Microbial Diversity and Metabolic Potential in the Stratified Sansha

Yongle Blue Hole in the South China Sea

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Table S1. Physiochemical	variables and n	nethods using	redundancy a	nalvsis
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Variables	Method		
Silicate	Molybdosilicic acid method (Fanning and Pilson, 1973)		
Nitrate, Nitrite	Standard pink azo dye method (Strickland and Parsons, 1972),		
Ammonium	Hypobromite oxidation method (Strickland and Parsons, 1972).		
Temperature, Salinity	A Conductivity Temperature Depth profiler (Sea-Bird SBE		
	911plus)		
Suspended particulate matter	Gravimetric analysis		
Dissolved organic carbon	High temperature catalytic oxidation using a Total Organic Carbon		
	analyzer (Shimadzu, Japan) (Sugimura and Suzuki, 1988)		
Particulate organic carbon,	A CHN elemental analyzer (Elementar Analysensysteme		
Particulate nitrogen	GmbH, Germany).		
Dissolved oxygen	Metrohm 848 Titrino plus sensor using the Winkler method		
	(Metrohm, Switzerland)		
Methane, nitrous oxide	Gas chromatography using gas-stripping methods (Zhang et al.,		
	2004, 2006)		
Sufide	Methylene blue method (State technology supervision bureau,		
	2007)		

Physiochemical variables and methods were based on our parallel study²⁰.

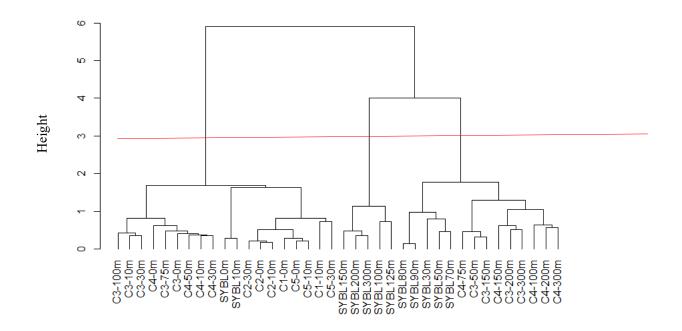


Figure S1. Cluster analysis of 1345 microbial genera from the Sansha Yongle Blue Hole and surrounding regions based on 16S rRNA amplicon sequences

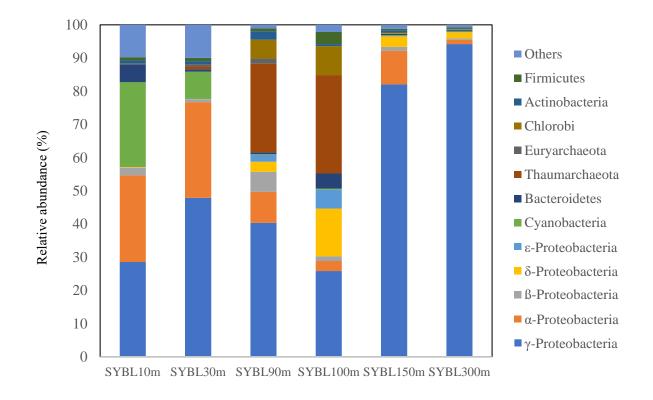


Figure S2. Relative abundance of microbial phyla from the Sansha Yongle Blue Hole based on metagenomic sequences

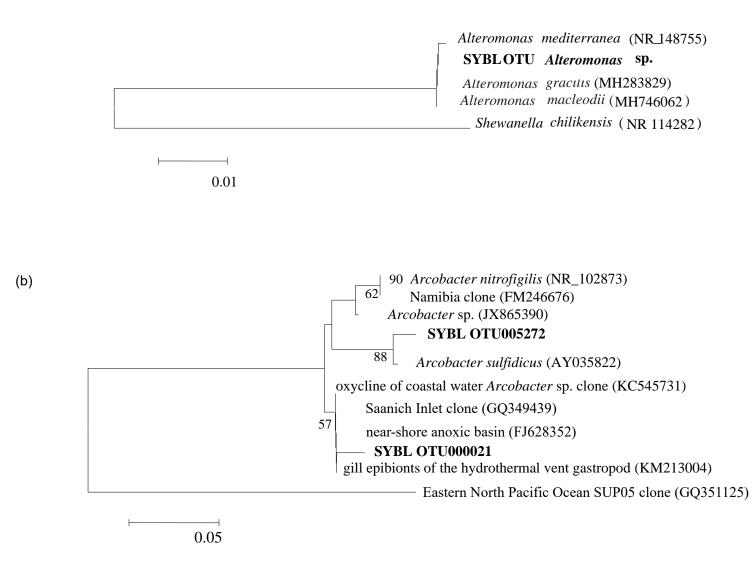


Figure S3. Phylogenetic trees of (a) nitrate-reducing bacteria and (b) sulfide-oxidizing bacteria, based on 16S rRNA sequences. Phylogenetic trees were constructed using the neighbor-joining (NJ) algorithm implemented in MEGA4⁹⁴. Bootstrapping was performed by resampling 1000 times. Bootstrap values <50% are not shown.