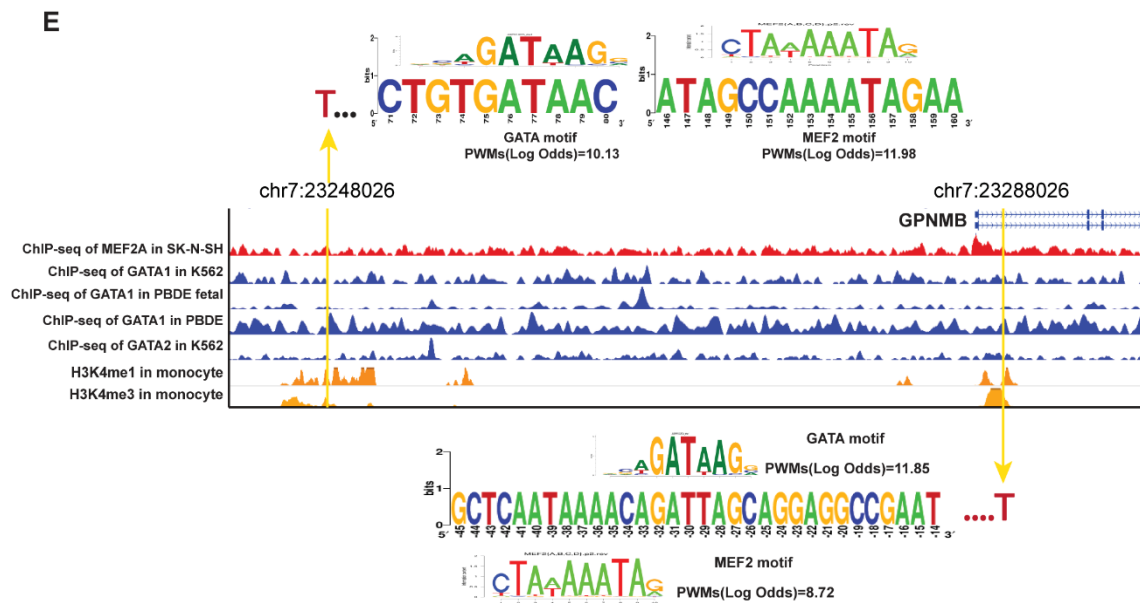
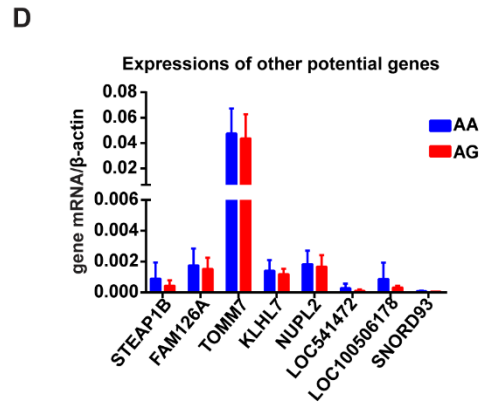
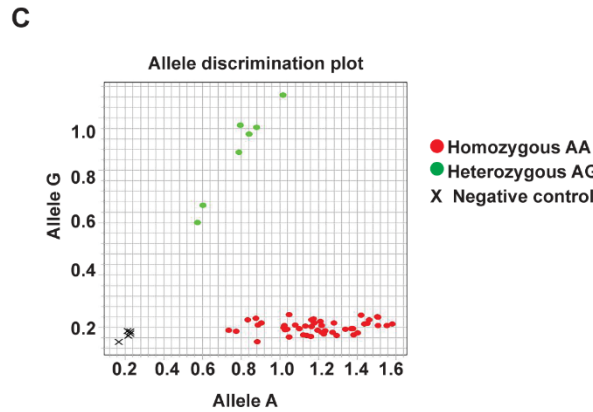
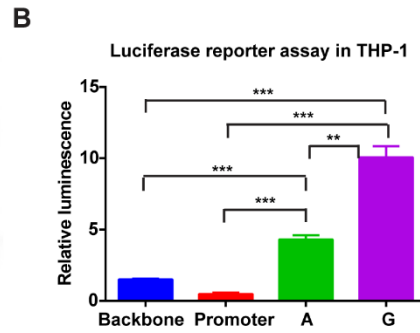
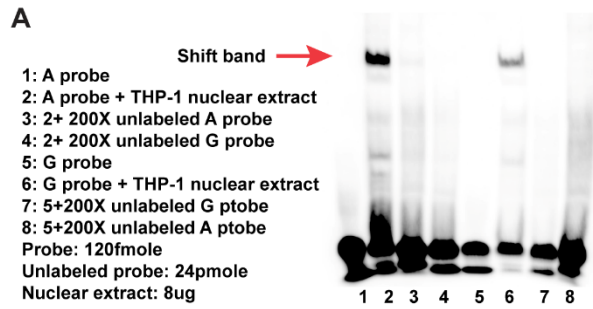


## Supplementary Material

### Supplementary Figure1

**A.** EMSA assay in THP-1 cells demonstrating increased protein binding to the probe with A allele compared to G allele. More DNA-protein complexes were shifted with the probe with allele A (lane 2) than allele G (lane 6); both unlabeled probes with A allele or G allele (24 pmol, 200X) can totally compete out the corresponding shifted band separately (lane 3 and lane 7); using 200X unlabeled probe with G allele can also compete out the shifted band formed with the probe with A allele (lane 4); unlabeled probe with A allele was also able to compete out the shifted band formed in the presence of probe with G allele (lane 8). **B.** The enhancer function of rs2069837 locus was also confirmed in THP-1 cells. Significantly increased luciferase expression was detected in both vectors with intronic sequence (A/G), with significantly higher expression in the construct with G allele than that with A allele ( $p < 0.05$ ). **C.** The allele discrimination plot of 48 healthy subjects genotyped for rs2069837. **D.** No differences were observed between macrophages with AA and AG in the expressions of other genes interacting with rs2069837 based on DNase hypersensitivity correlations maps. **E.** Other epigenetic markers (H3K4me1 and H3K4me3) in monocytes also indicated the locus upstream *GPNMB* and the locus in the first intron of *GPNMB* are regulatory regions. GATA binding motif (72bp downstream of chr7:23248026 and 26bp upstream of chr7: 23288026) and MEF2 binding motif (149bp downstream of chr7:23248026 and 32bp upstream of chr7:23288026) close to these two loci are predicted, which were also demonstrated in ChIP-seq data of some cell lines (no ChIP-seq data of GATA and MEF2 in monocytes were available).



**Supplementary Table 1. Primers used in RT-PCR**

<b>Gene</b>	<b>Forward primer</b>	<b>Reverse primer</b>
<i>Actin</i>	GTCAGGCAGCTCGTAGCTCT	GCCATGTACGTTG- CTATCCA
<i>GPNUMB</i>	CCTCGTGGGCTCAAATATAACAT	ACTGTCCTCTGACCATGCTGT
<i>IL6</i>	ACTCACCTCTTCAGAACGAATTG	CCATCTTTGGAAGGTTTCAGG-TTG
<i>STEAP1B</i>	CCCTTCTACTGGGCACAATACA	GCATGGCAGGAATAGTATGCTTT
<i>FAM126A</i>	GTTGTGGAGGAATGGTTGTCA	GACAGGTTCTAGCAACTCACTTT
<i>NUPL2</i>	GCTTTGGATTGTCTGAGAACCC	CAAGCCTCAATTCCTCTGGTG
<i>KLHL7</i>	GTCAAGCGAGTAACACATCTTCT	AACCTGATCCTCTGCTCTCAC
<i>TOMM7</i>	CATTCGCTGGGGCTTTATCC	TCTGCACCCCTCTTAAATCCC
<i>LOC541472</i>	AAATTACTGAAGCCCCTTGTT	ACTCTGCAAGATGCCACAAGG
<i>LOC100506178</i>	TGCAGAGCTGGTGGTCATAG	TTAGCGTGCACCAGAATGAG
<i>SNORD93</i>	ATCCTGGCCAAGGATGAGAACT	ATCCTGGCCTCAGGTAAATCCT

**Supplementary Table 2. Primers used in 3C assay**

<b>Primer</b>	<b>BsrDI restriction site</b>
P <sub>C</sub> : TTCTCTGATTGTCCCCCTTGA P <sub>R</sub> : CAGCATGTTTTGGGACATTG	chr7:22768898
P1: TTCTGTGGCTGGTTTTGGT P2: TTCCAGAGGTGGAGGCTTTA	chr7:23240228
P3: GGAAAAGGTTGCTTGATTGG P4: CAGGCTGTAGCCCCTAGTGA	chr7:23247996
P5: CTGGCGGTTGGTAAACTGA P6: GCTACTTGGGAGGCTGAGG	chr7:23255204
P7: GTCAGGCTGGTCTCGAACTG P8: AGAGCCTTTTACACCATTCCA	chr7:23254931
P9: TGGTGGTGTGTGCCTGTAGT P10: TTCATGTAATGTAAAGCGAATAATG	chr7:23282317
P11: CAGGAGAATGGCTTGAACCT P12: GACTCACTCCTTCTTGGCATCT	chr7:23286092
P13: GGTGGTGTGTGCCTGTAGT P14: TTCATGTAATGTAAAGCGAATAATG	chr7: 23288603
P15: GGTGAAGTCCCGTCCCTACT P16: GCTCATCAATGTGCCATTCT	chr7: 23297260

Primer pairs next to restriction sites were used to evaluate digestion efficiency (Pc and P<sub>R</sub>, P1 and P2, P3 and P4, P5 and P6, P7 and P8, P9 and P10, P11 and P12, P13 and P14, P15 and P16). The constant primer (Pc) paired with P2, P4, P6, P8, P10, P12, P14, P16 were used in 3C analysis. The following primers in *GAPDH* were used in loading control assessment (F: ACAGTCCATGCCATCACTGCC, R: CCTGCTTCACCACCTTCTTG). Pc: constant primer.

**Supplementary Table 3. Regulatory motifs altered on rs2069837**

<b>Position Weight Matrix ID</b> (Library from <a href="#">Kheradpour and Kellis, 2013</a> )	<b>Strand</b>	<b>Ref</b>	<b>Alt</b>	<b>Match on:</b> <b>Ref:</b> TAAGTATCTACTGTGTGCCAGGCACTTTAAATAAATATTGTGTCTAA TCTTCAAACAA <b>Alt:</b> TAAGTATCTACTGTGTGCCAGGCACTTTAGATAAATATTGTGTCTA ATCTTCAAACAA
<b>Arid3a_2</b>	+	11.6	6.1	NNNNTTRATYAAWHNH
<b>Arid5a</b>	-	11.5	10.5	DNYHBHAATATTRB
<b>Fox</b>	-	16	12.4	WAARYAAAYAWWV
<b>Foxa_known1</b>	-	12.9	10.2	HWRWRYAAAYA
<b>Foxa_known4</b>	-	14.7	10.6	WRARYAAAYAWKNMV
<b>Foxd3</b>	-	12.8	8.5	RAAHMAAYAWWY
<b>Foxf1</b>	-	14.1	10.1	YRHAYAAACAHNB
<b>Foxi1</b>	-	11.6	7.4	WAWRYAAAYAHVH
<b>Foxj1_1</b>	-	11.9	-0	TAAACAAACAHWD
<b>Foxj2_1</b>	-	14.5	12.7	HHNHMRRYAAAYAHHNNW
<b>Foxl1_1</b>	-	14.8	13.8	DHDVHATAAAYAHDDN
<b>HDAC2_disc2</b>	+	11.9	10.4	WRRGYMAAYA
<b>Hlx1</b>	-	11.4	8.1	DRTAWTYAAWTADKD
<b>Mef2_known5</b>	+	6.9	-5	HSTGTTTRCTAWAAATAGAWHMN
<b>Nkx6-2</b>	-	11.8	10.8	RWRDTAAWTABB
<b>TATA_known1</b>	+	12	9.1	NHDWWWTTWWAWWWDRN
<b>TATA_known3</b>	+	10.8	11.3	KATAAATW

Ref: reference allele; Alt: alternative allele; values in the table are log-odds scores of position weight matrices (PWMs). S = C or G, W = A or T, R = A or G, Y = C or T, K = G or T, M = A or C, N = any base pair.

**Supplementary Table 4. Predicted binding proteins in the rs2069837 locus using CIS-BP database**

Name	Motif ID	Family	Sequence	From	To	Direction	Score Allele A	Score Allele G
ARID3	M0106_1.02	ARID/BRIGHT	AAATAAATA	7	15	R	8.04	-
FOXA	M6241_1.02	Forkhead	TAAATAAATA	6	15	F	14.932	13.313
FOXD	M6241_1.02	Forkhead	TAAATAAATA	6	15	F	14.932	13.313
FOXI	M6241_1.02	Forkhead	TAAATAAATA	6	15	F	14.932	13.313
FOXJ	M6241_1.02	Forkhead	TAAATAAATA	6	15	F	14.932	13.313
FOXL	M6241_1.02	Forkhead	TAAATAAATA	6	15	F	14.932	13.313
MEF2	M6340_1.02	MADS box	ACTTTAAATAAATA	2	15	F	10.525	-
GATA	M3314_1.02	GATA	TTTAGATAAATAT	4	16	F	-	8.627

F: forward; "From" and "to" mean the position of the sequence CACTTTAA/GATAAATAT which matched the corresponding motif; The "score" evaluates each position in the sequence with all PWMs, using a standard log odds scoring method (Nat Biotechnol. 2011;29(6):480-3.).



**Supplementary Table 5. Binding proteins observed with mass spectrometry. The list is filtered to include only proteins detected using data included in Supplementary Tables 3 and 4 (See Methods).**

<b>Family/protein</b>	<b>Protein name</b>	<b>Coverage</b>	<b>Unique Peptides</b>	<b>MW [kDa]</b>	<b>PSMs (unlabeled)</b>	<b>PSMs (Mutated)</b>	<b>PSMs (Allele A)</b>	<b>PSMs (Allele G)</b>
<b>ARID</b>	Arid3A	25.29	6	62.9	1	1	7	5
	Arid3B	19.25	8	60.6	1	3	9	5
<b>Fox</b>	FOXJ3	20.25	7	68.9	2	6	10	5
	FOXA1	46.39	9	49.1	0	8	16	12
<b>MEF2</b>	MEF2A	7.49	1	54.8	0	0	<b>3</b>	<b>0</b>
	MEF2C	7.61	1	51.2	0	0	<b>3</b>	<b>0</b>
	MEF2D	8.06	2	55.9	0	0	<b>4</b>	<b>0</b>
<b>GATA</b>	GATA1	26.92	6	42.7	1	4	<b>2</b>	<b>8</b>
	GATA2	27.39	8	50	2	3	<b>2</b>	<b>12</b>
<b>TAF</b>	TAF1	4.64	7	212.5	1	5	1	3
	TAF4	14.74	8	110	2	7	3	4
<b>HDAC</b>	HDAC2	56.35	14	55.3	2	19	19	15
	HDAC5	25.70	6	121.9	0	4	<b>3</b>	<b>0</b>
	HDAC7	61.61	14	102.9	4	20	19	20

PSMs: peptide spectrum matches; MW: molecular weight; ARID: AT-rich interaction domain containing (ARID) family; Fox: forkhead box family; MEF2: myocyte enhancer factor-2; TAF: TATA box binding protein associated factor; HDAC: Histone deacetylase.

**Supplementary Table 6. Binding proteins detected by mass spectrometry that also participate in chromatin looping**

<b>Family/protein</b>	<b>Protein name</b>	<b>Coverage</b>	<b>Unique Peptides</b>	<b>MW [kDa]</b>	<b>PSMs (unlabeled)</b>	<b>PSMs (Mutated)</b>	<b>PSMs (Allele A)</b>	<b>PSMs (Allele G)</b>
<b>CTCF</b>	CTCF	41.67	22	82.7	3	16	12	11
<b>Cohesin complex</b>	SMC1A	62.53	81	143.1	2	66	52	66
	SMC3	61.54	79	141.5	4	63	61	64
	RAD21	63.07	30	71.6	2	20	15	26
	STAG1	14.78	10	144.3	3	7	8	6
	STAG2	39.88	30	141.2	5	23	25	19

PSMs: peptide spectrum matches; MW: molecular weight; CTCF: CCCTC-binding factor.

**Supplementary Table 7. Annotation analysis of the genes interacting with rs2069837 using DNase hypersensitivity data**

<b>Category</b>	<b>Term</b>	<b>Count</b>	<b>%</b>	<b>P-Value</b>	<b>Genes</b>
GOTERM_BP_DIRECT	GO:0045765~regulation of angiogenesis	2	16.6	0.015	<i>IL6, GPNMB</i>
GOTERM_BP_DIRECT	GO:1901215~negative regulation of neuron death	2	16.6	0.018	<i>IL6, GPNMB</i>
GOTERM_BP_DIRECT	GO:0070374~positive regulation of ERK1 and ERK2 cascade	2	16.6	0.080	<i>IL6, GPNMB</i>
GOTERM_MF_DIRECT	GO:0005515~protein binding	8	66.6	0.045	<i>KLHL7, IL6, TOMM7, MALSU1, IGF2BP3, NUPL2, GPNMB, FAM126A</i>

**Supplementary Table 8. Demographic information of the healthy subjects using in this study**

<b>Sample</b>	<b>Gender</b>	<b>Ethnicity</b>	<b>Age</b>	<b>Genotype of rs2069837</b>
1	Female	Hispanic	22	AA
3	Female	Caucasian	22	AA
4	Male	Caucasian	23	AA
5	Female	Caucasian	25	AA
6	Female	Caucasian	52	AA
7	Female	Caucasian	53	AA
2	Female	Caucasian	64	AA
8	Female	Hispanic	20	AG
9	Female	Caucasian	22	AG
10	Male	N/A	22	AG
11	Female	Caucasian	25	AG
12	Female	Caucasian	54	AG
13	Female	Caucasian	56	AG
14	Female	Caucasian	64	AG