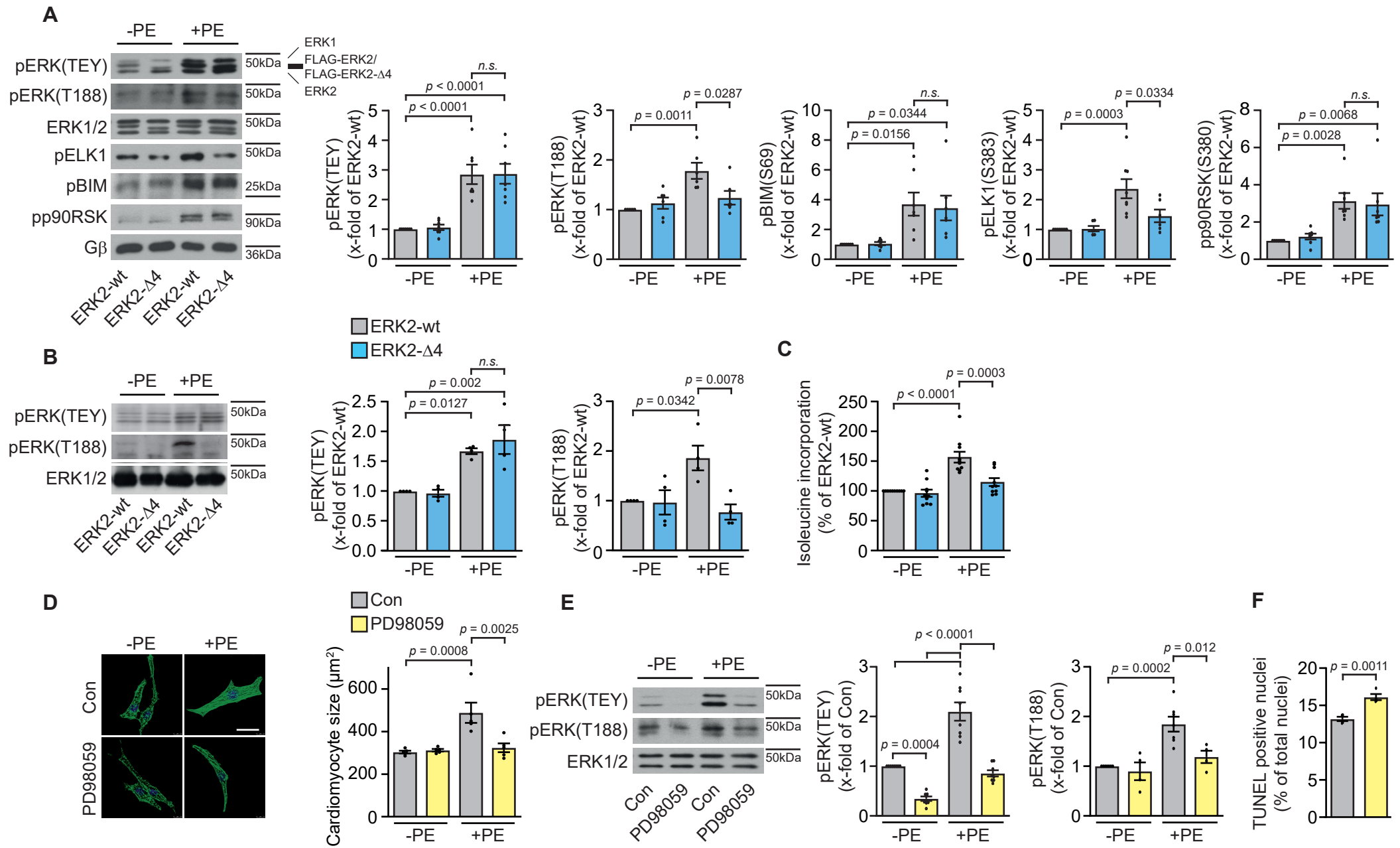


Supplementary Information accompanying:

Interference with ERK-dimerization at the nucleocytoplasmic interface targets pathological ERK1/2 signaling without cardiotoxic side-effects

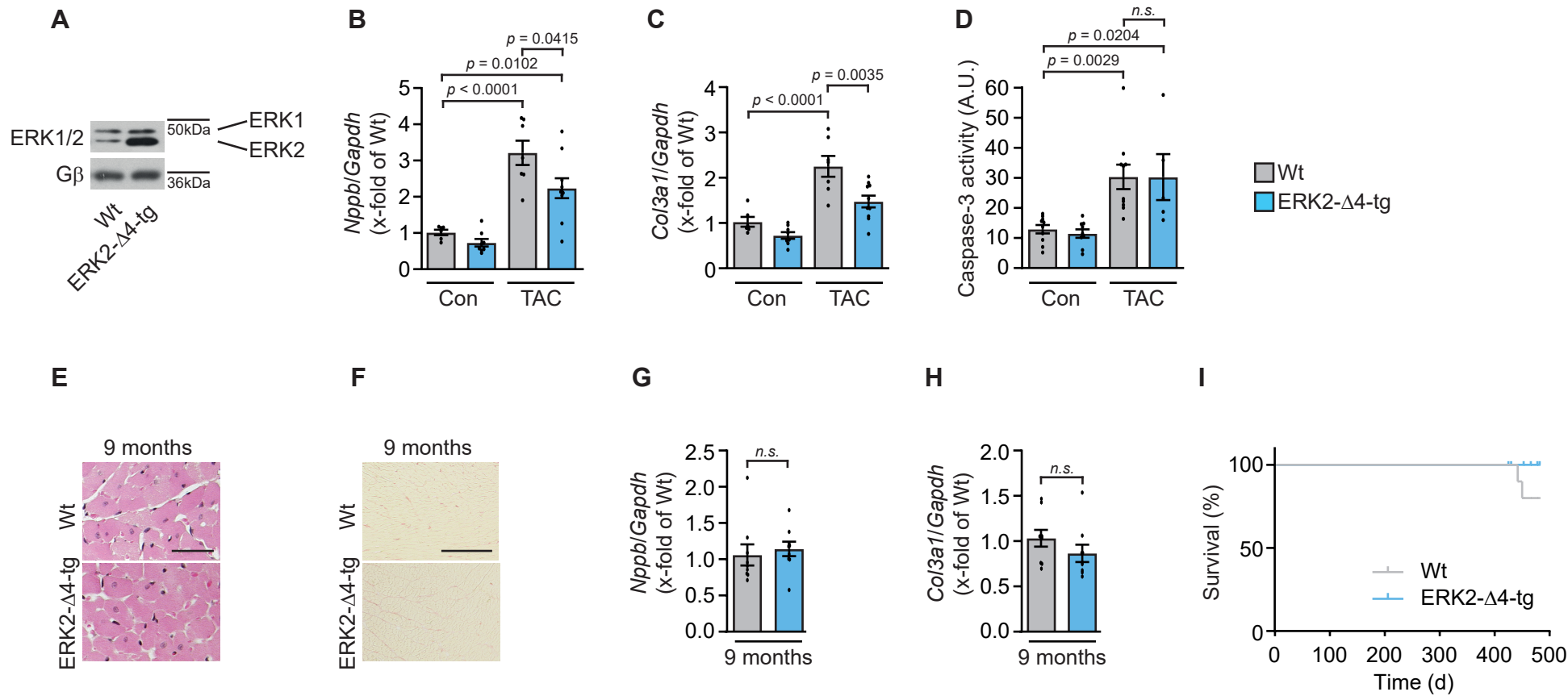
Angela Tomasovic#, Theresa Brand#, Constanze Schanbacher#, Sofia Kramer, Martin W. Hümmert, Patricio Godoy, Wolfgang Schmidt-Heck, Peter Nordbeck, Jonas Ludwig, Susanne Homann, Armin Wiegering, Timur Shaykhtudinov, Christoph Kratz, Ruth Knüchel, Hans-Konrad Müller-Hermelink, Andreas Rosenwald, Norbert Frey, Jutta Eichler, Dobromir Dobrev, Ali El-Armouche, Jan G. Hengstler, Oliver J. Müller, Karsten Hinrichs, Friederike Cuello, Alma Zerneck, Kristina Lorenz*

Supplementary Figures and Tables



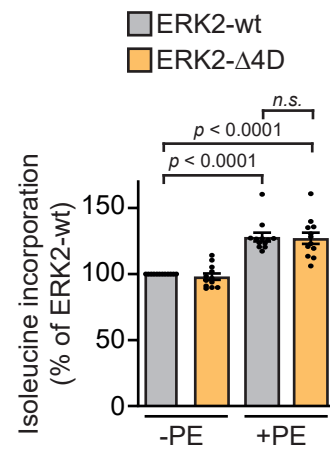
Supplementary Figure 1

Cells transduced with monomeric ERK2 show reduced ERK_{T188}-phosphorylation but normal ERK activation. (A-C) NRCM were transduced with Flag-ERK2-wt (ERK2-wt) or Flag-ERK2 Δ 174-177 (ERK2- Δ 4) and stimulated as indicated. (A) Immunoblot analysis of pERK(TEY) (n=7) and pERK(T188) (n=6) and phosphorylation levels of nuclear (pELK1[S383], n=8 for ERK2-wt and n=6 for ERK2- Δ 4) and cytosolic (pBIM[S69], n=7; pp90RSK[S380], n=7) ERK-targets in the absence (-PE) and presence of phenylephrine stimulation (+PE; 4 μ M, 10min). Shown are quantitative analyses and representative western blots. G β was used as additional loading control. (B) Immunoblot analysis of pERK(TEY) and pERK(T188) in the absence (-PE) and presence of phenylephrine stimulation (+PE; 4 μ M, 24h) (n=4). Shown are quantitative analyses and representative western blots. (C) [³H]Isoleucine incorporation assay (n=10). Cells were stimulated with PE as indicated (4 μ M, 24h). (D) Representative pictures of phalloidin-stained NRCM and cell area analysis of NRCM pretreated with **PD98059** (30 μ M, 1h) and stimulated with PE as indicated (4 μ M, 24h; n=5 and 40 cells per group and experiment; scale bar 20 μ m). (E) Western blot analyses of NRCM pretreated with **PD98059** (30 μ M, 1h) and stimulated with PE (4 μ M, 10min) as indicated for ERK(TEY)- and ERK(T188)-phosphorylation. Shown are quantitative analyses and representative western blots (pERK(TEY), n=8; pERK(T188), Con \pm PE n=7 and PD98059 \pm PE n=4). (F) TUNEL assay analysis of NRCM after H₂O₂ treatment (100 μ M, 1h; n=4 of at least 500 cells per experiment and group). Error bars are mean \pm s.e.m.; n numbers represent biologically independent experiments. For statistical analysis ordinary one-way ANOVA (A-E) or unpaired and two-sided Student's *t*-test (F) were applied, for (B) the Tukey test and for (A,C,D,E) Bonferroni was applied as *post hoc* test. Source data are provided as a Source Data file.



Supplementary Figure 2

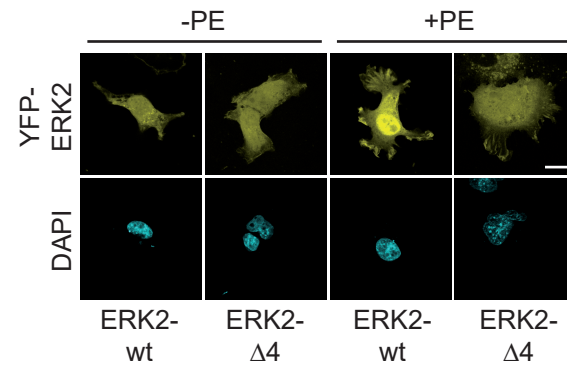
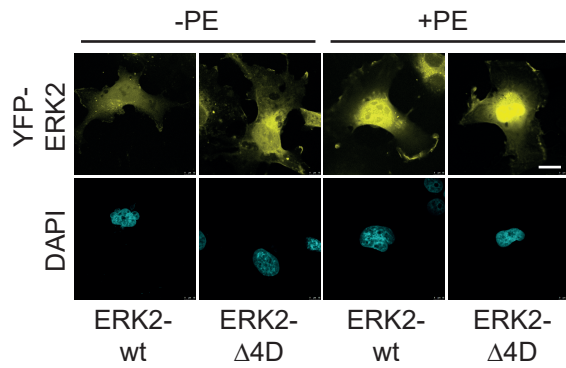
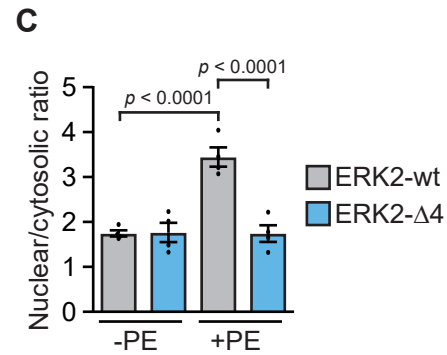
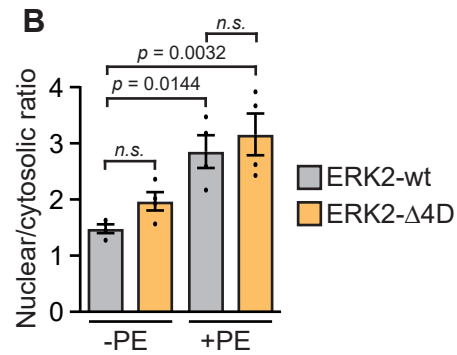
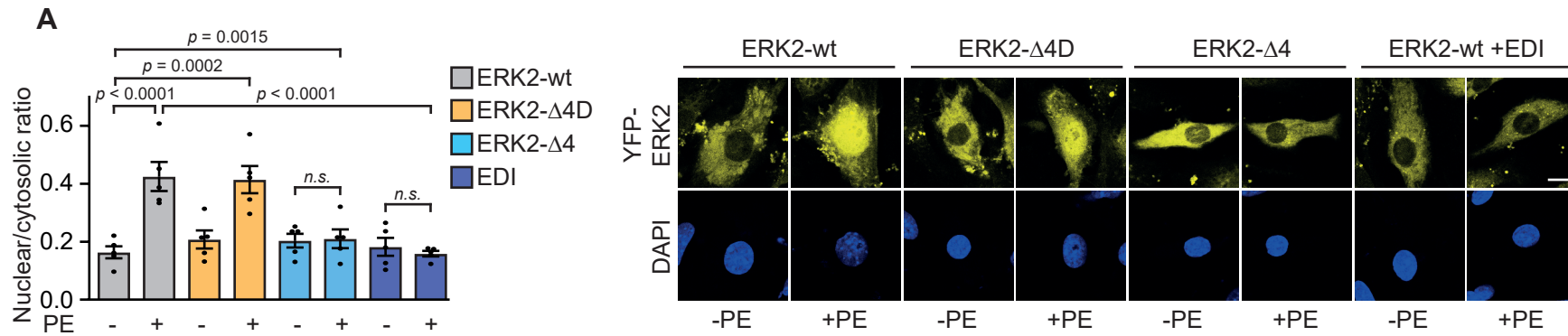
Monomeric ERK2 attenuates pathological cardiac remodeling but not physiological heart growth. (A) Representative western blot of ERK1/2 expression levels in wild-type (Wt) and ERK2 Δ 174-177 (ERK2- Δ 4-tg) transgenic mice. G β was used as an additional loading control. Western blot was reproduced five times with similar result. (B-D) Wt and ERK2- Δ 4-tg mice were analyzed for apoptosis and markers of remodeling before (Con) and 6 weeks after transverse aortic constriction (TAC). (B,C) mRNA expression levels of brain natriuretic peptide (*Nppb*; B) and collagen type III alpha 1 (*Col3a1*; C) normalized to glyceraldehyde-3-phosphate dehydrogenase (*Gapdh*). Wt Con, n=6; ERK2- Δ 4-tg Con, n=8; Wt TAC, n=7; ERK2- Δ 4-tg TAC, n=10 mice per group; n represent numbers of mice measured in triplicates. (D) Caspase-3 activity in heart lysates (Wt Con, n=10; ERK2- Δ 4-tg Con, n=9; Wt TAC, n=10; ERK2- Δ 4-tg TAC, n=5 mice per group). (E-I) 9 months old Wt and ERK2- Δ 4-tg were analyzed for cardiomyocyte size, interstitial fibrosis, markers of cardiac remodeling and survival. (E,F) Shown are representative histological H&E (scale bar 50 μ m, E) and Sirius Red-stained (scale bar 200 μ m, F) sections of left ventricular myocardium. Histological stainings were performed in Wt (n=9) and ERK2- Δ 4-tg (n=10). Quantifications: Supplementary Table 2. (G,H) mRNA expression levels of *Nppb* (G) and *Col3a1* (H) normalized to *Gapdh* (n=9; n represents the numbers of mice per group measured in triplicates). (I) Kaplan-Meier survival curves of Wt (n=14) and ERK2- Δ 4-tg (n=25 mice per group). Error bars are mean \pm s.e.m.. For statistical analysis ordinary one-way ANOVA (B-D) and Bonferroni as *post hoc* test or an unpaired and two-sided Student's *t*-test (G,H) were applied. For (I) an survival curve comparison was applied. Source data are provided as a Source Data file.



Supplementary Figure 3

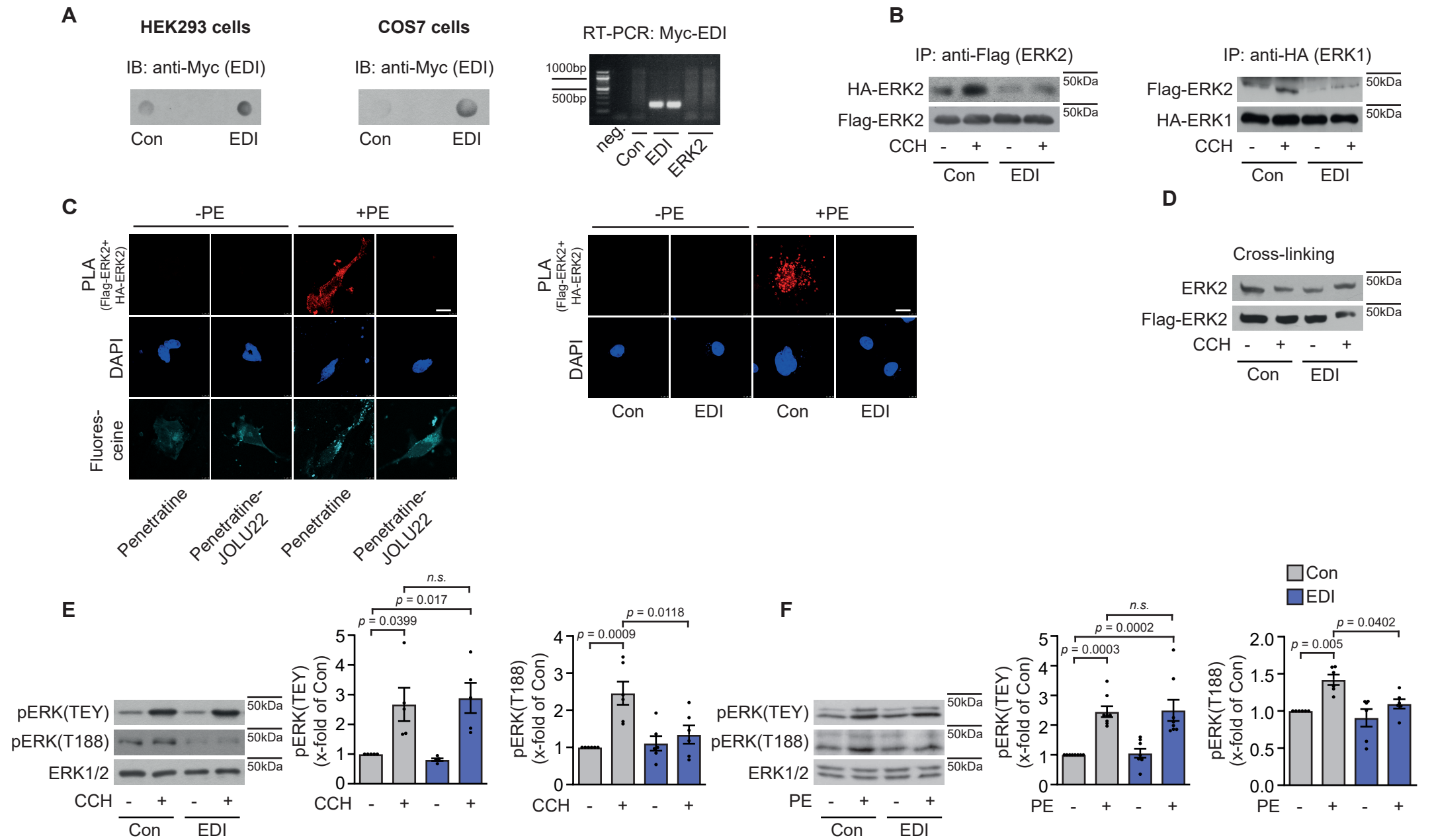
ERK2-Δ4D shows a comparable hypertrophic response to phenylephrine as ERK2-wt.

[³H]Isoleucine incorporation assay of NRCM transduced with Flag-tagged ERK2-wt or Flag-ERK2_{Δ174-177,T188D} (ERK2-Δ4D) stimulated with phenylephrine (PE; 4μM, 30h; n=12 experiments). Error bars are mean±s.e.m., n numbers represent biologically independent experiments. For statistical analysis ordinary one-way ANOVA and Bonferroni as *post hoc* test was applied. Source data are provided as a Source Data file.



Supplementary Figure 4

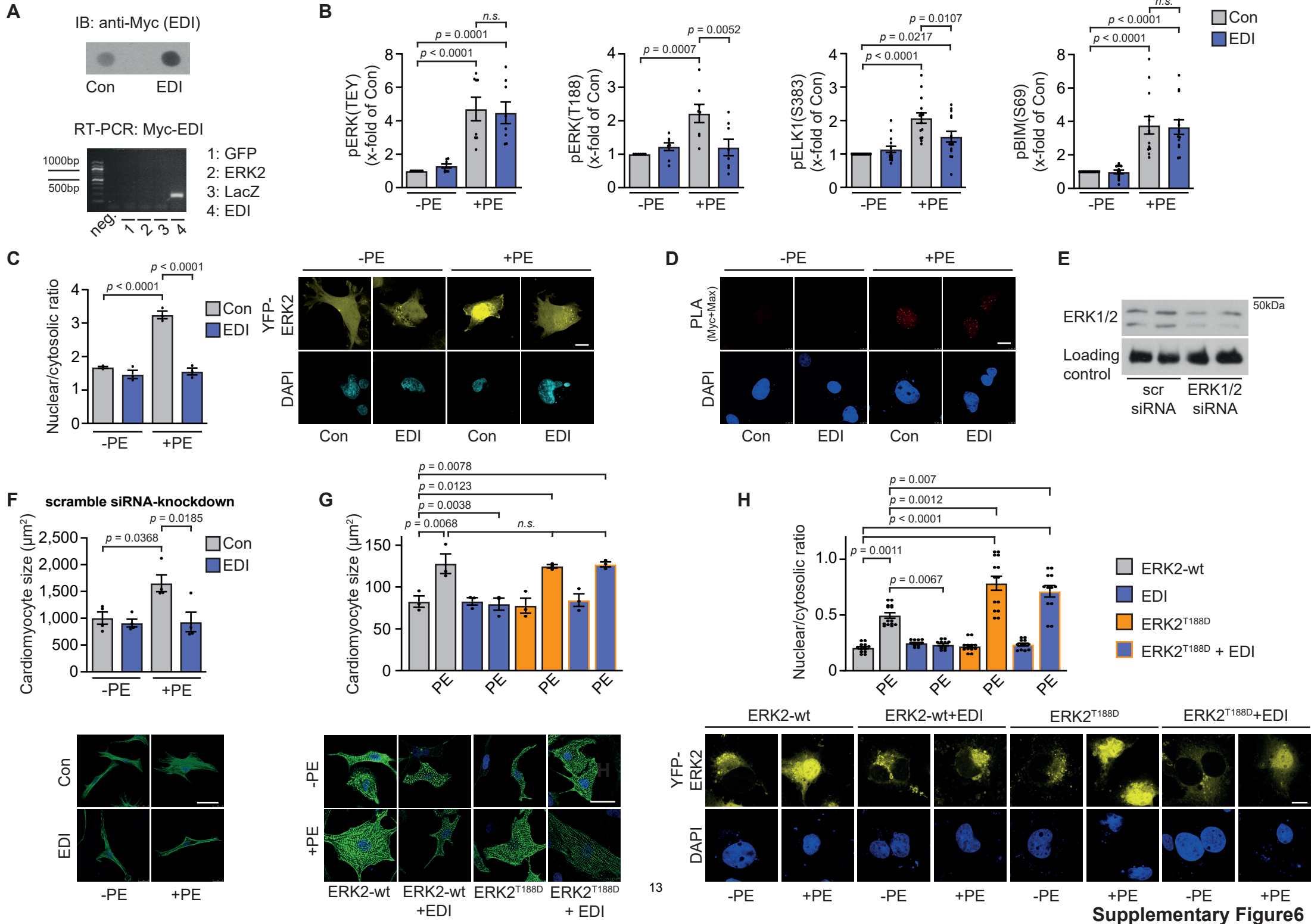
ERK_{T188}-phosphorylation is essential for nuclear accumulation of ERK2 and can be prevented by interference with ERK dimerization. (A) Quantitative analysis of nuclear-to-cytosolic ratios of YFP-tagged ERK2-wt (ERK2-wt), YFP-ERK2 Δ 174-177 (ERK2- Δ 4) and YFP-ERK2 Δ 174-177,T188D (ERK2- Δ 4D) and ERK2-wt+EDI in NRCM. Cells were stimulated with PE (4 μ M, 10min) as indicated. (30-100 cells per group of n=5). For illustration, representative confocal images displaying distribution of YFP-tagged ERK2 constructs (yellow) and DAPI-stained cell nuclei (blue) (scale bar 10 μ m) are shown. (B,C) Analysis of nuclear-to-cytosolic ratios of YFP-ERK2-wt, YFP-ERK2- Δ 4D and YFP-ERK2- Δ 4 in COS7 cells. Cells were stimulated with PE (10 μ M, 10min) as indicated. Shown are representative confocal images displaying distribution of YFP-tagged ERK2 constructs (yellow) and DAPI-stained cell nuclei (blue) (scale bar 10 μ m) and the respective quantitative analysis (60 cells per group of n=4). Error bars are mean \pm s.e.m.; n numbers represent biologically independent experiments. For statistical analysis ordinary one-way ANOVA and for (B,C) Bonferroni as *post hoc* test and for (A) Tukey test was applied. Source data are provided as a Source Data file.



Supplementary Figure 5

Characterization of a peptide that interferes with ERK dimerization. (A) Analysis of myc-ERK2₃₀₉₋₃₅₇ (EDI) expression in HEK293 (left panel) and COS7 cells (middle panel) by dot blot analysis or reverse transcriptase polymerase chain reaction (RT-PCR; right panel). Comparable transfection conditions were used for experiments in Supplementary Figure 5B-F and Supplementary Figure 6C. Dot blots and RT-PCR were reproduced three times with similar results. (B) Shown are representative co-immunoprecipitations performed using HEK293 cells transfected with HA- and Flag-ERK2 (left panel; n=5) or HA-ERK1 and Flag-ERK2 (right panel; n=3) and either pcDNA3 (Con) or myc-EDI (EDI). Flag-ERK2 or HA-ERK1 were immunoprecipitated (IP) and analyzed for co-immunoprecipitated HA-ERK2 or Flag-ERK2 in response to carbachol stimulation (CCH; 100 μ M, 10min) as indicated. (C) Representative confocal pictures of proximity ligation assays in COS7 cells. Cells were transfected with HA- and Flag-tagged ERK2-wt, and treated with either fluorescein-tagged penetratin as control or with fluorescein-tagged penetratin fused to ERK2₃₂₈₋₃₅₂ (JOLU22, left panel) or cells were transfected with HA- and Flag-tagged ERK2-wt and either myc-ERK₃₀₉₋₃₅₇ (EDI) or pcDNA3 as a control (Con) and stimulated with phenylephrine (PE; 4 μ M, 10min; n=4; scale bar 10 μ m). (D) Representative cross-linking experiment using HEK293 lysates, paraformaldehyde as crosslinking reagent and carbachol (CCH; 300 μ M, 10min) as stimulus for ERK1/2 activation. Transfection was performed as described for (B, left panel). ERK2 and its mobility shift were detected using anti-Flag and anti-ERK1/2 antibodies. Cross-linking experiment was reproduced six times with similar result. (E,F) Representative western blots and western blot analyses of ERK-phosphorylation levels in HEK293 (E) or COS7 (F) cells transfected with Flag-ERK2 and either pcDNA3 (Con) or myc-EDI (EDI) in the absence and presence of CCH (E; 300 μ M, 10min; pERK(TEY), n=5; pERK(T188), n=6) or PE (F; 4 μ M, 10min; pERK(TEY), n=8; pERK(T188), n=6). Error bars are mean \pm s.e.m.; n numbers represent biologically independent

experiments. For statistical analysis ordinary one-way ANOVA and Bonferroni as *post hoc* test was applied. Source data are provided as a Source Data file.

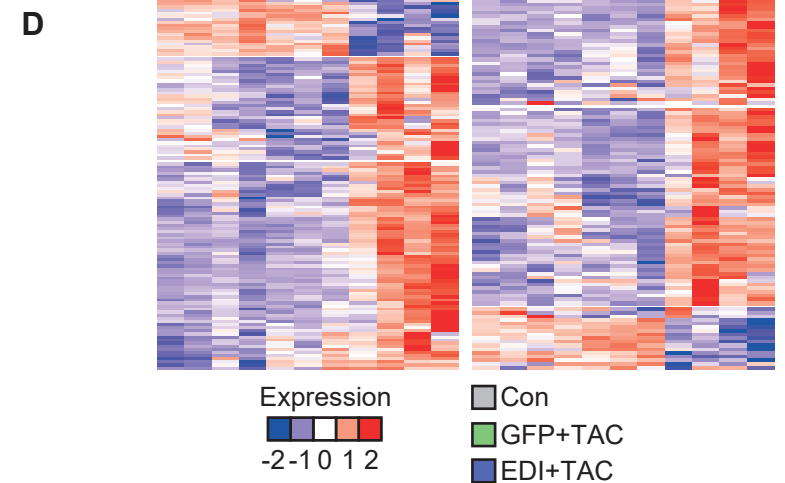
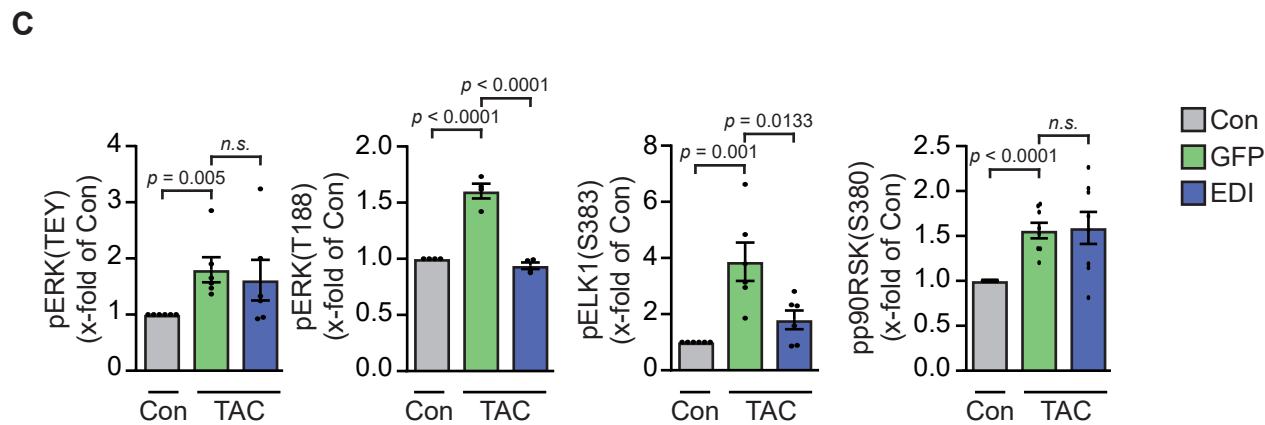
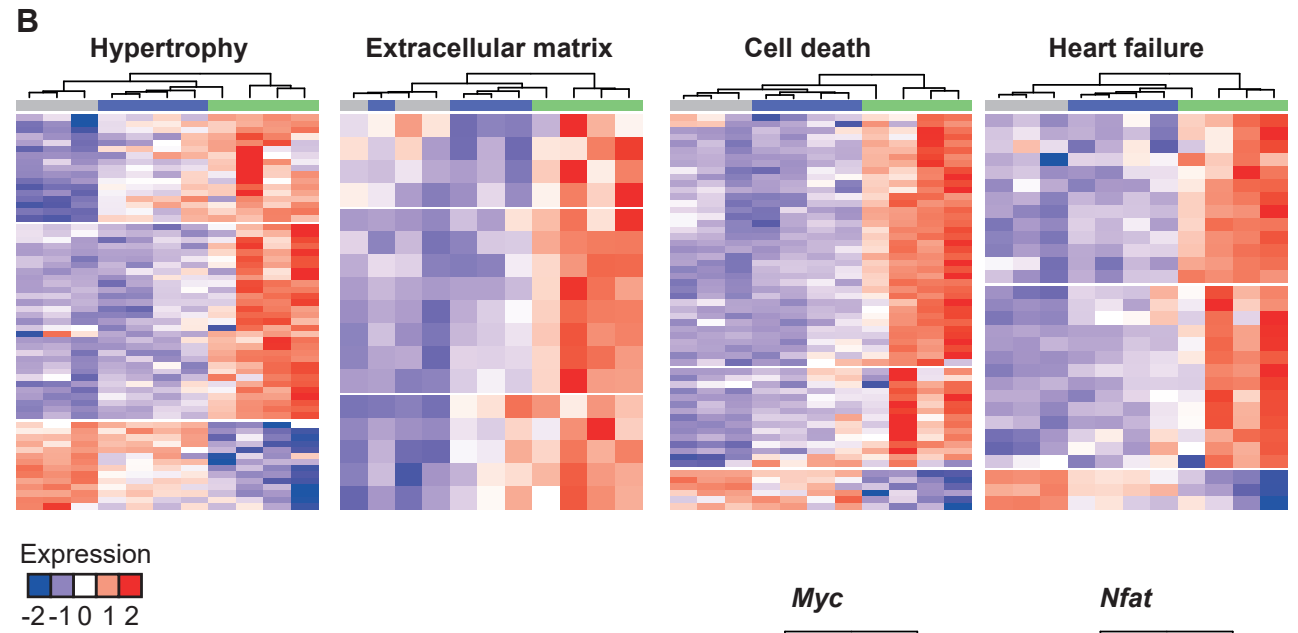
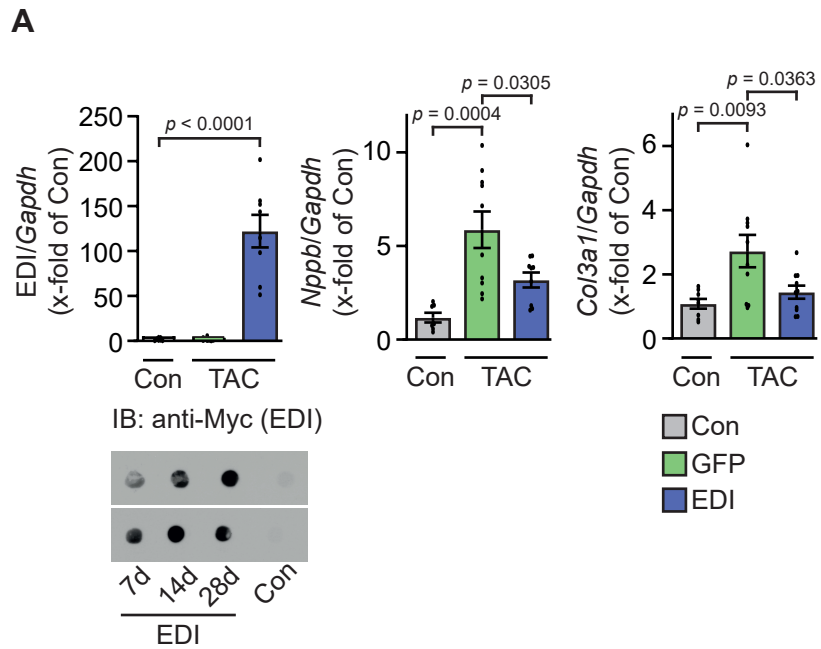


Supplementary Figure 6

Supplementary Figure 6

Functional analysis of myc-EDI. (A) Myc-EDI expression in NRCM detected by dot blot analysis and RT-PCR. NRCM were transduced with GFP (Con), myc-EDI, LacZ or Flag-ERK2-wt. Dot blot was reproduced 4 times with similar result and RT-PCR three times. (B) NRCM were transduced with LacZ (Con) or EDI virus, stimulated with PE (4 μ M, 10min) and analyzed for phosphorylation of ERK (pERK[TEY] and pERK[T188]), nuclear ELK1 (pELK[S383]) and cytosolic BIM (pBIM[S69]) by western blot (n=8, for pBIM n=13). (C) Nucleocytoplasmic distribution of YFP-ERK2-wt (yellow) in COS7 cells in the presence of EDI and PE (10 μ M, 10min); DAPI-stained nuclei (blue). Shown are representative pictures and quantitative analyses (60-70 cells per group of n=3; scale bar 10 μ m). (D) Representative confocal PLA pictures in COS7 detecting the interaction of Myc and Max in the absence and presence of EDI and PE (4 μ M, 10min; 15-20 cells per group and experiment of n=4; scale bar 10 μ m). (E) siRNA-mediated knock-down of ERK1/2 (ERK1/2 siRNA) in NRCM and scrambled siRNA (scr siRNA) analyzed by western blot (n=4). (F) NRCM size after transduction with control virus or EDI and treatment with scr siRNA and PE (4 μ M, 24h). Representative confocal pictures of phalloidin-stained cells and quantifications (55-87 cells per group of n=4; scale bar 20 μ m). (G) NRCM were transduced with Flag-tagged ERK2-wt (ERK2-wt) or Flag-tagged ERK2_{T188D} and EDI if indicated, and treated with PE (4 μ M, 24h). Representative phalloidin-stained NRCM (scale bar 25 μ m) and analyses of cell sizes (70-120 cells per group and experiment of n=3). (H) COS7 cells were transfected with YFP-tagged ERK2-wt (ERK2-wt), YFP-ERK2_{T188D} (ERK2_{T188D}) and EDI as indicated and stimulated with PE (10 μ M, 10min). Representative confocal images of YFP-ERK2 (yellow), DAPI-stained nuclei (blue; scale bar 10 μ m) and respective quantitative analyses (ERK2-wt, ERK2-wt+PE, ERK2_{T188D}, ERK2_{T188D}+PE, n=7; EDI, EDI+PE, ERK2_{T188D}+EDI, ERK2_{T188D}+EDI+PE, n=6; 30-50 cells per group of the indicated number of experiments). Error bars are mean \pm s.e.m.; n numbers represent biologically independent experiments. For statistical analysis ordinary one-

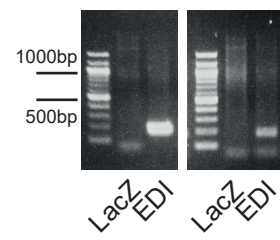
way ANOVA and as *post hoc* test Bonferroni was applied for (B,C,F) and for (G,H) the Tukey test. Source data are provided as a Source Data file.



Supplementary Figure 7

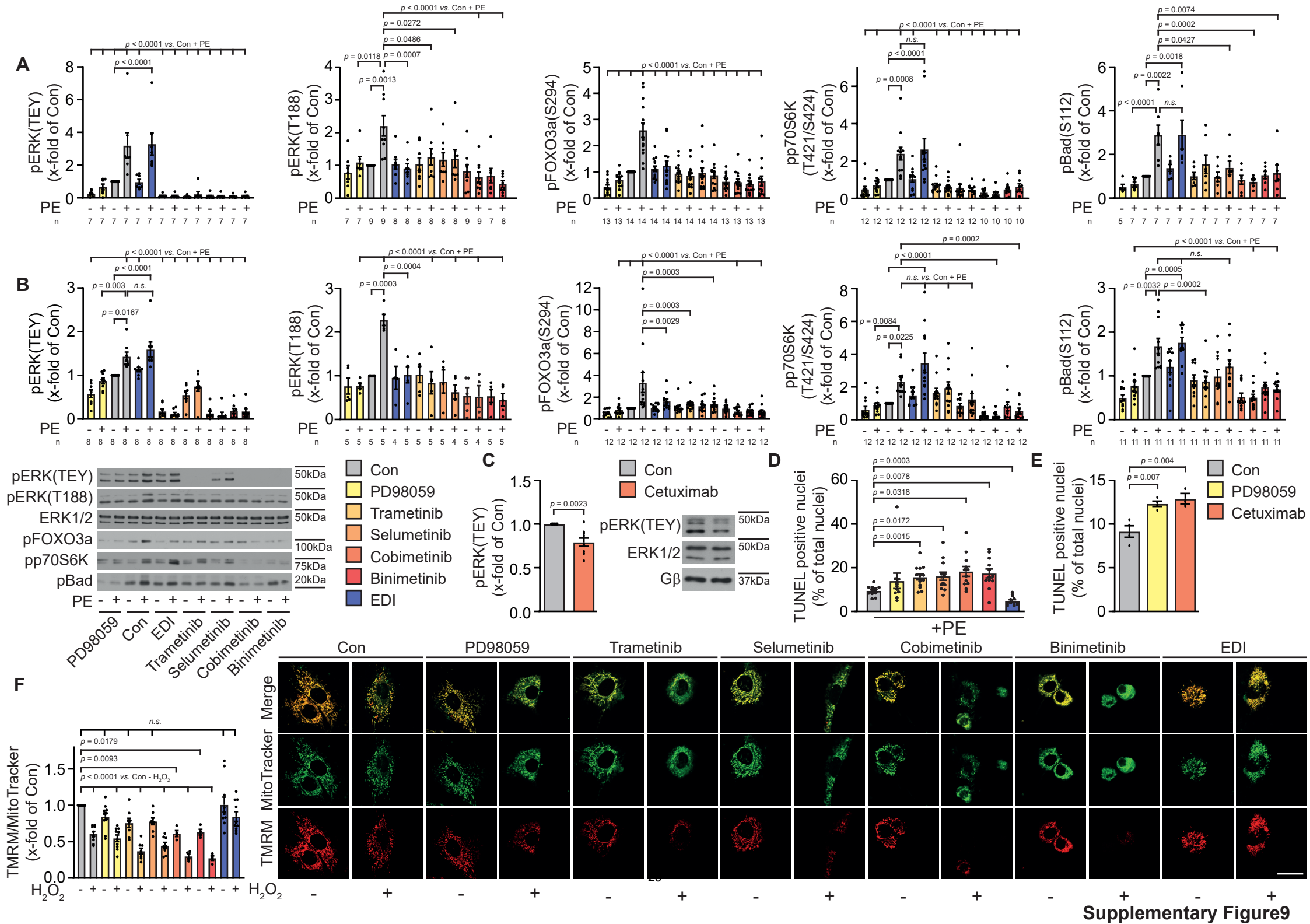
AAV9-EDI gene therapy reduces cardiac remodeling in response to TAC. (A-D) Cardiac characterization of wild-type mice before (Con; 8-week old C57/BL6 mice) and 4 weeks after transverse aortic constriction (TAC) and treatment with an adeno-associated virus serotype 9 (AAV9) vector encoding EDI (EDI) or eGFP as control (GFP) under the control of the CMV-enhanced myosin light chain promotor fragment (CMV-MLC0.26kb). **(A)** mRNA expression of myc-EDI (Con, n=7; GFP+TAC, n=11; EDI+TAC, n=8), brain natriuretic peptide (*Nppb*: Con, n=7; GFP+TAC, n=10; EDI+TAC, n=9) and collagen type III alpha 1 (*Col3a1*: Con, n=8; GFP+TAC, n=10; EDI+TAC, n=10) normalized to glyceraldehyde-3-phosphate dehydrogenase (*Gapdh*) and dot blot immunoblot analysis of myc-EDI expression (lower panel) for the indicated time points after AAV9-EDI injection (-TAC). n represents number of mice measured in triplicates. **(B)** Microarray gene expression analysis of left ventricular myocardial tissue (Con n=3, GFP and EDI n=4 mice per group). Heatmap visualization of transcriptional changes of genes related to hypertrophy, extracellular matrix, cell death and heart failure. For gene list and changes refer to Supplementary Tables 7-10. **(C)** Western blot analyses of heart lysates: pERK(TEY) (n=6) and pERK(T188) (n=4); pELK1(S383) (n=6; nuclear ERK-target) and pp90RSK(S380) (n=8; cytosolic ERK-target). **(D)** Heatmap visualization of transcriptional changes of genes related to *Nfat*- or *Myc*-signaling. Same Microarray gene expression analysis as in B. For gene list and changes refer to Supplementary Tables 11 and 12. Error bars are mean±s.e.m.; n numbers represent biologically independent animals. For statistical analysis ordinary one-way ANOVA and Tukey as *post hoc* test was applied. Source data are provided as a Source Data file.

LS174T HT29



Supplementary Figure 8

Detection of EDI in tumor cells. LS174T and HT29 colon cancer cells adenovirally transduced with LacZ or EDI. Detection of EDI-mRNA expression by RT-PCR. Shown are representative experiments of n=4. n numbers represent biologically independent experiments. Source data are provided as a Source Data file.



Supplementary Figure 9

Analysis of PD98059, newer generation MEK-inhibitors, cetuximab and EDI in cardiomyocytes. (A,B) NRCM (A) and H9c2 cells (B) were adenovirally transduced with EDI or LacZ (Con) and pretreated with **PD98059** (30 μ M), trametinib (15 μ M), selumetinib (1 μ M), cobimetinib (5 μ M) and binimetinib (10 μ M) for 1h as indicated. Shown are quantitative analyses of representative blots of Fig. 6A (A, NRCM) and H9c2 cells including representative blots (B) of pERK(TEY) and pERK(T188) and phosphorylation levels of nuclear (pFoxo3a[S294]) and cytosolic (pp70S6K[T421/S424]; pBad[S112]) ERK-targets in the absence and presence of PE (4 μ M, 10min). n numbers are given in the figures. (C) Representative western blots and western blot analysis of pERK(TEY) of untreated (Con) or cetuximab-treated NRCM (0.2 μ g/ μ l, 24h) n=9. (D, E) Analysis of TUNEL-positive cardiomyocyte nuclei. (D) NRCM untreated (Con, n=13) or treated with **PD98059** (30 μ M, n=13), trametinib (15 μ M, n=14), selumetinib (1 μ M, n=13), cobimetinib (5 μ M, n=12), binimetinib (10 μ M, n=12) for 1h or transduced with EDI (n=11) were stimulated with PE (30 μ M, 15min, \geq 100 cells per experiment and group). (E) NRCM treated with cetuximab (0.2 μ g/ μ l, 24h) or **PD98059** (30 μ M, 1h) and stimulated with H₂O₂ (100 μ M, 1h; Con, **PD98059**, n=4; cetuximab, n=3, \geq 100 cells per experiment and group). (F) H9c2 cells were adenovirally transduced with EDI (n=10) or LacZ (Con, n=10) and pretreated with **PD98059** (30 μ M, n=10), trametinib (15 μ M, n=8), selumetinib (1 μ M, n=8), cobimetinib (5 μ M, n=4) and binimetinib (10 μ M, n=4) for 1h as indicated. Assessment of mitochondrial membrane potential in the presence or absence of H₂O₂ stimulation (100 μ M, 15min). Shown are representative NRCM stained with TMRM and MitoTrackerGreen (scale bar 25 μ m) and respective quantifications (15-40 cells per experiment). Error bars are mean \pm s.e.m.; n numbers represent biologically independent experiments. For statistical analysis ANOVA was applied. For (A,B,E,F) Tukey test, for (C) paired two-sided Student's *t*-test was used. Source data are provided as a Source Data file.

Figure 2D

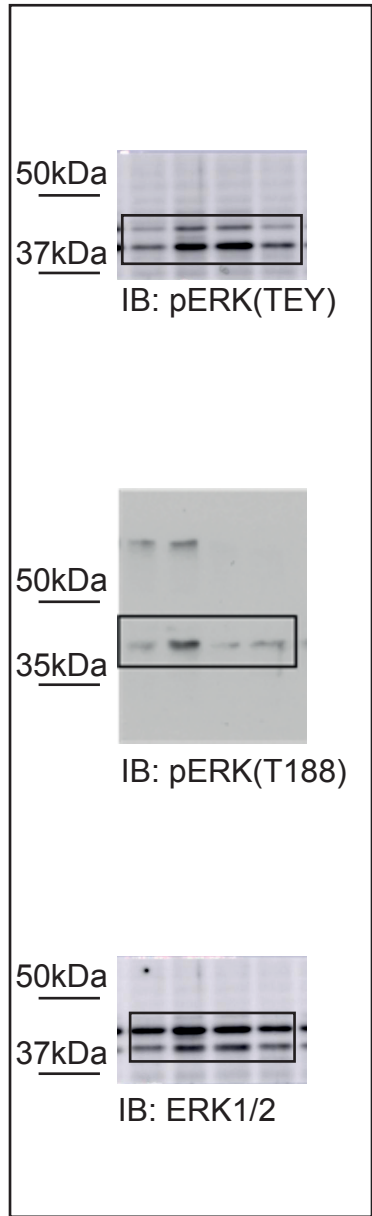


Figure 3C

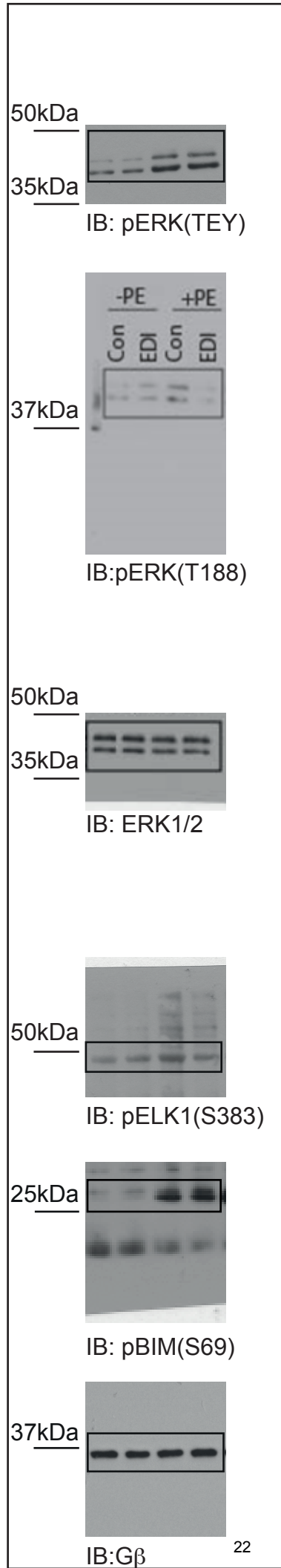


Figure 4H

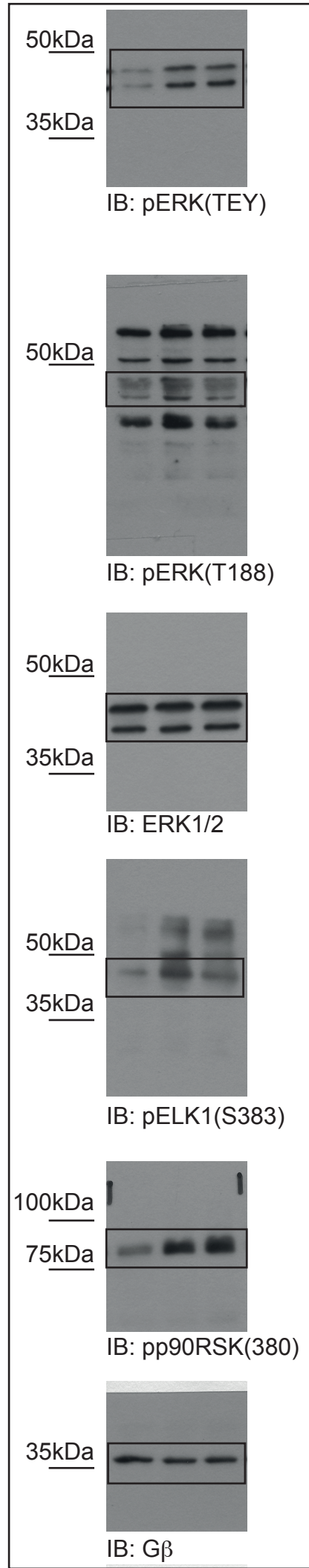


Figure 5B

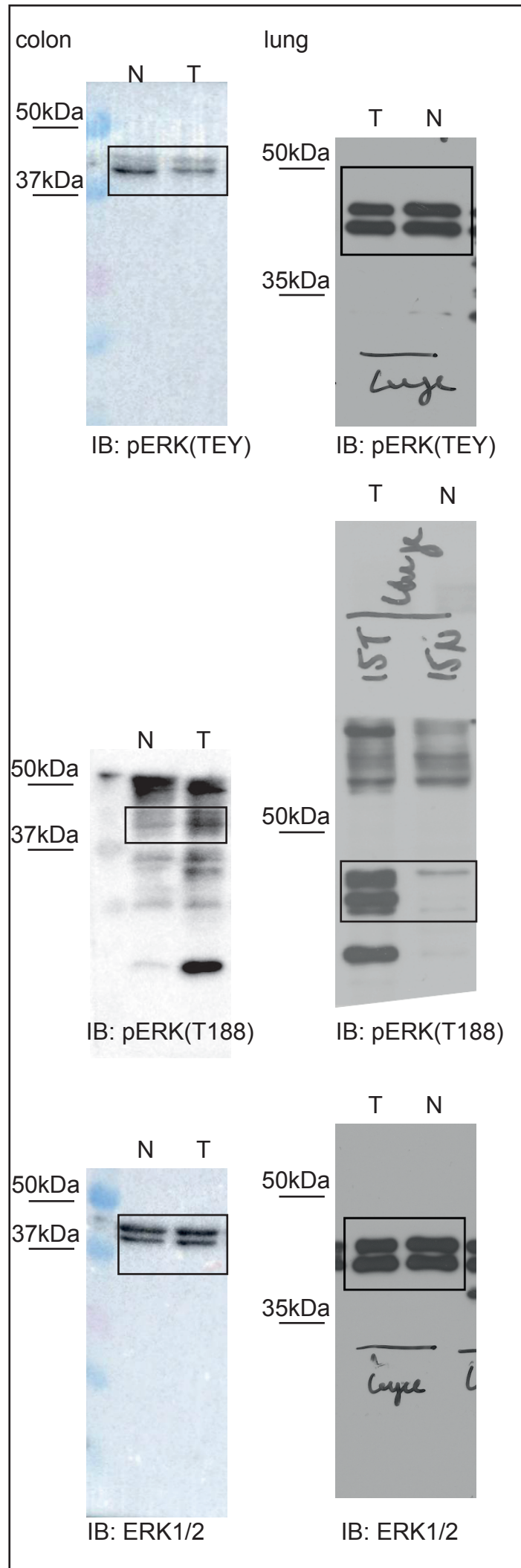


Figure 5D

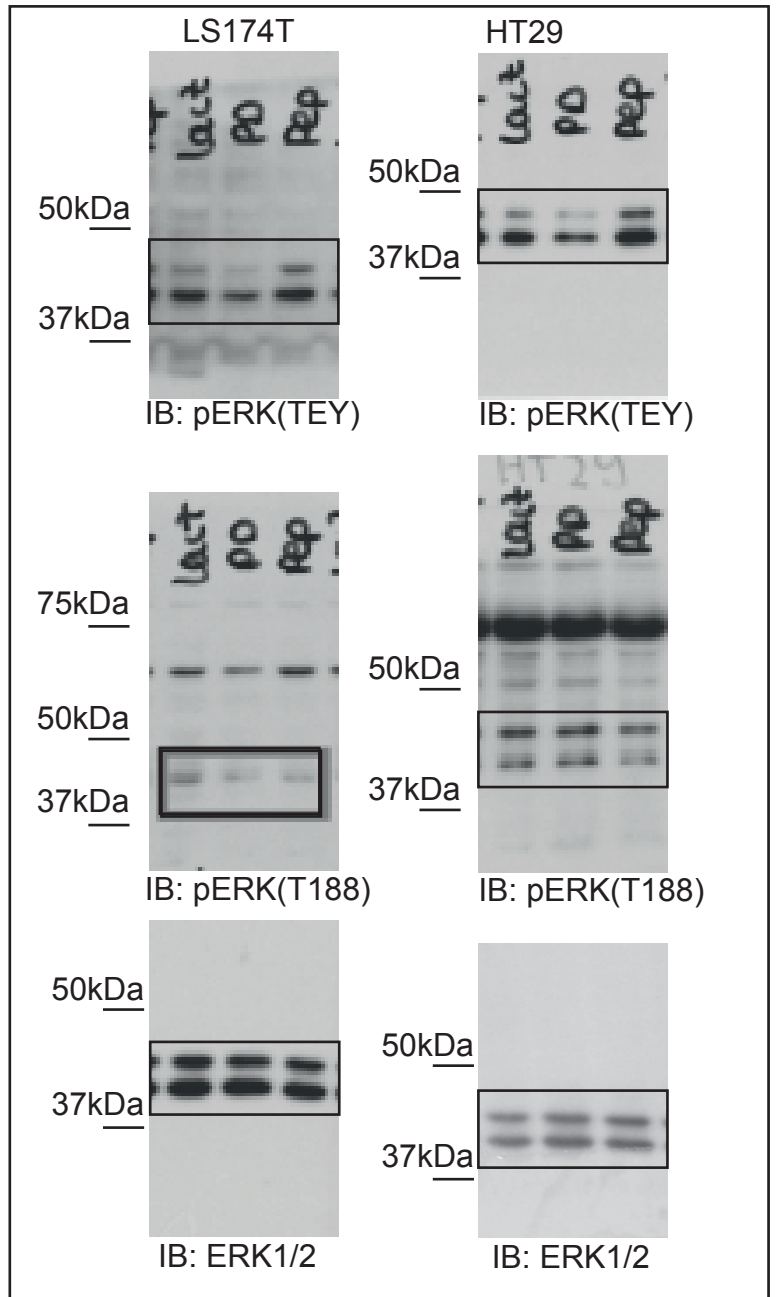


Figure 5E

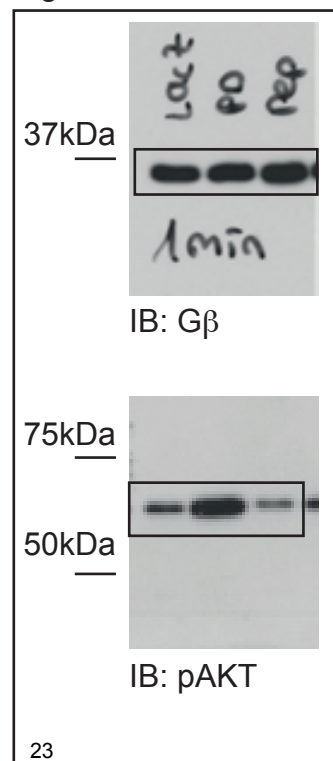
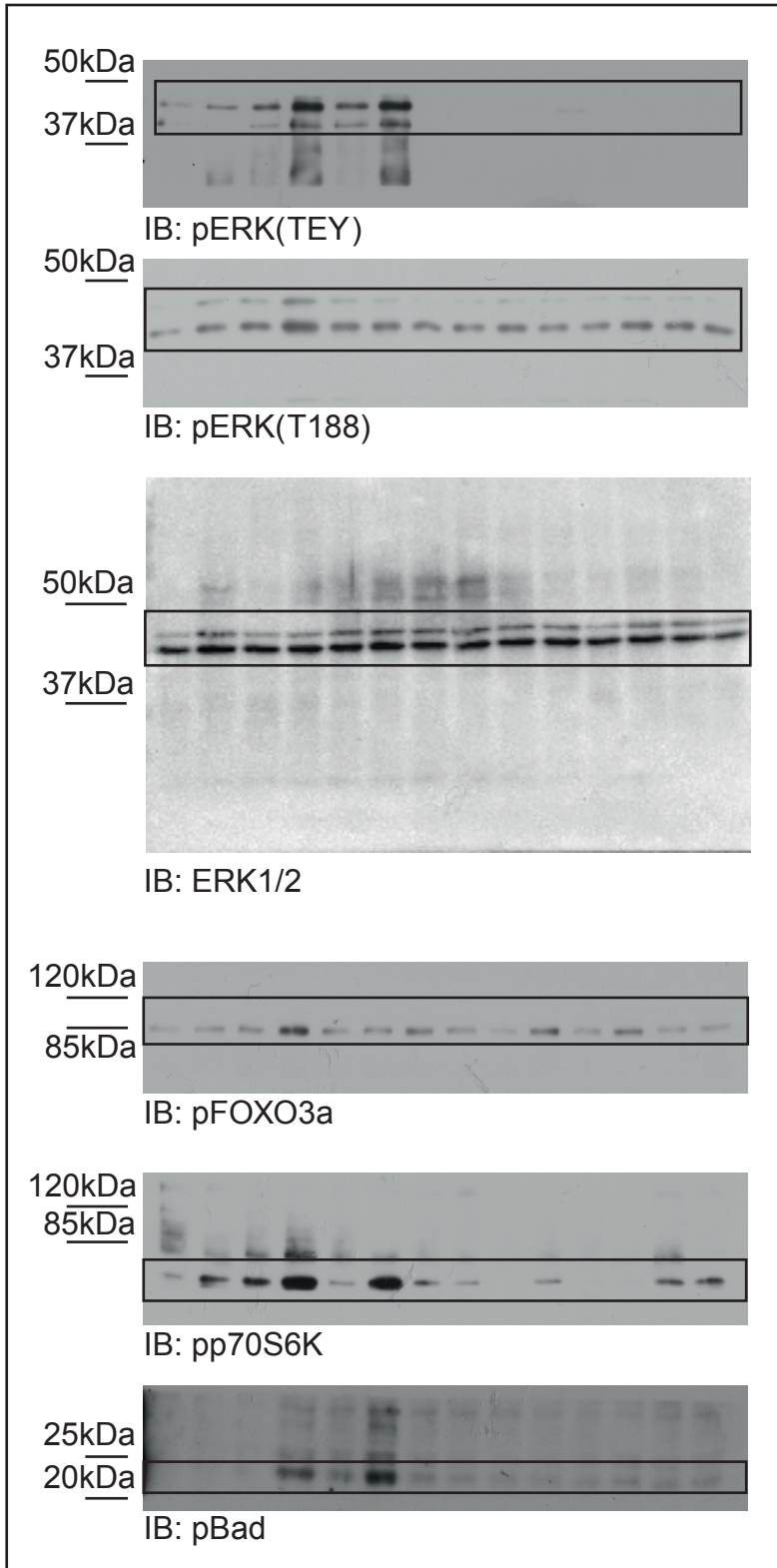
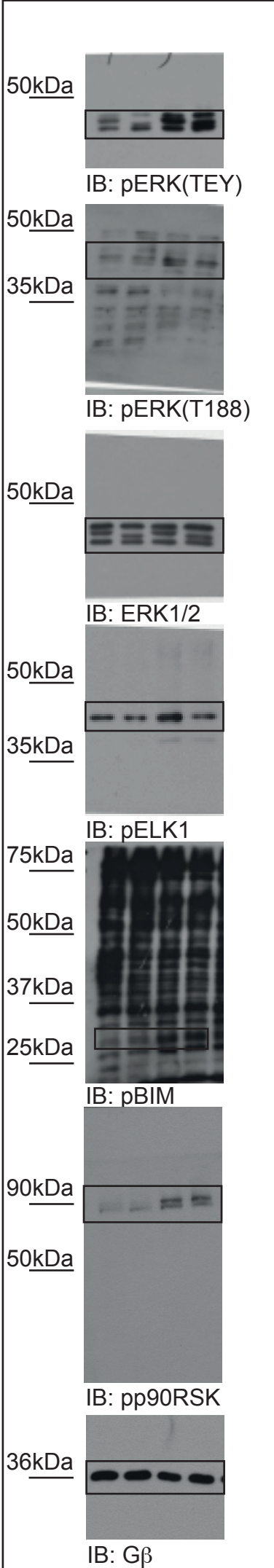


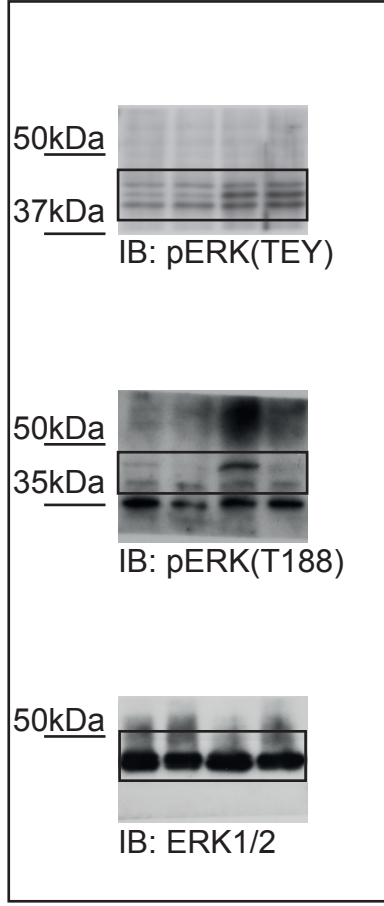
Figure 6A



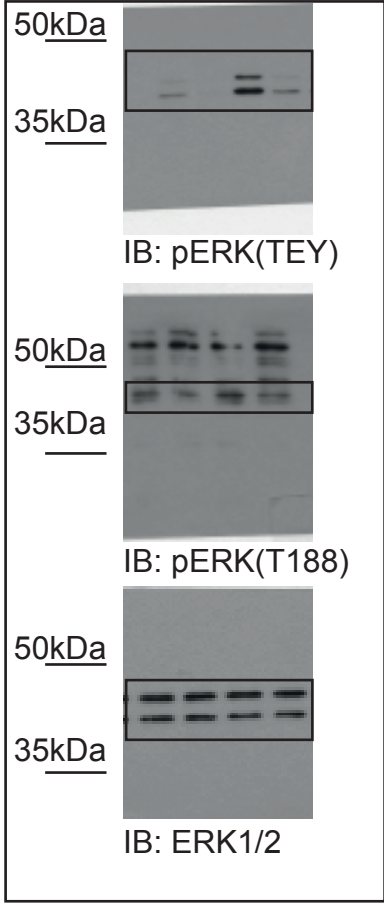
Supplementary Figure 1A



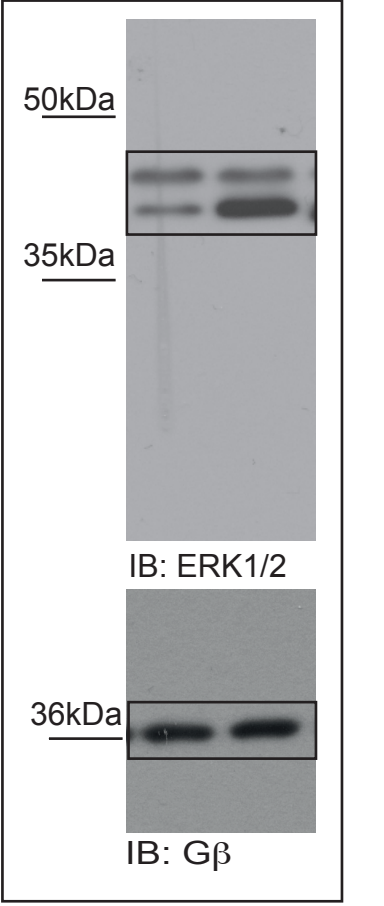
Supplementary Figure 1B



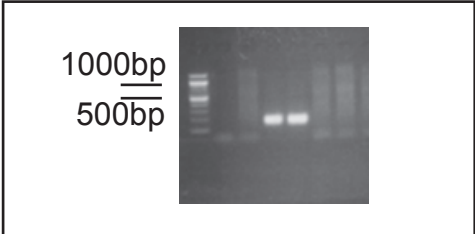
Supplementary Figure 1E



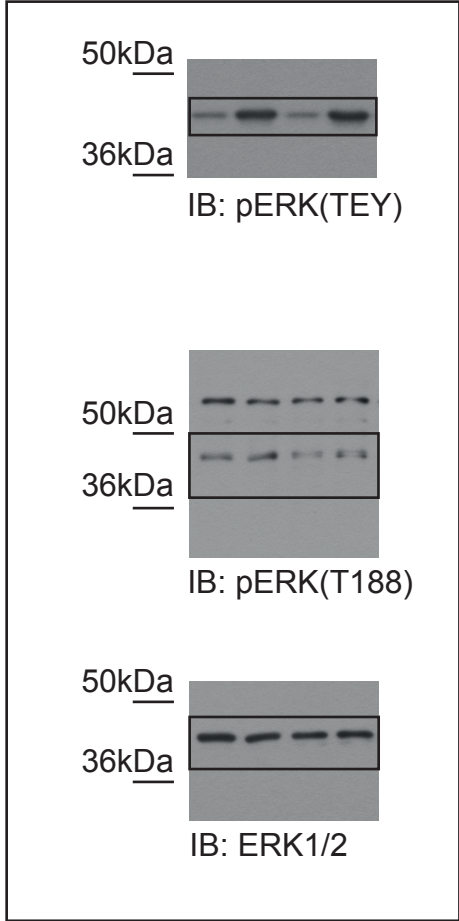
Supplementary Figure 2A



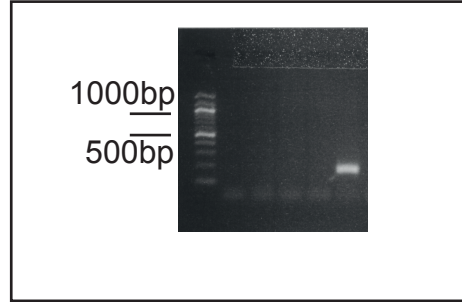
Supplementary Figure 5A



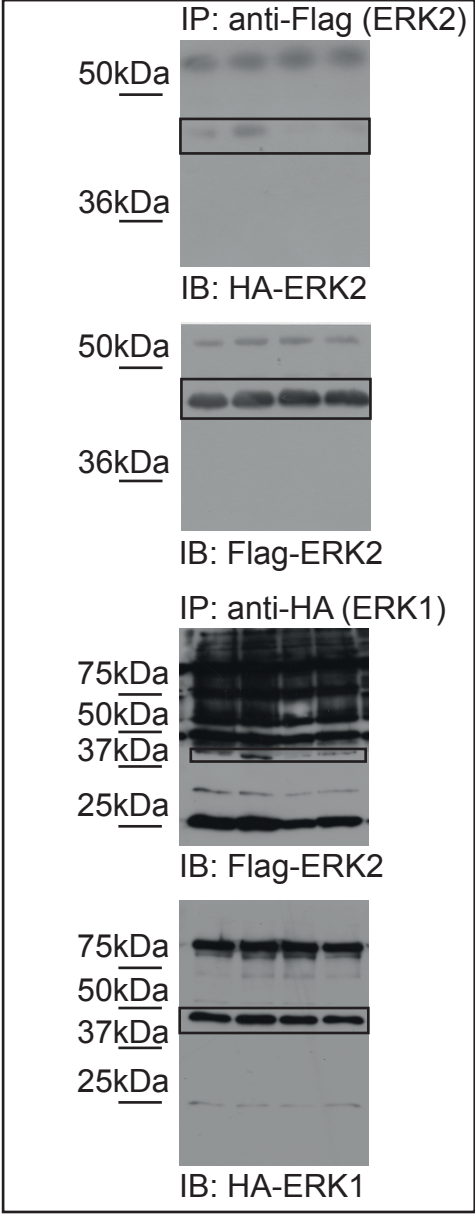
Supplementary Figure 5E



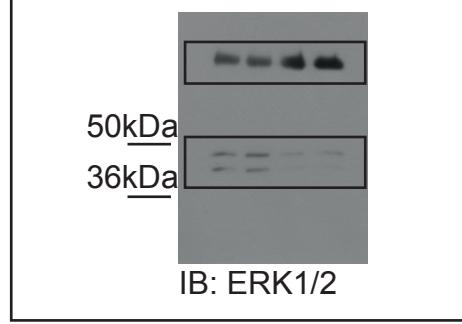
Supplementary Figure 6A



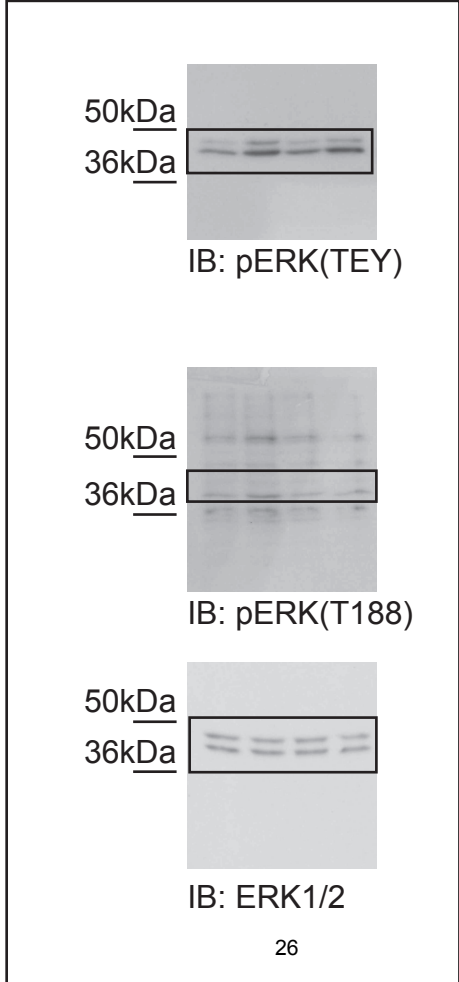
Supplementary Figure 5B



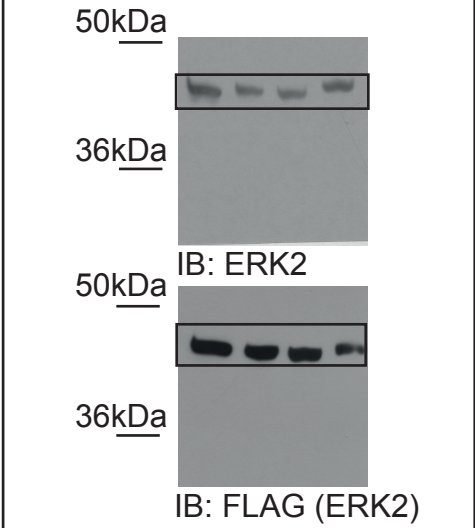
Supplementary Figure 6E

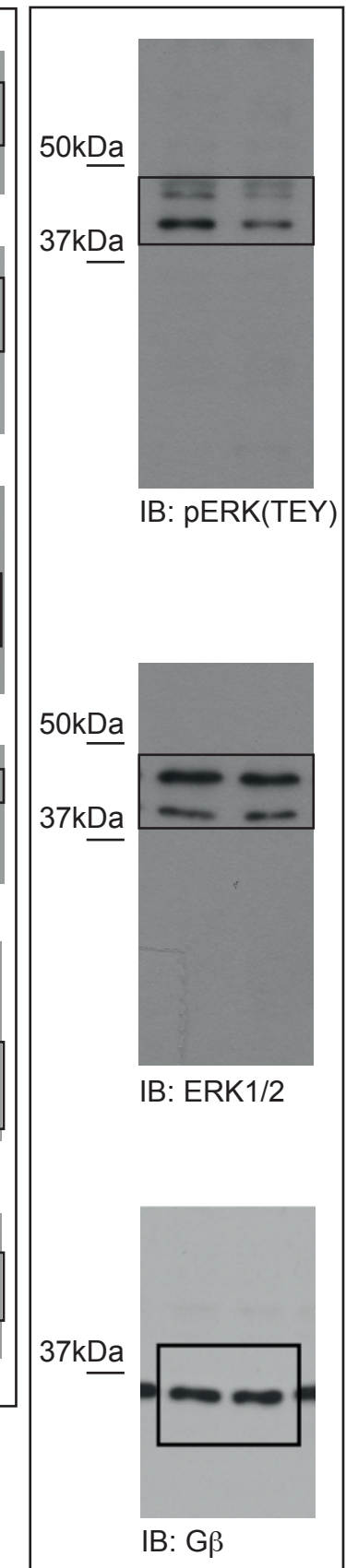
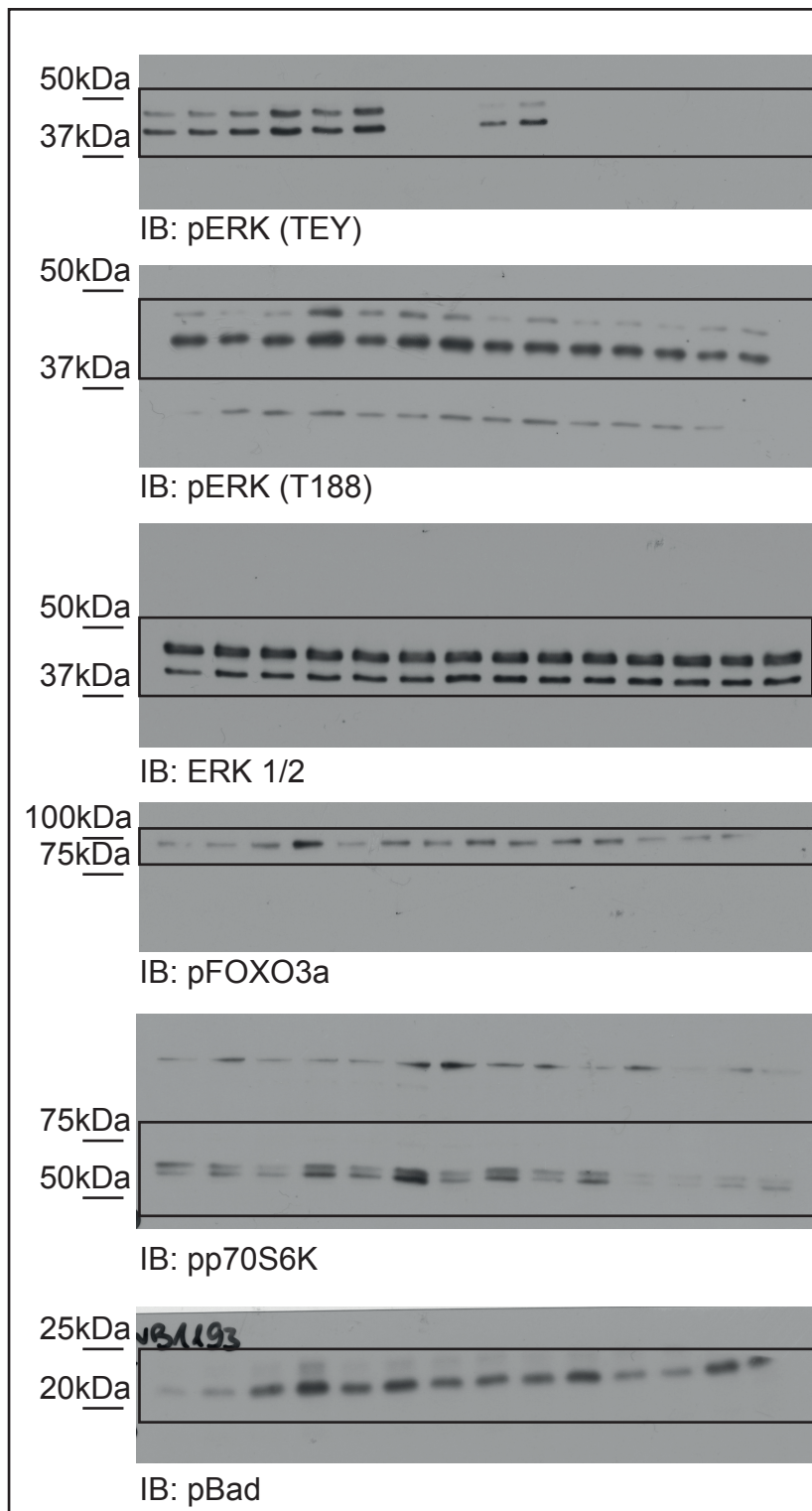
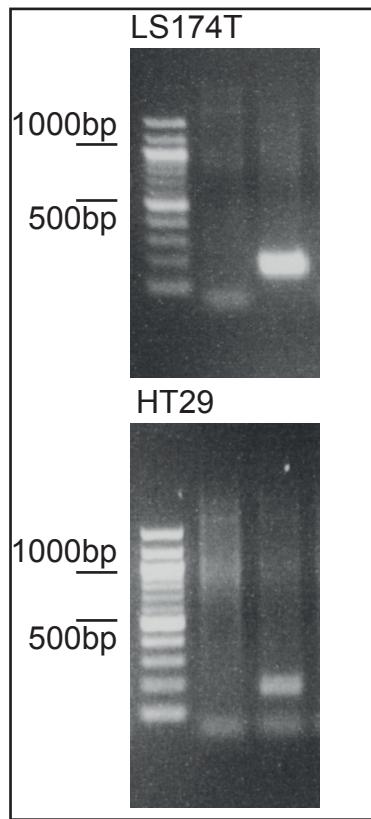


Supplementary Figure 5F



Supplementary Figure 5D





Supplementary Table 1
Characterization of cardiac function and histology of mice overexpressing monomeric ERK2 in response to TAC.

	Wt Con	ERK2-Δ4- tg Con	Wt TAC	ERK2-Δ4- tg TAC
IVS, end-diastolic (mm)	0.75 (±0.015)	0.78 (±0.014)	1.2 (±0.015)* <i>p</i> < 0.0001	1.0 (±0.014)*# <i>p</i> < 0.0001
LVPW, end-diastolic (mm)	0.77 (±0.020)	0.80 (±0.016)	1.1 (±0.023)* <i>p</i> < 0.0001	1.0 (±0.016)*# <i>p</i> < 0.0001
LVID, end-diastolic (mm)	3.3 (±0.097)	3.1 (±0.084)	3.2 (±0.074)	3.4 (±0.097)
FS (%)	38 (±1.4)	41 (±1.7)	36 (±2.0)	37 (±1.5)
EF (%)	69 (±1.8)	73 (±2.0)	66 (±2.8)	67 (1.8)
<i>n</i>-number	12	10	15	10
Heart rate (bpm)	489 (±15)	466 (±8.9)	496 (±8.8)	473 (±9.8)
Heart rate (dobutamine) (bpm)	532 (±17)	504 (±10)	518 (±8.1)	490 (±8.6)
LVP_{max} (mmHg)	94 (±3.6)	92 (±3.3)	191 (±5.8)* <i>p</i> < 0.0001	179 (±6.1)* <i>p</i> < 0.0001
LVP_{max} (dobutamine) (mmHg)	100 (±3.5)	102 (±3.3)	201 (±6.3)* <i>p</i> < 0.0001	192 (±3.8)* <i>p</i> < 0.0001
dp/dt_{min} (mmHg s⁻¹)	-6239 (±375)	-5751 (±353)	-9876 (±267)* <i>p</i> < 0.0001	-9538 (±433)* <i>p</i> < 0.0001
dp/dt_{min} (dobutamine) (mmHg s⁻¹)	-6795 (±340)	-7110 (±366)	-10432 (±336)* <i>p</i> < 0.0001	-10398 (±377)* <i>p</i> < 0.0001
dp/dt_{max} (mmHg s⁻¹)	6761 (±380)	6431 (±499)	8204 (±239)* <i>p</i> = 0.0107	8087 (±485)
dp/dt_{max} (dobutamine) (mmHg s⁻¹)	8334 (±378)	8459 (±527)	9122 (±325)	9253 (±391)
<i>n</i>-number	10	12	13	8

	Wt Con	ERK2-Δ4-tg Con	Wt TAC	ERK2-Δ4-tg TAC
HW/TL (mg mm⁻¹)	5.7 (±0.12)	5.6 (±0.099)	10.8 (±0.25)* <i>p</i> < 0.0001	9.0 (±0.47)*# <i>p</i> < 0.0001
HW/BW (mg g⁻¹)	4.2 (±0.10)	4.0 (±0.064)	7.1 (±0.16)* <i>p</i> < 0.0001	5.9 (±0.33)*# <i>p</i> < 0.0001
<i>n</i>-number	13	10	20	10
Interstitial fibrosis (A.U.)	0.015 (±0.0042) <i>n</i> =8	0.014 (±0.0015) <i>n</i> =7	0.043 (±0.010)* <i>p</i> = 0.0214 <i>n</i> =10	0.016 (±0.0021)*# <i>p</i> = 0.0173 <i>n</i> =10
Cardiomyocyte cross-sectional area (μm²)	201 (±11) <i>n</i> =8	192 (±11) <i>n</i> =9	308 (±24)*, <i>p</i> = 0.0003 <i>n</i> =10	227 (±10)*# <i>p</i> = 0.0064 <i>n</i> =9

Supplementary Table 1

Characterization of cardiac function and histology of mice overexpressing monomeric ERK2 in response to TAC. Characterization of wild-type (Wt) and ERK2 Δ 174-177 (ERK2-Δ4-tg) transgenic mice before and after TAC surgery (6 weeks). Shown are echocardiographic measurements of end-diastolic wall thicknesses (interventricular septal [IVS] and left ventricular posterior wall [LVPW]), left ventricular internal diameter (LVID), fractional shortening (FS), ejection fraction (EF) and heart rates; data derived from left ventricular cardiac catheterization with dobutamine dose response curves for dp/dt_{max} and dp/dt_{min} as determined for left ventricular pressure; heart weight-to-tibia length (HW/TL) and heart weight-to-body weight (HW/BW) ratios and data from analyses of fibrosis of Sirius Red-stained histological sections and data on cardiomyocyte cross-sectional area determined using H&E-stained histological sections of left ventricles. Values represent mean \pm s.e.m.; *n*-numbers represent biologically independent animals; * *vs.* Wt control mice, # *vs.* Wt TAC mice. For statistical analysis ordinary one-way ANOVA was applied; for LVID, end-diastolic and EF Tukey test

and for all other parameters Bonferroni was applied as *post hoc* test. Source data are provided as a Source Data file.

Supplementary Table 2**Characterization of cardiac function of Wt and ERK2- Δ 4 transgenic mice at the age of 9 months.**

	Wt 9 months	ERK2-Δ4- tg 9 months
IVS, end-diastolic (mm)	0.87 (\pm 0.016)	0.81 (\pm 0.036)
LVPW, end-diastolic (mm)	0.96 (\pm 0.017)	0.9 (\pm 0.025)
LVID, end-diastolic (mm)	3.5 (\pm 0.075)	3.5 (\pm 0.083)
FS (%)	38 (\pm 1.5)	35 (\pm 2.3)
EF (%)	69 (\pm 1.9)	65 (\pm 3.2)
Heart rate (bpm)	539 (\pm 8.5)	506 (\pm 16)
HW/TL (mg mm⁻¹)	7.6 (\pm 0.23)	7.1 (\pm 0.12)
HW/BW (mg g⁻¹)	3.9 (\pm 0.058)	3.7 (\pm 0.069)
<i>n</i>-number	9	9

Supplementary Table 2**Characterization of cardiac function of Wt and ERK2- Δ 4 transgenic mice at the age of 9**

months. Characterization of wild-type (Wt) and ERK2 Δ ₁₇₄₋₁₇₇ (ERK2- Δ 4) transgenic mice at the age of 9 months. Shown are echocardiographic measurements of end-diastolic wall thicknesses (interventricular septal [IVS] and left ventricular posterior wall [LVPW]), left ventricular internal diameter (LVID), fractional shortening (FS), ejection fraction (EF) and heart rates, heart weight-to-tibia length (HW/TL) and heart weight-to-body weight (HW/BW) ratios. Values represent mean \pm s.e.m.. *n*-numbers represent biologically independent animals. For statistical analysis unpaired and two-sided Student's *t*-test was applied. Source data are provided as a Source Data file.

Supplementary Table 3

Characterization of Wt and ERK2-Δ4 transgenic mice after 3 weeks of voluntary running.

	Wt Con	ERK2-Δ4- tg Con	Wt RUN	ERK2-Δ4- tg RUN
IVS, end-diastolic (mm)	0.73 (±0.018)	0.73 (±0.015)	0.82 (±0.016)* <i>p</i> = 0.0087	0.85 (±0.018)* <i>p</i> = 0.0003
LVPW, end-diastolic (mm)	0.71 (±0.022)	0.71 (±0.016)	0.82 (±0.019)* <i>p</i> = 0.0046	0.84 (±0.014)* <i>p</i> = 0.0002
LVID, end-diastolic (mm)	3.3 (±0.098)	3.1 (±0.047)	3.3 (±0.071)	3.2 (±0.069)
FS (%)	39 (±1.5)	39 (±0.84)	38 (±2.0)	39 (±1.7)
EF (%)	63 (±4.7)	71 (±1.1)	62 (±5.4)	70 (±2.0)
Heart rate (bpm)	524 (±18)	545 (±11)	541 (±14)	520 (±15)
Running distance			371078 (±40582)	351894 (±45195)
<i>n</i>-number	8	16	9	12
HW/TL (mg mm⁻¹)	5.7 (±0.12)	5.6 (±0.099)	6.2 (±0.092)* <i>p</i> = 0.0164	6.2 (±0.12)* <i>p</i> = 0.004
HW/BW (mg g⁻¹)	4.2 (±0.10)	4.0 (±0.064)	4.5 (±0.045)	4.6 (±0.17)
<i>n</i>-number	13	10	9	13
Interstitial fibrosis (A.U.)	0.90 (±0.63) <i>n</i> =5	0.53 (±0.12) <i>n</i> =5	0.87 (±0.34) <i>n</i> =6	0.91 (±0.39) <i>n</i> =6
Cardiomyocyte cross-sectional area (μm²)	290 (±22) <i>n</i> =7	287 (±10) <i>n</i> =7	386 (±10)* <i>p</i> = 0.001 <i>n</i> =6	368 (±12)* <i>p</i> = 0.004 <i>n</i> =8

Supplementary Table 3

Characterization of Wt and ERK2-Δ4 transgenic mice after 3 weeks of voluntary running.

Characterization of wild-type (Wt) and ERK2 Δ 174-177 (ERK2-Δ4-tg) transgenic mice before and after voluntary exercise in a running wheel (RUN) (3 weeks). Shown are echocardiographic measurements of end-diastolic wall thicknesses (interventricular septal [IVS] and left ventricular posterior wall [LVPW]), left ventricular internal diameter (LVID), fractional shortening (FS), ejection fraction (EF) and heart rates; heart weight-to-tibia length (HW/TL) and heart weight-to-body weight (HW/BW) ratios as well as and data from analyses of interstitial fibrosis of Sirius Red-stained histological sections and data on cardiomyocyte cross-sectional area determined using H&E-stained histological sections of left ventricles. Values represent mean \pm s.e.m., *n*-numbers represent biologically independent animals; * vs. Wt control mice. For statistical analysis ordinary one-way ANOVA was applied, Bonferroni was applied as *post hoc* test. Source data are provided as a Source Data file.

Supplementary Table 4

Analysis of Wt mice 4 weeks after TAC surgery and AAV9-EDI treatment.

	Wt Con	Wt + AAV9-GFP TAC	Wt + AAV-EDI
IVS, end-diastolic (mm)	0,64 (±0.013)	1,0 (±0.024)* <i>p</i> < 0.0001	0,80 (±0.019)*# <i>p</i> < 0.0001
LVPW, end-diastolic (mm)	0,69 (±0.014) <i>p</i> < 0.0001	0,99 (±0.030)* <i>p</i> < 0.0001	0,82 (±0.0090)*# <i>p</i> < 0.0001
LVID, end-diastolic (mm)	3,6 (±0.063)	3,9 (±0.060)* <i>p</i> = 0.011	3,6 (±0.055)# <i>p</i> = 0.0213
FS (% of LVID)	35 (±1.6)	22 (±1.2)* <i>p</i> < 0.0001	36 (±1.5)# <i>p</i> < 0.0001
EF (%)	64 (±2.1)	45 (±2.1)* <i>p</i> < 0.0001	66 (±1.9)# <i>p</i> < 0.0001
Heart rate (bpm)	509 (±12)	499 (±18)	531 (±17)
<i>n</i>-number	19	11	14
HW/TL (mg mm⁻¹)	5.2 (±0.17)	9.5 (±0.47)* <i>p</i> < 0.0001	7.6 (±0.29)*# * <i>p</i> = 0.0004 # <i>p</i> = 0.0013
HW/BW (mg g⁻¹)	4.3 (±0.12)	7.6 (±0.38)* <i>p</i> = 0.0054	3.2 (±0.93)# <i>p</i> < 0.0001
LW/TL (mg mm⁻¹)	7.7 (±0.12)	9.1 (±0.23)* <i>p</i> = 0.0018	7.9 (±0.30)# <i>p</i> = 0.0032
<i>n</i>-number	8	11	11
Interstitial fibrosis (A.U.)	0.38 (±0.065) <i>n</i> =7	0.79 (±0.13)* <i>p</i> = 0.0467 <i>n</i> =11	0.32 (±0.085)# <i>p</i> = 0.0076 <i>n</i> =11
Cardiomyocyte cross- sectional area (µm²)	175 (±5.2) <i>n</i> =9	297 (±12)* <i>p</i> < 0.0001 <i>n</i> =11	228 (±13)*# * <i>p</i> = 0.0102 # <i>p</i> = 0.0004 <i>n</i> =11

Supplementary Table 4

Analysis of Wt mice 4 weeks after TAC surgery and AAV9-EDI treatment. Basic echocardiographic analyses of untreated wild-type (Wt) and AAV9-EDI or AAV9-GFP treated mice after transverse aortic constriction (TAC; 4 weeks). Shown are echocardiographic measurements of end-diastolic wall thicknesses (interventricular septal [IVS] and left ventricular posterior wall [LVPW]), left ventricular internal diameter (LVID), fractional shortening (FS), ejection fraction (EF) and heart rates; heart weight to tibia length (HW/TL), heart weight-to-body weight (HW/BW) and lung weight-to-tibia length (LW/TL) ratios as well as data from analyses of interstitial fibrosis of Sirius Red-stained histological sections and data on cardiomyocyte cross-sectional area determined using H&E-stained histological sections of left ventricles. Values represent mean \pm s.e.m., *n*-numbers represent biologically independent animals; * *vs.* Wt control mice, # *vs.* AAV9-GFP TAC mice. For statistical analysis ordinary one-way ANOVA was applied; for HW/TL, HW/BW, LW/TL, interstitial fibrosis, cardiomyocyte cross-sectional area Bonferroni, for FS Sidak, for IVS, end-diastolic, LVPW, end-diastolic LVID, end-diastolic, EF and heart rate Tukey test was applied as *post hoc* test. Source data are provided as a Source Data file.

Supplementary Table 5
Analysis of wild-type mice after AAV9-EDI treatment.

	Wt Con	Wt + AAV9-GFP	Wt + AAV9-EDI
IVS, end-diastolic (mm)	0.65 (\pm 0.016)	0.70 (\pm 0.031)	0.69 (\pm 0.012)
LVPW, end-diastolic (mm)	0.66 (\pm 0.014)	0.65 (\pm 0.0010)	0.66 (\pm 0.012)
LVID, end-diastolic (mm)	3.5 (\pm 0.058)	3.7 (\pm 0.090)	3.4 (\pm 0.059)
FS (% of LVID)	40 (\pm 1.7)	40 (\pm 3.5)	46 (\pm 1.6)
EF (%)	71 (\pm 2.2)	71 (\pm 4.3)	78 (\pm 1.7)
Heart rate (bpm)	621 (\pm 15)	613 (\pm 39)	634 (\pm 26)
<i>n</i>-number	18	6	8
HW/TL (mg mm⁻¹)	5.8 (\pm 0.095)	6.1 (\pm 0.12)	6.1 (\pm 0.29)
HW/BW (mg mg⁻¹)	4.2 (\pm 0.10)	4.3 (\pm 0.067)	4.5 (\pm 0.15)
LW/TL (mg mm⁻¹)	8.0 (\pm 0.35)	9.2 (\pm 0.97)	8.9 (\pm 1.0)
<i>n</i>-number	11 (LW/TL12)	7	8
Interstitial fibrosis (A.U.)	0.09 (\pm 0.014) <i>n</i> =6	0.11 (\pm 0.023) <i>n</i> =6	0.09 (\pm 0.019) <i>n</i> =8
Cardiomyocyte cross- sectional area (μm²)	281 (\pm 21) <i>n</i> =9	310 (\pm 23) <i>n</i> =7	297 (\pm 21) <i>n</i> =8

Supplementary Table 5

Analysis of wild-type mice after AAV9-EDI treatment. Echocardiographic analyses of wild-type (Wt) and Wt mice 4 weeks after AAV9-EDI or AAV9-GFP treatment. Shown are echocardiographic measurements of end-diastolic wall thicknesses (interventricular septal

[IVS] and left ventricular posterior wall [LVPW]), left ventricular internal diameter (LVID), fractional shortening (FS), ejection fraction (EF) and heart rates; heart weight-to-tibia length (HW/TL), heart weight-to-body weight (HW/BW) and lung weight to tibia length (LW/TL) ratios as well as data from analyses of interstitial fibrosis of Sirius Red-stained histological sections and data on cardiomyocyte cross-sectional area determined using H&E-stained histological sections of left ventricles. Values represent mean \pm s.e.m.; *n*-numbers represent biologically independent animals. For statistical analysis ordinary one-way ANOVA and Bonferroni as *post hoc* test was applied. Tukey test was used as *post hoc* test. Source data are provided as a Source Data file.

Supplementary Table 6

Top 100 regulated genes in wild-type mice 4 weeks after TAC surgery and AAV9-EDI treatment. Table refers to heat map shown in Fig. 4G.

Gene symbol	Gene name	Fold change (<i>P</i> -value)		
		AAV9-GFP TAC vs Con	AAV9-EDI TAC vs Con	AAV9-EDI TAC vs AAV9-GFP TAC
<i>Cilp</i>	cartilage intermediate layer protein, nucleotide pyrophosphohydrolase	17.6 (1.37E-04)	3.37 (1.22E-01)	-5.21 (2.13E-03)
<i>Ltbp2</i>	latent transforming growth factor beta binding protein 2	9.91 (2.80E-03)	1.89 (7.39E-01)	-5.24 (1.42E-02)
<i>Postn</i>	periostin, osteoblast specific factor	8.35 (9.57E-05)	1.90 (2.66E-01)	-4.40 (9.31E-04)
<i>S100a9</i>	S100 calcium binding protein A9 (calgranulin B)	8.04 (5.03E-02)	4.96 (5.02E-01)	-1.62 (7.63E-01)
<i>Serpina3n</i>	serine (or cysteine) peptidase inhibitor, clade A, member 3N	5.95 (4.31E-02)	2.78 (6.57E-01)	-2.14 (4.40E-01)
<i>Crlf1</i>	cytokine receptor-like factor 1	5.70 (2.12E-03)	1.73 (6.60E-01)	-3.29 (1.38E-02)
<i>S100a8</i>	S100 calcium binding protein A8 (calgranulin A)	5.18 (1.13E-01)	3.32 (6.64E-01)	-1.56 (7.87E-01)
<i>Col8a1</i>	Collagen, type VIII, alpha 1	5.13 (1.60E-04)	1.52 (4.76E-01)	-3.37 (9.91E-04)
<i>Thbs4</i>	thrombospondin 4	5.06 (5.60E-03)	1.75 (7.03E-01)	-2.89 (3.76E-02)
<i>Mfap4</i>	microfibrillar-associated protein 4	4.78 (5.18E-04)	1.55 (5.71E-01)	-3.09 (2.74E-03)
<i>Cdkn1a</i>	cyclin-dependent kinase inhibitor 1A (P21)	4.46 (5.33E-03)	1.51 (7.79E-01)	-2.95 (2.36E-02)
<i>Mybpc2</i>	myosin binding protein C, fast-type	4.16 (3.41E-04)	2.73 (4.60E-02)	-1.52 (1.06E-01)
<i>Itgbl1</i>	Integrin, beta-like 1	4.02 (1.09E-03)	1.69 (4.65E-01)	-2.38 (1.26E-02)
<i>Prg4</i>	proteoglycan 4 (megakaryocyte stimulating factor, articular superficial zone protein)	3.84 (2.67E-02)	2.36 (5.12E-01)	-1.63 (4.70E-01)
<i>Ccl8</i>	chemokine (C-C motif) ligand 8	3.79 (3.58E-03)	1.99 (3.99E-01)	-1.91 (9.45E-02)
<i>Mt2</i>	metallothionein 2	3.72 (3.35E-03)	1.56 (6.68E-01)	-2.38 (2.56E-02)

Gene symbol	Gene name	Fold change (<i>P</i> -value)		
		AAV9-GFP TAC vs Con	AAV9-EDI TAC vs Con	AAV9-EDI TAC vs AAV9-GFP TAC
<i>Nppa</i>	natriuretic peptide type A	3.57 (1.37E-04)	2.02 (5.68E-02)	-1.77 (7.98E-03)
<i>Nppb</i>	natriuretic peptide type B	3.37 (5.11E-03)	1.65 (5.87E-01)	-2.04 (5.43E-02)
<i>Svep1</i>	Sushi, von Willebrand factor type A. EGF and pentraxin domain containing 1	3.33 (6.54E-04)	1.59 (3.86E-01)	-2.09 (9.13E-03)
<i>Tnfrsf12a</i>	tumor necrosis factor receptor superfamily. member 12a	3.01 (3.03E-03)	1.63 (4.75E-01)	-1.85 (4.48E-02)
<i>Ddit4</i>	DNA-damage-inducible transcript 4	2.72 (2.05E-02)	1.62 (6.62E-01)	-1.68 (2.12E-01)
<i>Myh7</i>	myosin. heavy polypeptide 7, cardiac muscle. beta	7.94 (4.39E-04)	5.8 (3.04E-02)	-1.37 (5.54E-01)
<i>Cnksr1</i>	connector enhancer of kinase suppressor of Ras 1	4.80 (1.08E-03)	3.30 (8.07E-02)	-1.45 (3.68E-01)
<i>Myot</i>	myotilin	4.27 (5.19E-04)	3.68 (3.04E-02)	-1.16 (7.64E-01)
<i>Acta1</i>	actin. alpha 1, skeletal muscle	4.26 (3.80E-05)	3.14 (3.75E-03)	-1.36 (6.72E-02)
<i>Cpxm2</i>	carboxypeptidase X 2 (M14 family)	4.12 (1.37E-04)	2.97 (1.83E-02)	-1.39 (1.27E-01)
<i>Gck</i>	glucokinase	3.73 (7.58E-04)	3.01 (4.89E-02)	-1.24 (5.53E-01)
<i>Wisp2</i>	WNT1 inducible signaling pathway protein 2	2.60 (4.10E-02)	1.89 (5.47E-01)	-1.38 (5.83E-01)
<i>Lcn2</i>	lipocalin 2	2.29 (4.32E-01)	2.37 (7.47E-01)	1.03 (9.87E-01)
<i>Mmp3</i>	matrix metalloproteinase 3	2.00 (3.07E-01)	1.92 (7.17E-01)	-1.04 (9.79E-01)
<i>C3</i>	complement component 3	1.78 (3.00E-01)	1.77 (6.87E-01)	-1.01 (9.93E-01)
<i>Ift122</i>	intraflagellar transport 122	1.71 (2.50E-01)	-1.97 (4.90E-01)	-3.38 (9.67E-03)
<i>Top2a</i>	topoisomerase (DNA) II alpha	1.82 (1.87E-01)	-1.62 (6.94E-01)	-2.95 (1.64E-02)
<i>Comp</i>	cartilage oligomeric matrix protein	6.88 (5.64E-03)	1.20 (9.12E-01)	-5.72 (8.24E-03)

Gene symbol	Gene name	Fold change (<i>P</i> -value)		
		AAV9-GFP TAC vs Con	AAV9-EDI TAC vs Con	AAV9-EDI TAC vs AAV9-GFP TAC
<i>Thbs1</i>	thrombospondin 1	5.64 (2.24E-04)	1.00 (9.99E-01)	-5.63 (6.09E-04)
<i>Timp1</i>	tissue inhibitor of metalloproteinase 1	5.92 (8.15E-04)	1.46 (7.56E-01)	-4.05 (3.21E-03)
<i>Gdf15</i>	growth differentiation factor 15	5.62 (4.79E-03)	1.27 (8.79E-01)	-4.43 (9.57E-03)
<i>Ctgf</i>	connective tissue growth factor	5.01 (1.37E-04)	1.39 (5.81E-01)	-3.61 (8.44E-04)
<i>Col5a2</i>	collagen. type V, alpha 2	4.73 (3.60E-04)	1.20 (8.49E-01)	-3.94 (9.82E-04)
<i>Fmod</i>	fibromodulin	4.49 (2.80E-03)	1.18 (8.88E-01)	-3.79 (5.12E-03)
<i>Mfap5</i>	microfibrillar associated protein 5	4.36 (2.62E-04)	1.50 (5.12E-01)	-2.92 (1.74E-03)
<i>Egr2</i>	early growth response 2	3.61 (1.58E-02)	-1.06 (9.70E-01)	-3.83 (1.11E-02)
<i>Apolo1</i>	apolipoprotein L domain containing 1	3.92 (3.80E-05)	1.17 (7.47E-01)	-3.36 (8.51E-05)
<i>Col3a1</i>	collagen. type III, alpha 1	3.21 (3.75E-03)	-1.21 (8.59E-01)	-3.9 (1.74E-03)
<i>Spp1</i>	secreted phosphoprotein 1	3.56 (1.56E-02)	1.07 (9.67E-01)	-3.33 (1.87E-02)
<i>Col8a2</i>	collagen. type VIII, alpha 2	3.54 (6.09E-03)	1.12 (9.18E-01)	-3.15 (9.29E-03)
<i>Col1a2</i>	collagen. type I, alpha 2	3.46 (4.84E-04)	1.05 (9.54E-01)	-3.30 (9.59E-04)
<i>1500015010Rik</i>	RIKEN cDNA 1500015010 gene	3.43 (3.72E-03)	1.12 (9.09E-01)	-3.07 (5.90E-03)
<i>Col1a1</i>	collagen. type I, alpha 1	3.05 (2.51E-03)	-1.11 (9.06E-01)	-3.38 (1.90E-03)
<i>Meox1</i>	mesenchyme homeobox 1	3.01 (2.61E-03)	-1.12 (8.92E-01)	-3.38 (1.77E-03)
<i>Pamr1</i>	peptidase domain containing associated with muscle regeneration 1	3.57 (1.19E-02)	1.49 (7.81E-01)	-2.39 (5.63E-02)
<i>Frzb</i>	frizzled-related protein	3.47 (7.88E-04)	1.31 (7.50E-01)	-2.65 (3.21E-03)

Gene symbol	Gene name	Fold change (<i>P</i> -value)		
		AAV9-GFP TAC vs Con	AAV9-EDI TAC vs Con	AAV9-EDI TAC vs AAV9-GFP TAC
<i>Fbn1</i>	fibrillin 1	3.19 (6.47E-04)	1.00 (9.98E-01)	-3.18 (9.82E-04)
<i>Fstl1</i>	follistatin-like 1	3.14 (3.22E-04)	1.13 (8.61E-01)	-2.79 (9.51E-04)
<i>Rcan1</i>	regulator of calcineurin 1	3.02 (4.64E-04)	1.05 (9.41E-01)	-2.88 (9.82E-04)
<i>Loxl2</i>	lysyl oxidase-like 2	2.72 (3.63E-03)	-1.11 (9.01E-01)	-3.02 (2.27E-03)
<i>Emp1</i>	epithelial membrane protein 1	2.99 (1.60E-04)	1.12 (8.42E-01)	-2.68 (6.72E-04)
<i>Bgn</i>	biglycan	3.10 (3.44E-04)	1.33 (5.90E-01)	-2.32 (2.00E-03)
<i>Rbp1</i>	retinol binding protein 1, cellular	3.00 (4.76E-03)	1.18 (8.68E-01)	-2.54 (1.01E-02)
<i>Sfrp2</i>	secreted frizzled-related protein 2	3.01 (1.82E-02)	1.24 (8.63E-01)	-2.41 (4.15E-02)
<i>Nmrk2</i>	nicotinamide riboside kinase 2	3.04 (1.21E-01)	1.33 (8.85E-01)	-2.28 (2.57E-01)
<i>Vcan</i>	versican	2.86 (5.19E-04)	1.08 (9.04E-01)	-2.66 (1.10E-03)
<i>Cthrc1</i>	collagen triple helix repeat containing 1	2.92 (9.51E-03)	1.17 (8.82E-01)	-2.48 (1.87E-02)
<i>Aldh 1a2</i>	aldehyde dehydrogenase family 1, subfamily A2	2.94 (7.48E-03)	1.22 (8.60E-01)	-2.40 (1.94E-02)
<i>Cebpd</i>	CCAAT/enhancer binding protein (C/EBP), delta	2.97 (1.68E-03)	1.48 (5.61E-01)	-2.00 (1.86E-02)
<i>Lox</i>	lysyl oxidase	2.88 (4.23E-03)	1.23 (8.32E-01)	-2.34 (1.18E-02)
<i>Serpine1</i>	serine (or cysteine) peptidase inhibitor, clade E, member 1	2.31 (2.67E-02)	-1.21 (8.62E-01)	-2.79 (9.48E-03)
<i>Col 15a1</i>	collagen. type XV, alpha 1	2.05 (2.00E-02)	-1.39 (6.88E-01)	-2.85 (2.78E-03)
<i>Serpinf1</i>	serine (or cysteine) peptidase inhibitor. clade F. member 1	2.82 (1.70E-03)	1.30 (7.36E-01)	-2.16 (8.31E-03)
<i>Gm 30873</i>	predicted gene, 30873	2.72 (8.10E-03)	1.37 (7.52E-01)	-1.98 (4.32E-02)

Gene symbol	Gene name	Fold change (<i>P</i> -value)		
		AAV9-GFP TAC vs Con	AAV9-EDI TAC vs Con	AAV9-EDI TAC vs AAV9-GFP TAC
<i>Tgfb2</i>	transforming growth factor. beta 2	2.58 (8.60E-03)	1.08 (9.38E-01)	-2.39 (1.19E-02)
<i>Ms 4a6d</i>	membrane-spanning 4-domains. subfamily A, member 6D	2.68 (8.81E-03)	1.30 (8.06E-01)	-2.07 (3.53E-02)
<i>Arntl</i>	aryl hydrocarbon receptor nuclear translocator-like	2.19 (5.24E-02)	-1.19 (8.82E-01)	-2.6 (1.79E-02)
<i>Gem</i>	GTP binding protein (gene overexpressed in skeletal muscle)	2.15 (7.46E-02)	1.00 (9.98E-01)	-2.14 (6.61E-02)
<i>Sprr1a</i>	small proline-rich protein 1A	1.92 (2.26E-01)	1.10 (9.50E-01)	-1.74 (2.97E-01)
<i>Nlrc5</i>	NLR family, CARD domain containing 5	1.69 (3.66E-01)	-1.13 (9.40E-01)	-1.91 (2.23E-01)
<i>Angpt1</i>	angiopoietin 1	-5.3 (1.37E-04)	-2.17 (8.41E-02)	2.44 (3.42E-03)
<i>Gm 19277</i>	predicted gene, 19277	-3.85 (1.41E-03)	-1.86 (3.58E-01)	2.07 (3.25E-02)
<i>Cngb3</i>	cyclic nucleotide gated channel beta 3	-3.21 (2.52E-03)	-1.97 (2.79E-01)	1.63 (1.23E-01)
<i>Snord 14e</i>	small nucleolar RNA, C/D box 14E	-2.92 (2.20E-02)	-1.83 (5.75E-01)	1.60 (3.16E-01)
<i>Gpr22</i>	G protein-coupled receptor 22	-4.21 (1.17E-03)	1.02 (9.88E-01)	4.29 (1.45E-03)
<i>Ces1d</i>	carboxylesterase 1D	-3.85 (4.80E-04)	-1.49 (5.38E-01)	2.58 (3.49E-03)
<i>Pfkfb1</i>	6-phosphofructo-2-kinase/fructose-2,6-biphosphatase 1	-3.48 (-.32E-04)	-1.23 (6.94E-01)	2.83 (6.17E-04)
<i>Il15</i>	interleukin 15	-3.12 (3.60E-04)	-1.35 (5.63E-01)	2.31 (2.35E-03)
<i>Inmt</i>	indolethylamine N-methyltransferase	-2.11 (3.58E-01)	1.49 (8.55E-01)	3.15 (1.14E-01)
<i>Mylk4</i>	myosin light chain kinase family. member 4	-2.78 (8.92E-04)	-1.06 (9.39E-01)	2.63 (1.74E-03)
<i>Acot1</i>	acyl-CoA thioesterase 1	-2.72 (9.53E-03)	-1.40 (7.47E-01)	1.95 (5.64E-02)
<i>Dbp</i>	D site albumin promoter binding protein	-2.48 (5.49E-02)	-1.02 (9.93E-01)	2.44 (5.12E-02)

Gene symbol	Gene name	Fold change (<i>P</i> -value)		
		AAV9-GFP TAC vs Con	AAV9-EDI TAC vs Con	AAV9-EDI TAC vs AAV9-GFP TAC
<i>Ppip 5k2</i>	diphosphoinositol pentakisphosphate kinase 2	-2.45 (8.55E-03)	-1.17 (8.62E-01)	2.10 (2.00E-02)
<i>Tmem 267</i>	transmembrane protein 267	-1.68 (4.56E-01)	-1.15 (9.37E-01)	1.46 (6.56E-01)
<i>Egr1</i>	early growth response 1	1.22 (8.28E-01)	-2.30 (5.78E-01)	-2.80 (9.52E-02)
<i>Fos</i>	FBJ osteosarcoma oncogene	-1.15 (8.40E-01)	-2.61 (2.95E-01)	-2.28 (7.39E-02)
<i>ligp1</i>	interferon inducible GTPase 1	-1.31 (7.52E-01)	-1.51 (8.27E-01)	-1.15 (9.15E-01)
<i>Irgm2</i>	immunity-related GTPase family M member 2	1.42 (6.79E-01)	-1.26 (8.99E-01)	-1.79 (4.38E-01)
<i>BC 023105</i>	cDNA sequence BC023105	1.21 (9.26E-01)	-1.38 (9.28E-01)	-1.68 (8.05E-01)
<i>Igtp</i>	interferon gamma induced GTPase	1.20 (8.54E-01)	-1.26 (8.97E-01)	-1.51 (6.39E-01)
<i>Saa3</i>	serum amyloid A 3	1.34 (6.21E-01)	1.13 (9.22E-01)	-1.18 (8.48E-01)
<i>Cyp 26b1</i>	cytochrome P450. family 26. subfamily b. polypeptide 1	-1.1 (9.29E-01)	1.13 (9.47E-01)	1.24 (8.53E-01)
<i>C1s1</i>	complement component 1. s subcomponent 1	1.10 (9.37E-01)	1.03 (9.93E-01)	-1.07 (9.71E-01)

Supplementary Table 6

Top 100 regulated genes in wild-type mice 4 weeks after TAC surgery and AAV9-EDI

treatment. The table refers to the heat map shown in Fig. 4G; gene array of untreated wild type (Wt; n=3) and AAV9-EDI (n=4) or AAV9-GFP (n=4) treated mice after transverse aortic constriction (TAC; 4 weeks). The table indicates the gene symbol, gene name and fold-changes and *p*-values; *n*-numbers represent samples of individual mice. For statistical analysis ANOVA with Tukey *post hoc* test was applied. Source data are provided under the following link:

<https://www.ebi.ac.uk/arrayexpress/experiments/E-MTAB-8110>.

Supplementary Table 7

Regulated genes associated with cardiac hypertrophy in wild-type mice 4 weeks after TAC surgery and AAV9-EDI treatment. Table refers to the heat map shown in Supplementary Figure 7B.

Gene symbol	Gene name	Fold change (<i>P</i> -value)		
		AAV9-GFP TAC vs Con	AAV9-EDI TAC vs Con	AAV9-EDI TAC vs AAV9-GFP TAC
<i>Postn</i>	periostin, osteoblast specific factor	8.34 (1.70E-05)	1.89 (1.20E-01)	-4.40 (4.37E-04)
<i>Myh7</i>	myosin, heavy polypeptide 7, cardiac muscle, beta	6.20 (7.95E-04)	4.53 (3.11E-02)	-1.37 (6.30E-01)
<i>Ctgf</i>	connective tissue growth factor	4.84 (3.12E-05)	1.34 (4.91E-01)	-3.61 (4.21E-04)
<i>S100a8</i>	S100 calcium binding protein A8 (calgranulin A)	4.61 (8.49E-02)	2.95 (5.68E-01)	-1.56 (7.68E-01)
<i>Cdkn1a</i>	cyclin-dependent kinase inhibitor 1A (P21)	4.61 (1.54E-03)	1.56 (6.35E-01)	-2.95 (1.76E-02)
<i>Mybpc2</i>	myosin binding protein C, fast-type	4.52 (7.20E-05)	2.97 (1.13E-02)	-1.52 (1.08E-01)
<i>Nppb</i>	natriuretic peptide type B	3.80 (1.55E-03)	1.86 (3.25E-01)	-2.04 (5.75E-02)
<i>Nppa</i>	natriuretic peptide type A	3.74 (3.31E-05)	2.12 (1.26E-02)	-1.77 (6.44E-03)
<i>Prg4</i>	proteoglycan 4 (megakaryocyte stimulating factor, articular superficial zone protein)	3.41 (2.08E-02)	2.10 (4.49E-01)	-1.63 (4.52E-01)
<i>Rcan1</i>	regulator of calcineurin 1	3.15 (1.02E-04)	1.09 (8.56E-01)	-2.88 (5.38E-04)
<i>Pi16</i>	peptidase inhibitor 16	2.75 (1.13E-03)	1.39 (5.37E-01)	-1.99 (1.69E-02)
<i>Fstl3</i>	follistatin-like 3	2.68 (1.71E-04)	1.74 (6.37E-02)	-1.54 (3.55E-02)
<i fn1<="" i=""></i>	fibronectin 1	2.65 (2.29E-04)	1.06 (9.05E-01)	-2.49 (9.27E-04)
<i>Nox4</i>	NADPH oxidase 4	2.62 (4.87E-03)	1.49 (5.18E-01)	-1.76 (9.01E-02)
<i>Tgfb2</i>	transforming growth factor, beta 2	2.48 (4.20E-03)	1.04 (9.60E-01)	-2.39 (8.45E-03)
<i>Fhl1</i>	four and a half LIM domains 1	2.48 (1.32E-03)	1.48 (3.56E-01)	-1.68 (4.29E-02)
<i>Errfi1</i>	ERBB receptor feedback inhibitor 1	2.41 (7.04E-03)	1.19 (8.21E-01)	-2.03 (3.12E-02)

Gene symbol	Gene name	Fold change (<i>P</i> -value)		
		AAV9-GFP TAC vs Con	AAV9-EDI TAC vs Con	AAV9-EDI TAC vs AAV9-GFP TAC
<i>Fgl2</i>	fibrinogen-like protein 2	2.20 (8.58E-03)	1.05 (9.47E-01)	-2.10 (1.81E-02)
<i>Eln</i>	elastin	2.20 (1.67E-04)	-1.00 (9.90E-01)	-2.21 (5.38E-04)
<i>Il6</i>	interleukin 6	2.16 (1.56E-03)	1.04 (9.47E-01)	-2.09 (3.70E-03)
<i>Cdk1</i>	cyclin-dependent kinase 1	2.13 (1.33E-02)	1.08 (9.16E-01)	-1.97 (3.09E-02)
<i>Mmp2</i>	matrix metalloproteinase 2	2.06 (1.45E-03)	1.06 (9.08E-01)	-1.94 (4.52E-03)
<i>Apod</i>	apolipoprotein D	2.04 (4.78E-04)	1.54 (8.51E-02)	-1.32 (1.04E-01)
<i>Hbegf</i>	heparin-binding EGF-like growth factor	2.02 (9.19E-04)	1.19 (6.72E-01)	-1.69 (9.32E-03)
<i>Ace</i>	angiotensin I converting enzyme (peptidyl-dipeptidase A) 1	1.94 (4.71E-04)	1.2 (5.47E-01)	-1.62 (6.00E-03)
<i>Edn3</i>	endothelin 3	1.93 (1.15E-04)	1.63 (1.52E-02)	-1.19 (1.92E-01)
<i>Tlr4</i>	toll-like receptor 4	1.90 (1.81E-02)	1.23 (7.38E-01)	-1.54 (1.28E-01)
<i>Bcl2</i>	B cell leukemia/lymphoma 2	1.90 (1.10E-03)	1.06 (8.96E-01)	-1.80 (3.71E-03)
<i>F2r</i>	coagulation factor II (thrombin) receptor	1.88 (1.89E-03)	-1.08 (8.49E-01)	-2.03 (1.82E-03)
<i>Igf1</i>	insulin-like growth factor 1	1.87 (5.32E-04)	1.02 (9.64E-01)	-1.84 (1.51E-03)
<i>Abat</i>	4-aminobutyrate aminotransferase	1.86 (3.07E-03)	1.89 (2.81E-02)	1.02 (9.75E-01)
<i>Col14a1</i>	collagen, type XIV, alpha 1	1.82 (2.33E-03)	1.00 (9.93E-01)	-1.81 (4.21E-03)
<i>Nr4a1</i>	nuclear receptor subfamily 4, group A, member 1	1.82 (2.04E-01)	-1.07 (9.51E-01)	-1.95 (1.61E-01)
<i>Plat</i>	plasminogen activator, tissue	1.79 (1.06E-03)	1.06 (8.78E-01)	-1.69 (4.35E-03)
<i>Lbp</i>	lipopolysaccharide binding protein	1.76 (1.73E-03)	1.34 (2.70E-01)	-1.32 (9.55E-02)
<i>Shisa3</i>	shisa family member 3	1.74 (4.06E-02)	1.08 (9.12E-01)	-1.61 (9.50E-02)

Gene symbol	Gene name	Fold change (<i>P</i> -value)		
		AAV9-GFP TAC vs Con	AAV9-EDI TAC vs Con	AAV9-EDI TAC vs AAV9-GFP TAC
<i>Tnfrsf6</i>	tumor necrosis factor alpha induced protein 6	1.66 (1.79E-03)	1.04 (9.14E-01)	-1.59 (5.31E-03)
<i>Foxc2</i>	forkhead box C2	1.58 (1.19E-02)	-1.01 (9.90E-01)	-1.59 (1.71E-02)
<i>Gpx3</i>	glutathione peroxidase 3	1.58 (6.52E-02)	1.14 (8.27E-01)	-1.39 (2.43E-01)
<i>Kif5b</i>	kinesin family member 5B	1.56 (1.40E-03)	1.12 (6.74E-01)	-1.39 (1.38E-02)
<i>Smad1</i>	SMAD family member 1	1.53 (3.08E-03)	-1.11 (7.34E-01)	-1.70 (1.66E-03)
<i>Clec4d</i>	C-type lectin domain family 4, member d	1.51 (5.76E-02)	1.16 (7.79E-01)	-1.31 (2.83E-01)
<i>Kcnk2</i>	potassium channel, subfamily K, member 2	1.43 (3.40E-02)	-1.13 (7.70E-01)	-1.61 (1.22E-02)
<i>Trf</i>	transferrin	1.43 (5.58E-02)	-1.13 (7.90E-01)	-1.61 (2.01E-02)
<i>Fetub</i>	fetuin beta	1.39 (1.13E-01)	1.32 (4.92E-01)	-1.05 (9.12E-01)
<i>Wisp1</i>	WNT1 inducible signaling pathway protein 1	1.36 (2.80E-01)	-1.18 (7.93E-01)	-1.61 (9.79E-02)
<i>Grk5</i>	G protein-coupled receptor kinase 5	1.34 (4.23E-01)	-1.16 (8.55E-01)	-1.55 (2.30E-01)
<i>Tmem100</i>	transmembrane protein 100	-1.40 (1.83E-01)	-1.17 (7.87E-01)	1.19 (6.24E-01)
<i>Parp1</i>	poly (ADP-ribose) polymerase family, member 1	-1.55 (1.00E-03)	-1.19 (3.94E-01)	1.30 (2.89E-02)
<i>Adrb1</i>	adrenergic receptor, beta 1	-1.59 (2.27E-02)	-1.40 (3.19E-01)	1.13 (6.87E-01)
<i>Tbx5</i>	T-box 5	-1.63 (1.33E-03)	-1.04 (9.03E-01)	1.57 (3.96E-03)
<i>Myocd</i>	myocardin	-1.64 (7.28E-05)	-1.25 (8.60E-02)	1.31 (5.79E-03)
<i>Corin</i>	corin	-1.64 (1.67E-03)	-1.03 (9.32E-01)	1.59 (4.52E-03)
<i>Tcap</i>	titin-cap	-1.66 (4.92E-03)	-1.04 (9.17E-01)	1.59 (1.28E-02)
<i>Amy1</i>	amylase 1, salivary	-2.05 (2.67E-03)	1.11 (8.40E-01)	2.28 (2.12E-03)

Gene symbol	Gene name	Fold change (<i>P</i> -value)		
		AAV9-GFP TAC vs Con	AAV9-EDI TAC vs Con	AAV9-EDI TAC vs AAV9-GFP TAC
<i>Rgs2</i>	regulator of G-protein signaling 2	-2.21 (1.61E-03)	-1.77 (8.71E-02)	1.25 (3.97E-01)
<i>Kcnd2</i>	potassium voltage-gated channel, Shal-related family, member 2	-2.23 (4.72E-04)	-1.49 (1.74E-01)	1.50 (3.97E-02)
<i>Penk</i>	preproenkephalin	-2.30 (6.02E-04)	-1.47 (2.26E-01)	1.57 (3.52E-02)
<i>Mylk4</i>	myosin light chain kinase family, member 4	-2.73 (2.69E-04)	-1.04 (9.47E-01)	2.63 (9.70E-04)
<i>Phkg1</i>	phosphorylase kinase gamma 1	-2.84 (4.16E-05)	-1.75 (2.89E-02)	1.62 (7.96E-03)
<i>Ces1d</i>	carboxylesterase 1D	-3.79 (1.25E-04)	-1.47 (3.88E-01)	2.58 (2.17E-03)

Supplementary Table 7

Regulated genes associated with cardiac hypertrophy in wild-type mice 4 weeks after TAC surgery and AAV9-EDI treatment. The table refers to the heat map shown in Supplementary Figure 7B; genes reported by Gene Ontology as associated with cardiac hypertrophy and regulated in untreated wild-type (Wt, n=3) versus AAV9-EDI (n=4) or AAV9-GFP (n=4) treated mice after transverse aortic constriction (TAC; 4 weeks). The table indicates the gene symbol, gene name and fold-changes and *p*-values; *n*-numbers represent samples of individual mice. For statistical analysis ANOVA with Tukey *post hoc* test was applied. Source data are provided under the following link: <https://www.ebi.ac.uk/arrayexpress/experiments/E-MTAB-8110>.

Supplementary Table 8

Regulated genes associated with extracellular matrix in wild-type mice 4 weeks after TAC surgery and AAV9-EDI treatment. The table refers to the heat map shown in Supplementary Figure 7B.

Gene symbol	Gene name	Fold change (<i>P</i> -value)		
		AAV9-GFP TAC vs Con	AAV9-EDI TAC vs Con	AAV9-EDI TAC vs AAV9-GFP TAC
<i>Postn</i>	periostin, osteoblast specific factor	8.34 (1.70E-05)	1.89 (1.20E-01)	-4.40 (4.37E-04)
<i>Mfap4</i>	microfibrillar-associated protein 4	4.76 (1.16E-04)	1.54 (4.00E-01)	-3.09 (1.61E-03)
<i>Mfap5</i>	microfibrillar associated protein 5	4.59 (5.25E-05)	1.57 (2.63E-01)	-2.92 (1.06E-03)
<i>Fstl3</i>	follistatin-like 3	2.68 (1.71E-04)	1.74 (6.37E-02)	-1.54 (3.55E-02)
<i>Fn1</i>	fibronectin 1	2.65 (2.29E-04)	1.06 (9.05E-01)	-2.49 (9.27E-04)
<i>Nox4</i>	NADPH oxidase 4	2.62 (4.87E-03)	1.49 (5.18E-01)	-1.76 (9.01E-02)
<i>Serpine 1</i>	serine (or cysteine) peptidase inhibitor, clade E, member 1	2.28 (1.23E-02)	-1.22 (8.02E-01)	-2.79 (6.36E-03)
<i>Eln</i>	elastin	2.20 (1.67E-04)	-1.00 (9.90E-01)	-2.21 (5.38E-04)
<i>Fbln2</i>	fibulin 2	2.04 (1.67E-03)	1.43 (2.92E-01)	-1.43 (8.42E-02)
<i>Hbegf</i>	heparin-binding EGF-like growth factor	2.02 (9.19E-04)	1.19 (6.72E-01)	-1.69 (9.32E-03)
<i>Ace</i>	angiotensin I converting enzyme (peptidyl-dipeptidase A) 1	1.94 (4.71E-04)	1.20 (5.47E-01)	-1.62 (6.00E-03)
<i>Plat</i>	plasminogen activator, tissue	1.79 (1.06E-03)	1.06 (8.78E-01)	-1.69 (4.35E-03)
<i>Xirp1</i>	xin actin-binding repeat containing 1	1.59 (1.21E-02)	1.01 (9.86E-01)	-1.58 (1.93E-02)
<i>Mfap2</i>	microfibrillar-associated protein 2	1.54 (2.11E-03)	1.21 (4.11E-01)	-1.28 (6.23E-02)
<i>Entpd1</i>	ectonucleoside triphosphate diphosphohydrolase 1	1.51 (1.14E-02)	1.33 (2.66E-01)	-1.13 (5.68E-01)
<i>Egr1</i>	early growth response 1	1.45 (5.96E-01)	-1.93 (6.04E-01)	-2.80 (9.68E-02)

Gene symbol	Gene name	Fold change (<i>P</i> -value)		
		AAV9-GFP TAC vs Con	AAV9-EDI TAC vs Con	AAV9-EDI TAC vs AAV9-GFP TAC
<i>Fbn2</i>	fibrillin 2	1.34 (1.12E-01)	-1.24 (6.01E-01)	-1.66 (1.39E-02)

Supplementary Table 8

Regulated genes associated with extracellular matrix in wild-type mice 4 weeks after TAC surgery and AAV9-EDI treatment.

The table refers to the heat map shown in Supplementary Figure 7B; genes reported by Gene Ontology as associated with extracellular matrix and regulated in untreated wild-type (Wt, n=3) versus AAV9-EDI (n=4) or AAV9-GFP (n=4) treated mice after transverse aortic constriction (TAC; 4 weeks). The table indicates the gene symbol, gene name and fold-changes and *p*-values; *n*-numbers represent samples of individual mice. For statistical analysis ANOVA with Tukey *post hoc* test was applied. Source data are provided under the following link: <https://www.ebi.ac.uk/arrayexpress/experiments/E-MTAB-8110>.

Supplementary Table 9

Regulated genes associated with cell death in wild-type mice 4 weeks after TAC surgery and AAV9-EDI treatment. Table refers to the heat map shown in Supplementary Figure 7B.

Gene symbol	Gene name	Fold change (<i>P</i> -value)		
		AAV9-GFP TAC vs Con	AAV9-EDI TAC vs Con	AAV9-EDI TAC vs AAV9-GFP TAC
<i>Postn</i>	periostin, osteoblast specific factor	8.34 (1.70E-05)	1.89 (1.20E-01)	-4.40 (4.37E-04)
<i>Myh7</i>	myosin, heavy polypeptide 7, cardiac muscle, beta	6.20 (7.95E-04)	4.53 (3.11E-02)	-1.37 (6.30E-01)
<i>Thbs1</i>	thrombospondin 1	5.55 (4.48E-05)	-1.01 (9.85E-01)	-5.63 (3.23E-04)
<i>Ctgf</i>	connective tissue growth factor	4.84 (3.12E-05)	1.34 (4.91E-01)	-3.61 (4.21E-04)
<i>Rcan1</i>	regulator of calcineurin 1	3.15 (1.02E-04)	1.09 (8.56E-01)	-2.88 (5.38E-04)
<i>Bgn</i>	biglycan	3.10 (7.20E-05)	1.33 (4.12E-01)	-2.32 (1.11E-03)
<i>Spp1</i>	secreted phosphoprotein 1	3.08 (1.65E-02)	-1.08 (9.48E-01)	-3.33 (1.78E-02)
<i>Col1a1</i>	collagen, type I, alpha 1	3.04 (7.57E-04)	-1.11 (8.78E-01)	-3.38 (1.06E-03)
<i>Sfrp2</i>	secreted frizzled-related protein 2	2.97 (7.65E-03)	1.23 (8.25E-01)	-2.41 (3.11E-02)
<i>Vcan</i>	versican	2.92 (1.11E-04)	1.10 (8.39E-01)	-2.66 (5.81E-04)
<i>Fn1</i>	fibronectin 1	2.65 (2.29E-04)	1.06 (9.05E-01)	-2.49 (9.27E-04)
<i>Nox4</i>	NADPH oxidase 4	2.62 (4.87E-03)	1.49 (5.18E-01)	-1.76 (9.01E-02)
<i>Cyr61</i>	cysteine rich protein 61	2.60 (1.05E-03)	-1.02 (9.81E-01)	-2.64 (1.88E-03)
<i>Tgfb2</i>	transforming growth factor, beta 2	2.48 (4.20E-03)	1.04 (9.60E-01)	-2.39 (8.45E-03)
<i>Stc1</i>	stanniocalcin 1	2.39 (8.01E-05)	1.51 (8.09E-02)	-1.58 (7.89E-03)
<i>Igfbp7</i>	insulin-like growth factor binding protein 7	2.35 (1.28E-05)	1.14 (5.47E-01)	-2.07 (2.39E-04)

Gene symbol	Gene name	Fold change (<i>P</i> -value)		
		AAV9-GFP TAC vs Con	AAV9-EDI TAC vs Con	AAV9-EDI TAC vs AAV9-GFP TAC
<i>Col4a1</i>	collagen, type IV, alpha 1	2.29 (3.77E-04)	-1.08 (8.62E-01)	-2.48 (5.97E-04)
<i>Nupr1</i>	nuclear protein transcription regulator 1	2.28 (5.80E-03)	1.10 (8.96E-01)	-2.08 (1.74E-02)
<i>Serpine1</i>	serine (or cysteine) peptidase inhibitor, clade E, member 1	2.28 (1.23E-02)	-1.22 (8.02E-01)	-2.79 (6.36E-03)
<i>Sulf1</i>	sulfatase 1	2.23 (2.59E-04)	1.11 (8.07E-01)	-2.01 (1.53E-03)
<i>Anxa1</i>	annexin A1	2.18 (1.49E-04)	1.21 (5.06E-01)	-1.80 (1.80E-03)
<i>Pla2g4a</i>	phospholipase A2, group IVA (cytosolic, calcium-dependent)	2.13 (1.63E-03)	1.01 (9.86E-01)	-2.11 (3.32E-03)
<i>Sparc</i>	secreted acidic cysteine rich glycoprotein	2.10 (1.41E-04)	-1.04 (9.13E-01)	-2.19 (4.38E-04)
<i>Casp12</i>	caspase 12	2.06 (6.50E-03)	1.29 (6.47E-01)	-1.59 (7.32E-02)
<i>Lrp1</i>	low density lipoprotein receptor-related protein 1	2.04 (6.97E-03)	1.12 (8.47E-01)	-1.82 (2.47E-02)
<i>Col4a2</i>	collagen, type IV, alpha 2	2.04 (5.00E-04)	-1.02 (9.66E-01)	-2.07 (1.06E-03)
<i>Pmepa1</i>	prostate transmembrane protein, androgen induced 1	2.03 (6.02E-04)	1.05 (9.11E-01)	-1.94 (1.86E-03)
<i>Igfbp6</i>	insulin-like growth factor binding protein 6	2.03 (1.79E-02)	1.40 (5.96E-01)	-1.45 (2.55E-01)
<i>Hbegf</i>	heparin-binding EGF-like growth factor	2.02 (9.19E-04)	1.19 (6.72E-01)	-1.69 (9.32E-03)
<i>Nuak1</i>	NUAK family, SNF1-like kinase, 1	2.00 (1.84E-04)	1.11 (7.58E-01)	-1.81 (1.14E-03)
<i>Tgfb3</i>	transforming growth factor, beta 3	1.96 (2.90E-04)	1.15 (6.73E-01)	-1.70 (2.54E-03)
<i>Tlr4</i>	toll-like receptor 4	1.90 (1.81E-02)	1.23 (7.38E-01)	-1.54 (1.28E-01)
<i>Bcl2</i>	B cell leukemia/lymphoma 2	1.90 (1.10E-03)	1.06 (8.96E-01)	-1.80 (3.71E-03)
<i>Haus8</i>	4HAUS augmin-like complex, subunit 8	1.90 (4.97E-03)	1.15 (7.78E-01)	-1.65 (2.54E-02)

Gene symbol	Gene name	Fold change (<i>P</i> -value)		
		AAV9-GFP TAC vs Con	AAV9-EDI TAC vs Con	AAV9-EDI TAC vs AAV9-GFP TAC
<i>Plat</i>	plasminogen activator, tissue	1.79 (1.06E-03)	1.06 (8.78E-01)	-1.69 (4.35E-03)
<i>Rtn4</i>	reticulon 4	1.76 (1.28E-03)	1.05 (8.90E-01)	-1.67 (4.19E-03)
<i>Vim</i>	vimentin	1.75 (7.20E-05)	1.01 (9.75E-01)	-1.74 (4.27E-04)
<i>Lum</i>	lumican	1.73 (8.97E-05)	1.11 (6.05E-01)	-1.55 (9.27E-04)
<i>Cyp1b1</i>	cytochrome P450, family 1, subfamily b, polypeptide 1	1.70 (2.59E-03)	1.09 (8.13E-01)	-1.55 (1.29E-02)
<i>Mmp14</i>	matrix metalloproteinase 14 (membrane-inserted)	1.67 (1.17E-03)	-1.05 (8.95E-01)	-1.75 (1.52E-03)
<i>Plcg2</i>	phospholipase C, gamma 2	1.64 (1.98E-03)	1.11 (7.55E-01)	-1.48 (1.31E-02)
<i>Tnip1</i>	TNFAIP3 interacting protein 1	1.60 (3.18E-03)	1.13 (7.18E-01)	-1.42 (2.53E-02)
<i>Axl</i>	AXL receptor tyrosine kinase	1.58 (8.73E-04)	1.14 (6.08E-01)	-1.39 (1.09E-02)
<i>Mvp</i>	major vault protein	1.56 (1.06E-02)	1.09 (8.32E-01)	-1.43 (4.29E-02)
<i>Sema3f</i>	sema domain, immunoglobulin domain (Ig), short basic domain, secreted, (semaphorin) 3F	1.55 (6.14E-03)	-1.00 (9.98E-01)	-1.55 (8.93E-03)
<i>Ccdc80</i>	coiled-coil domain containing 80	1.53 (9.39E-03)	1.08 (8.39E-01)	-1.42 (3.52E-02)
<i>Serp1g1</i>	serine (or cysteine) peptidase inhibitor, clade G, member 1	1.49 (8.98E-02)	1.17 (7.72E-01)	-1.27 (4.17E-01)
<i>Ccnd1</i>	cyclin D1	1.45 (2.35E-01)	-1.27 (7.30E-01)	-1.84 (5.44E-02)
<i>Cfh</i>	complement component factor h	1.43 (3.38E-01)	1.20 (8.28E-01)	-1.19 (7.66E-01)
<i>Creb3l2</i>	cAMP responsive element binding protein 3-like 2	1.42 (1.61E-02)	-1.06 (8.70E-01)	-1.50 (1.06E-02)
<i>Rhoc</i>	ras homolog family member C	1.41 (1.11E-01)	-1.27 (6.20E-01)	-1.78 (1.56E-02)
<i>Grk5</i>	G protein-coupled receptor kinase 5	1.34 (4.23E-01)	-1.16 (8.55E-01)	-1.55 (2.30E-01)

Gene symbol	Gene name	Fold change (<i>P</i> -value)		
		AAV9-GFP TAC vs Con	AAV9-EDI TAC vs Con	AAV9-EDI TAC vs AAV9-GFP TAC
<i>Ptgds</i>	prostaglandin D2 synthase (brain)	1.32 (2.52E-01)	1.75 (1.22E-01)	1.33 (2.93E-01)
<i>Herpud1</i>	homocysteine-inducible, endoplasmic reticulum stress-inducible, ubiquitin-like domain member 1	-1.51 (5.38E-02)	1.30 (5.35E-01)	1.96 (6.53E-03)
<i>Camk2a</i>	calcium/calmodulin-dependent protein kinase II alpha	-1.51 (3.07E-04)	-1.25 (1.05E-01)	1.21 (4.65E-02)
<i>As3mt</i>	arsenic (+3 oxidation state) methyltransferase	-1.58 (5.68E-05)	-1.06 (7.57E-01)	1.50 (5.05E-04)
<i>Sirt1</i>	sirtuin 1	-1.61 (6.79E-03)	-1.12 (7.78E-01)	1.45 (3.69E-02)
<i>Myocd</i>	myocardin	-1.64 (7.28E-05)	-1.25 (8.60E-02)	1.31 (5.79E-03)
<i>Zbtb16</i>	zinc finger and BTB domain containing 16	-1.80 (9.37E-03)	1.02 (9.78E-01)	1.82 (1.24E-02)

Supplementary Table 9

Regulated genes associated with cell death in wild-type mice 4 weeks after TAC surgery and AAV9-EDI treatment. The table refers to the heat map shown in Supplementary Figure 7B; genes reported by Gene Ontology as associated with cell death and regulated in untreated wild-type (Wt, n=3) versus AAV9-EDI (n=4) or AAV9-GFP (n=4) treated mice after transverse aortic constriction (TAC; 4 weeks). The table indicates the gene symbol, gene name and fold-changes and *p*-values; *n*-numbers represent samples of individual mice. For statistical analysis ANOVA with Tukey *post hoc* test was applied. Source data are provided under the following link: <https://www.ebi.ac.uk/arrayexpress/experiments/E-MTAB-8110>.

Supplementary Table 10

Regulated genes associated with heart failure in wild-type mice 4 weeks after TAC surgery and AAV9-EDI treatment. The table refers to the heat map shown in Supplementary Figure 7B.

Gene symbol	Gene name	Fold change (<i>P</i> -value)		
		AAV9-GFP TAC vs Con	AAV9-EDI TAC vs Con	AAV9-EDI TAC vs AAV9-GFP TAC
<i>Postn</i>	periostin, osteoblast specific factor	8.34 (1.70E-05)	1.89 (1.20E-01)	-4.40 (4.37E-04)
<i>Comp</i>	cartilage oligomeric matrix protein	6.67 (2.07E-03)	1.17 (9.11E-01)	-5.72 (5.41E-03)
<i>Thbs4</i>	thrombospondin 4	4.96 (1.93E-03)	1.71 (5.68E-01)	-2.89 (2.81E-02)
<i>Ctgf</i>	connective tissue growth factor	4.84 (3.12E-05)	1.34 (4.91E-01)	-3.61 (4.21E-04)
<i>Frzb</i>	frizzled-related protein	3.46 (1.89E-04)	1.30 (6.34E-01)	-2.65 (1.84E-03)
<i>Tnfrsf12a</i>	tumor necrosis factor receptor superfamily, member 12a	3.39 (9.75E-04)	1.84 (2.29E-01)	-1.85 (5.38E-02)
<i>Fstl1</i>	follistatin-like 1	3.21 (6.32E-05)	1.15 (7.63E-01)	-2.79 (5.18E-04)
<i>Lox</i>	lysyl oxidase	2.86 (1.35E-03)	1.22 (7.61E-01)	-2.34 (7.81E-03)
<i>Fn1</i>	fibronectin 1	2.65 (2.29E-04)	1.06 (9.05E-01)	-2.49 (9.27E-04)
<i>Nox4</i>	NADPH oxidase 4	2.62 (4.87E-03)	1.49 (5.18E-01)	-1.76 (9.01E-02)
<i>Cyr61</i>	cysteine rich protein 61	2.60 (1.05E-03)	-1.02 (9.81E-01)	-2.64 (1.88E-03)
<i>Wisp2</i>	WNT1 inducible signaling pathway protein 2	2.59 (1.93E-02)	1.88 (3.63E-01)	-1.38 (5.48E-01)
<i>Tgfb2</i>	transforming growth factor, beta 2	2.48 (4.20E-03)	1.04 (9.60E-01)	-2.39 (8.45E-03)
<i>Fhl1</i>	four and a half LIM domains 1	2.48 (1.32E-03)	1.48 (3.56E-01)	-1.68 (4.29E-02)
<i>Col4a1</i>	collagen, type IV, alpha 1	2.29 (3.77E-04)	-1.08 (8.62E-01)	-2.48 (5.97E-04)
<i>Serpine1</i>	serine (or cysteine) peptidase inhibitor, clade E, member 1	2.28 (1.23E-02)	-1.22 (8.02E-01)	-2.79 (6.36E-03)

Gene symbol	Gene name	Fold change (<i>P</i> -value)		
		AAV9-GFP TAC vs Con	AAV9-EDI TAC vs Con	AAV9-EDI TAC vs AAV9-GFP TAC
<i>Mmp2</i>	matrix metalloproteinase 2	2.06 (1.45E-03)	1.06 (9.08E-01)	-1.94 (4.52E-03)
<i>Fbln2</i>	fibulin 2	2.04 (1.67E-03)	1.43 (2.92E-01)	-1.43 (8.42E-02)
<i>Tgfb3</i>	transforming growth factor, beta 3	1.96 (2.90E-04)	1.15 (6.73E-01)	-1.70 (2.54E-03)
<i>Bcl2</i>	B cell leukemia/lymphoma 2	1.90 (1.10E-03)	1.06 (8.96E-01)	-1.80 (3.71E-03)
<i>Sfrp1</i>	secreted frizzled-related protein 1	1.73 (1.80E-02)	1.03 (9.54E-01)	-1.67 (3.41E-02)
<i>Lum</i>	lumican	1.73 (8.97E-05)	1.11 (6.05E-01)	-1.55 (9.27E-04)
<i>P4ha1</i>	procollagen-proline, 2-oxoglutarate 4-dioxygenase (proline 4-hydroxylase), alpha 1 polypeptide	1.69 (7.04E-03)	1.04 (9.43E-01)	-1.63 (1.56E-02)
<i>Tgm2</i>	transglutaminase 2, C polypeptide	1.64 (4.94E-04)	1.29 (1.59E-01)	-1.27 (4.71E-02)
<i>Stat3</i>	signal transducer and activator of transcription 3	1.48 (2.70E-05)	1.05 (7.14E-01)	-1.41 (3.23E-04)
<i>Dusp6</i>	dual specificity phosphatase 6	1.43 (1.15E-01)	-1.11 (8.49E-01)	-1.59 (5.68E-02)
<i>Grk5</i>	G protein-coupled receptor kinase 5	1.34 (4.23E-01)	-1.16 (8.55E-01)	-1.55 (2.30E-01)
<i>Maob</i>	monoamine oxidase B	-2.14 (6.74E-05)	-1.20 (4.40E-01)	1.79 (9.70E-04)
<i>Rgs2</i>	regulator of G-protein signaling 2	-2.21 (1.61E-03)	-1.77 (8.71E-02)	1.25 (3.97E-01)
<i>Rbfox1</i>	RNA binding protein, fox-1 homolog (<i>C. elegans</i>) 1	-2.54 (3.50E-05)	-1.51 (4.31E-02)	1.68 (1.89E-03)

Supplementary Table 10

Regulated genes associated with heart failure in wild-type mice 4 weeks after TAC surgery and AAV9-EDI treatment. The table refers to the heat map shown in Supplementary Figure 7B; genes reported by Gene Ontology as associated with heart failure and regulated in untreated wild-type (Wt, n=3) versus AAV9-EDI (n=4) or AAV9-GFP (n=4) treated mice after transverse aortic constriction (TAC; 4 weeks). The table indicates the gene symbol, gene name and fold-changes and *p*-values; *n*-numbers represent samples of individual mice. For statistical analysis ANOVA with Tukey *post hoc* test was applied. Source data are provided under the following link: <https://www.ebi.ac.uk/arrayexpress/experiments/E-MTAB-8110>.

Supplementary table 11

Regulated genes associated with Myc-related gene networks in wild-type mice 4 weeks after TAC surgery and AAV9-EDI treatment. Table refers to the heat map shown in Supplementary Figure 7D.

Gene symbol	Gene name	Fold change (<i>P</i> -value)		
		AAV9-GFP TAC vs Con	AAV9-EDI TAC vs Con	AAV9-EDI TAC vs AAV9-GFP TAC
<i>Myh7</i>	myosin. heavy polypeptide 7. cardiac muscle. beta	6.20 (7.95E-04)	4.53 (3.11E-02)	-1.37 (6.30E-01)
<i>Timp1</i>	tissue inhibitor of metalloproteinase 1	5.74 (2.19E-04)	1.42 (6.74E-01)	-4.05 (1.88E-03)
<i>Thbs1</i>	thrombospondin 1	5.55 (4.48E-05)	-1.01 (9.85E-01)	-5.63 (3.23E-04)
<i>Acta1</i>	actin. alpha 1. skeletal muscle	4.49 (5.78E-06)	3.31 (5.69E-04)	-1.36 (6.62E-02)
<i>Fmod</i>	fibromodulin	4.42 (8.99E-04)	1.17 (8.69E-01)	-3.79 (3.21E-03)
<i>Gck</i>	glucokinase	3.97 (1.84E-04)	3.19 (1.24E-02)	-1.24 (5.37E-01)
<i>Itgbl1</i>	integrin. beta-like 1	3.94 (3.00E-04)	1.65 (3.19E-01)	-2.38 (8.51E-03)
<i>Nppb</i>	natriuretic peptide type B	3.80 (1.55E-03)	1.86 (3.25E-01)	-2.04 (5.75E-02)
<i>Nppa</i>	natriuretic peptide type A	3.74 (3.31E-05)	2.12 (1.26E-02)	-1.77 (6.44E-03)
<i>Egr2</i>	early growth response 2	3.72 (5.47E-03)	-1.03 (9.78E-01)	-3.83 (7.52E-03)
<i>Frzb</i>	frizzled-related protein	3.46 (1.89E-04)	1.30 (6.34E-01)	-2.65 (1.84E-03)
<i>Mt2</i>	metallothionein 2	3.37 (2.39E-03)	1.41 (6.74E-01)	-2.38 (2.38E-02)
<i>Fbn1</i>	fibrillin 1	3.33 (1.49E-04)	1.05 (9.39E-01)	-3.18 (5.38E-04)
<i>Fstl1</i>	follistatin-like 1	3.21 (6.32E-05)	1.15 (7.63E-01)	-2.79 (5.18E-04)
<i>Emp1</i>	epithelial membrane protein 1	3.10 (3.50E-05)	1.16 (6.94E-01)	-2.68 (4.03E-04)
<i>Spp1</i>	secreted phosphoprotein 1	3.08 (1.65E-02)	-1.08 (9.48E-01)	-3.33 (1.78E-02)
<i>Cebpd</i>	CCAAT/enhancer binding protein (C/EBP). delta	2.96 (4.84E-04)	1.48 (3.84E-01)	-2.00 (1.31E-02)

Gene symbol	Gene name	Fold change (<i>P</i> -value)		
		AAV9-GFP TAC vs Con	AAV9-EDI TAC vs Con	AAV9-EDI TAC vs AAV9-GFP TAC
<i>Lox</i>	lysyl oxidase	2.86 (1.35E-03)	1.22 (7.61E-01)	-2.34 (7.81E-03)
<i>Uck2</i>	uridine-cytidine kinase 2	2.83 (1.49E-04)	1.19 (7.11E-01)	-2.38 (1.21E-03)
<i>Rbp1</i>	retinol binding protein 1. cellular	2.82 (2.45E-03)	1.11 (8.93E-01)	-2.54 (7.66E-03)
<i fn1<="" i=""></i>	fibronectin 1	2.65 (2.29E-04)	1.06 (9.05E-01)	-2.49 (9.27E-04)
<i>Ddit4</i>	DNA-damage-inducible transcript 4	2.64 (1.02E-02)	1.57 (5.37E-01)	-1.68 (1.86E-01)
<i>Tnc</i>	tenascin C	2.58 (9.19E-04)	1.06 (9.24E-01)	-2.44 (2.50E-03)
<i>Tgfb2</i>	transforming growth factor. beta 2	2.48 (4.20E-03)	1.04 (9.60E-01)	-2.39 (8.45E-03)
<i>Adamts 1</i>	a disintegrin-like and metallopeptidase (reprolysin type) with thrombospondin type 1 motif. 1	2.28 (1.92E-04)	1.41 (2.04E-01)	-1.61 (9.71E-03)
<i>Serpine 1</i>	serine (or cysteine) peptidase inhibitor. clade E. member 1	2.28 (1.23E-02)	-1.22 (8.02E-01)	-2.79 (6.36E-03)
<i>Cst6</i>	cystatin E/M	2.22 (3.38E-03)	1.47 (3.80E-01)	-1.51 (1.12E-01)
<i>Mgp</i>	matrix Gla protein	2.22 (1.15E-03)	1.41 (3.45E-01)	-1.57 (3.94E-02)
<i>Ccnb1</i>	cyclin B1	2.18 (6.79E-03)	1.06 (9.37E-01)	-2.06 (1.54E-02)
<i>Cdk1</i>	cyclin-dependent kinase 1	2.13 (1.33E-02)	1.08 (9.16E-01)	-1.97 (3.09E-02)
<i>Myc</i>	myelocytomatosis oncogene	2.12 (1.11E-02)	1.24 (7.46E-01)	-1.71 (6.87E-02)
<i>Sparc</i>	secreted acidic cysteine rich glycoprotein	2.10 (1.41E-04)	-1.04 (9.13E-01)	-2.19 (4.38E-04)
<i>Pmp22</i>	peripheral myelin protein 22	2.05 (5.60E-05)	1.09 (7.46E-01)	-1.87 (5.09E-04)
<i>Fbln2</i>	fibulin 2	2.04 (1.67E-03)	1.43 (2.92E-01)	-1.43 (8.42E-02)
<i>Csrp2</i>	cysteine and glycine-rich protein 2	2.00 (1.69E-03)	1.01 (9.78E-01)	-1.97 (3.50E-03)

Gene symbol	Gene name	Fold change (<i>P</i> -value)		
		AAV9-GFP TAC vs Con	AAV9-EDI TAC vs Con	AAV9-EDI TAC vs AAV9-GFP TAC
<i>Bcl2</i>	B cell leukemia/lymphoma 2	1.90 (1.10E-03)	1.06 (8.96E-01)	-1.8 (3.71E-03)
<i>Ccna2</i>	cyclin A2	1.90 (1.95E-02)	-1.05 (9.47E-01)	-1.99 (1.91E-02)
<i>Ccl6</i>	chemokine (C-C motif) ligand 6	1.88 (3.18E-02)	1.49 (4.69E-01)	-1.26 (5.61E-01)
<i>Uchl1</i>	ubiquitin carboxy-terminal hydrolase L1	1.88 (6.93E-03)	1.19 (7.30E-01)	-1.58 (4.64E-02)
<i>Ptgs2</i>	prostaglandin-endoperoxide synthase 2	1.85 (7.31E-04)	1.10 (7.81E-01)	-1.68 (4.03E-03)
<i>Icam1</i>	intercellular adhesion molecule 1	1.84 (1.89E-02)	1.16 (8.13E-01)	-1.58 (8.84E-02)
<i>Cd44</i>	CD44 antigen	1.75 (5.83E-03)	-1.05 (9.20E-01)	-1.84 (6.20E-03)
<i>Vim</i>	vimentin	1.75 (7.20E-05)	1.01 (9.75E-01)	-1.74 (4.27E-04)
<i>Cd9</i>	CD9 antigen	1.74 (1.41E-04)	1.18 (3.71E-01)	-1.47 (2.57E-03)
<i>Ecm1</i>	extracellular matrix protein 1	1.74 (1.60E-02)	1.05 (9.38E-01)	-1.67 (3.43E-02)
<i>Lum</i>	lumican	1.73 (8.97E-05)	1.11 (6.05E-01)	-1.55 (9.27E-04)
<i>Sfrp1</i>	secreted frizzled-related protein 1	1.73 (1.80E-02)	1.03 (9.54E-01)	-1.67 (3.41E-02)
<i>Zfp36</i>	zinc finger protein 36	1.73 (1.35E-03)	1.13 (7.23E-01)	-1.54 (1.03E-02)
<i>Asns</i>	asparagine synthetase	1.70 (4.10E-03)	1.25 (5.00E-01)	-1.36 (8.58E-02)
<i>Cdc20</i>	cell division cycle 20	1.70 (3.02E-02)	-1.14 (8.31E-01)	-1.93 (1.55E-02)
<i>Ptprc</i>	protein tyrosine phosphatase. receptor type. C	1.70 (8.53E-02)	1.03 (9.76E-01)	-1.66 (1.21E-01)
<i>Sgk1</i>	serum/glucocorticoid regulated kinase 1	1.70 (8.46E-03)	1.38 (3.27E-01)	-1.23 (3.53E-01)
<i>Lxn</i>	latexin	1.69 (1.45E-02)	-1.03 (9.50E-01)	-1.75 (1.48E-02)

Gene symbol	Gene name	Fold change (<i>P</i> -value)		
		AAV9-GFP TAC vs Con	AAV9-EDI TAC vs Con	AAV9-EDI TAC vs AAV9-GFP TAC
<i>Anxa5</i>	annexin A5	1.67 (6.42E-04)	1.08 (7.96E-01)	-1.54 (3.58E-03)
<i>Birc5</i>	baculoviral IAP repeat-containing 5	1.67 (8.02E-02)	-1.16 (8.30E-01)	-1.95 (3.39E-02)
<i>Pycr1</i>	pyrroline-5-carboxylate reductase 1	1.66 (4.47E-02)	1.12 (8.51E-01)	-1.48 (1.43E-01)
<i>Inhba</i>	inhibin beta-A	1.65 (8.15E-04)	1.09 (7.74E-01)	-1.51 (4.91E-03)
<i>Itga1</i>	integrin alpha 1	1.65 (1.31E-03)	-1.01 (9.68E-01)	-1.67 (2.05E-03)
<i>Tspo</i>	translocator protein	1.65 (3.49E-02)	1.03 (9.56E-01)	-1.60 (6.01E-02)
<i>Crip1</i>	cysteine-rich protein 1 (intestinal)	1.64 (2.34E-04)	1.14 (5.09E-01)	-1.44 (3.33E-03)
<i>Nfkbia</i>	nuclear factor of kappa light polypeptide gene enhancer in B cells inhibitor. alpha	1.60 (5.61E-03)	1.08 (8.44E-01)	-1.48 (2.03E-02)
<i>Tagln2</i>	transgelin 2	1.60 (9.24E-04)	-1.02 (9.47E-01)	-1.64 (1.55E-03)
<i>Serpine2</i>	serine (or cysteine) peptidase inhibitor. clade E. member 2	1.59 (7.27E-02)	-1.22 (7.49E-01)	-1.93 (1.99E-02)
<i>Axl</i>	AXL receptor tyrosine kinase	1.58 (8.73E-04)	1.14 (6.08E-01)	-1.39 (1.09E-02)
<i>Fbln5</i>	fibulin 5	1.58 (1.54E-03)	1.16 (5.98E-01)	-1.37 (2.31E-02)
<i>Klf4</i>	Kruppel-like factor 4 (gut)	1.58 (1.50E-03)	1.20 (4.43E-01)	-1.32 (3.67E-02)
<i>Pam</i>	peptidylglycine alpha-amidating monooxygenase	1.58 (3.95E-05)	1.25 (3.96E-02)	-1.26 (4.52E-03)
<i>Ctdspl</i>	CTD (carboxy-terminal domain. RNA polymerase II. polypeptide A) small phosphatase-like	1.56 (1.80E-02)	1.07 (8.81E-01)	-1.46 (5.57E-02)
<i>Alb</i>	albumin	1.54 (2.73E-01)	-1.04 (9.68E-01)	-1.61 (2.70E-01)
<i>Clu</i>	clusterin	1.54 (5.26E-03)	1.24 (3.93E-01)	-1.24 (1.71E-01)
<i>Hmox1</i>	heme oxygenase 1	1.53 (7.02E-03)	1.28 (3.45E-01)	-1.20 (2.86E-01)

Gene symbol	Gene name	Fold change (<i>P</i> -value)		
		AAV9-GFP TAC vs Con	AAV9-EDI TAC vs Con	AAV9-EDI TAC vs AAV9-GFP TAC
<i>Smad1</i>	SMAD family member 1	1.53 (3.08E-03)	-1.11 (7.34E-01)	-1.70 (1.66E-03)
<i>Abcc3</i>	ATP-binding cassette, sub-family C (CFTR/MRP), member 3	1.51 (2.22E-03)	1.18 (4.49E-01)	-1.27 (5.25E-02)
<i>Mrc1</i>	mannose receptor, C type 1	1.51 (1.22E-02)	1.01 (9.83E-01)	-1.5 (2.01E-02)
<i>Tns3</i>	tensin 3	1.50 (7.00E-04)	1.03 (9.17E-01)	-1.46 (1.88E-03)
<i>Actn1</i>	actinin, alpha 1	1.49 (5.90E-04)	1.01 (9.73E-01)	-1.48 (1.52E-03)
<i>Dbn1</i>	drebrin 1	1.49 (4.01E-03)	1.07 (8.17E-01)	-1.39 (1.88E-02)
<i>Oasl2</i>	2'-5' oligoadenylate synthetase-like 2	1.49 (2.55E-01)	-1.24 (7.96E-01)	-1.84 (8.99E-02)
<i>Ccnb2</i>	cyclin B2	1.48 (2.42E-01)	-1.19 (8.24E-01)	-1.77 (8.99E-02)
<i>Mt1</i>	metallothionein 1	1.48 (3.00E-02)	1.28 (4.92E-01)	-1.16 (5.51E-01)
<i>Mcm6</i>	minichromosome maintenance complex component 6	1.46 (1.96E-01)	-1.23 (7.66E-01)	-1.80 (5.31E-02)
<i>Ccnd1</i>	cyclin D1	1.45 (2.35E-01)	-1.27 (7.30E-01)	-1.84 (5.44E-02)
<i>Egr1</i>	early growth response 1	1.45 (5.96E-01)	-1.93 (6.04E-01)	-2.80 (9.68E-02)
<i>Rrm2</i>	ribonucleotide reductase M2	1.45 (1.99E-01)	-1.20 (7.99E-01)	-1.74 (6.71E-02)
<i>Serpinh1</i>	serine (or cysteine) peptidase inhibitor, clade H, member 1	1.45 (4.18E-02)	-1.30 (4.24E-01)	-1.89 (3.84E-03)
<i>Bcl6</i>	B cell leukemia/lymphoma 6	1.43 (1.21E-01)	-1.23 (7.09E-01)	-1.76 (2.59E-02)
<i>Cald1</i>	caldesmon 1	1.43 (6.61E-03)	-1.12 (6.89E-01)	-1.6 (2.18E-03)
<i>Dusp6</i>	dual specificity phosphatase 6	1.43 (1.15E-01)	-1.11 (8.49E-01)	-1.59 (5.68E-02)
<i>Fap</i>	fibroblast activation protein	1.43 (8.53E-02)	-1.16 (7.71E-01)	-1.66 (2.55E-02)

Gene symbol	Gene name	Fold change (<i>P</i> -value)		
		AAV9-GFP TAC vs Con	AAV9-EDI TAC vs Con	AAV9-EDI TAC vs AAV9-GFP TAC
<i>Chrn1</i>	cholinergic receptor, nicotinic, beta polypeptide 1 (muscle)	1.40 (3.33E-02)	-1.20 (6.05E-01)	-1.68 (5.35E-03)
<i>Ptn</i>	pleiotrophin	1.40 (1.52E-01)	-1.12 (8.44E-01)	-1.56 (6.97E-02)
<i>Akap12</i>	A kinase (PRKA) anchor protein (gravin) 12	1.39 (1.27E-02)	-1.17 (5.43E-01)	-1.62 (2.27E-03)
<i>Flna</i>	filamin, alpha	1.38 (4.61E-02)	-1.16 (6.90E-01)	-1.60 (9.78E-03)
<i>Wisp1</i>	WNT1 inducible signaling pathway protein 1	1.36 (2.80E-01)	-1.18 (7.93E-01)	-1.61 (9.79E-02)
<i>Usp18</i>	ubiquitin specific peptidase 18	1.35 (2.49E-01)	-1.19 (7.71E-01)	-1.61 (7.10E-02)
<i>Casp8</i>	caspase 8	1.34 (1.48E-01)	-1.17 (7.50E-01)	-1.57 (3.50E-02)
<i>Fbn2</i>	fibrillin 2	1.34 (1.12E-01)	-1.24 (6.01E-01)	-1.66 (1.39E-02)
<i>Itga6</i>	integrin alpha 6	1.34 (7.79E-02)	-1.31 (3.66E-01)	-1.76 (4.31E-03)
<i>Fos</i>	FBJ osteosarcoma oncogene	1.17 (8.25E-01)	-1.94 (5.43E-01)	-2.28 (1.48E-01)
<i>Klf10</i>	Kruppel-like factor 10	-1.16 (6.50E-01)	-1.51 (4.08E-01)	-1.30 (4.53E-01)
<i>Mgst1</i>	microsomal glutathione S-transferase 1	-1.27 (4.79E-01)	1.21 (7.83E-01)	1.53 (1.76E-01)
<i>Usp54</i>	ubiquitin specific peptidase 54	-1.35 (6.26E-02)	1.11 (7.83E-01)	1.50 (2.16E-02)
<i>Got1</i>	glutamic-oxaloacetic transaminase 1, soluble	-1.40 (4.01E-02)	1.17 (6.73E-01)	1.64 (8.26E-03)
<i>Cobll1</i>	Cobl-like 1	-1.43 (5.09E-02)	1.06 (8.93E-01)	1.52 (3.17E-02)
<i>Fkbp4</i>	FK506 binding protein 4	-1.43 (1.28E-03)	-1.08 (7.37E-01)	1.33 (8.95E-03)
<i>Noct</i>	nocturnin	-1.43 (7.49E-02)	1.05 (9.20E-01)	1.51 (5.64E-02)
<i>Idh1</i>	isocitrate dehydrogenase 1 (NADP+), soluble	-1.45 (9.78E-03)	1.18 (5.26E-01)	1.71 (1.75E-03)

Gene symbol	Gene name	Fold change (<i>P</i> -value)		
		AAV9-GFP TAC vs Con	AAV9-EDI TAC vs Con	AAV9-EDI TAC vs AAV9-GFP TAC
<i>Bckdhb</i>	branched chain ketoacid dehydrogenase E1. beta polypeptide	-1.54 (1.84E-04)	-1.05 (8.33E-01)	1.47 (1.04E-03)
<i>Pfas</i>	phosphoribosylformylglycinamide synthase (FGAR amidotransferase)	-1.54 (3.57E-03)	-1.28 (2.70E-01)	1.2 (2.24E-01)
<i>Ezr</i>	ezrin	-1.55 (1.59E-04)	-1.34 (2.64E-02)	1.15 (1.14E-01)
<i>Parp1</i>	poly (ADP-ribose) polymerase family. member 1	-1.55 (1.00E-03)	-1.19 (3.94E-01)	1.30 (2.89E-02)
<i>Dusp7</i>	dual specificity phosphatase 7	-1.56 (7.30E-05)	-1.16 (2.42E-01)	1.35 (1.90E-03)
<i>Ppl</i>	periplakin	-1.57 (1.86E-02)	-1.01 (9.84E-01)	1.55 (2.80E-02)
<i>Sumo3</i>	small ubiquitin-like modifier 3	-1.58 (1.23E-02)	-1.29 (4.49E-01)	1.23 (3.31E-01)
<i>Gpt</i>	glutamic pyruvic transaminase. soluble	-1.63 (6.47E-04)	1.01 (9.86E-01)	1.64 (1.35E-03)
<i>Acss1</i>	acyl-CoA synthetase short-chain family member 1	-1.68 (1.73E-03)	1.01 (9.86E-01)	1.69 (2.91E-03)
<i>Irx3</i>	Iroquois related homeobox 3	-1.69 (8.78E-03)	-1.48 (1.85E-01)	1.14 (6.44E-01)
<i>Bmper</i>	BMP-binding endothelial regulator	-1.71 (1.58E-02)	-1.16 (7.83E-01)	1.48 (8.35E-02)
<i>F3</i>	coagulation factor III	-1.77 (3.30E-03)	-1.19 (6.65E-01)	1.49 (3.65E-02)
<i>Ppat</i>	phosphoribosyl pyrophosphate amidotransferase	-1.79 (1.71E-04)	-1.06 (8.30E-01)	1.69 (9.27E-04)
<i>Rtn2</i>	reticulon 2 (Z-band associated protein)	-1.81 (5.78E-06)	-1.32 (9.67E-03)	1.37 (7.90E-04)
<i>H19</i>	H19. imprinted maternally expressed transcript	-1.97 (5.26E-04)	-1.69 (3.11E-02)	1.17 (4.31E-01)
<i>Efemp1</i>	epidermal growth factor-containing fibulin-like extracellular matrix protein 1	-2.15 (2.84E-02)	-1.34 (7.18E-01)	1.61 (2.14E-01)
<i>Lamb3</i>	laminin. beta 3	-2.74 (2.78E-05)	-1.35 (1.74E-01)	2.03 (5.38E-04)
<i>Pfkfb1</i>	6-phosphofructo-2-kinase/fructose-2,6-bisphosphatase 1	-3.66 (2.50E-05)	-1.29 (4.13E-01)	2.83 (3.89E-04)

Gene symbol	Gene name	Fold change (<i>P</i> -value)		
		AAV9-GFP TAC vs Con	AAV9-EDI TAC vs Con	AAV9-EDI TAC vs AAV9-GFP TAC
<i>Angpt1</i>	angiopoietin 1	-5.35 (2.78E-05)	-2.20 (2.64E-02)	2.44 (2.04E-03)

Supplementary Table 11

Regulated genes associated with Myc-related gene networks in wild-type mice 4 weeks after TAC surgery and AAV9-EDI treatment. The table refers to the heat map shown in Supplementary Figure 7D; genes reported by Gene Ontology as associated with Myc-related gene networks and regulated in untreated wild-type (Wt, n=3) versus AAV9-EDI (n=4) or AAV9-GFP (n=4) treated mice after transverse aortic constriction (TAC; 4 weeks). The table indicates the gene symbol, gene name and fold-changes and *p*-values; *n*-numbers represent samples of individual mice. For statistical analysis ANOVA with Tukey *post hoc* test was applied. Source data are provided under the following link: <https://www.ebi.ac.uk/arrayexpress/experiments/E-MTAB-8110>.

Supplementary Table 12

Regulated genes associated with NFAT-related gene networks in wild-type mice 4 weeks after TAC surgery and AAV9-EDI treatment. Table refers to the heat map shown in Supplementary Figure 7D.

Gene symbol	Gene name	Fold change (p-value)		
		AAV9-GFP TAC vs Con	AAV9-EDI TAC vs Con	AAV9-EDI TAC vs AAV9- GFP TAC
<i>Myh7</i>	myosin, heavy polypeptide 7, cardiac muscle, beta	6.20 (7.95E-05)	4.53 (3.11E-2)	-1.37 (6.30E-1)
<i>Gdf15</i>	growth differentiation factor 15	5.77 (1.40E-3)	1.3 (8.18E-1)	-4.43 (6.40E-3)
<i>Ctgf</i>	connective tissue growth factor	4.84 (3.12E-5)	1.34 (4.91E-1)	-3.61 (4.21E-4)
<i>Acta1</i>	actin, alpha 1, skeletal muscle	4.49 (5.78E-6)	3.31 (5.69E-4)	-1.36 (6.62E-2)
<i>Nppb</i>	natriuretic peptide type B	3.8 (1.55E-3)	1.86 (3.25E-1)	-2.04 (5.75E-2)
<i>Nppa</i>	natriuretic peptide type A	3.74 (3.31E-5)	2.12 (1.26E-2)	-1.77 (6.44E-3)
<i>Egr2</i>	early growth response 2	3.72 (5.47E-3)	-1.03 (9.78E-1)	-3.83 (7.52E-3)
<i>Ccl8</i>	chemokine (C-C motif) ligand 8	3.43 (2.27E-3)	1.8 (3.60E-1)	-1.91 (8.90E-2)
<i>Rcan1</i>	regulator of calcineurin 1	3.15 (1.02E-4)	1.09 (8.56E-1)	-2.88 (5.38E-4)
<i>Spp1</i>	secreted phosphoprotein 1	3.08 (1.65E-2)	-1.08 (9.48E-1)	-3.33 (1.78E-2)
<i fn1<="" i=""></i>	fibronectin 1	2.65 (2.29E-4)	1.06 (9.05E-1)	-2.49 (9.27E-4)
<i>Ddit4</i>	DNA-damage-inducible transcript 4	2.64 (1.02E-2)	1.57 (5.37E-1)	-1.68 (1.86E-1)
<i>Nox4</i>	NADPH oxidase 4	2.62 (4.87E-3)	1.49 (5.18E-1)	-1.76 (9.01E-2)
<i>Cyr61</i>	cysteine rich protein 61	2.6 (1.05E-3)	-1.02 (9.81E-1)	-2.64 (1.88E-3)
<i>Fhl1</i>	four and a half LIM domains 1	2.48 (1.32E-3)	1.48 (3.56E-1)	-1.68 (4.29E-2)

Gene symbol	Gene name	Fold change (p-value)		
		AAV9-GFP TAC vs Con	AAV9-EDI TAC vs Con	AAV9-EDI TAC vs AAV9-GFP TAC
<i>Tgfb2</i>	transforming growth factor, beta 2	2.48 (4.20E-3)	1.04 (9.60E-1)	-2.39 (8.45E-3)
<i>Dkk3</i>	dickkopf WNT signaling pathway inhibitor 3	2.43 (5.63E-5)	1.26 (3.66E-1)	-1.93 (9.27E-4)
<i>Serpine 1</i>	serine (or cysteine) peptidase inhibitor, clade E, member 1	2.28 (1.23E-2)	-1.22 (8.02E-1)	-2.79 (6.36E-3)
<i>Eln</i>	elastin	2.20 (1.67E-4)	-1.00 (9.90E-1)	-2.21 (5.38E-4)
<i>Fgl2</i>	fibrinogen-like protein 2	2.2 (8.58E-3)	1.05 (9.47E-1)	-2.1 (1.81E-2)
<i>Il6</i>	interleukin 6	2.16 (1.56E-3)	1.04 (9.47E-1)	-2.09 (3.70E-3)
<i>Cdk1</i>	cyclin-dependent kinase 1	2.13 (1.33E-2)	1.08 (9.16E-1)	-1.97 (3.09E-2)
<i>Myc</i>	myelocytomatosis oncogene	2.12 (1.11E-2)	1.24 (7.46E-1)	-1.71 (6.87E-2)
<i>Mmp2</i>	matrix metalloproteinase 2	2.06 (1.45E-3)	1.06 (9.08E-1)	-1.94 (4.52E-3)
<i>Nfkbiz</i>	nuclear factor of kappa light polypeptide gene enhancer in B cells inhibitor, zeta	2.06 (1.44E-2)	1.01 (9.84E-1)	-2.03 (2.34E-2)
<i>Mapk4</i>	mitogen-activated protein kinase 4	2.05 (1.14E-3)	1.5 (1.78E-1)	-1.37 (1.14E-1)
<i>Pmp22</i>	peripheral myelin protein 22	2.05 (5.60E-5)	1.09 (7.46E-1)	-1.87 (5.09E-4)
<i>Hbegf</i>	heparin-binding EGF-like growth factor	2.02 (9.19E-4)	1.19 (6.72E-1)	-1.69 (9.32E-3)
<i>Tgfb3</i>	transforming growth factor, beta 3	1.96 (2.90E-4)	1.15 (6.73E-1)	-1.70 (2.54E-3)
<i>Ace</i>	angiotensin I converting enzyme (peptidyl-dipeptidase A) 1	1.94 (4.71E-4)	1.20 (5.47E-1)	-1.62 (6.0E-3)
<i>Bcl2</i>	B cell leukemia/lymphoma 2	1.90 (1.10E-3)	1.06 (8.96E-1)	-1.8 (3.71E-3)
<i>Haus8</i>	4HAUS augmin-like complex, subunit 8	1.90 (4.97E-3)	1.15 (7.78E-1)	-1.65 (2.54E-2)
<i>Tlr4</i>	toll-like receptor 4	1.9 (1.81E-2)	1.23 (7.38E-1)	-1.54 (1.28E-1)

Gene symbol	Gene name	Fold change (p-value)		
		AAV9-GFP TAC vs Con	AAV9-EDI TAC vs Con	AAV9-EDI TAC vs AAV9-GFP TAC
<i>Ptgs2</i>	prostaglandin-endoperoxide synthase 2	1.85 (7.31E-4)	1.1 (7.81E-1)	-1.68 (4.03E-3)
<i>Spc25</i>	SPC25, NDC80 kinetochore complex component, homolog (S. cerevisiae)	1.85 (2.61E-2)	1.08 (9.14E-1)	-1.72 (6.14E-2)
<i>Icam1</i>	intercellular adhesion molecule 1	1.84 (1.89E-2)	1.16 (8.13E-1)	-1.58 (8.84E-2)
<i>Cd14</i>	CD14 antigen	1.83 (3.36E-4)	1.03 (9.38E-1)	-1.78 (1.06E-3)
<i>Ifit2</i>	interferon-induced protein with tetratricopeptide repeats 2	1.82 (8.43E-3)	1.08 (8.90E-1)	-1.69 (2.29E-2)
<i>Atf3</i>	activating transcription factor 3	1.81 (3.91E-2)	1.12 (8.81E-1)	-1.62 (1.09E-1)
<i>Plat</i>	plasminogen activator, tissue	1.79 (0,00106)	1.06 (0,878)	-1.69 (0,00435)
<i>Egr3</i>	early growth response 3	1.77 (2.64E-3)	1.06 (9.00E-1)	-1.68 (7.73E-3)
<i>Vim</i>	vimentin	1.75 (7.20E-5)	1.01 (9.75E-1)	-1.74 (4.27E-4)
<i>Zfp36</i>	zinc finger protein 36	1.73 (1.35E-3)	1.13 (7.23E-1)	-1.54 (1.03E-2)
<i>Lcp1</i>	lymphocyte cytosolic protein 1	1.72 (9.28E-3)	-1.21 (6.98E-1)	-2.08 (2.89E-3)
<i>Mmp3</i>	matrix metalloproteinase 3	1.72 (3.76E-1)	1.65 (7.01E-1)	-1.04 (9.79E-1)
<i>Met</i>	met proto-oncogene	1.71 (9.92E-4)	1.15 (6.66E-1)	-1.49 (1.00E-2)
<i>Cdc20</i>	cell division cycle 20	1.70 (3.02E-2)	-1.14 (8.31E-1)	-1.93 (1.55E-2)
<i>Sgk1</i>	serum/glucocorticoid regulated kinase 1	1.70 (8.46E-3)	1.38 (3.27E-1)	-1.23 (3.53E-1)
<i>Birc5</i>	baculoviral IAP repeat-containing 5	1,67 (8.02E-2)	-1,16 (8.30E-1)	-1,95 (3.39E-2)
<i>Mmp14</i>	matrix metalloproteinase 14 (membrane-inserted)	1,67 (1.17E-3)	-1,05 (8.95E-1)	-1,75 (1.52E-3)
<i>Osmr</i>	oncostatin M receptor	1,67 (4.90E-4)	1,21 (3.83E-1)	-1,38 (1.51E-2)

Gene symbol	Gene name	Fold change (p-value)		
		AAV9-GFP TAC vs Con	AAV9-EDI TAC vs Con	AAV9-EDI TAC vs AAV9-GFP TAC
<i>Socs3</i>	suppressor of cytokine signaling 3	1.67 (8.52E-4)	1.02 (9.59E-1)	-1.64 (2.12E-3)
<i>Inhba</i>	inhibin beta-A	1.65 (8.15E-4)	1.09 (7.74E-1)	-1.51 (4.91E-3)
<i>Itga1</i>	integrin alpha 1	1.65 (1.31E-3)	-1.01 (9.68E-1)	-1.67 (2.05E-3)
<i>Plcg2</i>	phospholipase C, gamma 2	1.64 (1.98E-3)	1.11 (7.55E-1)	-1.48 (1.31E-2)
<i>Tgm2</i>	transglutaminase 2, C polypeptide	1.64 (4.94E-4)	1.29 (1.59E-1)	-1.27 (04.71E-2)
<i>Ctsk</i>	cathepsin K	1.63 (5.49E-3)	1.04 (9.34E-1)	-1.58 (1.30E-2)
<i>Ereg</i>	epiregulin	1.63 (1.79E-3)	-1.03 (9.28E-1)	-1.69 (2.35E-3)
<i>Nfkbia</i>	nuclear factor of kappa light polypeptide gene enhancer in B cells inhibitor, alpha	1.6 (5.61E-3)	1.08 (8.44E-1)	-1.48 (2.03E-2)
<i>Ifitm3</i>	interferon induced transmembrane protein 3	1.58 (4.02E-3)	-1.03 (9.37E-1)	-1.62 (5.19E-3)
<i>Itgb2</i>	integrin beta 2	1.58 (5.74E-3)	1.09 (8.18E-1)	-1.45 (2.53E-2)
<i>Efemp2</i>	epidermal growth factor-containing fibulin-like extracellular matrix protein 2	1,56 (1.19E-2)	1,03 (9.54E-1)	-1,52 (2.54E-2)
<i>Tlr2</i>	toll-like receptor 2	1,55 (6.11E-3)	1,14 (7.00E-1)	-1,35 (5.63E-2)
<i>Cd74</i>	CD74 antigen (invariant polypeptide of major histocompatibility complex, class II antigen-associated)	1,53 (6.06E-2)	1,02 (9.75E-1)	-1,5 (9.01E-2)
<i>Cdca8</i>	cell division cycle associated 8	1,52 (2.61E-2)	-1,03 (9.44E-1)	-1,57 (2.47E-2)
<i>Mrc1</i>	mannose receptor, C type 1	1,51 (1.22E-2)	1,01 (9.83E-1)	-1,5 (2.01E-2)
<i>Rhoj</i>	ras homolog family member J	1,5 (5.52E-2)	-1,1 (8.49E-1)	-1,65 (2.94E-2)
<i>Ptger4</i>	prostaglandin E receptor 4 (subtype EP4)	1,48 (1.15E-1)	1,08 (9.03E-1)	-1,37 (2.62E-1)
<i>C3</i>	complement component 3	1,47 (5.00E-1)	1,46 (7.44E-1)	-1,01 (9.93E-1)

Gene symbol	Gene name	Fold change (p-value)		
		AAV9-GFP TAC vs Con	AAV9-EDI TAC vs Con	AAV9-EDI TAC vs AAV9-GFP TAC
<i>Jak3</i>	Janus kinase 3	1.46 (5.97E-2)	1.05 (9.20E-1)	-1.38 (1.34E-1)
<i>Ccnd1</i>	cyclin D1	1.45 (2.35E-1)	-1.27 (7.30E-1)	-1.84 (5.44E-2)
<i>Egr1</i>	early growth response 1	1.45 (5.96E-1)	-1.93 (6.04E-1)	-2.8 (9.68E-2)
<i>Rrm2</i>	ribonucleotide reductase M2	1.45 (1.99E-1)	-1.2 (7.99E-1)	-1.74 (6.71E-2)
<i>Aif1</i>	allograft inflammatory factor 1	1.44 (1.80E-1)	-1.25 (7.24E-1)	-1.79 (4.09Ee-2)
<i>Itga9</i>	integrin alpha 9	1.44 (1.92E-2)	-1.08 (8.40E-1)	-1.55 (1.04E-2)
<i>Cfh</i>	complement component factor h	1.43 (3.38E-1)	1.2 (8.28E-1)	-1.19 (7.66E-1)
<i>Dusp6</i>	dual specificity phosphatase 6	1.43 (1.15E-1)	-1.11 (8.49E-1)	-1.59 (5.68E-2)
<i>Cdca3</i>	cell division cycle associated 3	1.41 (4.47E-2)	-1.1 (8.15E-1)	-1.56 (1.90E-2)
<i>Isg20</i>	interferon-stimulated protein	1.41 (2.60E-2)	-1.1 (8.03E-1)	-1.55 (1.03E.2)
<i>Rhoc</i>	ras homolog family member C	1.41 (1.11E-1)	-1.27 (6.20E-1)	-1.78 (1.56E-2)
<i>Cenpe</i>	centromere protein E	1.39 (1.77E-2)	-1.11 (7.30E-1)	-1.55 (5.25E.3)
<i>Cdt1</i>	chromatin licensing and DNA replication factor 1	1.36 (2.88E-1)	-1.21 (7.81E-1)	-1.64 (8.92E-2)
<i>Itga6</i>	integrin alpha 6	1.34 (7.79E-2)	-1.31 (3.66E-1)	-1.76 (4.31E-3)
<i>Nusap1</i>	nucleolar and spindle associated protein 1	1.23 (4.49E-1)	-1.3 (6.29E-1)	-1.6 (6.26E-2)
<i>Cxcl9</i>	chemokine (C-X-C motif) ligand 9	1.18 (6.35E-1)	-1.3 (6.97E-1)	-1.53 (1.63E-1)
<i>Ccl2</i>	chemokine (C-C motif) ligand 2	1.15 (2.98E-1)	-1.02 (9.47E-1)	-1.17 (2.52E-1)
<i>Nfatc4</i>	nuclear factor of activated T cells, cytoplasmic, calcineurin dependent 4	1.12 (4.83E-2)	1.02 (8.94E-1)	-1.09 (1.26E-1)

Gene symbol	Gene name	Fold change (p-value)		
		AAV9-GFP TAC vs Con	AAV9-EDI TAC vs Con	AAV9-EDI TAC vs AAV9-GFP TAC
<i>Csf2</i>	colony stimulating factor 2 (granulocyte-macrophage)	1.11 (5.22E-1)	1.17 (6.07E-1)	1.05 (8.54E-1)
<i>Nfatc2</i>	nuclear factor of activated T cells, cytoplasmic, calcineurin dependent 2	1.03 (5.95E-1)	-1.01 (9.07E-1)	-1.05 (4.52E-1)
<i>Cxcl3</i>	chemokine (C-X-C motif) ligand 3	1.02 (8.57E-1)	1.08 (7.38E-1)	1.05 (7.42E-1)
<i>Nfat5</i>	nuclear factor of activated T cells 5	1.00 (9.69E-1)	1.03 (8.78E-1)	1.02 (8.74E-1)
<i>Nfatc1</i>	nuclear factor of activated T cells, cytoplasmic, calcineurin dependent 1	1.00 (9.73E-1)	-1.01 (9.55E-1)	-1.01 (9.36E-1)
<i>Nfatc3</i>	nuclear factor of activated T cells, cytoplasmic, calcineurin dependent 3	-1.11 (4.55E-1)	-1.07 (8.26E-1)	1.04 (8.90E-1)
<i>Gna12</i>	guanine nucleotide binding protein, alpha 12	-1.40 (4.21E-2)	1.18 (6.45E-1)	1.66 (7.43E-3)
<i>Mapk10</i>	mitogen-activated protein kinase 10	-1.46 (1.88E-3)	1.08 (7.78E-1)	1.57 (1.42E-3)
<i>Lifr</i>	leukemia inhibitory factor receptor	-1.5 (1.11E-3)	-1.07 (7.83E-1)	1.40 (6.29E-3)
<i>Hdac11</i>	histone deacetylase 11	-1.55 (1.96E-4)	-1.20 (1.92E-1)	1.29 (1.14E-2)
<i>Adrb1</i>	adrenergic receptor, beta 1	-1.59 (2.27E-2)	-1.40 (3.19E-1)	1.13 (6.87E-1)
<i>Cacnb2</i>	calcium channel, voltage-dependent, beta 2 subunit	-1.59 (3.87E-3)	-1.03 (9.44E-1)	1.55 (8.97E-3)
<i>Mme</i>	membrane metallo endopeptidase	-1.61 (2.48E-3)	-1.00 (9.96E-1)	1.61 (4.21E-3)
<i>Ctf1</i>	cardiotrophin 1	-1.64 (4.27E-4)	-1.04 (9.00E-1)	1.59 (1.52E-3)
<i>Myocd</i>	myocardin	-1.64 (7.28E-5)	-1.25 (8.60E-2)	1.31 (5.79E-3)
<i>Rgs2</i>	regulator of G-protein signaling 2	-2.21 (1.61E-3)	-1.77 (8.71E-2)	1.25 (3.97E-1)
<i>Kcnd2</i>	potassium voltage-gated channel, Shal-related family, member 2	-2.23 (4.72E-4)	-1.49 (1.74E-1)	1.50 (03.97E-2)
<i>Enpp2</i>	ectonucleotide pyrophosphatase/phosphodiesterase 2	-2.53 (2.84E-3)	-1.25 (7.24E-1)	2.02 (2.01E-2)

Gene symbol	Gene name	Fold change (p-value)		
		AAV9-GFP TAC vs Con	AAV9-EDI TAC vs Con	AAV9-EDI TAC vs AAV9-GFP TAC
<i>Il15</i>	interleukin 15	-3.25 (7.41E-5)	-1.41 (3.01E-1)	2.31 (1.53E-3)

Supplementary Table 12

Regulated genes associated with NFAT-related gene networks in wild-type mice 4 weeks after TAC surgery and AAV9-EDI treatment. The table refers to the heat map shown in Supplementary Figure 7D; genes reported by Gene Ontology as associated with NFAT-related gene networks and regulated in untreated wild-type (Wt, n=3) versus AAV9-EDI (n=4) or AAV9-GFP (n=4) treated mice after transverse aortic constriction (TAC; 4 weeks). The table indicates the gene symbol, gene name and fold-changes and *p*-values; *n*-numbers represent samples of individual mice. For statistical analysis ANOVA with Tukey *post hoc* test was applied. Source data are provided under the following link: <https://www.ebi.ac.uk/arrayexpress/experiments/E-MTAB-8110>.

Supplementary Table 13

Top 100 regulated genes in LS174T cells transduced with LacZ (Con) or EDI. The table refers to the heat map shown in Fig. 5F.

Gene symbol	Gene name	Fold change (<i>P</i>-value) AAV9-EDI vs AAV9-LacZ
<i>Kcng2</i>	potassium voltage-gated channel modifier subfamily G member 2	5.80 (1.10E-07)
<i>Reg1b</i>	regenerating family member 1 beta	4.18 (3.47E-07)
<i>Hist2h2be</i>	histone cluster 2 H2B family member e	3.99 (5.69E-06)
<i>Agpat4</i>	1-acylglycerol-3-phosphate O-acyltransferase 4	3.72 (2.62E-06)
<i>Fxyd4</i>	FXYP domain containing ion transport regulator 4	3.48 (2.89E-05)
<i>Hla-dma</i>	major histocompatibility complex, class II, DM alpha	3.40 (3.00E-04)
<i>Tff2</i>	trefoil factor 2	3.40 (1.96E-05)
<i>Hist1h2bd</i>	histone cluster 1 H2B family member d	3.24 (1.93E-05)
<i>Stc1</i>	stanniocalcin 1	3.24 (1.08E-08)
<i>Loc105375773</i>	-	3.12 (2.98E-06)
<i>Krt20</i>	keratin 20	3.08 (1.94E-03)
<i>Anxa10</i>	annexin A10	3.07 (4.47E-04)
<i>Linc00520</i>	long intergenic non-protein coding RNA 520	3.01 (2.15E-05)
<i>Dpqr1</i>	diffuse panbronchiolitis critical region 1	2.94 (2.69E-04)
<i>Loc102725220</i>	-	2.89 (8.29E-05)
<i>Ceacam5</i>	carcinoembryonic antigen related cell adhesion molecule 5	2.81 (2.84E-03)
<i>Loc102723684</i>	-	2.76 (2.63E-03)
<i>Tm4sf20</i>	transmembrane 4 L six family member 20	2.75 (2.15E-05)

Gene symbol	Gene name	Fold change (<i>P</i>-value) AAV9-EDI vs AAV9-LacZ
<i>Diras2</i>	DIRAS family GTPase 2	2.71 (1.66E-05)
<i>Mmp1</i>	matrix metalloproteinase 1	2.68 (7.74E-05)
<i>Tff1</i>	trefoil factor 1	2.61 (1.04E-03)
<i>Srpx</i>	sushi repeat containing protein, X-linked	2.57 (1.81E-05)
<i>Hist1h3d</i>	histone cluster 1 H3 family member d	2.53 (1.85E-03)
<i>Otub2</i>	OTU deubiquitinase, ubiquitin aldehyde binding 2	2.53 (1.09E-06)
<i>Ceacam6</i>	carcinoembryonic antigen related cell adhesion molecule 6	2.48 (2.00E-03)
<i>Ndrp1</i>	N-myc downstream regulated 1	2.47 (1.33E-05)
<i>Nts</i>	neurotensin	2.45 (5.81E-04)
<i>Dfb1</i>	defensin beta 1	2.42 (3.84E-05)
<i>Capn10-as1</i>	CAPN10 antisense RNA 1 (head to head)	2.42 (7.42E-06)
<i>Hmgcs2</i>	3-hydroxy-3-methylglutaryl-CoA synthase 2	2.34 (1.13E-03)
<i>Mgp</i>	matrix Gla protein	2.20 (3.37E-02)
<i>Ccl26</i>	C-C motif chemokine ligand 26	2.07 (1.05E-02)
<i>Itih2a</i>	integral membrane protein 2A	-2.43 (2.07E-04)
<i>Stil</i>	SCL/TAL1 interrupting locus	-2.43 (7.06E-05)
<i>Kif4a</i>	kinesin family member 4A	-2.44 (1.47E-06)
<i>Psrc1</i>	proline and serine rich coiled-coil 1	-2.46 (4.88E-05)
<i>Mir924hg</i>	MIR924 host gene	-2.48 (8.48E-04)

Gene symbol	Gene name	Fold change (<i>P</i>-value) AAV9-EDI vs AAV9-LacZ
<i>Ube2c</i>	ubiquitin conjugating enzyme E2 C	-2.49 (2.08E-07)
<i>Sacs</i>	sacsin molecular chaperone	-2.49 (4.17E-07)
<i>Kn11</i>	kinetochore scaffold 1	-2.49 (7.72E-07)
<i>Linc01224</i>	long intergenic non-protein coding RNA 1224	-2.5 (3.17E-05)
<i>Mastl</i>	microtubule associated serine/threonine kinase like	-2.53 (4.12E-05)
<i>Kif2c</i>	kinesin family member 2C	-2.54 (2.10E-06)
<i>Ncapg2</i>	non-SMC condensin II complex subunit G2	-2.54 (6.79E-08)
<i>Cdk1</i>	cyclin dependent kinase 1	-2.56 (3.68E-10)
<i>Cdca8</i>	cell division cycle associated 8	-2.58 (6.82E-08)
<i>Ccdc77</i>	coiled-coil domain containing 77	-2.58 (1.39E-08)
<i>Cenpf</i>	centromere protein F	-2.59 (1.31E-08)
<i>Cdc20</i>	cell division cycle 20	-2.60 (7.22E-07)
<i>Nek2</i>	NIMA related kinase 2	-2.60 (5.17E-07)
<i>Bora</i>	bora, aurora kinase A activator	-2.63 (6.24E-07)
<i>Brip1</i>	BRCA1 interacting protein C-terminal helicase 1	-2.70 (6.52E-07)
<i>Ncaph</i>	non-SMC condensin I complex subunit H	-2.70 (8.75E-08)
<i>C1orf112</i>	chromosome 1 open reading frame 112	-2.70 (2.24E-07)
<i>Prim1</i>	primase (DNA) subunit 1	-2.72 (1.58E-05)
<i>Asf1b</i>	anti-silencing function 1B histone chaperone	-2.72 (7.44E-05)

Gene symbol	Gene name	Fold change (<i>P</i>-value) AAV9-EDI vs AAV9-LacZ
<i>Ccna2</i>	cyclin A2	-2.72 (3.57E-10)
<i>Ascl2</i>	achaete-scute family bHLH transcription factor 2	-2.74 (9.50E-08)
<i>Cenpa</i>	centromere protein A	-2.75 (1.24E-08)
<i>Aspm</i>	abnormal spindle microtubule assembly	-2.75 (1.05E-06)
<i>Gen1</i>	GEN1, Holliday junction 5' flap endonuclease	-2.77 (1.39E-04)
<i>Cenph</i>	centromere protein H	-2.79 (6.52E-08)
<i>Mnd1</i>	meiotic nuclear divisions 1	-2.80 (8.58E-06)
<i>Nmu</i>	neuromedin U	-2.80 (1.83E-04)
<i>Pbk</i>	PDZ binding kinase	-2.80 (4.04E-09)
<i>Ccnb1</i>	cyclin B1	-2.82 (7.49E-08)
<i>Pif1</i>	PIF1 5'-to-3' DNA helicase	-2.84 (2.30E-05)
<i>Ttf2</i>	transcription termination factor 2	-2.85 (4.87E-06)
<i>Kif23</i>	kinesin family member 23	-2.85 (5.69E-06)
<i>Tmed10</i>	transmembrane p24 trafficking protein 10	-2.86 (1.85E-08)
<i>Taf5</i>	TATA-box binding protein associated factor 5	-2.87 (6.45E-10)
<i>Hjurp</i>	Holliday junction recognition protein	-2.88 (1.69E-07)
<i>Kif15</i>	kinesin family member 15	-2.88 (8.62E-08)
<i>Rad51ap1</i>	RAD51 associated protein 1	-2.90 (1.13E-07)
<i>Arhgap19</i>	Rho GTPase activating protein 19	-2.91 (4.38E-08)

Gene symbol	Gene name	Fold change (<i>P</i>-value) AAV9-EDI vs AAV9-LacZ
<i>Gins1</i>	GINS complex subunit 1	-2.92 (1.38E-08)
<i>Depdc1b</i>	DEP domain containing 1B	-2.93 (8.71E-06)
<i>Kif20a</i>	kinesin family member 20A	-2.93 (2.42E-08)
<i>Trappc4</i>	trafficking protein particle complex 4	-2.94 (2.23E-08)
<i>Kif14</i>	kinesin family member 14	-2.99 (6.32E-08)
<i>Hmmr</i>	hyaluronan mediated motility receptor	-3.02 (5.29E-09)
<i>Tex30</i>	testis expressed 30	-3.04 (7.54E-09)
<i>Mtfr2</i>	mitochondrial fission regulator 2	-3.06 (4.48E-09)
<i>Sgo2</i>	shugoshin 2	-3.14 (3.23E-07)
<i>Ncapg</i>	non-SMC condensin I complex subunit G	-3.19 (7.88E-08)
<i>Oip5</i>	Opa interacting protein 5	-3.21 (4.08E-09)
<i>Vrk1</i>	vaccinia related kinase 1	-3.21 (2.71E-08)
<i>Depdc1</i>	DEP domain containing 1	-3.22 (1.87E-08)
<i>Anln</i>	anillin actin binding protein	-3.22 (2.22E-08)
<i>Nuf2</i>	NUF2, NDC80 kinetochore complex component	-3.22 (4.08E-09)
<i>Ttk</i>	TTK protein kinase	-3.30 (8.60E-09)
<i>Iglv2-14</i>	immunoglobulin lambda variable 2-14	-3.31 (1.12E-05)
<i>Cenpe</i>	centromere protein E	-3.32 (2.84E-09)
<i>Dlgap5</i>	DLG associated protein 5	-3.41 (3.02E-09)

Gene symbol	Gene name	Fold change (<i>P</i>-value) AAV9-EDI vs AAV9-LacZ
<i>Kif11</i>	kinesin family member 11	-3.48 (4.08E-09)
<i>Arl6ip1</i>	ADP ribosylation factor like GTPase 6 interacting protein 1	-3.52 (5.95E-09)
<i>Spc25</i>	SPC25, NDC80 kinetochore complex component	-3.61 (1.19E-06)
<i>Ska1</i>	spindle and kinetochore associated complex subunit 1	-3.65 (3.26E-06)
<i>Dtymk</i>	deoxythymidylate kinase	-3.84 (2.32E-08)
<i>Ndc80</i>	NDC80, kinetochore complex component	-4.23 (3.89E-09)

Supplementary Table 13

Top 100 regulated genes in LS174T cells transduced with LacZ (Con) or EDI. The table refers to the heat map shown in Fig. 5E; microarray gene expression analysis of LS174 cells transduced with LacZ (Con) or EDI (n=3 per condition). The table indicates the gene symbol, gene name and fold-changes and *p*-values; *n*-numbers represent biologically independent experiments. For statistical analysis unpaired two-sided Student's *t*-test was applied. Source data are provided under the following link: <http://www.ebi.ac.uk/arrayexpress/experiments/E-MTAB-8111>.

Supplementary Table 14**Regulated genes associated with proliferation in LS174T cells transduced with LacZ (Con) or EDI.** The table refers to the heat map shown in Fig. 5E

Gene symbol	Gene name	Fold change (<i>P</i>-value) AAV9-EDI vs AAV9-LacZ
<i>Tspan1</i>	tetraspanin 1	1.64 (1.95E-02)
<i>Traip</i>	TRAF interacting protein	-2.20 (2.76E-06)
<i>Ndrp1</i>	N-myc downstream regulated 1	2.47 (1.33E-05)
<i>Cenpf</i>	centromere protein F	-2.59 1.31E-08
<i>Kif2c</i>	kinesin family member 2C	-2.54 (2.10E-06)
<i>Crip2</i>	cysteine rich protein 2	1.59 (1.58E-02)
<i>Cse1l</i>	chromosome segregation 1 like	-1.53 (6.94E-08)
<i>Dkc1</i>	dyskerin pseudouridine synthase 1	-1.58 (5.66E-07)
<i>Dnmt1</i>	DNA methyltransferase 1	-2.24 (9.14E-07)
<i>Dtymk</i>	deoxythymidylate kinase	-3.84 (2.32E-08)
<i>Ezh2</i>	enhancer of zeste 2 polycomb repressive complex 2 subunit	-1.59 (1.41E-04)
<i>Fabp1</i>	fatty acid binding protein 1	1.75 (6.99E-04)
<i>Tpx2</i>	TPX2, microtubule nucleation factor	-2.22 (1.34E-06)
<i>Foxm1</i>	forkhead box M1	-1.52 (7.87E-04)
<i>Angpt1</i>	angiopoietin 1	1.89 (1.75E-06)
<i>Grk5</i>	G protein-coupled receptor kinase 5	1.58 (1.66E-03)
<i>Hdgf</i>	heparin binding growth factor	-1.93 (1.93E-06)
<i>Birc5</i>	baculoviral IAP repeat containing 5	-2.05 (3.95E-06)

Gene symbol	Gene name	Fold change (<i>P</i>-value) AAV9-EDI vs AAV9-LacZ
<i>Rbpj</i>	recombination signal binding protein for immunoglobulin kappa J region	-1.77 (1.33E-05)
<i>Cxcr2</i>	C-X-C motif chemokine receptor 2	1.82 (7.34E-03)
<i>Il15</i>	interleukin 15	-1.86 (4.58E-04)
<i>Isg20</i>	interferon stimulated exonuclease gene 20	2.03 (1.21E-05)
<i>Itga2</i>	integrin subunit alpha 2	1.54 (5.87E-03)
<i>Trnp1</i>	TMF1-regulated nuclear protein 1	1.63 (8.15E-03)
<i>Mxd1</i>	MAX dimerization protein 1	1.65 (7.53E-04)
<i>Mcm7</i>	minichromosome maintenance complex component 7	-1.93 (6.07E-06)
<i>Mki67</i>	marker of proliferation Ki-67	-2.07 (5.77E-05)
<i>Mre11</i>	MRE11 homolog, double strand break repair nuclease	-1.92 (4.66E-05)
<i>Nasp</i>	nuclear autoantigenic sperm protein	-1.95 (2.35E-06)
<i>Ceacam6</i>	carcinoembryonic antigen related cell adhesion molecule 6	2.48 (2.00E-03)
<i>Hp1bp3</i>	heterochromatin protein 1 binding protein 3	-1.96 (2.22E-08)
<i>Sidt2</i>	SID1 transmembrane family member 2	2.34 (4.89E-08)
<i>Pcna</i>	proliferating cell nuclear antigen	-1.70 (1.36E-04)
<i>Ier5</i>	immediate early response 5	-1.51 (6.05E-04)
<i>Plac8</i>	placenta specific 8	1.78 (3.91E-03)
<i>Map3k20</i>	mitogen-activated protein kinase kinase kinase 20	-1.65 (3.07E-06)
<i>Plk1</i>	polo like kinase 1	-2.29 (4.42E-05)

Gene symbol	Gene name	Fold change (<i>P</i>-value) AAV9-EDI vs AAV9-LacZ
<i>Pola1</i>	DNA polymerase alpha 1, catalytic subunit	-1.55 (3.05E-05)
<i>Tipin</i>	TIMELESS interacting protein	-1.70 (7.63E-05)
<i>Pak1ip1</i>	PAK1 interacting protein 1	-1.58 (6.63E-03)
<i>Phip</i>	pleckstrin homology domain interacting protein	-1.67 (1.62E-05)
<i>Styk1</i>	serine/threonine/tyrosine kinase 1	1.64 (3.23E-04)
<i>Mcm10</i>	minichromosome maintenance 10 replication initiation factor	-2.03 (2.50E-06)
<i>Cdca7l</i>	cell division cycle associated 7 like	-1.57 (1.15E-04)
<i>Prox1</i>	prospero homeobox 1	1.63 (5.50E-04)
<i>Kif15</i>	kinesin family member 15	-2.88 (8.62E-08)
<i>Cnot6</i>	CCR4-NOT transcription complex subunit 6	-1.70 (5.40E-07)
<i>Rac2</i>	ras-related C3 botulinum toxin substrate 2 (rho family, small GTP binding protein Rac2)	1.54 (7.29E-03)
<i>Ccnd1</i>	cyclin D1	-1.57 (2.64E-04)
<i>Reg1b</i>	regenerating family member 1 beta	4.18 (3.47E-07)
<i>Rpa3</i>	replication protein A3	-1.73 (2.05E-05)
<i>Prdm1</i>	PR/SET domain 1	1.53 (4.21E-03)
<i>Stil</i>	SCL/TAL1 interrupting locus	-2.43 (7.06E-05)
<i>Ndrp4</i>	NDRG family member 4	1.63 (2.09E-04)
<i>Skp2</i>	S-phase kinase associated protein 2	-1.50 (1.04E-03)
<i>Brca1</i>	BRCA1, DNA repair associated	-2.26 (2.71E-08)

Gene symbol	Gene name	Fold change (<i>P</i>-value) AAV9-EDI vs AAV9-LacZ
<i>Stc1</i>	stanniocalcin 1	3.24 (1.08E-08)
<i>Tcf19</i>	transcription factor 19	-1.55 (4.45E-03)
<i>Bub1</i>	BUB1 mitotic checkpoint serine/threonine kinase	-2.45 (5.09E-07)
<i>Bub1b</i>	BUB1 mitotic checkpoint serine/threonine kinase B	-2.48 (5.05E-09)
<i>Tgfb1</i>	transforming growth factor beta induced	1.61 (5.78E-03)
<i>Tgfb3</i>	transforming growth factor beta receptor 3	-1.62 (4.08E-03)
<i>Tnfrsf1b</i>	TNF receptor superfamily member 1B	1.52 (2.77E-02)
<i>Ttk</i>	TTK protein kinase	-3.30 (8.60E-09)
<i>Ccr2</i>	C-C motif chemokine receptor 2	1.81 (4.36E-05)
<i>E2f8</i>	E2F transcription factor 8	-2.38 (1.58E-06)
<i>Nr4a3</i>	nuclear receptor subfamily 4 group A member 3	1.52 (1.78E-03)
<i>Fam83d</i>	family with sequence similarity 83 member D	-1.82 (1.01E-06)
<i>Cdc7</i>	cell division cycle 7	-2.13 (8.62E-08)
<i>Lgr5</i>	leucine rich repeat containing G protein-coupled receptor 5	-1.61 (1.97E-03)
<i>Ccnb1</i>	cyclin B1	-2.82 (7.49E-08)
<i>Usp13</i>	ubiquitin specific peptidase 13 (isopeptidase T-3)	-1.53 (5.73E-04)
<i>Prc1</i>	protein regulator of cytokinesis 1	-1.99 (1.34E-07)
<i>Aurkb</i>	aurora kinase B	-2.10 (4.11E-06)
<i>Klf4</i>	Kruppel like factor 4	1.57 (2.75E-02)

Gene symbol	Gene name	Fold change (<i>P</i>-value) AAV9-EDI vs AAV9-LacZ
<i>Lipg</i>	lipase G, endothelial type	-1.68 (3.50E-04)
<i>Kif20b</i>	kinesin family member 20B	-2.04 (4.17E-04)
<i>Dlgap5</i>	DLG associated protein 5	-3.41 (3.02E-09)
<i>Cdk1</i>	cyclin dependent kinase 1	-2.56 (3.68E-10)
<i>Melk</i>	maternal embryonic leucine zipper kinase	-2.02 (4.99E-09)
<i>Cdc20</i>	cell division cycle 20	-2.60 (7.22E-07)
<i>Arnt2</i>	aryl hydrocarbon receptor nuclear translocator 2	1.56 (7.24E-03)
<i>Kif14</i>	kinesin family member 14	-2.99 (6.32E-08)
<i>Cdc25c</i>	cell division cycle 25C	-2.27 (3.15E-04)
<i>Fgfbp1</i>	fibroblast growth factor binding protein 1	1.58 (1.35E-03)

Supplementary Table 14

Regulated genes associated with proliferation in LS174T cells transduced with LacZ (Con) or EDI. The table refers to the heat map shown in Fig. 5E; microarray gene expression analysis of LS174 cells transduced with LacZ (Con) or EDI (n=3 per condition). The table indicates the gene symbol, gene name and fold-changes and *p*-values; *n*-numbers represent biologically independent experiments. For statistical analysis unpaired two-sided Student's *t*-test was applied. Source data are provided under the following link: <http://www.ebi.ac.uk/arrayexpress/experiments/E-MTAB-8111>.

Supplementary Table 15

Regulated genes associated with cell cycle in LS174T cells transduced with LacZ (Con) or EDI. The table refers to the heat map shown in Fig. 5E.

Gene symbol	Gene name	Fold change (<i>P</i>-value) AAV9-EDI vs AAV9-LacZ
<i>Rgs2</i>	regulator of G-protein signaling 2	2.43 (6.15E-07)
<i>Trnp1</i>	TMF1-regulated nuclear protein 1	1.63 (8.15E-03)
<i>Nt5e</i>	5'-nucleotidase ecto	1.58 (5.38E-03)
<i>Zwint</i>	ZW10 interacting kinetochore protein	-1.52 (4.56E-05)
<i>Foxm1</i>	forkhead box M1	-1.52 (7.87E-04)
<i>Hace1</i>	HECT domain and ankyrin repeat containing E3 ubiquitin protein ligase 1	-1.53 (1.60E-03)
<i>Erh</i>	enhancer of rudimentary homolog (Drosophila)	-1.54 (6.14E-05)
<i>Lig1</i>	DNA ligase 1	-1.55 (3.19E-03)
<i>Prkcd</i>	protein kinase C delta	-1.55 (1.69E-02)
<i>Suv39h2</i>	suppressor of variegation 3-9 homolog 2	-1.55 (1.32E-03)
<i>Cdt1</i>	chromatin licensing and DNA replication factor 1	-1.56 (7.88E-05)
<i>Ccnd1</i>	cyclin D1	-1.57 (2.64E-04)
<i>Ccne2</i>	cyclin E2	-1.57 (1.60E-02)
<i>Cenps</i>	centromere protein S	-1.64 (2.56E-05)
<i>Rhno1</i>	RAD9-HUS1-RAD1 interacting nuclear orphan 1	-1.64 (7.26E-06)
<i>Ncapd3</i>	non-SMC condensin II complex subunit D3	-1.65 (3.07E-06)
<i>Timeless</i>	timeless circadian clock	-1.65 (1.66E-04)
<i>Kpna2</i>	karyopherin subunit alpha 2	-1.66 (1.18E-04)

Gene symbol	Gene name	Fold change (<i>P</i>-value) AAV9-EDI vs AAV9-LacZ
<i>Haus4</i>	HAUS augmin like complex subunit 4	-1.66 (5.38E-04)
<i>Usp37</i>	ubiquitin specific peptidase 37	-1.66 (8.18E-07)
<i>Esco2</i>	establishment of sister chromatid cohesion N-acetyltransferase 2	-1.67 (1.46E-03)
<i>Chaf1a</i>	chromatin assembly factor 1 subunit A	-1.68 (8.46E-05)
<i>Zwilch</i>	zwilch kinetochore protein	-1.68 (4.59E-08)
<i>Cntrl</i>	centriolin	-1.69 (1.33E-04)
<i>Katna1</i>	katanin catalytic subunit A1	-1.69 (3.17E-05)
<i>Chtf18</i>	chromosome transmission fidelity factor 18	-1.69 (1.54E-03)
<i>Tipin</i>	TIMELESS interacting protein	-1.70 (7.63E-05)
<i>Chaf1b</i>	chromatin assembly factor 1 subunit B	-1.72 (2.08E-05)
<i>Arpp19</i>	cAMP regulated phosphoprotein 19	-1.74 (6.57E-05)
<i>Eri1</i>	exoribonuclease 1	-1.75 (1.60E-07)
<i>Kntc1</i>	kinetochore associated 1	-1.77 (6.14E-06)
<i>Pimreg</i>	PICALM interacting mitotic regulator	-1.78 (2.96E-04)
<i>Cenpw</i>	centromere protein W	-1.80 (8.96E-06)
<i>Wrn</i>	Werner syndrome RecQ like helicase	-1.80 (4.08E-05)
<i>Fam83d</i>	family with sequence similarity 83 member D	-1.82 (1.01E-06)
<i>Nup37</i>	nucleoporin 37	-1.85 (3.16E-07)
<i>Nedd1</i>	neural precursor cell expressed, developmentally down-regulated 1	-1.87 (6.46E-07)

Gene symbol	Gene name	Fold change (<i>P</i>-value) AAV9-EDI vs AAV9-LacZ
<i>Nup43</i>	nucleoporin 43	-1.89 (5.05E-03)
<i>Cdca5</i>	cell division cycle associated 5	-1.90 (8.04E-06)
<i>Hells</i>	helicase, lymphoid-specific	-1.91 (2.70E-05)
<i>Ncapd2</i>	non-SMC condensin I complex subunit D2	-1.94 (2.41E-05)
<i>Nasp</i>	nuclear autoantigenic sperm protein	-1.95 (2.35E-06)
<i>Aurka</i>	aurora kinase A	-1.96 (1.39E-08)
<i>Smc2</i>	structural maintenance of chromosomes 2	-1.97 (2.42E-07)
<i>Mis18a</i>	MIS18 kinetochore protein A	-1.97 (2.71E-08)
<i>Knstrn</i>	kinetochore localized astrin/SPAG5 binding protein	-1.99 (3.76E-08)
<i>Haus1</i>	HAUS augmin like complex subunit 1	-2.01 (6.00E-05)
<i>Haus7</i>	HAUS augmin like complex subunit 7	-2.02 (8.53E-06)
<i>Cdc6</i>	cell division cycle 6	-2.02 (3.00E-04)
<i>Haus8</i>	HAUS augmin like complex subunit 8	-2.03 (3.26E-06)
<i>Kif20b</i>	kinesin family member 20B	-2.04 (4.17E-04)
<i>Birc5</i>	baculoviral IAP repeat containing 5	-2.05 (3.95E-06)
<i>Kifc1</i>	kinesin family member C1	-2.07 (2.54E-06)
<i>Mki67</i>	marker of proliferation Ki-67	-2.07 (5.77E-05)
<i>Mis18bp1</i>	MIS18 binding protein 1	-2.11 (5.61E-06)
<i>Smc4</i>	structural maintenance of chromosomes 4	-2.13 (3.24E-07)

Gene symbol	Gene name	Fold change (<i>P</i>-value) AAV9-EDI vs AAV9-LacZ
<i>Rad21</i>	RAD21 cohesin complex component	-2.13 (3.52E-07)
<i>Cdc7</i>	cell division cycle 7	-2.13 (8.62E-08)
<i>Cdca2</i>	cell division cycle associated 2	-2.14 (1.31E-07)
<i>Cdca3</i>	cell division cycle associated 3	-2.14 (1.40E-06)
<i>Cdknn3</i>	cyclin dependent kinase inhibitor 3	-2.15 (2.40E-07)
<i>Kif18b</i>	kinesin family member 18B	-2.16 (1.27E-06)
<i>Spag5</i>	sperm associated antigen 5	-2.18 (9.56E-07)
<i>Lin9</i>	lin-9 DREAM MuvB core complex component	-2.18 (2.45E-06)
<i>Tpx2</i>	TPX2, microtubule nucleation factor	-2.22 (1.34E-06)
<i>Pttg1</i>	pituitary tumor-transforming 1	-2.25 (3.10E-07)
<i>Cdc25c</i>	cell division cycle 25C	-2.27 (3.15E-04)
<i>Mad2l1</i>	mitotic arrest deficient 2 like 1	-2.33 (3.00E-07)
<i>Cenpj</i>	centromere protein J	-2.41 (7.38E-07)
<i>Bub1</i>	BUB1 mitotic checkpoint serine/threonine kinase	-2.45 (5.09E-07)
<i>Ccnb2</i>	cyclin B2	-2.45 (3.70E-07)
<i>Psrc1</i>	proline and serine rich coiled-coil 1	-2.46 (4.88E-05)
<i>Bub1b</i>	BUB1 mitotic checkpoint serine/threonine kinase B	-2.48 (5.05E-09)
<i>Ube2c</i>	ubiquitin conjugating enzyme E2 C	-2.49 (2.08E-07)
<i>Kn11</i>	kinetochore scaffold 1	-2.49 (7.72E-07)

Gene symbol	Gene name	Fold change (<i>P</i>-value) AAV9-EDI vs AAV9-LacZ
<i>Fbxo5</i>	F-box protein 5	-2.50 (7.99E-09)
<i>Mastl</i>	microtubule associated serine/threonine kinase like	-2.53 (4.12E-05)
<i>Kif2c</i>	kinesin family member 2C	-2.54 (2.10E-06)
<i>Ncapg2</i>	non-SMC condensin II complex subunit G2	-2.54 (6.79E-08)
<i>Cdk1</i>	cyclin dependent kinase 1	-2.56 (3.68E-10)
<i>Cenpf</i>	centromere protein F	-2.59 (1.31E-08)
<i>Nek2</i>	NIMA related kinase 2	-2.60 (5.17E-07)
<i>Cdc20</i>	cell division cycle 20	-2.60 (7.22E-07)
<i>Bora</i>	bora, aurora kinase A activator	-2.63 (6.24E-07)
<i>Ncaph</i>	non-SMC condensin I complex subunit H	-2.70 (8.75E-08)
<i>Ccna2</i>	cyclin A2	-2.72 (3.57E-10)
<i>Ccnb1</i>	cyclin B1	-2.82 (7.49E-08)
<i>Hjurp</i>	Holliday junction recognition protein	-2.88 (1.69E-07)
<i>Kif14</i>	kinesin family member 14	-2.99 (6.32E-08)
<i>Sgo2</i>	shugoshin 2	-3.14 (3.23E-07)
<i>Oip5</i>	Opa interacting protein 5	-3.21 (4.08E-09)
<i>Vrk1</i>	vaccinia related kinase 1	-3.21 (2.71E-08)
<i>Cenpe</i>	centromere protein E	-3.32 (2.84E-09)
<i>Kif11</i>	kinesin family member 11	-3.48 (4.08E-09)

Gene symbol	Gene name	Fold change (<i>P</i>-value) AAV9-EDI vs AAV9-LacZ
<i>Dtymk</i>	deoxythymidylate kinase	-3.84 (2.32E-08)
<i>Ndc80</i>	NDC80, kinetochore complex component	-4.23 (3.89E-09)

Supplementary Table 15

Regulated genes associated with cell cycle in LS174T cells transduced with LacZ (Con) or

EDI. The table refers to the heat map shown in Fig. 5E; microarray gene expression analysis of LS174 cells transduced with LacZ (Con) or EDI (n=3 per condition). The table indicates the gene symbol, gene name and fold-changes and *p*-values; *n*-numbers represent biologically independent experiments. For statistical analysis unpaired two-sided Student's *t*-test was applied. Source data are provided under the following link:

<http://www.ebi.ac.uk/arrayexpress/experiments/E-MTAB-8111>.