

Supplemental Material

Online Methods

Drug-hERG interaction function scale model

The wild-type drug-free hERG Markov model (Online Table I) previously described in ⁷⁷ is shown in main text Figure 3B. To simulate drug interactions with hERG, we used **simulated affinities**, (i.e. drug dissociation constants) K_{D_0} , and drug diffusion rates, D , both computed from the umbrella sampling (US) molecular dynamics (MD) simulations (Main text Figure 2) used to constrain the drug “on” (k_{o_d} , k_{od} , and k_{od_zw}) and “off” (r_{o_d} , r_{od} , and r_{od_zw}) model transition rates for open state (Online Table II). Based on available literature data ⁴⁴, K_{DI} was assumed to be 70-fold less than K_{D_0} in the dofetilide model. Then, using the relation, $k_{off} = k_{on} * K_{D_0}$, we optimized k_{i_d} and k_{id} for open-inactivated state of neutral and cationic forms of dofetilide (Online Table III).

There are two modes of drug bound channel states – neutral (cyan) and cationic (red) for dofetilide (Main text Figure 3B). The cationic and neutral drug fractions, f_1 and f_0 , are calculated using the following equations:

$$f_1 = \frac{1}{(1+10^{(pH-pK_a)})}; \quad f_0 = 1 - f_1 \quad [1]$$

Where $pH = 7.2$ and $pK_a = 7.0$ ³⁹

For moxifloxacin, there are three modes of drug bound channel states – neutral (cyan), cationic (red), and zwitterionic (purple) as shown in Main text Figure 3D. The drug fractions at $pH = 7.2$ based on ⁴⁰ are shown in Online Table IV.

Online Table I: Transition rates in the I_{Kr} model

Transition rates (ms^{-1})

Drug free Kr channel

C3→C2

$$ae = \frac{T}{T_{base}} e^{(24.335 + \frac{T_{base}}{T}(0.0112 \times V - 25.914))}$$

C2→C3

$$be = \frac{T}{T_{base}} e^{(13.688 + \frac{T_{base}}{T}(-0.0603 \times V - 15.707))}$$

C2→C1

$$ain = \frac{T}{T_{base}} e^{(22.746 + \frac{T_{base}}{T}(-25.914))}$$

C1→C2

$$bin = \frac{T}{T_{base}} e^{(13.193 + \frac{T_{base}}{T}(-15.707))}$$

C1→O

$$aa = \frac{T}{T_{base}} e^{(22.098 + \frac{T_{base}}{T}(0.0365 \times V - 25.914))}$$

O→C1

$$bb = \frac{T}{T_{base}} e^{(7.313 + \frac{T_{base}}{T}(-0.0399 \times V - 15.707))}$$

O→I

$$\beta i = \frac{T}{T_{base}} e^{(30.016 + \frac{T_{base}}{T}(0.0223 \times V - 30.88))} \times \left(\frac{5.4}{[K]^o}\right)^{0.4}$$

I→O

$$\alpha i = \frac{T}{T_{base}} e^{(30.061 + \frac{T_{base}}{T}(-0.0312 \times V - 33.243))}$$

Online Table II: Transition rates for Moxifloxacin model (see main text Figures 2E and 3D)

| | Open |
|---|--|
| Transition rates | <i>Neutral Drug</i> |
| k_{on} ($k_{\text{o_d}}$) | 6.6E+02 ($\mu\text{M}^{-1}\text{s}^{-1}$) * [drug] |
| k_{off} ($r_{\text{o_d}}$) | 4.9E+02 (s^{-1}) |
| | <i>Cationic Drug</i> |
| k_{on} (k_{od}) | 4.2E+02 ($\mu\text{M}^{-1}\text{s}^{-1}$) * [drug] |
| k_{off} (r_{od}) | 2.8E+06 (s^{-1}) |
| | <i>Zwitterionic Drug</i> |
| k_{on} ($k_{\text{od_zw}}$) | 3.4E+02 ($\mu\text{M}^{-1}\text{s}^{-1}$) * [drug] |
| k_{off} ($r_{\text{od_zw}}$) | 2.9E+06 (s^{-1}) |

Online Table III: Transition rates for Dofetilide model (see main text Figures 2E and 3B)

| | Open | Inactivated |
|---------------------------------------|--|--|
| Transition rates | <i>Neutral Drug</i> | |
| k_{on} (k_{o_d} , k_{i_d}) | 6.7E+02 ($\mu\text{M}^{-1}\text{s}^{-1}$) * [drug] | 1.7E+03 ($\mu\text{M}^{-1}\text{s}^{-1}$) * [drug] |
| k_{off} (r_{o_d} , r_{i_d}) | 1.1E+02 (s^{-1}) | 3.9 (s^{-1}) |
| | <i>Cationic Drug</i> | |
| k_{on} (k_{od} , k_{id}) | 5.3E+02 ($\mu\text{M}^{-1}\text{s}^{-1}$) * [drug] | 1.5E+06 ($\mu\text{M}^{-1}\text{s}^{-1}$) * [drug] |
| k_{off} (r_{od} , r_{id}) | 3.5E+04 (s^{-1}) | 1.4E+06 (s^{-1}) |

Online Table IV: Drug fraction for cationic, zwitterionic and neutral states in moxifloxacin model (Main text Figures 2E and 3D)

| States | Fractions |
|--------------------------------------|-----------|
| Moxifloxacin (\pm), zwitterionic | 83.99% |
| Moxifloxacin (+), cationic | 10.33% |
| Moxifloxacin (0), neutral | 5.68% |

Computed dofetilide concentrations

We used the population C_{max} (maximum plasma concentration) of dofetilide: 2.72 ng/mL⁵⁶, and converted it to nanomolar (nM) concentration ($\frac{2.72 \text{ ng/ml}}{441.567 \text{ g/mol}} \times 1000 \cong 6.16 \text{ nM}$) in the models, where 441.567 g/mol is dofetilide molar mass.

Computed moxifloxacin concentrations

We used the population C_{max} (maximum plasma concentration) of moxifloxacin: 2.5 mg/L⁵⁸, and converted it to micromolar (μM) concentration ($\frac{2.5 \mu\text{g/l}}{401.431 \text{ g/mol}} \times 1000 \cong 6.23 \mu\text{M}$) in the models, where 401.431 g/mol is moxifloxacin molar mass.

Simulation of TRIaD in dofetilide and control case in O'Hara-Rudy Human model

First, action potential duration (APD) *Triangulation* was calculated as the repolarization time from APD₃₀ to APD₉₀ from 1000 simulated cells with application of noise current. The noise current was calculated using the equation from ⁶⁷,

$$V_{t+\Delta t} = V_t - \frac{I(V_t)\Delta t}{C_m} + \xi n \sqrt{\Delta t} \quad [2]$$

where n is a random number between 0 and 1 from a Gaussian distribution, and Δt is the time step. ξ ($= 0.3$) is the diffusion coefficient, which defines the amplitude of noise ⁶⁷. The noise current was generated and applied to the membrane potential V_t throughout the simulated time course. *Reverse-use-dependence* was measured as APD₉₀ at steady state for each pacing cycle length (from 2 Hz to 0.5 Hz), and APD adaptation curves were constructed. *Beat-to-beat (bTb) Instability* was simulated by applying small amplitude inward currents randomly between -0.1 to -0.2 pA/pF for 50 ms over the course of the action potential plateau between 10 to 700 ms at 1 Hz for 1000 beats.

Fiber simulations

We simulated a transmural fiber composed of 165 O'Hara-Rudy human ventricular cells ⁵⁷ ($\Delta x = \Delta y = 100 \mu\text{m}$) connected by resistances to simulate gap junctions ⁹³. The fiber contains an endocardial region (cells 1 to 80) and epicardial region (cells 81 to 165), which showed a linear decrease in APDs ^{70, 71}. G_{Kr} was monotonically increased from 0.04 to 0.05. The heart was paced at 1 Hz to match the clinically observed QT intervals ~ 400 ms ⁹⁴⁻⁹⁶. AP simulations were carried out in epi/endocardial cells by changing various ion channel conductances ⁵⁷. The stimulus is applied to the first cell.

The fiber was paced at varying cycle length from 800 to 1400 ms for 200 beats (mean heart rates = 61 bpm) in order to match the clinical data (56.8 ± 6.4 bpm) ⁵⁶. Pseudo ECGs were computed from the transmembrane potential V_m using the integral

expression as in Gima and Rudy ⁹⁷. Heart rate corrected QT (QTc) was computed using Fridericia formula using the cubic root of RR interval ⁹⁸.

$$QT_c = \frac{QT}{\sqrt[3]{RR}} \quad [3]$$

Spatial APD dispersion was measured using the T-wave area indicator, which was calculated as the T-wave amplitude on the computed pseudo-ECGs. For this purpose, a 1-dimensional model of the transmural wedge preparation, as described in ⁸², was stimulated by applying a standard short-long protocol as follows: The transmural wedge preparation was stimulated by a train of pulses (S1) at 1 Hz pacing rate until the steady-state was reached followed by a premature beat (S1-S2 interval = 800 ms) and then a delayed beat (S3) was delivered after a long pause (S2-S3 interval = 5000 ms). T-wave area calculations were computed as follows:

$$\sum_{t=t_1}^{t_2} |ecg(mV)| \cdot \Delta t \quad [4]$$

where $\Delta t = 1$ ms, t_1 is the time where ECG equals to $T_{peak} - 0.9*(T_{peak} = \text{minimum of left side of T wave})$, and t_2 is the time where ECG equals to $T_{peak} - 0.9*(T_{peak} = \text{minimum of right side of T wave})$.

Frequency-dependent QT prolongation

The fiber was paced at 1Hz for 1000 beats (S1) and then a second stimulus (S2) was applied after a varying RR interval (between 550 and 1200 ms). The QT interval, in response to S2, was recorded. The same simulations were carried out 6 times for both control, 2.5 mg/L moxifloxacin, and 2.72 ng/mL dofetilide with noise currents, and the relative changes in slope of relationship of QT and preceding RR intervals were calculated.

Two-dimensional simulations

2D simulations were performed to determine if proarrhythmic phenomena observed in lower dimensions cause reentrant arrhythmias. Current flow is described by the following equation:

$$\frac{\partial V(x,y,t)}{\partial t} = D_x \frac{\partial^2 V(x,y,t)}{\partial x^2} + D_y \frac{\partial^2 V(x,y,t)}{\partial y^2} - \frac{I_{ion} - I_{stim}}{C_m} \quad [5]$$

where V is the membrane potential, x and y are distances in the longitudinal and transverse directions, respectively, D_x and D_y are diffusion coefficients in the x and y directions. We simulated a *heterogeneous* and a *homogenous* cardiac tissue on a 500 by 500 pixel grid with $\Delta x = \Delta y = 100 \mu\text{m}$. The heterogeneous tissue contains an endocardial region (fibers 1 to 180) and epicardial region (fibers 181 to 500). We also incorporated anisotropic effects by setting D_x and D_y such that the ratio of conduction velocities is 1:2⁹⁹. A typical S1-S2 protocol was used for Main Text Figure 7. The tissue was first paced (S1) in a $0.5 \text{ cm} \times 1.1 \text{ cm}$ area on the left edge of the endocardial region, and a premature stimulus (S2) was then applied in a $1.8 \text{ cm} \times 1.5 \text{ cm}$ area on the top left corner of the endocardial region. Small amplitude inward currents were randomly applied between -0.1 to -0.45 pA/pF on *each cell* in both *heterogeneous* and *homogenous* tissues after 0.5 ms.

Action potential duration (APD) mapping

We reconstructed the “human transmural myocardial wedge” based on data describing transmural action potential heterogeneity mapped from normal human left ventricle⁷¹ (Main Text Figure 8). First, the O’Hara-Rudy human model⁵⁷ was used to generate a G_{Kr} lookup table corresponding to APD₈₀. Next, experimental 2D APD₈₀ map (100×100 – Main Text Figure 8) was used to create a 2D G_{Kr} map using the G_{Kr} lookup table. Then the two-dimensional G_{Kr} values (100×100) were used to simulate APD₈₀ at pacing rate of 0.5 Hz. We then constructed **3D wedge** of 100 by 100 by 1 cells with $\Delta x = \Delta y = 200 \mu\text{m}$ and $\Delta z = 500 \mu\text{m}$ using this APD mapping data. Current flow is described by the following equation:

$$\frac{\partial V(x,y,z,t)}{\partial t} = D_x \frac{\partial^2 V(x,y,z,t)}{\partial x^2} + D_y \frac{\partial^2 V(x,y,z,t)}{\partial y^2} + D_z \frac{\partial^2 V(x,y,z,t)}{\partial z^2} - \frac{I_{ion} - I_{stim}}{C_m} \quad [6]$$

Where V is the membrane potential. D_x , D_y and D_z are diffusion coefficients in the x , y and z directions. Stimulus current I_{stim} is 150 mA/cm² for 0.5 ms. We also incorporated anisotropic effects by setting D_x , D_y and D_z such that the ratio of conduction velocities is 2:4:1 ⁹⁹.

Local sensitivity analysis

We calculated elasticity coefficients (sensitivities) for arrhythmia vulnerability parameters from the TRIaD based simulations. The protocol for arrhythmia vulnerability parameters from the TRIaD based simulations is the same as in Online Figure II in the presence of 2.72 ng/mL dofetilide and in Online Figure III with 2.5 mg/L moxifloxacin. The relative change in each arrhythmia vulnerability parameter in response to a parameter perturbation (local sensitivity) was calculated by following equation:

$$x\text{-elasticity of } y: \varepsilon = \frac{\partial \ln y}{\partial \ln x} \cong \frac{\ln(y_2) - \ln(y_1)}{\ln(x_2) - \ln(x_1)} = \frac{\ln\left(\frac{y_2}{y_1}\right)}{\ln\left(\frac{x_2}{x_1}\right)} = \frac{\ln\left(\frac{\text{outputs at+20\%}}{\text{outputs at-20\%}}\right)}{\ln\left(\frac{\text{rate at+20\%}}{\text{rate at-20\%}}\right)} \quad [7]$$

In each case, the model rate constants were increased and decreased by 20%. If $|\varepsilon| > 1$ (sensitive), y (model output) changes more than changes in x (the model rate). In contrast, $|\varepsilon| < 1$ (insensitive) indicates that y (model output) changes less than changes in x (the model rate) ^{100, 101}.

Simulated data of the arrhythmia vulnerability parameters from TRIaD for random forest machine learning application

1) *APD TRI*: APD triangulations were calculated from APD₃₀ to APD₉₀ from 1000 simulated cells with noise currents. The protocol is same as in **Simulation of TRId**. 2) *bTb instability*: 1000 simulated APDs₉₀ were recorded by adding noise currents into membrane potential calculations. The protocol is same as in **Simulation of TRId**. 3) *RUD*: APDs₉₀ were recorded from 1000 cells at slow pacing rate (BCL = 2000 ms) with noise currents. 4) *T-wave area*: The transmural fibers were stimulated by a standard short-long protocol, and T-wave areas were calculated (as described in **Fiber simulations**) from 1000 cases with noise currents.

Random forest machine learning algorithm

We applied a multivariate correlation-based filter selection (CFS) technique using a random forest machine learning algorithm for multiclass classification ¹⁰² to indicate the importance of each arrhythmia vulnerability parameter from **TRId** (*APD TRI*, *bTb instability*, *T-wave area and RUD*) towards the target (classification of control, different moxifloxacin doses and different dofetilide doses) (Online Figure IV). We calculated the existing correlation coefficients using Pearson's correlation coefficient Eq. [8] to explore the linear dependence of arrhythmia vulnerability parameters from **TRId**. Pearson's coefficient values vary between -1 and 1, where 1 is highly correlated (changes in x_1 are correlated with changes in x_2) and -1 is highly anticorrelated (changes in x_1 are negatively correlated with changes in x_2) ¹⁰³.

$$\rho(\vec{x}_1, \vec{x}_2) = \frac{cov(\vec{x}_1, \vec{x}_2)}{\sigma_{x_1} \sigma_{x_2}} \quad [8]$$

where \vec{x}_i is a vector consisting of each **TRId** parameter's observations, *cov* is covariance between two **TRId** parameters and σ is standard deviation of each **TRId** parameter observed in the simulations.

Random forest is an ensemble decision tree, which contains a collection of single decision trees. Each tree is built over random extraction of the observations from the dataset and the random extraction of the features (arrhythmia vulnerability parameters

in this case)¹⁰⁴. For each feature a series of questions are formulated so that the answer to those questions lead to the best possible separation of classes into groups that contain only one class or the majority of one class at each node. Therefore, the importance of each feature is indicated by purity in each branched out node, and impurity within a node for a particular feature indicates reduced importance in Eq. [10]. The importance of each feature is averaged across all the trees in random forest classifier to determine the final importance of the features¹⁰⁵ in Eq. [11].

$$n_{i_j} = w_j E_j - w_{L_j} E_{L_j} - w_{R_j} E_{R_j} \quad [9]$$

$$norm(f_{i_i}) = \frac{\Sigma_j n_{i_j} / \Sigma_k n_{i_k}}{\Sigma_m f_{i_m}} \quad [10]$$

$$RF(f_{i_n}) = \frac{\sum_n norm(f_{i_n})}{T} \quad [11]$$

Where n_{i_j} is the importance of node j splits on feature i , w_j is the weighted number of samples in node j , E_j is the impurity value of node j , L and R represents child node from left and right split on node j , f_{i_i} is the importance of feature i , k and m belong to nodes and features sets, $RF(f_{i_n})$ is the importance of feature i calculated from all trees in the RF model, $norm(f_{i_n})$ is the normalized feature importance i in tree n , and T is total number of trees.

In order to train the random forest classifier, we first performed feature scaling using Eq. [12] to assure that each feature in the dataset has zero-mean and unit-variance¹⁰⁶

$$\frac{\vec{x}_i - \bar{x}_i}{\sigma_{x_i}} \quad [12]$$

After feature scaling, we split our dataset into random training and testing subsets (using 80% of data for training and 20% of data for testing the performance of the trained classifier). Then we trained the random forest multiclass classifier using the training subset and set 20 trees and entropy at the same time as the measure of

impurity for the random forest classifier. Entropy (E) is a measure that controls how the decision tree decides which feature to choose first and where to split the data (Eq. [13]).

$$E = \sum_i -P_i \log_2(P_i) \quad [13]$$

Where the P_i is the fraction of examples in class i . After training the classifier, we used test dataset to evaluate the performance of the classifier, where classifier will classify test data; which has not seen them before, into specified classes and reported the percentage of correct responses as accuracy of the classifier.

Atomistic structural modeling

hERG open-state atomistic model generation

The 3D coordinates of hERG (PDB: 5VA2) obtained via cryogenic electron microscopy (cryo-EM) were used as a template⁸¹ for our wild-type open state model of voltage sensing and pore domains (residues 405-668). This structure is mostly complete except for some missing extracellular loops; namely, residues 434-451 (between helices S1 and S2), 512-519 (between helices S3 and S4), and 578-582, 598-602 (in the outer vestibule pore loop region) are unresolved. ROSETTA symmetry methods¹⁰⁷, and *de novo* loop modeling protocols¹⁰⁷⁻¹⁰⁹ were used to generate the missing loop regions.

General molecular dynamics (MD) simulation protocols

The CHARMM-GUI online toolkit¹¹⁰, CHARMM^{111, 112}, NAMD¹¹³, and Anton 2¹¹⁴ software programs were used to build and simulate the molecular systems in this study. In all atomistic simulations in this study all-atom biomolecular CHARMM force fields were used including CHARMM36 protein¹¹⁵ and lipid models,¹¹⁶ standard ion parameters¹¹⁷ and TIP3P water model.¹¹⁸ Dofetilide and moxifloxacin force field models were optimized based on CHARMM general force field (CGENFF) parameters¹¹⁹ as described below.

hERG – dofetilide and moxifloxacin as well as dofetilide – membrane simulations contained 1-palmitoyl-2-oleylphosphatidylcholine (POPC) lipid bilayer hydrated by a 0.15 M aqueous KCl solution. Moxifloxacin – membrane simulations contained a 1,2-dimyristoylphosphatidylcholine (DMPC) lipid bilayer hydrated by a 0.15 M aqueous KCl solution. The membrane normal axis was aligned along the z-axis in all cases. The hERG channel was placed in the bilayer center with its aqueous pore aligned with the membrane normal. All membrane – drug and hERG – drug MD simulations were carried out in an *NPT* ensemble with 1 atm pressure maintained by Langevin piston barostat¹²⁰, and 310 K, controlled by Nosé-Hoover thermostat^{121, 122}. MD simulations with applied voltage used to study ion conduction were carried out in the constant volume *NVT* ensemble. Tetragonal cells with periodic boundary conditions (PBC) were used in all the simulations, and the SHAKE algorithm¹²³ was employed to fix the bonds to all hydrogen atoms, allowing for the use of a 2 fs time step. Electrostatic interactions were computed via Particle Mesh Ewald¹²⁴, with a mesh grid of 1 Å.

For all hERG simulations in this study we used an extended staged MD equilibration protocol. For drug-free and drug “flooding” simulations we gradually reduced harmonic restraints on the backbone atoms of the whole protein, then pore domain and finally selectivity filter from 1.0 to 0.1 kcal/mol/Å² over the first 40 ns and then continued equilibration simulations for additional 50 ns without restraints. For enhanced sampling hERG – drug binding simulations we kept 1.0 kcal/mol/ Å² harmonic restraints on the selectivity filter backbone non-hydrogen and pore domain C_α atoms throughout initial 50 ns long equilibration with drug in bulk solvent and 90 ns long steered MD simulations for drug pulling into the channel pore. To avoid bias related to initial drug orientation and conformation, five independent steered MD runs with different starting points were used to randomly “seed” structures for umbrella sampling simulations. We gradually reduced restraints to 0.2 kcal/mol/Å² over the course of initial 5 ns for each umbrella sampling run and kept them throughout production simulations to prevent a random channel transition to a different conformational state.

hERG open-state atomistic model validation

Our hERG open state channel pore was found to be stable in a ~1 μ s long unbiased MD simulations, and potassium ion permeation was measured by applying transmembrane voltage during multi-microsecond simulations on Anton 2¹¹⁴. To do so, a uniform electric field was applied in z direction, and it can be computed as:

$$E_z = \frac{V}{L_z \cdot 43.5} \quad [14],$$

Where V is the voltage in mV, E_z is the z component of the electric field vector in kcal/(mol· \AA ·e), and L_z is the length of the unit cell in z direction in \AA . A factor of 43.5 was used to convert from mV/ \AA to kcal/(mol· \AA ·e). Seven instances of outward K⁺ conduction were observed (depicted in Online Figure V) during 0.3 μ s of a ~5 μ s long unrestrained hERG MD simulation under 750 mV applied voltage, with similar findings for a Kv1.2/2.1 chimera (PDB ID: 2R9R)¹²⁵ MD run under the same conditions. This confirmed that our hERG model represent an open conducting state of the channel.

Atomistic force field parameterization of drug models

Atomistic models for cationic and neutral dofetilide and moxifloxacin as well as zwitterionic moxifloxacin were optimized using initial guesses from generalized CHARMM force field (CGENFF) program^{126, 127} and the ffTK plugin¹²⁸ for the Visual Molecular Dynamics program (VMD)¹²⁹. Gas-phase quantum mechanical (QM) calculations utilizing Møller–Plesset (MP2) and Hartree-Fock (HF) perturbation theory and the 6-31(d) basis set in Gaussian 09¹³⁰ program were used to compute target data for molecular mechanical (MM) parameter optimization. Optimized charges (*Data Supplement Tables DS1 &DSII*) provide a good agreement with QM target dipole values. The optimized MM dipole moments are larger in magnitude compared to QM MP2/6-31G(d) dipole moments by 19% (9.7 vs. 8.1 Debye) for neutral dofetilide, 17% (10.8 vs. 9.2 Debye) for cationic dofetilide, 12% (41.3 vs. 37.0 Debye) for zwitterionic moxifloxacin, 20% (24.6 vs. 20.3 Debye) for cationic moxifloxacin, and 17% (9.9 vs 8.6 Debye) for neutral moxifloxacin, which are all close to the 20% threshold suggested for

CGENFF. The water interaction distances and energies were also in good agreement with QM values (*Data Supplement* Tables DSIII-DSVII). Dihedral angle parameter optimizations resulted in substantial improvement over CGENFF initial guesses, with optimized torsional energy minima within ~1 kcal/mol of QM values for dofetilide (*Data Supplement* Figure DSII) and within ~2 kcal/mol for moxifloxacin (*Data Supplement* Figures DSIII & DSIV). Final topology and parameters for dofetilide and moxifloxacin are provided at the end of this document in *Data Supplement* Tables DSVIII – DSXII.

Atomistic drug model validation

To validate developed drug force field parameters, all-atom umbrella sampling (US) MD simulations¹³¹ of drug partitioning across a hydrated lipid bilayers (POPC for dofetilide, and DMPC for moxifloxacin) were performed. For each drug model, 81 independent simulation windows were created, in which the center of mass (COM) of a randomly oriented drug molecule was placed at 1 Å intervals from z=–40 Å to z=40 Å with respect to COM of the membrane. In addition, for the membrane-spanning central windows, z=–20 Å to z=20 Å, additional simulations were performed, in which the drug was flipped about its z-axis in order to enhance sampling, and hence reduce asymmetries in the computed potential of mean force (PMF) profiles (Online Figure VI). For all US MD simulations, the COM of the drug was restrained along the z-axis with a force constant of 2.5 kcal/mol/Å², and an additional 5 kcal/mol/Å² cylindrical restraint was applied in order to prevent its drift in the xy plane. The PMF profiles were computed using the weighted histogram analysis method (WHAM)¹³² with error bars representing standard errors of mean computed from profile asymmetries with respect to z = 0 (error($W(z)$) = $|W(z) - W(-z)| / 2$). See Online Figure VI for convergence of unsymmetrized PMF profiles.

Symmetrized diffusion coefficient profiles (–40 Å to 0 Å shown in Main Text Figure 1D &1H) were obtained using Laplace transform of drug position autocorrelation function¹³³ as was described in our recent studies^{42, 43}. Water-membrane distribution coefficients log D_{MW} were computed as was done previously^{43, 134, 135}, resulting in 0.32±0.15 for

dofetilide and 0.13 ± 0.11 for moxifloxacin. The values for dofetilide were in reasonable agreement with experimental water-octanol (0.84^{136} , 0.96^{137}) and water-artificial membrane (2.08^{136}) values. The values for moxifloxacin agreed less favorably with an experimental value for DMPC membrane (2.4^{138}), but in a good agreement with water-octanol and water-artificial membrane values (1.12 and 1.57 , respectively 136). To compute the translocation rates of both drugs across lipid membranes we used Kramer's transition rate approximation, as was done previously $^{139, 140}$ using Laplace transform of position autocorrelation function 133 for computing diffusion coefficients at the z positions of the PMF profile peaks, and also computing PMF barrier heights and profile curvatures at the peaks and wells.

Drug binding to the hERG pore

Both drugs in each ionization state were found to spontaneously access through an intracellular gate and bind in the hERG channel pore during $2.5\text{ }\mu\text{s}$ long unbiased “flooding” MD simulations with multiple (11) drug molecules corresponding to $\sim 25\text{ mM}$ initial aqueous concentration. However, in most cases we did not observe any drug unbinding events, and had to use enhanced sampling simulations to compute affinities and rates as described below. The only exception is cationic moxifloxacin, for which we only observed transient drug binding and unbinding events at the intracellular end of the pore. We also did not observe any drug binding through lipid-facing fenestrations as in the case of Nav channels, $^{42, 135}$ justifying enhanced sampling simulations for drug binding through an intracellular channel gate. To do this, US MD simulations were run for 40 ns for each US window for dofetilide, and 70 ns per US window for moxifloxacin. US windows corresponded to harmonically restrained drug center of mass z positions, in 0.5 \AA increments from -50 to -5 \AA with respect to hERG selectivity filter (SF) C α atoms. The first 10 ns of each run was considered equilibration, and the rest was used to compute free energy, or potential of mean force (PMF) profiles (see Online Figure VII for convergence) using the weighted histogram analysis method (WHAM), 132 and diffusion coefficient profiles via Laplace transform of drug position autocorrelation function 133 . Error bars for PMF profiles in ain text Figure 2 B & D were computed as

standard errors of mean from block averaging analysis. For that we separated US simulation data into three blocks of 5 ns for dofetilide or 20 ns for moxifloxacin and computed PMF profiles from each of them.

Dissociation constants, K_D , were computed from PMF profiles using established methods¹⁴¹ and are shown in main text Figure 2E. For open state hERG binding, cationic and neutral dofetilide K_D were computed to be 65 μM and 0.16 μM , respectively, and 8600 μM , 6700 μM , and 0.74 μM for zwitterionic, cationic, and neutral moxifloxacin, respectively. For both drugs, there was much stronger binding of the neutral form.

To compute the drug-channel association or ingress “on” rate (k_{on}) for each drug binding to hERG, we assumed that the binding process is a purely diffusion-limited reaction (barrier-free), and that the reaction is fast once a drug molecule enters the reactive region. Diffusion coefficient, $\mathcal{D}(z)$, profiles for each drug going from the intracellular bulk aqueous solution region into the hERG pore were obtained using Laplace transform of drug position autocorrelation function¹³³ as was done in our recent studies on drug – membrane partitioning^{42, 43}. The rates, k_{on} , were computed from $\mathcal{D}(z)$ and free energy, $W(z)$, profiles using a formulation of the Debye-Smoluchowski equation^{49, 50}:

$$k_{\text{on}}^{-1} = \frac{1}{\pi R^2} \int_{z_{\text{out}}}^{z_{\text{bind}}} e^{\frac{W(z)}{k_B T}} \mathcal{D}(z)^{-1} dz \quad [15]$$

where R is the radius of a cylinder (10 Å) encompassing the channel pore and used as a flat-bottomed restraint for US MD simulations, k_B is Boltzmann constant, T is absolute temperature, and z is the reaction coordinate. Using computed K_D and k_{on} values, we can estimate drug – channel dissociation rate constant, k_{off} , as:

$$k_{\text{off}} = k_{\text{on}} \cdot K_D \quad [16].$$

Computed K_D , k_{on} and k_{off} values are listed in main text Figure 2E and were used as parameters for drug-hERG interaction function scale models (see sections above and main text).

We also compared computed drug ingress rates at maximum physiological plasma concentrations (C_{max}) of dofetilide (~6.16 nM, see above) and moxifloxacin (~6.23 μ M, see above) using $x \cdot C_{max} \cdot k_{on}$, where x is each drug ionization species fraction at pH = 7.2. For dofetilide, ingress rates at maximal plasma concentration were computed to be equal to 2.5 and 1.2 s^{-1} for cationic and neutral dofetilide, respectively, and 1,800, 270, and 230 s^{-1} for zwitterionic, cationic, and neutral moxifloxacin, respectively. Drug-membrane translocation rates, in comparison, were 8,000 s^{-1} for neutral dofetilide, 43,000 s^{-1} for cationic moxifloxacin, and $1.1 \cdot 10^6 s^{-1}$ for neutral moxifloxacin. For either drug, channel ingress rates are 2–3 orders of magnitude slower than membrane translocation rates, indicating that kinetics associated with open hERG – drug interactions will be rate-determining.

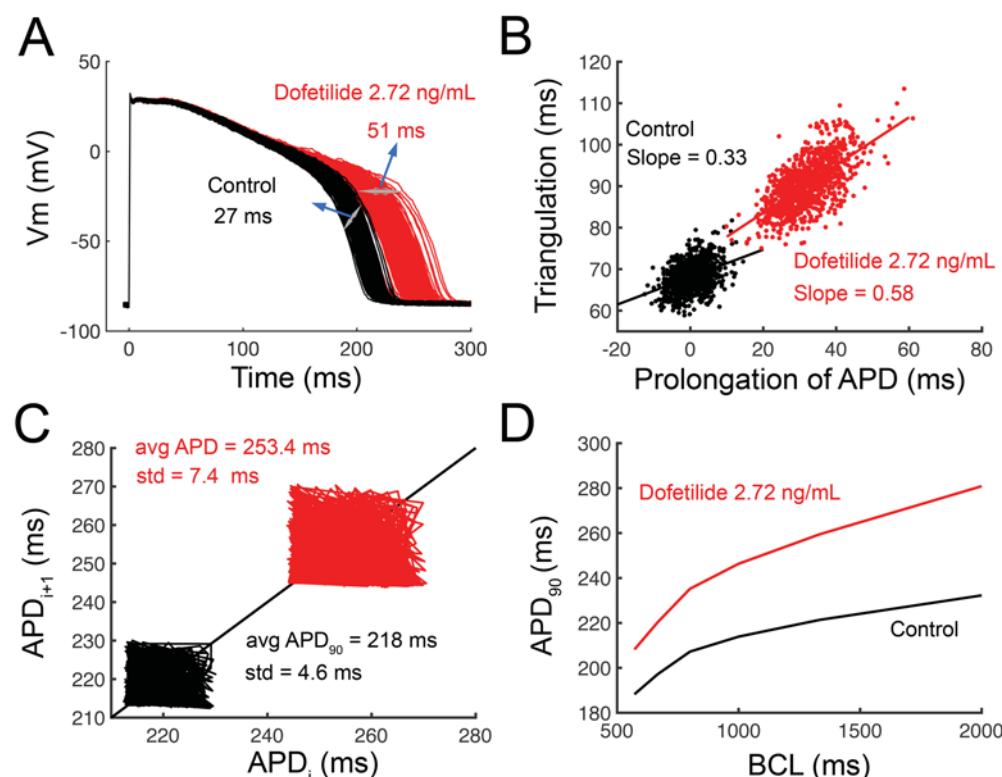
Drug binding pocket topologies within the hERG pore

The topologies of the binding pockets for the neutral and cationic forms of dofetilide in representative poses from US MD simulations are shown in Online Figure VIII A and VIII B, respectively. Neutral dofetilide binds at $z=-15.5$ Å with one end pointing up toward polar residues in the SF, stabilized by interactions near its base (S624) as well as a cluster of residues from S6 helix including S660, Y652 and F656 from multiple chains, which are all the residues that have been implicated in experimental drug binding studies¹⁴². Cationic dofetilide was observed to bind below the Y652 ring at $z=-20$ Å and is also coordinated by hydrophobic residues F656 and Y652 from multiple chains, with both terminal methanesulfonamide groups coordinated via hydrogen bonds with S660 residues (near the intracellular ends of S6 helices) from two chains (see Online Figure VIII B). In other words, both ends of the molecule point down toward the bulk aqueous solvent, whereas its cationic ammonium group in the middle points up towards the SF and forms a hydrogen bond with Y652. Zwitterionic moxifloxacin was found to bind at the intracellular end of the pore at $z=-22.5$ Å and coordinate with S6

residues Y652, F656, S660 as well as Q664 (see Online Figure IX A). Neutral moxifloxacin most tightly binds in the hERG pore at $z=-20$ Å and engages in the interactions with S6 helix residues F656, G657, and S660 from two chains (Online Figure IX B). Cationic moxifloxacin was found to bind hERG at $z=-21.5$ Å interacting with S6 residues N658, S660 and Q664 (Online Figure XI C).

Online Figures I - IX

Online Figure I

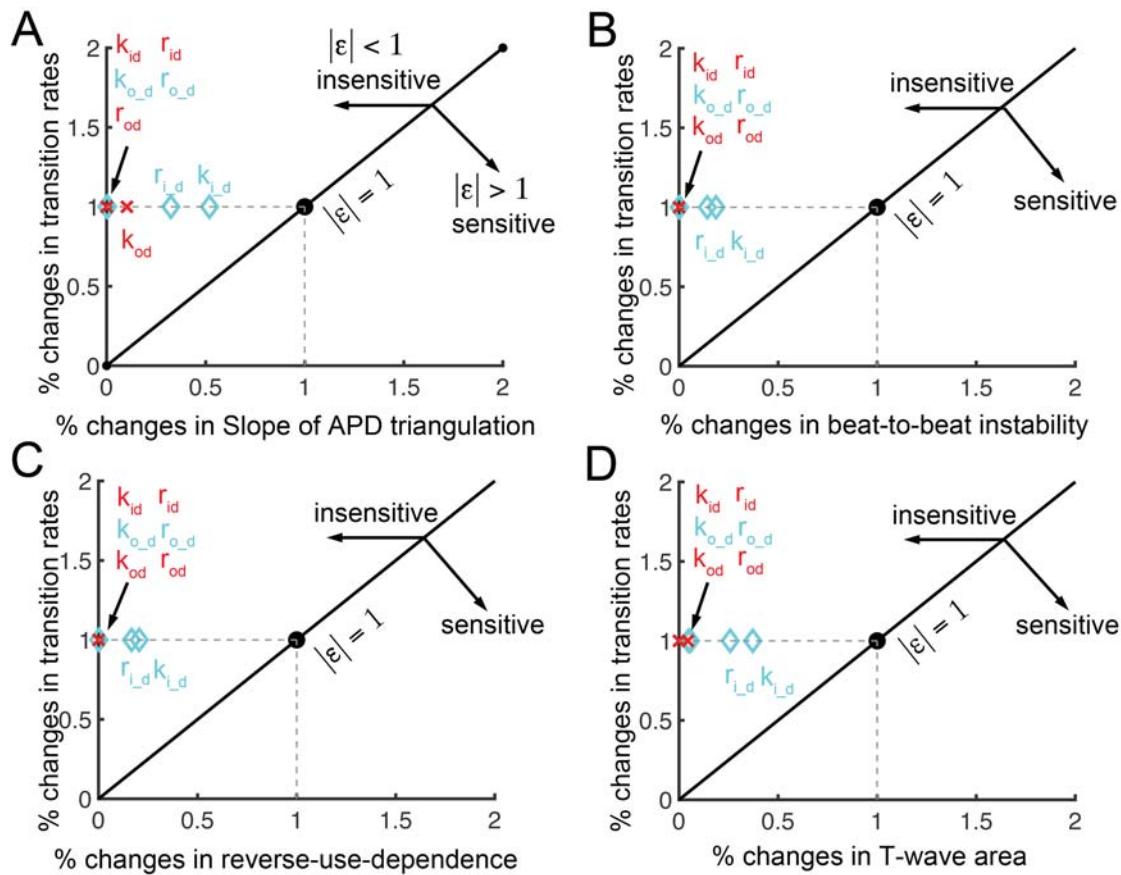


Computational screen of arrhythmia vulnerability in rabbit model. In panel A, **temporal action potential duration dispersion** was quantified in a cell population of

1000 individual simulated cardiac myocyte action potentials constructed by incorporating physiological noise^{67, 68}. Dispersion of APD was quantified as the difference between the maximum and minimum action potential duration. Dofetilide within the clinical dosing range has a clear effect to promote temporal action potential duration variability in the presence of the drug (Control – 27 ms; Dofetilide 2.72 ng/mL = 51 ms). Panel B illustrates the effect of dofetilide to promote ***triangulation*** of the action potential as a function of APD prolongation. In the absence of drug, control cells had a slope = 0.33, while Dofetilide 2.72 ng/mL increased the slope = 0.58. Panel C shows Poincaré plots of sequential APD pairs indicating beat-to-beat ***instability*** following the application of small electrical perturbations in the absence of drug or with 2.72 ng/mL dofetilide. Instability was assessed by applying small amplitude inward currents randomly between -0.1 to -0.2 pA/pF for 50 ms over the course of the action potential plateau at a pacing cycle length = 1000 ms. Finally, in panel D, ***reverse use dependence*** induced by dofetilide was evaluated. The action potential adaptation curves were generated using APD₉₀ values from human computational ventricular myocytes at steady-state at the indicated pacing frequencies. When dofetilide (red) was applied, there was a clear steepening of the APD adaptation curve compared to the baseline drug-free case (black).

Online Figure II

Dofetilide

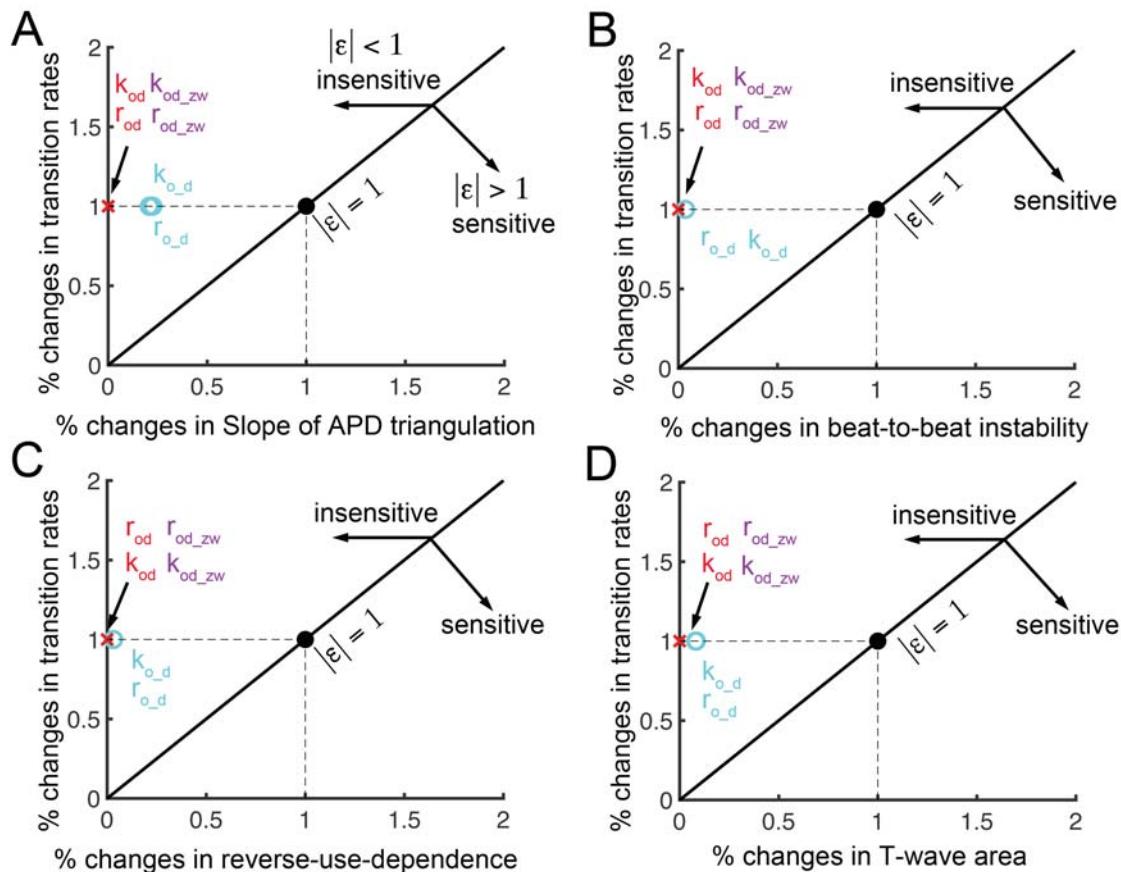


Local sensitivity analysis of arrhythmia vulnerability parameters from the TRIaD for Dofetilide

In order to assess the specific drug-channel interactions comprising the dofetilide structure-activity relationship and the link to proarrhythmia, we undertook sensitivity analysis to determine how sensitive the arrhythmia vulnerability parameters from the TRIaD based simulations are to our underlying model parameters. As shown in panel A, we first carried out an *in silico* test of the local sensitivity of the slope of the relationship between action potential *triangulation* and APD prolongation in O'Hara-Rudy computational myocytes plotted for a range of drug “on” (k_{o_d} , k_{od} , k_{i_d} , and k_{id}) and “off” (r_{o_d} , r_{od} , r_{i_d} and r_{id}) model transition rates for open and open-inactivated states by increasing and decreasing each rate at $\pm 20\%$ for neutral (cyan) and cationic (red), and calculated elasticity coefficients. The local sensitivity analysis showed that perturbation to the rate constant k_{i_d} (open-inactivated state binding) of neutral drug results in the greatest effect on the slope of APD triangulation. The elasticity value of k_{i_d} shown in panel A is 0.52, which suggests that if the rate constant k_{i_d} is increased by 1% then the slope of APD triangulation increases by approximately 0.52%, indicating relative insensitivity to even the most sensitive model parameter. Perturbation to all other rate constants also results in less than 1% changes in the slope of APD triangulation. Similarly, in panel B, simulated beat-to-beat *instability* of action potentials (average and standard deviation of APD₉₀ for each rate perturbation are shown) with respect to transition rate changes are demonstrated. And panel C shows relative changes in transition rates with respect to the APD₉₀ at a slow pacing rate, which reflect maximal reverse use dependence (BCL = 2000 ms). Again, binding and unbinding rate parameters of neutral drug (cyan) for inactivated channel state caused more changes in the model outputs compared to other parameters, but the model outputs are still relatively insensitive. Finally, panel D, shows higher elasticity values of the T-wave area to neutral drug (cyan) binding and unbinding rates for inactivated state. However, the analysis showed that the model was robust to perturbations (elasticity value < 1).

Online Figure III

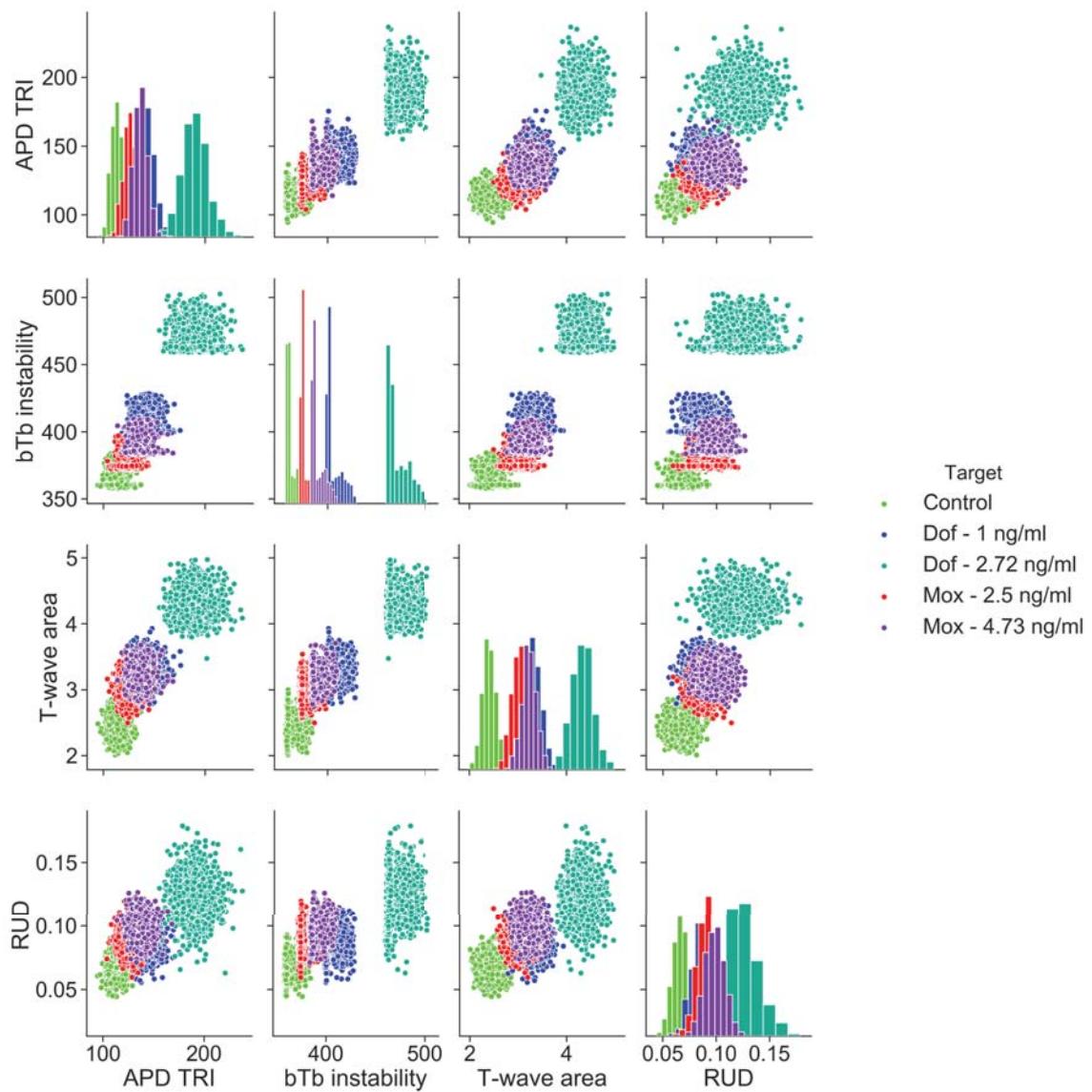
Moxifloxacin



Local sensitivity analysis of arrhythmia vulnerability parameters from the TRIaD for Moxifloxacin

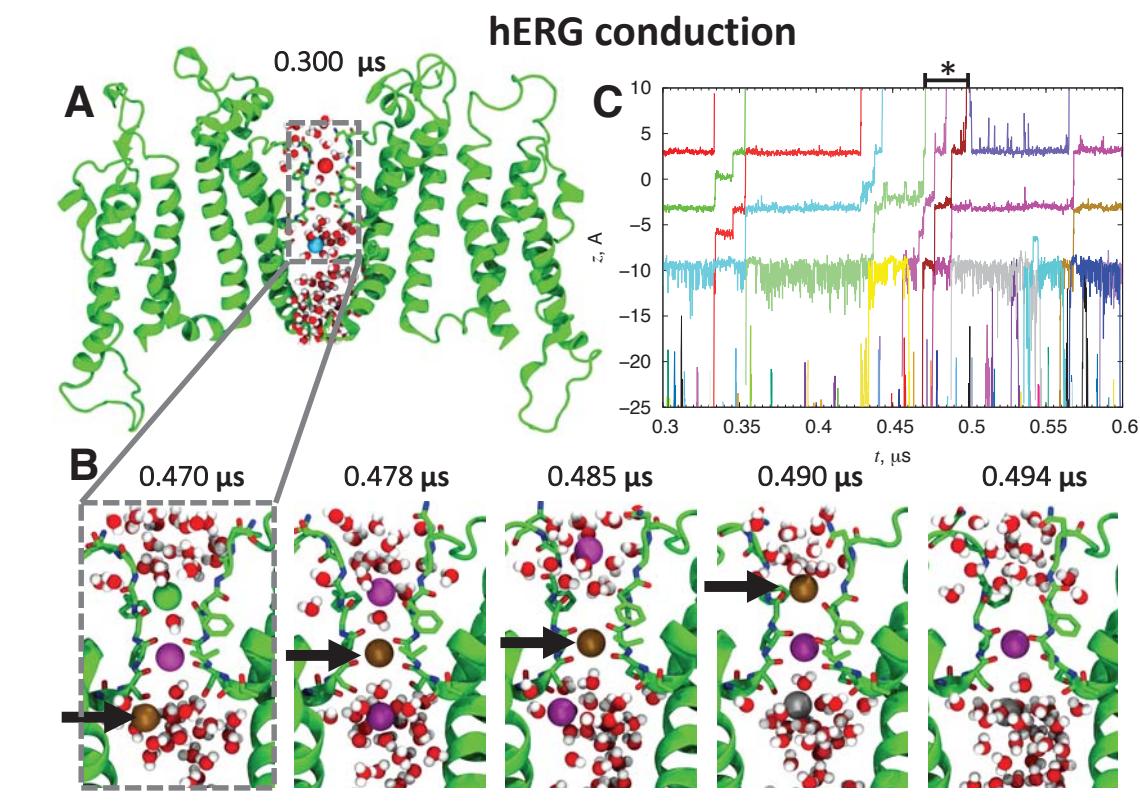
A) The sensitivity of the slope of the relationship between action potential *triangulation* in O'Hara-Rudy computational myocytes plotted for a range of drug "on" (k_{o_d} , k_{od} , and k_{od_zw}) and "off" (r_{o_d} , r_{od} , and r_{od_zw}) model transition rates for open and inactivated states by increasing and decreasing each rate at $\pm 20\%$ for neutral (cyan), cationic (red), and zwitterionic (purple). B) Sensitivity of simulated beat-to-beat *instability* of action potentials for a range of rate constants. C) The sensitivity to changes in drug transition rates of the recorded APD₉₀ at BCL of 2000 ms. D) Sensitivity of the T-wave area to model transition rates.

Online Figure IV



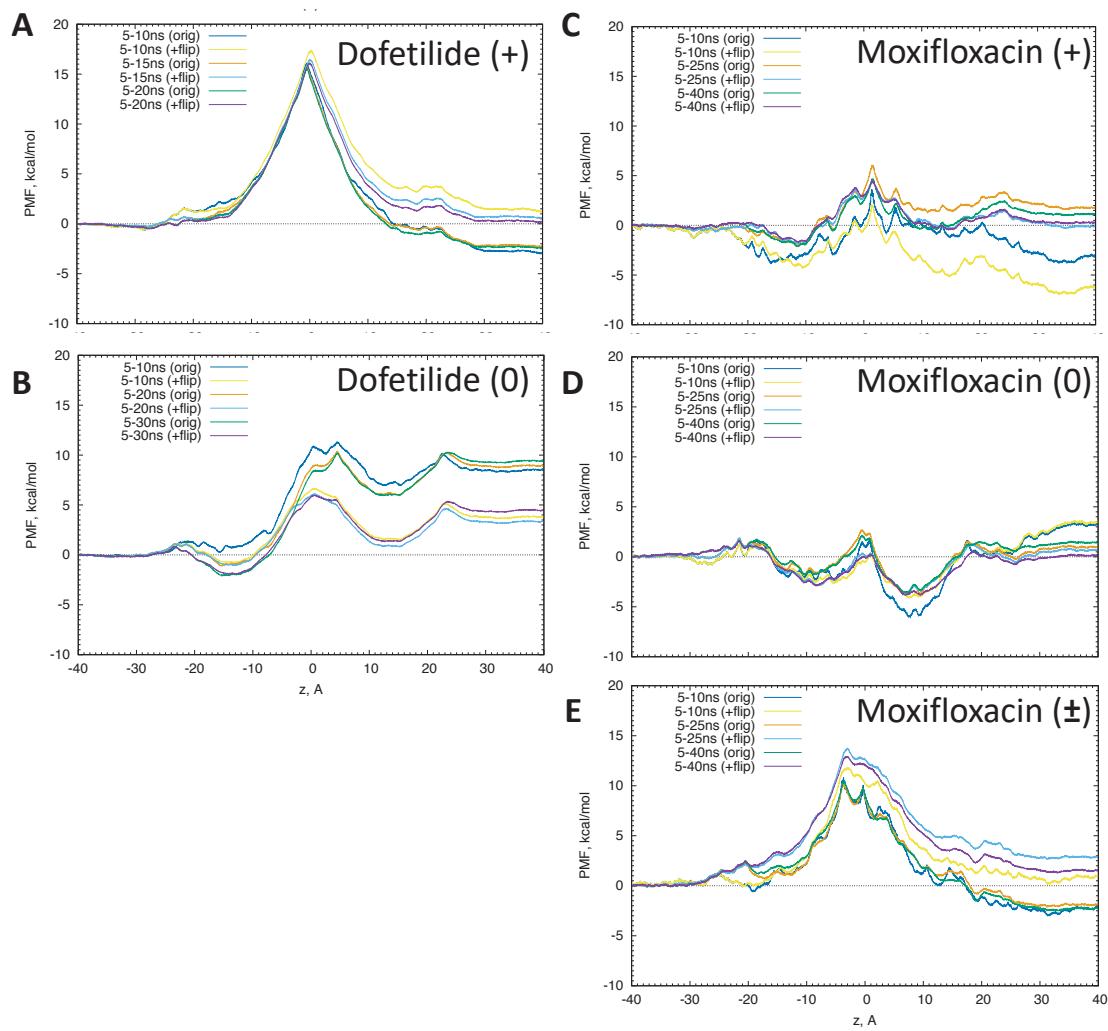
The scatter plots show the distribution of the arrhythmia vulnerability parameter from TRId (APD TRI, bTb instability, T-wave area and RUD) and relationship between the target (classification of control, different moxifloxacin doses and different dofetilide doses).

Online Figure V



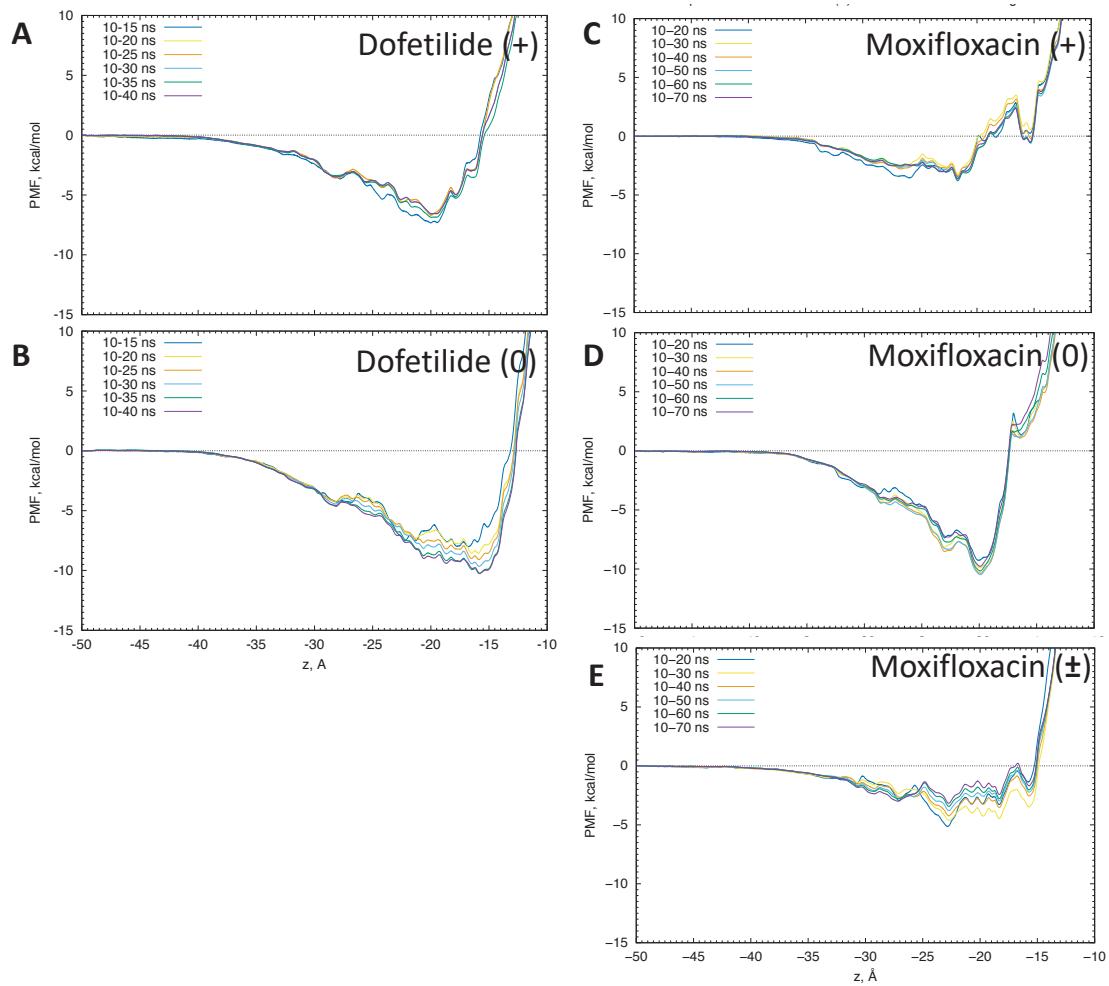
K⁺ conduction of atomistic open state hERG model under an applied 750 mV voltage. A 0.3 μs slice of a 5 μs trajectory, where most conduction events took place, is shown. **(A)** Initial frame in this time slice ($t = 0.300 \mu\text{s}$) showing two opposite protein chains (green ribbons with SF residues S624-G628, S6 helix Y652 and F656 residues shown as sticks with red O and blue N), pore ions (colored balls) and waters (red/white). **(B)** Close-up views of the channel SF in the same representation at different time points, showing a complete translocation of one ion (brown ball), indicated by an arrow. **(C)** Time series of ion z-positions (with respect to the SF backbone center of mass). Colors of the z profiles match those of the ions in panels **A**, **B**. Portion of the profiles corresponding to snapshots in panel **B** is indicated with an asterisk.

Online Figure VI



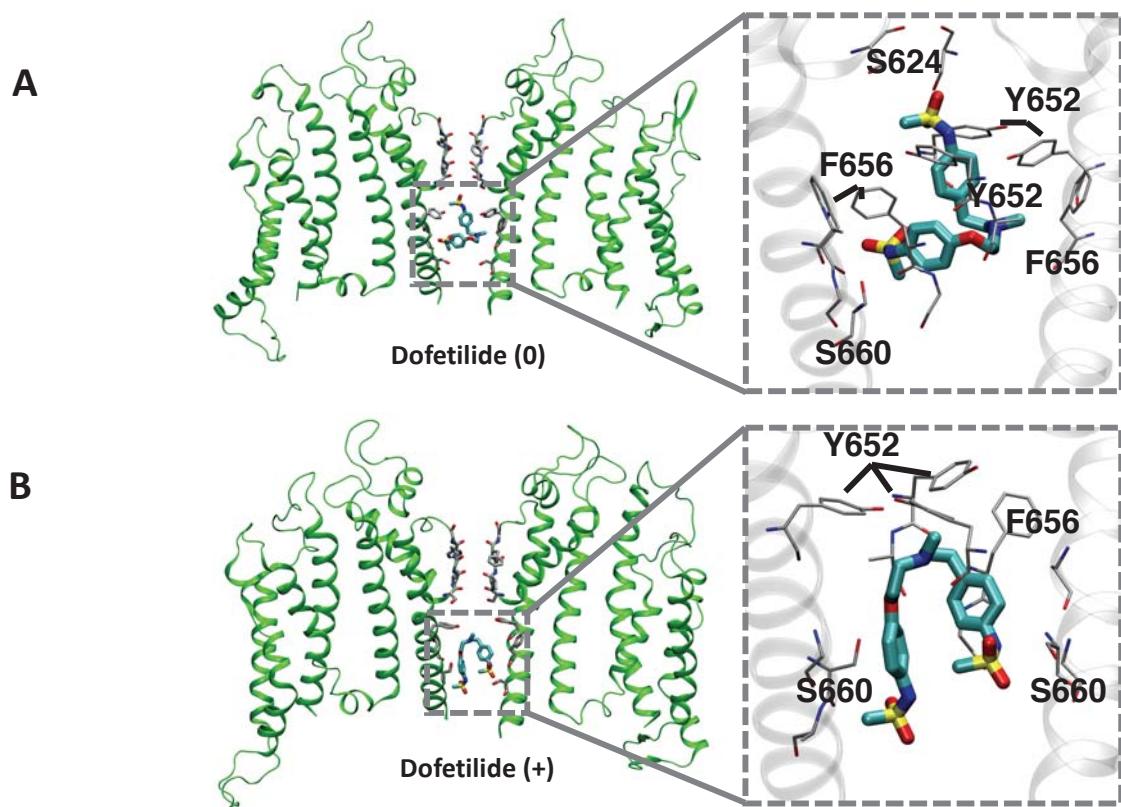
Non-symmetrized free energy or potential of mean force (PMF) profiles for drug models crossing a lipid bilayer computed from umbrella sampling MD simulations. Cationic (**A**) and neutral (**B**) dofetilide across a POPC bilayer, and cationic (**C**) neutral (**D**) and zwitterionic (**E**) moxifloxacin across a DMPC bilayer. They were computed using weighted histogram analysis method (WHAM) discarding first 5 ns for each umbrella sampling window. In some cases, additional simulations with initial drug orientation rotated around the z-axis (indicated as +flip) were performed to improve drug re-orientation sampling.

Online Figure VII



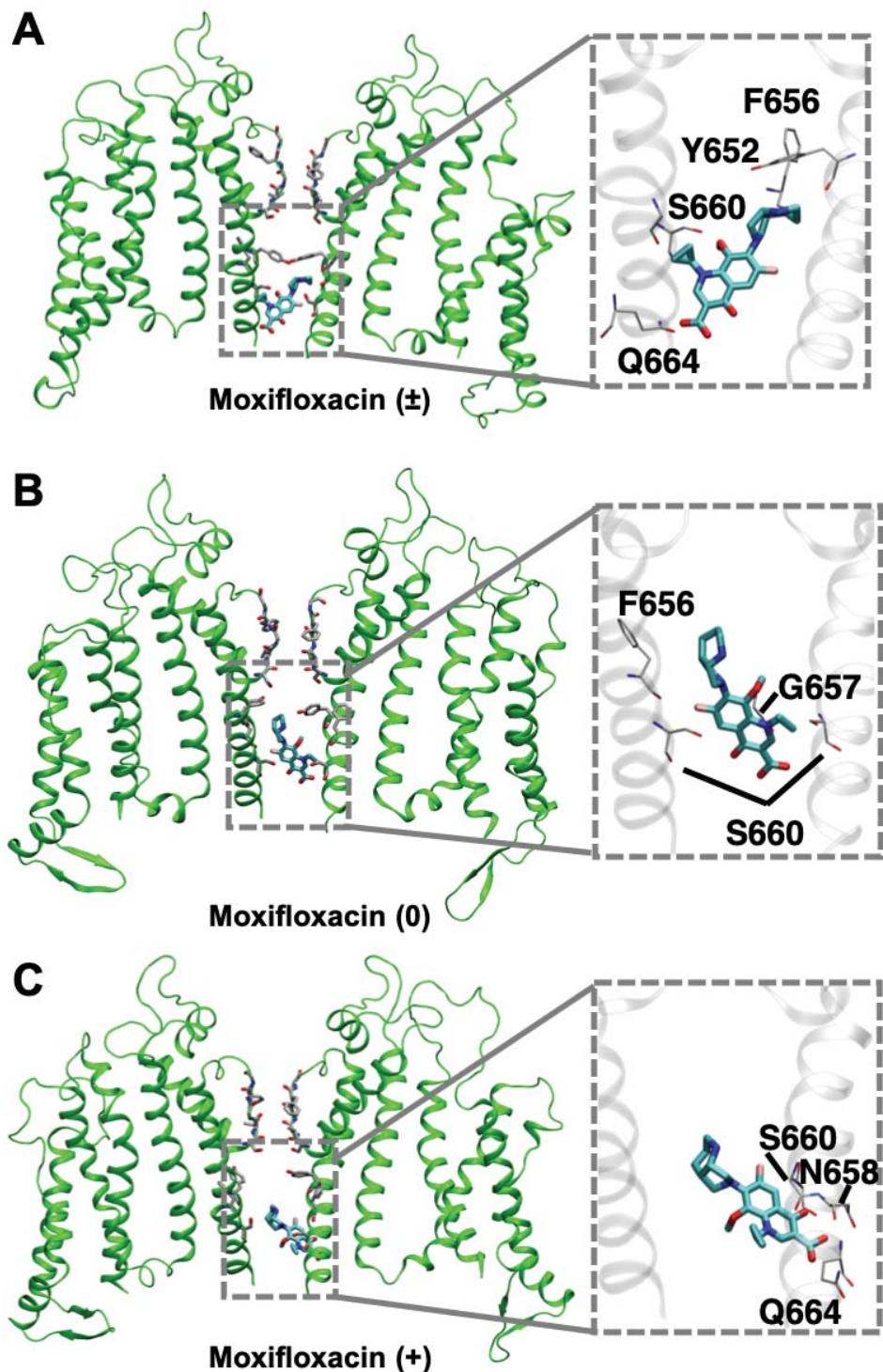
Convergence of free energy or potential of mean force (PMF) profiles for dofetilide and moxifloxacin binding to an open hERG channel model computed from umbrella sampling MD simulations. Cationic (**A**) and neutral (**B**) dofetilide profiles, and cationic (**C**) neutral (**D**) and zwitterionic (**E**) moxifloxacin profiles computed using weighted histogram analysis method (WHAM) discarding first 10 ns for each umbrella sampling window, and incrementally including data from additional 5 ns blocks up to 40 ns for dofetilide and 10 ns blocks up to 70 ns for moxifloxacin.

Online Figure VIII



Cationic and neutral dofetilide hERG binding from umbrella sampling MD simulations. Representative lowest free energy drug binding configurations for neutral (A) and cationic (B) dofetilide in the hERG pore. Two opposite hERG chains with a drug bound are shown on the left, whereas insets on the right show close-up views of hERG residues interacting with dofetilide (within 3.5 Å counting non-H atoms only). hERG is shown as ribbons (green or gray), interacting residues as thin sticks and dofetilide as thick sticks (C – cyan, O – red, N – blue, S – yellow, F – pink).

Online Figure IX



Zwitterionic (\pm), neutral (0) and cationic (+) moxifloxacin hERG binding from umbrella sampling MD simulations. See Online Fig. VIII for more details.

Data Supplement: Atomistic force field models for different ionization states of dofetilide and moxifloxacin

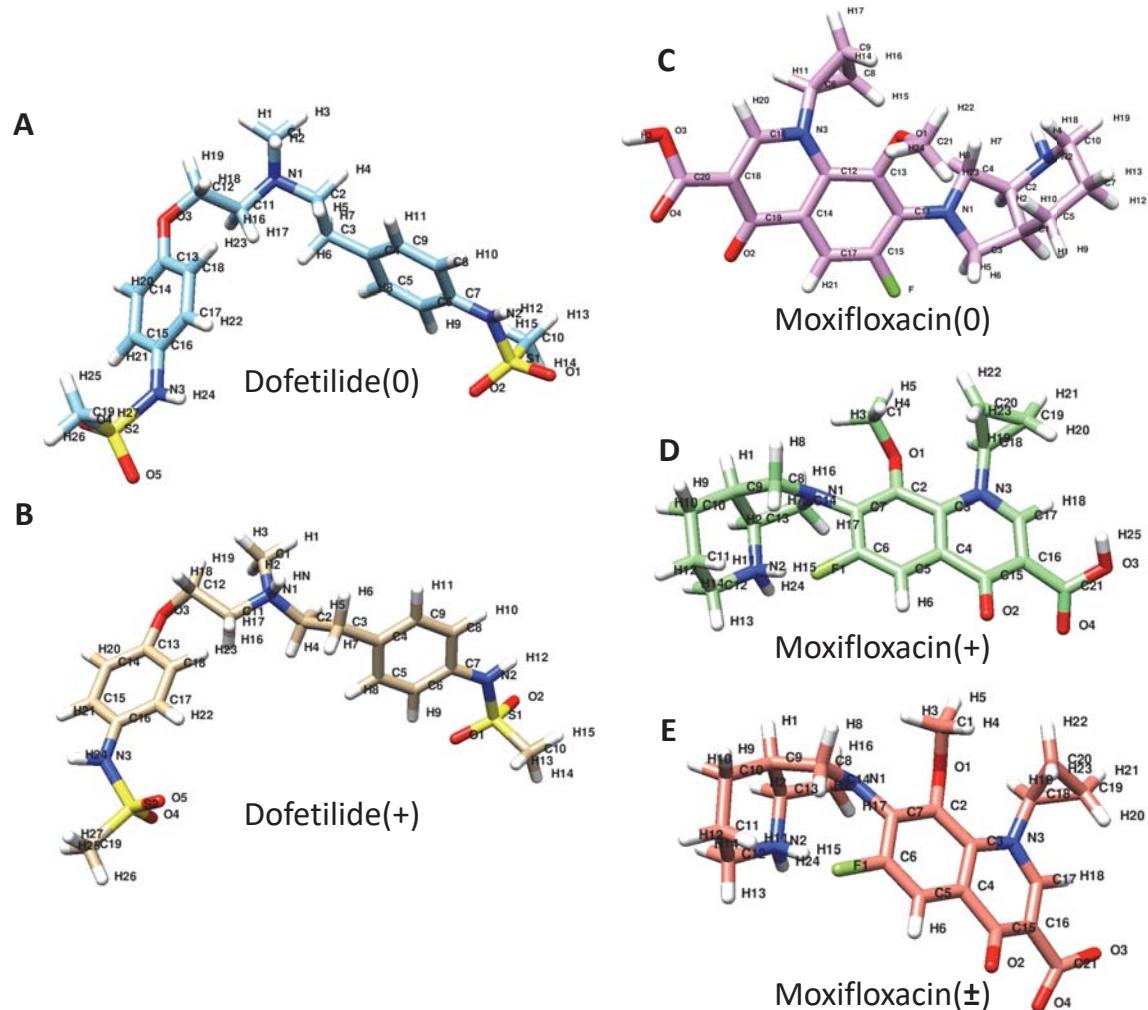
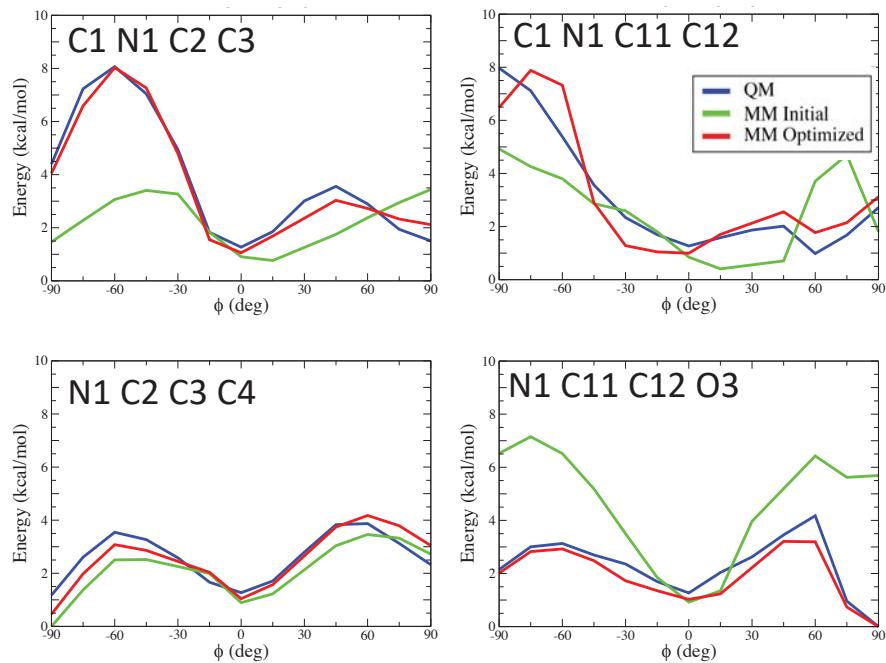


Figure DSI. Atom naming for different ionization states of dofetilide and moxifloxacin atomistic force field models. (A) Neutral dofetilide (DOFN or Dofetilide(0)). (B) Cationic dofetilide - DOFC or Dofetilide(+) (C) Neutral moxifloxacin - MOX0 or Moxifloxacin (0) (D) Cationic moxifloxacin - MOXC or Moxifloxacin (+), and (E) Zwitterionic moxifloxacin - MOXZ or Moxifloxacin (\pm)

A Dofetilide(0) Dihedral Scans



B Dofetilide(+) Dihedral Scan

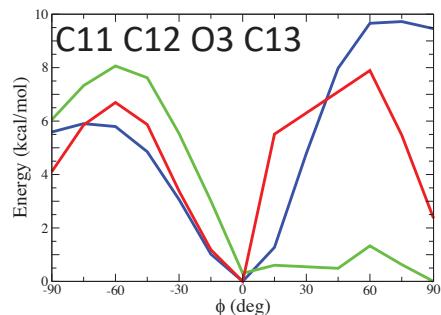


Figure DSII. Gas-phase torsional energy profiles for neutral and cationic dofetilide models from quantum mechanical (QM), initial and optimized molecular mechanics (MM) calculations. Atom names corresponding to the ones in *Data Supplement* Fig. DSIA and Fig.DSIB for (A) Dofetilide (0) or DOFN, and (B) Dofetilide (+) or DOFC, respectively, with corresponding topology and parameter files in the *Data Supplement* Tables DSVIII and DSIX.

Moxifloxacin (0) Dihedral Scans

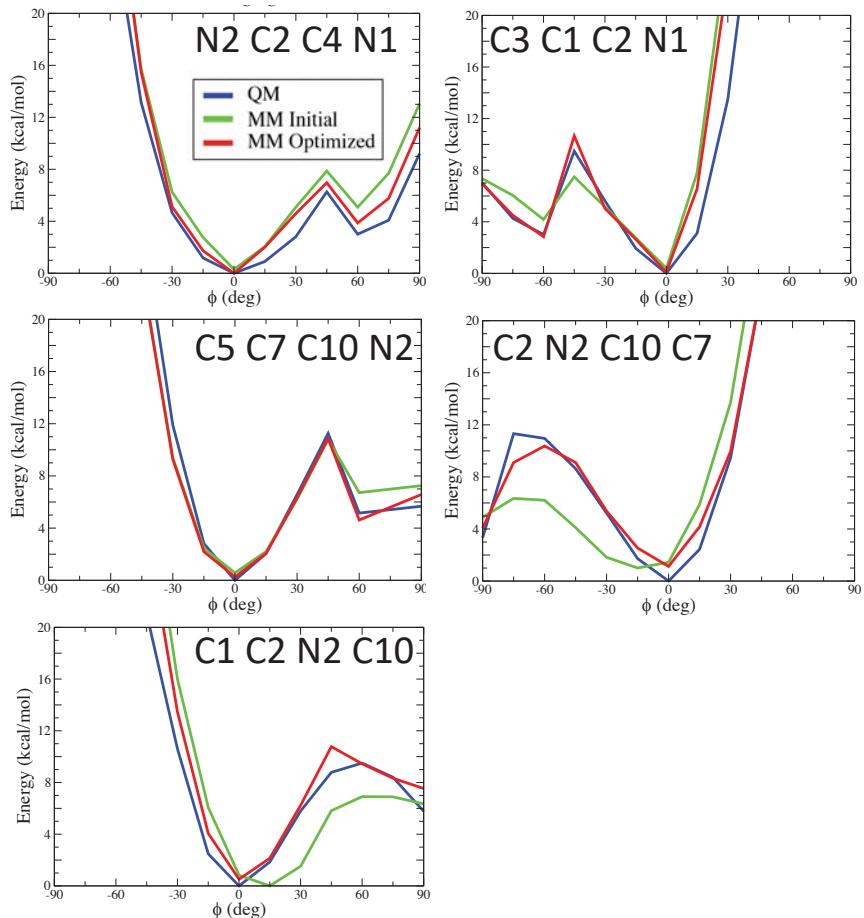


Figure DSIII. Gas-phase torsional energy profiles for neutral moxifloxacin drug model from quantum mechanical (QM), initial and optimized molecular mechanics (MM) calculations. Atom names corresponding to the ones in *Data Supplement Fig. DSIC* for Moxifloxacin (0) with corresponding topology and parameter files in the *Data Supplement Table DSXII*.

Moxifloxacin (\pm) Dihedral Scans

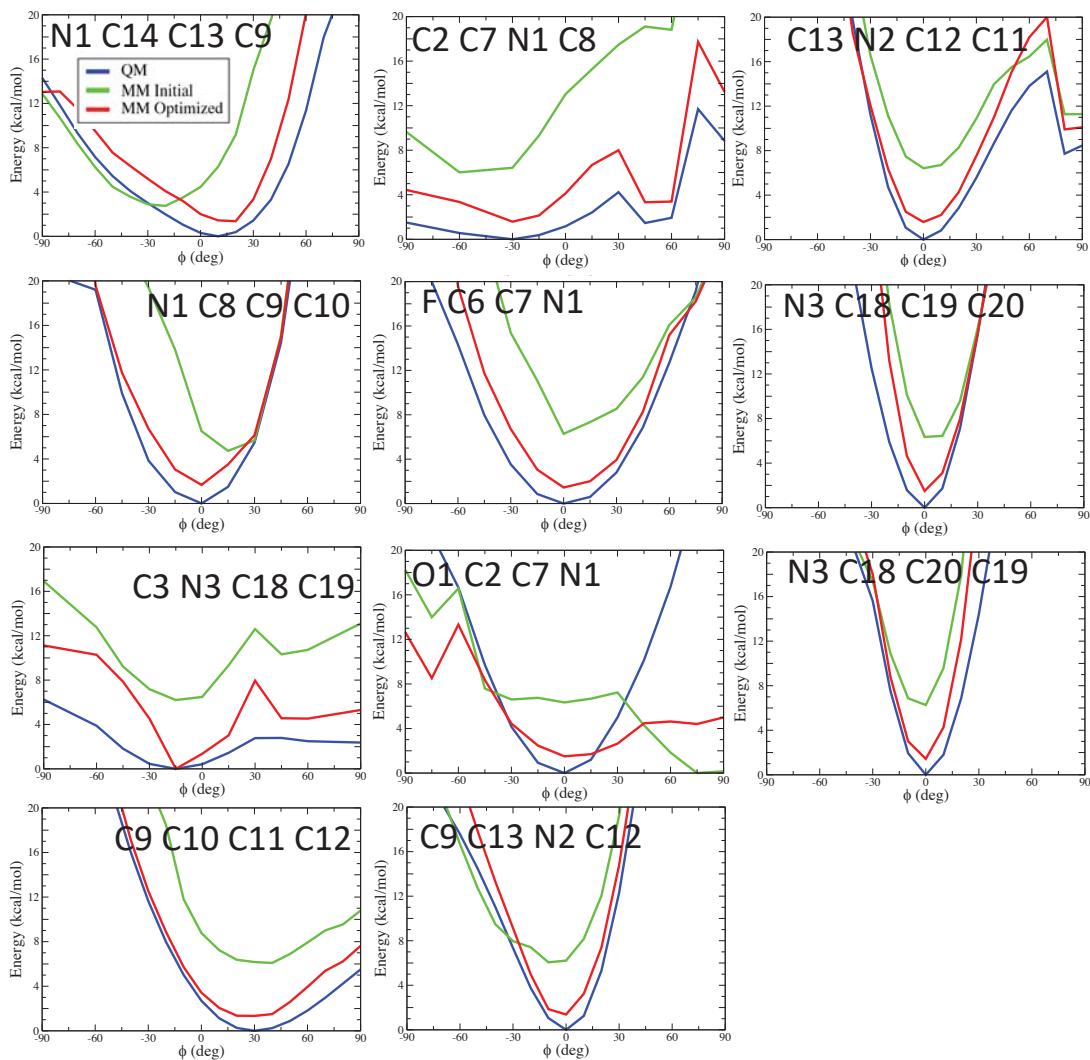


Figure DSIV. Gas-phase torsional energy profiles for zwitterionic moxifloxacin drug model from quantum mechanical (QM), initial and optimized molecular mechanics (MM) calculations. Atom names corresponding to the ones in Fig. DSIE for Moxifloxacin (\pm), with corresponding topology and parameter files in the *Data Supplement Table DSX*.

| Drug | Total Charge | Drug | Total Charge |
|------|--------------|------|--------------|
| DOFC | 1 | DOFN | 0 |
| C1 | 0.149 | C1* | -0.392 |
| N1* | -0.006 | N1* | -0.401 |
| C2* | -0.495 | C2* | 0.24 |
| C3* | -0.25 | C3* | -0.559 |
| C4* | 0.31 | C4* | 0.296 |
| C5 | -0.115 | C5 | -0.115 |
| C6 | -0.111 | C6 | -0.111 |
| C7 | 0.219 | C7 | 0.219 |
| C8 | -0.111 | C8 | -0.111 |
| C9 | -0.115 | C9 | -0.115 |
| N2 | -0.466 | N2 | -0.466 |
| S1 | 0.439 | S1 | 0.439 |
| O1 | -0.384 | O1 | -0.384 |
| O2 | -0.384 | O2 | -0.384 |
| C10 | -0.019 | C10 | -0.019 |
| C11 | 0.162 | C11* | 0.01 |
| C12 | -0.014 | C12 | -0.021 |
| O3 | -0.391 | O3 | -0.39 |
| C13 | 0.219 | C13 | 0.219 |
| C14 | -0.113 | C14 | -0.113 |
| C15 | -0.115 | C15 | -0.115 |
| C16 | 0.219 | C16 | 0.219 |
| C17 | -0.115 | C17 | -0.115 |
| C18 | -0.113 | C18 | -0.113 |
| N3 | -0.466 | N3 | -0.466 |
| S2 | 0.439 | S2 | 0.439 |
| O4 | -0.384 | O4 | -0.384 |
| O5 | -0.384 | O5 | -0.384 |
| C19 | -0.019 | C19 | -0.019 |
| H1 | 0.09 | H1 | 0.09 |
| H2 | 0.09 | H2 | 0.09 |
| H3 | 0.09 | H3 | 0.09 |
| H4 | 0.09 | H4 | 0.09 |
| H5 | 0.09 | H5 | 0.09 |
| H6 | 0.09 | H6 | 0.09 |
| H7 | 0.09 | H7 | 0.09 |
| H8 | 0.115 | H8 | 0.115 |
| H9 | 0.115 | H9 | 0.115 |
| H10 | 0.115 | H10 | 0.115 |
| H11 | 0.115 | H11 | 0.115 |
| H12 | 0.323 | H12 | 0.323 |
| H13 | 0.09 | H13 | 0.09 |
| H14 | 0.09 | H14 | 0.09 |
| H15 | 0.09 | H15 | 0.09 |
| H16 | 0.09 | H16 | 0.09 |
| H17 | 0.09 | H17 | 0.09 |
| H18 | 0.09 | H18 | 0.09 |
| H19 | 0.09 | H19 | 0.09 |
| H20 | 0.115 | H20 | 0.115 |
| H21 | 0.115 | H21 | 0.115 |
| H22 | 0.115 | H22 | 0.115 |
| H23 | 0.115 | H23 | 0.115 |
| H24 | 0.323 | H24 | 0.323 |
| H25 | 0.09 | H25 | 0.09 |
| H26 | 0.09 | H26 | 0.09 |
| H27 | 0.09 | H27 | 0.09 |
| HN | 0.318 | | |

Table DSI. Partial atomic charges for cationic (DOFC) and neutral (DOFN) dofetilide models.

(Optimized charge values are shown by asterisk)

| Drug | Total Charge | Drug | Total Charge | Drug | Total Charge |
|------|--------------|------|--------------|------|--------------|
| MOXC | 1.00 | MOXZ | 0.00 | MOX0 | 0.00 |
| C1 | -0.1 | C1 | -0.1 | F * | -0.145 |
| O1 | -0.391 | O1 | -0.391 | O1 | -0.391 |
| C2* | 0.297 | C2* | 0.275 | O2* | -0.425 |
| C3* | 0.395 | C3* | 0.296 | O3 | -0.513 |
| C4 | -0.234 | C4* | -0.386 | O4 | -0.437 |
| C5 | 0.021 | C5 | 0.04 | N1* | -0.094 |
| C6* | 0.006 | C6* | 0.061 | N2* | -0.761 |
| C7* | 0.118 | C7* | 0.278 | N3* | -0.177 |
| N1* | -0.507 | N1* | -0.69 | C1* | -0.086 |
| C8* | 0.169 | C8* | 0.222 | C2* | 0.018 |
| C9* | -0.114 | C9* | -0.216 | C3* | -0.019 |
| H1 | 0.09 | H1 | 0.09 | C4* | 0.096 |
| C10 | -0.19 | C10 | -0.19 | C5 | -0.178 |
| C11 | -0.161 | C11 | -0.177 | C6* | 0.094 |
| C12* | 0.17 | C12* | 0.127 | C7* | -0.171 |
| N2* | -0.344 | N2* | -0.409 | C8 | -0.18 |
| C13* | 0.245 | C13* | 0.342 | C9 | -0.18 |
| H2 | 0.09 | H2 | 0.09 | C10* | 0.078 |
| C14* | -0.026 | C14* | -0.045 | C11* | -0.15 |
| F1* | -0.282 | F1* | -0.258 | C12* | 0.027 |
| C15* | 0.304 | C15* | 0.469 | C13* | 0.216 |
| O2* | -0.449 | O2* | -0.609 | C14* | 0.097 |
| C16* | 0.118 | C16* | -0.203 | C15* | -0.135 |
| C17 | 0.095 | C17* | 0.187 | C16 | 0.016 |
| N3* | -0.402 | N3* | -0.41 | C17 | 0.04 |
| C18* | 0.099 | C18* | 0.092 | C18* | 0.261 |
| C19 | -0.18 | C19 | -0.18 | C19* | 0.026 |
| C20 | -0.18 | C20 | -0.18 | C20* | 0.314 |
| C21* | 0.32 | C21* | 0.805 | C21 | -0.1 |
| O3 | -0.513 | O3 | -0.76 | H1 | 0.09 |
| O4 | -0.437 | O4 | -0.76 | H2 | 0.09 |
| H3 | 0.09 | H3 | 0.09 | H3 | 0.429 |
| H4 | 0.09 | H4 | 0.09 | H4 | 0.342 |
| H5 | 0.09 | H5 | 0.09 | H5 | 0.09 |
| H6 | 0.161 | H6 | 0.177 | H6 | 0.09 |
| H7 | 0.09 | H7 | 0.09 | H7 | 0.09 |
| H8 | 0.09 | H8 | 0.09 | H8 | 0.09 |
| H9 | 0.09 | H9 | 0.09 | H9 | 0.09 |
| H10 | 0.09 | H10 | 0.09 | H10 | 0.09 |
| H11 | 0.09 | H11 | 0.09 | H11 | 0.09 |
| H12 | 0.09 | H12 | 0.09 | H12 | 0.09 |
| H13 | 0.09 | H13 | 0.09 | H13 | 0.09 |
| H14 | 0.09 | H14 | 0.09 | H14 | 0.09 |
| H15 | 0.329 | H15* | 0.341 | H15 | 0.09 |
| H16 | 0.09 | H16 | 0.09 | H16 | 0.09 |
| H17 | 0.09 | H17 | 0.09 | H17 | 0.09 |
| H18 | 0.105 | H18 | 0.111 | H18 | 0.09 |
| H19 | 0.09 | H19 | 0.09 | H19 | 0.09 |
| H20 | 0.09 | H20 | 0.09 | H20 | 0.111 |
| H21 | 0.09 | H21 | 0.09 | H21 | 0.177 |
| H22 | 0.09 | H22 | 0.09 | H22 | 0.09 |
| H23 | 0.09 | H23 | 0.09 | H23 | 0.09 |
| H24 | 0.329 | H24 | 0.341 | H24 | 0.09 |
| H25 | 0.429 | | | | |

Table DSII. Partial atomic charges for cationic (MOXC), zwitterionic (MOXZ), and neutral (MOX0) moxifloxacin models.

(Optimized charge values are shown by asterisk)

Table DSIII. Gas-phase neutral dofetilide (DOFN) - water interactions.

| | Water interaction distance (Å) | | | Water interaction energy (kcal/mol) | | |
|-------------|--------------------------------|------|------------|-------------------------------------|--------|------------|
| | QM | MM | Difference | QM | MM | Difference |
| N1 | 3.077 | 3.13 | 0.05 | -5.839 | -6.017 | -0.178 |
| N2 | 3.205 | 3.31 | 0.10 | -3.572 | -2.582 | 0.990 |
| N3 | 3.26 | 3.21 | -0.05 | -4.137 | -5.059 | -0.922 |
| O1 | 3.003 | 2.95 | -0.05 | -6.363 | -6.377 | -0.014 |
| O1 | 2.986 | 2.94 | -0.05 | -6.34 | -6.396 | -0.056 |
| O1 | 3.049 | 2.95 | -0.10 | -5.384 | -5.409 | -0.025 |
| O2 | 3.053 | 2.95 | -0.10 | -6.019 | -6.677 | -0.658 |
| O2 | 2.977 | 2.93 | -0.05 | -6.822 | -7.164 | -0.342 |
| O2 | 3.065 | 2.97 | -0.10 | -5.515 | -5.753 | -0.238 |
| O3 | 3.01 | 2.91 | -0.10 | -4.847 | -5.452 | -0.605 |
| O4 | 3.039 | 2.99 | -0.05 | -5.764 | -6.268 | -0.504 |
| O4 | 2.987 | 2.94 | -0.05 | -6.254 | -6.867 | -0.613 |
| O4 | 3.018 | 2.97 | -0.05 | -5.721 | -6.023 | -0.302 |
| O5 | 3.045 | 3.00 | -0.05 | -5.844 | -6.039 | -0.195 |
| O5 | 2.974 | 2.92 | -0.05 | -6.761 | -6.833 | -0.072 |
| O5 | 3.044 | 2.94 | -0.10 | -5.713 | -5.814 | -0.101 |
| H8 | 2.515 | 2.82 | 0.30 | -2.324 | -2.02 | 0.304 |
| H9 | 3.28 | 3.68 | 0.40 | -3.321 | -2.796 | 0.525 |
| H10 | 2.375 | 2.78 | 0.40 | -3.877 | -3.12 | 0.757 |
| H11 | 2.504 | 2.80 | 0.30 | -3.24 | -2.828 | 0.412 |
| H12 | 1.989 | 2.04 | 0.05 | -7.541 | -6.935 | 0.606 |
| H20 | 2.512 | 2.86 | 0.35 | -1.634 | -0.797 | 0.837 |
| H21 | 2.538 | 2.94 | 0.40 | -0.309 | 0.884 | 1.193 |
| H22 | 2.43 | 2.78 | 0.35 | -4.092 | -3.323 | 0.769 |
| H24 | 2.012 | 2.01 | 0.00 | -7.342 | -6.946 | 0.396 |
| RMSE | 0.211 | | | 0.527 | | |

Table DSIV. Gas-phase cationic dofetilide (DOFC) – water interactions.

| | Water interaction distance (Å) | | | Water interaction energy (kcal/mol) | | |
|-------------|--------------------------------|------|------------|-------------------------------------|---------|------------|
| | QM | MM | Difference | QM | MM | Difference |
| N2 | 3.237 | 3.29 | 0.05 | -2.26 | -1.975 | 0.285 |
| N3 | 3.148 | 3.20 | 0.05 | -3.155 | -2.233 | 0.922 |
| O1 | 3.086 | 2.99 | -0.10 | -8.934 | -9.013 | -0.079 |
| O1 | 2.973 | 2.92 | -0.05 | -6.092 | -6.75 | -0.658 |
| O1 | 3.067 | 2.97 | -0.10 | -7.698 | -7.875 | -0.177 |
| O2 | 3.037 | 2.99 | -0.05 | -6.907 | -7.176 | -0.269 |
| O2 | 3.006 | 2.96 | -0.05 | -5.22 | -5.922 | -0.702 |
| O2 | 3.081 | 2.98 | -0.10 | -4.433 | -5.068 | -0.635 |
| O3 | 3.114 | 2.91 | -0.20 | -1.865 | -3.155 | -1.290 |
| O4 | 7.296 | 7.30 | 0.00 | -0.851 | -0.618 | 0.233 |
| O5 | 4.481 | 4.88 | 0.40 | -3.364 | -1.704 | 1.660 |
| O5 | 3.04 | 2.94 | -0.10 | -3.51 | -4.124 | -0.614 |
| O5 | 3.092 | 2.94 | -0.15 | -3.383 | -4.898 | -1.515 |
| H8 | 2.529 | 2.83 | 0.30 | -5.074 | -4.223 | 0.851 |
| H9 | 2.839 | 3.24 | 0.40 | -5.555 | -3.318 | 2.237 |
| H10 | 2.356 | 2.76 | 0.40 | -5.84 | -4.289 | 1.551 |
| H12 | 1.981 | 2.03 | 0.05 | -9.487 | -7.887 | 1.600 |
| H20 | 2.331 | 2.73 | 0.40 | -6.588 | -4.648 | 1.940 |
| H21 | 2.356 | 2.76 | 0.40 | -6.025 | -3.831 | 2.194 |
| H22 | 2.207 | 2.61 | 0.40 | -7.815 | -5.072 | 2.743 |
| H23 | 2.298 | 2.70 | 0.40 | -7.732 | -5.814 | 1.918 |
| H24 | 1.943 | 1.99 | 0.05 | -12.94 | -10.529 | 2.411 |
| RMSE | 0.246 | | | 1.443 | | |

Table DSV. Gas-phase zwitterionic moxifloxacin (MOXZ) – water interactions.

| | Water interaction distance (Å) | | | Water interaction energy (kcal/mol) | | |
|-------------|--------------------------------|------|--------|-------------------------------------|--------|--------|
| | QM | MM | Delta | QM | MM | Delta |
| N3 | 3.451 | 3.30 | -0.150 | -6.045 | -7.362 | -1.317 |
| O1 | 3.013 | 2.91 | -0.100 | -6.199 | -6.409 | -0.21 |
| O2 | 2.913 | 2.86 | -0.050 | -11.398 | -13.53 | -2.132 |
| O3 | 2.797 | 2.80 | 0.000 | -15.015 | -14.01 | 1.005 |
| O4 | 2.829 | 2.83 | 0.000 | -14.72 | -14.64 | 0.08 |
| H1 | 2.433 | 2.78 | 0.350 | -5.325 | -4.465 | 0.86 |
| H2 | 2.318 | 2.67 | 0.350 | -7.508 | -7.329 | 0.179 |
| H3 | 6.061 | 6.26 | 0.200 | -3.634 | -3.073 | 0.561 |
| H4 | 3.266 | 3.47 | 0.200 | -3.118 | -2.912 | 0.206 |
| H5 | 2.569 | 2.87 | 0.300 | -2.865 | -1.924 | 0.941 |
| H6 | 2.219 | 2.57 | 0.350 | -2.191 | -1.147 | 1.044 |
| H7 | 3.021 | 3.42 | 0.400 | -4.813 | -3.817 | 0.996 |
| H8 | 2.524 | 2.77 | 0.250 | -4.191 | -4.296 | -0.105 |
| H9 | 2.467 | 2.82 | 0.350 | -4.823 | -3.801 | 1.022 |
| H10 | 2.541 | 2.84 | 0.300 | -4.946 | -4.429 | 0.517 |
| H11 | 3.713 | 4.11 | 0.400 | -4.112 | -3.784 | 0.328 |
| H12 | 2.426 | 2.78 | 0.350 | -5.159 | -4.063 | 1.096 |
| H13 | 2.252 | 2.65 | 0.400 | -8.578 | -7.249 | 1.329 |
| H14 | 2.33 | 2.73 | 0.400 | -7.392 | -6.804 | 0.588 |
| H16 | 2.477 | 2.78 | 0.300 | -4.51 | -3.864 | 0.646 |
| H17 | 2.331 | 2.73 | 0.400 | -9.584 | -8.85 | 0.734 |
| H18 | 0.282 | 0.68 | 0.400 | 0.587 | 1.797 | 1.21 |
| H19 | 2.936 | 3.04 | 0.100 | -0.2 | -0.037 | 0.163 |
| H21 | 2.613 | 2.91 | 0.300 | -1.411 | -0.59 | 0.821 |
| H22 | 2.784 | 2.93 | 0.150 | -2.16 | -1.418 | 0.742 |
| H23 | 2.504 | 2.85 | 0.350 | -2.829 | -2.334 | 0.495 |
| H24 | 1.871 | 1.97 | 0.100 | -17.35 | -17.13 | 0.22 |
| RMSE | 0.290 | | | 0.864 | | |

Table DSVI. Gas-phase cationic moxifloxacin (MOXC) – water interactions.

| | Water interaction distance (Å) | | | Water interaction energy (kcal/mol) | | |
|-------------|--------------------------------|------|--------|-------------------------------------|--------|--------|
| | QM | MM | Delta | QM | MM | Delta |
| N3 | 3.811 | 3.41 | -0.400 | -1.131 | -2.43 | -1.296 |
| 01 | 3.087 | 3.09 | 0.000 | -2.324 | -2.64 | -0.315 |
| 02 | 4.746 | 4.35 | -0.400 | -2.647 | -3.07 | -0.418 |
| 02 | 3.723 | 3.52 | -0.200 | -5.188 | -6.24 | -1.053 |
| 02 | 3.023 | 2.92 | -0.100 | -6.031 | -7.03 | -0.994 |
| 03 | 3.133 | 3.08 | -0.050 | -4.107 | -4.92 | -0.813 |
| 04 | 3.091 | 2.99 | -0.100 | -9.64 | -9.60 | 0.037 |
| 04 | 3.09 | 2.99 | -0.100 | -5.311 | -5.88 | -0.564 |
| 04 | 3.074 | 2.92 | -0.150 | -5.62 | -6.89 | -1.273 |
| H1 | 2.39 | 2.74 | 0.350 | -7.217 | -6.61 | 0.608 |
| H2 | 2.266 | 2.67 | 0.400 | -9.652 | -8.86 | 0.796 |
| H4 | 2.647 | 2.85 | 0.200 | -6.009 | -5.27 | 0.742 |
| H5 | 2.454 | 2.80 | 0.350 | -5.801 | -3.81 | 1.987 |
| H6 | 2.196 | 2.55 | 0.350 | -4.854 | -3.32 | 1.536 |
| H7 | 2.91 | 3.31 | 0.400 | -6.713 | -4.76 | 1.949 |
| H8 | 2.499 | 2.75 | 0.250 | -6.254 | -6.30 | -0.048 |
| H9 | 2.418 | 2.77 | 0.350 | -6.332 | -5.37 | 0.958 |
| H10 | 2.468 | 2.77 | 0.300 | -6.686 | -6.15 | 0.541 |
| H11 | 3.267 | 3.67 | 0.400 | -4.74 | -3.21 | 1.531 |
| H12 | 2.36 | 2.76 | 0.400 | -6.914 | -5.60 | 1.314 |
| H13 | 2.238 | 2.64 | 0.400 | -9.438 | -8.12 | 1.321 |
| H14 | 2.285 | 2.69 | 0.400 | -9.318 | -8.51 | 0.805 |
| H16 | 2.436 | 2.74 | 0.300 | -6.831 | -6.08 | 0.755 |
| H17 | 2.325 | 2.73 | 0.400 | -9.275 | -8.39 | 0.883 |
| H18 | 2.213 | 2.56 | 0.350 | -7.808 | -6.02 | 1.792 |
| H19 | 2.443 | 2.84 | 0.400 | -3.756 | -2.358 | 1.398 |
| H21 | 2.437 | 2.84 | 0.400 | -4.805 | -2.951 | 1.854 |
| H22 | 2.634 | 2.93 | 0.300 | -5.4 | -3.401 | 1.999 |
| H23 | 2.488 | 2.89 | 0.400 | -4.997 | -4.293 | 0.704 |
| H24 | 1.87 | 1.97 | 0.100 | -18.08 | -17.68 | 0.400 |
| H25 | 1.999 | 1.95 | -0.050 | -7.705 | -8.912 | -1.207 |
| RMSE | 0.312 | | | 1.165 | | |

Table DSVII. Gas-phase neutral moxifloxacin (MOX0) – water interactions.

| | Water interaction distance (Å) | | | Water interaction energy (kcal/mol) | | |
|-------------|--------------------------------|-------|-------|-------------------------------------|--------|--------|
| | QM | MM | Delta | QM | MM | Delta |
| N3 | 5.521 | 5.271 | -0.25 | -0.197 | -0.79 | -0.595 |
| O1 | 5.114 | 5.014 | -0.1 | -1.026 | -1.23 | -0.203 |
| O2 | 4.489 | 4.889 | 0.4 | -1.315 | -0.85 | 0.466 |
| O2 | 4.481 | 4.481 | 0 | -6.081 | -5.58 | 0.501 |
| O2 | 2.972 | 2.922 | -0.05 | -7.7 | -7.78 | -0.075 |
| O3 | 5.118 | 4.718 | -0.4 | -1.264 | -1.61 | -0.346 |
| O4 | 2.914 | 2.864 | -0.05 | -7.85 | -7.29 | 0.563 |
| O4 | 4.218 | 4.268 | 0.05 | -7.061 | -6.27 | 0.787 |
| O4 | 3.011 | 2.911 | -0.1 | -7.089 | -7.37 | -0.284 |
| H1 | 2.686 | 2.936 | 0.25 | -1.424 | -0.77 | 0.658 |
| H2 | 2.662 | 2.862 | 0.2 | -1.862 | -1.41 | 0.454 |
| H3 | 1.852 | 1.952 | 0.1 | -8.475 | -8.02 | 0.453 |
| H4 | 2.255 | 2.305 | 0.05 | -3.717 | -3.67 | 0.052 |
| H5 | 2.604 | 2.854 | 0.25 | -1.395 | -0.97 | 0.428 |
| H6 | 2.649 | 2.849 | 0.2 | -1.719 | -1.25 | 0.465 |
| H7 | 3.884 | 4.134 | 0.25 | -2.494 | -1.88 | 0.614 |
| H8 | 2.812 | 2.862 | 0.05 | -2.287 | -2.35 | -0.061 |
| H9 | 2.788 | 2.888 | 0.1 | -1.467 | -1.34 | 0.130 |
| H11 | 2.394 | 2.794 | 0.4 | -3.805 | -3.16 | 0.641 |
| H12 | 2.783 | 2.983 | 0.2 | -0.842 | -0.35 | 0.494 |
| H13 | 2.846 | 2.896 | 0.05 | -1.231 | -1.47 | -0.239 |
| H14 | 2.615 | 2.915 | 0.3 | -2.14 | -1.60 | 0.545 |
| H17 | 2.644 | 2.894 | 0.250 | -2.027 | -1.544 | 0.483 |
| H19 | 2.766 | 2.916 | 0.15 | -1.304 | -1.354 | -0.050 |
| H20 | 2.36 | 2.71 | 0.35 | -2.321 | -1.977 | 0.344 |
| H21 | 2.24 | 2.54 | 0.3 | -1.66 | -1.432 | 0.228 |
| H22 | 2.553 | 2.903 | 0.35 | -2.51 | -1.555 | 0.955 |
| H24 | 2.593 | 2.943 | 0.35 | -3.35 | -2.661 | 0.689 |
| RMSE | 0.225 | | | 0.475 | | |

Table DSVIII. Optimized force field topology and parameters for cationic dofetilide

Topology file: top_dof1.rtf

```
* Initial topologies generated by
* CHARMM General Force Field (CGenFF) program version 1.0.0
* For use with CGenFF version 3.0.1
36 1

! "penalty" is the highest penalty score of the associated parameters.
! Penalties lower than 10 indicate the analogy is fair; penalties between 10
! and 50 mean some basic validation is recommended; penalties higher than
! 50 indicate poor analogy and mandate extensive validation/optimization.
```

```
=====
! Dofetilide(+)
=====
```

| RESI | DOF1 | 1.000 |
|-------|------|-----------------------|
| GROUP | | ! CHARGE CH_PENALTY |
| ATOM | C1 | CG334 0.149 |
| ATOM | N1 | NG3P1 -0.006 |
| ATOM | C2 | CG324 -0.495 |
| ATOM | C3 | CG321 -0.250 |
| ATOM | C4 | CG2R61 0.310 |
| ATOM | C5 | CG2R61 -0.115 ! 8.929 |
| ATOM | C6 | CG2R61 -0.111 ! 0.000 |
| ATOM | C7 | CG2R61 0.219 ! 0.000 |
| ATOM | C8 | CG2R61 -0.111 ! 0.000 |
| ATOM | C9 | CG2R61 -0.115 ! 8.929 |
| ATOM | N2 | NG311 -0.466 ! 0.000 |
| ATOM | S1 | SG3O2 0.439 ! 0.000 |
| ATOM | O1 | OG2P1 -0.384 ! 0.000 |
| ATOM | O2 | OG2P1 -0.384 ! 0.000 |
| ATOM | C10 | CG331 -0.019 ! 0.000 |
| ATOM | C11 | CG324 0.162 ! 1.152 |
| ATOM | C12 | CG321 -0.014 ! 1.152 |
| ATOM | O3 | OG3O1 -0.391 ! 0.861 |
| ATOM | C13 | CG2R61 0.219 ! 0.850 |
| ATOM | C14 | CG2R61 -0.113 ! 0.000 |
| ATOM | C15 | CG2R61 -0.115 ! 0.000 |
| ATOM | C16 | CG2R61 0.219 ! 0.000 |
| ATOM | C17 | CG2R61 -0.115 ! 0.000 |
| ATOM | C18 | CG2R61 -0.113 ! 0.000 |
| ATOM | N3 | NG311 -0.466 ! 0.000 |
| ATOM | S2 | SG3O2 0.439 ! 0.000 |
| ATOM | O4 | OG2P1 -0.384 ! 0.000 |

| | | | | |
|------|-----|-------|----------|-------|
| ATOM | O5 | OG2P1 | -0.384 ! | 0.000 |
| ATOM | C19 | CG331 | -0.019 ! | 0.000 |
| ATOM | H1 | HGA3 | 0.090 ! | 0.000 |
| ATOM | H2 | HGA3 | 0.090 ! | 0.000 |
| ATOM | H3 | HGA3 | 0.090 ! | 0.000 |
| ATOM | H4 | HGA2 | 0.090 ! | 2.455 |
| ATOM | H5 | HGA2 | 0.090 ! | 2.455 |
| ATOM | H6 | HGA2 | 0.090 ! | 0.000 |
| ATOM | H7 | HGA2 | 0.090 ! | 0.000 |
| ATOM | H8 | HGR61 | 0.115 ! | 0.000 |
| ATOM | H9 | HGR61 | 0.115 ! | 0.000 |
| ATOM | H10 | HGR61 | 0.115 ! | 0.000 |
| ATOM | H11 | HGR61 | 0.115 ! | 0.000 |
| ATOM | H12 | HGP1 | 0.323 ! | 0.000 |
| ATOM | H13 | HGA3 | 0.090 ! | 0.000 |
| ATOM | H14 | HGA3 | 0.090 ! | 0.000 |
| ATOM | H15 | HGA3 | 0.090 ! | 0.000 |
| ATOM | H16 | HGA2 | 0.090 ! | 0.075 |
| ATOM | H17 | HGA2 | 0.090 ! | 0.075 |
| ATOM | H18 | HGA2 | 0.090 ! | 0.000 |
| ATOM | H19 | HGA2 | 0.090 ! | 0.000 |
| ATOM | H20 | HGR61 | 0.115 ! | 0.000 |
| ATOM | H21 | HGR61 | 0.115 ! | 0.000 |
| ATOM | H22 | HGR61 | 0.115 ! | 0.000 |
| ATOM | H23 | HGR61 | 0.115 ! | 0.000 |
| ATOM | H24 | HGP1 | 0.323 ! | 0.000 |
| ATOM | H25 | HGA3 | 0.090 ! | 0.000 |
| ATOM | H26 | HGA3 | 0.090 ! | 0.000 |
| ATOM | H27 | HGA3 | 0.090 ! | 0.000 |
| ATOM | HN | HGP2 | 0.318 ! | 0.000 |

| | | |
|------|----|-----|
| BOND | C1 | N1 |
| BOND | C1 | H1 |
| BOND | C1 | H2 |
| BOND | C1 | H3 |
| BOND | N1 | C2 |
| BOND | N1 | C11 |
| BOND | N1 | HN |
| BOND | C2 | C3 |
| BOND | C2 | H4 |
| BOND | C2 | H5 |
| BOND | C3 | C4 |
| BOND | C3 | H6 |
| BOND | C3 | H7 |
| BOND | C4 | C9 |
| BOND | C4 | C5 |
| BOND | C5 | C6 |
| BOND | C5 | H8 |
| BOND | C6 | C7 |

```
BOND C6 H9
BOND C7 C8
BOND C7 N2
BOND C8 C9
BOND C8 H10
BOND C9 H11
BOND N2 S1
BOND N2 H12
BOND S1 O1
BOND S1 O2
BOND S1 C10
BOND C10 H13
BOND C10 H14
BOND C10 H15
BOND C11 C12
BOND C11 H16
BOND C11 H17
BOND C12 O3
BOND C12 H18
BOND C12 H19
BOND O3 C13
BOND C13 C18
BOND C13 C14
BOND C14 C15
BOND C14 H20
BOND C15 C16
BOND C15 H21
BOND C16 C17
BOND C16 N3
BOND C17 C18
BOND C17 H22
BOND C18 H23
BOND N3 S2
BOND N3 H24
BOND S2 O4
BOND S2 O5
BOND S2 C19
BOND C19 H25
BOND C19 H26
BOND C19 H27
```

```
END
```

Parameter file: par_dof1.prm

* Initial parameters generated by analogy by

```

* CHARMM General Force Field (CGenFF) program version 1.0.0
* For use with CGenFF version 3.0.1

! Penalties lower than 10 indicate the analogy is fair; penalties between 10
! and 50 mean some basic validation is recommended; penalties higher than
! 50 indicate poor analogy and mandate extensive validation/optimization.

!=====
! Dofetilide(+)
!=====

BONDS

ANGLES
CG2R61 CG321 CG324      51.80    107.50 ! ZINC00 , from CG2R61 CG321 CG314, penalty= 0.6
CG324 CG321 OG301      75.70    110.10 ! ZINC00 , from CG324 CG321 OG302, penalty= 0.5

DIHEDRALS
CG2R61 CG2R61 CG321 CG324      0.2300  2    180.00 ! ZINC00 , from CG2R61 CG2R61 CG321 CG314,
penalty= 0.6
CG2R61 CG321 CG324 NG3P1      0.2000  3     0.00 ! ZINC00 , from NG3P3 CG314 CG321 CG2R61,
penalty= 5.5
CG2R61 CG321 CG324 HGA2      0.0400  3     0.00 ! ZINC00 , from CG2R61 CG321 CG321 HGA2,
penalty= 1
OG301 CG321 CG324 NG3P1      3.3000  1    180.00 ! ZINC00 , from OG302 CG321 CG324 NG3P0,
penalty= 1.7
OG301 CG321 CG324 NG3P1      -0.4000  3    180.00 ! ZINC00 , from OG302 CG321 CG324 NG3P0,
penalty= 1.7
OG301 CG321 CG324 HGA2      0.1900  3     0.00 ! ZINC00 , from OG301 CG321 CG321 HGA2,
penalty= 1
CG324 CG321 OG301 CG2R61      2.9990  1    180.00
CG324 CG321 OG301 CG2R61      2.4130  2    180.00
CG324 CG321 OG301 CG2R61      2.3380  3     0.00

```

Table DSIX. Optimized force field topology and parameters for neutral dofetilide

Topology file: top_dof0.rtf

```
* Initial topologies generated by
* CHARMM General Force Field (CGenFF) program version 1.0.0
* For use with CGenFF version 3.0.1
36 1

! "penalty" is the highest penalty score of the associated parameters.
! Penalties lower than 10 indicate the analogy is fair; penalties between 10
! and 50 mean some basic validation is recommended; penalties higher than
! 50 indicate poor analogy and mandate extensive validation/optimization.

!=====
! Dofetilide(0)
!=====

RESI DOF0      0.000
GROUP          ! CHARGE   CH_PENALTY
ATOM C1        CG331    -0.392
ATOM N1        NG301    -0.401
ATOM C2        CG321    0.240
ATOM C3        CG321    -0.559
ATOM C4        CG2R61   0.296
ATOM C5        CG2R61   -0.115 !
ATOM C6        CG2R61   -0.111 !
ATOM C7        CG2R61   0.219 !
ATOM C8        CG2R61   -0.111 !
ATOM C9        CG2R61   -0.115 !
ATOM N2        NG311    -0.466 !
ATOM S1        SG3O2    0.439 !
ATOM O1        OG2P1    -0.384 !
ATOM O2        OG2P1    -0.384 !
ATOM C10       CG331    -0.019 !
ATOM C11       CG321    0.010
ATOM C12       CG321    -0.021 !
ATOM O3        OG301    -0.390 !
ATOM C13       CG2R61   0.219 !
ATOM C14       CG2R61   -0.113 !
ATOM C15       CG2R61   -0.115 !
ATOM C16       CG2R61   0.219 !
ATOM C17       CG2R61   -0.115 !
ATOM C18       CG2R61   -0.113 !
ATOM N3        NG311    -0.466 !
```

| | | | | |
|------|-----|-------|----------|-------|
| ATOM | S2 | SG3O2 | 0.439 ! | 0.000 |
| ATOM | O4 | OG2P1 | -0.384 ! | 0.000 |
| ATOM | O5 | OG2P1 | -0.384 ! | 0.000 |
| ATOM | C19 | CG331 | -0.019 ! | 0.000 |
| ATOM | H1 | HGA3 | 0.090 ! | 3.536 |
| ATOM | H2 | HGA3 | 0.090 ! | 3.536 |
| ATOM | H3 | HGA3 | 0.090 ! | 3.536 |
| ATOM | H4 | HGA2 | 0.090 ! | 3.536 |
| ATOM | H5 | HGA2 | 0.090 ! | 3.536 |
| ATOM | H6 | HGA2 | 0.090 ! | 0.480 |
| ATOM | H7 | HGA2 | 0.090 ! | 0.480 |
| ATOM | H8 | HGR61 | 0.115 ! | 0.000 |
| ATOM | H9 | HGR61 | 0.115 ! | 0.000 |
| ATOM | H10 | HGR61 | 0.115 ! | 0.000 |
| ATOM | H11 | HGR61 | 0.115 ! | 0.000 |
| ATOM | H12 | HGP1 | 0.323 ! | 0.000 |
| ATOM | H13 | HGA3 | 0.090 ! | 0.000 |
| ATOM | H14 | HGA3 | 0.090 ! | 0.000 |
| ATOM | H15 | HGA3 | 0.090 ! | 0.000 |
| ATOM | H16 | HGA2 | 0.090 ! | 3.536 |
| ATOM | H17 | HGA2 | 0.090 ! | 3.536 |
| ATOM | H18 | HGA2 | 0.090 ! | 0.480 |
| ATOM | H19 | HGA2 | 0.090 ! | 0.480 |
| ATOM | H20 | HGR61 | 0.115 ! | 0.000 |
| ATOM | H21 | HGR61 | 0.115 ! | 0.000 |
| ATOM | H22 | HGR61 | 0.115 ! | 0.000 |
| ATOM | H23 | HGR61 | 0.115 ! | 0.000 |
| ATOM | H24 | HGP1 | 0.323 ! | 0.000 |
| ATOM | H25 | HGA3 | 0.090 ! | 0.000 |
| ATOM | H26 | HGA3 | 0.090 ! | 0.000 |
| ATOM | H27 | HGA3 | 0.090 ! | 0.000 |

| | | |
|------|----|-----|
| BOND | C1 | N1 |
| BOND | C1 | H1 |
| BOND | C1 | H2 |
| BOND | C1 | H3 |
| BOND | N1 | C2 |
| BOND | N1 | C11 |
| BOND | C2 | C3 |
| BOND | C2 | H4 |
| BOND | C2 | H5 |
| BOND | C3 | C4 |
| BOND | C3 | H6 |
| BOND | C3 | H7 |
| BOND | C4 | C9 |
| BOND | C4 | C5 |
| BOND | C5 | C6 |
| BOND | C5 | H8 |
| BOND | C6 | C7 |

BOND C6 H9
BOND C7 C8
BOND C7 N2
BOND C8 C9
BOND C8 H10
BOND C9 H11
BOND N2 S1
BOND N2 H12
BOND S1 O1
BOND S1 O2
BOND S1 C10
BOND C10 H13
BOND C10 H14
BOND C10 H15
BOND C11 C12
BOND C11 H16
BOND C11 H17
BOND C12 O3
BOND C12 H18
BOND C12 H19
BOND O3 C13
BOND C13 C18
BOND C13 C14
BOND C14 C15
BOND C14 H20
BOND C15 C16
BOND C15 H21
BOND C16 C17
BOND C16 N3
BOND C17 C18
BOND C17 H22
BOND C18 H23
BOND N3 S2
BOND N3 H24
BOND S2 O4
BOND S2 O5
BOND S2 C19
BOND C19 H25
BOND C19 H26
BOND C19 H27

END

Parameter file: par_dof0.prm

```
* Initial parameters generated by analogy by
* CHARMM General Force Field (CGenFF) program version 1.0.0
* For use with CGenFF version 3.0.1

! Penalties lower than 10 indicate the analogy is fair; penalties between 10
! and 50 mean some basic validation is recommended; penalties higher than
! 50 indicate poor analogy and mandate extensive validation/optimization.

BONDS
CG321 NG301    263.00      1.4740 ! ZINC00 , from CG321 NG311, penalty= 5
CG331 NG301    255.00      1.4630 ! ZINC00 , from CG331 NG311, penalty= 5

ANGLES
CG321 CG321 NG301    43.70     112.20 ! ZINC00 , from CG331 CG321 NG311, penalty= 1.5
NG301 CG321 HGA2      32.40     109.50   50.00     2.13000 ! ZINC00 , from NG311 CG321 HGA2,
penalty= 0.6
NG301 CG331 HGA3      30.50     109.70   50.00     2.14000 ! ZINC00 , from NG311 CG331 HGA3,
penalty= 0.6
CG321 NG301 CG321    42.452    104.717
CG321 NG301 CG331    59.281    106.264

DIHEDRALS
CG2R61 CG321 CG321 NG301      0.3650  3      0.00
NG301 CG321 CG321 OG301      3.0000  1     180.00
NG301 CG321 CG321 OG301      0.9850  2      0.00
NG301 CG321 CG321 HGA2      0.1600  3      0.00 ! ZINC00 , from NG311 CG321 CG331 HGA3,
penalty= 6.6
CG321 CG321 NG301 CG321      1.7910  1      0.00
CG321 CG321 NG301 CG321      0.7430  2      0.00
CG321 CG321 NG301 CG321      0.1570  3      0.00
CG321 CG321 NG301 CG331      0.5940  1      0.00
CG321 CG321 NG301 CG331      0.1350  2      0.00
CG321 CG321 NG301 CG331      1.0280  3      0.00
HGA2 CG321 NG301 CG321      0.3570  3     180.00
HGA2 CG321 NG301 CG331      0.7040  3      0.00
HGA3 CG331 NG301 CG321      0.8520  3     180.00
```

Table DSX. Optimized force field topology and parameters for zwitterionic moxifloxacin (CHARMM stream including CGENFF parameters)

```

* Toppar stream file generated by
*
read rtf card append
* Initial topologies generated by
* CHARMM General Force Field (CGenFF) program version 1.0.0
* For use with CGenFF version 3.0.1
*
36 1

=====
! Moxifloxacin - Zwitterionic
=====

RESI MOX      0.000
GROUP          ! CHARGE   CH_PENALTY
ATOM C1      CG331  -0.100 !    0.025
ATOM O1      OG301  -0.391 !    2.180
ATOM C2      CG2R61  0.275
ATOM C3      CG2R62  0.296
ATOM C4      CG2R62 -0.386
ATOM C5      CG2R61  0.040 !    6.996
ATOM C6      CG2R66  0.061
ATOM C7      CG2R61  0.278
ATOM N1      NG3C51 -0.690
ATOM C8      CG3C52  0.222
ATOM C9      CG3RC1 -0.216
ATOM H1      HGA1    0.090 !    2.316
ATOM C10     CG321  -0.190 !    5.785
ATOM C11     CG321  -0.177 !    5.331
ATOM C12     CG324   0.127
ATOM N2      NG3P2  -0.409
ATOM C13     CG3RC1  0.342
ATOM H2      HGA1    0.090
ATOM C14     CG3C52 -0.045
ATOM F1      FGR1    -0.258
ATOM C15     CG2R63  0.469
ATOM O2      OG2D4  -0.609
ATOM C16     CG2R62 -0.203
ATOM C17     CG2R62  0.187
ATOM N3      NG2R61 -0.410
ATOM C18     CG3C31  0.092
ATOM C19     CG3C31 -0.180 !    7.029
ATOM C20     CG3C31 -0.180 !    7.029
ATOM C21     CG2O3   0.805
ATOM O3      OG2D2  -0.760 !
ATOM O4      OG2D2  -0.760 !
ATOM H3      HGA3    0.090 !
ATOM H4      HGA3    0.090 !
ATOM H5      HGA3    0.090 !
ATOM H6      HGR62  0.177 !
ATOM H7      HGA2    0.090 !

```

| | | | | |
|------|-----|-------|---------|-------|
| ATOM | H8 | HGA2 | 0.090 ! | 1.375 |
| ATOM | H9 | HGA2 | 0.090 ! | 0.000 |
| ATOM | H10 | HGA2 | 0.090 ! | 0.000 |
| ATOM | H11 | HGA2 | 0.090 ! | 0.000 |
| ATOM | H12 | HGA2 | 0.090 ! | 0.000 |
| ATOM | H13 | HGA2 | 0.090 ! | 0.975 |
| ATOM | H14 | HGA2 | 0.090 ! | 0.975 |
| ATOM | H15 | HGP2 | 0.341 | |
| ATOM | H16 | HGA2 | 0.090 ! | 2.637 |
| ATOM | H17 | HGA2 | 0.090 ! | 2.637 |
| ATOM | H18 | HGR62 | 0.111 ! | 6.687 |
| ATOM | H19 | HGA1 | 0.090 ! | 4.940 |
| ATOM | H20 | HGA2 | 0.090 ! | 2.500 |
| ATOM | H21 | HGA2 | 0.090 ! | 2.500 |
| ATOM | H22 | HGA2 | 0.090 ! | 2.500 |
| ATOM | H23 | HGA2 | 0.090 ! | 2.500 |
| ATOM | H24 | HGP2 | 0.341 | |

| | | |
|------|-----|-----|
| BOND | C1 | O1 |
| BOND | C1 | H3 |
| BOND | C1 | H4 |
| BOND | C1 | H5 |
| BOND | O1 | C2 |
| BOND | C2 | C7 |
| BOND | C2 | C3 |
| BOND | C3 | N3 |
| BOND | C3 | C4 |
| BOND | C4 | C5 |
| BOND | C4 | C15 |
| BOND | C5 | C6 |
| BOND | C5 | H6 |
| BOND | C6 | C7 |
| BOND | C6 | F1 |
| BOND | C7 | N1 |
| BOND | N1 | C14 |
| BOND | N1 | C8 |
| BOND | C8 | C9 |
| BOND | C8 | H7 |
| BOND | C8 | H8 |
| BOND | C9 | H1 |
| BOND | C9 | C13 |
| BOND | C9 | C10 |
| BOND | C10 | C11 |
| BOND | C10 | H9 |
| BOND | C10 | H10 |
| BOND | C11 | C12 |
| BOND | C11 | H11 |
| BOND | C11 | H12 |
| BOND | C12 | N2 |
| BOND | C12 | H13 |
| BOND | C12 | H14 |
| BOND | N2 | C13 |
| BOND | N2 | H15 |
| BOND | N2 | H24 |
| BOND | C13 | H2 |
| BOND | C13 | C14 |

```

BOND C14 H16
BOND C14 H17
BOND C15 O2
BOND C15 C16
BOND C16 C17
BOND C16 C21
BOND C17 N3
BOND C17 H18
BOND N3 C18
BOND C18 C20
BOND C18 C19
BOND C18 H19
BOND C19 C20
BOND C19 H20
BOND C19 H21
BOND C20 H22
BOND C20 H23
BOND C21 O3
BOND C21 O4
IMPR C15 C4 C16 O2
IMPR C21 O4 O3 C16

END

read param card flex append
* Parameters generated by analogy by
* CHARMM General Force Field (CGenFF) program version 2.2.0
*
! Penalties lower than 10 indicate the analogy is fair; penalties between 10
! and 50 mean some basic validation is recommended; penalties higher than
! 50 indicate poor analogy and mandate extensive validation/optimization.

BONDS
CG2O3 CG2R62 200.00 1.5000 ! ***** , from CG2O3 CG2R61, PENALTY= 5
CG2O3 OG2D2 525.00 1.2600 ! PROT adm jr. 7/23/91, acetic acid
CG2R61 CG2R61 305.00 1.3750 ! PROT benzene, JES 8/25/89
CG2R61 CG2R62 394.00 1.3750 ! YTHY, 2,4(1H,3H)-quinazolininedione, isg
CG2R61 CG2R66 305.00 1.3700 ! NAMODEL difluorotoluene
CG2R61 NG3C51 406.429 1.370
CG2R61 OG301 230.00 1.3820 ! COMPDS peml
CG2R61 HGR62 340.00 1.0800 ! NA, DFT
CG2R62 CG2R62 420.00 1.3500 ! NA nad/ppi, jjp1/adm jr. 7/95
CG2R62 CG2R63 302.00 1.4030 ! NA T, adm jr. 11/97
CG2R62 NG2R61 302.00 1.3430 ! NA C, adm jr. 11/97
CG2R62 HGR62 350.00 1.0900 ! NA C,U, JWK
CG2R63 OG2D4 660.00 1.2340 ! NA U,A,G par_a4 adm jr. 10/2/91
CG2R66 FGR1 400.00 1.3580 ! NAMODEL difluorotoluene
CG321 CG321 222.50 1.5300 ! PROT alkane update, adm jr., 3/2/92
CG321 CG324 222.50 1.5300 ! FLAVOP PIP1,2,3
CG321 CG3RC1 222.50 1.5240 ! CARBOCY carbocyclic sugars
CG321 HGA2 309.00 1.1110 ! PROT alkane update, adm jr., 3/2/92
CG324 NG3P2 200.00 1.4900 ! PIP, piperidine
CG324 HGA2 284.50 1.1000 ! FLAVOP PIP1,2,3
CG331 OG301 360.00 1.4150 ! diethylether, alex
CG331 HGA3 322.00 1.1110 ! PROT alkane update, adm jr., 3/2/92

```

| | | | | | |
|--------|--------|---------|--------|---|---|
| CG3C31 | CG3C31 | 240.00 | 1.5010 | ! | PROTMOD cyclopropane |
| CG3C31 | NG2R61 | 352.828 | 1.448 | | |
| CG3C31 | HGA1 | 340.00 | 1.0830 | ! | PROTMOD cyclopropane |
| CG3C31 | HGA2 | 340.00 | 1.0830 | ! | PROTMOD cyclopropane |
| CG3C52 | CG3RC1 | 222.50 | 1.5240 | ! | CARBOCY carbocyclic sugars |
| CG3C52 | NG3C51 | 400.00 | 1.4780 | ! | PRLD, pyrrolidine; 2PRL, 2-pyrroline, kevo |
| CG3C52 | HGA2 | 307.00 | 1.1000 | ! | THF, THF neutron diffr., 5/30/06, viv |
| CG3RC1 | CG3RC1 | 222.50 | 1.5230 | ! | CARBOCY carbocyclic sugars |
| CG3RC1 | NG3P2 | 164.854 | 1.497 | | |
| CG3RC1 | HGA1 | 309.00 | 1.1110 | ! | CARBOCY carbocyclic sugars |
| NG3P2 | HGP2 | 460.00 | 1.0060 | ! | PROT AcProNH2, ProNH2, AcProNHCH3 RLD 4/23/93 |

ANGLES

| | | | | | | | | |
|--------|--------|--------|--------|---------|-------|--|---|---|
| CG2R62 | CG2O3 | OG2D2 | 40.00 | 116.00 | 50.00 | 2.35300 | ! | ***** , from CG2R61 CG2O3 OG2D2, PENALTY= 0.5 |
| OG2D2 | CG2O3 | OG2D2 | 100.00 | 128.00 | 70.00 | 2.25870 | ! | PROT adm jr. 7/23/91, correction, ACETATE (KK) |
| CG2R61 | CG2R61 | CG2R62 | 40.00 | 119.00 | 35.00 | 2.41620 | ! | YTHY, 2,4(1H,3H)-quinazolinedione, isg |
| CG2R61 | CG2R61 | CG2R66 | 40.00 | 119.00 | 35.00 | 2.41620 | ! | NAMODEL difluorotoluene |
| CG2R61 | CG2R61 | NG3C51 | 40.00 | 120.00 | ! | ***** , from CG2R61 CG2R61 NG311, PENALTY= 3 | | |
| CG2R61 | CG2R61 | OG301 | 110.00 | 120.00 | ! | BIPHENYL ANALOGS, peml | | |
| CG2R62 | CG2R61 | CG2R66 | 40.00 | 119.00 | 35.00 | 2.41620 | ! | ***** , from CG2R61 CG2R61 CG2R66, PENALTY= 0.5 |
| CG2R62 | CG2R61 | OG301 | 110.00 | 120.00 | ! | ***** , from CG2R61 CG2R61 OG301, PENALTY= 0.5 | | |
| CG2R62 | CG2R61 | HGR62 | 30.00 | 120.00 | 22.00 | 2.15250 | ! | ***** , from CG2R61 CG2R61 HGR62, PENALTY= 0.5 |
| CG2R66 | CG2R61 | NG3C51 | 40.00 | 120.00 | ! | ***** , from CG2R61 CG2R61 NG311, PENALTY= 4.5 | | |
| CG2R66 | CG2R61 | HGR62 | 30.00 | 121.50 | 22.00 | 2.15250 | ! | NAMODEL difluorotoluene |
| CG2O3 | CG2R62 | CG2R62 | 45.00 | 119.00 | ! | ***** , from CG2O3 CG2R61 CG2R61, PENALTY= 5.5 | | |
| CG2O3 | CG2R62 | CG2R63 | 45.00 | 119.00 | ! | ***** , from CG2O3 CG2R61 CG2R61, PENALTY= 8 | | |
| CG2R61 | CG2R62 | CG2R62 | 40.00 | 121.00 | ! | RIN, coumarin, isg | | |
| CG2R61 | CG2R62 | CG2R63 | 120.00 | 122.30 | ! | YTHY, 2,4(1H,3H)-quinazolinedione, isg | | |
| CG2R61 | CG2R62 | NG2R61 | 23.00 | 116.10 | ! | YTHY, 2,4(1H,3H)-quinazolinedione, isg | | |
| CG2R62 | CG2R62 | CG2R63 | 120.00 | 116.70 | ! | NA T | | |
| CG2R62 | CG2R62 | NG2R61 | 85.00 | 122.90 | ! | NA C | | |
| CG2R62 | CG2R62 | HGR62 | 42.00 | 119.00 | ! | NA nadh/ppi, jjpl/adm jr. 7/95 | | |
| NG2R61 | CG2R62 | HGR62 | 44.00 | 115.00 | ! | NA C, h6 | | |
| CG2R62 | CG2R63 | CG2R62 | 10.00 | 120.80 | ! | 4PYO, 4(1H)-pyridinone; from CG2R62 CG2R62 CG2R62; isg | | |
| CG2R62 | CG2R63 | OG2D4 | 100.00 | 124.60 | ! | NA T, o4 | | |
| CG2R61 | CG2R66 | CG2R61 | 40.00 | 122.50 | 35.00 | 2.41620 | ! | NAMODEL difluorotoluene |
| CG2R61 | CG2R66 | FGR1 | 60.00 | 118.75 | ! | NAMODEL difluorotoluene | | |
| CG321 | CG321 | CG324 | 58.35 | 110.50 | 11.16 | 2.56100 | ! | FLAVOP PIP1,2,3 |
| CG321 | CG321 | CG3RC1 | 53.35 | 111.00 | 8.00 | 2.56100 | ! | CARBOCY carbocyclic sugars |
| CG321 | CG321 | HGA2 | 26.50 | 110.10 | 22.53 | 2.17900 | ! | PROT alkane update, adm jr., 3/2/92 |
| CG324 | CG321 | HGA2 | 26.50 | 110.10 | 22.53 | 2.17900 | ! | FLAVOP PIP1,2,3 |
| CG3RC1 | CG321 | HGA2 | 34.50 | 110.10 | 22.53 | 2.17900 | ! | CARBOCY carbocyclic sugars |
| HGA2 | CG321 | HGA2 | 35.50 | 109.00 | 5.40 | 1.80200 | ! | PROT alkane update, adm jr., 3/2/92 |
| CG321 | CG324 | NG3P2 | 40.00 | 110.00 | ! | PIP, piperidine | | |
| CG321 | CG324 | HGA2 | 26.50 | 111.80 | 22.53 | 2.17900 | ! | FLAVOP PIP1,2,3 |
| NG3P2 | CG324 | HGA2 | 45.00 | 102.30 | 35.00 | 2.10100 | ! | PIP, piperidine |
| HGA2 | CG324 | HGA2 | 35.50 | 109.00 | 5.40 | 1.80200 | ! | PIP1,2,3 |
| OG301 | CG331 | HGA3 | 45.90 | 108.89 | ! | MEOB, Methoxybenzene, cacha | | |
| HGA3 | CG331 | HGA3 | 35.50 | 108.40 | 5.40 | 1.80200 | ! | PROT alkane update, adm jr., 3/2/92 |
| CG3C31 | CG3C31 | CG3C31 | 77.35 | 111.00 | 8.00 | 2.56100 | ! | PROTMOD cyclopropane |
| CG3C31 | CG3C31 | NG2R61 | 30.184 | 117.044 | | | | |
| CG3C31 | CG3C31 | HGA1 | 23.00 | 117.10 | 22.53 | 2.17900 | ! | PROTMOD cyclopropane |
| CG3C31 | CG3C31 | HGA2 | 23.00 | 117.10 | 22.53 | 2.17900 | ! | PROTMOD cyclopropane |
| NG2R61 | CG3C31 | HGA1 | 75.892 | 114.188 | | | | |
| HGA2 | CG3C31 | HGA2 | 23.00 | 117.00 | 5.40 | 1.80200 | ! | PROTMOD cyclopropane |
| CG3RC1 | CG3C52 | NG3C51 | 84.00 | 107.60 | ! | ***** , from CG3C52 CG3C52 NG3C51, PENALTY= 1.1 | | |

CG3RC1 CG3C52 HGA2 34.50 110.10 22.53 2.17900 ! CARBOCY carbocyclic sugars
 NG3C51 CG3C52 HGA2 54.00 109.00 !v 107.7 PRLD, pyrrolidine; 110.8 2PRL, 2-pyrroline; 110.4 3PRL, 3-pyrroline; 111.4 2IMI, 2-imidazoline; 111.7 2PRZ, 2-pyrazoline, kevo
 HGA2 CG3C52 HGA2 38.50 106.80 5.40 1.80200 ! THF, 10/17/05 viv
 CG321 CG3RC1 CG3C52 58.35 113.50 11.16 2.56100 ! BAM1, bile acid steroidal C-D ring, cacha, 02/08
 CG321 CG3RC1 CG3RC1 53.35 111.00 8.00 2.56100 ! CARBOCY carbocyclic sugars
 CG321 CG3RC1 HGA1 34.50 110.10 22.53 2.17900 ! CARBOCY carbocyclic sugars
 CG3C52 CG3RC1 CG3RC1 53.35 111.00 8.00 2.56100 ! CARBOCY carbocyclic sugars
 CG3C52 CG3RC1 NG3P2 162.274 108.021
 CG3C52 CG3RC1 HGA1 34.50 110.10 22.53 2.17900 ! CARBOCY carbocyclic sugars
 CG3RC1 CG3RC1 NG3P2 97.279 109.977 8.00 2.5610
 CG3RC1 CG3RC1 HGA1 34.50 110.10 22.53 2.17900 ! CARBOCY carbocyclic sugars
 NG3P2 CG3RC1 HGA1 105.536 105.525
 CG2R62 NG2R61 CG2R62 30.00 120.00 ! NA nad/ppi, jjp1/adm jr. 7/95
 CG2R62 NG2R61 CG3C31 45.00 118.40 ! ***** , from CG2R62 NG2R61 CG3C51, PENALTY= 6.9
 CG2R61 NG3C51 CG3C52 93.282 116.898
 CG3C52 NG3C51 CG3C52 140.00 103.70 !v 102.9 PRLD, pyrrolidine; 105.4 3PRL, 3-pyrroline, kevo
 CG324 NG3P2 CG3RC1 26.558 115.625
 CG324 NG3P2 HGP2 30.00 110.80 27.00 2.07400 ! PIP, piperidine
 CG3RC1 NG3P2 HGP2 97.752 110.057 27.00 2.0740
 HGP2 NG3P2 HGP2 51.00 107.50 ! PROT AcProNH₂, ProNH₂, AcProNHCH₃ RLD 4/23/93
 CG2R61 OG301 CG331 65.00 108.00 ! MEOB, Methoxybenzene, cacha

DIHEDRALS

OG2D2 CG2O3 CG2R62 CG2R62 3.1000 2 180.00 ! ***** , from OG2D2 CG2O3 CG2R61 CG2R61, PENALTY= 5.5
 OG2D2 CG2O3 CG2R62 CG2R63 3.1000 2 180.00 ! ***** , from OG2D2 CG2O3 CG2R61 CG2R61, PENALTY= 8
 CG2R62 CG2R61 CG2R61 CG2R66 3.1000 2 180.00 ! ***** , from CG2R61 CG2R61 CG2R61 CG2R66, PENALTY= 0.5
 CG2R62 CG2R61 CG2R61 NG3C51 3.1000 2 180.00 ! ***** , from CG2R61 CG2R61 CG2R61 NG311, PENALTY= 3.5
 CG2R66 CG2R61 CG2R61 OG301 3.1000 2 180.00 ! ***** , from CG2R61 CG2R61 CG2R61 OG301, PENALTY= 1.5
 NG3C51 CG2R61 CG2R61 OG301 2.4440 2 180.00
 CG2R61 CG2R61 CG2R62 CG2R62 0.5000 2 180.00 ! RIN, coumarin, isg
 CG2R61 CG2R61 CG2R62 NG2R61 7.0000 2 180.00 ! YTHY, 2,4(1H,3H)-quinazolinedione, isg
 CG2R66 CG2R61 CG2R62 CG2R62 0.5000 2 180.00 ! ***** , from CG2R61 CG2R61 CG2R62 CG2R62, PENALTY= 1.5
 CG2R66 CG2R61 CG2R62 CG2R63 3.1000 2 180.00 ! ***** , from CG2R61 CG2R61 CG2R62 CG2R63, PENALTY= 1.5
 OG301 CG2R61 CG2R62 CG2R62 3.1000 2 180.00 ! ***** , from CG2R61 CG2R61 CG2R61 OG301, PENALTY= 5.5
 OG301 CG2R61 CG2R62 NG2R61 2.4000 2 180.00
 HGR62 CG2R61 CG2R62 CG2R62 3.1000 2 180.00 ! ***** , from HGR61 CG2R61 CG2R62 CG2R62, PENALTY= 1
 HGR62 CG2R61 CG2R62 CG2R63 1.0000 2 180.00 ! ***** , from HGR61 CG2R61 CG2R61 CG2R62 CG2R63, PENALTY= 1
 CG2R61 CG2R61 CG2R66 CG2R61 3.1000 2 180.00 ! NAMODEL difluorotoluene
 CG2R61 CG2R61 CG2R66 FGR1 4.5000 2 180.00 ! NAMODEL difluorotoluene
 CG2R62 CG2R61 CG2R66 CG2R61 3.1000 2 180.00 ! ***** , from CG2R61 CG2R61 CG2R66 CG2R61, PENALTY= 0.5
 CG2R62 CG2R61 CG2R66 FGR1 4.5000 2 180.00 ! ***** , from CG2R61 CG2R61 CG2R66 FGR1, PENALTY= 0.5
 NG3C51 CG2R61 CG2R66 CG2R61 3.1000 2 180.00 ! ***** , from CG2R61 CG2R61 CG2R61 NG311, PENALTY= 8
 NG3C51 CG2R61 CG2R66 FGR1 2.4970 2 180.00
 HGR62 CG2R61 CG2R66 CG2R61 4.2000 2 180.00 ! NAMODEL difluorotoluene
 HGR62 CG2R61 CG2R66 FGR1 2.4000 2 180.00 ! NAMODEL difluorotoluene
 CG2R61 CG2R61 NG3C51 CG3C52 1.3590 2 0.00
 CG2R61 CG2R61 NG3C51 CG3C52 2.4490 4 0.00
 CG2R66 CG2R61 NG3C51 CG3C52 0.3300 2 180.00
 CG2R66 CG2R61 NG3C51 CG3C52 0.2440 4 0.00
 CG2R61 CG2R61 OG301 CG331 1.5800 2 180.00 ! MEOB, Methoxybenzene update, yxu
 CG2R61 CG2R61 OG301 CG331 0.2000 4 180.00 ! MEOB, Methoxybenzene update, yxu
 CG2R62 CG2R61 OG301 CG331 1.5800 2 180.00 ! ***** , from CG2R61 CG2R61 OG301 CG331, PENALTY= 0.5
 CG2R62 CG2R61 OG301 CG331 0.2000 4 180.00 ! ***** , from CG2R61 CG2R61 OG301 CG331, PENALTY= 0.5
 CG2O3 CG2R62 CG2R62 NG2R61 2.5000 2 180.00 ! ***** , from CG2O1 CG2R62 CG2R62 NG2R61, PENALTY= 7.5
 CG2O3 CG2R62 CG2R62 HGR62 1.0000 2 180.00 ! ***** , from CG2O1 CG2R62 CG2R62 HGR63, PENALTY= 9.5

CG2R61 CG2R62 CG2R62 CG2R61 2.5000 2 180.00 ! RIN, coumarin, isg
 CG2R61 CG2R62 CG2R62 CG2R63 3.7000 2 180.00 ! YTHY, 2,4(1H,3H)-quinazolinedione, isg
 CG2R61 CG2R62 CG2R62 NG2R61 3.5000 2 180.00 ! YTHY, 2,4(1H,3H)-quinazolinedione, isg
 CG2R63 CG2R62 CG2R62 NG2R61 3.0000 2 180.00 ! NA T
 CG2R63 CG2R62 CG2R62 HGR62 1.0000 2 180.00 ! NA bases
 CG203 CG2R62 CG2R63 CG2R62 3.0000 2 180.00
 CG203 CG2R62 CG2R63 OG2D4 2.4980 2 180.00
 CG2R61 CG2R62 CG2R63 CG2R62 1.6000 2 180.00 ! ***** , from CG2R62 CG2R62 CG2R63 CG2R62, PENALTY= 1.5
 CG2R61 CG2R62 CG2R63 OG2D4 1.0000 2 180.00 ! YTHY, 2,4(1H,3H)-quinazolinedione, isg
 CG2R62 CG2R62 CG2R63 CG2R62 1.6000 2 180.00 ! 4PYO, 4(1H)-pyridinone, isg
 CG2R62 CG2R62 CG2R63 OG2D4 1.0000 2 180.00 ! NA bases
 CG2R61 CG2R62 NG2R61 CG2R62 4.0000 2 180.00 ! ***** , from CG2R67 CG2R62 NG2R61 CG2R62, PENALTY= 0.5
 CG2R61 CG2R62 NG2R61 CG3C31 11.0000 2 180.00 ! ***** , from CG2R62 CG2R62 NG2R61 CG3C51, PENALTY= 8.4
 CG2R62 CG2R62 NG2R61 CG2R62 4.0000 2 180.00 ! NA nad/ppi, jjp1/adm jr. 7/95
 CG2R62 CG2R62 NG2R61 CG3C31 11.0000 2 180.00 ! ***** , from CG2R62 CG2R62 NG2R61 CG3C51, PENALTY= 6.9
 HGR62 CG2R62 NG2R61 CG2R62 5.6000 2 180.00 ! 4PYO, 4(1H)-pyridinone, isg
 HGR62 CG2R62 NG2R61 CG3C31 0.3000 2 180.00 ! ***** , from HGR62 CG2R62 NG2R61 CG3C51, PENALTY= 6.9
 CG324 CG321 CG321 CG3RC1 0.4520 3 0.00
 CG324 CG321 CG321 HGA2 0.1950 3 0.00 ! FLAVOP PIP1,2,3
 CG3RC1 CG321 CG321 HGA2 0.1950 3 0.00 ! LIPID alkanes
 HGA2 CG321 CG321 HGA2 0.2200 3 0.00 ! LIPID alkanes
 CG321 CG321 CG324 NG3P2 1.0000 3 0.00 ! PIP, piperidine ! @@@ Kenno: 0.1950 -> 1.0000
 CG321 CG321 CG324 HGA2 0.1950 3 0.00 ! FLAVOP PIP1,2,3
 HGA2 CG321 CG324 NG3P2 0.1950 3 0.00 ! PIP, piperidine
 HGA2 CG321 CG324 HGA2 0.1950 3 0.00 ! FLAVOP PIP1,2,3
 CG321 CG321 CG3RC1 CG3C52 0.2000 3 0.00 ! BAM1, bile acid steroidal C-D ring, cacha, 02/08
 CG321 CG321 CG3RC1 CG3RC1 0.1500 3 0.00 ! CARBOCY carbocyclic sugars
 CG321 CG321 CG3RC1 HGA1 0.1500 3 0.00 ! CARBOCY carbocyclic sugars
 HGA2 CG321 CG3RC1 CG3C52 0.1950 1 0.00 ! BAM1, bile acid steroidal C-D ring, cacha, 02/08
 HGA2 CG321 CG3RC1 CG3RC1 0.1500 3 0.00 ! CARBOCY carbocyclic sugars
 HGA2 CG321 CG3RC1 HGA1 0.1500 3 0.00 ! CARBOCY carbocyclic sugars
 CG321 CG324 NG3P2 CG3RC1 0.3190 1 180.00
 CG321 CG324 NG3P2 CG3RC1 2.2270 2 0.00
 CG321 CG324 NG3P2 CG3RC1 1.4290 3 180.00
 CG321 CG324 NG3P2 HGP2 0.1000 3 0.00 ! PIP, piperidine
 HGA2 CG324 NG3P2 CG3RC1 1.1970 3 0.00
 HGA2 CG324 NG3P2 HGP2 0.1000 3 0.00 ! PIP, piperidine
 HGA3 CG331 OG301 CG2R61 0.0850 3 0.00 ! MEOB, Methoxybenzene, cacha
 CG3C31 CG3C31 CG3C31 NG2R61 1.2340 3 180.00
 CG3C31 CG3C31 CG3C31 HGA1 0.1000 6 0.00 ! AMCP, aminomethyl cyclopropane; from PROTMOD hf/cyclopropane; jhs
 CG3C31 CG3C31 CG3C31 HGA2 0.1000 6 0.00 ! PROTMOD hf/cyclopropane
 NG2R61 CG3C31 CG3C31 HGA2 2.2320 3 180.00
 HGA1 CG3C31 CG3C31 HGA2 0.2000 5 180.00 ! AMCP, aminomethyl cyclopropane; from PROTMOD hf/cyclopropane; jhs
 HGA2 CG3C31 CG3C31 HGA2 0.2000 5 180.00 ! PROTMOD hf/cyclopropane
 CG3C31 CG3C31 NG2R61 CG2R62 2.3350 3 0.00
 HGA1 CG3C31 NG2R61 CG2R62 2.4770 3 0.00
 NG3C51 CG3C52 CG3RC1 CG321 1.5720 3 180.0
 NG3C51 CG3C52 CG3RC1 CG3RC1 0.9650 3 0.002
 NG3C51 CG3C52 CG3RC1 NG3P2 1.4040 3 0.00
 NG3C51 CG3C52 CG3RC1 HGA1 1.0890 3 0.00
 HGA2 CG3C52 CG3RC1 CG321 0.1950 1 0.00 ! BAM1, bile acid steroidal C-D ring, cacha, 02/08
 HGA2 CG3C52 CG3RC1 CG3RC1 0.1500 3 0.00 ! CARBOCY carbocyclic sugars
 HGA2 CG3C52 CG3RC1 NG3P2 2.4440 3 0.00
 HGA2 CG3C52 CG3RC1 HGA1 0.1500 3 0.00 ! CARBOCY carbocyclic sugars

```

CG3RC1 CG3C52 NG3C51 CG2R61      1.2050  3   180.00
CG3RC1 CG3C52 NG3C51 CG3C52      0.1800  3   0.00 ! ***** , from CG3C52 CG3C52 NG3C51 CG3C52, PENALTY= 1.1
HGA2  CG3C52 NG3C51 CG2R61      1.6800  3   0.00
HGA2  CG3C52 NG3C51 CG3C52      0.0000  3   0.00 ! 3PRL, 3-pyrroline, kevo
CG321 CG3RC1 CG3RC1 CG3C52      0.1500  3   0.00 ! CARBOCY carbocyclic sugars
CG321 CG3RC1 CG3RC1 NG3P2       0.3810  3   180.00
CG321 CG3RC1 CG3RC1 HGA1        0.1500  3   0.00 ! CARBOCY carbocyclic sugars
CG3C52 CG3RC1 CG3RC1 CG3C52      0.1500  3   0.00 ! CARBOCY carbocyclic sugars
CG3C52 CG3RC1 CG3RC1 NG3P2       0.8780  3   180.0
CG3C52 CG3RC1 CG3RC1 HGA1        0.1500  3   0.00 ! CARBOCY carbocyclic sugars
NG3P2  CG3RC1 CG3RC1 HGA1        2.0850  3   180.00
HGA1  CG3RC1 CG3RC1 HGA1        0.1500  3   0.00 ! CARBOCY carbocyclic sugars
CG3C52 CG3RC1 NG3P2  CG324     1.8470  3   180.00
CG3C52 CG3RC1 NG3P2  HGP2       1.3210  3   0.00
CG3RC1 CG3RC1 NG3P2  CG324     0.2800  3   180.00
CG3RC1 CG3RC1 NG3P2  HGP2       1.3090  3   0.00
HGA1  CG3RC1 NG3P2  CG324     0.7100  3   0.00
HGA1  CG3RC1 NG3P2  HGP2       2.4770  3   180.00

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IMPROPERs

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CG2O3 OG2D2 OG2D2 CG2R62      96.0000  0   0.00 ! ***** , from CG2O3 OG2D2 OG2D2 CG2R61, PENALTY= 0.5
CG2R63 CG2R62 CG2R62 OG2D4      15.0000  0   0.00 ! 4PYO, 4(1H)-pyridinone, isg

```

```

END
RETURN

```

Table DSXI. Optimized force field topology and parameters for cationic moxifloxacin (CHARMM stream including CGenFF parameters)

```

* Toppar stream file generated by
*
read rtf card append
* Topologies generated by
* CHARMM General Force Field (CGenFF) program version 2.2.0
*
36 1
=====
! Moxifloxacin - Cationic
=====

RESI MOXC      1.000 !
GROUP          ! CHARGE  CH_PENALTY
ATOM C1        CG331  -0.100 !    0.025
ATOM O1        OG301  -0.391 !    2.180
ATOM C2        CG2R61  0.297
ATOM C3        CG2R62  0.395
ATOM C4        CG2R62  -0.234 !    9.831
ATOM C5        CG2R61  0.021 !
ATOM C6        CG2R66  0.006
ATOM C7        CG2R61  0.118
ATOM N1        NG3C51 -0.507
ATOM C8        CG3C52  0.169

```

ATOM C9 CG3RC1 -0.114
 ATOM H1 HGA1 0.090 ! 2.316
 ATOM C10 CG321 -0.190 ! 7.285
 ATOM C11 CG321 -0.161
 ATOM C12 CG324 0.170
 ATOM N2 NG3P2 -0.344
 ATOM C13 CG3RC1 0.245
 ATOM H2 HGA1 0.090 ! 4.911
 ATOM C14 CG3C52 -0.026
 ATOM F1 FGR1 -0.282
 ATOM C15 CG2R63 0.304
 ATOM O2 OG2D4 -0.449
 ATOM C16 CG2R62 0.118
 ATOM C17 CG2R62 0.095 ! 8.365
 ATOM N3 NG2R61 -0.402
 ATOM C18 CG3C31 0.099
 ATOM C19 CG3C31 -0.180 ! 7.029
 ATOM C20 CG3C31 -0.180 ! 7.029
 ATOM C21 CG2O2 0.320
 ATOM O3 OG311 -0.513 ! 4.026
 ATOM O4 OG2D1 -0.437 ! 3.884
 ATOM H3 HGA3 0.090 ! 0.000
 ATOM H4 HGA3 0.090 ! 0.000
 ATOM H5 HGA3 0.090 ! 0.000
 ATOM H6 HGR62 0.161 ! 0.075
 ATOM H7 HGA2 0.090 ! 1.375
 ATOM H8 HGA2 0.090 ! 1.375
 ATOM H9 HGA2 0.090 ! 0.000
 ATOM H10 HGA2 0.090 ! 0.000
 ATOM H11 HGA2 0.090 ! 0.000
 ATOM H12 HGA2 0.090 ! 0.000
 ATOM H13 HGA2 0.090 ! 0.975
 ATOM H14 HGA2 0.090 ! 0.975
 ATOM H15 HGP2 0.329
 ATOM H16 HGA2 0.090 ! 2.637
 ATOM H17 HGA2 0.090 ! 2.637
 ATOM H18 HGR62 0.105 ! 4.797
 ATOM H19 HGA1 0.090 ! 4.940
 ATOM H20 HGA2 0.090 ! 2.500
 ATOM H21 HGA2 0.090 ! 2.500
 ATOM H22 HGA2 0.090 ! 2.500
 ATOM H23 HGA2 0.090 ! 2.500
 ATOM H24 HGP2 0.329
 ATOM H25 HGP1 0.429 ! 0.219

BOND H23 C20
 BOND H7 C8
 BOND H8 C8
 BOND H9 C10
 BOND H4 C1
 BOND H6 C5
 BOND O2 C15
 BOND H11 C11
 BOND F1 C6
 BOND C5 C6
 BOND C5 C4

```

BOND C8   N1
BOND C8   C9
BOND C20  H22
BOND C20  C19
BOND C20  C18
BOND C6   C7
BOND H12  C11
BOND H20  C19
BOND C15  C4
BOND C15  C16
BOND C4   C3
BOND H3   C1
BOND C11  C10
BOND C11  C12
BOND C10  C9
BOND C10  H10
BOND C7   N1
BOND C7   C2
BOND C1   H5
BOND C1   O1
BOND C19  C18
BOND C19  H21
BOND C3   C2
BOND C3   N3
BOND N1   C14
BOND C2   O1
BOND C9   H1
BOND C9   C13
BOND C16  C17
BOND C16  C21
BOND N3   C18
BOND N3   C17
BOND C18  H19
BOND C17  H18
BOND H15  N2
BOND H25  O3
BOND C21  O3
BOND C21  O4
BOND C12  H13
BOND C12  N2
BOND C12  H14
BOND N2   C13
BOND N2   H24
BOND C14  C13
BOND C14  H16
BOND C14  H17
BOND C13  H2
IMPR C15   C4     C16     O2
IMPR C21   C16     O4     O3

END

```

```

read param card flex append
* Parameters generated by analogy by
* CHARMM General Force Field (CGenFF) program version 2.2.0
*

```

! Penalties lower than 10 indicate the analogy is fair; penalties between 10
 ! and 50 mean some basic validation is recommended; penalties higher than
 ! 50 indicate poor analogy and mandate extensive validation/optimization.

BONDS

| | | | | |
|--------|--------|---------|--------|--|
| CG2O2 | CG2R62 | 254.00 | 1.4800 | ! ***** , from CG2O2 CG2R61, PENALTY= 5 |
| CG2O2 | OG2D1 | 750.00 | 1.2200 | ! PROT adm jr. 5/02/91, acetic acid pure solvent; LIPID methyl acetate |
| CG2O2 | OG311 | 230.00 | 1.4000 | ! PROT adm jr. 5/02/91, acetic acid pure solvent |
| CG2R61 | CG2R61 | 305.00 | 1.3750 | ! PROT benzene, JES 8/25/89 |
| CG2R61 | CG2R62 | 394.00 | 1.3750 | ! YTHY, 2,4(1H,3H)-quinazolinedione, isg |
| CG2R61 | CG2R66 | 305.00 | 1.3700 | ! NAMODEL difluorotoluene |
| CG2R61 | NG3C51 | 406.429 | 1.370 | |
| CG2R61 | OG301 | 230.00 | 1.3820 | ! COMPDS peml |
| CG2R61 | HGR62 | 340.00 | 1.0800 | ! NA, DFT |
| CG2R62 | CG2R62 | 420.00 | 1.3500 | ! NA nad/ppi, jjp1/adm jr. 7/95 |
| CG2R62 | CG2R63 | 302.00 | 1.4030 | ! NA T, adm jr. 11/97 |
| CG2R62 | NG2R61 | 302.00 | 1.3430 | ! NA C, adm jr. 11/97 |
| CG2R62 | HGR62 | 350.00 | 1.0900 | ! NA C,U, JWK |
| CG2R63 | OG2D4 | 660.00 | 1.2340 | ! NA U,A,G par_a4 adm jr. 10/2/91 |
| CG2R66 | FGR1 | 400.00 | 1.3580 | ! NAMODEL difluorotoluene |
| CG321 | CG321 | 222.50 | 1.5300 | ! PROT alkane update, adm jr., 3/2/92 |
| CG321 | CG324 | 222.50 | 1.5300 | ! FLAVOP PIP1,2,3 |
| CG321 | CG3RC1 | 222.50 | 1.5240 | ! CARBOCY carbocyclic sugars |
| CG321 | HGA2 | 309.00 | 1.1110 | ! PROT alkane update, adm jr., 3/2/92 |
| CG324 | NG3P2 | 200.00 | 1.4900 | ! PIP, piperidine |
| CG324 | HGA2 | 284.50 | 1.1000 | ! FLAVOP PIP1,2,3 |
| CG331 | OG301 | 360.00 | 1.4150 | ! diethylether, alex |
| CG331 | HGA3 | 322.00 | 1.1110 | ! PROT alkane update, adm jr., 3/2/92 |
| CG3C31 | CG3C31 | 240.00 | 1.5010 | ! PROTMOD cyclopropane |
| CG3C31 | NG2R61 | 352.828 | 1.448 | |
| CG3C31 | HGA1 | 340.00 | 1.0830 | ! PROTMOD cyclopropane |
| CG3C31 | HGA2 | 340.00 | 1.0830 | ! PROTMOD cyclopropane |
| CG3C52 | CG3RC1 | 222.50 | 1.5240 | ! CARBOCY carbocyclic sugars |
| CG3C52 | NG3C51 | 400.00 | 1.4780 | ! PRLD, pyrrolidine; 2PRL, 2-pyrroline, kevo |
| CG3C52 | HGA2 | 307.00 | 1.1000 | ! THF, THF neutron diffr., 5/30/06, viv |
| CG3RC1 | CG3RC1 | 222.50 | 1.5230 | ! CARBOCY carbocyclic sugars |
| CG3RC1 | NG3P2 | 164.854 | 1.497 | |
| CG3RC1 | HGA1 | 309.00 | 1.1110 | ! CARBOCY carbocyclic sugars |
| NG3P2 | HGP2 | 460.00 | 1.0060 | ! PROT AcProNH2, ProNH2, AcProNHCH3 RLD 4/23/93 |
| OG311 | HGP1 | 545.00 | 0.9600 | ! PROT EMB 11/21/89 methanol vib fit; og tested on MeOH EtOH,... |

ANGLES

| | | | | | | | |
|--------|--------|--------|--------|--------|--|---------|---|
| CG2R62 | CG2O2 | OG2D1 | 70.00 | 123.10 | 20.00 | 2.44200 | ! ***** , from CG2R61 CG2O2 OG2D1, PENALTY= 0.5 |
| CG2R62 | CG2O2 | OG311 | 40.00 | 113.90 | 30.00 | 2.37000 | ! ***** , from CG2R61 CG2O2 OG311, PENALTY= 0.5 |
| OG2D1 | CG2O2 | OG311 | 50.00 | 123.00 | 210.00 | 2.26200 | ! PROT adm jr, 10/17/90, acetic acid vibrations |
| CG2R61 | CG2R61 | CG2R62 | 40.00 | 119.00 | 35.00 | 2.41620 | ! YTHY, 2,4(1H,3H)-quinazolinedione, isg |
| CG2R61 | CG2R61 | CG2R66 | 40.00 | 119.00 | 35.00 | 2.41620 | ! NAMODEL difluorotoluene |
| CG2R61 | CG2R61 | NG3C51 | 40.00 | 120.00 | ! ***** , from CG2R61 CG2R61 NG311, PENALTY= 3 | | |
| CG2R61 | CG2R61 | OG301 | 110.00 | 120.00 | ! BIPHENYL ANALOGS, peml | | |
| CG2R62 | CG2R61 | CG2R66 | 40.00 | 119.00 | 35.00 | 2.41620 | ! ***** , from CG2R61 CG2R61 CG2R66, PENALTY= 0.5 |
| CG2R62 | CG2R61 | OG301 | 110.00 | 120.00 | ! ***** , from CG2R61 CG2R61 OG301, PENALTY= 0.5 | | |
| CG2R62 | CG2R61 | HGR62 | 30.00 | 120.00 | 22.00 | 2.15250 | ! ***** , from CG2R61 CG2R61 HGR62, PENALTY= 0.5 |
| CG2R66 | CG2R61 | NG3C51 | 40.00 | 120.00 | ! ***** , from CG2R61 CG2R61 NG311, PENALTY= 4.5 | | |
| CG2R66 | CG2R61 | HGR62 | 30.00 | 121.50 | 22.00 | 2.15250 | ! NAMODEL difluorotoluene |
| CG2O2 | CG2R62 | CG2R62 | 10.00 | 131.80 | ! ***** , from CG2O1 CG2R62 CG2R62, PENALTY= 2.5 | | |

CG2O2 CG2R62 CG2R63 10.00 131.80 ! ***** , from CG2O1 CG2R62 CG2R62, PENALTY= 4
 CG2R61 CG2R62 CG2R62 40.00 121.00 ! RIN, coumarin, isg
 CG2R61 CG2R62 CG2R63 120.00 122.30 ! YTHY, 2,4(1H,3H)-quinazolinedione, isg
 CG2R61 CG2R62 NG2R61 23.00 116.10 ! YTHY, 2,4(1H,3H)-quinazolinedione, isg
 CG2R62 CG2R62 CG2R63 120.00 116.70 ! NA T
 CG2R62 CG2R62 NG2R61 85.00 122.90 ! NA C
 CG2R62 CG2R62 HGR62 42.00 119.00 ! NA nadh/ppi, jjp1/adm jr. 7/95
 NG2R61 CG2R62 HGR62 44.00 115.00 ! NA C, h6
 CG2R62 CG2R63 CG2R62 10.00 120.80 ! 4PYO, 4(1H)-pyridinone; from CG2R62 CG2R62 CG2R62; isg
 CG2R62 CG2R63 OG2D4 100.00 124.60 ! NA T, o4
 CG2R61 CG2R66 CG2R61 40.00 122.50 35.00 2.41620 ! NAMODEL difluorotoluene
 CG2R61 CG2R66 FGR1 60.00 118.75 ! NAMODEL difluorotoluene
 CG321 CG321 CG324 58.35 110.50 11.16 2.56100 ! FLAVOP PIP1,2,3
 CG321 CG321 CG3RC1 53.35 111.00 8.00 2.56100 ! CARBOCY carbocyclic sugars
 CG321 CG321 HGA2 26.50 110.10 22.53 2.17900 ! PROT alkane update, adm jr., 3/2/92
 CG324 CG321 HGA2 26.50 110.10 22.53 2.17900 ! FLAVOP PIP1,2,3
 CG3RC1 CG321 HGA2 34.50 110.10 22.53 2.17900 ! CARBOCY carbocyclic sugars
 HGA2 CG321 HGA2 35.50 109.00 5.40 1.80200 ! PROT alkane update, adm jr., 3/2/92
 CG321 CG324 NG3P2 40.00 110.00 ! PIP, piperidine
 CG321 CG324 HGA2 26.50 111.80 22.53 2.17900 ! FLAVOP PIP1,2,3
 NG3P2 CG324 HGA2 45.00 102.30 35.00 2.10100 ! PIP, piperidine
 HGA2 CG324 HGA2 35.50 109.00 5.40 1.80200 ! PIP1,2,3
 OG301 CG331 HGA3 45.90 108.89 ! MEOB, Methoxybenzene, cacha
 HGA3 CG331 HGA3 35.50 108.40 5.40 1.80200 ! PROT alkane update, adm jr., 3/2/92
 CG3C31 CG3C31 CG3C31 77.35 111.00 8.00 2.56100 ! PROTMOD cyclopropane
 CG3C31 CG3C31 NG2R61 30.184 117.044
 CG3C31 CG3C31 HGA1 23.00 117.10 22.53 2.17900 ! PROTMOD cyclopropane
 CG3C31 CG3C31 HGA2 23.00 117.10 22.53 2.17900 ! PROTMOD cyclopropane
 NG2R61 CG3C31 HGA1 75.892 114.188
 HGA2 CG3C31 HGA2 23.00 117.00 5.40 1.80200 ! PROTMOD cyclopropane
 CG3RC1 CG3C52 NG3C51 84.00 107.60 ! ***** , from CG3C52 CG3C52 NG3C51, PENALTY= 1.1
 CG3RC1 CG3C52 HGA2 34.50 110.10 22.53 2.17900 ! CARBOCY carbocyclic sugars
 NG3C51 CG3C52 HGA2 54.00 109.00 !v 107.7 PRLD, pyrrolidine; 110.8 2PRL, 2-pyrroline; 110.4 3PRL, 3-pyrroline; 111.4 2IMI, 2-imidazoline; 111.7 2PRZ, 2-pyrazoline, kevo
 HGA2 CG3C52 HGA2 38.50 106.80 5.40 1.80200 ! THF, 10/17/05 viv
 CG321 CG3RC1 CG3C52 58.35 113.50 11.16 2.56100 ! BAM1, bile acid steroidal C-D ring, cacha, 02/08
 CG321 CG3RC1 CG3RC1 53.35 111.00 8.00 2.56100 ! CARBOCY carbocyclic sugars
 CG321 CG3RC1 HGA1 34.50 110.10 22.53 2.17900 ! CARBOCY carbocyclic sugars
 CG3C52 CG3RC1 CG3RC1 53.35 111.00 8.00 2.56100 ! CARBOCY carbocyclic sugars
 CG3C52 CG3RC1 NG3P2 162.274 108.021
 CG3C52 CG3RC1 HGA1 34.50 110.10 22.53 2.17900 ! CARBOCY carbocyclic sugars
 CG3RC1 CG3RC1 NG3P2 97.279 109.977 8.00 2.5610
 CG3RC1 CG3RC1 HGA1 34.50 110.10 22.53 2.17900 ! CARBOCY carbocyclic sugars
 NG3P2 CG3RC1 HGA1 105.536 105.525
 CG2R62 NG2R61 CG2R62 30.00 120.00 ! NA nad/ppi, jjp1/adm jr. 7/95
 CG2R62 NG2R61 CG3C31 45.00 118.40 ! ***** , from CG2R62 NG2R61 CG3C51, PENALTY= 6.9
 CG2R61 NG3C51 CG3C52 48.288 107.844
 CG3C52 NG3C51 CG3C52 140.00 103.70 !v 102.9 PRLD, pyrrolidine; 105.4 3PRL, 3-pyrroline, kevo
 CG324 NG3P2 CG3RC1 26.558 115.625
 CG324 NG3P2 HGP2 30.00 110.80 27.00 2.07400 ! PIP, piperidine
 CG3RC1 NG3P2 HGP2 97.752 110.057 27.00 2.0740
 HGP2 NG3P2 HGP2 51.00 107.50 ! PROT AcProNH₂, ProNH₂, AcProNHCH₃ RLD 4/23/93
 CG2R61 OG301 CG331 65.00 108.00 ! MEOB, Methoxybenzene, cacha
 CG2O2 OG311 HGP1 55.00 115.00 ! PROT adm jr. 5/02/91, acetic acid pure solvent

DIHEDRALS

| | | | | | | |
|--------|--------|--------|--------|---------|---|---|
| OG2D1 | CG2O2 | CG2R62 | CG2R62 | 1.0250 | 2 | 180.00 ! ***** , from OG2D1 CG2O2 CG2R61 CG2R61, PENALTY= 5.5 |
| OG2D1 | CG2O2 | CG2R62 | CG2R63 | 1.0250 | 2 | 180.00 ! ***** , from OG2D1 CG2O2 CG2R61 CG2R61, PENALTY= 8 |
| OG311 | CG2O2 | CG2R62 | CG2R62 | 1.0250 | 2 | 180.00 ! ***** , from OG311 CG2O2 CG2R61 CG2R61, PENALTY= 5.5 |
| OG311 | CG2O2 | CG2R62 | CG2R63 | 1.0250 | 2 | 180.00 ! ***** , from OG311 CG2O2 CG2R61 CG2R61, PENALTY= 8 |
| CG2R62 | CG2O2 | OG311 | HGP1 | 0.9750 | 1 | 180.00 ! ***** , from CG2R61 CG2O2 OG311 HGP1, PENALTY= 0.5 |
| CG2R62 | CG2O2 | OG311 | HGP1 | 2.7000 | 2 | 180.00 ! ***** , from CG2R61 CG2O2 OG311 HGP1, PENALTY= 0.5 |
| CG2R62 | CG2O2 | OG311 | HGP1 | 0.0500 | 3 | 180.00 ! ***** , from CG2R61 CG2O2 OG311 HGP1, PENALTY= 0.5 |
| CG2R62 | CG2O2 | OG311 | HGP1 | 0.2500 | 6 | 180.00 ! ***** , from CG2R61 CG2O2 OG311 HGP1, PENALTY= 0.5 |
| OG2D1 | CG2O2 | OG311 | HGP1 | 2.0500 | 2 | 180.00 ! PROT adm jr, 10/17/90, acetic Acid C-OH rotation barrier |
| CG2R62 | CG2R61 | CG2R61 | CG2R66 | 3.1000 | 2 | 180.00 ! ***** , from CG2R61 CG2R61 CG2R61 CG2R66, PENALTY= 0.5 |
| CG2R62 | CG2R61 | CG2R61 | NG3C51 | 3.1000 | 2 | 180.00 ! ***** , from CG2R61 CG2R61 CG2R61 NG311, PENALTY= 3.5 |
| CG2R66 | CG2R61 | CG2R61 | OG301 | 3.1000 | 2 | 180.00 ! ***** , from CG2R61 CG2R61 CG2R61 OG301, PENALTY= 1.5 |
| NG3C51 | CG2R61 | CG2R61 | OG301 | 2.4440 | 2 | 180.00 |
| CG2R61 | CG2R61 | CG2R62 | CG2R62 | 0.5000 | 2 | 180.00 ! RIN, coumarin, isg |
| CG2R61 | CG2R61 | CG2R62 | NG2R61 | 7.0000 | 2 | 180.00 ! YTHY, 2,4(1H,3H)-quinazolinedione, isg |
| CG2R66 | CG2R61 | CG2R62 | CG2R62 | 0.5000 | 2 | 180.00 ! ***** , from CG2R61 CG2R61 CG2R62 CG2R62, PENALTY= 1.5 |
| CG2R66 | CG2R61 | CG2R62 | CG2R63 | 3.1000 | 2 | 180.00 ! ***** , from CG2R61 CG2R61 CG2R62 CG2R63, PENALTY= 1.5 |
| OG301 | CG2R61 | CG2R62 | CG2R62 | 3.1000 | 2 | 180.00 ! ***** , from CG2R61 CG2R61 CG2R61 OG301, PENALTY= 5.5 |
| OG301 | CG2R61 | CG2R62 | NG2R61 | 2.4000 | 2 | 180.00 |
| HGR62 | CG2R61 | CG2R62 | CG2R62 | 3.1000 | 2 | 180.00 ! ***** , from HGR61 CG2R61 CG2R62 CG2R62, PENALTY= 1 |
| HGR62 | CG2R61 | CG2R62 | CG2R63 | 1.0000 | 2 | 180.00 ! ***** , from HGR61 CG2R61 CG2R62 CG2R63, PENALTY= 1 |
| CG2R61 | CG2R61 | CG2R66 | CG2R61 | 3.1000 | 2 | 180.00 ! NAMODEL difluorotoluene |
| CG2R61 | CG2R61 | CG2R66 | FGR1 | 4.5000 | 2 | 180.00 ! NAMODEL difluorotoluene |
| CG2R62 | CG2R61 | CG2R66 | CG2R61 | 3.1000 | 2 | 180.00 ! ***** , from CG2R61 CG2R61 CG2R66 CG2R61, PENALTY= 0.5 |
| CG2R62 | CG2R61 | CG2R66 | FGR1 | 4.5000 | 2 | 180.00 ! ***** , from CG2R61 CG2R61 CG2R66 FGR1, PENALTY= 0.5 |
| NG3C51 | CG2R61 | CG2R66 | CG2R61 | 3.1000 | 2 | 180.00 ! ***** , from CG2R61 CG2R61 CG2R61 NG311, PENALTY= 8 |
| NG3C51 | CG2R61 | CG2R66 | FGR1 | 2.5800 | 2 | 180.00 ! ***** , from NG311 CG2R61 CG2R61 OG3R60, PENALTY= 36.2 |
| HGR62 | CG2R61 | CG2R66 | CG2R61 | 4.2000 | 2 | 180.00 ! NAMODEL difluorotoluene |
| HGR62 | CG2R61 | CG2R66 | FGR1 | 2.4000 | 2 | 180.00 ! NAMODEL difluorotoluene |
| CG2R61 | CG2R61 | NG3C51 | CG3C52 | 1.3590 | 2 | 0.00 |
| CG2R61 | CG2R61 | NG3C51 | CG3C52 | 2.4490 | 4 | 0.00 |
| CG2R66 | CG2R61 | NG3C51 | CG3C52 | 0.3300 | 2 | 180.00 |
| CG2R66 | CG2R61 | NG3C51 | CG3C52 | 0.2440 | 4 | 0.00 |
| CG2R61 | CG2R61 | OG301 | CG331 | 1.5800 | 2 | 180.00 ! MEOB, Methoxybenzene update, yxu |
| CG2R61 | CG2R61 | OG301 | CG331 | 0.2000 | 4 | 180.00 ! MEOB, Methoxybenzene update, yxu |
| CG2R62 | CG2R61 | OG301 | CG331 | 1.5800 | 2 | 180.00 ! ***** , from CG2R61 CG2R61 OG301 CG331, PENALTY= 0.5 |
| CG2R62 | CG2R61 | OG301 | CG331 | 0.2000 | 4 | 180.00 ! ***** , from CG2R61 CG2R61 OG301 CG331, PENALTY= 0.5 |
| CG2O2 | CG2R62 | CG2R62 | NG2R61 | 2.5000 | 2 | 180.00 ! ***** , from CG2O1 CG2R62 CG2R62 NG2R61, PENALTY= 2.5 |
| CG2O2 | CG2R62 | CG2R62 | HGR62 | 1.0000 | 2 | 180.00 ! ***** , from CG2O1 CG2R62 CG2R62 HGR63, PENALTY= 4.5 |
| CG2R61 | CG2R62 | CG2R62 | CG2R61 | 2.5000 | 2 | 180.00 ! RIN, coumarin, isg |
| CG2R61 | CG2R62 | CG2R62 | CG2R63 | 3.7000 | 2 | 180.00 ! YTHY, 2,4(1H,3H)-quinazolinedione, isg |
| CG2R61 | CG2R62 | CG2R62 | NG2R61 | 3.5000 | 2 | 180.00 ! YTHY, 2,4(1H,3H)-quinazolinedione, isg |
| CG2R63 | CG2R62 | CG2R62 | NG2R61 | 3.0000 | 2 | 180.00 ! NA T |
| CG2R63 | CG2R62 | CG2R62 | HGR62 | 1.0000 | 2 | 180.00 ! NA bases |
| CG2O2 | CG2R62 | CG2R63 | CG2R62 | 3.0000 | 2 | 180.00 |
| CG2O2 | CG2R62 | CG2R63 | OG2D4 | 2.4980 | 2 | 180.00 |
| CG2R61 | CG2R62 | CG2R63 | CG2R62 | 1.6000 | 2 | 180.00 ! ***** , from CG2R62 CG2R62 CG2R63 CG2R62, PENALTY= 1.5 |
| CG2R61 | CG2R62 | CG2R63 | OG2D4 | 1.0000 | 2 | 180.00 ! YTHY, 2,4(1H,3H)-quinazolinedione, isg |
| CG2R62 | CG2R62 | CG2R63 | CG2R62 | 1.6000 | 2 | 180.00 ! 4PYO, 4(1H)-pyridinone, isg |
| CG2R62 | CG2R62 | CG2R63 | OG2D4 | 1.0000 | 2 | 180.00 ! NA bases |
| CG2R61 | CG2R62 | NG2R61 | CG2R62 | 4.0000 | 2 | 180.00 ! ***** , from CG2R67 CG2R62 NG2R61 CG2R62, PENALTY= 0.5 |
| CG2R61 | CG2R62 | NG2R61 | CG3C31 | 11.0000 | 2 | 180.00 ! ***** , from CG2R62 CG2R62 NG2R61 CG3C51, PENALTY= 8.4 |
| CG2R62 | CG2R62 | NG2R61 | CG2R62 | 4.0000 | 2 | 180.00 ! NA nad/ppi, jjp1/adm jr. 7/95 |
| CG2R62 | CG2R62 | NG2R61 | CG3C31 | 11.0000 | 2 | 180.00 ! ***** , from CG2R62 CG2R62 NG2R61 CG3C51, PENALTY= 6.9 |
| HGR62 | CG2R62 | NG2R61 | CG2R62 | 5.6000 | 2 | 180.00 ! 4PYO, 4(1H)-pyridinone, isg |
| HGR62 | CG2R62 | NG2R61 | CG3C31 | 0.3000 | 2 | 180.00 ! ***** , from HGR62 CG2R62 NG2R61 CG3C51, PENALTY= 6.9 |

CG324 CG321 CG321 CG3RC1 0.4520 3 0.00
 CG324 CG321 CG321 HGA2 0.1950 3 0.00 ! FLAVOP PIP1,2,3
 CG3RC1 CG321 CG321 HGA2 0.1950 3 0.00 ! LIPID alkanes
 HGA2 CG321 CG321 HGA2 0.2200 3 0.00 ! LIPID alkanes
 CG321 CG321 CG324 NG3P2 1.0000 3 0.00 ! PIP, piperidine ! @@@ Kenno: 0.1950 -> 1.0000
 CG321 CG321 CG324 HGA2 0.1950 3 0.00 ! FLAVOP PIP1,2,3
 HGA2 CG321 CG324 NG3P2 0.1950 3 0.00 ! PIP, piperidine
 HGA2 CG321 CG324 HGA2 0.1950 3 0.00 ! FLAVOP PIP1,2,3
 CG321 CG321 CG3RC1 CG3C52 0.2000 3 0.00 ! BAM1, bile acid steroidal C-D ring, cacha, 02/08
 CG321 CG321 CG3RC1 CG3RC1 0.1500 3 0.00 ! CARBOCY carbocyclic sugars
 CG321 CG321 CG3RC1 HGA1 0.1500 3 0.00 ! CARBOCY carbocyclic sugars
 HGA2 CG321 CG3RC1 CG3C52 0.1950 1 0.00 ! BAM1, bile acid steroidal C-D ring, cacha, 02/08
 HGA2 CG321 CG3RC1 CG3RC1 0.1500 3 0.00 ! CARBOCY carbocyclic sugars
 HGA2 CG321 CG3RC1 HGA1 0.1500 3 0.00 ! CARBOCY carbocyclic sugars
 CG321 CG324 NG3P2 CG3RC1 0.3190 1 180.00
 CG321 CG324 NG3P2 CG3RC1 2.2270 2 0.00
 CG321 CG324 NG3P2 CG3RC1 1.4290 3 180.00
 CG321 CG324 NG3P2 HGP2 0.1000 3 0.00 ! PIP, piperidine
 HGA2 CG324 NG3P2 CG3RC1 1.1970 3 0.00
 HGA2 CG324 NG3P2 HGP2 0.1000 3 0.00 ! PIP, piperidine
 HGA3 CG331 OG301 CG2R61 0.0850 3 0.00 ! MEOB, Methoxybenzene, cacha
 CG3C31 CG3C31 CG3C31 NG2R61 1.2340 3 180.00
 CG3C31 CG3C31 CG3C31 HGA1 0.1000 6 0.00 ! AMCP, aminomethyl cyclopropane; from PROTMOD
 hf/cyclopropane; jhs
 CG3C31 CG3C31 CG3C31 HGA2 0.1000 6 0.00 ! PROTMOD hf/cyclopropane
 NG2R61 CG3C31 CG3C31 HGA2 2.2320 3 180.00
 HGA1 CG3C31 CG3C31 HGA2 0.2000 5 180.00 ! AMCP, aminomethyl cyclopropane; from PROTMOD
 hf/cyclopropane; jhs
 HGA2 CG3C31 CG3C31 HGA2 0.2000 5 180.00 ! PROTMOD hf/cyclopropane
 CG3C31 CG3C31 NG2R61 CG2R62 2.3350 3 0.00
 HGA1 CG3C31 NG2R61 CG2R62 2.4770 3 0.00
 NG3C51 CG3C52 CG3RC1 CG321 1.5720 3 180.00
 NG3C51 CG3C52 CG3RC1 CG3RC1 0.9650 3 0.00
 NG3C51 CG3C52 CG3RC1 NG3P2 1.4040 3 0.00
 NG3C51 CG3C52 CG3RC1 HGA1 1.0890 3 0.00
 HGA2 CG3C52 CG3RC1 CG321 0.1950 1 0.00 ! BAM1, bile acid steroidal C-D ring, cacha, 02/08
 HGA2 CG3C52 CG3RC1 CG3RC1 0.1500 3 0.00 ! CARBOCY carbocyclic sugars
 HGA2 CG3C52 CG3RC1 NG3P2 2.4440 3 0.00
 HGA2 CG3C52 CG3RC1 HGA1 0.1500 3 0.00 ! CARBOCY carbocyclic sugars
 CG3RC1 CG3C52 NG3C51 CG2R61 1.2050 3 180.00
 CG3RC1 CG3C52 NG3C51 CG3C52 0.1800 3 0.00 ! ***** , from CG3C52 CG3C52 NG3C51 CG3C52, PENALTY= 1.1
 HGA2 CG3C52 NG3C51 CG2R61 1.6800 3 0.00
 HGA2 CG3C52 NG3C51 CG3C52 0.0000 3 0.00 ! 3PRL, 3-pyrroline, kevo
 CG321 CG3RC1 CG3RC1 CG3C52 0.1500 3 0.00 ! CARBOCY carbocyclic sugars
 CG321 CG3RC1 CG3RC1 NG3P2 0.3810 3 180.00
 CG321 CG3RC1 CG3RC1 HGA1 0.1500 3 0.00 ! CARBOCY carbocyclic sugars
 CG3C52 CG3RC1 CG3RC1 CG3C52 0.1500 3 0.00 ! CARBOCY carbocyclic sugars
 CG3C52 CG3RC1 CG3RC1 NG3P2 0.8780 3 180.00
 CG3C52 CG3RC1 CG3RC1 HGA1 0.1500 3 0.00 ! CARBOCY carbocyclic sugars
 NG3P2 CG3RC1 CG3RC1 HGA1 2.0850 3 180.00
 HGA1 CG3RC1 CG3RC1 HGA1 0.1500 3 0.00 ! CARBOCY carbocyclic sugars
 CG3C52 CG3RC1 NG3P2 CG324 1.8470 3 180.00
 CG3C52 CG3RC1 NG3P2 HGP2 1.3210 3 0.00
 CG3RC1 CG3RC1 NG3P2 CG324 0.2800 3 180.00
 CG3RC1 CG3RC1 NG3P2 HGP2 1.3090 3 0.00
 HGA1 CG3RC1 NG3P2 CG324 0.7100 3 0.00

```

HGA1 CG3RC1 NG3P2 HGP2      2.4770  3   180.00

IMPROPERs
CG2O2 CG2R62 OG2D1 OG311    53.0000  0    0.00 ! ***** , from CG2O2 CG2R61 OG2D1 OG311, PENALTY= 0.5
CG2R63 CG2R62 CG2R62 OG2D4   15.0000  0    0.00 ! 4PYO, 4(1H)-pyridinone, isg

END
RETURN

```

**Table DSXII. Optimized force field topology and parameters for neutral moxifloxacin
(CHARMM stream including CGenFF parameters)**

```

* Toppar stream file generated by
*
read rtf card append
* Initial topologies generated by
* CHARMM General Force Field (CGenFF) program version 1.0.0
* For use with CGenFF version 3.0.1
*
36 1
=====
! Moxifloxacin - Neutral
=====
```

| RESI | MOXO | 0.000 |
|-------|------|-----------------------|
| GROUP | | ! CHARGE CH_PENALTY |
| ATOM | F | FGR1 -0.145 |
| ATOM | O1 | OG301 -0.391 ! 2.180 |
| ATOM | O2 | OG2D4 -0.425 |
| ATOM | O3 | OG311 -0.513 ! 4.025 |
| ATOM | O4 | OG2D1 -0.437 ! 3.884 |
| ATOM | N1 | NG3C51 -0.094 |
| ATOM | N2 | NG311 -0.761 |
| ATOM | N3 | NG2R61 -0.177 |
| ATOM | C1 | CG3RC1 -0.086 |
| ATOM | C2 | CG3RC1 0.018 |
| ATOM | C3 | CG3C52 -0.019 |
| ATOM | C4 | CG3C52 0.096 |
| ATOM | C5 | CG321 -0.178 ! 3.307 |
| ATOM | C6 | CG3C31 0.094 |
| ATOM | C7 | CG321 -0.171 |
| ATOM | C8 | CG3C31 -0.180 ! 7.029 |
| ATOM | C9 | CG3C31 -0.180 ! 7.029 |
| ATOM | C10 | CG321 0.078 |
| ATOM | C11 | CG2R61 -0.150 |
| ATOM | C12 | CG2R62 0.027 |
| ATOM | C13 | CG2R61 0.216 |
| ATOM | C14 | CG2R62 0.097 |
| ATOM | C15 | CG2R66 -0.135 |
| ATOM | C16 | CG2R62 0.016 ! 8.806 |
| ATOM | C17 | CG2R61 0.040 ! 6.996 |
| ATOM | C18 | CG2R62 0.261 |
| ATOM | C19 | CG2R63 0.026 |
| ATOM | C20 | CG202 0.314 |
| ATOM | C21 | CG331 -0.100 ! 0.025 |
| ATOM | H1 | HGA1 0.090 ! 2.026 |
| ATOM | H2 | HGA1 0.090 ! 3.009 |
| ATOM | H3 | HGP1 0.429 ! 0.219 |
| ATOM | H4 | HGPAM1 0.342 ! 3.537 |
| ATOM | H5 | HGA2 0.090 ! 1.375 |
| ATOM | H6 | HGA2 0.090 ! 1.375 |
| ATOM | H7 | HGA2 0.090 ! 1.596 |
| ATOM | H8 | HGA2 0.090 ! 1.596 |
| ATOM | H9 | HGA2 0.090 ! 0.000 |
| ATOM | H10 | HGA2 0.090 ! 0.000 |
| ATOM | H11 | HGA1 0.090 ! 4.940 |
| ATOM | H12 | HGA2 0.090 ! 0.450 |
| ATOM | H13 | HGA2 0.090 ! 0.450 |
| ATOM | H14 | HGA2 0.090 ! 2.500 |
| ATOM | H15 | HGA2 0.090 ! 2.500 |
| ATOM | H16 | HGA2 0.090 ! 2.500 |
| ATOM | H17 | HGA2 0.090 ! 2.500 |
| ATOM | H18 | HGA2 0.090 ! 2.500 |
| ATOM | H19 | HGA2 0.090 ! 2.500 |
| ATOM | H20 | HGR62 0.111 ! 4.793 |
| ATOM | H21 | HGR62 0.177 ! 0.253 |
| ATOM | H22 | HGA3 0.090 ! 0.000 |
| ATOM | H23 | HGA3 0.090 ! 0.000 |
| ATOM | H24 | HGA3 0.090 ! 0.000 |

BOND F C15
BOND O1 C13
BOND O1 C21
BOND O2 C19
BOND O3 C20
BOND O4 C20
BOND N1 C3
BOND N1 C4
BOND N1 C11
BOND N2 C2
BOND N2 C10
BOND N3 C6
BOND N3 C12
BOND N3 C16
BOND C1 C2
BOND C1 C3
BOND C1 C5
BOND C1 H1
BOND C2 C4
BOND C2 H2
BOND C5 C7
BOND C6 C8
BOND C6 C9
BOND C7 C10
BOND C8 C9
BOND C11 C13
BOND C11 C15
BOND C12 C13
BOND C12 C14
BOND C14 C17
BOND C14 C19
BOND C15 C17
BOND C16 C18
BOND C18 C19
BOND C18 C20
BOND O3 H3
BOND N2 H4
BOND C3 H5
BOND C3 H6
BOND C4 H7
BOND C4 H8
BOND C5 H9
BOND C5 H10
BOND C6 H11
BOND C7 H12
BOND C7 H13
BOND C8 H14
BOND C8 H15
BOND C9 H16
BOND C9 H17
BOND C10 H18
BOND C10 H19
BOND C16 H20
BOND C17 H21
BOND C21 H22

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BOND C21 H23
BOND C21 H24
IMPR C19    C14     C18     O2
IMPR C20    C18     O4      O3

END

read param card flex append
* Parameters generated by analogy by
* CHARMM General Force Field (CGenFF) program version 2.2.0
*
! Penalties lower than 10 indicate the analogy is fair; penalties between 10
! and 50 mean some basic validation is recommended; penalties higher than
! 50 indicate poor analogy and mandate extensive validation/optimization.

BONDS
CG2O2 CG2R62 254.00   1.4800 ! ***** , from CG2O2 CG2R61, PENALTY= 5
CG2O2 OG2D1  750.00   1.2200 ! PROT adm jr. 5/02/91, acetic acid pure solvent; LIPID methyl acetate
CG2O2 OG311  230.00   1.4000 ! PROT adm jr. 5/02/91, acetic acid pure solvent
CG2R61 CG2R61 305.00   1.3750 ! PROT benzene, JES 8/25/89
CG2R61 CG2R62 394.00   1.3750 ! YTHY, 2,4(1H,3H)-quinazolinedione, isg
CG2R61 CG2R66 305.00   1.3700 ! NAMODEL difluorotoluene
CG2R61 NG3C51 433.937  1.330
CG2R61 OG301  230.00   1.3820 ! COMPDS peml
CG2R61 HGR62  340.00   1.0800 ! NA, DFT
CG2R62 CG2R62 420.00   1.3500 ! NA nad/ppi, jjpl/adm jr. 7/95
CG2R62 CG2R63 302.00   1.4030 ! NA T, adm jr. 11/97
CG2R62 NG2R61 302.00   1.3430 ! NA C, adm jr. 11/97
CG2R62 HGR62  350.00   1.0900 ! NA C,U, JWK
CG2R63 OG2D4  660.00   1.2340 ! NA U,A,G par_a4 adm jr. 10/2/91
CG2R66 FGR1   400.00   1.3580 ! NAMODEL difluorotoluene
CG321 CG321  222.50   1.5300 ! PROT alkane update, adm jr., 3/2/92
CG321 CG3RC1 222.50   1.5240 ! CARBOCY carbocyclic sugars
CG321 NG311  263.00   1.4740 ! AMINE aliphatic amines
CG321 HGA2   309.00   1.1110 ! PROT alkane update, adm jr., 3/2/92
CG331 OG301  360.00   1.4150 ! diethylether, alex
CG331 HGA3   322.00   1.1110 ! PROT alkane update, adm jr., 3/2/92
CG3C31 CG3C31 240.00   1.5010 ! PROTMOD cyclopropane
CG3C31 NG2R61 259.453  1.372
CG3C31 HGA1   340.00   1.0830 ! PROTMOD cyclopropane
CG3C31 HGA2   340.00   1.0830 ! PROTMOD cyclopropane
CG3C52 CG3RC1 222.50   1.5240 ! CARBOCY carbocyclic sugars
CG3C52 NG3C51 400.00   1.4780 ! PRLD, pyrrolidine; 2PRL, 2-pyrroline, kevo
CG3C52 HGA2   307.00   1.1000 ! THF, THF neutron diffr., 5/30/06, viv
CG3RC1 CG3RC1 222.50   1.5230 ! CARBOCY carbocyclic sugars
CG3RC1 NG311  248.581  1.399
CG3RC1 HGA1   309.00   1.1110 ! CARBOCY carbocyclic sugars
NG311 HGPAM1 447.80   1.0190 ! AMINE aliphatic amines
OG311 HGP1   545.00   0.9600 ! PROT EMB 11/21/89 methanol vib fit; og tested on MeOH EtOH, ...

ANGLES
CG2R62 CG2O2 OG2D1  70.00   123.10  20.00   2.44200 ! ***** , from CG2R61 CG2O2 OG2D1, PENALTY= 0.5
CG2R62 CG2O2 OG311  40.00   113.90  30.00   2.37000 ! ***** , from CG2R61 CG2O2 OG311, PENALTY= 0.5
OG2D1 CG2O2 OG311  50.00   123.00  210.00  2.26200 ! PROT adm jr, 10/17/90, acetic acid vibrations
CG2R61 CG2R61 CG2R62 40.00   119.00  35.00   2.41620 ! YTHY, 2,4(1H,3H)-quinazolinedione, isg

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CG2R61 CG2R61 CG2R66 40.00 119.00 35.00 2.41620 ! NAMODEL difluorotoluene
 CG2R61 CG2R61 NG3C51 40.00 120.00 ! ***** , from CG2R61 CG2R61 NG311, PENALTY= 3
 CG2R61 CG2R61 OG301 110.00 120.00 ! BIPHENYL ANALOGS, peml
 CG2R62 CG2R61 CG2R66 40.00 119.00 35.00 2.41620 ! ***** , from CG2R61 CG2R61 CG2R66, PENALTY= 0.5
 CG2R62 CG2R61 OG301 110.00 120.00 ! ***** , from CG2R61 CG2R61 OG301, PENALTY= 0.5
 CG2R62 CG2R61 HGR62 30.00 120.00 22.00 2.15250 ! ***** , from CG2R61 CG2R61 HGR62, PENALTY= 0.5
 CG2R66 CG2R61 NG3C51 40.00 120.00 ! ***** , from CG2R61 CG2R61 NG311, PENALTY= 4.5
 CG2R66 CG2R61 HGR62 30.00 121.50 22.00 2.15250 ! NAMODEL difluorotoluene
 CG2O2 CG2R62 CG2R62 10.00 131.80 ! ***** , from CG2O1 CG2R62 CG2R62, PENALTY= 2.5
 CG2O2 CG2R62 CG2R63 10.00 131.80 ! ***** , from CG2O1 CG2R62 CG2R62, PENALTY= 4
 CG2R61 CG2R62 CG2R62 40.00 121.00 ! RIN, coumarin, isg
 CG2R61 CG2R62 CG2R63 120.00 122.30 ! YTHY, 2,4(1H,3H)-quinazolinedione, isg
 CG2R61 CG2R62 NG2R61 23.00 116.10 ! YTHY, 2,4(1H,3H)-quinazolinedione, isg
 CG2R62 CG2R62 CG2R63 120.00 116.70 ! NA T
 CG2R62 CG2R62 NG2R61 85.00 122.90 ! NA C
 CG2R62 CG2R62 HGR62 42.00 119.00 ! NA nadh/ppi, jjp1/adm jr. 7/95
 NG2R61 CG2R62 HGR62 44.00 115.00 ! NA C, h6
 CG2R62 CG2R63 CG2R62 10.00 120.80 ! 4PYO, 4(1H)-pyridinone; from CG2R62 CG2R62 CG2R62; isg
 CG2R62 CG2R63 OG2D4 100.00 124.60 ! NA T, o4
 CG2R61 CG2R66 CG2R61 40.00 122.50 35.00 2.41620 ! NAMODEL difluorotoluene
 CG2R61 CG2R66 FGR1 60.00 118.75 ! NAMODEL difluorotoluene
 CG321 CG321 CG321 58.35 113.60 11.16 2.56100 ! PROT alkane update, adm jr., 3/2/92
 CG321 CG321 CG3RC1 53.35 111.00 8.00 2.56100 ! CARBOCY carbocyclic sugars
 CG321 CG321 NG311 43.70 110.00 ! K2Cn, cgenff_compromise, kevo
 CG321 CG321 HGA2 26.50 110.10 22.53 2.17900 ! PROT alkane update, adm jr., 3/2/92
 CG3RC1 CG321 HGA2 34.50 110.10 22.53 2.17900 ! CARBOCY carbocyclic sugars
 NG311 CG321 HGA2 32.40 109.50 50.00 2.13000 ! PEI polymers, kevo
 HGA2 CG321 HGA2 35.50 109.00 5.40 1.80200 ! PROT alkane update, adm jr., 3/2/92
 OG301 CG331 HGA3 45.90 108.89 ! MEOB, Methoxybenzene, cacha
 HGA3 CG331 HGA3 35.50 108.40 5.40 1.80200 ! PROT alkane update, adm jr., 3/2/92
 CG3C31 CG3C31 CG3C31 77.35 111.00 8.00 2.56100 ! PROTMOD cyclopropane
 CG3C31 CG3C31 NG2R61 89.072 120.511
 CG3C31 CG3C31 HGA1 23.00 117.10 22.53 2.17900 ! PROTMOD cyclopropane
 CG3C31 CG3C31 HGA2 23.00 117.10 22.53 2.17900 ! PROTMOD cyclopropane
 NG2R61 CG3C31 HGA1 12.460 104.276
 HGA2 CG3C31 HGA2 23.00 117.00 5.40 1.80200 ! PROTMOD cyclopropane
 CG3RC1 CG3C52 NG3C51 84.00 107.60 ! ***** , from CG3C52 CG3C52 NG3C51, PENALTY= 1.1
 CG3RC1 CG3C52 HGA2 34.50 110.10 22.53 2.17900 ! CARBOCY carbocyclic sugars
 NG3C51 CG3C52 HGA2 54.00 109.00 !v 107.7 PRLD, pyrrolidine; 110.8 2PRL, 2-pyrroline; 110.4 3PRL, 3-pyrroline; 111.4 2IMI, 2-imidazoline; 111.7 2PRZ, 2-pyrazoline, kevo
 HGA2 CG3C52 HGA2 38.50 106.80 5.40 1.80200 ! THF, 10/17/05 viv
 CG321 CG3RC1 CG3C52 58.35 113.50 11.16 2.56100 ! BAM1, bile acid steroidal C-D ring, cacha, 02/08
 CG321 CG3RC1 CG3RC1 53.35 111.00 8.00 2.56100 ! CARBOCY carbocyclic sugars
 CG321 CG3RC1 HGA1 34.50 110.10 22.53 2.17900 ! CARBOCY carbocyclic sugars
 CG3C52 CG3RC1 CG3RC1 53.35 111.00 8.00 2.56100 ! CARBOCY carbocyclic sugars
 CG3C52 CG3RC1 NG311 82.560 114.667
 CG3C52 CG3RC1 HGA1 34.50 110.10 22.53 2.17900 ! CARBOCY carbocyclic sugars
 CG3RC1 CG3RC1 NG311 77.024 111.797 8.00 2.5610
 CG3RC1 CG3RC1 HGA1 34.50 110.10 22.53 2.17900 ! CARBOCY carbocyclic sugars
 NG311 G3RC1 HGA1 42.880 109.227
 CG2R62 NG2R61 CG2R62 30.00 120.00 ! NA nadh/ppi, jjp1/adm jr. 7/95
 CG2R62 NG2R61 CG3C31 45.00 118.40 ! ***** , from CG2R62 NG2R61 CG3C51, PENALTY= 6.9
 CG321 NG311 CG3RC1 107.450 114.812 5.00 2.4217
 CG321 NG311 HGPAM1 35.00 111.00 ! compromise between PEI0 on the one hand and OBTZ AOBT on the other hand, kevo & xxwy
 CG3RC1 NG311 HGPAM1 89.535 107.894

| | | | | |
|--------|--------|--------|--------|--|
| CG2R61 | NG3C51 | CG3C52 | 48.288 | 107.844 |
| CG3C52 | NG3C51 | CG3C52 | 140.00 | 103.70 !v 102.9 PRLD, pyrrolidine; 105.4 3PRL, 3-pyrroline, kevo |
| CG2R61 | OG301 | CG331 | 65.00 | 108.00 ! MEOB, Methoxybenzene, cacha |
| CG2O2 | OG311 | HGP1 | 55.00 | 115.00 ! PROT adm jr. 5/02/91, acetic acid pure solvent |

DIHEDRALS

| | | | | |
|--------|--------|--------|--------|--|
| OG2D1 | CG2O2 | CG2R62 | CG2R62 | 1.0250 2 180.00 ! ***** , from OG2D1 CG2O2 CG2R61 CG2R61, PENALTY= 5.5 |
| OG2D1 | CG2O2 | CG2R62 | CG2R63 | 1.0250 2 180.00 ! ***** , from OG2D1 CG2O2 CG2R61 CG2R61, PENALTY= 8 |
| OG311 | CG2O2 | CG2R62 | CG2R62 | 1.0250 2 180.00 ! ***** , from OG311 CG2O2 CG2R61 CG2R61, PENALTY= 5.5 |
| OG311 | CG2O2 | CG2R62 | CG2R63 | 1.0250 2 180.00 ! ***** , from OG311 CG2O2 CG2R61 CG2R61, PENALTY= 8 |
| CG2R62 | CG2O2 | OG311 | HGP1 | 0.9750 1 180.00 ! ***** , from CG2R61 CG2O2 OG311 HGP1, PENALTY= 0.5 |
| CG2R62 | CG2O2 | OG311 | HGP1 | 2.7000 2 180.00 ! ***** , from CG2R61 CG2O2 OG311 HGP1, PENALTY= 0.5 |
| CG2R62 | CG2O2 | OG311 | HGP1 | 0.0500 3 180.00 ! ***** , from CG2R61 CG2O2 OG311 HGP1, PENALTY= 0.5 |
| CG2R62 | CG2O2 | OG311 | HGP1 | 0.2500 6 180.00 ! ***** , from CG2R61 CG2O2 OG311 HGP1, PENALTY= 0.5 |
| OG2D1 | CG2O2 | OG311 | HGP1 | 2.0500 2 180.00 ! PROT adm jr, 10/17/90, acetic Acid C-OH rotation barrier |
| CG2R62 | CG2R61 | CG2R61 | CG2R66 | 3.1000 2 180.00 ! ***** , from CG2R61 CG2R61 CG2R61 CG2R66, PENALTY= 0.5 |
| CG2R62 | CG2R61 | CG2R61 | NG3C51 | 3.1000 2 180.00 ! ***** , from CG2R61 CG2R61 CG2R61 NG311, PENALTY= 3.5 |
| CG2R66 | CG2R61 | CG2R61 | OG301 | 3.1000 2 180.00 ! ***** , from CG2R61 CG2R61 CG2R61 OG301, PENALTY= 1.5 |
| NG3C51 | CG2R61 | CG2R61 | OG301 | 2.4440 2 180.00 |
| CG2R61 | CG2R61 | CG2R62 | CG2R62 | 0.5000 2 180.00 ! RIN, coumarin, isg |
| CG2R61 | CG2R61 | CG2R62 | NG2R61 | 7.0000 2 180.00 ! YTHY, 2,4(1H,3H)-quinazolinedione, isg |
| CG2R66 | CG2R61 | CG2R62 | CG2R62 | 0.5000 2 180.00 ! ***** , from CG2R61 CG2R61 CG2R62 CG2R62, PENALTY= 1.5 |
| CG2R66 | CG2R61 | CG2R62 | CG2R63 | 3.1000 2 180.00 ! ***** , from CG2R61 CG2R61 CG2R62 CG2R63, PENALTY= 1.5 |
| OG301 | CG2R61 | CG2R62 | CG2R62 | 3.1000 2 180.00 ! ***** , from CG2R61 CG2R61 CG2R61 OG301, PENALTY= 5.5 |
| OG301 | CG2R61 | CG2R62 | NG2R61 | 2.4000 2 180.00 |
| HGR62 | CG2R61 | CG2R62 | CG2R62 | 3.1000 2 180.00 ! ***** , from HGR61 CG2R61 CG2R62 CG2R62, PENALTY= 1 |
| HGR62 | CG2R61 | CG2R62 | CG2R63 | 1.0000 2 180.00 ! ***** , from HGR61 CG2R61 CG2R61 CG2R62 CG2R63, PENALTY= 1 |
| CG2R61 | CG2R61 | CG2R66 | CG2R61 | 3.1000 2 180.00 ! NAMODEL difluorotoluene |
| CG2R61 | CG2R61 | CG2R66 | FGR1 | 4.5000 2 180.00 ! NAMODEL difluorotoluene |
| CG2R62 | CG2R61 | CG2R66 | CG2R61 | 3.1000 2 180.00 ! ***** , from CG2R61 CG2R61 CG2R66 CG2R61, PENALTY= 0.5 |
| CG2R62 | CG2R61 | CG2R66 | FGR1 | 4.5000 2 180.00 ! ***** , from CG2R61 CG2R61 CG2R66 FGR1, PENALTY= 0.5 |
| NG3C51 | CG2R61 | CG2R66 | CG2R61 | 3.1000 2 180.00 ! ***** , from CG2R61 CG2R61 CG2R61 NG311, PENALTY= 8 |
| NG3C51 | CG2R61 | CG2R66 | FGR1 | 2.5800 2 180.00 ! ***** , from NG311 CG2R61 CG2R61 OG3R60, PENALTY= 36.2 |
| HGR62 | CG2R61 | CG2R66 | CG2R61 | 4.2000 2 180.00 ! NAMODEL difluorotoluene |
| HGR62 | CG2R61 | CG2R66 | FGR1 | 2.4000 2 180.00 ! NAMODEL difluorotoluene |
| CG2R61 | CG2R61 | NG3C51 | CG3C52 | 1.3590 2 0.00 |
| CG2R61 | G2R61 | NG3C51 | CG3C52 | 2.4490 4 0.00 |
| CG2R66 | CG2R61 | NG3C51 | CG3C52 | 0.3300 2 180.00 |
| CG2R66 | CG2R61 | NG3C51 | CG3C52 | 0.2440 4 0.00 |
| CG2R61 | CG2R61 | OG301 | CG331 | 1.5800 2 180.00 ! MEOB, Methoxybenzene update, yxu |
| CG2R61 | CG2R61 | OG301 | CG331 | 0.2000 4 180.00 ! MEOB, Methoxybenzene update, yxu |
| CG2R62 | CG2R61 | OG301 | CG331 | 1.5800 2 180.00 ! ***** , from CG2R61 CG2R61 OG301 CG331, PENALTY= 0.5 |
| CG2R62 | CG2R61 | OG301 | CG331 | 0.2000 4 180.00 ! ***** , from CG2R61 CG2R61 OG301 CG331, PENALTY= 0.5 |
| CG2O2 | CG2R62 | CG2R62 | NG2R61 | 2.5000 2 180.00 ! ***** , from CG2O1 CG2R62 CG2R62 NG2R61, PENALTY= 2.5 |
| CG2O2 | CG2R62 | CG2R62 | HGR62 | 1.0000 2 180.00 ! ***** , from CG2O1 CG2R62 CG2R62 HGR63, PENALTY= 4.5 |
| CG2R61 | CG2R62 | CG2R62 | CG2R61 | 2.5000 2 180.00 ! RIN, coumarin, isg |
| CG2R61 | CG2R62 | CG2R62 | CG2R63 | 3.7000 2 180.00 ! YTHY, 2,4(1H,3H)-quinazolinedione, isg |
| CG2R61 | CG2R62 | CG2R62 | NG2R61 | 3.5000 2 180.00 ! YTHY, 2,4(1H,3H)-quinazolinedione, isg |
| CG2R63 | CG2R62 | CG2R62 | NG2R61 | 3.0000 2 180.00 ! NA T |
| CG2R63 | CG2R62 | CG2R62 | HGR62 | 1.0000 2 180.00 ! NA bases |
| CG2O2 | CG2R62 | CG2R63 | CG2R62 | 3.0000 2 180.00 |
| CG2O2 | CG2R62 | CG2R63 | OG2D4 | 2.4980 2 180.00 |
| CG2R61 | CG2R62 | CG2R63 | CG2R62 | 1.6000 2 180.00 ! ***** , from CG2R62 CG2R62 CG2R63 CG2R62, PENALTY= 1.5 |
| CG2R61 | CG2R62 | CG2R63 | OG2D4 | 1.0000 2 180.00 ! YTHY, 2,4(1H,3H)-quinazolinedione, isg |
| CG2R62 | CG2R62 | CG2R63 | CG2R62 | 1.6000 2 180.00 ! 4PYO, 4(1H)-pyridinone, isg |
| CG2R62 | CG2R62 | CG2R63 | OG2D4 | 1.0000 2 180.00 ! NA bases |

CG2R61 CG2R62 NG2R61 CG2R62 4.0000 2 180.00 ! ***** , from CG2R67 CG2R62 NG2R61 CG2R62, PENALTY= 0.5
 CG2R61 CG2R62 NG2R61 CG3C31 11.0000 2 180.00 ! ***** , from CG2R62 CG2R62 NG2R61 CG3C51, PENALTY= 8.4
 CG2R62 CG2R62 NG2R61 CG2R62 4.0000 2 180.00 ! NA nad/ppi, jjp1/adm jr. 7/95
 CG2R62 CG2R62 NG2R61 CG3C31 11.0000 2 180.00 ! ***** , from CG2R62 CG2R62 NG2R61 CG3C51, PENALTY= 6.9
 HGR62 CG2R62 NG2R61 CG2R62 5.6000 2 180.00 ! 4PYO, 4(1H)-pyridinone, isg
 HGR62 CG2R62 NG2R61 CG3C31 0.3000 2 180.00 ! ***** , from HGR62 CG2R62 NG2R61 CG3C51, PENALTY= 6.9
 CG321 CG321 CG321 CG3RC1 0.1500 3 0.00 ! CARBOCY carbocyclic sugars
 CG321 CG321 CG321 NG311 3.0000 1 180.00
 CG321 CG321 CG321 NG311 0.5000 2 0.00
 CG321 CG321 CG321 HGA2 0.1950 3 0.00 ! LIPID alkanes
 CG3RC1 CG321 CG321 HGA2 0.1950 3 0.00 ! LIPID alkanes
 NG311 CG321 CG321 HGA2 0.1950 3 0.00 ! K2Cn, cgenff_compromise, kevo
 HGA2 CG321 CG321 HGA2 0.2200 3 0.00 ! LIPID alkanes
 CG321 CG321 CG3RC1 CG3C52 0.2000 3 0.00 ! BAM1, bile acid steroidal C-D ring, cacha, 02/08
 CG321 CG321 CG3RC1 CG3RC1 0.1500 3 0.00 ! CARBOCY carbocyclic sugars
 CG321 CG321 CG3RC1 HGA1 0.1500 3 0.00 ! CARBOCY carbocyclic sugars
 HGA2 CG321 CG3RC1 CG3C52 0.1950 1 0.00 ! BAM1, bile acid steroidal C-D ring, cacha, 02/08
 HGA2 CG321 CG3RC1 CG3RC1 0.1500 3 0.00 ! CARBOCY carbocyclic sugars
 HGA2 CG321 CG3RC1 HGA1 0.1500 3 0.00 ! CARBOCY carbocyclic sugars
 CG321 CG321 NG311 CG3RC1 2.1190 1 0.00
 CG321 CG321 NG311 CG3RC1 2.9060 2 0.00
 CG321 CG321 NG311 CG3RC1 0.0660 3 0.00
 CG321 CG321 NG311 HGPAM1 0.3000 3 0.00 ! K2Cn, cgenff_compromise, kevo
 HGA2 CG321 NG311 CG3RC1 0.9290 3 180.00
 HGA2 CG321 NG311 HGPAM1 0.0500 3 0.00 ! PEI0, OBTZ, AOBT, kevo & xxwy
 HGA3 CG331 OG301 CG2R61 0.0850 3 0.00 ! MEOB, Methoxybenzene, cacha
 CG3C31 CG3C31 CG3C31 NG2R61 1.2340 3 180.00
 CG3C31 CG3C31 CG3C31 HGA1 0.1000 6 0.00 ! AMCP, aminomethyl cyclopropane; from PROTMOD
 hf/cyclopropane; jhs
 CG3C31 CG3C31 CG3C31 HGA2 0.1000 6 0.00 ! PROTMOD hf/cyclopropane
 NG2R61 CG3C31 CG3C31 HGA2 2.2320 3 180.00
 HGA1 CG3C31 CG3C31 HGA2 0.2000 5 180.00 ! AMCP, aminomethyl cyclopropane; from PROTMOD
 hf/cyclopropane; jhs
 HGA2 CG3C31 CG3C31 HGA2 0.2000 5 180.00 ! PROTMOD hf/cyclopropane
 CG3C31 CG3C31 NG2R61 CG2R62 2.3350 3 0.00
 HGA1 CG3C31 NG2R61 CG2R62 2.4770 3 0.00
 NG3C51 CG3C52 CG3RC1 CG321 1.5720 3 180.00
 NG3C51 CG3C52 CG3RC1 CG3RC1 0.9650 3 0.00
 NG3C51 CG3C52 CG3RC1 NG311 2.9350 3 0.00
 NG3C51 CG3C52 CG3RC1 HGA1 1.0890 3 0.00
 HGA2 CG3C52 CG3RC1 CG321 0.1950 1 0.00 ! BAM1, bile acid steroidal C-D ring, cacha, 02/08
 HGA2 CG3C52 CG3RC1 CG3RC1 0.1500 3 0.00 ! CARBOCY carbocyclic sugars
 HGA2 CG3C52 CG3RC1 NG311 0.8650 3 180.00
 HGA2 CG3C52 CG3RC1 HGA1 0.1500 3 0.00 ! CARBOCY carbocyclic sugars
 CG3RC1 CG3C52 NG3C51 CG2R61 1.2050 3 180.00
 CG3RC1 CG3C52 NG3C51 CG3C52 0.1800 3 0.00 ! ***** , from CG3C52 CG3C52 NG3C51 CG3C52, PENALTY= 1.1
 CG3RC1 CG3C52 NG3C51 CG2R61 1.2050 3 180.00
 HGA2 CG3C52 NG3C51 CG3C52 0.0000 3 0.00 ! 3PRL, 3-pyrroline, kevo
 CG321 CG3RC1 CG3RC1 CG3C52 0.1500 3 0.00 ! CARBOCY carbocyclic sugars
 CG321 CG3RC1 CG3RC1 NG311 1.6840 3 0.00
 CG321 CG3RC1 CG3RC1 HGA1 0.1500 3 0.00 ! CARBOCY carbocyclic sugars
 CG3C52 CG3RC1 CG3RC1 CG3C52 0.1500 3 0.00 ! CARBOCY carbocyclic sugars
 CG321 CG3RC1 CG3RC1 NG311 1.6840 3 0.00
 CG3C52 CG3RC1 CG3RC1 HGA1 0.1500 3 0.00 ! CARBOCY carbocyclic sugars
 NG311 CG3RC1 CG3RC1 HGA1 0.5290 3 0.00
 HGA1 CG3RC1 CG3RC1 HGA1 0.1500 3 0.00 ! CARBOCY carbocyclic sugars

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CG3C52 CG3RC1 NG311 CG321      0.5830  3     0.00
CG3C52 CG3RC1 NG311 HGPAM1    0.7110  3   180.00
CG3RC1 CG3RC1 NG311 CG321      0.6760  3     0.00
CG3RC1 CG3RC1 NG311 HGPAM1    2.9830  3   180.00
HGA1   CG3RC1 NG311 CG321      2.6450  3     0.00
HGA1   CG3RC1 NG311 HGPAM1    0.7990  3     0.00

IMPROPERs
CG2O2  CG2R62 OG2D1  OG311      53.0000  0     0.00 ! *****, from CG2O2 CG2R61 OG2D1 OG311, PENALTY= 0.5
CG2R63 CG2R62 CG2R62 OG2D4     15.0000  0     0.00 ! 4PYO, 4(1H)-pyridinone, isg

END
RETURN
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