

**Supplement to:**

**Differential impact on motility and biofilm dispersal of closely related phosphodiesterases in *Pseudomonas aeruginosa***

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JSW and IT are last authors on this work

**Table S1: Bacterial strains and plasmids**

Strains and plasmids	Genotype or phenotypes <sup>a</sup>	Reference
<b><i>E. coli</i></b>		
S17-1( $\lambda$ pir)	<i>TpR SmR recA, thi, pro, hsdR-M+RP4; 2-Tc:Mu: Km Tn7</i>	Biomedal corp.
DH5 $\alpha$	<i>spuE44 <math>\Delta</math>lacU169(<math>\phi</math>80lacZ<math>\Delta</math>M15) hsdR17 <math>\lambda</math>pir recA1 endA1 gyrA96 thi-1 relA1</i>	Lab collection
<b><i>P. aeruginosa</i></b>		
PAO1	Wild-Type, C.Manoil lab, University of Washington	57
<b>KO mutants</b>		
$\Delta$ pa0285	PAO1 mutant ( <i>pa0285::aacCl</i> ); Gm <sup>R</sup>	This study
$\Delta$ pa0290	PAO1 mutant ( <i>pa0290::aacCl</i> ); Gm <sup>R</sup>	This study
$\Delta$ pa0338	PAO1 mutant ( <i>pa0338::aacCl</i> ); Gm <sup>R</sup>	This study
$\Delta$ pa0575	PAO1 mutant ( <i>pa0575::aacCl</i> ); Gm <sup>R</sup>	This study
$\Delta$ pa0847	PAO1 mutant ( <i>pa0847::aacCl</i> ); Gm <sup>R</sup>	This study
$\Delta$ pa0861 ( $\Delta$ rbdA)	PAO1 mutant ( <i>pa0861::aacCl</i> ); Gm <sup>R</sup>	This study
$\Delta$ pa1181 ( $\Delta$ yegeE)	PAO1 mutant ( <i>pa1181::aacCl</i> ); Gm <sup>R</sup>	This study
$\Delta$ pa2072	PAO1 mutant ( <i>pa2072::aacCl</i> ); Gm <sup>R</sup>	This study
$\Delta$ pa4601 ( $\Delta$ morA)	PAO1 mutant ( <i>pa4601::aacCl</i> ); Gm <sup>R</sup>	This study
$\Delta$ pa4959 ( $\Delta$ fimX)	PAO1 mutant ( <i>pa4959::aacCl</i> ); Gm <sup>R</sup>	This study
$\Delta$ pa5017 ( $\Delta$ dipA)	PAO1 mutant ( <i>pa5017::aacCl</i> ); Gm <sup>R</sup>	This study
$\Delta$ pa5442	PAO1 mutant ( <i>pa5442::aacCl</i> ); Gm <sup>R</sup>	This study
$\Delta$ pa1443 ( $\Delta$ fliM)	PAO1 mutant ( <i>pa4959::aacCl</i> ); Gm <sup>R</sup>	This study
$\Delta$ pa4525 ( $\Delta$ pilA)	PAO1 mutant ( <i>pa5017::aacCl</i> ); Gm <sup>R</sup>	This study
$\Delta$ pa1443 $\Delta$ pa4525 ( $\Delta$ pilA $\Delta$ fliM)	PAO1 mutant ( <i>pa1443::aacCl,pa4525::Neo</i> ); Gm <sup>R</sup> ; Km <sup>R</sup>	This study
<b>KO mutants with c-di-GMP reporter gfp vector</b>		
PAO1 Tn7CdrA::gfp	PAO1 WT with mini-Tn7-P <sub>cdrA</sub> -RBSII-gfp(Mut3)-T <sub>0</sub> -T <sub>1</sub> ;Tet <sup>R</sup>	This study
$\Delta$ pa0285 Tn7CdrA::gfp	PAO1 mutant ( <i>pa0285::aacCl</i> ) with mini-Tn7-P <sub>cdrA</sub> -RBSII-gfp(Mut3)-T <sub>0</sub> -T <sub>1</sub> ;Gm <sup>R</sup> ;Tet <sup>R</sup>	This study
$\Delta$ pa0290 Tn7CdrA::gfp	PAO1 mutant ( <i>pa0290::aacCl</i> ) with mini-Tn7-P <sub>cdrA</sub> -RBSII-gfp(Mut3)-T <sub>0</sub> -T <sub>1</sub> ;Gm <sup>R</sup> ;Tet <sup>R</sup>	This study
$\Delta$ pa0338 Tn7CdrA::gfp	PAO1 mutant ( <i>pa0338::aacCl</i> ) with mini-Tn7-P <sub>cdrA</sub> -RBSII-gfp(Mut3)-T <sub>0</sub> -T <sub>1</sub> ;Gm <sup>R</sup> ;Tet <sup>R</sup>	This study
$\Delta$ pa0575 Tn7CdrA::gfp	PAO1 mutant ( <i>pa0575::aacCl</i> ) with mini-Tn7-P <sub>cdrA</sub> -RBSII-gfp(Mut3)-T <sub>0</sub> -T <sub>1</sub> ;Gm <sup>R</sup> ;Tet <sup>R</sup>	This study
$\Delta$ pa0847 Tn7CdrA::gfp	PAO1 mutant ( <i>pa0847::aacCl</i> ) with mini-Tn7-P <sub>cdrA</sub> -RBSII-gfp(Mut3)-T <sub>0</sub> -T <sub>1</sub> ;Gm <sup>R</sup> ;Tet <sup>R</sup>	This study
$\Delta$ pa0861 Tn7CdrA::gfp	PAO1 mutant ( <i>pa0861::aacCl</i> ) with mini-Tn7-P <sub>cdrA</sub> -RBSII-gfp(Mut3)-T <sub>0</sub> -T <sub>1</sub> ;Gm <sup>R</sup> ;Tet <sup>R</sup>	This study
$\Delta$ pa1181 Tn7CdrA::gfp	PAO1 mutant ( <i>pa1181::aacCl</i> ) with mini-Tn7-P <sub>cdrA</sub> -RBSII-gfp(Mut3)-T <sub>0</sub> -T <sub>1</sub> ;Gm <sup>R</sup> ;Tet <sup>R</sup>	This study
$\Delta$ pa2072 Tn7CdrA::gfp	PAO1 mutant ( <i>pa2072::aacCl</i> ) with mini-Tn7-P <sub>cdrA</sub> -RBSII-gfp(Mut3)-T <sub>0</sub> -T <sub>1</sub> ;Gm <sup>R</sup> ;Tet <sup>R</sup>	This study
$\Delta$ pa4601 Tn7CdrA::gfp	PAO1 mutant ( <i>pa4601::aacCl</i> ) with mini-Tn7-P <sub>cdrA</sub> -RBSII-gfp(Mut3)-T <sub>0</sub> -T <sub>1</sub> ;Gm <sup>R</sup> ;Tet <sup>R</sup>	This study
$\Delta$ pa4959 Tn7CdrA::gfp	PAO1 mutant ( <i>pa4959::aacCl</i> ) with mini-Tn7-P <sub>cdrA</sub> -RBSII-gfp(Mut3)-T <sub>0</sub> -T <sub>1</sub> ;Gm <sup>R</sup> ;Tet <sup>R</sup>	This study
$\Delta$ pa5017 Tn7CdrA::gfp	PAO1 mutant ( <i>pa5017::aacCl</i> ) with mini-Tn7-P <sub>cdrA</sub> -RBSII-gfp(Mut3)-T <sub>0</sub> -T <sub>1</sub> ;Gm <sup>R</sup> ;Tet <sup>R</sup>	This study
$\Delta$ pa5442 Tn7CdrA::gfp	PAO1 mutant ( <i>pa5442::aacCl</i> ) with mini-Tn7-P <sub>cdrA</sub> -RBSII-gfp(Mut3)-T <sub>0</sub> -T <sub>1</sub> ;Gm <sup>R</sup> ;Tet <sup>R</sup>	This study
<b>Plasmids</b>		
pPS856	Ap <sup>R</sup> , Gm <sup>R</sup> ; 0.83-kb blunt-ended SacI fragment from pUCGM	37
pEX100T	Ap <sup>R</sup> ; oriT <sup>+</sup> sacB <sup>+</sup> gene replacement vector	37
pCdrA::gfp	pUCP22Not-P <sub>cdrA</sub> -RBSII-gfp(Mut3)-T <sub>0</sub> -T <sub>1</sub> , Amp <sup>R</sup> Tet <sup>R</sup>	41
pCR <sup>TM</sup> 4-TOPO <sup>TM</sup>	Km <sup>R</sup> and Amp <sup>R</sup> TOPO cloning expression vector	Invitrogen <sup>TM</sup>

**Table S2: Primer sequences**

Primer	Sequence (5'→3')
<b>Used for KO mutant generation and sequencing</b>	
<i>Δpa0285</i>	
PA0285-up-F	ATG <u>CCCCGGG</u> CCTGACCGACAATCTCGAC
PA0285-up-R	ATGCG <u>AATTC</u> GTTCCAGATCTCTCCGCTC
PA0285-dn-F	ATGCA <u>AGCTT</u> AGTCACCGAAAGCGCGG
PA0285-dn-R	ATG <u>CCCCGGG</u> TCAGTCTTCCGGCAGCG
SeqPA0285-F	GTAAGTCCACCAGTTGC
SeqPA0285-R	CCATCCAGGCTTCTGAG
<i>Δpa0290</i>	
PA0290-up-F	ATG <u>CCCCGGG</u> GCGAATGGTCTACGTCAGC
PA0290-up-R	ATGCG <u>AATTC</u> CAGGAACGCCAACGGTATC
PA0290-dn-F	ATGCA <u>AGCTT</u> GCGACCTGTTCGGGC
PA0290-dn-R	ATG <u>CCCCGGG</u> TCAGCCCACGACGATG
SeqPA0290-F	GTATCGCTCGCAACTG
SeqPA0290-R	CCTTCTGGCGAGTTACC
<i>Δpa0338</i>	
PA0338-up-F	ATG <u>CCCCGGG</u> GTGCGGCGCTTACGC
PA0338-up-R	ATGCG <u>AATTC</u> CTCGGGATGGATGATGC
PA0338-dn-F	ATGCA <u>AGCTT</u> CCTGTTCGGCCACTCC
PA0338-dn-R	ATG <u>CCCCGGG</u> TCAGCAACAGGCCACG
SeqPA0338-F	CAAGGTGAAGTGGCTGC
SeqPA0338-R	CTCCAGCGTGTTCATCG
<i>Δpa0575</i>	
PA0575-up-F	ATG <u>CCCCGGG</u> GATCATCATCCTGGCACG
PA0575-up-R	ATGCG <u>AATTC</u> GGACGATGGGAAGACCG
PA0575-dn-F	ATGCA <u>AGCTT</u> GCCGAGGATTGCGG
PA0575-dn-R	ATG <u>CCCCGGG</u> CACCGTGAGCTGCATG
SeqPA0575-F	GACGAGTGACGAACGAAC
SeqPA0575-R	GTTGCCATCAGATAAGGAC
<i>Δpa0847</i>	
PA0847-up-F	ATG <u>CCCCGGG</u> CCGTCGCCGAGGAC
PA0847-up-R	ATGCG <u>AATTC</u> GTCGTAGCTGGCATCCC
PA0847-dn-F	ATGCA <u>AGCTT</u> CGCGCCATGCAGAG
PA0847-dn-R	ATG <u>CCCCGGG</u> CGCCAGGCGGTAG
SeqPA0847-F	GGAACTTATCCATGCGGTTG
SeqPA0847-R	GTGGCTGAACTGCTGCATC
<i>Δpa0861</i>	
PA0861-up-F	ATG <u>CCCCGGG</u> ATGAGGCAGAACCGGACTC
PA0861-up-R	ATGCG <u>AATTC</u> AGCTCGATGGCGCG
PA0861-dn-F	ATGCA <u>AGCTT</u> GGTTGCCAACCTGGCC
PA0861-dn-R	ATG <u>CCCCGGG</u> CTACCGGAGGTTCTGTCCC
SeqPA0861-F	CAGCCACTAGACTCCTACTG
SeqPA0861-R	CAGTCGACGATCAGTTGC
<i>Δpa1181</i>	
PA1181-up-F	ATG <u>CCCCGGG</u> TTCGCGTGCTCTGGG
PA1181-up-R	ATGCG <u>AATTC</u> AGGCCATCGGCAGGTAC
PA1181-dn-F	ATGCA <u>AGCTT</u> CTTTCGAGCTGACCGAGACG
PA1181-dn-R	ATG <u>CCCCGGG</u> TCAGCCCAACTCCTGGC
SeqPA1181-F	CTGAGCCTGGACGATTC
SeqPA1181-R	CTGCACATGAGTTTCGACG

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*Δpa2072*  
PA2072-up-F ATGCCCCGGGATGCTTGCTTCAGCGATTG  
PA2072-up-R ATGCGAATTCTCGACGAAGACCATCACC  
PA2072-dn-F ATGCAAGCTTAGGCGAGGTGATGGTCTC  
PA2072-dn-R ATGCCCCGGGTGAGCAGGAACCCCTTGC  
SeqPA2072-F ATGAAGGTCGAGGTGGTG  
SeqPA2072-R TGACGCCTCGTTGTTATTGC

*Δpa4601*  
PA4601-up-F ATGCCCCGGGTTGCGGACTCTACGACAG  
PA4601-up-R ATGCGAATTCAACTCGCTACGGGTCTCG  
PA4601-dn-F ATGCAAGCTTATCCTGATGAGTGACGTCCG  
PA4601-dn-R ATGCCCCGGGTCAGCCCTCGTTGAACATG  
SeqPA4601-F GACAAAAGTGCCTCGAC  
SeqPA4601-R GTCAAATCACGGCTGAAC

*Δpa4959*  
PA4959-up-F ATGCCCCGGGCACGTACGACGGCGAAC  
PA4959-up-R ATGCGAATTCGTTCTGACTTTCCTCAGCAG  
PA4959-dn-F ATGCAAGCTTGAGGTGCTCCTGCGC  
PA4959-dn-R ATGCCCCGGGGGTCTGGACGAAGGAG  
SeqPA4959-F GTTCGAATACCGCTCG  
SeqPA4959-R TGGACGGCATCCTCTAC

*Δpa5017*  
PA5017-up-F ATGCCCCGGGCGACTACCCTCTGCGC  
PA5017-up-R ATGCGAATTCGTGCCGTAGGCTTCGC  
PA5017-dn-F ATGCAAGCTTGCCGATCGGCAAGTG  
PA5017-dn-R ATGCCCCGGGCCATGGCGATCACCG  
SeqPA5017-F CAATCGGCATAATGCACC  
SeqPA5017-R CAGAACCATGGCTCTCG

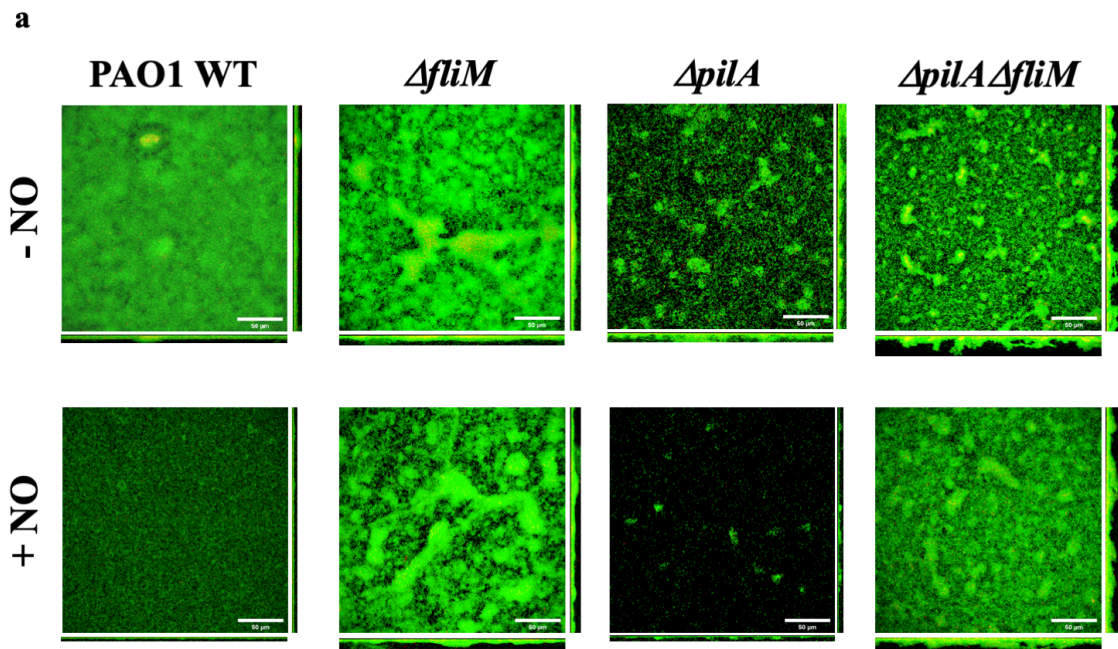
*Δpa5442*  
PA5442-up-F ATGCCCCGGGATGACCGTCCATGTTCGAG  
PA5442-up-R ATGCGAATTCCGAACACCAGCACCTGC  
PA5442-dn-F ATGCAAGCTTCGGCGCAACTGGAGC  
PA5442-dn-R ATGCCCCGGGTCAACGAGCCTGGCG  
SeqPA5442-F GTCGTTGAGGGAGTTTCAG  
SeqPA5442-R GACCAATCGGGATAGCAG

*Δpa1443*  
PA1443-up-F ATGCCCCGGGAGCAAGCTGAAGCTGATC  
PA1443-up-R ATGCGAATTCTTTCATCCTGGGAAAGCAG  
PA1443-dn-F ATGCAAGCTTTACAGATTCTCGAAGCGG  
PA1443-dn-R ATGCCCCGGGAGCTTCTTGATGCGTTTCG  
SeqPA1443-F AGCAAGCTGAAGCTGATC  
SeqPA1443-R AGCTTCTTGATGCGTTTCG

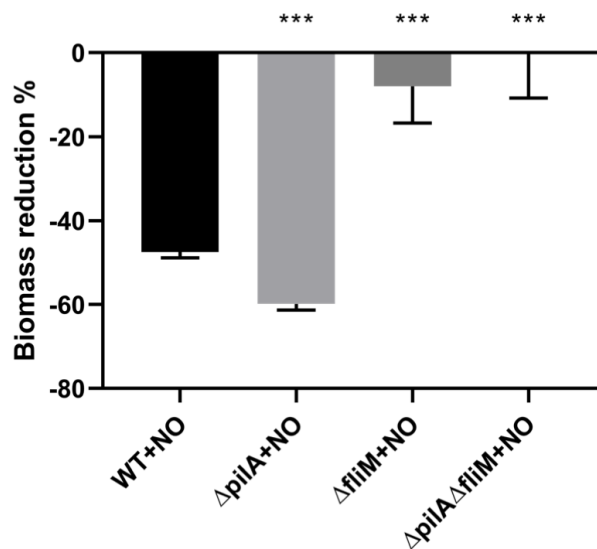
*Δpa4525*  
PA4525-up-F ATGCCCCGGGTTGTCCAGGCTATTCAGG  
PA4525-up-R ATGCGAATTCACCACGATCATCAGTTTCGATC  
PA4525-dn-F ATGCAAGCTTTACCTTTACCTTCAGACTG  
PA4525-dn-R ATGCCCCGGGTGACCAAGGATGTCAGG  
SeqPA4525-F TTGTCCAGGCTATTCAGG  
SeqPA4525-R TGACCAAGGATGTCAGG

Gentamicin resistance and kanamycin resistance gene cassettes  
Gm-F ATGCGAATTCGAATTGACATAAGCCTGTTCCGG  
Gm-R ATGCAAGCTTGAATTGGCCGCGG  
Km-F ATGCGAATTCAATGTCAGCTACTGGGCTATC  
Km-R ATGCAAGCTTCTGATGCGGTGTGAAATAC

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**b** **NO-induced biofilm dispersal**



**Figure S1. NO-induced biofilm dispersal of flagellum and pili deletion mutants. (A)** Confocal laser scanning microscopy images used for phenotypic analysis of 72 hrs mature biofilms before or after 2 hrs treatment with 250  $\mu$ M SNP for the flagellum mutant  $\Delta$ *fliM*, the pili mutant  $\Delta$ *pilA* and the double mutant  $\Delta$ *pilA* $\Delta$ *fliM* compared with PAO1 WT (scale bar 50  $\mu$ m). **(B)** COMSTAT analysis of biomass reduction after treatment normalised against WT, highlighting differences in biomass reduction for the mutants. The Student's T-test was used to determine significances, where \*\*\* denotes a confidence level of  $p < 0.01$ , and \*\* denotes a confidence level of  $0.01 < p < 0.05$ . Data acquired from 3 independent experiments.