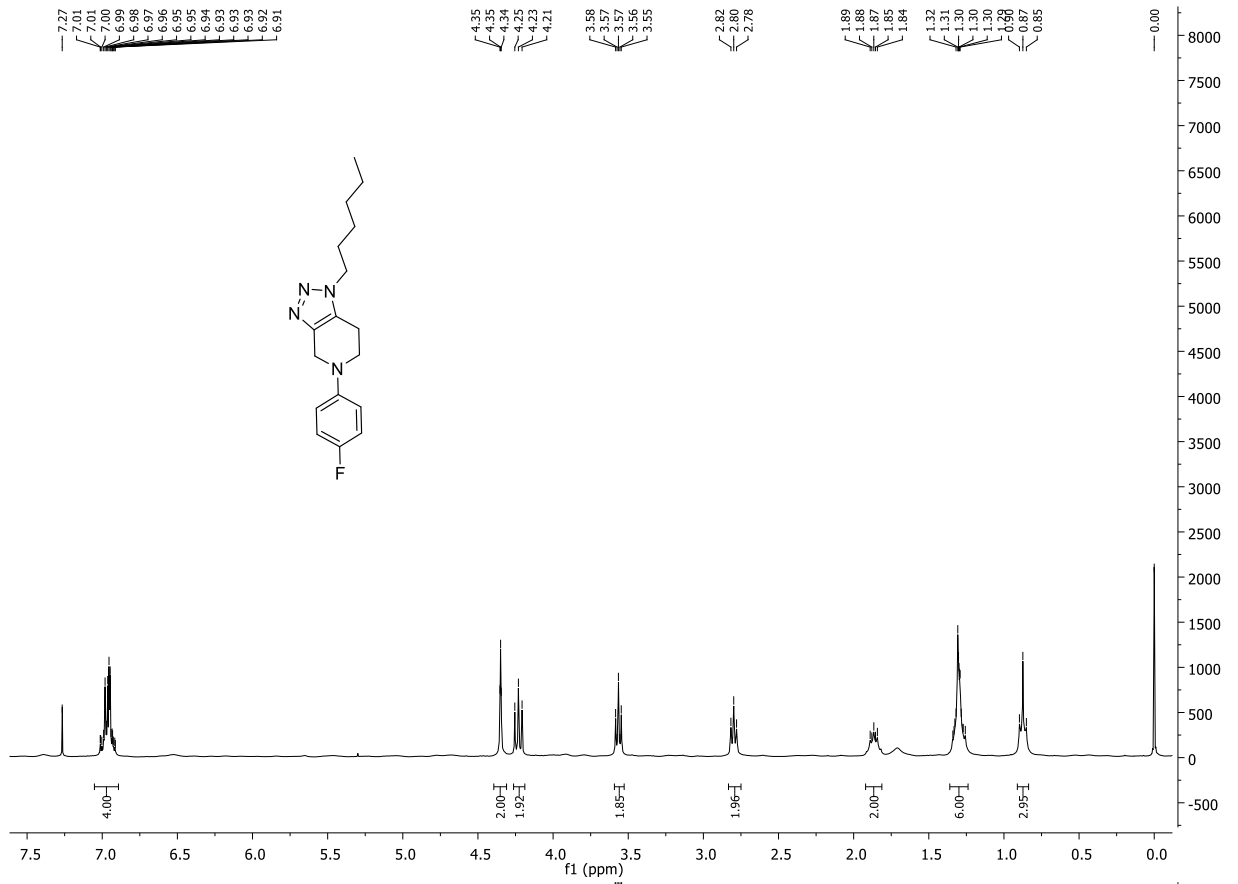


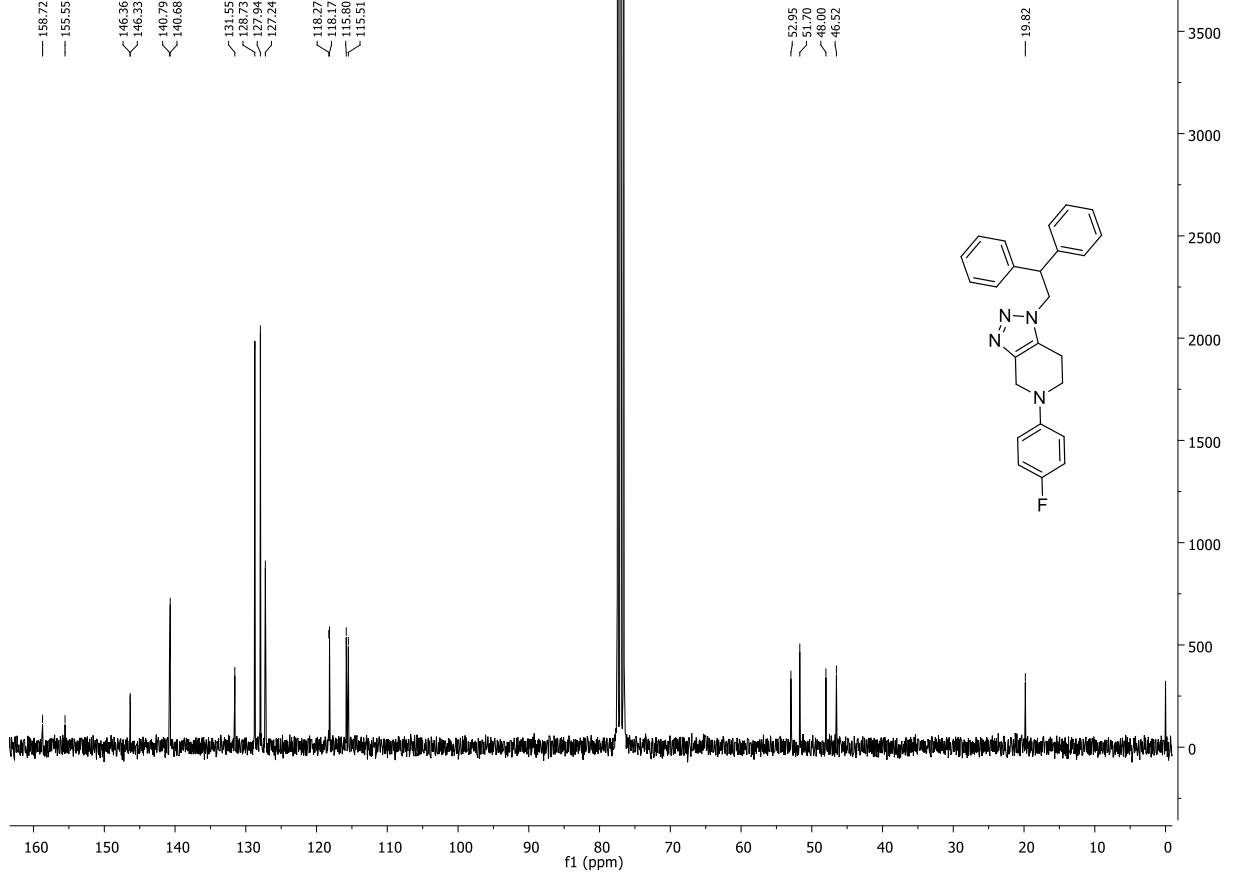
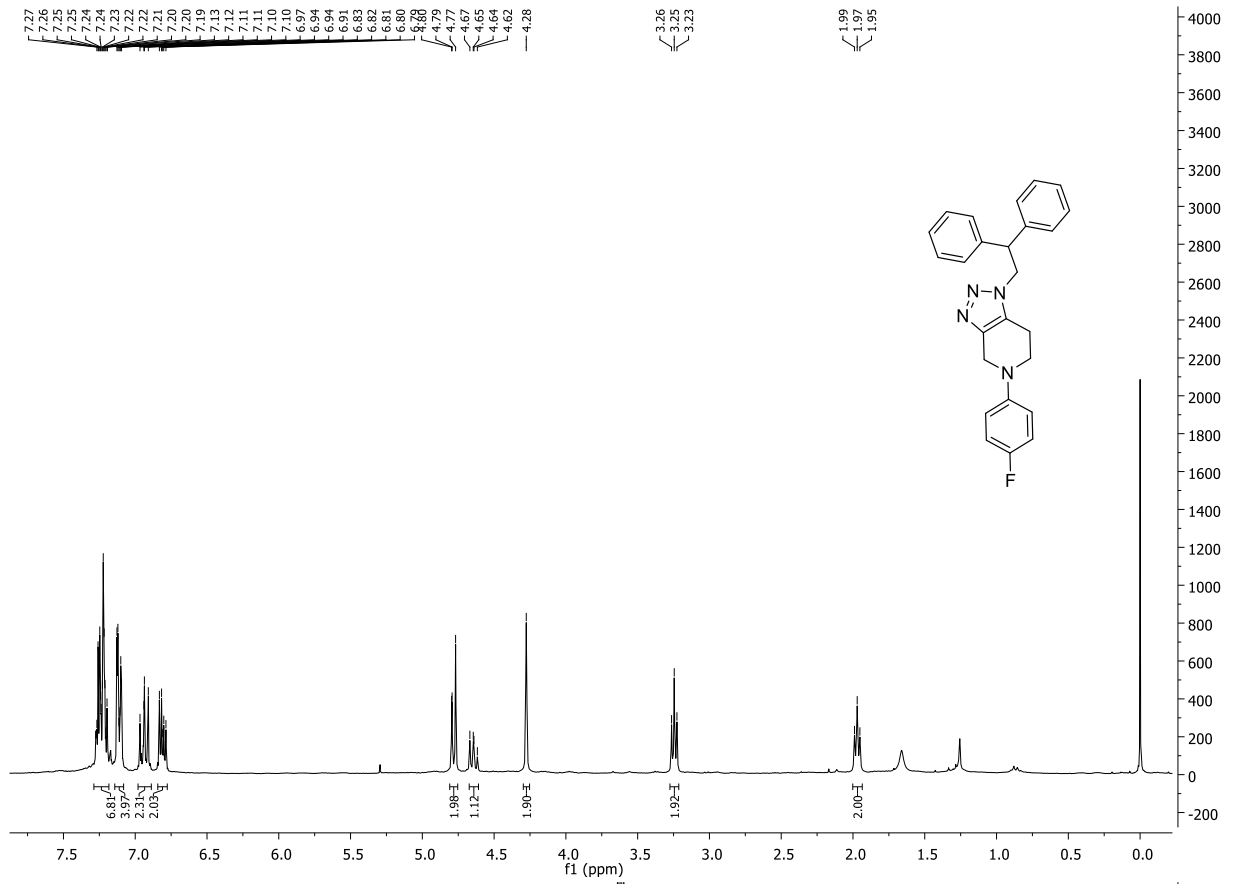












18e	μM	>100	>100	>100	>100	>100	16	45
18f	μM	>100	>100	>100	>100	>100	>100	8,9
18g	μM	>100	>100	>100	>100	>100	>100	>100
18h	μM	100	>100	>100	>100	>100	>100	>100
18i	μM	100	>100	>100	>100	>100	>100	11,95
18j	μM	>100	>100	>100	>100	>100	>100	>100
18k	μM	>100	>100	>100	>100	>100	>100	>100
18l	μM	≥100	>100	>100	>100	>100	>100	>100
Brivudin	μM	>250	0,01	250	0,1	3,8	-	-
Cidofovir	μM	>250	4,5	3,4	2,8	85	22	-
Acyclovir	μM	>250	0,6	0,6	2	>250	-	-
Ganciclovir	μM	>100	0,01	0,01	0,2	>100	-	-
Zalcitabine	μM	>250	-	-	-	-	50	-
Alovudine	μM	>250	-	-	-	-	22	-
UDA	μg/ml	>100	-	-	-	-	-	1,8

^a Required to cause a microscopically detectable alteration of normal cell morphology.

^b Required to reduce virus-induced cytopathogenicity by 50 %.

Data indicating antiviral activity are shown in red font, and marked in yellow if the SI (ratio of MCC to EC50) is five or higher.

Note that the SI cannot be accurately calculated for compounds showing no cytotoxicity at the highest concentration tested (100μM).

1.2. Molecular modeling studies

The initial structures of the modeled structures were constructed using MarvinSketch v.16.2.15 developed by ChemAxon Ltd.⁴, energy minimized by applying a semi-empirical (AM1) and *ab initio* (HF/6-311 +G*) methods as implemented in the Gaussian03 software.⁵ Prior to docking analyses, all ligands were subjected to conformational enumeration using the OMEGA software developed by OpenEye Scientific Software,⁶ applying an energy threshold of 1k kcal/mol. The corresponding biological activity data for the training set was retrieved from the ChEMBL database (Doc IDs: ChEMBL2439951 and ChEMBL2311314).^{7,8}

In order to perform molecular docking studies, crystallographic structures of 3CL^{pro} in complex with the corresponding ligands were retrieved from Protein Data Bank (PDB codes: 3V3M and 4MDS, respectively). Ionization states of side chains in the receptor were assigned using the ProteinPlus Server, from the University of Hamburg.⁹ With the purpose of exhaustively exploring the surface energy landscape associated to the binding to the 3CL^{pro} catalytic site, all molecular docking assays were performed using identically parameterized receptor structures and two different docking engines: FRED3 and AutodockVina.¹⁰ In the first case, a fast rigid exhaustive docking approach was used based on the ligands conformers libraries previously generated, and ranking the ligands bound poses using the ChemGauss3 scoring function. When using AutodockVina, individual runs were performed for each ligand conformer, ranking the

resulting docked poses using the AutoDock built-in scoring function. In all cases, the cubic search space was placed on the geometrical center of ligand bound to 3CL^{pro} (pdb code: 3V3M or 4MDS), with the length, width and height being set to extend 30% from the bound ligand. In order to identify the lowest energy binding mode for each ligand, a free energy of binding analysis was performed on the corresponding docked structures obtained by using both software packages, applying the molecular mechanics Poisson-Boltzmann surface area (MMPBSA) approach as implemented in the MMPBSA.py tool.¹¹ The lowest total interaction energy was extracted for further MD analysis. Intermolecular interaction analyses were performed on the resulting complexes using the VIDA¹² and LigPot+¹³ software packages.

Molecular dynamics simulations were performed using the Amber18 software package.¹⁴ Starting from the lowest energy binding modes predicted by molecular docking, the *ff14SB* and GAFF force fields implemented were used to parameterize 3CL^{pro} and the corresponding ligands, respectively.^{15,16} Complexes were solvated using a pre-equilibrated TIP3P explicit water model, applying a solvent box with boundaries at a minimum distance from the solute of 10 Å in each direction. After standard minimization procedures (5000 steps, first stage: solute restrained; 5000 steps, second stage: unrestrained system), the minimized complexes were heated under constant volume conditions from 0 to 300 K in a 0.5 ns timeframe, applying restraints on the solute. Next, the systems were equilibrated during 1 ns, after which the production phase under constant pressure and temperature conditions was performed for additional 10 ns. The trajectories were obtained applying restraints to the backbone of 3CL^{pro}. In all cases, a 2 fs time step was used, with the SHAKE algorithm being applied to constrain all covalent bonds involving hydrogen atoms. A 10 Å cutoff value was used to calculate non-bonded interactions. The cpptraj module of Amber16 was used to analyze the hydrogen bond interaction on the 3CL^{pro}-ligand complexes during the 10 ns of production stage.

MD trajectories were generated using CUDA designed code (*pmemd.cuda*), and computed using computational resources provided by the CCAD – Universidad Nacional de Córdoba (<http://ccad.unc.edu.ar/>). In particular, the Mendieta cluster was used which is part of SNCAD – MinCyT, República Argentina. In addition, GPU infrastructure was provided by NVIDIA Corporation, specifically through the donation of the Titan Xp GPU used for this research.

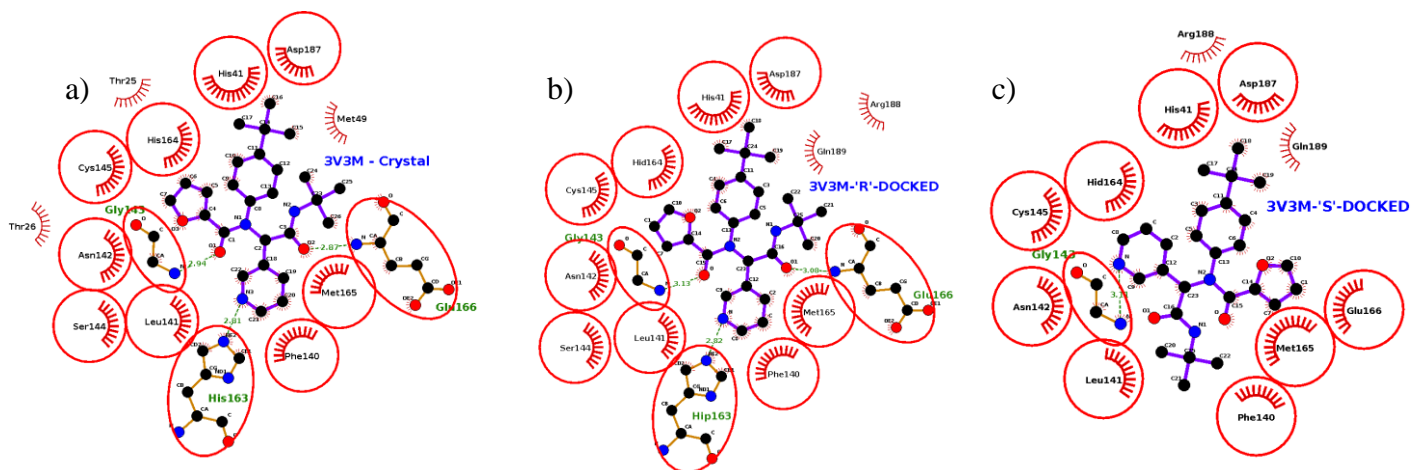


Figure S1. Intermolecular interactions observed in a) the crystallographic structure deposited under pdb code: 3V3M; b) docked position of the R enantiomer; c) docked position of the S enantiomer.

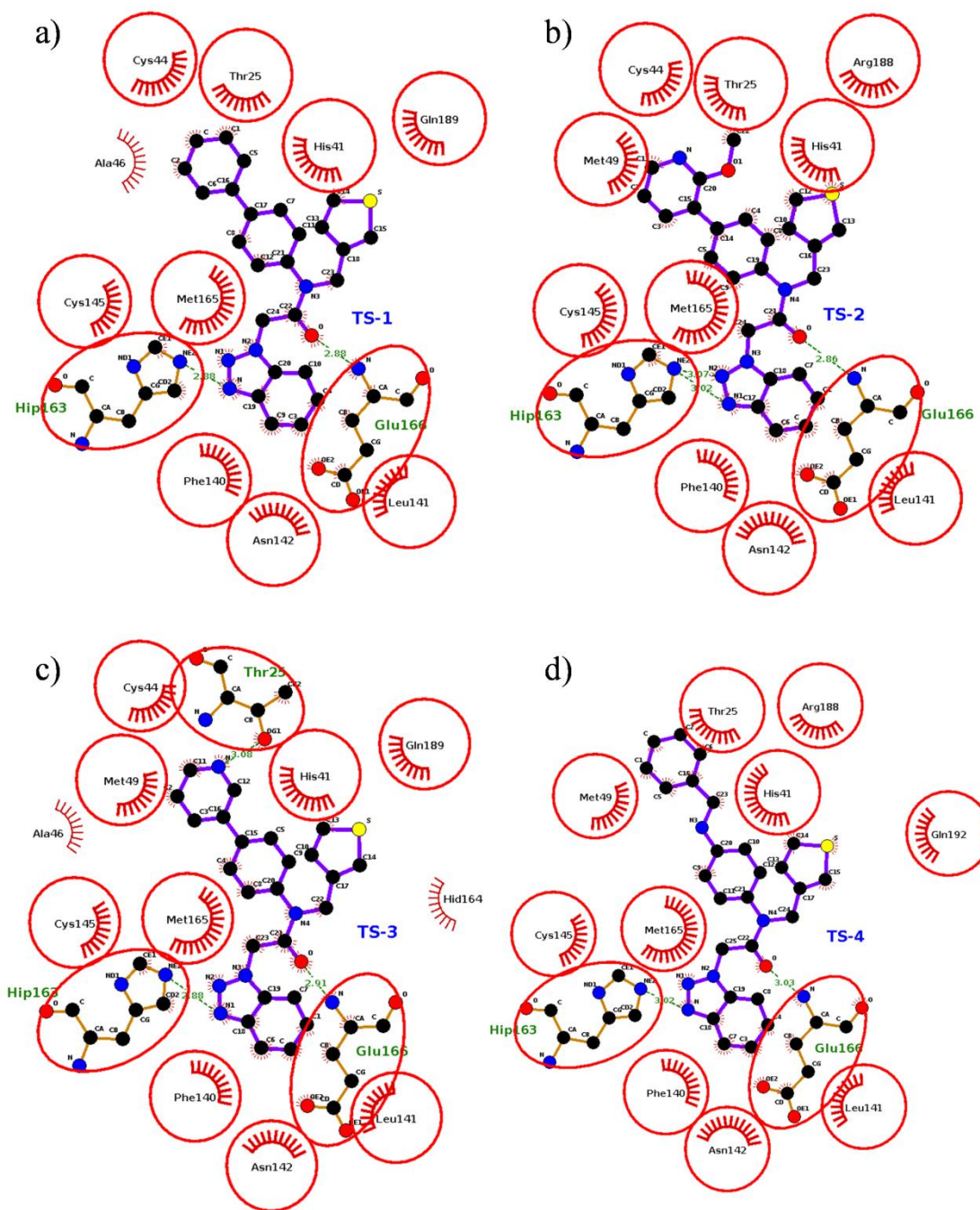


Figure S2. Intermolecular interaction pattern obtained from the molecular docking of compounds 1-4 of the training set (TS-1 - TS-4). Crystal structure template pdb code: 4MDS.

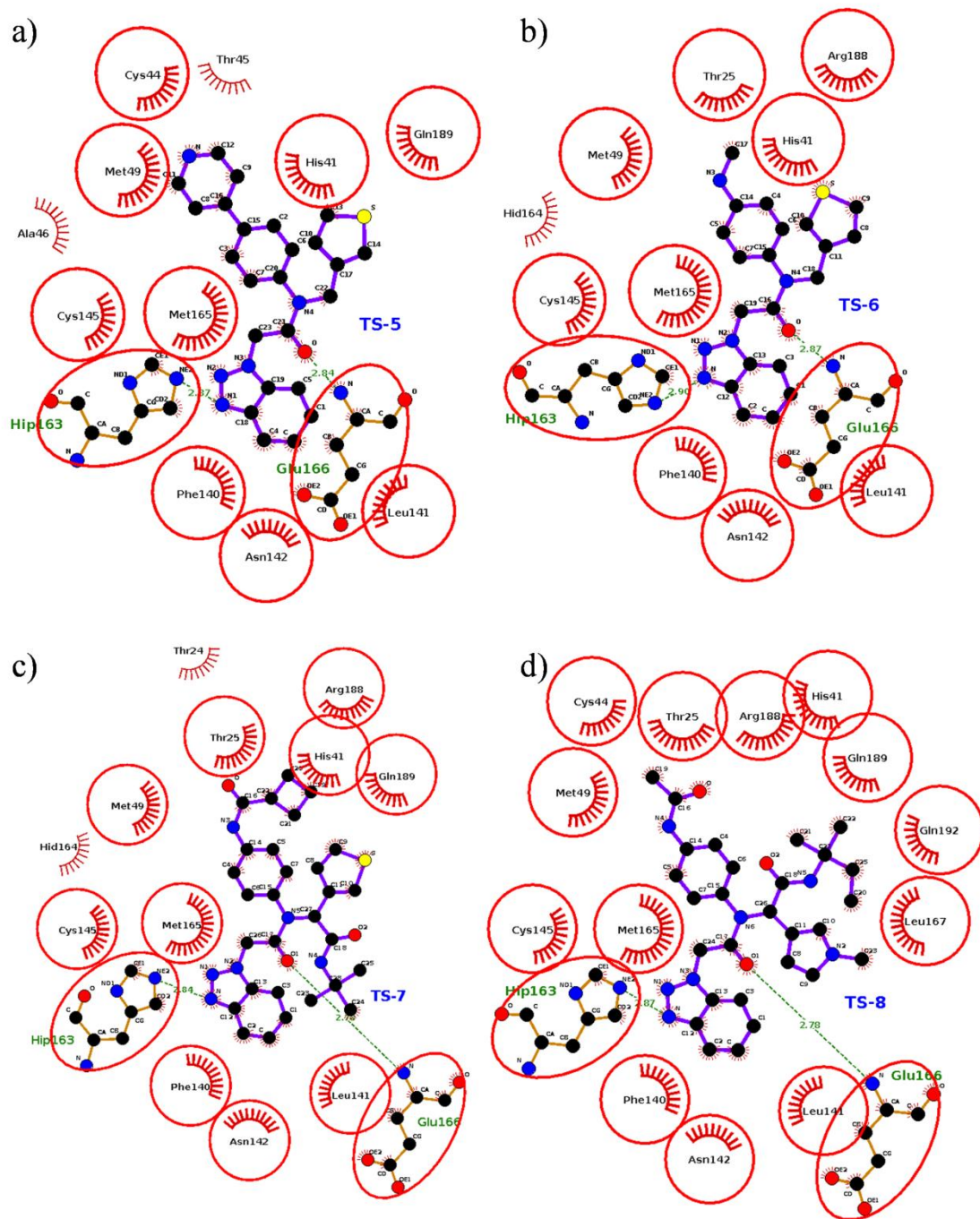


Figure S3. Intermolecular interaction pattern obtained from the molecular docking of compounds 5-8 of the training set (TS-5 - TS-8). Crystal structure template pdb code: 4MDS.

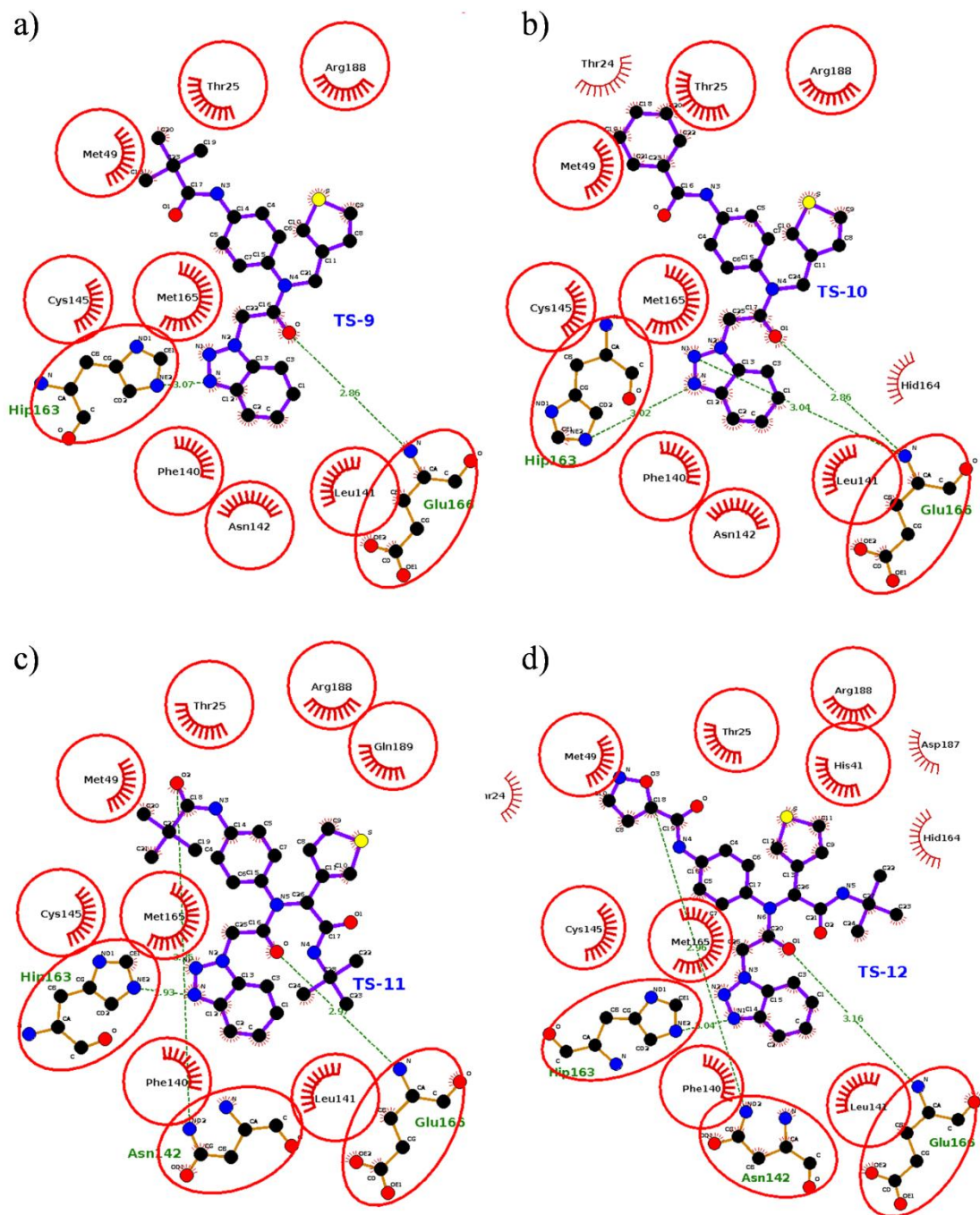


Figure S4. Intermolecular interaction pattern obtained from the molecular docking of compounds 9-12 of the training set (TS-9 - TS-12). Crystal structure template pdb code: 4MDS.

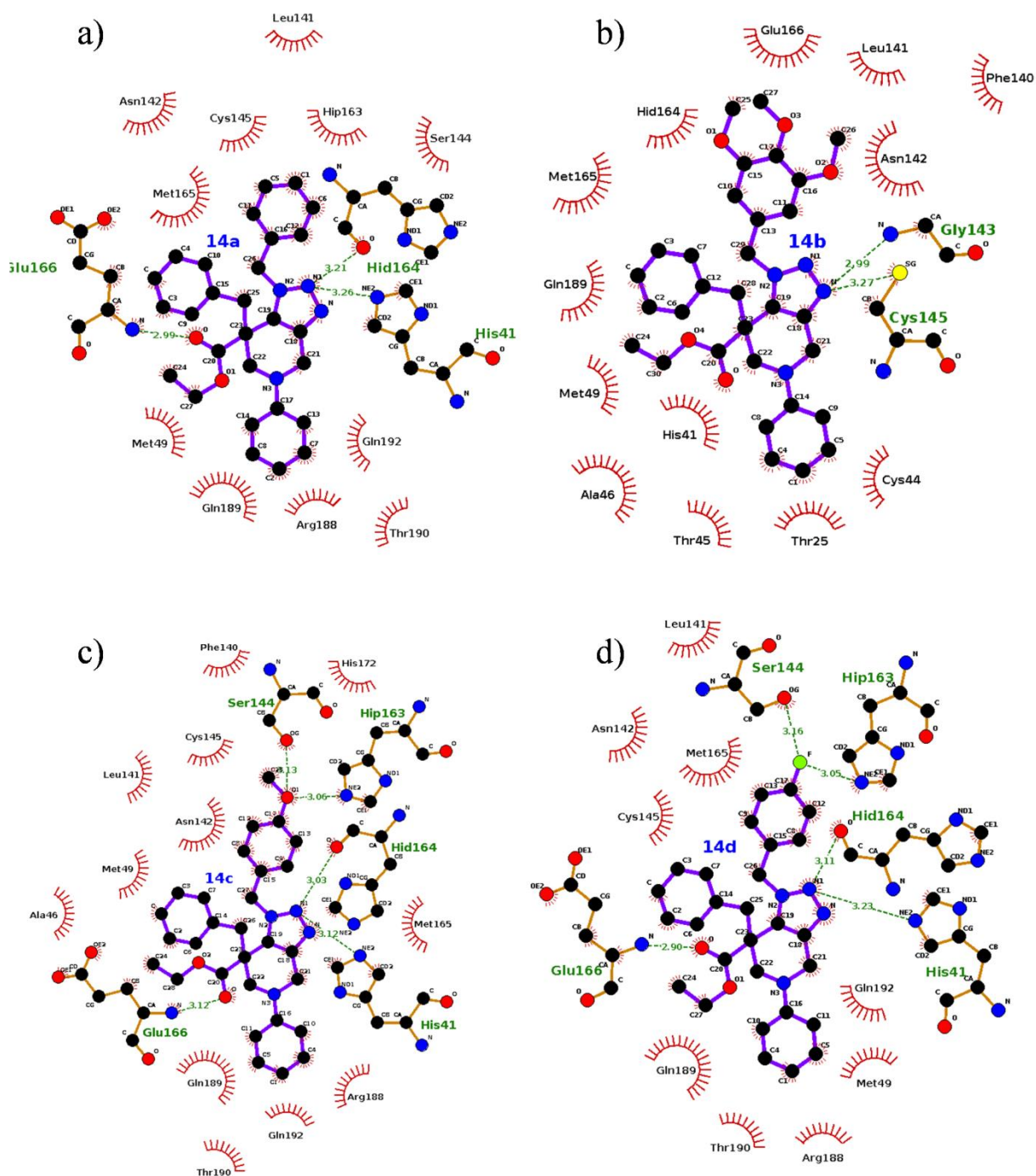


Figure S5. Lowest energy binding modes obtained by molecular docking simulation between 3CL^{PRO} (pdb code: 4MDS) and compounds 14a-14d.

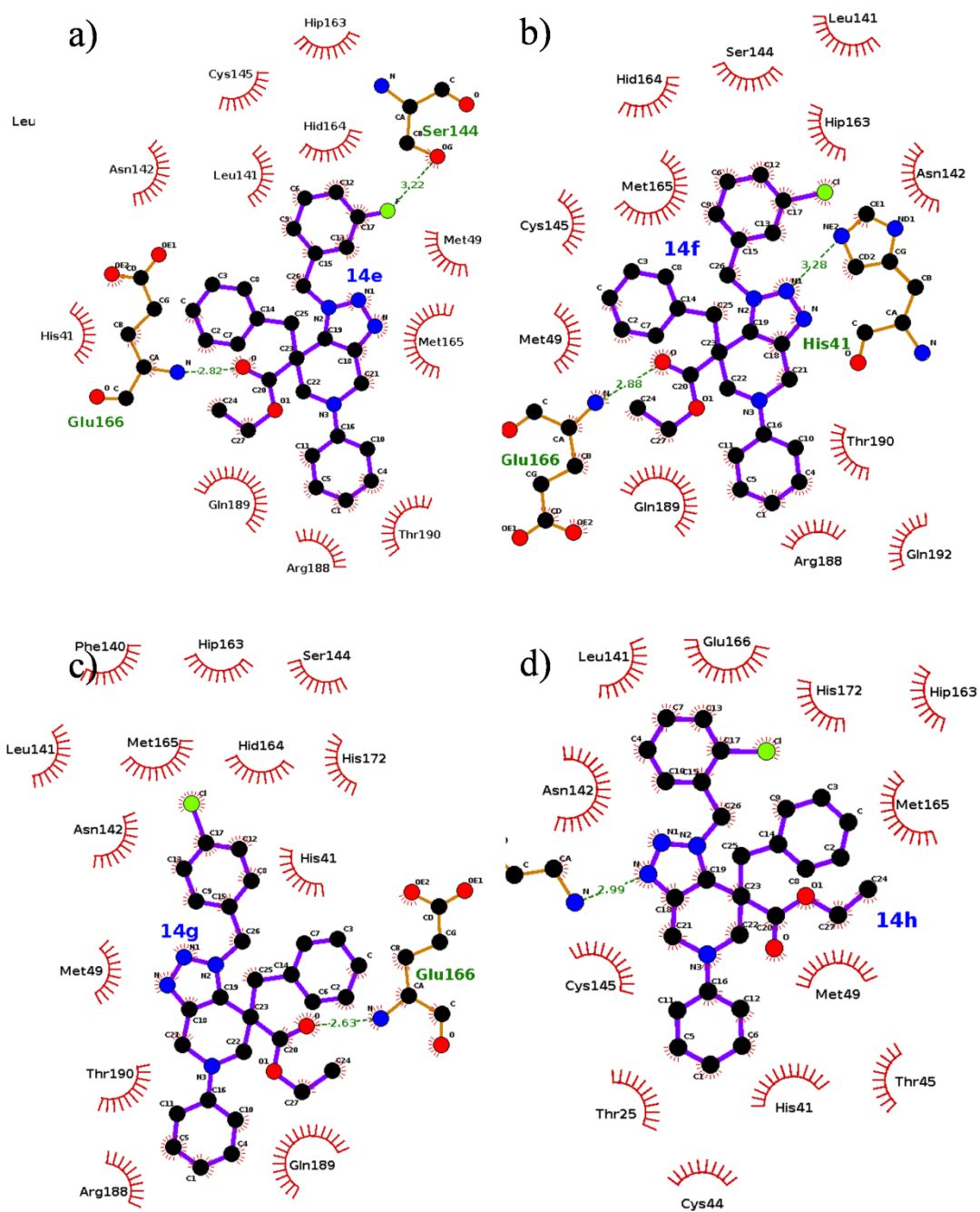


Figure S6. Lowest energy binding modes obtained by molecular docking simulation between 3CL^{pro} (pdb code: 4MDS) and compounds 14e-14h.

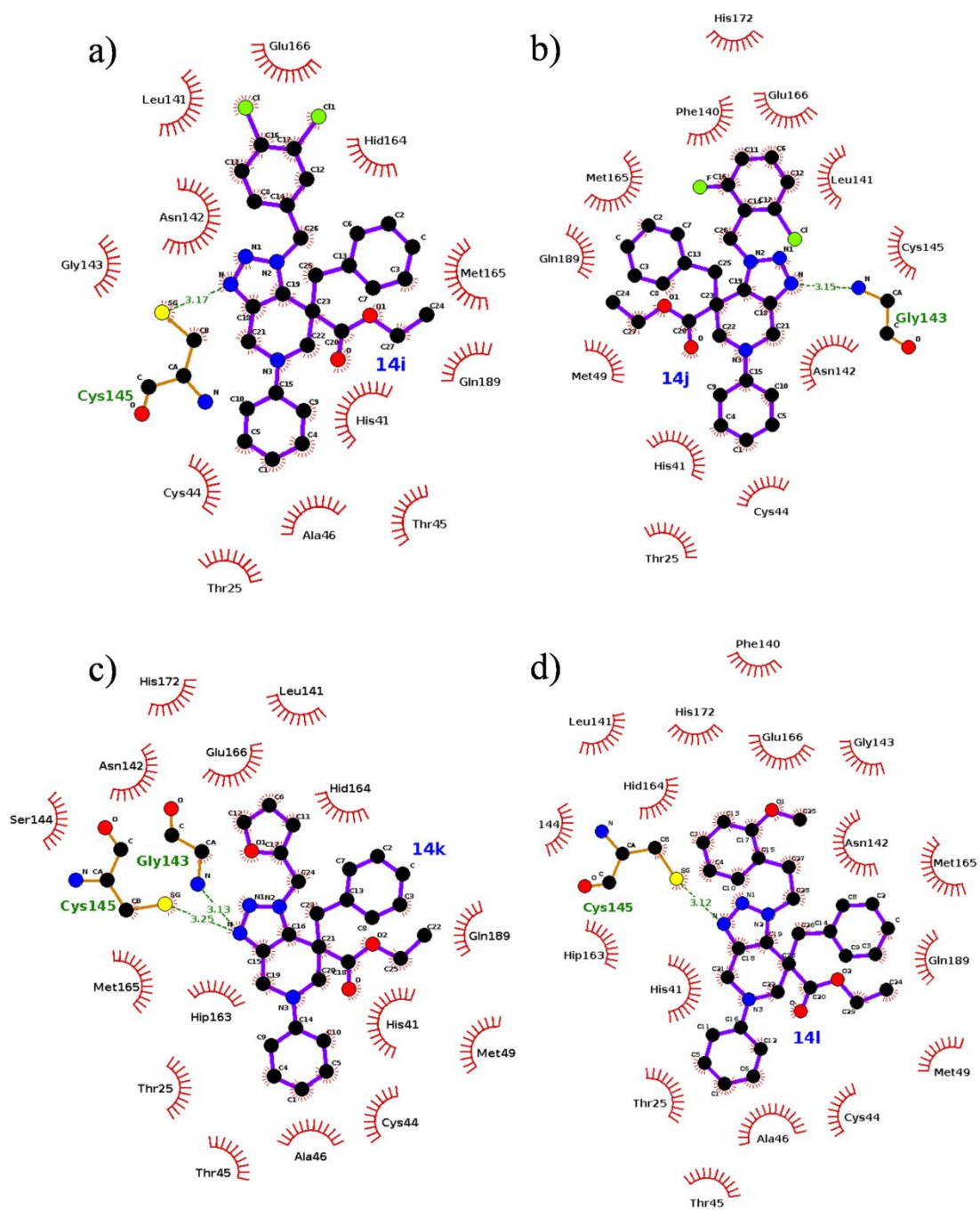


Figure S7. Lowest energy binding modes obtained by molecular docking simulation between 3CL^{PRO} (pdb code: 4MDS) and compounds 14i-14l.

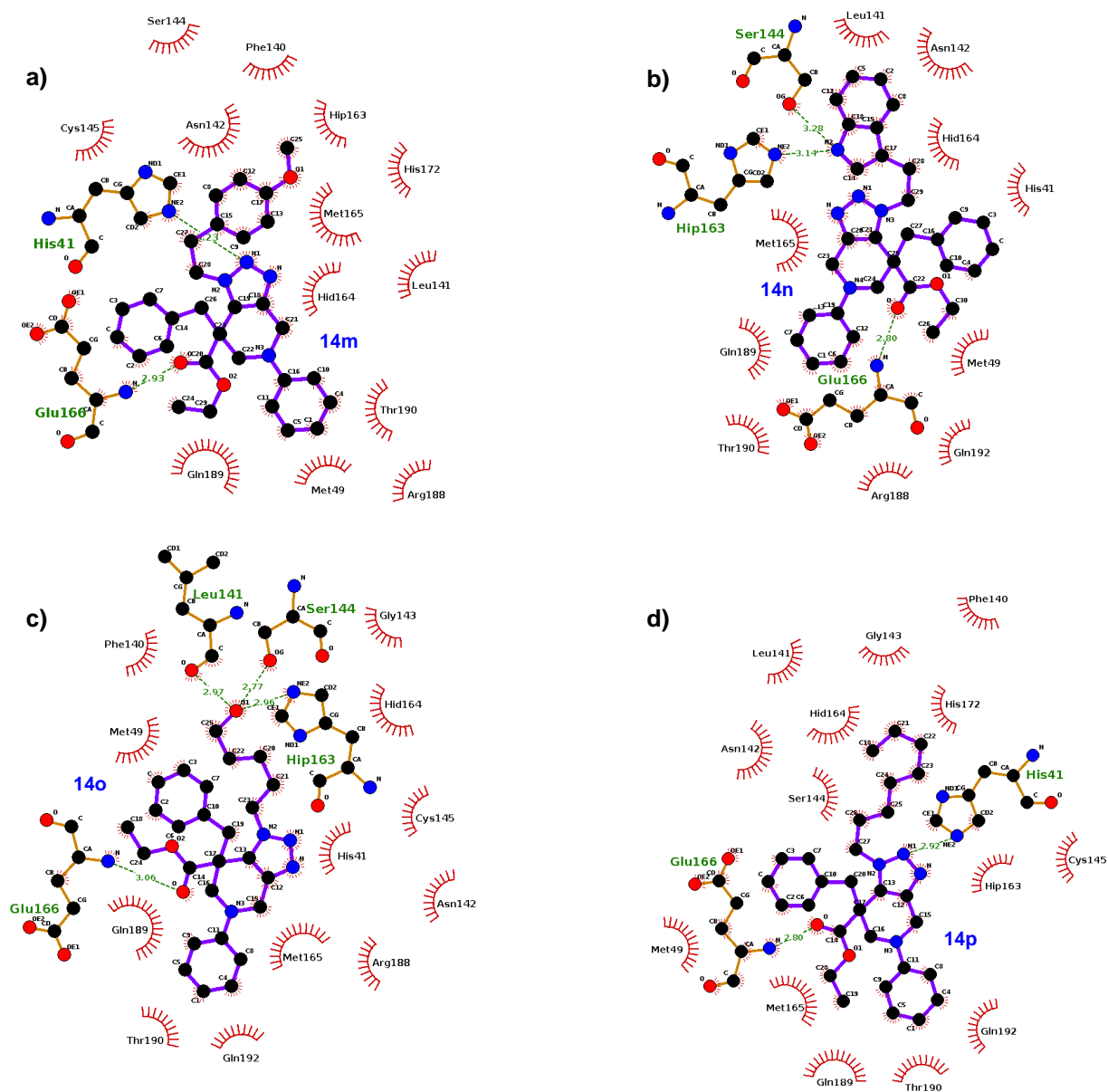


Figure S8. Lowest energy binding modes obtained by molecular docking simulation between 3CL^{pro} (pdb code: 4MDS) and compounds 14m-14p.

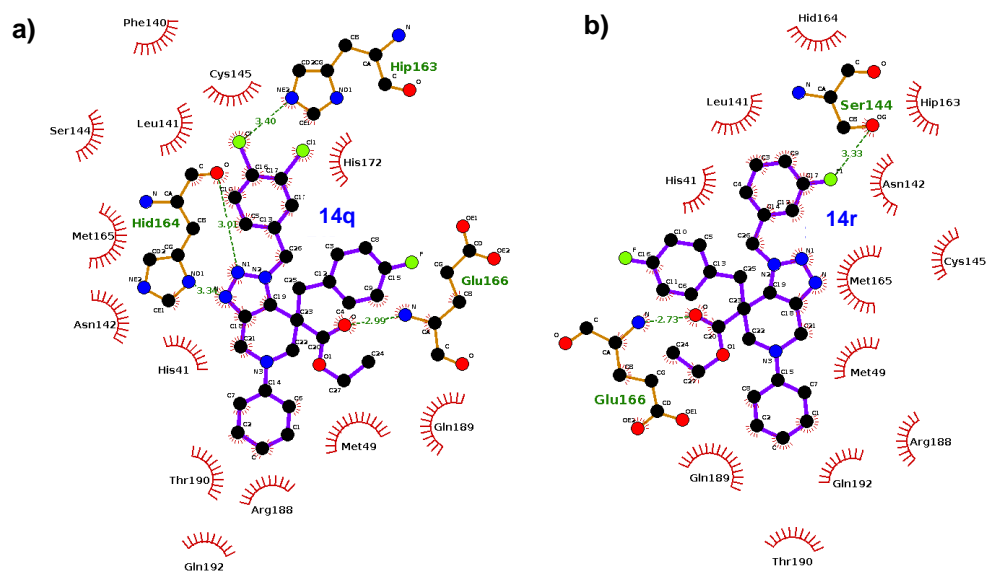


Figure S9. Lowest energy binding modes obtained by molecular docking simulation between 3CL^{pro} (pdb code: 4MDS) and compounds **14q-14r**.

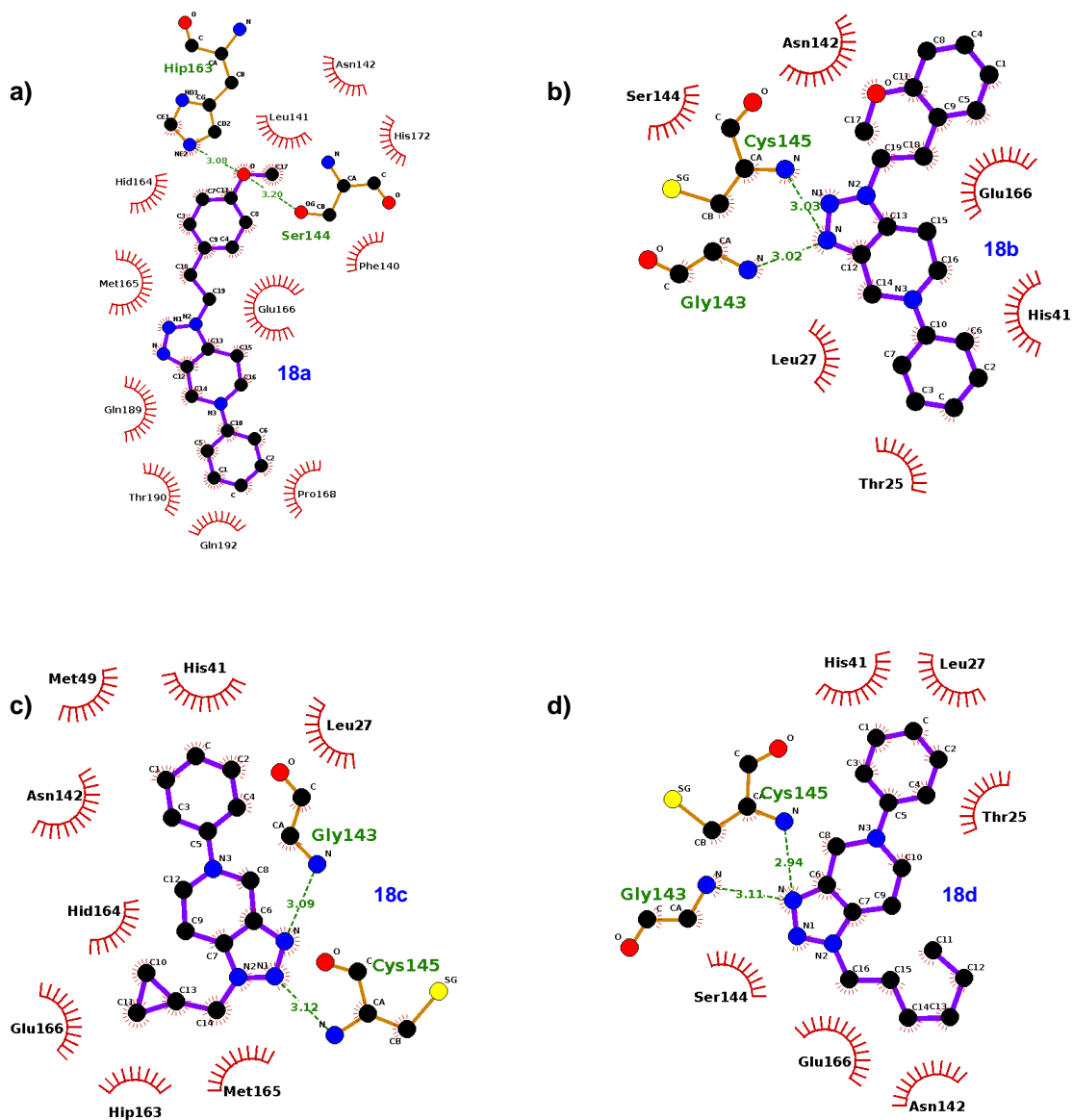


Figure S10. Lowest energy binding modes obtained by molecular docking simulation between 3CL^{pro} (pdb code: 4MDS) and compounds 18a-18d.

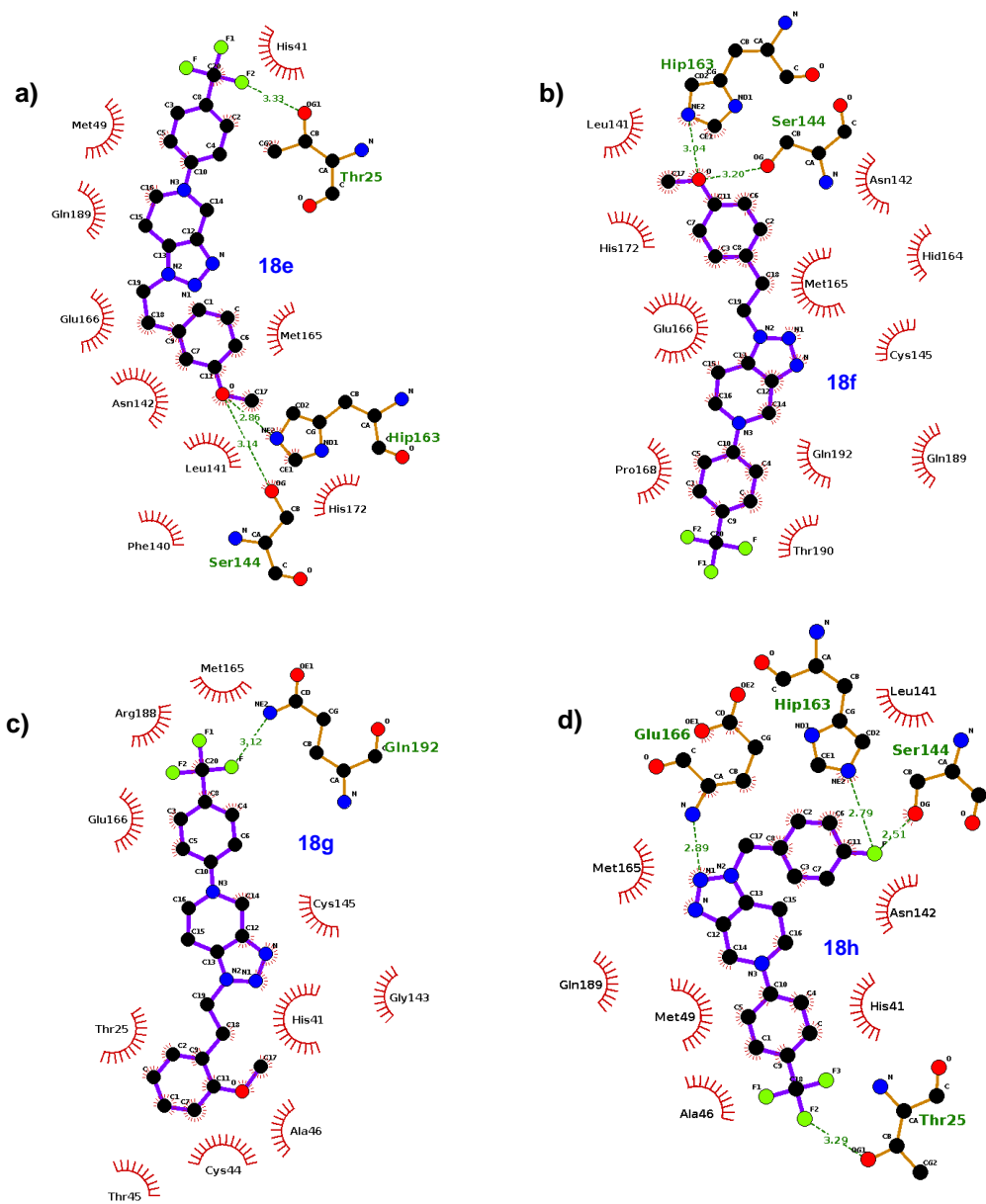


Figure S11. Lowest energy binding modes obtained by molecular docking simulation between 3CL^{pro} (pdb code: 4MDS) and compounds 18e-18h.

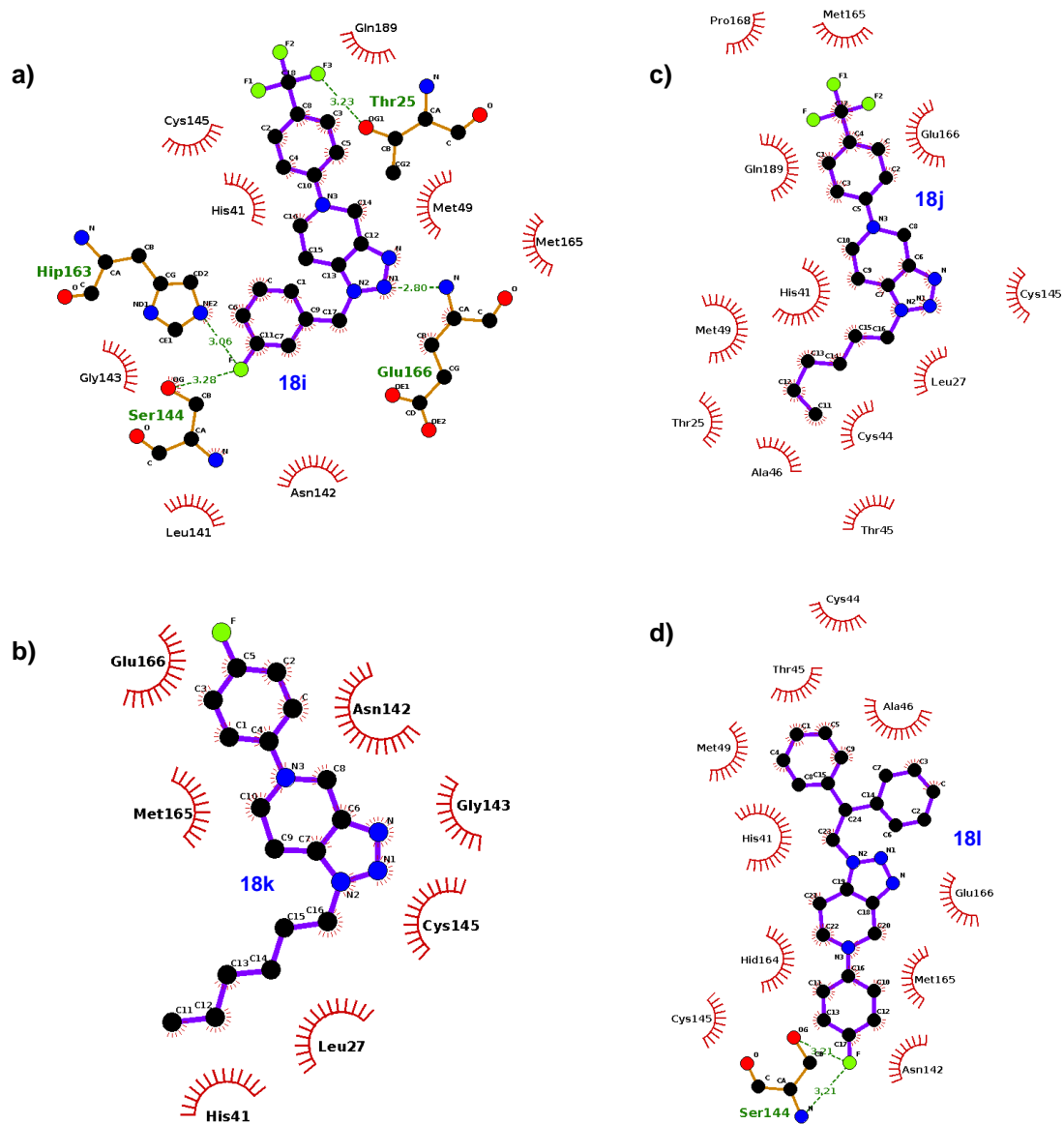


Figure S12. Lowest energy binding modes obtained by molecular docking simulation between 3CL^{pro} (pdb code: 4MDS) and compounds **18i-18l**.

Table S1. HB persistence value (%) for the training set.

Entry	Glu166 (% HB)	His163 (% HB)	Gly143(% HB)	Asn142(% HB)
1- 3V3M-R	82	39	96	-
2- 3V3M-S	60	0	0	-
3- TS-1	99	36	-	-
4- TS-2	99	34	-	-
5- TS-3	99	13	-	-
6- TS-4	99	25	-	-
7- TS-5	99	25	-	-
8- TS-6	99	32	-	-
9- TS-7 (R)	82	15	-	46
10- TS-8 (R)	99	48	-	-
11- TS-9	99	28	-	40
12- TS-10	99	10	-	-
13- TS-11 (R)	99	21	-	41
14- TS-12 (R)	49	10	-	72

Table S2. HB persistence value (%) for the fused 1,2,3-triazole derivatives.

Entry	Compound	Glu166 (% HB)	His163 (% HB)	Gln189 (% HB)	Thr25 (% HB)	Active
1	14a	37	-	-	-	
2	14b	-	-	-	-	
3	14c	99	88	-	-	
4	14d	99	57			
5	14e	99	-	-	-	
6	14f	41	-	-	-	
7	14g	98	7	-	-	
8	14h	-	71	-	-	
9	14i	63	56	-	-	
10	14j	-	46			
11	14k	-	3	-	-	
12	14l	99	-	-	-	
13	14m	94	70	-	-	
14	14n	97	69	-	-	
15	14o	79	50	-	-	
16	14p	92	-	-	-	
17	14q	90	47	-		
18	14r	61	-	-	-	
19	18a	-	12	35	-	
20	18b	-	-	-	-	
21	18c	-	-	-	-	
22	18d	-	-	-	-	
23	18e	-	46	-	-	
24	18f	40	94	-	55	
25	18g	-	-	-	-	
26	18h	98	-	-	-	
27	18i	99	97	52	-	
28	18j	-	-	-	-	
29	18k	-	-	-	-	
30	18l	48	-	-	-	

Table S3. Molecular structures of training set (TS1-TS4).

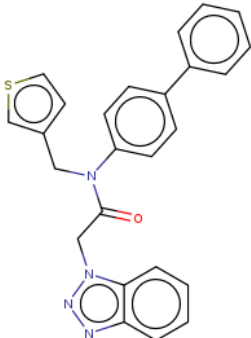
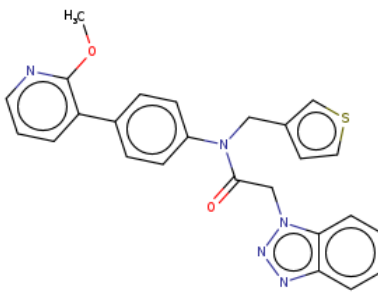
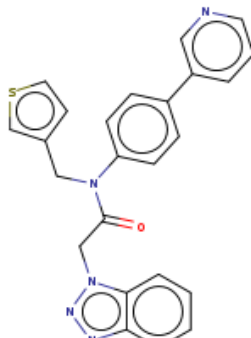
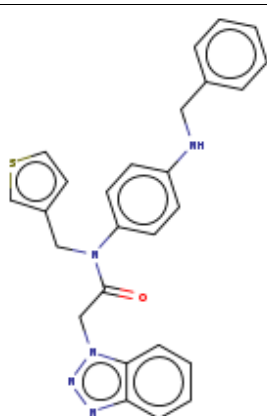
Compound	Structure	IC50 (nM)	Ref.
TS-1		51	7
TS-2		700	7
TS-3		970	7
TS-4		1500	7

Table S4. Molecular structures of training set (TS5-TS8).

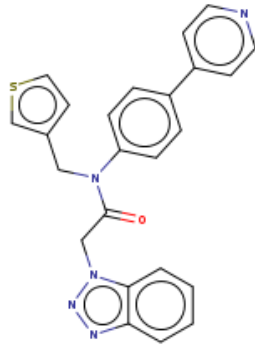
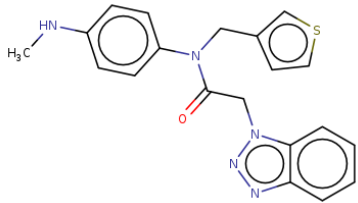
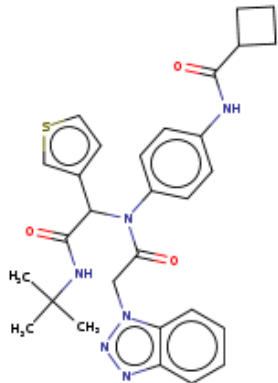
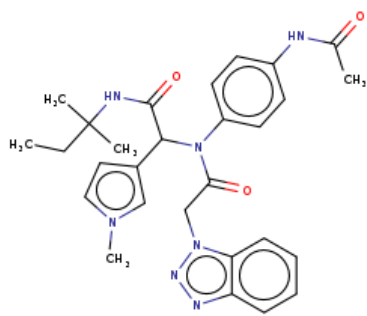
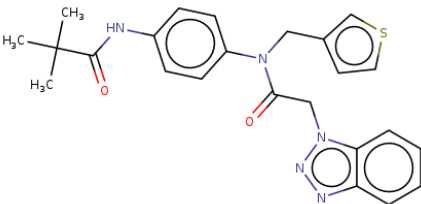
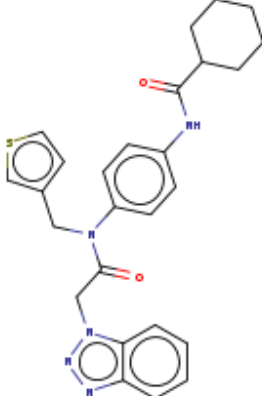
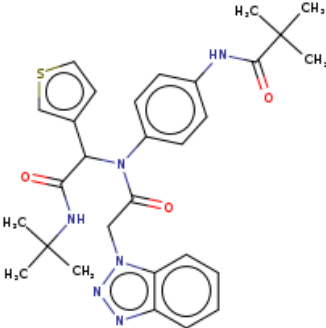
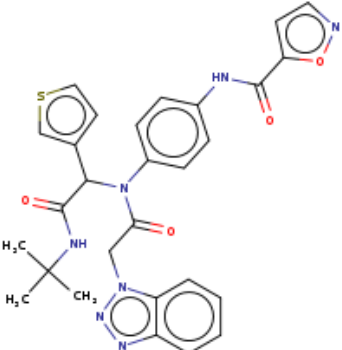
Compound	Structure	IC50 (nM)	Ref.
TS-5		2000	7
TS-6		2100	7
TS-7		3800	7
TS-8		6200	7

Table S5. Molecular structures of training set (TS9-TS12).

Compound	Structure	IC50 (nM)	Ref.
TS-9		13300	7
TS-10		22100	7
TS-11		22500	7
TS-12		26000	7

References

1. Pannecouque C, Daelemans D, De Clercq E. Tetrazolium-based colorimetric assay for the detection of HIV replication inhibitors: revisited 20 years later. *Nat Protoc.* 2008; 3: 427-34. <http://doi.org/10.1038/nprot.2007.517>.
2. De Clercq E, Holý A, Rosenberg I, Sakuma T, Balzarini J, Maudgal PC. A novel selective broad-spectrum anti-DNA virus agent. *Nature.* 1986; 323:464-7. <https://doi.org/10.1038/323464a0>.
3. Balzarini J, Naesens L, Slachmuylders J, Niphuis H, Rosenberg I, Holý A, Schellekens H, De Clercq E. 9-(2-Phosphonylmethoxyethyl)adenine (PMEA) effectively inhibits retrovirus replication in vitro and simian immunodeficiency virus infection in rhesus monkeys. *AIDS.* 1991; 5: 21-8.
4. MarvinSketch, 2017. MarvinSketch v.16.2.15. ChemAxon Ltd.
5. Gaussian 03, Revision C.02, Frisch MJ, Trucks GW, Schlegel HB, Scuseria GE, Robb MA, Cheeseman JR.; Montgomery Jr. JA, Vreven T, Kudin KN, Burant JC, Millam JM, Iyengar SS, Tomasi J, Barone V, Mennucci B, Coss, M, Scalmani G, Rega N, Petersson GA, Nakatsuji H, Hada M, Ehara M, Toyota K, Fukuda R, Hasegawa J, Ishida M, Nakajima T, Honda Y, Kitao O, Nakai H, Klene M, Li X, Knox JE, Hratchian HP, Cross JB, Bakken V, Adamo C, Jaramillo J, Gomperts R, Stratmann RE, Yazyev O, Austin AJ, Cammi R, Pomelli C, Ochterski JW, Ayala PY, Morokuma K, Voth GA, Salvador P, Dannenberg JJ, Zakrzewski VG, Dapprich S, Daniels AD, Strain MC, Farkas O, Malick DK, Rabuck AD, Raghavachari K, Foresman JB, Ortiz JV, Cui Q, Baboul AG, Clifford S, Cioslowski J, Stefanov BB, Liu G, Liashenko A, Piskorz P, Komaromi I, Martin RL, Fox DJ, Keith T, Al-Laham MA, Peng CY, Nanayakkara A, Challacombe M, Gill PMW, Johnson B, Chen W, Wong MW, Gonzalez C, Pople JA, Gaussian, Inc., Wallingford CT, 2004.Open
6. OMEGA 3.0.0.1: OpenEye Scientific Software, Santa Fe, NM. <http://www.eyesopen.com>.
7. Turlington M, Chun A, Tomar S, Egger A, Grum-Tokars V, Jacobs J, Daniels JS, Dawson E, Saldanha A, Chase P, Baez-Santos YM, Lindsley CW, Hodder P, Mesecar AD, Stauffer SR. Discovery of N-(benzo[1,2,3]triazol-1-yl)-N-(benzyl)acetamido)phenyl carboxamides as severe acute respiratory syndrome coronavirus (SARS-CoV) 3CLpro inhibitors: Identification of ML300 and noncovalent nanomolar inhibitors with an induced-fit binding. *Bioorg Med Chem Lett.* 2013; 23: 6172-6177. <http://doi.org/10.1016/j.bmcl.2013.08.112>.
8. Jacobs J, Grum-Tokars V, Zhou Y, Turlington M, Saldanha SA, Chase P, Egger A, Dawson ES, Baez-Santos YM, Tomar S, Mielech AM, Baker SC, Lindsley CW, Hodder P, Mesecar A, Stauffer SR. Discovery, synthesis, and structure-based optimization of a series of N-(tert-Butyl)-2-(N-arylamido)-2-(pyridin-3-yl)acetamides (ML188) as potent noncovalent small molecule inhibitors of the severe acute respiratory syndrome coronavirus (SARS-CoV) 3CL. *J Med Chem.* 2013; 56: 534-546. <https://doi.org/10.1021/jm301580n>.

9. Fährrolfes R, Bietz S, Flachsenberg F, Meyder A, Nittinger E, Otto, T, Volkamer, A, Rarey M. ProteinsPlus: a web portal for structure analysis of macromolecules. *Nucleic Acids Res.* 2017; 45: W337-W343. <https://doi.org/10.1093/nar/gkx333>.
10. Trott O., Olson AJ., AutoDock Vina: improving the speed and accuracy of docking with a new scoring function, efficient optimization and multithreading, *J Comput Chem.* 2010; 31: 455-461. <https://doi.org/10.1002/jcc.21334>.
11. Miller BR, McGee TD, Swails JM, Homeyer N, Gohlke H, Roitberg AE. MMPBSA.py: An Efficient Program for End-State Free Energy Calculations. *J Chem Theory Comput.* 2012; 8: 3314-3321. <https://doi.org/10.1021/ct300418h>.
12. VIDA. 4.3.0, 2017. OpenEye Scientific Software, Santa Fe, NM. <http://www.eyesopen.com>.
13. Wallace AC, Laskowski RA, Thornton JM. LIGPLOT: a program to generate schematic diagrams of protein-ligand interactions *J Chem Inf Model.* 2011; 51: 2778-2786. <https://doi.org/10.1021/ci200227u>.
14. A. Case, IY. Ben-Shalom, SR. Brozell, DS. Cerutti, TE. Cheatham, III, VWD. Cruzeiro, TA. Darden, RE. Duke, D. Ghoreishi, MK. Gilson, H. Gohlke, AW. Goetz, D. Greene, R Harris, N. Homeyer, S. Izadi, A. Kovalenko, T. Kurtzman, TS. Lee, S. LeGrand, P. Li, C. Lin, J. Liu, T. Luchko, R. Luo, DJ. Mermelstein, KM. Merz, Y. Miao, G. Monard, C. Nguyen, H. Nguyen, I. Omelyan, A. Onufriev, F. Pan, R. Qi, DR. Roe, A. Roitberg, C. Sagui, S. Schott-Verdugo, J. Shen, CL. Simmerling, J. Smith, R. Salomon-Ferrer, J. Swails, RC. Walker, J. Wang, H. Wei, RM. Wolf, X. Wu, L. Xiao, DM. York and PA. Kollman (2018), AMBER 2018, University of California, San Francisco.
15. Maier JA, Martinez C, Kasavajhala K, Wickstrom L, Hauser KE, Simmerling C. ff14SB: Improving the accuracy of protein side chain and backbone parameters from ff99SB. *J Chem Theory Comput.* 2015; 11: 3696-3713. <https://doi.org/10.1021/acs.jctc.5b00255>.
16. Wang J, Wolf RM, Caldwell JW, Kollman PA, Case DA. Development and testing of a general AMBER force field. *J Comput Chem.* 2004; 25: 1157-1174. <https://doi.org/10.1002/jcc.20035>.